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Cation-responsive cavity expansion of valinomycin revealed by cryogenic ion trap infrared spectroscopy†

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Valinomycin (VM) is a natural K⁺-selective ionophore that transports K⁺ through the cell membrane. VM captures K⁺ in its central cavity with a C₃-symmetric β-turn-like backbone. Although the binding affinity is drastically decreased for the VM-sodium (Na⁺VM) complex with respect to K⁺VM, VM holds relatively high affinity to Rb⁺ and Cs⁺. The high affinity for larger ions irrespective of ionic size seems to conflict with the expected optimal size matching model and raises questions on what factors determine ion selectivity. A combination of infrared spectroscopy with supporting computational calculations reveals that VM can accommodate larger Rb⁺ and Cs⁺ by flexibly changing its cavity size with the elongation of its folded β-turn-like backbone. The high affinity to Rb⁺ and Cs⁺ can be ascribed to a size-dependent cavity expansion. These findings provide a new perspective on molecular recognition and selectivity beyond the conventional size matching model.

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Introduction

Ionophores provide one mechanism for transporting ions in biological systems and in many cases with size-specificity. The transport of biologically active ions *via* the cell membrane disrupts the membrane potential, leading to cell apoptosis. This makes ionophores promising candidates as antimicrobial, antibacterial, anticancer, and antifungal agents.^{1–3} Valinomycin (VM) (Fig. 1) is a natural K⁺-selective ionophore (~10⁴ higher than Na⁺)^{4,5} consisting of six alternating ester and amide COs, and has displayed antibacterial⁶ and antiviral⁷ activities. The mechanism of the K⁺ selectivity has been extensively studied by numerous experimental methods.^{8–10} These studies all agree that when binding to a K⁺ ion, VM forms the C₃-symmetric structure, called a “bracelet”, in which K⁺ has near

octahedral coordination by six ester carbonyls (see Fig. 1b). The main chain is folded in a β-turn-like manner, in which six equivalent intramolecular hydrogen bonds are formed between the amide NH group in the *i*-th residue and the amide CO group in the (*i* + 3)-th residue. All the hydrophobic side chains point outwardly, suggesting that the structure is adopted inside hydrophobic lipid membranes when carrying an ion.

The ion selectivity of ionophores has been traditionally described by the optimal size matching model.^{11,12} In this model, an ionophore strongly binds to a metal when the cavity size of the ionophore is a close match to the ionic size of the metal. A representative example is a well-known K⁺-selective ionophore, 18-crown-6 ether,^{11,13,14} though the ion selectivity

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Fig. 1 (a) Valinomycin. (b) Bracelet structure of K⁺VM. Color codes: gray = C, blue = N, red = O, white = H, and purple = K. Blue dotted lines show intramolecular H-bonds. Hydrogen atoms not involved in H-bonds were omitted for clarity.

mechanism of crown ether has not been perfectly clarified and some arguments have been raised in terms of microscopic solvation.^{15–17} This crown ether has specific bonding to K^+ (Table S1, ESI†) and this is ascribed to an optimal cavity size, matching the ionic radius for K^+ , and thus allowing K^+ to be secured inside the cavity.^{11,18} The larger ions Rb^+ and Cs^+ cannot fit inside and reside above the cavity, in contrast to the conformation of the $K^+(18\text{-crown-6})$ complex.^{11,16,17} This suboptimal binding of Rb^+ and Cs^+ leads to 5–10 times lower binding affinity in comparison to K^+ .^{11,16,17} A notable characteristic of VM is relatively high affinity to Rb^+ and Cs^+ , the equilibrium binding constant for these two ions closely brackets the value for K^+ , as shown in Table S1 (ESI†).⁴ Infrared (IR) and nuclear magnetic resonance spectroscopy indicate similar values for vibrational frequencies, chemical shifts, and coupling constants among K^+ , Rb^+ , and Cs^+ , suggesting the same structural motif (bracelet structure) for these three ions.^{8,10} In this respect, the size matching model does not appear to work for VM because of the ionic size-independent affinity for larger alkali metal ions (ionic radius: $K^+ = 1.33 \text{ \AA}$, $Rb^+ = 1.48 \text{ \AA}$, $Cs^+ = 1.69 \text{ \AA}$). It would be interesting to characterize the mechanism of ion capture by VM for different-sized metal ions, which appears to be at variance to the conventional understanding of ion selectivity.

