



Cite this: *Soft Matter*, 2022, 18, 2500

Received 2nd January 2022,
Accepted 9th March 2022

DOI: 10.1039/d2sm00005a

rsc.li/soft-matter-journal

Weak polyelectrolyte brushes

Mark Geoghegan 

I review experimental developments in the growth and application of surface-grafted weak polyelectrolytes (brushes), concentrating on their surface, tribological, and adhesive and bioadhesive properties, and their role as actuators.

1 Introduction

A polymer brush is a layer of surface-grafted polymers. These are generally end-grafted, but that does not need to be the case. By chemically attaching the chains to a surface, the layer can remain stable even in a good solvent. This gives them an important advantage over films cast from solvents in coatings technologies. Further benefits arise, for example, in colloidal stabilization due to the limited swelling available for the

polymers, which are already extended in forming a brush.¹ Hydrophilic polymer brushes are particularly useful in aqueous media to combat fouling.^{2,3}

Polyelectrolytes are a class of polymers which contain ionized or ionizable groups. Examples of weak polyelectrolytes include poly(methacrylic acid) [PMAA] and poly(2-vinylpyridine) (P2VP) whereas polystyrene sulfonic acid and poly(allylamine hydrochloride) are examples of strong polyelectrolytes. The strong acid and base retain their charge in aqueous solution over a wide range of pH, whilst the weak acid (here, PMAA) is neutral at low pH and the weak base (P2VP) is neutral at high pH. The much more pH-sensitive behaviour of weak polyelectrolytes has the potential for significant utility in nanoscience because the charged state of a polyelectrolyte brush is intimately related to its conformation. Change the charge and, under many conditions, the conformation changes too. This conformational change brings with it a concomitant capacity to do mechanical work, which offered great promise to the nascent field of soft nanotechnology at the beginning of the millennium.

Weak polyelectrolytes have been known as “smart” polymers for some time due to their pH responsive behaviour. For example, PMAA may be used as a coating in some oral medications because it can protect the drug from the harsh acidic environment of the stomach.⁴ In the gut, the pH is significantly higher, and the acid can dissolve, releasing the cargo. Similarly, dried hydrogels of sodium polyacrylate are used in nappies (diapers), and their capacity for absorbance allows for a five-hundred-fold increase in mass before the hydrogels become saturated and leak.

This review will cover the different synthesis routes of weak polyelectrolyte brushes and their characterization. Different topologies and their importance will also be discussed. Since polyelectrolytes have a rich conformational space covered by pH and added salt,⁵ some effort will be spent covering how these parameters control brush conformation. The physical

School of Engineering, Newcastle University, Merz Court, Newcastle-upon-Tyne NE1 7RU, UK. E-mail: mark.geoghegan@newcastle.ac.uk



Mark Geoghegan

Professor Mark Geoghegan has held the Roland Cookson Chair in Engineering Materials at Newcastle University since 2019. He obtained his BA in Physics at the University of Oxford in 1988 and an MSc on cold gaseous plasmas at the University of Birmingham in 1990. He undertook his PhD at the University of Cambridge under the supervision of (now) Professor Richard Jones. He submitted his doctoral thesis on polymer blend films in 1994. He spent two years at CEA-Saclay

working as a postdoc with Dr François Boué before being awarded a Humboldt fellowship at the Universität Freiburg. Two years later, he moved to Bayreuth to undertake another postdoctoral fellowship in Bayreuth under the supervision of Professor Georg Krausch. In 2000 he moved to the University of Sheffield where he remained for nineteen years, being awarded a personal chair in 2011.



properties of the surfaces thus created through contact mechanics will be covered. The review will conclude with a discussion of the various applications of polyelectrolyte brushes, where actuation is required. Applications in areas of bioadhesion are of growing importance and these will also be included. Related areas of research involving strong polyelectrolytes and polyzwitterions are not covered.

2 Synthetic routes

Brushes may either be made *in situ* (grafting from) or the polymer may be synthesized first and then a functional end group used to attach the polymer to the surface (grafting to). Both have advantages and disadvantages. The grafting-to method allows for much better characterization of the polymer, because it lends itself to techniques such as gel permeation chromatography and light scattering to find the size of the polymer prior to grafting. However, it is difficult to obtain dense and thick brushes using grafting-to techniques because the first chains to arrive at the surface form a mushroom conformation (Fig. 1).⁶ At a grafting certain density (the mushroom-to-brush transition), polymers face an entropic barrier preventing them from reaching the substrate, because they need to elongate to navigate through the chains already at the surface. Possible routes to increase the density of brushes grafted to the substrate are discussed below, as are attempts to characterize brushes grown from the substrate.

Grafting-from techniques

The use of controlled polymerizations dominates the synthesis of polymer brushes grafted from surfaces. These are used because of their anticipated low dispersities. Free radical polymerization has been used with success to produce weak polyelectrolyte brush layers,⁷ but controlled polymerization methods are not significantly less convenient given that much of the effort in growing brushes involves depositing a suitable initiating monolayer.

Atom-transfer radical polymerization. ATRP is probably the most convenient way of growing brushes from surfaces due to

the relatively undemanding synthetic requirements.⁸ ATRP is a controlled radical polymerization in which the growing polymer is dormant until it transfers a halide to a metal salt, activating the polymer (Scheme 1) which rapidly reacts with the next monomer before being temporarily capped again. The long-lived dormant state allows for controlled growth, encouraging a relatively uniform polymer and restricting reactions between active end groups, which could otherwise be a problem in dense brushes.

The direct synthesis of polyelectrolyte brushes using ATRP is routinely performed with polycations, such as poly[2-(dimethylamino)ethyl methacrylate] (PDMAEMA)⁹ or poly[2-(diethylamino)ethyl methacrylate] (PDEAEMA).¹⁰ On the other hand, the growth of carboxylic polyanions in this manner is not easily achieved because they easily associate with the catalyst.¹¹ Standard polyanions such as poly(acrylic acid) (PAA) and PMAA can be synthesized by polymerizing *tert*-butyl acrylate or *tert*-butyl methacrylate followed by a hydrolysis step.^{8,12} However, it has been shown that it is possible to grow PMAA brushes directly by using methacrylic acid sodium salt.^{13,14}

The main development of ATRP is that of Activators ReGenerated by Electron Transfer (ARGET) ATRP, which reduces the quantity of heavy metal catalyst required by the addition of a reducing agent, often ascorbic acid. This agent reduces Cu(II) to Cu(I), which is the active catalyst. Since oxygen is generally responsible for the oxidation to Cu(II), the sensitivity of ATRP to oxygen is reduced, which is another advantage of the ARGET technique. ARGET ATRP is not routinely used in the production of polyelectrolyte brushes and it has been demonstrated that the tertiary amine groups present in PDMAEMA and, one supposes, PDEAEMA act effectively as reducing agents.¹⁵

New developments in ATRP have largely passed by those interested in the synthesis of polyelectrolyte brushes. For example, Initiators for Continuous Activator Regeneration (ICAR) ATRP is another method of synthesizing polymers in an aqueous environment¹⁶ and (like ARGET ATRP) reducing the amount of metal catalyst. This has not been embraced by those creating polyelectrolyte brushes. Indeed the metal catalyst can be eliminated using more specialized forms of ATRP,¹⁷ but this is currently rather niche. Ultimately, however, metal catalysts have an environmental footprint and create problems regarding the cleaning of the brushes. If ATRP is to be routinely used in biological systems, then perhaps more effort will need to be directed towards reducing or eliminating the metal catalysts that are used.

Nitroxide-mediated polymerization. More formally known as aminoxyl-mediated radical polymerization,¹⁸ NMP is a common route to creating brushes from vinyl monomers, such as styrene.^{19,20} It is similar to ATRP in that it uses the reversible

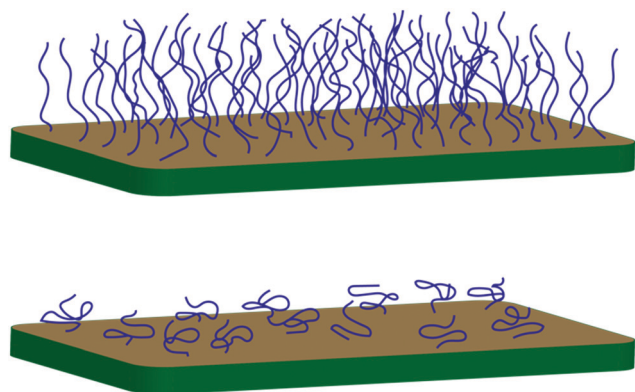
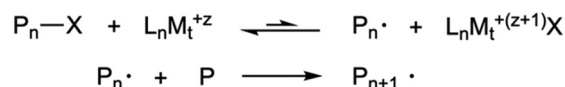
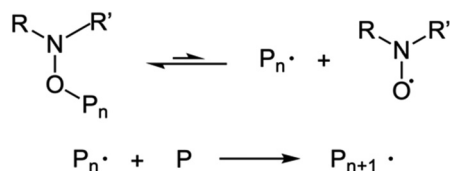


Fig. 1 If the polymer grafting points are further away from each other than the size of the polymer chain in a self-avoiding work configuration, the polymers exhibit a mushroom conformation (bottom). If they are closer together, then a brush conformation is observed (top).



Scheme 1 ATRP process. The polymer is activated by the metal (M_t) complexed ligand (L_n), which allows the chain to grow.



