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A carboxylesterase-activatable near-infrared phototheranostic probe for tumor fluorescence imaging and photodynamic therapy†

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Phototheranostic probes have been proven to be a promising option for cancer diagnosis and treatment. However, near-infrared phototheranostic probes with specific tumor microenvironment responsiveness are still in demand. In this paper, a carboxylesterase (CES)-responsive near-infrared phototheranostic probe was developed by incorporating 6-acetamidohexanoic acid into a hemicyanine dye through an ester bond. The probe exhibits highly sensitive and selective fluorescence enhancement towards CES because CES-catalyzed cleavage of the ester bond leads to the release of the fluorophore. By virtue of its near-infrared analytical wavelengths and high sensitivity, the probe has been employed for endogenous CES activatable fluorescence imaging of tumor cells. Moreover, under 660 nm laser irradiation, the probe can generate toxic reactive oxygen species and efficiently kill tumor cells, with low cytotoxicity in dark. As far as we know, the probe was the first CES-responsive phototheranostic probe with both near-infrared analytical wavelengths and photosensitive capacity, which may be useful in the real-time and *in situ* imaging of CES as well as imaging-guided photodynamic therapy of tumors. Therefore, the proposed probe may have wide application prospect in cancer theranostics.

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Introduction

Phototheranostics, characterized by integrating photoinduced imaging and concurrent therapy into a system, has attracted much attention in the field of cancer theranostics. Among optical diagnostics, fluorescence imaging has the advantages of excellent sensitivity, easy operation, fast response and so on. In particular, fluorescence imaging with near-infrared (NIR) wavelength (650–900 nm) is more favorable for biomedical diagnosis due to the deeper tissue penetration, less photo-damage, and lower background fluorescence disturbance.^{1–5} For photoinduced therapeutics, photodynamic therapy (PDT) is capable of killing cancer cells selectively by generating venous singlet oxygen under laser irradiation, without noticeable damage to normal tissues in dark.^{6–8} The merits, including minimal invasiveness, precise controllability and negligible drug resistance, have made PDT a promising therapeutic option for cancer. Therefore, developing novel phototheranostic

probes with fluorescence imaging-guided PDT is rather beneficial to tumor diagnosis and efficient treatment.

Moreover, the phototheranostic probe responding to specific tumor microenvironment is the ideal agent for cancer diagnosis and therapy because of the improved tumor specificity. Such phototheranostic probe is generally activated by an endogenous initiator in tumor cells, such as hypoxia,^{9–11} acidic pH,^{12–14} abundant thiol species,^{15–17} reactive oxygen species,^{18,19} and some overexpressed enzymes.^{20–23} Carboxylesterase (CES) is one of the overexpressed enzymes in various tumor cells, which can catalyze the hydrolysis of carboxylic esters and be in action in the growth, invasion, as well as metastasis of malignant tumors.^{24–26} CES has thus been regarded as a potential target for tumor theranostics, and several CES-activated prodrugs for chemotherapy have been reported.^{27–32} Besides, some CES-responsive fluorescence probes for single imaging were proposed.^{33–45} These probes were generally designed by incorporating varied substrates with carboxyl groups into fluorophores with hydroxyl groups, which displayed rational sensitivity and selectivity toward CES. They have been used for bioimaging at cellular level or *in vivo*, verifying the feasibility of such sensing strategy. However, most of these probes had relatively low emission wavelength (<650 nm), and CES-responsive photo-theranostic probe with NIR analytical wavelengths is still absent.

Herein, a CES-responsive phototheranostic probe HCH-AcAH (Scheme 1) was developed for activatable NIR

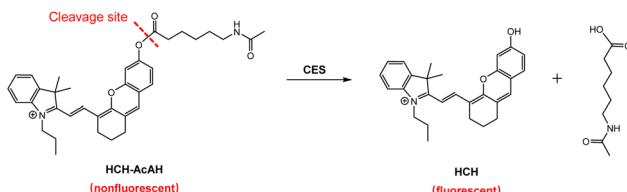
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Scheme 1 The structure and response mechanism of HCH-AcAH to CES.

fluorescence imaging and PDT of tumor cells. HCH-AcAH was synthesized by incorporating 6-acetamidohexanoic acid (AcAH) into the hemicyanine skeleton HCH *via* an ester bond. HCH was selected owing to its NIR analytical wavelengths, tunable fluorescence signal, superior photosensitive capacity, and high stability.^{46–48} The probe has almost no fluorescence signal due to the esterification of hydroxyl in HCH, which is desirable for sensitive detection. However, reaction with CES triggers the cleavage of ester bond, accompanied with the release of the fluorophore HCH and concomitant fluorescence enhancement, which results in the development of a sensitive and selective means for CES detection. Moreover, the released HCH also possesses photodynamic capacity, thereby damaging tumor cells. HCH-AcAH has been used not only for activatable fluorescence imaging of tumor cells with overexpressed CES, but also to efficiently killing tumor cells *via* photodynamic effect.