Cryogenic ion trap IR spectroscopy^{19–26} is an essential tool in determining gas-phase molecular structures of ion–ionophore complexes. This technique employs a combination of electrospray mass spectrometry and laser spectroscopy. Mass spectrometry allows the isolation of a specific ion–ionophore target molecule and eliminates contaminants such as uncomplexed VM, complexes with other ions, or higher complexes ($M^+(VM)_2$, $(M^+)_2(VM)_3$, \dots etc.). In solution-phase measurements, spectroscopic signatures of the target molecule are often perturbed by solvent molecules or contaminated by the above-mentioned undesired molecules. Cryogenic ion trap IR spectroscopy enables clear-cut spectroscopic assignments by eliminating such interference from solvents and contaminants. In fact, the gas-phase conformation of VM-potassium (hereafter denoted K^+VM) has been unambiguously assigned to the “bracelet” structure by cryogenic ion trap IR spectroscopy,²⁷ which resembles the crystal structure of K^+VM and the conformation in polar solvents. In the present study, this approach is applied to the other alkali metal ions to understand the mechanism of efficient capture of large ions (Rb^+ and Cs^+) by VM.

Experimental and theoretical methods

IR photodissociation (IRPD) spectra of M^+VM were measured in a cryogenic ion trap setup (Fig. S1, ESI†).^{26,27} In short, methanol solutions of VM (Wako, 10^{-5} M) and the desired alkali metal chloride (Wako, $2 \times 10^{-5} \text{ M}$) were electrosprayed *via* a glass capillary heated to $60 \text{ }^\circ\text{C}$ (where the solvent is driven off), and the ion–neutral complexes were allowed to enter the vacuum apparatus. The specific ions of interest were mass-selected by a quadrupole mass spectrometer, deflected by a

quadrupole bender, and then guided into a cryogenic quadrupole ion trap (QIT). The QIT was cooled to 4 K by a closed cycle He refrigerator. A mixture of hydrogen (20%) and helium buffer gas was introduced to the QIT *via* a pulsed nozzle and cooled down by collisions with the QIT’s gold-coated copper electrodes. The complex ions were trapped and cooled to $\sim 10 \text{ K}$ by collisions with helium gas. Hydrogen molecules were condensed onto the cold ions, forming a variety of weakly-bound hydrogen-attached cluster ions. The cluster ions were then irradiated with a tunable IR laser. Absorption of a photon triggers the dissociation of the weakly-bound hydrogen molecules from the cluster ions, yielding the parent M^+VM complex ion as a photofragment. The IRPD spectrum of the trapped ions, which is negligibly affected by the attached hydrogen, was measured by monitoring the fragment yield, recorded by a time-of-flight mass spectrometer as a function of the wavenumber of the IR laser, thus providing the IR absorption spectrum of the M^+VM complex.

Calculated vibrational frequencies presented in this study were obtained from density functional theory (DFT) calculations using the dispersion-corrected B3LYP-D3BJ functional and the lanl2dz (Rb, Cs)/6-31G(d,p) (C, N, O, H) basis sets.²⁷ The DFT calculations were performed using the Gaussian 16 package.²⁸ Initial structures were prepared, based on the geometry data of the crystal structures of K^+VM (bracelet-type conformation⁹) and bare VM (twisted bracelet- and propeller-type conformations^{29,30}), and are shown in Fig. S2 (ESI†). All the optimized structures were confirmed to be local minima with no imaginary vibrational frequencies. All of the calculated frequencies have been corrected for anharmonicity with a scaling factor of 0.961, identical to the previous report on the K^+VM complex.²⁷ This value was a good compromise between the fingerprint region (slightly underestimated) and the NH stretch regions (slightly overestimated) in K^+VM .²⁷ The calculated IR spectra were convoluted with a Lorentz lineshape function with a half width of 3 cm^{-1} . The binding energy (E_{BE}) between metal and VM was calculated at the B3LYP-D3BJ/def2-SVP level using eqn (1).

$$E_{BE} = (G_M + G_{VM}) - (G_{MVM} + BSSE) \quad (1)$$

The BSSE value corresponds to the correction term of basis set superposition error (BSSE). This value was calculated by the counterpoise method. G_M , G_{VM} , and G_{MVM} represent Gibbs free energy for a metal ion, VM, and the complex of the metal and VM at 298 K, respectively.