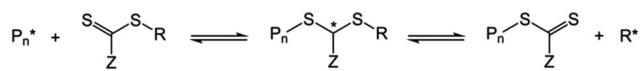


Scheme 2 NMP process.

activation and deactivation of growing polymer chains, although here it requires nitroxide radicals (Scheme 2). It has been used for the synthesis of both anionic and cationic polyelectrolyte brushes^{21–23} and has the useful advantage of not requiring a metal catalyst. It does however require elevated temperatures, generally above 100 °C. The initiating group is a nitroxide, which attaches reversibly to the chain end. The attachment is far faster than the detachment. However, when the nitroxide detaches, the remaining carbon radical (the growing polymer chain) reacts with further monomers before being capped again.^{19,24}

Reversible addition–fragmentation chain transfer. Like ATRP and NMP, RAFT is a controlled radical polymerization. Unlike ATRP and NMP, the deactivation process in RAFT releases a radical that can initiate another polymerization reaction. This deactivation process is provided by the chain transfer agent, which is in substantial excess of a standard (free radical) initiator (Scheme 3).²⁵ (For brushes the initiator is usually immobilized on the surface, with the chain transfer agent free in the solution.) The deactivation process is rapid, so most growing chains are dormant, which gives the reaction a controlled nature. RAFT is readily amenable to the synthesis of polymers that can be synthesized by free radical polymerization and do not undergo side reactions with the chain transfer agent. Acrylates and acrylamides are commonly grown with this method. Polycation brushes of PDMAEMA have been synthesized by RAFT,^{26,27} but the use of RAFT for the growth of weak polyelectrolytes has not so often been reported, although recent developments may change this; for example, oxygen-initiated RAFT shows some promise.^{28,29}

Advantages and disadvantages. RAFT and ATRP both have a large range of polymers that can be synthesized, and many solvent environments can be accommodated too. ATRP does not require extreme levels of purity and can be performed under ambient conditions. Weaknesses commonly associated with ATRP such as air sensitivity or a need to limit metal catalyst use can be addressed by a derivative technique. For these reasons it is anticipated that ATRP will dominate the synthesis of weak polyelectrolyte (and other) brushes in the foreseeable future. Nevertheless, the creation of mixed or binary brushes,^{30,31} which comprise two or more distinct

Scheme 3 RAFT process. The chain transfer agent reacts with an activated radical or growing polymer to extend the chain. The notation here follows that of Perrier.²⁵

polymers to form the brush, and (for the purposes of this review) contain at least one weak polyelectrolyte component can readily be achieved by grafting-to methods.^{32,33} Grafting-from approaches of mixed brushes can be achieved using different syntheses: for example ATRP and NMP has been used to create mixed brushes of poly(acrylic acid) and polystyrene.^{34,35} (Binary brushes can be created using only ATRP, by the use of a protecting species for some of the initiator followed by its photoremoval to allow the second polymer to be grown.³⁶) Weak polyelectrolyte brushes that can be synthesized by RAFT can also be synthesized by ATRP. RAFT chain transfer agents are quite expensive, which may well be why ATRP is favoured, although it is probable that a community used to using ATRP sees no need to change. RAFT does have some appeal for the synthesis of block copolymers, due to the relative ease of creating uniform materials, and this indeed has been demonstrated with a diblock copolymer brush containing a weak polyelectrolyte block.²⁶

Although many researchers prefer to use their favoured techniques, the development of chemical routes to grafting polymers from surfaces is ongoing. Clean chemistries that limit the use of metals are particularly under development and whichever dominates in the future will depend on such benefits as well as reliability and ease of synthesis. Of course, new developments are by no means restricted to the routes discussed here.³⁷

Alternatives to grafting-from techniques

Although individual chains grafted to surfaces do not usually present dense brush layers due to the entropic barrier inherent in grafting-to approaches, it is possible to graft multiple chains to a surface. This has been demonstrated by the creation of combs of PDMAEMA pre-synthesized from a poly(methyl methacrylate) (PMMA) backbone. After deposition in a Langmuir trough, the hydrophobic PMMA sits on the water surface, while the PDMAEMA chains extend into the water.³⁸ This layer can be compressed to achieve a desired grafting density and can be later deposited on a substrate of choice to create a brush layer.

An alternative approach involves the self-assembly of polymers to form micelles with a polyelectrolyte corona. This has been achieved with diblock copolymers of the temperature-responsive polymer poly(*N*-isopropylacrylamide) (PNIPAm) and PDMAEMA. Here the polymers can self-assemble above 40 °C to form micelles with the hydrophobic PNIPAm block forming the core of the micelle and the PDMAEMA a rather dense corona.³⁹ P2VP coronas have been grown from polyferrocenyldimethylsilane cores⁴⁰ using crystallization-driven self-assembly.⁴¹ These micellar brushes form by hydrogen bonding or electrostatic interaction to a variety of surfaces and have been functionalized to demonstrate, for example, the separation of an oil from an aqueous emulsion.

The biopolymer hyaluronic acid is grown *in vivo* using a hyaluronan synthase enzyme. This process has been replicated to form hyaluronic acid brushes, which have been shown to reach layers of up to 15 μm in thickness,⁴² which is perhaps fifty times greater than the thickest brush layers readily grown by standard synthetic routes.



Characterization

The characterization of polymer brushes requires primarily confirmation that the intended polymer is grown on the surface; the molar mass of the polymer; and the grafting density. A measurement of the thickness of the brush in the dry state links the molar mass, M_w , and grafting density, σ , by

$$h = \frac{\sigma M_w}{\rho N_A},$$

where N_A is the Avogadro constant and ρ the density of the polymer. If h is known, then only one of M_w or σ needs to be measured to determine the other. Direct measurement of grafting density is extremely difficult,⁴³ so grafting density is usually calculated from knowledge of the molar mass, which is not in itself easy to determine. A common route to obtaining the molar mass is to grow the polymer *ex situ* (from an unconstrained initiator in solution) under the same conditions as the brush and assume that these constitute the same polymer.⁸ This has been questioned from computer simulations which reveal that brushes grow faster in solution than from surfaces.⁴⁴ Experimental work on a variety of polymers show that PMMA grown in solution can be up to ~30% longer than those densely grafted from the surface.⁴⁵ For less dense polymers there is little difference in molar mass between solution-grown and surface-grafted polymers. The most accurate means of measuring molar mass is to de-graft the polymer and characterize it using standard techniques,^{45,46} but this method requires large substrates to obtain enough polymer to use for chromatographic measurements.

It is possible to measure the polymer molar mass *in situ* using an atomic force microscope (AFM). Here, the AFM tip is retracted from the surface of the polymer brush in solution. The length of tip retraction before it detaches from the brush can be correlated with the molar mass of the polymer. This technique is only suitable for solvent-swollen brushes and has had some success with neutral brushes.⁴⁷ Although this AFM-based method (force spectroscopy) can be readily applied to weak polyelectrolyte brushes, it is more challenging because of the effect of pH^{23,48} and ionic strength^{48,49} on polymer conformation. Experiments on weak polyacids found good agreement between gel permeation chromatography data on polymers grown from free initiators and force spectroscopy on brushes at neutral pH. However, at low pH the collapsed brushes presented complex spectra and at high pH the highly charged brushes adhered poorly to the AFM tip.²³ Poor adhesion of polycationic brushes to AFM tips was proposed as a reason for difficulty in determining the conformation in high ionic strength solutions.⁴⁸ These polycationic brushes (PDMAEMA) were grafted from a poly(methyl methacrylate) macroinitiator using ATRP,³⁸ and could be characterized by size-exclusion chromatography and so lend themselves well to a comparison of this nature. The pH dependence of PDMAEMA chain length determined by force spectroscopy,⁴⁸ gave a similar result to that for PAA.²³ This means that force spectroscopy can be used to provide a determination of polyelectrolyte molar mass (and dispersity) providing that the polymers are soluble

but not too charged, which typically means near neutral pH. This unsatisfactory recommendation is compounded by the best PAA measurements being obtained in 100 mM NaCl in order to control polymer–AFM tip interactions.²³

Ultimately, force spectroscopy works as a means of chain length (molar mass) characterization, but so much preparation is needed that calibration is critical for such a route to be pursued. Given the difficulties involved, if chromatographic characterization of the product of a synthesis from free initiator is not undertaken, then it is common for grafted-from polymers to be assumed to take a surface density of 0.5 chains per nm², following from earlier work.^{20,50} This assumption is not absurd and can reasonably be applied to polyelectrolyte monomers. If the monomer is not overly bulky, then assuming a 0.5 chains per nm² surface density is reasonable, given that the grafting density is set early in the synthesis and can only be reduced if the initiator layer is compromised, which may be by a protecting species,^{51–54} by mixing the initiator layer with inactive species,⁵⁵ by using a Langmuir–Blodgett trough,^{56–58} or by otherwise reducing surface initiator concentration.⁵⁰

Conformation

Brushes are defined by their density and molar mass. A polymer generally undertakes a self-avoiding random walk in a good solvent. This means that its size scales as $N^{0.6}$,^{59,60} where N is the number of steps (often taken to mean monomers). This self-avoiding walk creates a greater power law dependence than a standard random walk ($N^{0.5}$) due to excluded volume interactions between the monomers. A surface perturbs this (self-avoiding) random walk only a little, by ensuring that the grafted chain end lies at the edge of the polymer. This is the ‘mushroom’ conformation. When the grafting points are separated by less than the width of the mushroom (Fig. 1), the chains become compressed and are forced away from the substrate to form brushes.^{61,62} The height of the brush is then expected to increase monotonically with grafting density.

