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Opportunities and limitations of membrane-based preconcentration for rapid and continuous diagnostic applications†

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Modern rapid diagnostic tests have continued to improve in performance and convenience, including the proliferation of point-of-care and the emergence of continuous wearable diagnostics. Frequently, rapid diagnostic tests are challenged in their sensitivity and their limit of detection, especially for disease detection in its early stages where analyte concentration can be lower, leading to false negative test results. In response, preconcentration methods have been explored to increase analyte concentration in the collected sample. In theory, a particularly attractive approach is simple membrane preconcentration, whereby the membrane retains the analyte through size-selective filtration while removing water from the sample. This article reviews two leading methods for membrane-based preconcentration, high-pressure pneumatic and osmotic preconcentration, which have the greatest potential because they can operate with the speed (minutes) required for rapid diagnostic tests. Using these simple yet effective methods, analyte concentration can be rapidly increased by a factor of 100× or more, both in buffer and in biofluids. Data from previous studies and new data found only in this article now allows preliminary conclusions on the benefits and limitations of each method. Preconcentration amounts of 10× were achieved for bovine serum albumin and glucose using osmotic pressure, while preconcentration amounts of >160× were achieved for pregnancy hormone and >70× for flu using pneumatic pressure. While each method is more limited in utility than originally speculated, each method will be shown to have merit for niche applications of both point-of-care and continuous diagnostics, with particular value found in early disease detection.

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Introduction

Rapid diagnostics are now ubiquitous; there are thousands of tests used to diagnose everything from the simpler ailments of daily life, such as urinary tract infections¹ and strep throat,² to complex disorders, such as diabetes³ and cancer.⁴ However, there are caveats to nearly every diagnostic test, including low accuracy in many cases.^{5,6} Low accuracy can either be the result of a lack of specificity (false positive) or lack of sensitivity (false negative).⁷ A test's sensitivity is a measure of how often a test provides a 'true positive' result in comparison to a gold standard method of testing. For example, the sensitivity of rapid flu tests is often determined by comparing the number of positive tests to the number of actual flu cases as determined using a much slower approach of viral culture or more complex approach of polymerase chain reaction (PCR).⁸ The sensitivity of many rapid

diagnostic tests, especially rapid tests such as point-of-care (POC) tests^{9–11} is often fairly poor due to low analyte load in the body, such as during the early stages of a viral infection. Therefore, researchers have been working for decades to determine methods which increase diagnostic test sensitivity.¹² An alternative, arguably simpler, method of effectively increasing the test sensitivity is to increase the amount of analyte in the sample. Even though the detection limit of the assay remains the same, the amount of analyte in the sample could be brought above the limit of detection without modification of the test itself.

Sample preconcentration has been pursued by many research groups,^{13–26} but currently there is no universal method being used in the market without drawbacks in time consumption, cost, or the required use of bulky lab equipment. For example, a paper-based method has been demonstrated to preconcentrate influenza ribonucleic acid to increase the accuracy of Influenza A detection.²⁷ While the device was low-cost and increased the concentration of the viral load by 10×, the device required 45 minutes to operate, making it impractical for many instances of clinical use where a rapid answer is required. Another example demonstration sought to preconcentrate the bacteria in an

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aqueous sample for PCR using magnetic ionic liquids and an external magnetic field.²⁸ The process demonstrated impressive preconcentration quantities of nearly 50× in under 10 minutes; however, this method required the use of several separate steps and peripheral equipment, rendering it impractical for many use cases of rapid diagnostics.

In response, our research group has been investigating several methods of sample preconcentration in recent years which use a membrane filter to concentrate analyte as the bulk fluid in the sample is removed. The basics of the process are quite simple. An initial volume of fluid is decreased by moving most of the fluid through a membrane filter, which has pores small enough to retain the biomolecule of interest in the smaller volume of fluid, thereby increasing the potency of the fluid sample. This process can be used in devices which are quite small, or extremely large. For example, microchips with embedded membrane filtration modules have been used to concentrate many biomolecules, such as proteins, viruses, and bacteria;^{13,14,21,29} on the other hand, water treatment facilities often use huge sheets of membrane to filter salt water for drinking purposes.^{30,31} The methods used to drive fluid through the membrane can vary quite a bit in functionality and complexity. For example, some methods include osmotic pressure,²⁹ electrokinetics,^{20,21} and pneumatic pressure.³²

In our work, methods of fluid removal included osmosis,²⁹ vacuum,³² wicking, and pneumatic pressure.³² Our goal was to find a method which is rapid, reliable, and cost effective to create a device which could easily be integrated into rapid diagnostic tests or continuous diagnostics which require rapid preconcentration to enable 'real time' data. Based on our cumulative findings, in this article we will discuss the major challenges facing membrane-based concentration and important elements that must be considered for such a device to be successful. We will focus on the two preconcentration methods that we found to be most successful in detail, high-pressure pneumatic and osmotic driven preconcentration. We will further provide speculation on the types of analytes/diagnostic tests for which these membrane-based preconcentration approaches have the most potential for improving clinical outcomes in rapid diagnostics.

Considerations/issues with membrane-based concentration

We will first review several aspects of membrane-based preconcentration that must be considered regardless of application. Membrane-based preconcentration, in practice, requires very high pressures (10 000's to 100 000's of Pascals). The need for high pressures to generate high flow rates for removal of water from the sample then creates additional challenges that compromise the ability to maintain that high pressure, leading to loss of analyte at or near the membrane surface.

Practical devices require very large pressures

The first and foremost item to consider with any membrane preconcentration module for diagnostics is flux, or the amount of fluid which moves through a unit area of the membrane over a unit time (for example: mL cm⁻² min⁻¹).³³ A preconcentration module is impractical and unlikely to be used if it takes too long to complete its run-time for use with a rapid diagnostic test. Generally, if the preconcentration takes too long (10's of minutes to hours) then in many applications it is simpler to just use a conventional benchtop diagnostic technique with higher sensitivity and lower limit of detection (*e.g.*, enzyme-linked immunoassay, or ELISA). Therefore, preconcentration methods which can be completed in several minutes are the most desirable in real applications.

