Check for updates

The radical scavenging activity of glycozolidol in physiological environments: a quantum chemical study†

Le Trung Hieu,[a] Mai Van Bay,[b] Nguyen Thi Hoa,[c] Adam Mechler [ID][d] and Quan V. Vo [ID] *[c]

Glycozolidol was isolated from the root of *Glycosmis pentaphylla* (6-hydroxy-2-methoxy-3-methylcarbazole, **GLD**). This molecule attracted considerable interest due to its beneficial biological activities that likely stem from its antioxidant activity; yet, the radical scavenging action of **GLD** has not been investigated thus far. In this study, DFT calculations were used to estimate the radical scavenging activity of **GLD** against a variety of biologically significant radical species in physiological environments. The findings demonstrated that **GLD** exerts significant antiradical activity in water at pH = 7.40 and in pentyl ethanoate (as a model of lipidic media) with $k_{overall} = 8.23 \times 10^6$ and 3.53×10^4 M^{-1} s^{-1}, respectively. In aqueous solution, the sequential proton-loss electron transfer mechanism made the highest contribution to the activity, whereas in nonpolar solvents the formal hydrogen transfer mechanism dominated the activity. **GLD** is predicted to have strong antiradical activity against CH$_3$O$^{\bullet}$, CH$_3$OO$^{\bullet}$, CCl$_3$OO$^{\bullet}$, NO$_2$, SO$_4^{\bullet-}$, DPPH and ABTS$^{+\bullet}$ $k_{app} \approx 10^9$ M^{-1} s^{-1} and $k_f \approx 10^6$ M^{-1} s^{-1}. The results suggest that **GLD** is a good radical scavenger in physiological environments.

1. Introduction

Antioxidants attract considerable interest owing to their roles in important biological processes, and therefore their preferential inclusion in food and pharmaceutical products.[1,2] They protect against oxidative deterioration in the body and thus against oxidative stress-induced pathological processes such as cancer, aging, hypo sexuality, hyperlipidemia, cardiovascular disease, inflammation, and many others.[1,3,4]

It is estimated that more than 10 000 individual phytochemicals have been identified in plants, including polysaccharides, phenolics, triterpenoids, steroids, carotenoids, vitamins, essential oils, and alkaloids. Among the great structural diversity of phytochemicals, alkaloid components have attracted considerable attention for known or suspected activity in treating various diseases.[4,5] Thus, highly substituted carbazole alkaloids are active against *e.g.* neurodegenerative diseases, cancer, tuberculosis and Human Immunodeficiency Virus

infections.[6,7] Among these compounds, glycozolidol (6-hydroxy-2-methoxy-3-methylcarbazole) (Fig. 1) was isolated from the roots of *Glycosmis pentaphylla* and since confirmed to exert antibacterial (against both Gram-positive and Gram-negative bacteria) and antifungal activities.[8,9] Glycozolidol belongs to a class of natural aromatic nitrogen heterocyclic alkaloids. Based on the oxygenation pattern of tricyclic carbazole alkaloids, glycozolidol could provide robust antioxidant activity due to the presence of amine and quinone groups, however, there are no reports on the mechanism and/or kinetics of the radical scavenging activity of glycozolidol. Herein, we explore the effects of solvent environments and molecular structures on the antioxidant activity and oxidation resistance of glycozolidol against a range of free radicals by using thermodynamic and kinetic calculations.

2. Computational details

All density functional theory (DFT) computations in this work were performed with the Gaussian 16 suite of programs[10] using
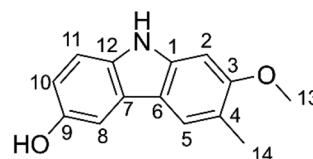
[a]University of Sciences, Hue University, Thua Thien Hue 530000, Vietnam

[b]The University of Danang, University of Science and Education, Danang 550000, Vietnam

[c]The University of Danang, University of Technology and Education, Danang 550000, Vietnam. E-mail: vvquan@ute.udn.vn

[d]Department of Biochemistry and Chemistry, La Trobe University, Victoria 3086, Australia

Fig. 1 Chemical structure of glycozolidol (**GLD**).

the M06-2X functional and the 6-311++G(d,p) basis set.[11] The M06-2X functional is one of the most reliable methods for computing the thermodynamics and kinetics of radical reactions.[12,13] This method is often used to evaluate the radical scavenging activity of natural and synthesized compounds[12,14–17] due to its reliable predictions when compared to experimental data.[18–22] The solvent effects of water and pentyl ethanoate were predicted using the SMD technique,[23] following a well-established practice in modelling the radical scavenging activity of antioxidants.[12,21,24]

The proton affinity (PA), bond dissociation enthalpy (BDE), and ionization energy (IE) values were calculated as follows.[21]

$$PA = H(\textbf{GLD}^-) + H(H^+) - H(\textbf{GLD}–H) \qquad (1)$$

$$BDE = H(\textbf{GLD}^{\textbf{·}}) + H(H^{\textbf{·}}) - H(\textbf{GLD}–H) \qquad (2)$$

$$IE = H(\textbf{GLD}–H^{+\textbf{·}}) + H(e^-) - H(\textbf{GLD}–H) \qquad (3)$$

where H(e), H(H$^+$), H(H$^{\textbf{·}}$), H($\textbf{GLD}$–H$^{+\textbf{·}}$), H($\textbf{GLD}^-$), H($\textbf{GLD}^{\textbf{·}}$), and H($\textbf{GLD}$–H) are the relative enthalpies of the electron, proton, hydrogen atom, cation-radical, anion, radical and neutral molecule, respectively.