Polyelectrolyte brushes also increase in thickness with increased grafting density, but, given that solvent quality is dramatically affected by pH and salt concentration, there are other parameters to consider when discussing their conformation.⁶³ Changes in pH generally cause swelling or collapse in weak polyelectrolyte brushes,^{7,64–66} but the addition of salt complicates an analysis of their conformation.

Salt contributes more ions to the brush in addition to those already there. Given the accepted definition of *counterion* in colloid chemistry,⁶⁷ salt ions with charges opposite to those of the polymer chain can also act as counterions. However, when considering chemical equilibrium, there is an important difference, because, by adding salt, some “native” counterions may be displaced from the brush, which may in turn cause the polyelectrolyte to become more charged. At low salt concentrations, chains do become more ionized and this results in an increase in brush swelling.⁶⁸ Here, each chain can be considered as a series of blobs of a Debye length in size. Within each blob the chain exhibits salt-free behaviour, but on larger length scales the chains are extended to counter the addition of salt, causing



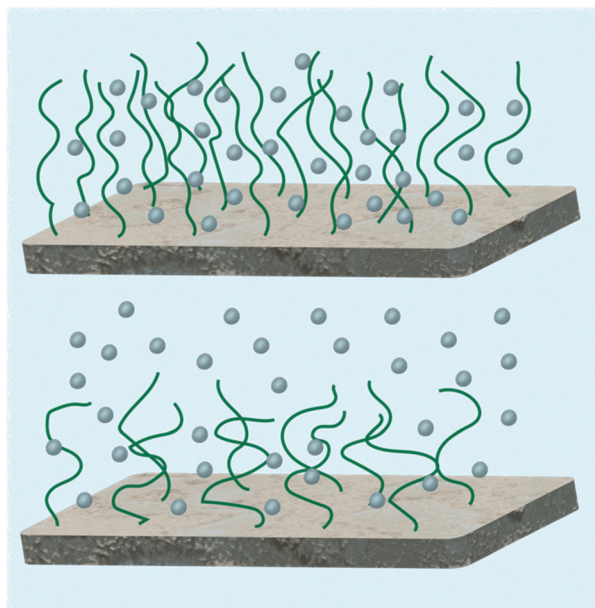


Fig. 2 Osmotic brushes (top) retain counterions within the brush whereas Pincus brushes (bottom) do not, but have a layer of counterions above the brush.

an osmotic swelling. This behaviour has been given a fuller theoretical foundation based on self-consistent field theory.⁶⁹ These brushes are referred to as osmotic brushes, but for brushes with reduced grafting density (or less charge) osmotic effects are reduced and the brush is often referred to as a *Pincus brush* (Fig. 2).

For Pincus brushes, the shielding increases with increasing ion concentration, which acts to reduce the osmotic pressure in the brush and cause a concomitant reduction in thickness.^{70,71}

At large ionic strength, the shielding of charges is such that excluded volume interactions dominate electrostatic behaviour.⁵ This is known as the quasi-neutral brush regime because the presence of charges is irrelevant to the polymer conformation. Experimentally, there are examples of abrupt brush collapse at salt concentrations approaching 1 M.^{72–76} Other experiments present data that show a more gradual decline in brush thickness over two orders of magnitude of increasing salt concentration,^{7,77–80} again up to 1 M salt. With these conflicting results, it is only possible to assert that weak polyelectrolyte brushes tend to have a collapsed structure at high salt concentrations.

Osmotic brushes have been proposed to grow with ionic strength as $I^{1/3}$,⁶⁸ but experimental support for this prediction is limited⁷ with other studies offering a power law exponent of 0.1 or less.^{72,73,75–77,79,80} The origin of this small exponent is not clear and is likely to depend on several factors. For example, the behaviour may be restricted to a limited range of salt concentration, which makes an accurate assessment of power law or other behaviour difficult. Furthermore, brush swelling and collapse are known to depend on the nature of the salt used. While experiments on PMAA showed qualitatively the same behaviour for different salt cations,⁸⁰ different polycations exhibited different behaviours when the anions were varied.^{78,81,82}

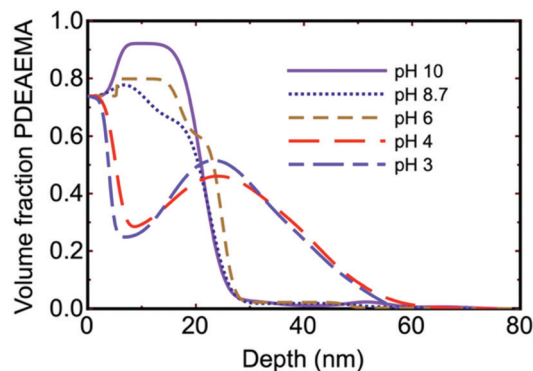


Fig. 3 Brush conformation as a function of pH for a PDEAEMA brush as obtained using neutron reflectometry.⁸³ The dry brush is 13 nm thick. The silicon substrate is equivalent to a volume fraction of 0.74.

In the polycation studies brush collapse was more pronounced for chaotropic ions.

Although brush conformation as a function of pH is expected to exhibit a monotonic increase in thickness as the polymers become more solvated due to increased charging, experiments show some subtlety in composition-depth profiles obtained by neutron reflectometry. As an example (Fig. 3), neutron reflectometry experiments on PDEAEMA⁸³ and PDMAEMA³⁸ brushes showed unusual composition depth profiles, whereby the brush concentration increased with distance from the substrate. Different explanations have been provided for this, noting that, in the middle of the film, there is increased polymer swelling, *i.e.*, a reduced polymer volume fraction. Consequently, there is a greater polymer density both close to the substrate and towards the solution interface. These regions of less swelling (greater polymer density) neighbour either the substrate or the bulk medium, which have fewer charges and therefore limit chain stretching.⁸⁴ Alternatively, it could be considered that the brush contained intrinsic entanglements that caused knotting as the brush swells,^{85,86} rather like trying to comb greasy hair. This would leave a high density of brush near the solution interface if swelling were initiated at the substrate. This explanation was provided for the earlier measurements of brushes created by ATRP from initiator-covered wafers,⁸³ but it is not clear that it can explain the conformation of polymers created from a hydrophobic macroinitiator.³⁸ It has also been proposed that hydrophobic interactions caused kinetic trapping of the brush near the solvent interface.⁸⁷ Recent experiments on DMAEMA copolymerized with a non-polyelectrolyte, 2-(2-methoxyethoxy)ethyl methacrylate, also show similar concentration profiles,^{88,89} and the behaviour has even been observed using ellipsometry⁹⁰ and a quartz crystal microbalance (QCM)⁹¹ for PDMAEMA and PAA respectively. There have been other attempts to explain the volume fraction profiles of weak polyelectrolytes, with good comparison being made between self-consistent field theory⁹² and neutron reflectometry experiments.⁹³ These neutron experiments did not reveal pronounced swelling in the middle of the brush, however. Theoretical work and simulations suggest that solvent entropy is a factor that needs greater consideration.^{63,94}



Other experiments of PDMAEMA concentration profiles have also shown non-monotonic behaviour that may be related to the measurements described above. Certainly, a strong adsorption of the brush at the substrate is noted,⁹⁵ which is common to other measurements.^{83,89} (This strong adsorption may be electrostatic as it is present on sapphire surfaces at pH 9, but not at pH 3,⁹⁶ which straddle the substrate isoelectric point.⁹⁷) Given that, the conformation of weak polyelectrolyte brushes as a function of pH is not fully understood, a full understanding of the effects of salt is still some way off.

Electric fields

First experiments on PDMAEMA brushes showed that an electric field applied perpendicular to the brush could control its thickness.⁹⁹ When a positive voltage was applied to the substrate (relative to the aqueous medium above the brush), brushes were swollen. This potential at the substrate repelled the polycations, causing swelling. Subsequent experiments using PDEAEMA, but a different experimental geometry produced similar findings (Fig. 4).⁹⁸ In this case the data were interpreted as showing that the electric field effectively changed the pH, which is reasonable, although it is to be anticipated that the removal of the electro-neutrality requirement in an electric field may cause additional effects such as a conformational phase separation of the brushes,¹⁰⁰ a phenomenon that is expected to increase with grafting density.¹⁰¹

The behaviour of weak polyelectrolyte brushes in electric fields is sensitive to different parameters, and so comparing different experiments is rather tricky. It is noted, for example, that small changes in solution pH have resulted in a significant difference in the applied potential to bring about conformational change in PAA brushes.^{102,103} This was interpreted as being due to the distance from the pK_a ,¹⁰³ but full studies of the effect of grafting density or chain molar mass are necessary to be confident of any interpretation.