Results and discussion

Fig. 2 and Fig. S3 (ESI†) display IRPD spectra of bare VM-alkali metal ion complexes. The observed bands were assigned based on previous studies²⁷ of K^+VM and its monohydrated complexes: NH bends at $1520\text{--}1560 \text{ cm}^{-1}$, amide C=O stretches (amide I) at $1630\text{--}1690 \text{ cm}^{-1}$, ester C=O stretches at $1700\text{--}1780 \text{ cm}^{-1}$, CH stretches at $2800\text{--}3100 \text{ cm}^{-1}$, and amide NH stretches (amide A) at $3200\text{--}3400 \text{ cm}^{-1}$ (frequencies are given in Table S2, ESI†). The



Fig. 2 IRPD spectra of Li^+ , Na^+ , Rb^+ , and Cs^+ VM complexes. The calculated IR spectrum of the most stable conformer at 298 K is shown with its molecular structure for each IRPD spectrum. The black solid curves in the calculated spectra are made by convolution of stick red lines. Color codes: gray = C, blue = N, red = O, white = H, pink = Li, cyan = Na, orange = Rb, light green = Cs. The IRPD spectrum of K^+ complex is shown in Fig. S3 in ESI†

spectral features in the Li^+VM complex are clearly distinct from the other four complexes and are discussed below. The spectral features in the IRPD spectra of Na^+VM , Rb^+VM and Cs^+VM are very similar to that of K^+VM (see Fig. S3 in ESI†). Specifically, the NH stretches and ester $\text{C}=\text{O}$ stretches appear as a single peak and the amide $\text{C}=\text{O}$ bands have a main peak and a shoulder. This is in light of the presence of six NH/amide CO/ester CO bonds for each complex. This strongly suggests that M^+VM ($\text{M}^+ = \text{Na}^+$, Rb^+ , Cs^+) has a similar molecular structure to K^+VM , in which VM forms a C_3 symmetric bracelet structure with intra-molecular H-bonds. To confirm this, we performed structural optimizations and harmonic frequency calculations for M^+VM at the B3LYP-D3BJ/6-31G(d,p) level. The computation level was of sufficient accuracy to reproduce the experimental results in the IRPD spectrum of K^+VM .²⁷ For the larger ions such as Rb^+ and

Cs^+ , all the initial structures converged to the same bracelet structure as K^+VM (Fig. 2). The features of the single band for NH stretch, ester CO stretch, amide CO stretches (peak and shoulder) and NH bend are well reproduced by the calculated IR spectrum of the bracelet conformer (Fig. 2). The reason why only the amide CO stretches are split into two bands is discussed later. For the smaller Na^+ , the C_3 -symmetric bracelet-type conformer (Conf. A) was energetically most stable at 298 K, while a few metastable conformers (Conf. B–D) were obtained (Fig. 3). Conf. B resembles Conf. A, but one of the distances between Na^+ and ester ($\text{C}=\text{O}$) is elongated by ~ 0.3 Å (Table S3, ESI†) and the C_3 -symmetry is slightly broken. The simplicities of the NH and CO bands of Conf. A are in good agreement with the experimental ones. Conf. B has a splitting of the NH stretch bands, which is inconsistent with the experimental IRPD spectrum.



Fig. 3 Calculated IR spectra of conformers A–D for Na⁺VM with the experimental IRPD spectrum. Parentheses indicate the relative Gibbs free energy for each conformer at 298 K.

The other two conformers Conf. C and D also give several bands of the NH stretches, CO stretches and NH bends, deviating from the obtained IRPD spectrum. The relatively high free energies ($>30 \text{ kJ mol}^{-1}$) of Conf. C and D make their presence in the cluster ion ensemble unlikely. In the previous ion mobility measurements on Na⁺VM,³¹ the gas-phase conformation was ascribed not to the C_3 -symmetric bracelet structure (like Conf. A) but to an asymmetric one with five coordination of the ester CO to Na⁺, structurally similar to Conf. B. However, the collision cross sections for the symmetric and asymmetric conformers were the same, and the structural assignment to the asymmetric conformer was based on the conformational sampling using molecular dynamics simulations with a classical force field. This apparent inconsistency with our findings could arise from the use of the classical force field. In this study, the gas-phase conformation of Na⁺VM can be unambiguously assigned to Conf. A from the vibrational signature in the IRPD spectrum. In conclusion, VM can adopt the symmetrical “bracelet” structure regardless of ionic size from Na⁺ (0.95 Å) to Cs⁺ (1.69 Å).