Overall, there were four membrane-based preconcentration methods which were considered over the course of our research group's work (Table 1):¹ wicking of fluid through the membrane using a fibrous material such as paper or a cotton ball,² pulling fluid through the membrane using a vacuum,³ moving fluid through the membrane using osmotic pressure, and⁴ driving fluid through the membrane using pneumatic pressure. Table 1 demonstrates the main characteristics of each of these four methods of membrane-based preconcentration.

In our own early work, we initially explored what we expected to be the simplest and yet still effective approaches: wicking pressure and vacuum pressure. Although wicking using materials with small capillary features can result in extremely high pressure gradients (Young-Laplace pressure) and high initial water fluxes, having small capillary features also creates a significant hydraulic resistance to fluid flow. Simply, hydraulic resistance quickly starts to dominate and diminish any advantages brought by the high capillary wicking pressure. Furthermore, many wicking materials such as paper have a rough surface and will have limited contact area with the membrane, further reducing the water flux regardless of wicking pressure. Vacuum pressure is more complex than wicking pressure (requires a pump or vacuum chamber), but it does not suffer from the resistance to fluid flow that exists for wicking pressure. However, vacuum pressure is limited to atmospheric pressure (101.3 kPa) which when coupled with the hydraulic resistance of membranes results in preconcentration times that require hours to days.

In our pursuit of membrane preconcentration for rapid and continuous diagnostics, we eventually concluded that much higher pressures, up to hundreds of thousands of Pascals, would be required. This led to a focus on the only two methods we believe to be feasible to maintain such high pressures during preconcentration: pneumatic pressure and osmotic pressure. Each of these methods is now presented with a brief description of operation and challenges for each. For rapid diagnostics (Fig. 1a), pneumatic preconcentration



Table 1 High-level comparison of various methods of membrane-based preconcentration

	Pneumatic pressure	Osmotic pressure	Wicking pressure	Vacuum pressure
Pressure gradient / flux	>5.7 MPa	>23 MPa	Challenging	101.3 kPa
Time to complete preconcentration	Minutes	Minutes	Hours	Hours – days
Complexity	Low	Low	Low	Medium

with looser membrane molecular weight cut-offs (MWCO's) (kDa's or greater) was found to be preferred because it is the only combination of pressure and membrane which can quickly concentrate large (mL's) initial sample volumes into a pre-concentrated sample of 10's to 100's of μL remaining for the diagnostic assay. For continuous diagnostics, biosensors are required and existing continuous biosensors (enzymatic^{34–37} and aptamer^{38–40}) are limited mainly to small molecule analytes. If the analyte is a small molecule, the membrane must be much tighter (small MWCO) which increases the hydraulic resistance of the membrane, which in turn requires even higher pressures that are most easily generated by osmotic pressure.

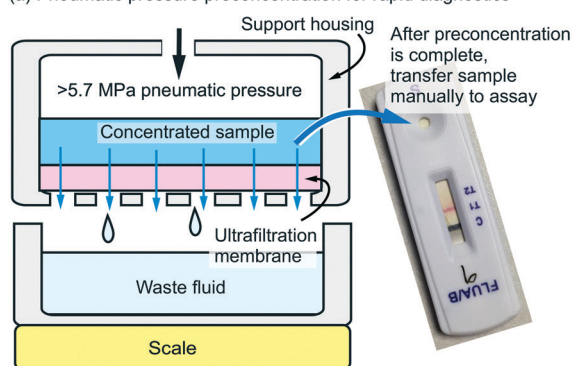
Basics of membrane preconcentration with high pneumatic and osmotic pressures

Pneumatic pressure for rapid diagnostics. Fig. 1a is a schematic of pneumatic pressure preconcentration for rapid diagnostic tests. A fluid sample is placed into the top chamber or sample cup of the device. A pneumatic pressure is applied to the sample using compressed gas, and the bulk of the fluid is forced through the ultrafiltration membrane into a waste collection container. As the bulk fluid is removed from the sample solution, the analyte remains in the top chamber of the device and becomes concentrated. Retaining the analyte in the sample chamber requires that the membrane have a MWCO that is less than the molecular weight of the analyte. A precise amount of preconcentration

can be achieved for each test using feedback from a scale weighing the amount of waste fluid, which then turns off the device once enough waste fluid is collected. After the process is complete, the pre-concentrated sample can be manually transferred to the desired assay using a syringe or micropipette.

Osmotic pressure for continuous diagnostics. Fig. 1b is a schematic describing the process of osmotic pressure preconcentration. Sample fluid such as environmental water from a river or sweat from a body is continuously introduced through a microfluidic channel beneath a forward osmosis membrane. A reservoir of draw solution, which is 10–100 \times the osmolality of the sample fluid, sits above the membrane. Due to the osmotic pressure difference, water from the sample solution begins to flow through the membrane into the draw solution reservoir to restore osmotic equilibrium.⁴¹ Therefore, as the sample fluid flows through the channel, the analyte becomes more concentrated. The concentrated sample solution eventually flows across the sensor, and a continuous reading of the analyte concentration can be obtained. If the osmolarities of both the incoming sample and the draw solution are known, this approach can be rendered more predictable in the amount of preconcentration achieved by allowing the sample stream and the osmotic draw solution to equilibrate in osmolality before reaching the sensor. However, allowing for equilibrium in osmotic strengths limits the input flow rate²⁹ and can impact the sensor performance (by changing pH or salinity).