Biological brushes

Biomacromolecules often have complex architectures, but sometimes this complexity hides simple phenomena. For example, the pH-dependent behaviour of *Rhodococcus* sp. RC291 attached to a colloidal probe on an AFM cantilever has been successfully

modelled by considering that the extracellular substances act like a weak polyelectrolyte brush.¹⁰⁴

Given the insights into biomacromolecules that may arise from synthetic weak polyelectrolytes, it is worth noting that hyaluronic acid brushes, for example, have been created that can be analysed using the expected behaviour for polyelectrolytes,⁴² but other work has shown that hyaluronic acid on the surface of chondrocytes is much less tractable to simple modelling.¹⁰⁵ Nevertheless, hyaluronic acid brushes have been the subject of numerous studies.^{106–110} It is perhaps worth noting that the salt dependence of initially swollen hyaluronic acid brushes was more like that of strong polyelectrolytes,¹⁰⁶ which may simply reflect the low pK_a (≈ 3.0) of hyaluronic acid.¹¹¹ Biomacromolecular weak polyelectrolyte brushes are not restricted to polysaccharides, with one study of a protein brush demonstrating a pH dependence of height commensurate with a weak polyacid.¹¹² Generally, however, proteins exhibit more complicated behaviour due to their polyampholytic nature.¹¹³

Contact mechanics

Friction and lubrication. The hydrated nature of swollen polymer brushes conveys remarkable properties of lubrication, with polyelectrolytes well known for this quality. The first demonstration of polyelectrolyte lubrication was for a strong polyelectrolyte,¹¹⁴ and the best results have been for polyzwitterionic brushes.^{115,116} In many circumstances it is not important which is used as the important point is the hydration of the layer. However, under compression and shear stress, weak polyelectrolytes will not retain their charge so effectively. Nevertheless, weak polyelectrolytes do have lubrication qualities controlled by pH or the addition of salt, whereas lubrication in strong polyelectrolytes depends on the nature of the counterions.¹¹⁷

Friction is an energy loss process when two surfaces move relative to each other. Its description is largely attributed to da Vinci, but it was formalized at the end of the seventeenth century, some 200 years later, by Amontons.¹¹⁸ This is the well-known law that the force due to friction is linearly proportional to the load applied. Limitations are shown on the nanoscale where Amontons' law needs to be supplemented by terms that are dependent upon the contact area.¹¹⁹ Such area-dependent behaviour is readily tested for brushes in contact with a nanoscale asperity such as an AFM tip. Here, the frictional force is the sum of a load dependent term and a shear term. This area dependence is not visible in the load dependent term because the contact area scales with load. The shear term includes, for example, lateral forces and so effectively describes deviations from Amontons' law.

For weak polyelectrolytes, there is a variation across pH, which provides information about the mechanical properties of the brush. The brush may be analysed in terms of Amontons' law, or alternatively a formalized contact mechanics approach may be used. If the interaction is soft and elastically deformable, the JKR (Johnson–Kendall–Roberts) approach¹²¹ is used, whereas for more rigid or stiff interactions, an approach based on DMT

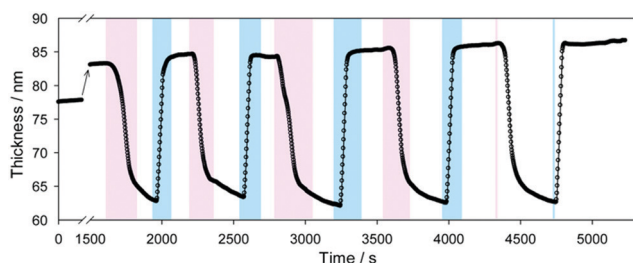


Fig. 4 The swelling and collapse of a PDEAEMA brush as an applied voltage across the brush is changed from +5 V (pink) to −5 V (blue). Figure reproduced with permission from the original article.⁹⁸ Copyright 2013 American Chemical Society.



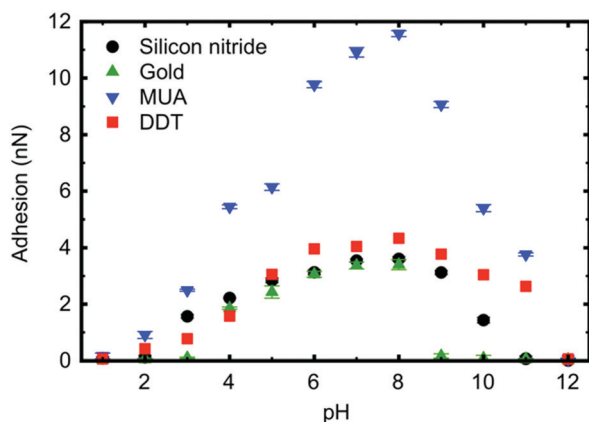


Fig. 5 The adhesion of a PDMAEMA brush to a silicon nitride AFM tip as received or coated with mercaptoundecanoic acid (MUA), dodecanethiol (DDT), or gold. This is the maximum pull-off force as the AFM tip retracts from the brush. The figure is taken from the original article,¹²⁰ and is reproduced under a CC BY 3.0 licence.

(Derjaguin–Muller–Toporov) theory¹²² is required. Both models have their origin in Hertzian mechanics¹²³ corrected for adhesion and can be seen as limiting behaviours, which may be unified.¹²⁴

In intermediate pH solutions, a PDMAEMA brush has a soft elastic contact with AFM tips coated with hydrophobic and hydrophilic monolayers (Fig. 5).¹²⁰ These experiments were performed using a form of scanning force microscopy known as friction force microscopy.^{125–127} The extremes of pH are harder to predict, with mercaptoundecanoic acid-coated tips exhibiting elastic behaviour in contact with the brush over $3 \leq \text{pH} \leq 11$, but the PDMAEMA brush presents a stiffer interaction (DMT behaviour) in contact with a dodecanethiol-coated tip at low pH. Furthermore, a gold-coated tip increasingly exhibits DMT behaviour with added salt, the concentration dependence of which is different for different anions.⁸¹ At the extremes of pH (1, 2, and 12), these PDMAEMA brushes obey Amontons' law.

Other experiments have shown a linear relationship between force and applied load for gold coated colloidal probes of 10 μm nominal diameter.¹²⁸ Such discrepancies show the importance of geometry and length scale in measuring friction.

The FFM approach to friction is consistent with an adhesion-based approach where Hertzian (non-adhesive) contact mechanics is equivalent to Amontons' behaviour once a correction^{129,130} is made for the multiple asperities that are important for the linear relationship between force and load predicted by Amontons. The shear term of the friction behaviour is mirrored by adhesive terms, which are dependent upon the elasticity of the layer.¹²⁴

Mechanical properties

Little work has been done to understand the mechanical properties of weak polyelectrolyte brushes partly because of their perceived inaccessibility to experimental investigation and partly because there is little work on the mechanical properties of polymer brushes in general. The mechanical properties of a polymer film are largely dominated by its

substrate, and it is difficult to find any application of these brushes whereby the properties of the film are separate from those of the substrate. However, it can often be necessary for the mechanical properties of the film to match the substrate, especially when the substrate itself is soft, *e.g.*, for bioengineering applications.^{131,132}

The most important mechanical property that may be measured is an elastic modulus, which is obtained using indentation tests. Measurements using an AFM are possible, but determining the area of contact is challenging, although for non-adhesive contacts and relatively thick films, a methodology is available.¹³³ Measurements on thin films (~ 50 nm) of poly[(aminopropyl)siloxane], a weak polyelectrolyte,¹³⁴ have been performed by interfacial force microscopy (a form of nanoindentation), assuming a non-adhesive contact.¹³⁵ These measurements have all been performed in dry conditions, and so do not test the responsive (pH-dependent) properties of the brush.

Assessing the mechanical properties of brushes is intrinsically difficult because they are determined by perturbing the film, which is complicated by the film often being a similar thickness to the depth of any indentation. In such cases the substrate contributes to the measured modulus, causing an overestimation of the modulus for substrates harder than the film and underestimating it for relatively soft substrates. This problem was exemplified by work considering the mechanical properties of PAA brushes using a colloidal probe technique.¹³⁶ Here, without substrate correction, a PAA brush in mildly acidic conditions (pH 4.0) had a greater (Young's) modulus than at neutral pH. With correction, the behaviour was inverted, with the PAA brush modulus increasing from ~ 200 Pa at pH 4.0 to ~ 1.1 kPa at pH 7.4. Consequently, this highly hydrated brush has much greater incompressibility than that at pH 4, which is supported by earlier QCM measurements.¹³⁷

Experiments in which charged brushes have a smaller modulus than neutral brushes have been reported for both a polycation and polyanion, where a surface forces apparatus was used.¹³⁸ Here, similar brushes were brought into contact, and the compression was measured. In this case very large compressibility moduli of up to 40 MPa were obtained, which is a surprising result. Also, that the charged brush had the lower modulus, while again surprising, is not inconsistent with the recognition that elasticity can vary at the extremes of pH, depending on how the system is measured.^{120,139} It is therefore unwise to declare that the modulus of such nanoscale systems is solely dependent upon pH, because it is also dependent on the experimental geometry of the measurement being performed.

Actuation

The pH-dependent conformational transition of weak polyelectrolytes lends them well to applications as nanoactuators. After all, a chain that can change from being collapsed to swollen can do mechanical work. This is not a new subject and it is not restricted to polyelectrolytes,¹⁴⁰ but weak polyelectrolytes are attractive candidate materials for exploiting such behaviour. The response of polyelectrolyte brushes to changes in pH is



usually not rapid, but it can be made to occur on the time scale of a few minutes.¹⁴¹

An appealing and simple demonstration of the capacity of weak polyelectrolyte brushes to actuate is by placing them in an oscillatory chemical reaction.¹⁴² Here a chemical reaction occurs away from equilibrium, where the products of one reaction produce reactants for the reverse process. When this occurs in a redox system, protons are produced and consumed, realizing pH-oscillatory behaviour. This has been demonstrated for PAA brushes by QCM.¹³⁷ Oscillatory chemical reactions can be put into reverse by a process of mechanotransduction. Here, work is done on the brushes to cause a chemical effect. Some work has been done on strong polyelectrolytes,^{143–145} but this subject has remained largely dormant in recent years.