A close look at the IRPD spectra reveals significant blueshifts in the NH stretches with increasing ionic size. Because the NH stretches involve the intramolecular H-bonds in the bracelet structure, the blueshift indicates a weakening of the H-bonds between amide COs and NHs, as the ionic radius increases. Another important finding is that ester CO stretches display a similar blue-shift. Because ester COs directly coordinate with the alkali metal ion, the blueshift strongly suggests weaker binding to larger ions. In fact, the calculated binding energy between VM and the metal ion also decreases for larger ions (Table S4, ESI[†]). The observed blueshifts in the NH stretch and

ester CO stretch are well reproduced by the calculated IR spectra. In the calculated bracelet-type structures, both the distances of the ester metal–O(C) and H-bonding distances of amide (N)H–O(C) are elongated with increasing ionic radius (Tables S3 and S4, ESI[†]). These results indicate that the cavity of valinomycin is expanded with the accommodation of larger ions. In other words, VM possesses some degrees of flexibility to bind ions with a wide range of size, due to its β -turn-like folded structure constructed by non-covalent H-bonds (Fig. 1).

The complete structural assignment has led us to consider the reason for the split bands of amide CO stretches despite the single bands of amide NH and ester CO stretches. The vibrational modes of these stretches are coupled (Fig. S4–S7, ESI[†]). We take the example of K⁺VM because the others have the same vibrational modes. The amide NHs are almost aligned on a plane and can be classified as D_{6h} symmetry (Fig. S4, ESI[†]). The six vibrational modes of amide NH stretches are assigned to IR-inactive a_{1g} , e_{2g} , and b_{1u} and IR-active e_{1u} modes, which is consistent with the observed single band of amide NH stretches (Fig. S5, ESI[†]). This is in parallel with the fact that only ν_{20} with e_{1u} symmetry is the IR-active CH stretch in benzene.³² On the other hand, the amide COs no longer reside on a plane and its symmetry is degraded to D_{3d} (Fig. S4, ESI[†]). The vibrational modes of amide COs are assigned to IR-active e_u , a_{2u} and IR-inactive e_g , a_{1g} modes, which well accounts for the split bands of amide CO stretches (Fig. S6, ESI[†]). The symmetry in the configuration of the ester COs can be grouped to D_{3d} as well (Fig. S4, ESI[†]) and the vibrational modes are assigned to IR-active (e_u , a_{2u}) and IR-inactive (e_g , a_{1g}) modes (Fig. S7, ESI[†]). Different from the amide CO stretches, the IR-active e_u modes of ester CO stretches are much more intense than the a_{2u} mode, which is the plausible reason for the single band of ester COs. The IR intensities depend on the magnitude of the vibrational transition dipole moment (VTM), which is due to the displacement of vibrating C=O as well as the polarization of C=O caused by the electric field (E) by a charged alkali metal ion. The latter effect becomes prominent when VTM is parallel to E , as evident in the OH stretches of Cs⁺(H₂O)₁.³³ The VTMs in the e_u modes are closely parallel to E generated by K⁺ (Fig. S7, ESI[†]), which well accounts for the enhancement of the e_u modes.

For the case of Li⁺VM, the amide NH and ester/amide CO bands split into several bands, indicating the breakdown of the C_3 symmetry. The conformational search of Li⁺VM provided an energetically stable conformer A in which Li⁺ is coordinated by four ester COs, losing the C_3 symmetry. Other conformers are obtained as metastable species though they have high energy gaps ($>20 \text{ kJ mol}^{-1}$) to the most stable Conf. A (Fig. 4 and Fig. S8, ESI[†]). The calculated IR spectra for the conformers with $\Delta G < 30 \text{ kJ mol}^{-1}$ are shown in Fig. 4. The spectral pattern in the Li⁺VM matches closely to the calculated IR spectrum of Conf. A. The calculated IR spectra of the other conformers do not match as well; Conf. B does not match the split of amide CO stretches; Conf. C does not match the intensity pattern of the NH stretches; the NH stretches in Conf. D are considerably blue-shifted compared to the experimental spectrum. The conformation of the gas-phase Li⁺VM is thus ascribed to Conf. A.