Experimental

Reagents and apparatus

CES was purchased from Sigma-Aldrich. IR-780 iodide and AcAH were purchased from J&K Scientific Co., Ltd (Beijing, China). Resorcinol and propidium iodide (PI) were obtained from Aladdin Biochemical Technology Co., Ltd (Shanghai, China). Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum (FBS), trypsin, glutathione (GSH), 2,7-dichlorodihydrofluorescein diacetate (DCFH-DA), 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT), and calcein acetoxymethyl ester (calcein-AM) were obtained from Solarbio Science & Technology Co., Ltd (Beijing, China). All the inorganic salts used were purchased from Beijing Chemical Industry Co., Ltd (Beijing, China). Human cervical cancer cell line (HeLa cell) was derived from Cell Bank of Chinese Academy of Sciences (Shanghai, China). Ultrapure water (over 19 MΩ cm) was produced by Heal Force (China).

¹H and ¹³C NMR spectra were scanned on an AVANCE III spectrometer (Bruker, Germany). High-resolution electrospray ionization mass spectrometry (HR-ESI-MS) was carried out using APEX IV FTMS instrument (Bruker Daltonics). An F-7000 fluorescence spectrophotometer (Hitachi, Japan) was used to record fluorescence spectra. A UH5300 UV-visible spectrophotometer (Hitachi, Japan) was employed to collect the absorption spectra. A SpectraMax190 microplate reader (Molecular Devices, USA) was used to determine the absorbance in MTT assay. Fluorescence imaging was conducted with an Eclipse Ts2 fluorescence microscope (Nikon, Japan). Confocal fluorescence

imaging was performed on an FV 3100-IX83 confocal laser scanning microscope (Olympus, Japan).

Synthesis and characterization of the probe HCH-AcAH

The synthesis route of the probe HCH-AcAH was shown in Scheme S1†. HCH was prepared according to our previous method.³⁴ Then, HCH (82 mg, 0.2 mmol) was added to a mixture of AcAH (52 mg, 0.3 mmol), 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydro-chloride (69 mg, 0.36 mmol) and 4-dimethylaminopyridine (4 mg, 0.32 mmol) in CH₂Cl₂ (6 mL). The reaction mixture was stirred for 1 h at ambient temperature. After evaporating the solvent under reduced pressure, the residue was further purified by column chromatography with CH₂Cl₂/CH₃OH as eluents, affording HCH-AcAH as a purple solid (28 mg, yield 24%). HR-ESI-MS: *m/z* calcd for HCH-AcAH (C₃₆H₄₂N₂O₄⁺, M⁺), 567.3217; found, 567.3214 (Fig. S1†). ¹H and ¹³C NMR spectra were shown in Fig. S2 and S3,† respectively. ¹H NMR (400 MHz, 298 K, CDCl₃): δ 8.68 (d, 1H, *J* = 15.2 Hz), 7.75 (s, 1H), 7.61 (d, 1H, *J* = 7.6 Hz), 7.48 (m, 4H), 7.23 (s, 1H), 7.14 (s, 1H), 7.04 (m, 1H), 6.62 (d, 1H, *J* = 15.2 Hz), 4.50 (t, 2H, *J* = 6.8 Hz), 3.28 (q, 2H, *J* = 6.8 Hz), 2.76 (m, 4H), 2.64 (t, 2H, *J* = 7.2 Hz), 2.44 (s, 2H), 2.07 (s, 3H), 1.99 (m, 4H), 1.83 (s, 6H), 1.66 (m, 2H), 1.50 (m, 2H), 1.11 (t, 3H, *J* = 7.2 Hz). ¹³C NMR (100 MHz, 298 K, CDCl₃): δ 178.6, 171.8, 171.1, 160.5, 153.1, 150.6, 146.5, 142.2, 141.3, 132.0, 131.6, 129.9, 129.4, 128.4, 128.1, 122.9, 119.5, 115.1, 113.2, 109.7, 105.5, 51.2, 47.6, 39.3, 34.3, 29.5, 29.1, 28.3, 26.4, 24.5, 24.4, 23.4, 21.6, 20.3, 11.6.

General procedure for CES detection

A certain amount of HCH-AcAH was dissolved in DMSO to prepare the storage solution (1 mM). For detecting the response of HCH-AcAH to CES, HCH-AcAH (10 μ M) was mixed with different concentrations of CES in PBS (pH 7.4). After incubation at 37 °C for 6 min, the reaction system was subjected to fluorescence measurement with $\lambda_{\text{ex/em}} = 680/710$ nm. As a control, HCH-AcAH in the absence of CES was determined under the same conditions.