By charging or neutralizing a brush-coated surface, it is possible to control electrochemical behaviour at that surface.¹⁴⁶ A charged PMAA brush has been shown to attract redox probes,¹⁴⁷ which is a route for electron transfer. This feature can be removed by lowering pH.

The actuating possibilities of weak polyelectrolyte brushes were postulated as giving them a significant role as components in a “molecular toolbox for soft nanotechnology”.¹² The realization of this vision has (at least thus far) failed and it is worth speculating why. Polyelectrolyte brushes are nanoscale objects and therefore need to be addressed at the nanoscale. They need to be carefully grown using rather routine synthetic techniques, for which experiments at scale have not been performed. The work that a single polymer chain has been shown to produce during a collapse and expansion cycle is of the order of 2 eV,¹⁴⁸ which limits potential applications. Nevertheless, it is still possible that early promise of polyelectrolyte brushes for adhesion could still be fulfilled and furthermore, responsive polymers have significant applications in biosensing.^{149,150}

Adhesion

Different categories of adhesives include structural adhesives,¹⁵¹ including, for example, epoxies or cyanoacrylates, for high-load applications and pressure-sensitive adhesives,^{152,153} where a high density of van der Waals interactions gives sufficient strength for temporary and low-load bonds. Coulombic interactions are not commercially important in adhesives, but charged polymers have two important advantages. They are predominantly water-based and furthermore, they are environmentally controllable.

Water-based adhesion of soft materials had historically been rather difficult to quantify,¹⁵⁴ but these problems have been addressed in recent years and experiments are now quite tractable.^{155–157} The first experiments showed that a polycationic brush would adhere to a polyanion (PMAA) hydrogel in water and that, when the pH was reduced, the adhesion would fail.¹⁵⁶ Deductive reasoning using contact mechanics experiments with oppositely charged polyelectrolyte brushes suggests that this adhesion was dominated by electrostatic forces.¹³⁹ These forces are so strong that their failure in water is cohesive, *i.e.* one of the components fractures rather than the adhesive interface itself.¹⁵⁸ However, replacing the PMAA gel with a double-network

hydrogel created a robust system that could continuously be attached and detached in water without switching pH.¹⁵⁹

Other attempts to use brushes for adhesive applications have used different bonding. The pH-dependent adhesion of PAA with poly(*N,N*-dimethylacrylamide) (PDMAc) hydrogels was attributed to hydrogen bonding. Electrostatic interactions are readily eliminated since PDMAc is not a polybase even if it often exhibits pH dependent behaviour.¹⁶⁰ Reversible adhesion has been shown to also work with strong polyelectrolyte¹⁵⁵ and polyzwitterionic¹⁶¹ brushes, but in these cases debonding was achieved by the addition of salt, a method that works equally well for weak polyelectrolyte brushes.¹⁶²

Adhesion between the PAA brush and the PDMAc hydrogel fails with increasing pH because as the polyanion becomes charged it loses its H-bond donor. This system therefore showed significant adhesion only at low pH (~ 2).¹⁵⁷ PMAA adheres to PDMAEMA only in the neutral pH region (~ 7),¹³⁹ and its failure at the extremes of pH mean that hydrogen bonding cannot explain the adhesion and so it can be concluded that this system adheres electrostatically. The electrostatic adhesion between PMAA and PDMAEMA¹⁵⁶ (and PMAA and PDMAEMA¹⁵⁸) is a little greater than the hydrogen bonding between PAA and PDMAc,¹⁵⁷ which probably reflects water competing for hydrogen bonding sites in the latter case.

The challenges of making progress with polyelectrolyte adhesion using brushes are significant. The adhesion is likely to be strong and robust, at least where electrostatic bonding dominates but the synthesis of brushes from surfaces is not appealing to the end-user. Pre-synthesized polymers that will self-assemble to form a brush layer is one approach that could be useful,⁵⁸ but the efficient self-organization of a polyelectrolyte brush is not an easy challenge. The ability to make these adhesive layers detach is appealing because of the opportunities it opens in recycling technologies.¹⁶³ However, the equilibration time of solutions required to change the pH has been prohibitively large thus far,^{156,157} although technologies can be improved.¹⁵⁹ Replacing the hydrogel with another brush layer could improve equilibration speed, as might patterning the surface to create channels to expedite solvent flow.

Bioadhesion

Polyelectrolytes have possibilities for interacting with biological cells, and their responsive nature allows adhesion and repulsion to be tuned depending on the local environment. As a rule, polyanions such as PAA and PMAA have reasonably good biocompatibility whereas polycations are more likely to be cytotoxic. Nevertheless, both have advantages in biological applications. PMAA is hydrophobic at low pH and can under such conditions attach to a target, with an increase in pH allowing, for example, the release of a cargo.¹⁶⁴ Conversely, because bacterial surfaces are typically negatively charged, polycations can readily attach to them. PDMAEMA does have the capacity to damage organisms,¹⁶⁵ which may help in biofouling applications, but an important use may well be for the delivery of molecules to a target. PDMAEMA-grafted nanoparticles have been shown to deliver a cargo of indocyanine



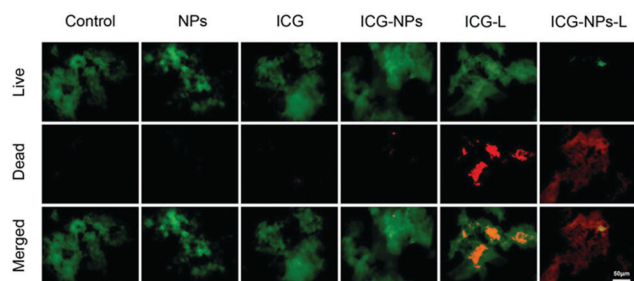


Fig. 6 Live (green)–dead (red) staining of *P. gingivalis*. When PDMAEMA-grafted nanoparticles (NPs) are present there is no significant change from the control experiment. This is also the case in the presence of ICG or ICG-loaded nanoparticles. Cells are killed if ICG is present and laser (L) irradiated. However, the effectiveness of this is at its greatest when the ICG is loaded in the nanoparticles, which attach to the bacteria. The figure is taken from the original article,¹⁶⁶ and is reproduced under a CC BY 4.0 licence.

green (ICG, a molecule used for phototherapy applications) to kill a pathogen, *Porphyromonas gingivalis* (Fig. 6).¹⁶⁶

Although charge is a useful means of interacting with organisms, pH can be used with weak polyelectrolytes to turn on and off adhesion. This phenomenon has been used with *Staphylococcus epidermidis* deposited on uncharged PAA brushes.¹⁶⁷ When the pH is increased, the charged PAA rejects the bacteria from the surface. The challenge in these experiments is to ensure that sufficient conformational change is possible in environments with high salt concentrations. Currently the use of temperature-responsive polymer brushes (not without their own problems) seems more appropriate for such purposes.¹⁶⁸

Bacteria are straightforward organisms for adhesion to polyelectrolyte brushes because they can be treated as negatively charged colloids. Eukaryotic cells are much more complex and the range of interactions that dictate their behaviour in contact with cationic and anionic weak polyelectrolyte brushes has been shown to be difficult to unravel.¹⁶⁹ Nevertheless here PAA brushes were shown to be adherent to keratinocyte cells which shows that weak polyelectrolytes may have some utility in their interactions with mammalian cells.

Bottlebrushes

The bottlebrush polymer geometry is a new area in which significant interest is being shown. These are essentially molecular brushes, because chains are grafted from a backbone rather like the comb geometry, although significantly more densely.^{170,171} They have structural similarities to articular biopolymers such as lubricin¹⁷² or aggrecan,^{173,174} which have impressive lubrication and load-bearing properties. Bottlebrushes are not easy to graft from a surface and experiments are currently dominated by polymers grafted to the surface. Work on charged polymers is largely focussed on polyzwitterions, because of their highly lubricious nature.^{175,176} Surface-grafted weak polyelectrolyte bottlebrushes are less studied but are likely to have interesting properties that are expected to be developed in the coming years.

3 Conclusions and outlook

Over the past decade there has been a significant amount of work developing polyelectrolyte brushes in areas of adhesion and lubrication. It is likely that much of this will continue, but the lack of new developments in actuation has been noticeable, despite early enthusiasm at the turn of the millennium. It is likely that future focus will lie in the interaction of polyelectrolyte brushes with biological materials and the further development of new geometries, such as bottlebrushes.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

I am grateful to the reviewers for numerous useful comments and suggestions in the preparation of the manuscript.