The quantum mechanics-based test for overall free radical scavenging activity (QM-ORSA) protocol[20] was used to complete the kinetic calculations. The rate constants (k) were calculated using conventional transition state theory (TST) at 1 M standard state, 298.15 K, following eqn (4).[21,25–31]

$$k = \sigma\kappa\frac{k_B T}{h}e^{-(\Delta G^{\neq})/RT} \qquad (4)$$

where s is the reaction symmetry number,[32,33] k stands for tunneling corrections that were calculated using Eckart barrier,[34] k_B is the Boltzmann constant, h is the Planck constant, $\Delta G^{\neq}$ is Gibbs free energy of activation.

The Marcus theory was used to calculate the reaction barriers of single electron transfer (SET) reactions in media.[35,36] The eqn (5) and (6) were used to compute the Gibbs free energy change of reaction $\Delta G^{\neq}$ for the SET reaction.

$$\Delta G^{\neq}_{SET} = \frac{\lambda}{4}\left(1 + \frac{\Delta G^0_{SET}}{\lambda}\right)^2 \qquad (5)$$

$$\lambda \approx \Delta E_{SET} - \Delta G^0_{SET} \qquad (6)$$

Where ΔG^0_{SET} represents the conventional Gibbs free energy change of the reaction and ΔE represents the nonadiabatic energy difference between reactants and vertical products for SET.[37,38]

For rate constants around the diffusion limit, a modification was made.[20] Collins–Kimball theory was used to calculate the apparent rate constants (k_{app}) for an irreversible bimolecular diffusion-controlled reaction in solvents at 298.15 K,[39] and the literature was used to estimate the steady-state Smoluchowski rate constant (k_D).[20,40]

$$k_{app} = \frac{k_{TST}k_D}{k_{TST} + k_D} \qquad (7)$$

$$k_D = 4\pi R_{AB}D_{AB}N_A \qquad (8)$$

$D_{AB} = D_A + D_B$ (denotes the mutual diffusion coefficient of A and B),[39,41] where D_A or D_B is obtained using the Stokes–Einstein formulation eqn (9).[42,43]

$$D_{A \text{ or } B} = \frac{k_B T}{6\pi h a_{A \text{ or } B}} \qquad (9)$$

η is the viscosity of the solvents (*i.e.* η(pentyl ethanoate) = 8.62×10^{-4} Pa s, η(H$_2$O) = 8.91×10^{-4} Pa s) and a is the radius of the solute.

The existence of just one imaginary frequency served as a distinguishing feature across all of the transition stages. Intrinsic coordinate calculations were carried out to ensure that each transition state has an accurate connection to both the pre- and post-complexes.

To avoid over-penalizing entropy losses in solution, Okuno's corrections were applied,[44] which were then updated with the free volume theory utilizing the Benson correction.[20,45–47]

Values for pK_a are computed in accordance with Galano *et al.* p$K_a = m\Delta G_{(BA)} + C_0$, where m and C_0 are fitted parameters taken directly from ref. 48 (the M06-2X/6-311++G(d,p) approach), and ΔG(BA) is the difference in Gibbs free energy between the conjugated base (B) and the corresponding acid (A).

3. Results and discussion

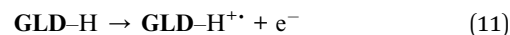### 3.1 The thermodynamic study

The **GLD** (**GLD**–H) can react with free radicals (R$^{\textbf{·}}$) *via* either of the three main pathways:
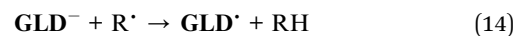
- Formal hydrogen transfer (FHT)[21,49,50]

$$\textbf{GLD}–H + R^{\textbf{·}} \rightarrow \textbf{GLD}^{\textbf{·}} + RH \qquad (10)$$
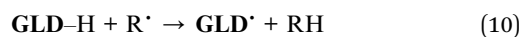
- Sequential electron transfer–proton transfer (SETPT)[51,52]

$$\textbf{GLD}–H \rightarrow \textbf{GLD}–H^{+\textbf{·}} + e^- \qquad (11)$$

$$\textbf{GLD}–H^{+\textbf{·}} + R^- \rightarrow \textbf{GLD}^{\textbf{·}} + RH \qquad (12)$$

- Sequential proton loss electron transfer (SPLET)[52–55]

$$\textbf{GLD}–H \rightarrow \textbf{GLD}^- + H^+ \qquad (13)$$

$$\textbf{GLD}^- + R^{\textbf{·}} \rightarrow \textbf{GLD}^{\textbf{·}} + RH \qquad (14)$$

In the initial step, the characteristic thermodynamic parameters (BDE, PA, IE) that define the energy barrier of the first step of each mechanism were calculated for all bonds in the HOO$^{\textbf{·}}$ radical scavenging activity of **GLD** in the gas phase (the standard medium for computational chemistry), in pentyl ethanoate for a lipid medium, and in water at pH 7.4 for an aqueous physiological environment. Table 1 provides an overview of the findings.