ROS evaluation

To evaluate the ROS generation ability of the reaction system *in vitro*, DCFH-DA was utilized as the ROS indicator. The reaction system was first made by reacting HCH-AcAH (1 μ M) with CES (200 U L⁻¹) at 37 °C for 6 min. Then, the activated DCFH-DA (1 μ M) was added, and the mixture was exposed to 660 nm laser irradiation. The fluorescence spectra were monitored with $\lambda_{\text{ex}} = 485$ nm. For comparison, DCFH-DA alone was observed under the same condition.

Confocal fluorescence imaging of endogenous CES in HeLa cells

HeLa cells were inoculated and grew with adherence for 24 h. The culture medium was discarded and the cells were cleaned with PBS (pH 7.4) three times. Then, the cells were incubated with HCH-AcAH (10 μ M) for 10 min. For the inhibitor study, the cells were pretreated with inhibitor triphenyl phosphate (TPP,



10 μM) for 1 h before incubation with HCH-AcAH. The imaging was carried out on a confocal laser scanning microscope with 635 nm excitation and 650–750 nm emission.

Cytotoxicity evaluation

The dark and photo toxicity of HCH-AcAH to cells was determined by standard MTT assay.¹⁰ After digestion and centrifugation, HeLa cells were collected, counted, and inoculated in 96-well plates with a density of 7000–10000 cells per well. Cells were incubated at 37 °C for 24 hours. Different concentrations of HCH-AcAH (1–30 μM) in DMEM were added to each well and co-incubated with cells for 12 h. The media were replaced by fresh DMEM. Then, the cells in the group with laser were exposed to 660 nm laser irradiation (0.25 W cm⁻²) for 10 min, while the control group was placed in dark. After removing the old media, 100 μL MTT solution (0.5 mg mL⁻¹) was added and incubated for another 4 h. The supernatant was discarded and DMSO (100 μL) was added. Finally, the absorbance at 490 nm was measured, and the cell viability was calculated.

Calcein-AM/PI staining of cells

Adherent HeLa cells were incubated with HCH-AcAH (10 μM) at 37 °C for 10 min. After washed with PBS, the group with laser was irradiated by 660 nm laser (0.25 W cm⁻²) for 10 min, while the control group without laser was kept away from light. Then, cells in each group were treated by calcein-AM (4 μM) and PI (100 $\mu\text{g mL}^{-1}$). After 20 min, the cells were washed and photographed with a fluorescence microscope.

Results and discussion

Spectral properties of HCH-AcAH reacting with CES

The spectral changes of HCH-AcAH before and after reacting with CES were first studied. In the UV-vis absorption spectra (Fig. 1A), it was observed that HCH-AcAH had a maximum absorption peak at 585 nm, with two shoulder peaks at 553 and 628 nm, respectively (curve *a*). After interacting with CES, the maximum absorption peak shifted to 680 nm (curve *b*), along with the color of the solution changing from purple to blue (inset in Fig. 1A). Meanwhile, HCH-AcAH itself was nearly nonfluorescent due to the blocking of hydroxyl in HCH.⁴⁶ However, addition of CES induced strong fluorescence signal at 710 nm (Fig. 1B). Such significant fluorescence enhancement

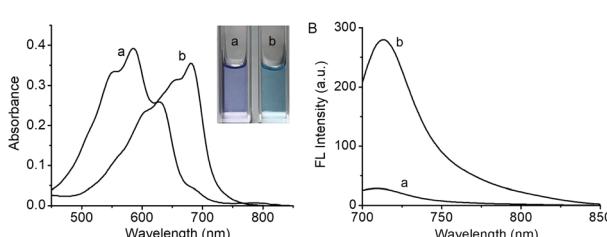


Fig. 1 The absorption (A) and fluorescence (B) spectra of HCH-AcAH (10 μM) before (a) and after (b) reacting with CES (200 U L⁻¹). $\lambda_{\text{ex}} = 680$ nm.

was highly favorable for the sensitive detection of CES. Moreover, both the absorption and fluorescence spectra of HCH-AcAH after reacting with CES resembled those of the fluorophore HCH (Fig. S4†), hinting that the reaction with CES triggered the release of HCH, thereby leading to the fluorescence enhancement. The formation of HCH was also verified by HR-ESI-MS analysis (Fig. S5†). The mass spectrum of the reaction solution showed a prominent peak at *m/z* = 412.2271, indicating the generation of the fluorophore HCH.

The influences of reaction time, temperature, and pH on the reaction system were then explored. The reaction kinetic curves (Fig. S6A†) showed that higher concentrations of CES induced quicker reaction rates and stronger fluorescence signals, and the fluorescence reached an approximate plateau within 6 min. By contrast, the fluorescence of HCH-AcAH itself had little change during the same period, suggesting the good stability of the probe in an aqueous environment. The fluorescence signal of either the probe or the reaction system was barely affected by the temperature increasing from 20 to 45 °C (Fig. S6B†). In addition, the strongest fluorescence enhancement was achieved at pH 7 (Fig. S6C†), which was attributed to the effects of pH on the fluorescence signal of HCH-AcAH and HCH, as well as the enzymatic activity of CES. The above results indicated that HCH-AcAH was suitable for the quick detection of CES at normal physiological conditions (37 °C, pH 7.4), which was thus employed in this work.