Notes and references

- 1 S. T. Milner, *Science*, 1991, **251**, 905–914.
- 2 Y. Higaki, M. Kobayashi, D. Murakami and A. Takahara, *Polym. J.*, 2018, **48**, 325–331.
- 3 W. Yang and F. Zhou, *Biosurf. Biotribol.*, 2017, **3**, 97–114.
- 4 L. Liu, W. Yao, Y. Rao, X. Lu and J. Gao, *Drug Delivery*, 2017, **24**, 569–581.
- 5 O. V. Borisov, E. B. Zhulina and T. M. Birshtein, *Macromolecules*, 1994, **27**, 4795–4803.
- 6 R. A. L. Jones and R. W. Richards, *Polymers at Surfaces and Interfaces*, Cambridge, Cambridge, UK, 1999.
- 7 M. Biesalski, D. Johannsmann and J. R  he, *J. Chem. Phys.*, 2002, **117**, 4988–4994.
- 8 K. Matyjaszewski, P. J. Miller, N. Shukla, B. Immaraporn, A. Gelman, B. B. Luokala, T. M. Siclovan, G. Kickelbick, T. Vallant, H. Hoffmann and T. Pakula, *Macromolecules*, 1999, **32**, 8716–8724.
- 9 R. R. Bhat, M. R. Tomlinson and J. Genzer, *Macromol. Rapid Commun.*, 2004, **25**, 270–274.
- 10 P. D. Topham, J. R. Howse, C. J. Crook, A. J. Parnell, M. Geoghegan, R. A. L. Jones and A. J. Ryan, *Polym. Int.*, 2006, **55**, 808–815.
- 11 J. Qiu and K. Matyjaszewski, *Acta Polym.*, 1997, **48**, 169–180.
- 12 A. J. Ryan, C. J. Crook, J. R. Howse, P. Topham, R. A. L. Jones, M. Geoghegan, A. J. Parnell, L. Ruiz-P  rez, S. J. Martin, A. Cadby, A. Menelle, J. R. P. Webster, A. J. Gleeson and W. Bras, *Faraday Discuss.*, 2005, **128**, 55–74.
- 13 P. Chapman, R. E. Ducker, C. R. Hurley, J. K. Hobbs and G. J. Leggett, *Langmuir*, 2015, **31**, 5935–5944.
- 14 A. Y. Sankhe, S. M. Husson and S. M. Kilbey II, *J. Polym. Sci., Part A: Polym. Chem.*, 2007, **45**, 566–575.
- 15 H. Dong and K. Matyjaszewski, *Macromolecules*, 2008, **41**, 6868–6870.



- 16 D. Konkolewicz, A. J. D. Magenau, S. E. Averick, A. Simakova, H. He and K. Matyjaszewski, *Macromolecules*, 2012, **45**, 4461–4468.
- 17 E. H. Discekici, C. W. Pester, N. J. Treat, J. Lawrence, K. M. Mattson, B. Narupai, E. P. Toumayan, Y. Luo, A. J. McGrath, P. G. Clark, J. Read de Alaniz and C. J. Hawker, *ACS Macro Lett.*, 2016, **5**, 258–262.
- 18 A. D. Jenkins, R. G. Jones and G. Moad, *Pure Appl. Chem.*, 2010, **82**, 483–491.
- 19 C. J. Hawker, A. W. Bosman and E. Harth, *Chem. Rev.*, 2001, **101**, 3661–3688.
- 20 M. Husseman, E. E. Malmström, M. McNamara, M. Mate, D. Mecerreyes, D. G. Benoit, J. L. Hedrick, P. Mansky, E. Huang, T. P. Russell and C. J. Hawker, *Macromolecules*, 1999, **32**, 1424–1431.
- 21 O. Garcia-Valdez, T. Brescacin, J. Arredondo, J. Bouchard, P. G. Jessop, P. Champagne and M. F. Cunningham, *Polym. Chem.*, 2017, **8**, 4124–4131.
- 22 J. Glasing, J. Bouchard, P. G. Jessop, P. Champagne and M. F. Cunningham, *Polym. Chem.*, 2017, **8**, 6000–6012.
- 23 L. Sonnenberg, J. Parvole, O. Borisov, L. Billon, H. E. Gaub and M. Seitz, *Macromolecules*, 2006, **39**, 281–288.
- 24 G. Moad and E. Rizzardo, in *Nitroxide Mediated Polymerization: From Fundamentals to Applications in Materials Science*, ed. D. Gimes, Royal Society of Chemistry, Cambridge, 2016, vol. 19, pp. 1–44.
- 25 S. Perrier, *Macromolecules*, 2017, **50**, 7433–7447.
- 26 F. Chen, Y. Zhu, W. Li, J. Yang, P. Fan, Z. Fei, M. Zhong, L. Chang and T. Kuang, *Appl. Surf. Sci.*, 2019, **464**, 273–279.
- 27 B. Wang, Z. Ye, Y. Tang, Y. Han, Q. Lin, H. Liu, H. Chen and K. Nan, *Int. J. Nanomed.*, 2017, **12**, 111–125.
- 28 M. K. Cho, H. J. Seo, J. H. Lee, W. K. Cho and K. Son, *Polym. Chem.*, 2021, **12**, 7023–7030.
- 29 J. H. Lee, H. J. Seo, S. Y. Lee, W. K. Cho and K. Son, *Macromol. Chem. Phys.*, 2021, **222**, 2100270.
- 30 D. Li, L. Xu, J. Wang and J. E. Gautrot, *Adv. Healthcare Mater.*, 2021, **10**, 2000953.
- 31 M. Li and C. W. Pester, *Polymers*, 2020, **12**, 1553.
- 32 A. Bratek-Sicki, P. Eloy, M. Morga and C. Dupont-Gillain, *Langmuir*, 2018, **34**, 3037–3048.
- 33 N. Houbenov, S. Minko and M. Stamm, *Macromolecules*, 2003, **36**, 5897–5901.
- 34 T. L. Fox, S. Tang, J. M. Horton, H. A. Holdaway, B. Zhao, L. Zhu and P. L. Stewart, *Langmuir*, 2015, **31**, 8680–8688.
- 35 D. Li, X. Sheng and B. Zhao, *J. Am. Chem. Soc.*, 2005, **127**, 6248–6256.
- 36 A. Johnson, J. Madsen, P. Chapman, A. Alswieleh, O. Al-Jaf, P. Bao, C. R. Hurley, M. L. Cartron, S. D. Evans, J. K. Hobbs, C. N. Hunter, S. P. Armes and G. J. Leggett, *Chem. Sci.*, 2017, **8**, 4517–4526.
- 37 C. Liu, F. Cheng, B. Liu, D. Gao, G. Cheng, C. Li, H. Wang and W. He, *Langmuir*, 2022, **38**, 1001–1010.
- 38 M. R. Tomlinson, F. Cousin and M. Geoghegan, *Polymer*, 2009, **50**, 4829–4836.
- 39 L. Xu, Z. Zhu, O. V. Borisov, E. B. Zhulina and S. A. Sukhishvili, *Phys. Rev. Lett.*, 2009, **103**, 118301.
- 40 J. Cai, C. Li, N. Kong, Y. Lu, G. Lin, X. Wang, Y. Yao, I. Manners and H. Qiu, *Science*, 2019, **366**, 1095–1098.
- 41 W. Xu, Y. Zheng and P. Pan, *J. Polym. Sci.*, 2022, DOI: 10.1002/pol.20210789.
- 42 J. L. Faubel, R. P. Patel, W. Wei, J. E. Curtis and B. K. Brettmann, *ACS Macro Lett.*, 2019, **8**, 1323–1327.
- 43 L. Michalek, L. Barner and C. Barner-Kowollik, *Adv. Mater.*, 2018, **30**, 1706321.
- 44 S. Turgman-Cohen and J. Genzer, *J. Am. Chem. Soc.*, 2011, **133**, 17567–17569.
- 45 C. Kang, R. Crockett and N. D. Spencer, *Polym. Chem.*, 2016, **7**, 302–309.
- 46 R. R. Patil, S. Turgman-Cohen, J. Šrogl, D. Kiserow and J. Genzer, *ACS Macro Lett.*, 2015, **4**, 251–254.
- 47 D. Goodman, J. N. Kizhakkedathu and D. E. Brooks, *Langmuir*, 2004, **20**, 6238–6245.
- 48 A. AL-Baradi, M. R. Tomlinson, Z. J. Zhang and M. Geoghegan, *Polymer*, 2015, **67**, 111–117.
- 49 J. D. Willott, T. J. Murdoch, G. B. Webber and E. J. Wanless, *Macromolecules*, 2016, **49**, 2327–2338.
- 50 B. Lego, M. François, W. G. Skene and S. Giasson, *Langmuir*, 2009, **25**, 5313–5321.
- 51 S. Alang Ahmad, G. J. Leggett, A. Hucknall and A. Chilkoti, *Biointerphases*, 2011, **6**, 8–15.
- 52 C. Schuh, S. Santer, O. Prucker and J. Rühe, *Adv. Mater.*, 2009, **21**, 4706–4710.
- 53 M. Steenackers, R. Jordan, A. Küller and M. Grunze, *Adv. Mater.*, 2009, **21**, 2921–2925.
- 54 Z. J. Zhang, M. Moxey, A. Alswieleh, S. P. Armes, A. L. Lewis, M. Geoghegan and G. J. Leggett, *Langmuir*, 2017, **33**, 706–713.
- 55 S. Christau, T. Möller, Z. Yenice, J. Genzer and R. von Klitzing, *Langmuir*, 2014, **30**, 13033–13041.
- 56 C. Devaux, J. P. Chapel, E. Beyou and P. Chaumont, *Eur. Phys. J. E: Soft Matter Biol. Phys.*, 2002, **7**, 345–352.
- 57 M. Ejaz, S. Yamamoto, K. Ohno, Y. Tsujii and T. Fukuda, *Macromolecules*, 1998, **31**, 5934–5936.
- 58 W. D. Seddon, L. Alfheid, A. D. F. Dunbar, M. Geoghegan and N. H. Williams, *Langmuir*, 2020, **36**, 13843–13852.
- 59 P.-G. de Gennes, *Scaling Concepts in Polymer Physics*, Cornell University Press, Ithaca, 1979.
- 60 P. J. Flory, *Principles of Polymer Chemistry*, Cornell University Press, Ithaca, 1953.
- 61 S. Alexander, *J. Phys.*, 1977, **38**, 983–987.
- 62 P. G. de Gennes, *Macromolecules*, 1980, **13**, 1069–1075.
- 63 J. D. Willott, T. J. Murdoch, G. B. Webber and E. J. Wanless, *Prog. Polym. Sci.*, 2017, **64**, 52–75.
- 64 S. W. An, P. N. Thirtle, R. K. Thomas, F. L. Baines, N. C. Billingham, S. P. Armes and J. Penfold, *Macromolecules*, 1999, **32**, 2731–2738.
- 65 J. Zhang, H. Cai, L. Tang and G. Liu, *Langmuir*, 2018, **34**, 12419–12427.
- 66 E. B. Zhulina and O. V. Borisov, *J. Chem. Phys.*, 1997, **107**, 5952–5967.
- 67 D. H. Everett, *Pure Appl. Chem.*, 1972, **31**, 577–638.
- 68 E. B. Zhulina, T. M. Birshtein and O. V. Borisov, *Macromolecules*, 1995, **28**, 1491–1499.