(a) Pneumatic pressure preconcentration for rapid diagnostics



(b) Osmotic preconcentration for continuous diagnostics

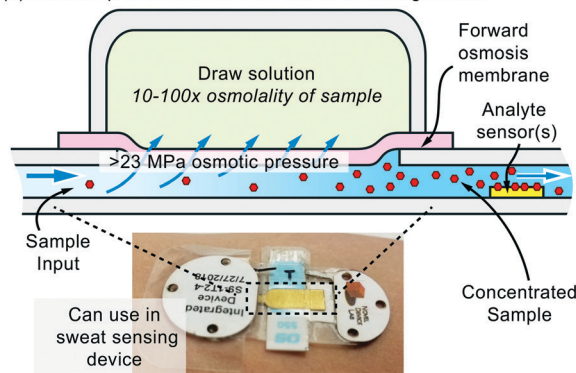


Fig. 1 Schematics of two viable membrane-based preconcentration methods: a) pneumatic-driven preconcentration for rapid diagnostics – compressed gas is used to pressurize the device and force the bulk of the sample fluid through an ultrafiltration membrane, concentrating the analyte in the sample. The sample is then transferred to a rapid diagnostic test device; b) osmotic preconcentration – an osmotic draw solution with a higher osmolality (salt concentration) than the sample solution is used to draw fluid through a forward osmosis membrane, concentrating the analyte as it flows through the channel. An example application for osmotic preconcentration is continuous sweat biosensing where most analytes of interest are highly diluted in the sweat.²⁷



Challenges with caking (reversible fouling) and pore blocking (irreversible fouling)

Fouling, or deposition of molecules into the pores or onto the surface of the membrane, is one of the biggest barriers to success in any membrane filtering process.⁴² The two primary types of fouling are pore blocking, also referred to as 'irreversible fouling', and caking, also referred to as 'reversible fouling'. Pore blocking, seen in Fig. 2a, occurs when analyte particles are lodged into the pores of the membrane such that it partially or completely blocks fluid flow through that pore. When this occurs, it is extremely difficult to remove the particles. If a large amount of pore blocking occurs, fluid will struggle to move through the membrane and additional particles will begin to build up on the membrane surface in a process called caking. Caking, seen in Fig. 2b, can also occur independently of pore blocking and can be reversed using methods such as membrane cleaning or swiping.^{43–46} The positive effects of membrane swiping are demonstrated later in this article (see 'Example success for pneumatic preconcentration'). Some caking can even be prevented by pre-filtering the sample fluid with a larger-pore membrane to remove the largest unwanted proteins and particles. However, since some amount of fouling from concentration polarization, pore blocking, or caking is inevitable, models have been created by various groups to better quantify the flux decrease which will occur due to fouling in particular environments and conditions.^{47–49}

Over time, all membrane filtration processes will result in fouling due to pore blocking and/or caking, and care must be taken to decrease these effects as much as possible. There are some simple, feasible fouling improvement strategies that can be used. For example, if attempting to concentrate a larger analyte, a membrane with larger pores can be chosen such that smaller solutes, such as ions or small molecules, can easily pass through the pores of the membrane, reducing pore blocking. Additionally, if attempting to concentrate smaller analytes, one can mix the fluid during

preconcentration using mechanical means such as vibration to prevent analyte from accumulating at the membrane surface and causing caking. Caking also has a strong dependence on the size or molecular weight of the particles which will be present in the sample solution. Small particles are more likely to get trapped in membrane pores and cause irreversible pore blocking. Larger particles are less likely to block membrane pores but have a higher affinity to cake onto the membrane and block fluid flow.⁵⁰ Therefore, the size of particles in the sample fluid plays a substantial role in the amount of analyte which is near or caked onto the membrane surface. As seen in Fig. 2c, bulkier particles have a larger cross-sectional area and therefore the applied pressure results in a larger applied force in the direction of fluid flow. In addition, larger particles have smaller diffusion coefficients, and therefore less diffusion away from the membrane surface, resulting in a concentration build up at the membrane surface. Smaller particles, on the other hand, have less cross-sectional area, less applied force towards the membrane, and larger diffusion coefficients. This results in more diffusion away from the membrane, making them much less susceptible to caking. As a rough estimate, assuming a spherical shape for a solute, the applied force due to the pressure applied is simply proportional to the square of the radius of the particle. Diffusion is directly associated with molecular weight, as shown in the Stokes-Einstein relation:⁵¹

$$D = \frac{kT}{6\pi\eta R_H} \quad (1)$$

where D is the diffusion coefficient ($\text{m}^2 \text{s}^{-1}$), k is the Boltzmann constant ($\text{kg m}^2 \text{s}^{-2} \text{K}^{-1}$), T is the temperature (K), η is the fluid viscosity ($\text{kg m}^{-1} \text{s}^{-1}$), and R_H is the hydrodynamic radius of the analyte (m). However, both of the above predictions for applied force and diffusion are just estimates which in practice must be measured for a given solute.^{52,53}

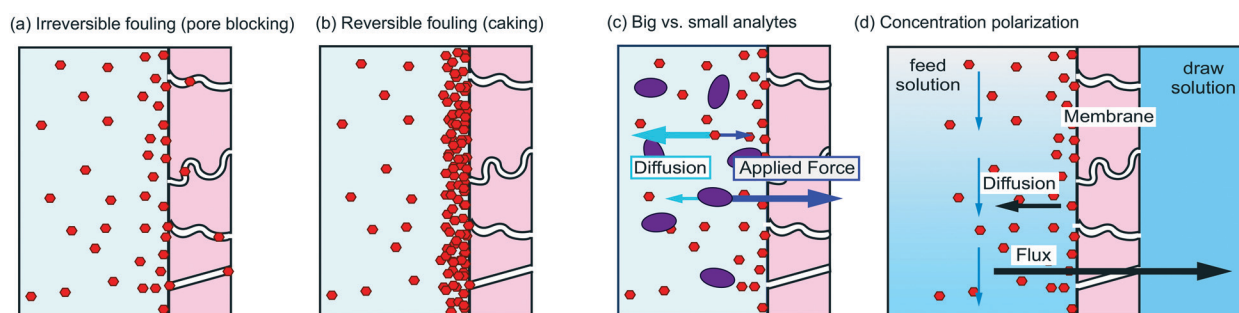


Fig. 2 Illustrations of prominent issues facing membrane-based concentration processes: a) pore blocking occurs when analyte becomes lodged onto the surface of pores or into membrane pores, decreasing flux over time; b) caking occurs to some extent in all types of membrane-based filtration processes and is increased after pore blocking has occurred, but can be reversed by membrane cleaning; c) analyte size must be considered in choosing which membrane-based preconcentration process to use, as this affects the amount of analyte which will be present near the membrane surface, ultimately affecting fouling and flux; d) concentration polarization, common in osmotic preconcentration, causes analyte to build up at the membrane surface, decreasing the net flux which occurs.