Under the above conditions, the fluorescence spectra of HCH-AcAH reacting with varying concentrations of CES were monitored. As depicted in Fig. 2, higher concentrations of CES induced larger fluorescence enhancement, and a good linearity could be achieved in the range of 5–40 U L⁻¹, with a linear equation of $\Delta F = 3.75 \times C + 19.92$ ($R = 0.988$), where ΔF means the fluorescence difference with and without CES. Using the formula $3s/m$ (s is the standard deviation of blank determinations, $n = 11$, and m is the slope of the linear equation), the detection limit was 0.712 U L⁻¹, which was lower than the existed NIR fluorescent probes for CES.^{39,40}

Next, the selectivity of HCH-AcAH for CES was evaluated by testing some probable coexisting substances covering inorganic salts (CaCl₂, FeCl₃, CuSO₄, and MgCl₂), amino acids (cysteine, tyrosine, proline, and methionine), ascorbic acid, GSH, and several enzymes (nitroreductase and hyaluronidase). The results (Fig. S7†) manifested that species except for CES scarcely

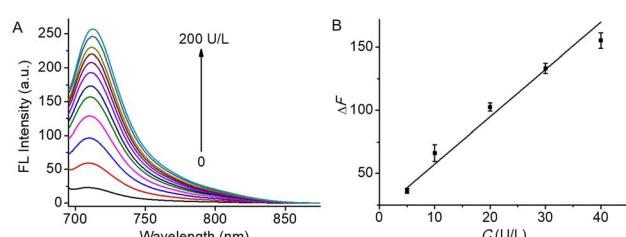


Fig. 2 (A) Fluorescence responses of HCH-AcAH (10 μM) to varying concentrations of CES (0, 5, 10, 20, 30, 40, 50, 60, 70, 80, 100, and 200 U L⁻¹). (B) Linear fitting curve of ΔF toward the content (C) of CES from 5 to 40 U L⁻¹. $\lambda_{\text{ex/em}} = 680/710$ nm.



affected the fluorescence signal of the probe, implying the excellent specificity of probe HCH-AcAH towards CES, which was ascribed to the specific cleavage of the ester bond by CES.

To further verify that the enhanced fluorescence resulted from the CES-mediated hydrolyzation, the effect of TPP, an inhibitor of CES,⁴⁹ was studied on the enzyme's activity. As was seen in Fig. S8†, 100 nM and 500 nM TPP reduced 45 and 71% of the fluorescence intensity compared with the fluorescence in the absence of TPP (curve *b*), respectively, demonstrating that the fluorescence off-on response was indeed triggered by CES-catalyzed cleavage.

In order to evaluate the photodynamic effect of the reaction system, DCFH-DA was utilized to indicate the formation of ROS. As depicted in Fig. 3, upon the addition of DCFH-DA and irradiation with 660 nm laser, the reaction system showed gradually increased fluorescence at 525 nm with a dependency on the irradiation time. As a control, the fluorescence of DCFH-DA itself scarcely varied under the same condition, verifying the laser-induced generation of ROS in the reaction system of HCH-AcAH and CES. In addition, HCH-AcAH itself also induced the formation of ROS (Fig. S9†). Even so, due to the precise controllability of the laser irradiation, HCH-AcAH may have great potential in the selective PDT of tumor with overexpressed CES.

Fluorescence imaging of endogenous CES in living cells

Encouraged by the high sensitivity and selectivity as well as the NIR analytical wavelengths of HCH-AcAH, we further explored its applicability in the real-time imaging of endogenous CES in living cells. It has been reported that CES was overexpressed in various tumors, including cervical cancer and liver cancer.^{50–52} And cervical cancer cell line HeLa has been used as the model cell in many studies on CES.^{33,39–41} Hence HeLa cells were chosen in the following cellular study. As shown in Fig. 4, HeLa cells alone emitted no background fluorescence, which profited from the long excitation wavelength (635 nm). However, upon

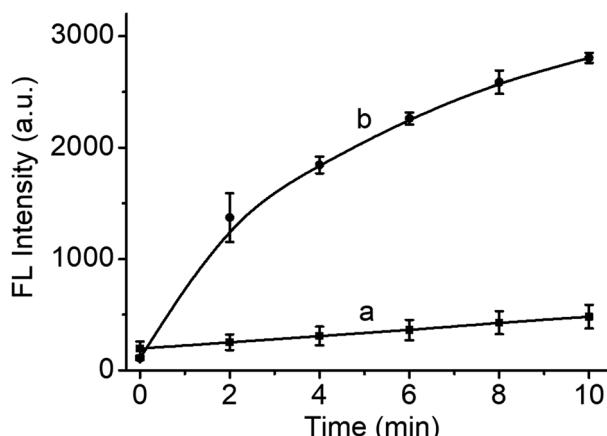


Fig. 3 Fluorescence changes of DCFH-DA in PBS (a) or in the reaction solution (b) of HCH-AcAH (1 μ M) and CES (200 $U\ L^{-1}$) upon 660 nm laser ($0.25\ W\ cm^{-2}$) for different irradiation time (0–10 min). $\lambda_{ex/em} = 485/525\ nm$.