The BDEs ranged from 82.8 to 141.9 kcal mol^{-1}, whereas the PA and IE values were in the range of 84.7–343.6 and 99.2–165.2

Table 1 The computed thermodynamic parameters (BDE, PA, IE in kcal mol^{-1}) of **GLD** in the studied media (G: the gas phase; P: pentyl ethanoate; W: water)

Positions	BDE			PA			IE		
	G	P	W	G	P	W	G	P	W
C2–H	136.7	116.0	132.0				165.2	122.1	99.2
C5–H	114.4	114.9	115.3						
C8–H	115.6	116.3	129.2						
C10–H	116.6	138.6	116.7						
C11–H	141.9	115.9	132.8						
C13–H	99.9	101.4	101.9						
C14–H	92.7	92.8	92.6						
O9–H	85.2	83.1	82.8	343.6	116.2	84.7			
N–H	91.6	92.5	90.4						

kcal mol^{-1}, respectively. The BDE values varied somewhat randomly, while the PAs and IEs reduced in correlation to the dielectric constant of solvents. The lowest BDE of C–H bond was observed at C14–H (BDE = 92.7, 92.8 and 92.6 kcal mol^{-1} in the gas phase, pentyl ethanoate and water, respectively) that is still higher than BDEs of N–H and O9–H by about 0.3–9.8 kcal mol^{-1}. The lowest BDE was found at the O9–H bond (85.2, 83.1 and 82.8 kcal mol^{-1} in the gas phase, pentyl ethanoate and water, respectively). The SPLET and SETPT mechanisms are not favorable in the gas phase or pentyl ethanoate solvent due to the higher PA and IE values compared with the BDE(O9–H). Thus it is expected that the O9–H bond will play a dominant role in the radical scavenging activity of **GLD** following the FHT reaction in all of the studied media; however, in the aqueous solution, the

Table 2 The computed ΔG^0 (in kcal mol^{-1}) of the HOO$^{\bullet}$ + **GLD** following the formal hydrogen transfer (FHT), proton loss (PL) and single electron transfer (SET) reactions in the studied media (G: the gas phase; P: pentyl ethanoate; W: water)

Positions	FHT			PL			SET		
	G	P	W	G	P	W	G	P	W
C2–H	48.3	25.8	41.3				143.1	60.7	54.3
C5–H	24.4	25.9	23.7						
C8–H	26.5	26.5	38.6						
C10–H	27.2	51.5	25.2						
C11–H	53.1	27.1	42.0						
C13–H	10.9	12.9	10.6						
C14–H	4.1	4.8	1.7						
O9–H	−3.6	−4.3	−7.3	188.4	130.2	89.3			
N–H	3.0	3.7	0.1						

SPLET mechanism may have a major contribution in the radical scavenging activity if the spontaneous dissociation of the acidic protons eliminates the energy barrier of the first step.

The Gibbs free energies of the **GLD** + HOO$^{\bullet}$ reaction following the proton loss (PL-the first step of SPLET), the FHT, and single electron transfer (SET-the first step of SETPT) mechanisms were calculated in order to eliminate pathways that are not spontaneous thermodynamically, and rate constants were only calculated for the spontaneous reactions. The results are shown in Table 2. The results showed that the process was spontaneous only according to the FHT mechanism, especially at the O9–H bond ($\Delta G^0 = -7.3$ to -3.6 kcal mol^{-1}). The ΔG^0 values of the H-abstraction of the N–H bond were 3.0, 3.7 and 0.1 kcal mol^{-1} in the gas phase, pentyl ethanoate and water, respectively, while the lowest ΔG^0 for the C–H bonds was observed at the C14–H bond with 4.1, 4.8 and 1.7 kcal mol^{-1} in the gas phase, pentyl ethanoate and water, respectively. The FHT reactions of the other C–H bonds were not spontaneous ($\Delta G^0 = 10.6$–53.1 kcal mol^{-1}). Similarly, neither the proton loss nor SET reactions were spontaneous in any of the investigated solvents. Thus, in nonpolar environments the HOO$^{\bullet}$ radical scavenging activity follows the FHT pathway, however in polar media such as water at pH = 7.40, the deprotonation of **GLD** needs also be addressed.