Challenges for continuous osmotic preconcentration: concentration polarization

In addition to pore blocking and caking, osmotic preconcentration can be further limited by reduced water flux through the membrane due to concentration polarization (Fig. 2d).^{31,41,50} Generally, the flux through a membrane can be estimated using the following equation:

$$J_w = A(\Delta\pi - \Delta P) \quad (2)$$

where J_w is flux across the membrane (in $\text{L m}^{-2} \text{s}^{-1}$), A is the hydraulic permeability of the membrane (in m s^{-1}), $\Delta\pi$ is the osmotic pressure difference across the membrane (in Pa), and ΔP is the difference in hydrostatic pressure across the membrane (in Pa). Note that in our specific case of osmotic preconcentration, ΔP is zero, and can be eliminated from this equation.

Without considering concentration polarization, the osmotic pressure difference across the membrane can be estimated by:

$$\pi_{\text{Total}} = \pi_{\text{Draw}} - \pi_{\text{Feed}} = iRT(M_{\text{Draw}} - M_{\text{Feed}}) \quad (3)$$

where i is the Van't Hoff factor (*ex.* $i = 2$ for NaCl), R is the ideal gas constant ($8.314 \text{ m}^3 \text{ Pa K}^{-1} \text{ mol}^{-1}$), T is the temperature (K), M_{Draw} is the molar concentration of the draw solution (in mol L^{-1}) and M_{Feed} is the molar concentration of the sample solution (in mol L^{-1}). However, concentration polarization, which is always present in osmotic preconcentration, reduces the osmotic pressure as predicted by the following equation:

$$\pi_{\text{Total}} = iRT(M_{\text{Draw}}e^{-J_w K} - M_{\text{Feed}}e^{J_w K}) \quad (4)$$

where k is the mass transfer coefficient (in m s^{-1}), and K is the solute resistivity for diffusion.⁵⁴ This variable, K , is dependent on multiple characteristics of the membrane and draw solute, and can be estimated by:

$$K = \frac{t\tau}{D\varepsilon} \quad (5)$$

where t is the membrane support layer thickness (in m), τ is the support layer tortuosity (unitless), D is the diffusion coefficient of the draw solute (in $\text{m}^2 \text{s}^{-1}$), and ε is the membrane support layer porosity (unitless).

As water flows through the membrane into the draw solution reservoir, a build-up of solutes in the sample fluid accumulates near the surface of the membrane (Fig. 2d). This larger concentration of solutes is called the boundary layer. The larger concentration of solutes in the boundary layer can approach the concentration of solutes in the draw solution, which decreases the osmotic pressure difference across the membrane, significantly decreasing flux and increasing the time needed for preconcentration. The negative effects of concentration polarization cannot be completely avoided but can be diminished by mixing the fluid during concentration

or increasing the turbidity of the flow to disperse solutes that are concentrating near the membrane surface.

Concentration polarization is generally not an issue in pneumatic pressure preconcentration because, as stated previously, a membrane with a larger MWCO is utilized, allowing the smaller molecular weight solutes that dominate total osmolality to simply pass through the membrane (the osmotic pressure difference caused by larger particles in the solution can be considered negligible in comparison).

Example success for osmotic concentration

As previously stated, investigation of analyte preconcentration using a high osmolality draw solution has been investigated in our laboratory. As the results of this study (all of the work in this section) have already been published, the full details of the study have been omitted and can be found in the original publication.²⁹ However, the main testing procedure and results are summarized here in order to allow for a thoughtful discussion on the method's potential in diagnostics.

Membrane and analyte selection

Glucose (~ 180 Da) was chosen as an analyte for osmotic preconcentration testing, as it is a small sugar which is simple to measure and which has high biological relevance. A review of commonly used membrane types was conducted, and a plot was made overlaying various membranes' MWCOs with certain analytes to demonstrate membrane/analyte size compatibility (Fig. 3a).²⁹ Considering glucose's small size, the forward osmosis Rainstick (FTS H20) membrane was selected for testing in this study because the membrane blocks even the passage of common salts (NaCl, KCl, *etc.*). Bovine serum albumin (BSA) was also used as an analyte in later testing because it is a sticky protein (~ 66.5 kDa) which could be used to test osmotic preconcentration's limits with larger analytes that may be more prone to fouling the membrane.

Testing procedures

A representative osmotic preconcentration device prototype is shown in Fig. 3b. Because the forward osmosis membrane is very tight, even with very high osmotic pressures of >23 MPa, the water flux through the membrane is low. Therefore, a long 10 cm channel was experimentally required to allow for complete preconcentration of the sample fluid. The osmotic preconcentration devices were fabricated using a rapid method, the design-cut-assemble approach, in which the device is assembled using layer by layer lamination. Complete fabrication details can be found in the original article.²⁹

Glucose was preconcentrated by using $1\times$ PBS (phosphate buffered saline) as the draw solution, and $0.1\times$ PBS in the