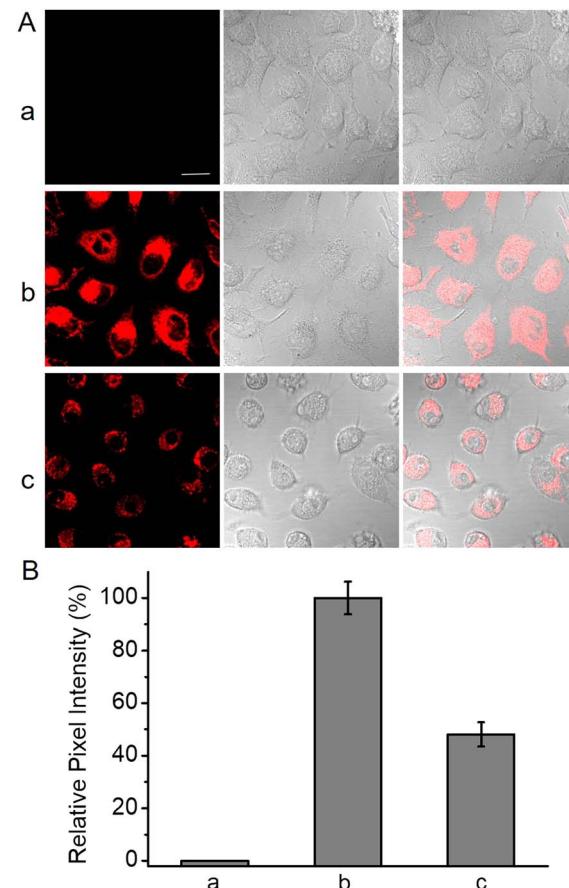


Fig. 4 (A) Fluorescence images of HeLa cells. (a) HeLa cells only (control), (b) HeLa cells incubated with HCH-AcAH (10 μ M) for 10 min, (c) HeLa cells were pretreated with TPP (10 μ M) for 1 h before incubation with HCH-AcAH (10 μ M) for 10 min. Scale bar, 20 μ m. (B) Relative pixel intensity of the corresponding images in panel A (the pixel intensity from image (b) is defined as 100%). The results are the mean \pm standard deviation of three separate measurements.

treatment with HCH-AcAH, bright red fluorescence was observed in the cytoplasm of HeLa cells, with negligible fluorescence in the nucleus. The reason for this phenomenon may be the location behaviors of CES³⁵ and the generated fluorophore HCH, which mainly resided in the cytoplasm instead of nucleus (Fig. S10†). Moreover, the fluorescence signal in cells with TPP (inhibitor of CES) pre-incubation was largely depressed (group c in Fig. 4), revealing the enhanced fluorescence in HeLa cells must be ascribed to the endogenous CES. Thus, the probe has great potential in imaging tumor cells with overexpressed CES.

PDT capacity of HCH-AcAH at the cellular level

Given the ROS generation and cell membrane permeability of HCH-AcAH, its PDT capacity at the cellular level was investigated. With DCFH-DA as the ROS imaging agent, we monitored the ROS formation in HCH-AcAH treated cells (Fig. S11†). The cells with either HCH-AcAH incubation or laser irradiation showed no remarkable fluorescence, indicating the probe or laser alone could not generate ROS. However, upon exposure to

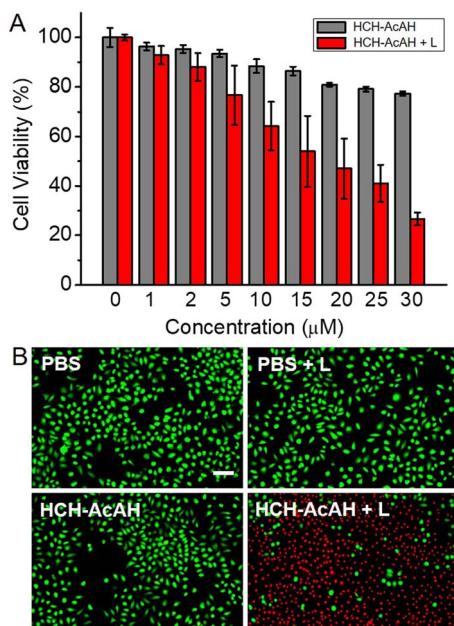


Fig. 5 (A) Viability of HCH-AcAH treated HeLa cells without or with 660 nm laser irradiation (0.25 W cm^{-2} , 10 min). The viability of cells in the absence of HCH-AcAH and laser irradiation was defined as 100%. (B) Fluorescence images of calcein-AM/PI staining HeLa cells after different treatments. Scale bar, 100 μm .