3.2 The kinetic study

3.2.1. Acid–base equilibrium. The conjugate base form often exhibits much higher activity than the protonated form of acidic species in aqueous environments.[12,17] To identify the most probable radical scavenging activity, the protonation state of **GLD** was initially examined at physiological pH. The **GLD** structure permits protonation at the N–H and O9–H bonds in accordance with reactions (1) and (2); hence, the pK_a values of **GLD** were determined based on the published literature[48] and are depicted in Fig. 2.

$$R_2NH_2^+ \leftrightarrows R_2NH + H^+ \tag{15}$$

$$ROH \leftrightarrows RO^- + H^+ \tag{16}$$
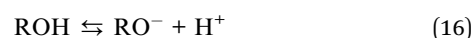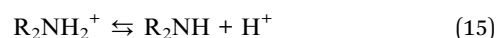
The computed pK_a values for the amine were p$K_{a1} = -2.56$ (for the cation form of the N–H bond) and p$K_{a2} = 10.64$ (for the O9–H bond). In an aqueous solution with a pH of 7.4, **GLD** exists in two states: neutral (HA, 99.9%) and anion (A$^-$, 0.1%). As a consequence of this, both states were considered during the kinetic analysis of the HOO$^{\bullet}$ radical activity of **GLD** in water
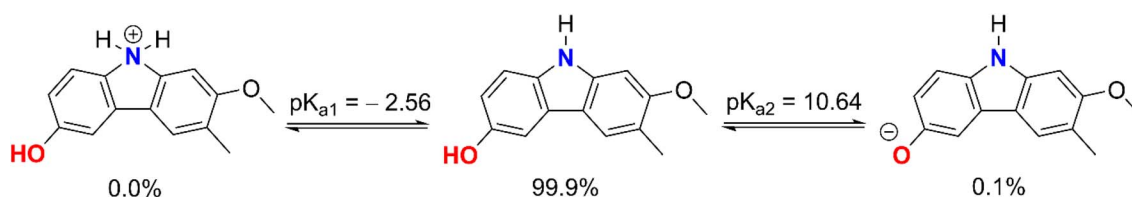
Fig. 2 Possible protonation states of **GLD** at pH = 7.40.

Table 3 Computed $\Delta G^{\neq}$ in kcal mol^{-1}, tunneling correction (k), Γ in %, and k_{Eck}, k_{app}, k_f, and $k_{overall}$ in M^{-1} s^{-1} of **GLD** + HOO$^{\bullet}$ reactions

Solv.	Mechanisms		$\Delta G^{\neq}$	κ	k_{Eck}	k_{app}	$k_f{}^a$	$k_{overall}$	Γ
G	FHT	N–H	15.8	570.5	9.64×10^3			8.53×10^5	1.1
		O9–H	12.0	82.3	8.43×10^5				98.9
P	FHT	N–H	18.8	2915		2.90×10^2		3.53×10^4	0.8
		O9–H	13.8	76.4		3.50×10^4			99.2
W	SET (A$^-$)		1.6	15.3^b		7.90×10^9	7.90×10^6	8.23×10^6	96.0
	FHT	N–H	17.6	4218		3.04×10^3	3.03×10^4		0.0
	(HA)	O9–H	12.8	116.5		3.30×10^5	3.30×10^5		4.0

^{a} $k_f = f.k_{app}$; f(A$^-$) = 0.001, f(HA) = 0.999. ^{b} The nuclear reorganization energy (λ, in kcal mol^{-1}).

at a pH value of 7.4, whereas in nonpolar media (*i.e.* the gas phase and pentyl ethanoate) the neutral state was used to compute the rate constants of the HOO$^{\bullet}$ + **GLD** reaction.

3.2.2. The kinetic study. The thermodynamic calculations (Tables 1 and 2) showed that the HOO$^{\bullet}$ antiradical activity of **GLD** in the nonpolar media is dominated by the hydrogen transfer reaction of the N–H and O9–H bonds ($\Delta G^0 \approx 0$ kcal mol^{-1}). The H-abstraction of the C14–H bond ($\Delta G^0 = 1.7$–4.8 kcal mol^{-1}) should be omitted due to the lower HOO$^{\bullet}$ radical scavenging activity of C–H bonds compared with the N–H and O9–H bonds.[17,51] However, in the aqueous solution, the SET reaction of the anion state should be considered.[12,24,50]

Therefore, the total rate constant ($k_{overall}$) of **GLD** antiradical activity against the HOO$^{\bullet}$ radical in the gas phase, pentyl ethanoate and water can be calculated using eqn (17) and (18). Table 3 and Fig. 3 show the final results.

