


Cite this: *RSC Adv.*, 2022, 12, 23839

Received 17th July 2022
Accepted 15th August 2022

DOI: 10.1039/d2ra04408k

rsc.li/rsc-advances

Enzymatic synthesis of vitexin glycosides and their activity†

Tingting Xu,^a Ziyun Fan,^b Junqiao Lou,^b Qi Du,^b Yue Kong,^b Yujia Lu^b
and Xueming Wu^{ID}*^b

Vitexin is an active component of many traditional Chinese medicines, and is found in various plants. The low solubility of vitexin limits its pharmaceutical usage. In this study, solvent-stable β -fructosidase was used to glycosylate vitexin in organic solvents. The β -fructosidase showed high activity and stability in 30–80% (v/v) ethyl acetate with 90–99% yields of vitexin glycosides. Highly efficient synthesis of β -D-fructofuranosyl-(2 \rightarrow 6)-vitexin (1.04 g L⁻¹) and β -D-difructofuranosyl-(2 \rightarrow 6)-vitexin (0.45 g L⁻¹) was attained in 50% (v/v) ethyl acetate solvent system from 1.5 g L⁻¹ vitexin. Two novel vitexin glycosides showed higher anti-tumor activities compared to that of vitexin by employing a human breast cancer cytotoxicity assay.

1. Introduction

Vitexin (apigenin-8-C-glucoside) is a bioactive ingredient found in many traditional Chinese medicines. As a C-glycosylated flavone, vitexin was isolated from various plants, such as hawthorn, mung bean, pearl millet, *Passiflora*, pigeon pea, bamboo, wheat leaves, mimosa, chaste tree, *etc.*^{1–5} It was reported to have anti-tumor,^{6–8} anti-oxidant,⁹ anti-viral,¹⁰ and hepatoprotective activities.^{11,12} Thus, vitexin is a potential phytochemical constituent for pharmaceutical usage. However, the poor aqueous solubility of vitexin is a serious issue which limits its pharmaceutical application. Glycosylation has been used to increase the solubility and bioactivity of various natural products.^{13,14} However, the chemical method requires glycosyl activation and complicated reaction steps to control regioselectivity. In contrast, enzymatic glycosylation has attracted extensive interest because of its exquisite selectivity and mild reaction conditions.^{15,16} There are two types of enzymes used for glycosylation: glycosyltransferases and glycosidases.^{17,18} Glycosyltransferases catalyze the formation of glycosidic bonds needed to supply an activated sugar donor, such as uridine-diphosphate glucose (UDP-G), which constrained its practical application. Glycosylation with glycosidases is under thermodynamic control using a non-activated sugar donor, which made these enzymes attractive in the low-cost synthesis of glycosides.¹⁸ Enzymatic synthesis of vitexin glucosides has been

reported by Chang;¹⁹ two glycosidases and four glycosyltransferases were selected to glycosylate vitexin. The glycosyltransferase (BtGT_16345) from *Bacillus thuringiensis* showed glycosylation activity, catalyzing the conversion of vitexin-4'-O- β -glucoside (17.5% yield) and vitexin-5-O- β -glucoside (18.6% yield).¹⁹ However, the glycosidases (amylase and maltogenic amylase) did not catalyze the glycosylation of vitexin.¹⁹ The reported enzymatic glycosylation of vitexin is not efficient with lower yields mainly due to the poor solubility issue of the substrate. The above results indicate that efficient glycosylation of vitexin is still more challenging for glycosyltransferases and glycosidases.

Interestingly, enzymatic glycosylation in organic solvents provides some industrially attractive advantages, such as reversing the thermodynamic equilibrium of glycosidases hydrolysis reactions, improving the aqueous solubility of hydrophobic substrates, eliminating microbial contamination, and lowering industrial cost process.^{20,21} By using this approach, solvent-stable glycosidase (β -fructosidase from *Arthrobacter nicotianae*) showed high activity and stability in hydrophilic organic solvents with more than 90% yield of puerarin glycosides.²⁰ In this study, we demonstrate that the solvent-stable β -fructosidase can be efficiently used for the glycosylation of vitexin, and evaluate the anti-tumor activities of vitexin glycosides.

2. Materials and methods

2.1 Materials and reagents

Vitexin was purchased from Zelang (Jiangsu, China). Methanol used for HPLC analysis was of chromatographic grade from Sinopharm (Shanghai, China). n-butanol, ethyl acetate, hexane, ethanol, methanol, DMSO were of analytical grade and

^aSchool of Medicine & Holistic Integrative Medicine, Nanjing University of Chinese Medicine, Nanjing, 210023, China

^bState Key Laboratory Cultivation Base for TCM Quality and Efficacy, School of Pharmacy, Nanjing University of Chinese Medicine, Nanjing, 210023, China. E-mail: xuemingwu@njucm.edu.cn

† Electronic supplementary information (ESI) available. See <https://doi.org/10.1039/d2ra04408k>



purchased from Beijing Chemical Factory (Beijing, China). Water used for deionized by an osmosis Milli-Q system (Millipore, USA). All other chemicals were of analytical grade and purchased from Sunshine (Nanjing, China). The β -fructosidase was purified from *Arthrobacter nicotianae* XM6 (CCTCC M2010164).²⁰

2.2 Enzymatic glycosylation of vitexin

The glycosylation reaction (1.0 L) was conducted in 1/15 M $\text{Na}_2\text{HPO}_4/\text{KH}_2\text{PO}_4$ buffer (pH 6.47) containing 0.25 U mL^{-1} β -fructosidase, 10% (w/v) sucrose and 1.5 g L^{-1} vitexin and 10–80% (v/v) organic solvent. The reaction mixture was conducted at 35 °C with shaking at 180 rpm for 24 h, then the reaction was stopped at 100 °C for 5 min.