660 nm laser for 10 min, the HCH-AcAH incubated cells exhibited intense green fluorescence, which is the direct evidence of ROS formation in cells. Whereafter, the cell viability was assessed with MTT and calcein-AM/PI staining assays. As can be seen in Fig. 5A, HeLa cells incubated with HCH-AcAH without laser kept relatively high viability (about 80%) even at the concentration of 30 μM , suggesting the low cytotoxicity of the probe in dark. However, once the HCH-AcAH treated cells were irradiated by 660 nm laser, the survival rates significantly reduced with concentration-dependency, which was attributed to the PDT effect of the probe. Consistent results were obtained from calcein-AM/PI staining (Fig. 5B). HCH-AcAH or laser alone could barely affect the cell viability owing to the intense green and negligible red fluorescence in cells, implying the low dark cytotoxicity of HCH-AcAH and high safety of laser used in this work. On the contrary, laser irradiation on HCH-AcAH treated cells induced bright red intracellular fluorescence, indicative of dead cells.¹⁰ These observations suggested that the probe HCH-AcAH possessed excellent PDT capacity at the cellular level.

Conclusions

In summary, the first NIR phototheranostic probe HCH-AcAH was developed for CES-activatable tumor fluorescence imaging and concomitant PDT. The probe showed highly sensitive and selective NIR fluorescence enhancement ($\lambda_{\text{ex/em}} = 680/710 \text{ nm}$) to CES, with the detection limit of 0.712 U L^{-1} . The response mechanism relied on the CES-catalyzed cleavage of ester bond, along with the liberation of HCH. Moreover, significant laser-induced ROS generation was achieved in the reaction system

of HCH-AcAH and CES. Compared with the reported fluorescence probes for CES, the proposed probe exhibited NIR analytical wavelengths, low detection limit, and prominent photosensitive capacity (Table S1†). These merits made HCH-AcAH useful in the real-time and *in situ* imaging of CES as well as imaging-guided PDT of tumors. The experiments at cellular level verified that HCH-AcAH could indeed be applied in endogenous CES-activatable fluorescence imaging of tumor cells. In addition, HCH-AcAH displayed low cytotoxicity in dark while conspicuous PDT-mediated cell death was observed upon 660 nm laser irradiation. The simplicity and great performance of HCH-AcAH may make it potential in cancer diagnosis and treatment.

Author contributions

L. H. Li: conceptualization, writing-review & editing, funding acquisition, supervision; Q. Zhang: methodology, writing-original draft; J. J. Li: software; Y. F. Tian: validation; J. Y. Li: validation; W. Liu: resources, project administration; H. P. Diao: funding acquisition, supervision.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- Z. M. Yang, Z. J. Zhang, Z. Q. Lei, D. Wang, H. C. Ma and B. Z. Tang, *ACS Nano*, 2021, **15**, 7328.
- A. Antonucci, M. Reggente, C. Roullier, A. J. Gillen, N. Schuergers, V. Zubkovs, B. P. Lambert, M. Mouhib, E. Carara, L. Dini and A. A. Boghossian, *Nat. Nanotechnol.*, 2022, **12**, 1111.
- Q. Zhang, L. Wu, S. Z. Liu, Q. J. Chen, L. P. Zeng, X. Z. Chen and Q. Zhang, *Cancer Cell Int.*, 2020, **20**, 570.
- A. Baghdasaryan, F. F. Wang, F. Q. Ren, Z. R. Ma, J. C. Li, X. T. Zhou, L. Grigoryan, C. Xu and H. J. Dai, *Nat. Commun.*, 2022, **12**, 5613.
- T. Setsuko and J. Takashi, *RSC Adv.*, 2020, **10**, 28171.
- S. S. Hafiz, M. Xavierselvan, S. Gokalp, D. Labadini, S. Barros, J. Duong, M. Foster and S. Mallidi, *ACS Appl. Nano Mater.*, 2022, **5**, 6125.



7 W. C. Shen, T. T. Hu, X. Y. Liu, J. J. Zha, F. Q. Meng, Z. K. Wu, Z. L. Cui, Y. Yang, H. Li, Q. H. Zhang, L. Gu, R. Z. Liang and C. L. Tan, *Nat. Commun.*, 2022, **13**, 3384.

8 W. Kang, Y. Tian, Y. Zhao, X. D. Yin and Z. G. Teng, *RSC Adv.*, 2022, **12**, 16927.

9 F. Xu, H. D. Li, Q. C. Yao, H. Y. Ge, J. L. Fan, W. Sun, J. Y. Wang and X. J. Peng, *Chem. Sci.*, 2019, **10**, 10586.

10 L. H. Li, L. Ding, X. Y. Zhang, D. N. Wen, M. Zhang, W. Liu, H. J. Wang, B. Wang, L. L. Yan, L. X. Guo and H. P. Diao, *Spectrochim. Acta, Part A*, 2022, **267**, 120579.