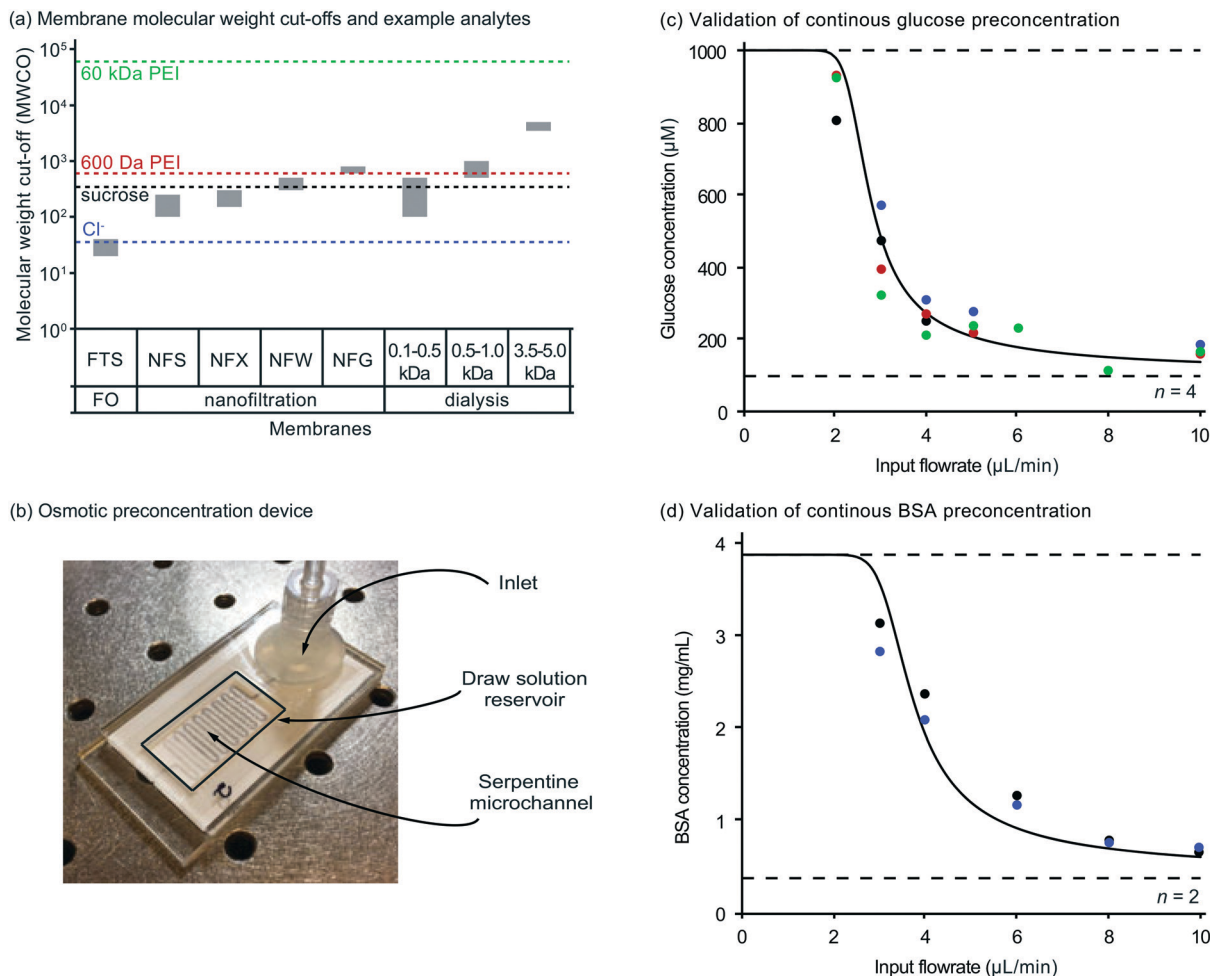


Fig. 3 Continuous osmotic pre-concentration device and results: a) various membrane types and their MWCOs are plotted with various analytes and their molecular weights to show compatible membrane/analyte combinations; b) completed device fabricated using cut-design-assemble approach with compact serpentine microchannel; c) concentration of glucose achieved using various input flowrates overlaid with theoretical model prediction (four different colors represent different test runs); d) concentration of BSA achieved using various input flowrates overlaid with theoretical model prediction (two different colors represent different test runs).²⁸

sample solution (10× osmolality difference). In this manner, the sample solution, containing a glucose concentration of 100 μM , would ideally be concentrated 10× to 1 mM. In a similar manner, a 10× PBS draw solution was used to attempt to concentrate a sample solution of BSA with a starting concentration of 0.39 mg mL^{-1} and 1× PBS to a final concentration of 3.9 mg mL^{-1} , a 10× increase. A syringe and the device channel were filled with the sample solution being tested using techniques which avoided the formation of air bubbles (details can be found in original publication²⁹) and the syringe was connected to the tubing at the inlet of the device. Then, a syringe pump was used to infuse the sample solution into the device at various flow rates. Samples of the concentrated solution were taken at the outlet of the device for later concentration quantification. In a fully integrated, commercial device, the sensors for the diagnostic assay could be placed in the channel itself for continuous monitoring.

Results

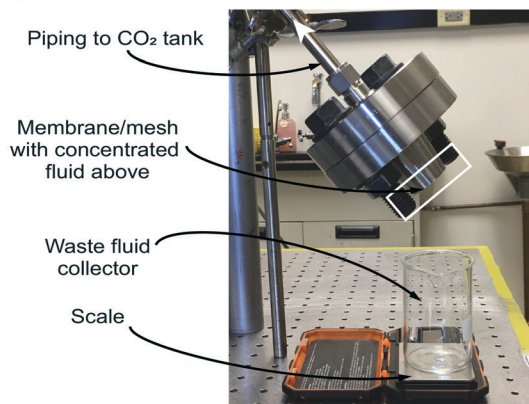
For laboratory testing, to determine the amount of glucose pre-concentration at each flow rate, electrochemical glucose test strips were used due to their extremely low volume requirement of 1 μL . BSA concentrations were tested using a UV-vis spectrophotometer (NanoDrop One), which also required only 1 μL of solution. The results of these tests for glucose and BSA can be seen in Fig. 3c and d, respectively. These results are plotted over the predicted concentration curve using a theoretical model, the details of which can be found in the original publication.²⁹ The decreasing slope of the predicted curves is due to the fact that lower flow rates are required to achieve full osmotic equilibration and therefore maximum concentration before fluid exits the channel. The pre-concentration results for glucose matched the theoretical predictions extremely closely. The BSA concentration results also followed the same overall curve as the predictive model but did not fit the theoretical curve as



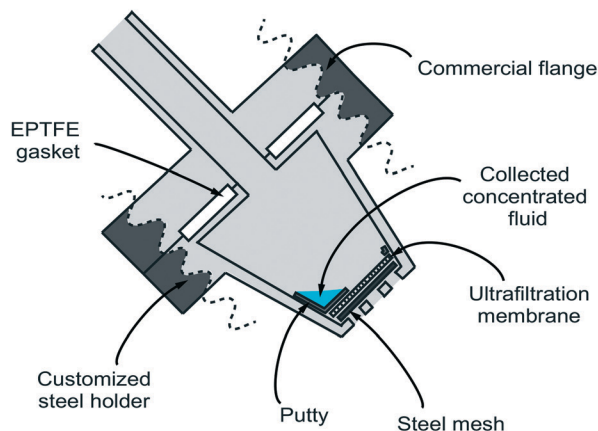
well as glucose did. This is potentially because BSA is sticky and likely adhered to the channel walls and the membrane. The lack of adherence of the BSA curve to the theoretical

curve may also be the result of increased fouling and caking onto the membrane due to the larger size of BSA, as was discussed in the previous section of this article (Fig. 2c).