Nonpolar environments:

$$k_{overall} = k_{app}(\text{FHT(O9–H)-neutral}) + k_{app}(\text{FHT(N–H)-neutral}) \quad (17)$$

Water at physiological pH:

$$k_{overall} = k_f(\text{SET-anion}) + k_f(\text{FHT(O9–H)-neutral}) + k_f(\text{FHT(N–H)-neutral}) \quad (18)$$
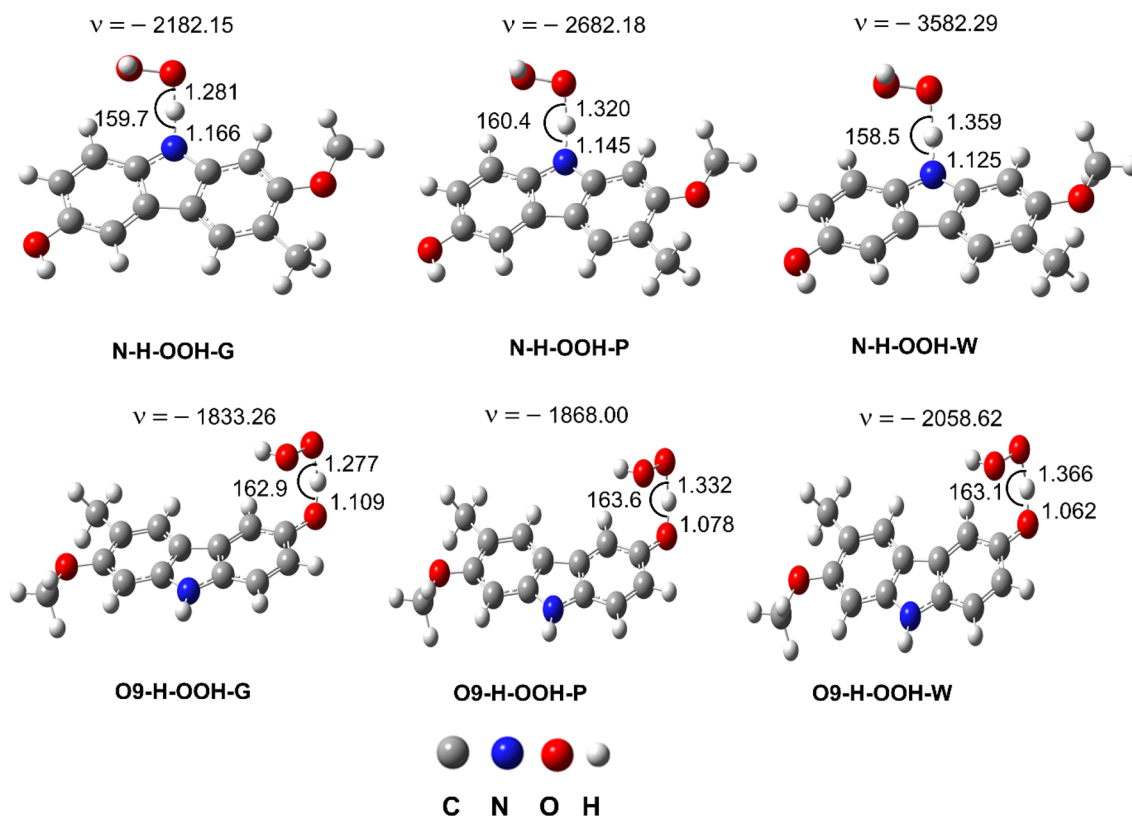
Fig. 3 The TS structures of the HOO$^{\bullet}$ + **GLD** reaction following the FHT pathway; imaginary frequencies given in cm^{-1}, bond lengths in Å and bond angles in °.

Gas-phase kinetic computations revealed that the HOO$^\bullet$ antiradical activity of **GLD** mainly proceeds *via* H-abstraction of the O9–H bond ($k_{(O9-H)} = 8.43 \times 10^5$ M^{-1} s^{-1}, $\Gamma = 98.9\%$). The contribution of FHT reactions of the N–H bond was negligible ($\Gamma = 1.1\%$). This is in agreement with thermodynamic data (Table 1). Similar trend was also observed in the lipid medium, H-abstraction at the O9–H bond contributed 99.2% ($k_{(O9-H)} = 3.50 \times 10^4$ M^{-1} s^{-1}) to the overall rate constant ($k_{overall} = 3.53 \times 10^4$ M^{-1} s^{-1}), whereas, the contribution of the N–H bond was only 0.8% ($k_{(N-H)} = 2.90 \times 10^2$ M^{-1} s^{-1}) in the $k_{overall}$. In aqueous solution, SPLET was the predominant mechanism with $k_f = 7.90 \times 10^6$ M^{-1} s^{-1} ($\Delta G^{\neq} = 1.6$ kcal mol^{-1}), however, the FHT reaction of the O9–H and N–H bond still contributed approximately 4.0% of the $k_{overall}$. Thus in the polar environment the HOO$^\bullet$ antiradical activity of **GLD** is roughly 233 times faster than in the lipid environment ($k_{overall} = 8.23 \times 10^6$ *vs.* 3.53×10^4 M^{-1} s^{-1}, respectively). Nevertheless, in the lipid environment the HOO$^\bullet$ radical scavenging activity of **GLD** is still higher than that of typical antioxidants such as BHT ($k_{overall} = 1.70 \times 10^4$ M^{-1} s^{-1}),[14] trolox ($k_{overall} = 3.40 \times 10^3$ M^{-1} s^{-1}),[19] ascorbic acid ($k_{overall} = 5.71 \times 10^3$ M^{-1} s^{-1}),[20] or resveratrol ($k_{overall} = 1.31 \times 10^4$ M^{-1} s^{-1}).[24] In the polar environment, **GLD** is about 33 and 92 times more active than BHT ($k_{overall} = 2.51 \times 10^5$ M^{-1} s^{-1}),[14] and trolox ($k = 8.96 \times 10^4$ M^{-1} s^{-1}),[19] respectively, but less active than ascorbic acid ($k = 9.97 \times 10^7$ M^{-1} s^{-1}),[20] and resveratrol ($k = 5.62 \times 10^7$ M^{-1} s^{-1}).[24] Thus **GLD** is a promising radical scavenger in physiological environments.