- 69 E. B. Zhulina and O. V. Borisov, *Langmuir*, 2011, **27**, 10615–10633.
- 70 N. A. Kumar and C. Seidel, *Macromolecules*, 2005, **38**, 9341–9350.
- 71 P. Pincus, *Macromolecules*, 1991, **24**, 2912–2919.
- 72 D. Bendejacq, V. Ponsinet and M. Joanicot, *Eur. Phys. J. E: Soft Matter Biol. Phys.*, 2004, **13**, 3–13.
- 73 E. P. K. Currie, A. B. Sieval, G. J. Fleer and M. A. Cohen Stuart, *Langmuir*, 2000, **16**, 8324–8333.
- 74 G. Ferrand-Drake del Castillo, R. L. N. Hailes and A. B. Dahlin, *J. Phys. Chem. Lett.*, 2020, **11**, 5212–5218.
- 75 A. J. Parnell, S. J. Martin, C. C. Dang, M. Geoghegan, R. A. L. Jones, C. J. Crook, J. R. Howse and A. J. Ryan, *Polymer*, 2009, **50**, 1005–1014.
- 76 T. Wu, P. Gong, I. Szleifer, P. Vlček, V. Šubr and J. Genzer, *Macromolecules*, 2007, **40**, 8756–8764.
- 77 X. Guo and M. Ballauff, *Phys. Rev. E: Stat., Nonlinear, Soft Matter Phys.*, 2001, **64**, 051406.
- 78 J. D. Willott, B. A. Humphreys, G. B. Webber, E. J. Wanless and W. M. de Vos, *Langmuir*, 2019, **35**, 2709–2718.
- 79 J. D. Willott, T. J. Murdoch, B. A. Humphreys, S. Edmondson, G. B. Webber and E. J. Wanless, *Langmuir*, 2014, **30**, 1827–1836.
- 80 H. Zhang and J. Rühle, *Macromolecules*, 2005, **38**, 4855–4860.
- 81 M. Raftari, Z. J. Zhang, S. R. Carter, G. J. Leggett and M. Geoghegan, *Tribol. Lett.*, 2018, **66**, 11.
- 82 J. D. Willott, T. J. Murdoch, B. A. Humphreys, S. Edmondson, E. J. Wanless and G. B. Webber, *Langmuir*, 2015, **31**, 3707–3717.
- 83 M. Geoghegan, L. Ruiz-Pérez, C. C. Dang, A. J. Parnell, S. J. Martin, J. R. Howse, R. A. L. Jones, R. Golestanian, P. D. Topham, C. J. Crook, A. J. Ryan, D. S. Sivia, J. R. P. Webster and A. Menelle, *Soft Matter*, 2006, **2**, 1076–1080.
- 84 F. von Goeler and M. Muthukumar, *Macromolecules*, 1995, **28**, 6608–6617.
- 85 L. Ruiz-Pérez, PhD thesis, University of Sheffield, 2006.
- 86 M. P. Weir and A. J. Parnell, *Polymers*, 2011, **3**, 2107–2132.
- 87 R. Toomey and M. Tirrell, *Annu. Rev. Phys. Chem.*, 2008, **59**, 493–517.
- 88 E. C. Johnson, I. J. Gresham, S. W. Prescott, A. Nelson, E. J. Wanless and G. B. Webber, *Polymer*, 2021, **214**, 123287.
- 89 E. C. Johnson, J. D. Willott, I. J. Gresham, T. J. Murdoch, B. A. Humphreys, S. W. Prescott, A. Nelson, W. M. de Vos, G. B. Webber and E. J. Wanless, *Langmuir*, 2020, **36**, 12460–12472.
- 90 S. Rauch, P. Uhlmann and K.-J. Eichhorn, *Anal. Bioanal. Chem.*, 2013, **405**, 9061–9069.
- 91 K. Ehtiati, S. Z. Moghaddam, A. E. Dugaard and E. Thormann, *Langmuir*, 2020, **36**, 2339–2348.
- 92 J. P. Mahalik, Y. Yang, C. Deodhar, J. F. Ankner, B. S. Lokitz, S. M. Kilbey, B. G. Sumpter and R. Kumar, *J. Polym. Sci., Part B: Polym. Phys.*, 2016, **54**, 956–964.
- 93 M. Moglianetti, J. R. P. Webster, S. Edmondson, S. P. Armes and S. Titmuss, *Langmuir*, 2010, **26**, 12684–12689.
- 94 R. J. Nap, M. Tagliazucchi and I. Szleifer, *J. Chem. Phys.*, 2014, **140**, 024910.
- 95 H. Jia, A. Wildes and S. Titmuss, *Macromolecules*, 2012, **45**, 305–312.
- 96 M. Moglianetti, J. R. P. Webster, S. Edmondson, S. P. Armes and S. Titmuss, *Langmuir*, 2011, **27**, 4489–4496.
- 97 N. Li, R. K. Thomas and A. R. Rennie, *J. Colloid Interface Sci.*, 2012, **378**, 152–158.
- 98 G. J. Dunderdale and J. P. A. Fairclough, *Langmuir*, 2013, **29**, 3628–3635.
- 99 M. P. Weir, S. Y. Heriot, S. J. Martin, A. J. Parnell, S. A. Holt, J. R. P. Webster and R. A. L. Jones, *Langmuir*, 2011, **27**, 11000–11007.
- 100 B. M. Okrugin, R. P. Richter, F. A. M. Leermakers, I. M. Neelov, E. B. Zhulina and O. V. Borisov, *Polymers*, 2020, **12**, 898.
- 101 Y.-F. Ho, T. N. Shendruk, G. W. Slater and P.-Y. Hsiao, *Langmuir*, 2013, **29**, 2359–2370.
- 102 O. V. Borisova, L. Billon, R. P. Richter, E. Reimhult and O. V. Borisov, *Langmuir*, 2015, **31**, 7684–7694.
- 103 V. Sénéchal, H. Saadaoui, J. Rodriguez-Hernandez and C. Drummond, *Langmuir*, 2017, **33**, 4996–5005.
- 104 Y. Pen, Z. J. Zhang, A. L. Morales-García, M. Mears, D. S. Tarmey, R. G. Edyvean, S. A. Banwart and M. Geoghegan, *Biochim. Biophys. Acta, Biomembr.*, 2015, **1848**, 518–526.
- 105 H. Boehm, T. A. Mundinger, C. H. J. Boehm, V. Hagel, U. Rauch, J. P. Spatz and J. E. Curtis, *Soft Matter*, 2009, **5**, 4331–4337.
- 106 S. Attili, O. V. Borisov and R. P. Richter, *Biomacromolecules*, 2012, **13**, 1466–1477.
- 107 S. Attili and R. P. Richter, *Langmuir*, 2012, **28**, 3206–3216.
- 108 J. L. Faubel, W. Wei and J. E. Curtis, *ACS Nano*, 2021, **15**, 4268–4276.
- 109 R. P. Richter, K. K. Hock, J. Burkhardtmeier, H. Boehm, P. Bingen, G. Wang, N. F. Steinmetz, D. J. Evans and J. P. Spatz, *J. Am. Chem. Soc.*, 2007, **129**, 5306–5307.
- 110 J. Seror, L. Zhu, R. Goldberg, A. J. Day and J. Klein, *Nat. Commun.*, 2015, **6**, 6497.
- 111 R. L. Cleland, J. L. Wang and D. M. Detweiler, *Macromolecules*, 1982, **15**, 386–395.
- 112 N. Srinivasan, M. Bhagawati, B. Ananthanarayanan and S. Kumar, *Nat. Commun.*, 2014, **5**, 5145.
- 113 A.-S. Yang and B. Honig, *J. Mol. Biol.*, 1993, **231**, 459–474.
- 114 U. Raviv, S. Glasson, N. Kampf, J.-F. Gohy, R. Jérôme and J. Klein, *Nature*, 2003, **425**, 163–165.
- 115 M. Chen, W. H. Briscoe, S. P. Armes, H. Cohen and J. Klein, *Eur. Polym. J.*, 2011, **47**, 511–523.
- 116 M. Chen, W. H. Briscoe, S. P. Armes and J. Klein, *Science*, 2009, **323**, 1698–1701.
- 117 J. Yu, N. E. Jackson, X. Xu, Y. Morgenstern, Y. Kaufman, M. Ruths, J. J. de Pablo and M. Tirrell, *Science*, 2018, **360**, 1434–1438.
- 118 G. Amontons, *Hist. Acad. R. Sci.*, 1699, 206–227.
- 119 B. Weber, T. Suhina, T. Junge, L. Pastewka, A. M. Brouwer and D. Bonn, *Nat. Commun.*, 2018, **9**, 888.
- 120 M. Raftari, Z. Zhang, S. R. Carter, G. J. Leggett and M. Geoghegan, *Soft Matter*, 2014, **10**, 2759–2766.