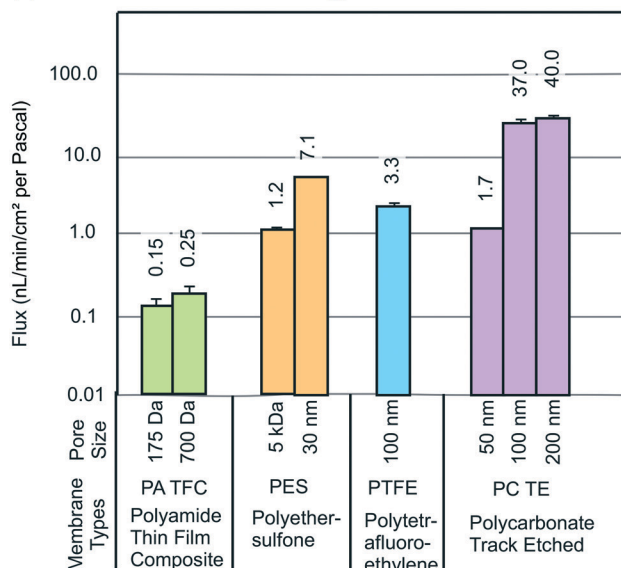
(a1) Pneumatic pressure preconcentration setup



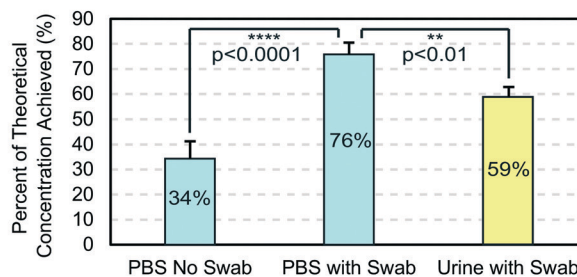
(a2) Pneumatic pressure preconcentration setup closeup inside holder (not drawn to scale)



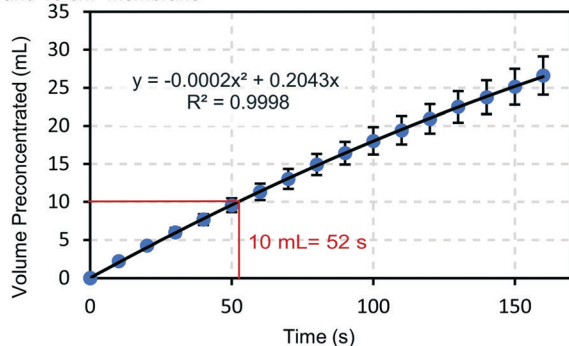
(b) Flux data for various membrane types



(c) Pneumatic pressure preconcentration results



(d) Total volume of PBS preconcentrated over time using 5.7 MPa and 10 cm² membrane



(e) Total volume of urine preconcentrated over time using 5.7 MPa and 10 cm² membrane

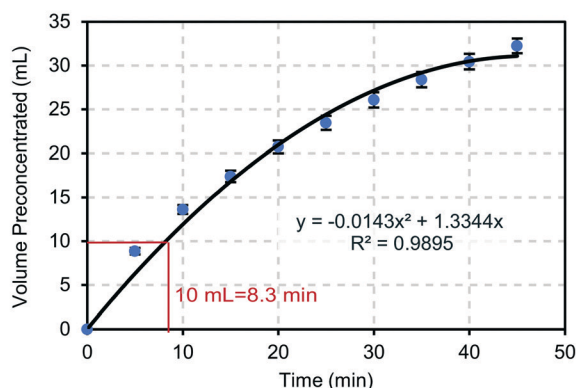


Fig. 4 Pneumatic pressure preconcentration device and results: a1) photo of the completed setup; a2) schematic of components inside steel holder; b) membrane flux characterization for membranes with various pore sizes (adapted from Drexelius 2020); c) concentration results for pneumatic pressure setup using PBS and urine; d) flux of PBS through a membrane which is 10 cm² and using 5.7 MPa (red box shows time needed to concentrate 10 mL of PBS); e) flux of urine through a membrane which is 10 cm² and using 5.7 MPa (red box shows time needed to concentrate 10 mL of urine).



Example success for pneumatic preconcentration

Next, the pneumatic preconcentration technique, which involved applying large amounts of positive pressure using compressed gas, was used. It should be noted that all of the work summarized in this section of the article is original work which has not been published.

Test setup

For testing the pneumatic preconcentration technique, a setup was created which used a carbon dioxide tank as the pressure source. Carbon dioxide was chosen for testing in this study because it is one of the most ubiquitous gases used as a pressure source on the market, and because carbon dioxide can be stored as a liquid while providing a large 5.7 MPa vapor pressure at room temperature. The carbon dioxide tank was coupled to a regulator and release valve, which was then routed to the concentration setup shown in Fig. 4a.¹⁻² The concentration setup itself consisted of a commercial flange as well as a custom designed steel holder for the preconcentration membrane. The flange and steel holder were bolted together along with an expanded polytetrafluoroethylene (EPTFE) gasket. The membrane itself was cut to an appropriate size using a laser cutter and placed into the bottom of the custom steel holder. In addition, a steel mesh (120 mesh, 304 stainless steel) was placed underneath the membrane to preserve the membrane integrity while pressure was being applied. A thin layer of putty (Dicor Butyl Tape, Part No. BT-1834-1) was used to occlude a small portion on one side of the membrane to stop the concentration process when the fluid volume dipped below a certain amount (the putty simply blocks further water flux). Without the putty, one would have to precisely time when the experiment would be stopped else all the water in the sample fluid would be removed. With the putty the device can be easily designed to have an 'auto-stop' such that a predictable final preconcentrated sample volume is achieved each time. The entire steel holder/flange piece was tilted to an angle of 45 degrees to position the device with respect to gravity such that the putty would catch the final concentrated volume of sample fluid. A beaker was placed below the steel holder to catch the waste fluid, and a scale was placed underneath the beaker in order to weigh the amount of waste fluid leaving the device. The weight measurement of the waste fluid then allowed the fluid flux to be monitored throughout the preconcentration process.