3.3 The antiradical activity of GLD against ordinary free radicals in aqueous solution following the SET reaction

Although HOO$^\bullet$ scavenging activity is a valuable comparative metric, the antiradical activity against different radicals often vary in a broad range. Therefore, the radical scavenging activity of **GLD** was subsequently modeled against a variety of common free radicals, including HOO$^\bullet$, CH$_3$OO$^\bullet$, CCl$_3$OO$^\bullet$, HO$^\bullet$, CH$_3$O$^\bullet$, CCl$_3$O$^\bullet$, NO, NO$_2$, O$_2^{\bullet-}$, SO$_4^{\bullet-}$, N$_3^\bullet$, ABTS$^{+\bullet}$ and DPPH. The hydroperoxyl radical scavenging activity of **GLD** ($\Gamma = 96.0\%$) is determined by the SET mechanism. Consequently, in this investigation, the antiradical activity against these radicals in water at pH = 7.4 was evaluated using the SET mechanism and the findings are shown in Table 4.

GLD should have high activity against CH$_3$O$^\bullet$, CH$_3$OO$^\bullet$, CCl$_3$OO$^\bullet$, NO$_2$, SO$_4^{\bullet-}$, DPPH and ABTS$^{+\bullet}$ radicals with $k_{app} \approx 10^9$ M^{-1} s^{-1} and $k_f \approx 10^6$ M^{-1} s^{-1}, whereas NO, O$_2^{\bullet-}$, HO$^\bullet$, CCl$_3$O$^\bullet$, and N$_3^\bullet$ radicals cannot be eliminated by **GLD** under the studied conditions. By the SET reaction, **GLD** is less active against CCl$_3$O$^\bullet$, and N$_3^\bullet$ radicals than fraxin[55] or usnic acid,[56] but more effective against HOO$^\bullet$ and CH$_3$OO$^\bullet$ radicals. For HO$^\bullet$, however, the prediction of low activity suggests that SET is not the correct mechanism; highly reactive radicals are known to follow alternative pathways (FHT or radical adduct formation to the neutral species) that for these radicals are essentially barrierless in aqueous media. Thus our results also highlight the limits of generalizations from one detailed study.[50]

4. Conclusion

The antiradical activity of **GLD** against HOO$^\bullet$, CH$_3$OO$^\bullet$, CCl$_3$OO$^\bullet$, HO$^\bullet$, CH$_3$O$^\bullet$, CCl$_3$O$^\bullet$, NO, NO$_2$, O$_2^{\bullet-}$, SO$_4^{\bullet-}$, N$_3^\bullet$, ABTS$^{\bullet+}$ and DPPH was studied using DFT calculations. In the physiological environments **GLD** exhibited significant antiradical activity. The overall rate constants for the antiradical trapping of **GLD** in water at pH = 7.40 and pentyl ethanoate were $k_{overall} = 8.23 \times 10^6$ and 3.53×10^4 M^{-1} s^{-1}, respectively. The SPLET mechanism made contributions to the activity in water at pH 7.40, however, the FHT mechanism characterized the activity in nonpolar solvents. Additionally, it was found that **GLD** has strong antiradical activity against CH$_3$O$^\bullet$, CH$_3$OO$^\bullet$, CCl$_3$OO$^\bullet$, NO$_2$, SO$_4^{\bullet-}$, DPPH and ABTS$^{+\bullet}$ $k_{app} \approx 10^9$ M^{-1} s^{-1} and $k_f \approx 10^6$ M^{-1} s^{-1}. According to the computed results, **GLD** is more effective at trapping HOO$^\bullet$ than reference antioxidants like trolox and BHT in the physiological environment. The results suggest that **GLD** can join the long list of phytochemicals with good radical scavenging activity in physiological environments, emphasizing yet again the importance of varied plant sources in diets and in dietary supplements to maintain health and prevent disease.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

Table 4 Calculated kinetic data between GLD–O9-ANION (A$^-$) and the selected radicals ($\Delta G^{\neq}$, λ in kcal mol^{-1}; k_D, k_{app} and k_f in M^{-1} s^{-1})[a]