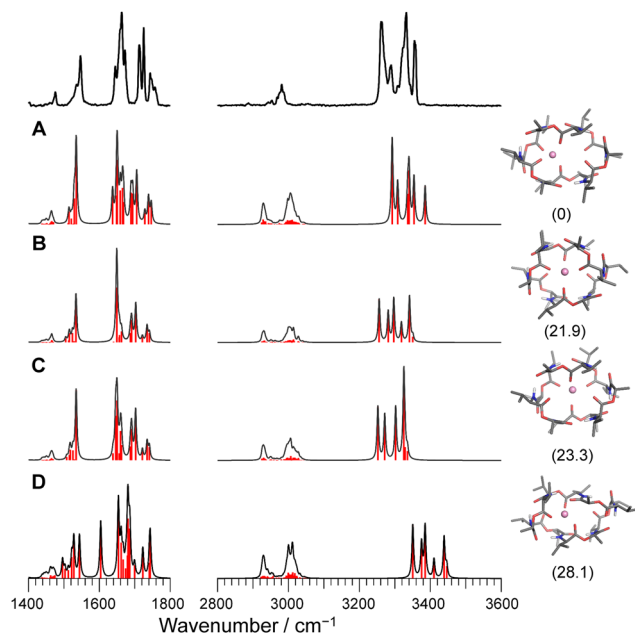


Fig. 4 Calculated IR spectra of conformers A–D for Li^+VM with the experimental IRPD spectrum. Parentheses indicate the relative Gibbs free energy for each conformer at 298 K.

The fact that Li^+VM does not adopt the C_3 -symmetric bracelet structure, but a distorted structure (Conf. A), is consistent with the low affinity of VM to Li^+ (Table S1, ESI[†]).

Conclusion

In summary, we have measured IRPD spectra of M^+VM ($\text{M} = \text{Li}, \text{Na}, \text{Rb}, \text{Cs}$) and found that VM can capture Na^+ , Rb^+ , and Cs^+ with the six ester COs forming the C_3 -symmetric “bracelet” structure. This symmetric structure was also observed in K^+VM .²⁷ The ability to capture different-sized metals with the same structural motif relies on the structural flexibility of VM. VM forms an intramolecular H-bond within its folded β -turn-like backbone. This affords flexibility to VM and enables VM to have the same bracelet motif regardless of ionic size ($\text{Na}^+ - \text{Cs}^+$). This bracelet structure is the same as the proposed structure of the Rb^+VM and Cs^+VM complexes in polar solvents.^{8,10} The conformational resemblance suggests minimal perturbations to the conformations of Rb^+VM and Cs^+VM gas-phase structures when placed in the solvent environment. The relatively high affinity of VM to Rb^+ and Cs^+ in polar solvents can be explained by the cation-size dependent cavity expansion of VM. On the other hand, the proposed conformation for Na^+VM in polar solvents indicates less than six-fold coordination to Na^+ ,¹⁰ deviating from the gas-phase structure. This strongly suggests that the low affinity of VM to Na^+ (Table S1, ESI[†]) arises from the solvation effect on the conformation of Na^+VM . This will be addressed in the near future by the structural analysis for the solvated clusters of Na^+VM , which will be performed by incorporating a second ion trap for the generation of solvated clusters.^{27,34} For the smallest Li^+ , VM no longer holds C_3

symmetry with less than six-fold coordination of the ester COs. This also explains the low affinity between VM and Li^+ . Conventionally, the selectivity in molecular recognition has been described by optimal size matching, in which a guest molecule strongly binds to a host molecule when the size of the guest pairs with the size of the host. In contrast, VM can flexibly change its cavity size to fit to the various sizes of metal ions. These findings shed new light on the molecular recognition in host–guest chemistry and demonstrate the potential of cryogenic ion trap IR spectroscopy to clarify unique properties of molecular recognition in host–guest supramolecules.

Conflicts of interest

There are no conflicts of interest to declare.

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References

- 1 B. C. Pressman, *Annu. Rev. Biochem.*, 1976, **45**, 501–530.
- 2 V. Kaushik, J. S. Yakisich, A. Kumar, N. Azad and A. K. V. Iyer, *Cancers*, 2018, **10**, 360.
- 3 Q. Wu, J. Patocka and K. Kuca, *Mini-Rev. Med. Chem.*, 2019, **19**, 206–214.
- 4 R. M. Izatt, J. S. Bradshaw, S. A. Nielsen, J. D. Lamb, J. J. Christensen and D. Sen, *Chem. Rev.*, 1985, **85**, 271–339.
- 5 A. Casnati, A. Pochini, R. Ungaro, C. Bocchi, F. Ugozzoli, R. J. M. Egberink, H. Struijk, R. Lugtenberg, F. De Jong and D. N. Reinhoudt, *Chem. – Eur. J.*, 1996, **2**, 436–445.
- 6 S. Huang, Y. Liu, W. Q. Liu, P. Neubauer and J. Li, *Microorganisms*, 2021, **9**, 780.
- 7 D. Zhang, Z. Ma, H. Chen, Y. Lu and X. Chen, *Biomed. J.*, 2020, **43**, 414–423.
- 8 D. J. Patel, *Biochemistry*, 1973, **12**, 496–501.
- 9 K. Neupert-Laves and M. Dobler, *Helv. Chim. Acta*, 1975, **58**, 432–442.
- 10 F. Wang, C. Zhao and P. L. Polavarapu, *Biopolymers*, 2004, **75**, 85–93.
- 11 C. J. Pedersen and H. K. Frensdorff, *Angew. Chem., Int. Ed. Engl.*, 1972, **11**, 16–25.
- 12 S. Varma, D. Sabo and S. B. Rempe, *J. Mol. Biol.*, 2008, **376**, 13–22.