Testing procedures

For initial pneumatic pressure preconcentration testing, 1× PBS (phosphate buffered saline) was chosen as the sample fluid type to emulate the extraction buffer fluid used in many nasopharyngeal swab rapid diagnostic tests. Influenza A nucleoprotein was chosen as the target analyte, as influenza is historically one of the most prevalent and deadly viruses.

Many rapid influenza tests use a nasopharyngeal swab and extraction buffer to release the nucleoprotein, and therefore test sensitivities could be improved with preconcentration. Multiple membranes were tested in the lab to determine their flux (see original publication for membrane characterization procedure detail³²), and plotted as seen in Fig. 4b. The membrane chosen for use in this study was a 5 kDa polyethersulfone (PES) membrane, as it could easily reject all of the nucleoprotein (~56 kDa) and still maintain a high flux.

For each test, PBS was spiked with influenza A nucleoprotein to a concentration of 16 ng mL⁻¹. 10 mL of the fluid was deposited into the custom steel holder before the holder was bolted onto the commercial flange. A transparent blast shield was placed around the setup for safety with this research-only device, the carbon dioxide tank was opened, and the system became rapidly pressurized to 5.7 MPa. The weight of fluid which had passed through the membrane and fallen into the waste beaker was recorded every 10 seconds, and later converted to a flux value and normalized to the amount of pressure, in mL min⁻¹ cm⁻² per Pascal.

Recordings continued every 10 seconds in this manner until flux had completely stopped as the final sample volume settled over the putty-covered membrane. At this point, the carbon dioxide tank was shut, and the release valve opened to depressurize the setup. The final volume of the concentrated sample was measured (in the range of 50–200 μL's). Using this volume, the maximum theoretical concentration amount was calculated. (e.g. 10 mL initial volume/100 μL final volume = 100×).

After these tests were complete, because of analyte loss due to caking, an additional step was introduced at the end of the concentration process to try to improve analyte recovery. After the system was depressurized, a cotton swab was used to wipe the membrane to mechanically disturb analyte which had been caked onto the surface. The cotton swab was then placed into a microcentrifuge tube with the concentrated fluid sample and centrifuged to release the recovered analyte back into the fluid. In real-world application, a more automated method will likely be required, such as piezoelectric vibration of the membrane during preconcentration to continuously prevent caking. The results are reported in the next section.

A second set of tests was also conducted using urine to determine the potential of the pneumatic pressure preconcentration with real biofluid samples. Human chorionic gonadotropin (HCG), or pregnancy hormone, was used as the target analyte for these tests, as it is one of the most commonly used rapid tests available for urine. Urine samples (purchased from Innovative Research, Inc.) were spiked with HCG to a concentration of 20 mIU mL⁻¹. Similarly to the PBS–influenza tests, 10 mL of spiked fluid was placed into the concentration setup, and the test procedure was repeated as before. Recordings of waste fluid weight were taken every five minutes rather than every 10 seconds (due to the lower flux of urine and its numerous



fouling solutes, Fig. 2). Again, since a much lower analyte recovery was expected using a biofluid which has more solutes to cause caking on the membrane, all of the membranes were swabbed with a cotton swab after testing to recover as much analyte as possible. The cotton swabs were then spun in a centrifuge with the final fluid sample as before to release the analyte from the swab back into the sample.

Concentration results for the influenza/PBS and HCG/urine tests were determined using ELISA kits. Using these results, the concentration amounts for each test were determined. From these values, the “% of theoretical concentration achieved” was calculated by dividing the actual concentration values by the maximum theoretical concentration values for each test. For example, if the actual concentration as measured by ELISA for a test was 100×, and the maximum theoretical concentration was 200×, then the ‘% of theoretical concentration achieved’ would be $100 \times / 200 \times = 50\%$.

Flux values recorded at each timepoint were normalized into values of $\text{mL min}^{-1} \text{cm}^{-2} \text{Pa}^{-1}$, by dividing the flux values by the exposed surface area of the membrane (2.29 cm^2) and the amount of pressure applied (5.7 MPa). The average value of the fluxes which were observed at each time point were calculated, and then used to determine how much fluid would be able to travel through a membrane of practical size (10 cm^2) over time at 5.7 MPa.

Results

The concentration test results can be seen in Fig. 4c. The concentration results show that swabbing the membrane and releasing analyte trapped on the surface greatly increases ($\sim 2\times$, from 34% to 76%) the amount of preconcentration which is achievable. Simply, the cotton swab recovers analyte lost due to caking. The ‘percent of maximum theoretical concentration achieved’ with the biofluid, urine, is also very high at 59%, but is lower than PBS samples which had been swabbed. This is likely because there are many more interferents and other small molecules present in urine which foul the membrane and bind to the analyte, making releasing the analyte back into the sample fluid more difficult. Impressive preconcentration amounts up to $165\times$ were achieved, even using urine (all preconcentration data can be found in the ESI†).