- 121 K. L. Johnson, K. Kendall and A. D. Roberts, *Proc. R. Soc. London, Ser. A*, 1971, **324**, 301–313.
- 122 B. V. Derjaguin, V. M. Muller and Y. P. Toporov, *J. Colloid Interface Sci.*, 1975, **53**, 314–326.
- 123 H. Hertz, *J. Reine Angew. Math.*, 1881, **92**, 156–171.
- 124 R. W. Carpick, D. F. Ogletree and M. Salmeron, *J. Colloid Interface Sci.*, 1999, **211**, 395–400.
- 125 R. W. Carpick and M. Salmeron, *Chem. Rev.*, 1997, **97**, 1163–1194.
- 126 G. J. Leggett, N. J. Brewer and K. S. L. Chong, *Phys. Chem. Chem. Phys.*, 2005, **7**, 1107–1120.
- 127 E. Liams, S. D. Connell, S. N. Ramakrishna and A. Sarkar, *Nanoscale*, 2020, **12**, 2292–2308.
- 128 N. Nordgren and M. W. Rutland, *Nano Lett.*, 2009, **9**, 2984–2990.
- 129 J. F. Archard, *Proc. R. Soc. London, Ser. A*, 1957, **243**, 190–205.
- 130 M. Ciavarella, J. Joe, A. Papangelo and J. R. Barber, *J. R. Soc., Interface*, 2019, **16**, 20180738.
- 131 F. J. O'Brien, *Mater. Today*, 2011, **14**, 88–95.
- 132 L. A. Osório, E. Silva and R. E. Mackay, *Bioengineering*, 2021, **8**, 113.
- 133 J. Domke and M. Radmacher, *Langmuir*, 1998, **14**, 3320–3325.
- 134 M. Etienne and A. Walcarius, *Talanta*, 2003, **59**, 1173–1188.
- 135 H. Cabibil, H. Celio, J. Lozano, J. M. White and R. Winter, *Langmuir*, 2001, 2160–2166.
- 136 P. C. Nalam, H.-S. Lee, N. Bhatt, R. W. Carpick, D. M. Eckmann and R. J. Composto, *ACS Appl. Mater. Interfaces*, 2017, **9**, 12936–12948.
- 137 G. Liu and G. Zhang, *J. Phys. Chem. B*, 2008, **112**, 10137–10141.
- 138 K. Kurihara, *Adv. Colloid Interface Sci.*, 2010, **158**, 130–138.
- 139 M. Raftari, Z. J. Zhang, S. R. Carter, G. J. Leggett and M. Geoghegan, *Macromolecules*, 2015, **48**, 6272–6279.
- 140 S. A. Prokhorova, A. Kopyshev, A. Ramakrishnan, H. Zhang and J. Rühle, *Nanotechnology*, 2003, **14**, 1098–1108.
- 141 G. Ferrand-Drake del Castillo, G. Emilsson and A. Dahlin, *J. Phys. Chem. C*, 2018, **122**, 27516–27527.
- 142 A. Isakova and K. Novakovic, *Eur. Polym. J.*, 2017, **95**, 430–439.
- 143 O. Azzaroni, B. Trappmann, P. van Rijn, F. Zhou, B. Kong and W. T. S. Huck, *Angew. Chem., Int. Ed.*, 2006, **45**, 7440–7443.
- 144 J. Bünsow, J. Erath, P. M. Biesheuvel, A. Fery and W. T. S. Huck, *Angew. Chem., Int. Ed.*, 2011, **50**, 9629–9632.
- 145 J. Bünsow, T. S. Kelby and W. T. S. Huck, *Acc. Chem. Res.*, 2010, **43**, 466–474.
- 146 J. Anthi, V. Kolivoška, B. Holubová and H. Vaisocherová-Lísalová, *Biomater. Sci.*, 2021, **9**, 7379–7391.
- 147 G. Panzarasa, M. Dübner, V. Pifferi, G. Soliveri and C. Padeste, *J. Mater. Chem. C*, 2016, **4**, 6287–6294.
- 148 S. Zou, M. A. Hempenius, H. Schönherr and G. J. Vancso, *Macromol. Rapid Commun.*, 2005, **27**, 103–108.
- 149 L. Hu, Q. Zhang, X. Li and M. J. Serpe, *Mater. Horiz.*, 2020, **6**, 1774–1793.
- 150 G. Kocak, C. Tuncer and V. Bütün, *Polym. Chem.*, 2017, **8**, 144–176.
- 151 R. A. Pethrick, *Proc. Inst. Mech. Eng., Part L*, 2015, **229**, 349–379.
- 152 C. Creton, *MRS Bull.*, 2003, **28**, 434–439.
- 153 S. Sun, M. Li and A. Liu, *Int. J. Adhes. Adhes.*, 2013, **41**, 98–106.
- 154 C. Loskofsky, F. Song and B. Zhang Newby, *J. Adhes.*, 2006, **82**, 713–730.
- 155 M. Kobayashi, M. Terada and A. Takahara, *Soft Matter*, 2011, **7**, 5717–5722.
- 156 R. La Spina, M. R. Tomlinson, L. Ruiz-Pérez, A. Chiche, S. Langridge and M. Geoghegan, *Angew. Chem., Int. Ed.*, 2007, **46**, 6460–6463.
- 157 G. Sudre, L. Olanier, Y. Tran, D. Hourdet and C. Creton, *Soft Matter*, 2012, **8**, 8184–8193.
- 158 L. Alfheid, R. La Spina, M. R. Tomlinson, A. R. Hall, W. D. Seddon, N. H. Williams, F. Cousin, S. Gorb and M. Geoghegan, *J. Adhes.*, 2018, **94**, 58–76.
- 159 L. Alfheid, W. D. Seddon, N. H. Williams and M. Geoghegan, *Soft Matter*, 2016, **12**, 5022–5028.
- 160 Z. Zhang, M. R. Tomlinson, R. Golestanian and M. Geoghegan, *Nanotechnology*, 2008, **19**, 035505.
- 161 M. Kobayashi and A. Takahara, *Polym. Chem.*, 2013, **4**, 4987–4992.
- 162 L. Alfheid, N. H. Williams and M. Geoghegan, *J. Appl. Polym. Sci.*, 2020, **137**, 49130.
- 163 J. M. Garcia and M. L. Robertson, *Science*, 2017, **358**, 870–872.
- 164 E. Koh and Y. Taek Lee, *Appl. Surf. Sci.*, 2021, **562**, 150133.
- 165 W. Yandi, S. Mieszkina, A. di Fino, P. Martin-Tanchereau, M. E. Callow, J. A. Callow, L. Tyson, A. S. Clare and T. Ederth, *Biofouling*, 2016, **32**, 609–625.
- 166 E. Shi, L. Bai, L. Mao, H. Wang, X. Yang, Y. Wang, M. Zhang, C. Li and Y. Wang, *J. Nanobiotechnol.*, 2021, **19**, 413.
- 167 V. Yadav, Y. A. Jaimes-Lizcano, N. K. Dewangan, N. Park, T.-H. Li, M. L. Robertson and J. C. Conrad, *ACS Appl. Mater. Interfaces*, 2017, **9**, 44900–44910.
- 168 J. Collett, A. Crawford, P. V. Hatton, M. Geoghegan and S. Rimmer, *J. R. Soc., Interface*, 2007, **4**, 117–126.
- 169 E. J. Cozens, D. Kong, N. Roohpour and J. E. Gautrot, *Soft Matter*, 2020, **16**, 505–522.
- 170 R. Verduzco, X. Li, S. L. Pesek and G. E. Stein, *Chem. Soc. Rev.*, 2015, **44**, 2405–2420.
- 171 Y. Xu, F. Plamper, M. Ballauff and A. H. E. Müller, *Adv. Polym. Sci.*, 2010, **228**, 1–38.
- 172 G. D. Jay and K. A. Waller, *Matrix Biol.*, 2014, **39**, 17–24.
- 173 C. Kiani, L. Chen, Y. J. Wu, A. J. Yee and B. B. Yang, *Cell Res.*, 2002, **12**, 19–32.
- 174 L. Ng, A. J. Grodzinsky, P. Patwari, J. Sandy, A. Plaas and C. Ortiz, *J. Struct. Biol.*, 2003, **143**, 242–257.
- 175 X. Banquy, J. Burdyńska, D. W. Lee, K. Matyjaszewski and J. Israelachvili, *J. Am. Chem. Soc.*, 2014, **136**, 6199–8202.
- 176 J. Faivre, B. R. Shrestha, G. Xie, M. Olszewski, V. Adibnia, F. Moldovan, A. Montembault, G. Sudre, T. Delair, L. David, K. Matyjaszewski and X. Banquy, *Chem. Mater.*, 2018, **30**, 4140–4149.