Radicals	$\Delta G^{\neq}$	λ	k_D	k_{app}	k_f
HO$^\bullet$	42.5	3.4	8.40×10^9	4.30×10^{-19}	4.30×10^{-22}
CH$_3$O$^\bullet$	2.1	4.5	7.90×10^9	7.60×10^9	7.60×10^6
CCl$_3$O$^\bullet$	24.1	21.2	7.50×10^9	1.30×10^{-5}	1.30×10^{-8}
HOO$^\bullet$	1.6	15.3	8.10×10^9	7.90×10^9	7.90×10^6
CH$_3$OO$^\bullet$	2.2	14.7	8.00×10^9	7.60×10^9	7.60×10^6
CCl$_3$OO$^\bullet$	1.4	16.8	7.60×10^9	7.50×10^9	7.50×10^6
NO	71.8	14.3	8.20×10^9	1.50×10^{-40}	1.50×10^{-43}
NO$_2$	0.0	27.7	8.00×10^9	8.00×10^9	8.00×10^6
O$_2^{\bullet-}$	36.8	17.1	8.00×10^9	7.10×10^{-15}	7.10×10^{-18}
SO$_4^{\bullet-}$	14.1	17.6	7.70×10^9	2.90×10^2	2.90×10^{-1}
N$_3^\bullet$	52.3	2.4	7.90×10^9	3.00×10^{-26}	3.00×10^{-29}
DPPH	0.8	18.8	7.40×10^9	7.40×10^9	7.40×10^6
ABTS$^{\bullet+}$	0.0	11.8	7.40×10^9	7.40×10^9	7.40×10^6

[a] $k_f = f.k_{app}$; $f(A^-) = 0.001$.

References

1 A. Bensid, N. El Abed, A. Houicher, J. M. Regenstein and F. Özogul, *Crit. Rev. Food Sci. Nutr.*, 2022, **62**, 2985–3001.

2 C. Nirmala, M. S. Bisht, H. K. Bajwa and O. Santosh, *Trends Food Sci. Technol.*, 2018, **77**, 91–99.

3 İ. Gulcin, *Arch. Toxicol.*, 2020, **94**, 651–715.

4 L. Khandokar, M. S. Bari, V. Seidel and M. A. Haque, *J. Ethnopharmacol.*, 2021, **278**, 114313.

5 P. K. Teja, P. Patel, D. Bhavsar, C. Bindusri, K. Jadhav and S. K. Chauthe, *Phytochemistry*, 2021, **190**, 112865.

6 M. A. Tan, N. Sharma and S. S. A. An, *Antioxidants*, 2022, **11**, 493.

7 R. S. Ramsewak, M. G. Nair, G. M. Strasburg, D. L. DeWitt and J. L. Nitiss, *J. Agric. Food Chem.*, 1999, **47**, 444–447.

8 P. Bhattacharyya, P. Chakrabartty and B. Chowdhury, *Phytochemistry*, 1985, **24**, 882–883.

9 H. Greger, O. Hofer, H. Kählig and G. Wurz, *Tetrahedron*, 1992, **48**, 1209–1218.

10 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, G. Z. J. Bloino, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery Jr, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, *Gaussian 16*, Gaussian, Inc., Wallingford CT, 2016.

11 Y. Zhao and D. G. Truhlar, *Theor. Chem. Acc.*, 2008, **120**, 215–241.

12 A. Galano and J. R. Alvarez-Idaboy, *J. Comput. Chem.*, 2014, **35**, 2019–2026.

13 Y. Zhao and D. G. Truhlar, *J. Phys. Chem. A*, 2008, **112**, 1095–1099.

14 H. Boulebd, *Int. J. Chem. Kinet.*, 2021, 1–8.

15 H. Boulebd, D. M. Pereira, I. A. Khodja, N. T. Hoa, A. Mechler and Q. V. Vo, *J. Mol. Liq.*, 2022, **346**, 118277.

16 M. Carreon-Gonzalez, A. Vivier-Bunge and J. R. Alvarez-Idaboy, *J. Comput. Chem.*, 2019, **40**, 2103–2110.

17 Q. V. Vo, N. T. Hoa, P. C. Nam, D. T. Quang and A. Mechler, *ACS Omega*, 2020, **5**, 24106–24110.

18 J. R. l. Alvarez-Idaboy and A. Galano, *J. Phys. Chem. B*, 2012, **116**, 9316–9325.

19 M. E. Alberto, N. Russo, A. Grand and A. Galano, *Phys. Chem. Chem. Phys.*, 2013, **15**, 4642–4650.

20 A. Galano and J. R. Alvarez-Idaboy, *J. Comput. Chem.*, 2013, **34**, 2430–2445.

21 H. Boulebd, A. Mechler, N. T. Hoa and Q. V. Vo, *New J. Chem.*, 2020, **44**, 9863–9869.

22 Q. V. Vo, N. T. Hoa and A. Mechler, *Polym. Degrad. Stab.*, 2021, **185**, 109483.

23 A. V. Marenich, C. J. Cramer and D. G. Truhlar, *J. Phys. Chem. B*, 2009, **113**, 6378–6396.

24 C. Iuga, J. R. l. Alvarez-Idaboy and N. Russo, *J. Org. Chem.*, 2012, **77**, 3868–3877.

25 M. G. Evans and M. Polanyi, *Trans. Faraday Soc.*, 1935, **31**, 875–894.

26 H. Eyring, *J. Chem. Phys.*, 1935, **3**, 107–115.

27 D. G. Truhlar, W. L. Hase and J. T. Hynes, *J. Phys. Chem. A*, 1983, **87**, 2664–2682.

28 T. Furuncuoglu, I. Ugur, I. Degirmenci and V. Aviyente, *Macromolecules*, 2010, **43**, 1823–1835.

29 E. Vélez, J. Quijano, R. Notario, E. Pabón, J. Murillo, J. Leal, E. Zapata and G. Alarcón, *J. Phys. Org. Chem.*, 2009, **22**, 971–977.