11 H. P. Lee and A. K. Gaharwar, *Adv. Sci.*, 2020, **7**, 2000863.

12 R. Wang, K. H. Kim, J. Yoo, X. S. Li, N. Kwon, Y. H. Jeon, S. H. Shin, S. S. Han, D. S. Lee and J. Yoon, *ACS Nano*, 2022, **16**, 3045.

13 M. J. Zhao, Q. Zeng, X. P. Li, D. Xing and T. Zhang, *Nano Res.*, 2022, **15**, 716.

14 X. Xie, K. L. Jiang, B. W. Li, S. L. Hou, H. L. Tang, B. H. Shao, Y. Ping and Q. Q. Zhang, *Biomaterials*, 2022, **286**, 121598.

15 Z. Yang, W. P. Fan, W. Tang, Z. Y. Shen, Y. L. Dai, J. B. Song, Z. T. Wang, Y. Liu, L. S. Lin, L. L. Shan, Y. J. Liu, O. Jacobson, P. F. Rong, W. Wang and X. Y. Chen, *Angew. Chem., Int. Ed.*, 2018, **57**, 14101.

16 Y. Q. Zhang, J. Fang, S. Y. Ye, Y. Zhao, A. N. Wang, Q. L. Mao, C. X. Cui, Y. L. Feng, J. C. Li, S. N. Li, M. Y. Zhang and H. B. Shi, *Nat. Commun.*, 2022, **13**, 1685.

17 G. D. Ren, Z. C. Wang, Y. F. Tian, J. Y. Li, Y. Y. Ma, L. Zhou, C. W. Zhang, L. X. Guo, H. P. Diao, L. H. Li, L. Lu, S. F. Ma, Z. F. Wu, L. L. Yan and W. Liu, *Biomed. Pharmacother.*, 2022, **153**, 113506.

18 J. X. Wang, W. Y. Kong, H. S. Jin, C. L. Li, Q. Luo, Y. Luo, C. P. Yuan, J. Lu, L. Zhang and X. J. Liu, *Colloids Surf.*, 2022, **218**, 112750.

19 Z. Y. Xu, J. Q. Qian, C. Meng, Y. Liu, Q. Ding, H. M. Wu, P. Li, F. S. Ran, G. Q. Liu, Y. Y. Wang and Y. Ling, *Theranostics*, 2022, **12**, 2535.

20 X. Zhou, H. D. Li, C. Shi, F. Xu, Z. Zhang, Q. C. Yao, H. Ma, W. Sun, K. Shao, J. J. Du, S. R. Long, J. L. Fan, J. Y. Wang and X. J. Peng, *Biomaterials*, 2020, **253**, 120089.

21 J. Yuan, Q. H. Zhou, S. Xu, Q. P. Zuo, W. Li, X. X. Zhang, T. B. Ren, L. Yuan and X. B. Zhang, *Angew. Chem., Int. Ed.*, 2022, e202206169.

22 F. Y. Zhou, S. K. Yang, C. Zhao, W. W. Liu, X. F. Yao, H. Yu, X. L. Sun and Y. Liu, *Theranostics*, 2021, **11**, 7045.

23 J. Zheng, Y. Z. Liu, F. L. Song, L. Jiao, Y. N. Wu and X. J. Peng, *Chem. Commun.*, 2020, **56**, 5819.

24 B. J. Aguilar, A. T. Nkembo, R. Duverna, R. A. Poku, F. Amissah, S. Y. Ablordeppéy and N. S. Lamango, *Eur. J. Med. Chem.*, 2014, **81**, 323.

25 H. N. He, L. Sun, J. X. Ye, E. G. Liu, S. H. Chen, Q. L. Liang, M. C. Shin and V. C. Yang, *J. Controlled Release*, 2016, **240**, 67.

26 A. Sharma, E. J. Kim, S. Mun, M. S. Ji, B. G. Chung and J. S. Kim, *Dyes Pigm.*, 2019, **163**, 62.

27 X. P. Ma, X. M. Huang, Z. Moore, G. Huang, J. A. Kilgore, Y. G. Wang, S. Hammer, N. S. Williams, D. A. Boothman and J. M. Gao, *J. Controlled Release*, 2015, **200**, 201.

28 P. E. Saw, H. R. Yao, C. H. Lin, W. Tao, O. C. Farokhzad and X. D. Xu, *Nano Lett.*, 2019, **19**, 5967.

29 M. R. Hellmich, C. Chao, K. Modis, Y. Ding, J. R. Zatarain, K. Thanki, M. Maskey, N. Druzhyna, A. A. Untereiner, A. Ahmad, Y. Xue, H. Y. Chen, W. K. Russell, J. M. Wang, J. Zhou and C. Szabo, *Biomolecules*, 2021, **11**, 1073.