- 13 C. J. Pedersen, *J. Am. Chem. Soc.*, 1967, **89**, 2495–2496.
- 14 C. J. Pedersen, *J. Am. Chem. Soc.*, 1967, **89**, 7017–7036.
- 15 J. D. Rodriguez, T. D. Vaden and J. M. Lisy, *J. Am. Chem. Soc.*, 2009, **131**, 17277–17285.
- 16 J. D. Rodriguez and J. M. Lisy, *J. Am. Chem. Soc.*, 2011, **133**, 11136–11146.
- 17 Y. Inokuchi, T. Ebata, T. R. Rizzo and O. V. Boyarkin, *J. Am. Chem. Soc.*, 2014, **136**, 1815–1824.
- 18 R. M. Izatt, R. E. Terry, D. P. Nelson, Y. Chan, D. J. Eatough, J. S. Bradshaw, L. D. Hansen and J. J. Christensen, *J. Am. Chem. Soc.*, 1976, **98**, 7626–7630.
- 19 O. V. Boyarkin, S. R. Mercier, A. Kamariotis and T. R. Rizzo, *J. Am. Chem. Soc.*, 2006, **128**, 2816–2817.
- 20 N. S. Nagornova, T. R. Rizzo and O. V. Boyarkin, *Science*, 2012, **336**, 320–323.
- 21 E. Garand, M. Z. Kamrath, P. A. Jordan, A. B. Wolk, C. M. Leavitt, A. B. McCoy, S. J. Miller and M. A. Johnson, *Science*, 2012, **335**, 694–698.
- 22 J. Jašík, J. Žabka, J. Roithová and D. Gerlich, *Int. J. Mass Spectrom.*, 2013, **354–355**, 204–210.
- 23 J. G. Redwine, Z. A. Davis, N. L. Burke, R. A. Oglesbee, S. A. McLuckey and T. S. Zwier, *Int. J. Mass Spectrom.*, 2013, **348**, 9–14.
- 24 M. Broquier, S. Soorkia and G. Grégoire, *Phys. Chem. Chem. Phys.*, 2015, **17**, 25854–25862.
- 25 Y. Inokuchi, K. Soga, K. Hirai, M. Kida, F. Morishima and T. Ebata, *J. Phys. Chem. A*, 2015, **119**, 8512–8518.
- 26 S. Ishiuchi, H. Wako, D. Kato and M. Fujii, *J. Mol. Spectrosc.*, 2017, **332**, 45–51.
- 27 E. Sato, K. Hirata, J. M. Lisy, S. Ishiuchi and M. Fujii, *J. Phys. Chem. Lett.*, 2021, **12**, 1754–1758.
- 28 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson and H. Nakatsuji, *et al.*, *Gaussian 16*, Gaussian, Inc.; Wallingford, CT, 2016.
- 29 I. L. Karle, *J. Am. Chem. Soc.*, 1975, **97**, 4379–4386.
- 30 I. L. Karle and J. L. Flippen-Anderson, *J. Am. Chem. Soc.*, 1988, **110**, 3253–3257.
- 31 T. Wyttenbach, J. J. Batka, J. Gidden and M. T. Bowers, *Int. J. Mass Spectrom.*, 1999, **193**, 143–152.
- 32 R. H. Page, Y. R. Shen and Y. T. Lee, *J. Chem. Phys.*, 1988, **88**, 5362–5376.
- 33 J. M. Lisy, *Int. Rev. Phys. Chem.*, 1997, **16**, 267–289.
- 34 B. M. Marsh, J. M. Voss and E. Garand, *J. Chem. Phys.*, 2015, **143**, 204201.