30 E. Dzib, J. L. Cabellos, F. Ortíz-Chi, S. Pan, A. Galano and G. Merino, *Int. J. Quantum Chem.*, 2019, **119**, e25686.

31 E. Dzib, J. L. Cabellos, F. Ortiz-Chi, S. Pan, A. Galano and G. Merino, *Eyringpy 1.0.2*, Cinvestav, 2018, Mérida, Yucatán.

32 E. Pollak and P. Pechukas, *J. Am. Chem. Soc.*, 1978, **100**, 2984–2991.

33 A. Fernández-Ramos, B. A. Ellingson, R. Meana-Pañeda, J. M. Marques and D. G. Truhlar, *Theor. Chem. Acc.*, 2007, **118**, 813–826.

34 C. Eckart, *Phys. Rev.*, 1930, **35**, 1303.

35 R. A. Marcus, *Annu. Rev. Phys. Chem.*, 1964, **15**, 155–196.

36 R. A. Marcus, *Rev. Mod. Phys.*, 1993, **65**, 599.

37 S. F. Nelsen, S. C. Blackstock and Y. Kim, *J. Am. Chem. Soc.*, 1987, **109**, 677–682.

38 S. F. Nelsen, M. N. Weaver, Y. Luo, J. R. Pladziewicz, L. K. Ausman, T. L. Jentzsch and J. J. O'Konek, *J. Phys. Chem. A*, 2006, **110**, 11665–11676.

39 F. C. Collins and G. E. Kimball, *J. Colloid Sci.*, 1949, **4**, 425–437.

40 M. Von Smoluchowski, *Z. Phys. Chem.*, 1917, **92**, 129–168.

41 D. G. Truhlar, *J. Chem. Educ.*, 1985, **62**, 104.

42 A. Einstein, *Ann. Phys.*, 1905, **17**, 549–560.

43 G. G. Stokes, *Mathematical and Physical Papers*, University Press, Cambridge, 1905.

44 Y. Okuno, *Chem.–Eur. J.*, 1997, **3**, 212–218.

45 S. Benson, *The Foundations Of Chemical Kinetics*, Malabar, Florida, 1982.

46 C. Iuga, J. R. Alvarez-Idaboy and A. Vivier-Bunge, *J. Phys. Chem. B*, 2011, **115**, 12234–12246.

47 J. R. Alvarez-Idaboy, L. Reyes and N. Mora-Diez, *Org. Biomol. Chem.*, 2007, **5**, 3682–3689.

48 A. Galano, A. Pérez-González, R. Castañeda-Arriaga, L. Muñoz-Rugeles, G. Mendoza-Sarmiento, A. Romero-Silva, A. Ibarra-Escutia, A. M. Rebollar-Zepeda, J. R. León-Carmona, M. A. Hernández-Olivares and J. R. Alvarez-Idaboy, *J. Chem. Inf. Model.*, 2016, **56**, 1714–1724.

49 A. Galano and J. Raúl Alvarez-Idaboy, *Int. J. Quantum Chem.*, 2019, **119**, e25665.

50 Q. V. Vo, M. V. Bay, P. C. Nam, D. T. Quang, M. Flavel, N. T. Hoa and A. Mechler, *J. Org. Chem.*, 2020, **85**, 15514–15520.

51 K. U. Ingold and D. A. Pratt, *Chem. Rev.*, 2014, **114**, 9022–9046.

52 A. Galano, G. Mazzone, R. Alvarez-Diduk, T. Marino, J. R. Alvarez-Idaboy and N. Russo, *Annu. Rev. Food Sci. Technol.*, 2016, **7**, 335–352.

53 G. Wang, Y. Xue, L. An, Y. Zheng, Y. Dou, L. Zhang and Y. Liu, *Food Chem.*, 2015, **171**, 89–97.

54 L. Estévez, N. Otero and R. A. Mosquera, *J. Phys. Chem. B*, 2010, **114**, 9706–9712.

55 P. C. Nam, N. M. Thong, N. T. Hoa, D. T. Quang, L. P. Hoang, A. Mechler and Q. V. Vo, *RSC Adv.*, 2021, **11**, 14269–14275.

56 N. T. Hoa, M. Van Bay, A. Mechler and Q. V. Vo, *ACS Omega*, 2020, **5**, 17715–17720.