30 Q. Huang, X. Liu, H. Y. Wang, X. R. Liu, Q. Z. Zhang, K. Li, Y. Chen, Q. Zhu, Y. Q. Shen and M. H. Sui, *Acta Biomater.*, 2022, **137**, 262.

31 C. Gao, X. L. Liang, S. Y. Mo, N. S. Zhang, D. S. Sun and Z. F. Dai, *ACS Appl. Mater. Interfaces*, 2018, **10**, 3219.

32 J. B. Zhuang, N. Li, Y. L. Zhang, B. L. Li, H. Q. Wen, X. C. Zhang, T. Y. Zhang, N. Zhao and B. Z. Tang, *CCS Chem.*, 2021, **3**, 1964.

33 Y. Y. Zhang, W. Chen, D. Feng, W. Shi, X. H. Li and H. M. Ma, *Analyst*, 2012, **137**, 716.

34 K. Kailass, O. Sadovski, W. R. Zipfel and A. A. Beharry, *J. Med. Chem.*, 2022, **65**, 8855.

35 J. N. Dai, Y. D. Hou, J. C. Wu and B. X. Shen, *ChemistrySelect*, 2020, **5**, 11185.

36 Y. Shu, C. Q. Huang, H. J. Liu, F. Y. Hu, H. M. Wen, J. Liu, X. Z. Wang, C. X. Shan and W. Li, *Spectrochim. Acta, Part A*, 2022, **281**, 121529.

37 J. Q. Dong, J. B. Gao and Y. Wang, *Dyes Pigm.*, 2022, **205**, 110549.

38 H. Zhou, J. B. Tang, J. Zhang, B. C. Chen, J. F. Kan, W. F. Zhang, J. Zhou and H. M. Ma, *J. Mater. Chem. B*, 2019, **7**, 2989.

39 M. Y. Li, C. Zhai, S. Y. Wang, W. X. Huang, Y. G. Liu and Z. Li, *RSC Adv.*, 2019, **9**, 40689.

40 J. M. Wang, Z. D. Teng, L. Zhang, Y. X. Yang, J. Qian, T. Cao, Y. P. Cao, W. W. Qin, Y. Liu and H. C. Guo, *ACS Sens.*, 2020, **5**, 3264.

41 L. Peng, S. D. Xu, X. K. Zheng, X. M. Cheng, R. Y. Zhang, J. Liu, B. Liu and A. J. Tong, *Anal. Chem.*, 2017, **89**, 3162.

42 J. Wang, W. B. Xu, Z. C. Yang, Y. C. Yan, X. X. Xie, N. Qu, Y. Wang, C. Y. Wang and J. L. Hua, *ACS Appl. Mater. Interfaces*, 2018, **10**, 31088.

43 Y. Y. Sun, X. N. Zhou, L. Y. Sun, X. X. Zhao, Y. R. He, G. Gao, W. N. Han and J. Zhou, *Chin. Chem. Lett.*, 2022, **33**, 4229.

44 Y. Liu, Z. X. He, Y. T. Yang, X. H. Li, Z. F. Li and H. M. Ma, *Biosens. Bioelectron.*, 2022, **211**, 114392.

45 S. Y. Liu, X. T. Zou, Y. Guo and X. Gao, *Anal. Chim. Acta*, 2022, **1221**, 340126.

46 L. H. Li, W. Shi, X. F. Wu, Q. Y. Gong, X. H. Li and H. M. Ma, *Biosens. Bioelectron.*, 2016, **81**, 395.

47 L. Yuan, W. Y. Lin, S. Zhao, W. S. Gao, B. Chen, L. W. He and S. S. Zhu, *J. Am. Chem. Soc.*, 2012, **134**, 13510.

48 L. H. Li, M. Zhang, L. Ding, G. D. Ren, X. Y. Hou, W. Liu, H. J. Wang, B. Wang, L. L. Yan and H. P. Diao, *Spectrochim. Acta, Part A*, 2021, **252**, 119518.

49 A. L. Eiden, P. E. Kaufman, F. M. Oi, M. J. Dark, J. R. Bloomquist and R. J. Miller, *Med. Vet. Entomol.*, 2007, **31**, 243.

50 J. G. Wang, Q. Q. Chen, N. Tian, W. P. Zhu, H. Zou, X. S. Wang, X. K. Li, X. L. Fan, G. Y. Jiang and B. Z. Tang, *J. Mater. Chem. B*, 2018, **6**, 1595.



51 K. Na, M. Kim, C. Y. Kim, J. S. Lim, J. Y. Cho, H. Shin, H. J. Lee, B. J. Kang, D. H. Han, H. Kim, J. H. Baik, M. S. Lange, J. Karl and Y. Paik, *J. Proteome Res.*, 2020, **19**, 4867.

52 P. R. Jensen, S. C. Serra, L. Miragoli, M. Karlsson, C. Cabella, L. Poggi, L. Venturi, F. Tedoldi and M. H. Lerche, *Int. J. Cancer*, 2015, **136**, E117.