The flux results can be seen in Fig. 4d and e for PBS and urine, respectively. When using PBS, one can concentrate 10 mL of fluid in only 52 seconds if using a membrane which is 10 cm^2 and a carbon dioxide pressure source. If using urine, one can concentrate the same amount of fluid in 8.3 minutes, which is much slower but still adequate for a rapid diagnostic test. When comparing the flux data of PBS and urine, the effect of fouling on membrane preconcentration can be seen. While the flux curve for PBS is rather straight and shows only a slight decrease in fluid flow over time, the flux curve for urine shows a large decrease in fluid flow over

time, as represented by the flattening of the curve. However, a large volume of urine would still be able to be concentrated before the flux decreases significantly, implying that real biofluids remain a viable option for pneumatic preconcentration. It should be mentioned that fluxes have the potential to be even higher if using an alternative source of gas with a higher vapor pressure, such as oxygen or nitrogen. These gases can be purchased for use in mini cartridges, which have filling pressures up to 3000 psi, potentially increasing the flux $>3\times$. The preconcentration time can also be reduced by using a larger membrane area, something not performed here because it likely requires optimization for each application due to analyte-specific caking.

Summary of utility and challenges for membrane-based preconcentration

Based on preliminary results, both osmotic pressure and pneumatic pressure methods of preconcentration have potential applied value. However, each method has its own caveats and is more strongly suited to work under certain conditions, a summary of which can be seen in Table 2. Pneumatic pressure preconcentration is more suited for rapid diagnostic applications such as point-of-care tests, whereas osmotic preconcentration is more suited to in-line, continuous monitoring of analytes.

As stated in a previous section, for rapid diagnostics, pneumatic preconcentration with looser membrane MWCO’s (kDa or greater) provides a combination of pressure and membrane which can quickly preconcentrate large (mL’s) initial sample volumes into a preconcentrated sample of 10’s to 100’s of μL for the diagnostic assay. Continuous diagnostics are mainly limited mainly to small molecule analytes,⁵⁵ where the membrane must be much tighter (small MWCO) which increases the hydraulic resistance of the membrane, requiring even higher pressures that are most easily generated by osmotic pressure. Furthermore, the osmotic preconcentration is better suited to a continuous operating format because it would be highly complex to continuously introduce sample into a high-pressure chamber.

Although preconcentration has tremendous potential for use in clinical diagnostics, there are some additional application-specific challenges. Firstly, there are some biofluids such as saliva or blood which may be too viscous to be successfully filtered. In cases such as this, it is not impossible to work with the fluid, but fluid pretreatment may be necessary before preconcentration occurs, adding an extra step to the process for the clinician or home user. For example, for saliva there are devices available which use a fiberglass-like substance to filter the mucins and other large viscosity-increasing solutes out of the saliva.⁵⁶ However, this pretreated fluid would still be much more viscous than buffer and would still preconcentrate slowly (10’s of minutes or more). Multiple studies have found that saliva may be a viable fluid for the diagnoses of respiratory viruses as well as



Table 2 Comparison of pros, cons, and potential diagnostic applications for pneumatic pressure and osmotic pressure preconcentration

	Pneumatic pressure	Osmotic pressure
General	<ul style="list-style-type: none"> • Point-of-care tests • Larger molecules • Viral and disease detection in urine/buffer, toxins in water 	<ul style="list-style-type: none"> • Continuous/in-line monitoring • Small molecules • Continuous detection of dilute analytes in biofluids such as sweat and toxin detection in water and food processing
Positive aspects	<ul style="list-style-type: none"> • Point-of-care tests • Easier to re-use device casing • More portable 	<ul style="list-style-type: none"> • Continuous/in-line monitoring • Larger potential pressure gradient • Continuous
Negative aspects	<ul style="list-style-type: none"> • Caking may require mechanical agitation • Adds an extra step for user to point-of-care tests 	<ul style="list-style-type: none"> • Concentration polarization • Pore blocking
		<ul style="list-style-type: none"> • Generally more complex (requires sample dispenser, etc.)

other conditions.^{57–63} One potential solution to using more viscous/analyte-heavy fluids such as saliva, plasma, or whole blood may be using conical pored membranes. These membranes are created by etching the cylindrical pores found on track-etched membranes even further by using methods such as chemical etching or plasma etching.^{64–66} The shape of the conical pore, including the cone base diameter and cone angle, can be controlled by continuously etching one side of the membrane until the desired shape is achieved.⁶⁵ Studies have found that membranes with these pores have a significantly decreased resistance, and therefore a higher rate of fluid transport.^{64,67} This means that preconcentration could theoretically be achieved using less applied pressure,⁶⁷ increasing the number of potential applications for this preconcentration technology.

Another potential factor to consider when preconcentrating fluid samples, particularly biofluids, is how the sample will be affected by the preconcentration process itself. For example, concentrating urine with a tight membrane such as a forward osmosis membrane can alter pH and salinity which can then interfere with the measurement sensor or assay. Each of these types of changes may affect the functionality of the assay, as the assays are often developed to perform most accurately at very specific conditions which are similar to the natural form of the biofluid. It may be possible to add a post-treatment step to the preconcentrated sample, such as adding powdered buffer to restore the original pH of the solution. However, it is extremely costly for companies to alter any process for tests because they must obtain FDA approval for the new process as well as all test components. The FDA must also determine that the new version of the test performs equal to, or better than before. Therefore, diagnostic companies will ideally require significant performance gains and a simple route forward for FDA.

Conclusions

In conclusion, when considering the work of others^{15,17,19–21,27,28} and our groups work,^{29,32} membrane preconcentration remains attractive due to its simplicity compared other preconcentration techniques.^{13,14,16,18}

However, the applied value of membrane-based preconcentration is limited by the alternative of being able to use a more sensitive laboratory or clinic-based test. Therefore we speculate that membrane-based preconcentration's primary value is limited to applications where portability and detection speed are important. Even within that applications' space, numerous challenges such as interferences, membrane caking, and other practical issues will further limit the utility of this approach. Although it seems a potentially ubiquitous solution for creating higher sensitivity diagnostic tests in theory, membrane preconcentration will require further development before achieving greater practical use in rapid and continuous diagnostic applications. That said, in this work, we conclude that pneumatic and osmotic driven preconcentration have the most promise, with clear demonstrations of their utility including use with real biofluids.

Conflicts of interest

There are no conflicts to declare.

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