2.3 Kinetic analysis of β -fructosidase

The kinetic parameters were determined in the reaction mixture containing vitexin as substrate and 50 mM sucrose in $\text{Na}_2\text{HPO}_4/\text{KH}_2\text{PO}_4$ buffer (1/15 M, pH 6.47). Substrate concentration was varied in the range 20–200 μM . The purified β -fructosidase (0.25 U mL^{-1}) was then added, and the reaction was incubated at 35 °C for 10 min, and 30 min for organic solvent reaction systems. Then, the reaction was stopped by adding 900 μL of methanol to 100 μL of reaction solution. The reaction products were analyzed by HPLC. One unit (U) of activity was equivalent to 1 μM of vitexin converted by the amount of enzyme per minute at described conditions. Each analysis was carried out in triplicate.

2.4 Analysis, isolation and identification of products

In general, products were analyzed by reversed-phase liquid chromatography on Agilent 1260 HPLC, using a XBridge C18 column (5 μm , 4.6 \times 250 mm). The HPLC analysis method was performed using the following gradient: 0.0–13.0 min, 10–100% B; 13.0–20.0 min, 100% B; 20.0–20.1 min, 100–10% B; 20.1–23.0 min, equilibration with 10% B. The mobile phase consisted of solvents A (water) and B (methanol). The column was operated with a constant flow rate of 0.8 mL min^{-1} at 30 °C and a sample injection volume of 20 μL . The liquid chromatography hyphenated with the high-resolution electrospray ionization mass spectrometry (HR-ESI-MS) was run on an Agilent 6546 Accurate-Mass Q-TOF LC/MS instrument with a Poroshell 120 EC-C18 column (2.7 μm , 4.5 \times 50 mm). The LC-HR/MS analysis of products was performed using the “A + B” mobile phase as specified below: 0.0–13.0 min, 10–100% B. The column was operated with a constant flow rate of 0.3 mL min^{-1} at 30 °C. Glycosylation products were purified by semipreparative HPLC on an Agilent Technologies 1260 Infinity LC System (Agilent, California, USA) by an Agilent Eclipse XDB-C18 column (5 μm , 9.4 \times 250 mm) at a flow rate of 2.0 mL min^{-1} . For structure elucidation, NMR spectra were recorded on a Bruker AV-600 NMR spectrometer (Bruker, Fällanden, Switzerland). Samples were dissolved in $\text{DMSO}-d_6$ at room temperature with tetramethylsilane (Me_4Si) as the chemical shift reference.

2.5 Determination of aqueous solubility

The aqueous solubility of vitexin and vitexin glucosides was determined as follows. Each compound was vortexed in double-deionized H_2O for 30 min at 25 °C. The mixture was centrifuged at 10 000 $\times g$ for 20 min at 25 °C. The supernatant was filtered through a 0.2 μm nylon membrane, mixed with an equal volume of methanol, and analyzed by HPLC. The concentration of these compounds was determined on the basis of the peak areas, using calibration curves prepared by HPLC analyses of authentic samples.

2.6 Cell lines and culture conditions

Human breast cancer cells MCF-7 and MDA-MB-231 were purchased from the Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences (Shanghai, China). Two cells were maintained in 90% DMEM (HyClone, USA) supplemented with 2 mM glutamine, 100 U mL^{-1} penicillin, 100 $\mu\text{g mL}^{-1}$ streptomycin and 10% heat-inactivated fetal bovine serum at 37 °C in a humidified incubator with 5% CO_2 .

2.7 Growth inhibition assay

The cytotoxicity effects of β -D-fructofuranosyl-(2 \rightarrow 6)-vitexin, β -D-difructofuranosyl-(2 \rightarrow 6)-vitexin and vitexin were determined by MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide; solarbio, China) assay.²² Approximately 1×10^4 cells suspended in 100 μL of growth medium were added to each well of a 96 well plates and were incubated. The cells were treated with vitexin, β -D-fructofuranosyl-(2 \rightarrow 6)-vitexin and β -D-difructofuranosyl-(2 \rightarrow 6)-vitexin at final concentrations of 10, 20, 40, 80, 160, 250 μM for 48 h. Cells treated with dimethyl sulfoxide (DMSO) alone (0.01%) were used as control. Each concentration was tested with 6 replicate wells. After 48 h, the medium containing the treatments was removed, and 100 μL of MTT solution (0.5 mg mL^{-1}) was added to each well and incubated at 37 °C for 3 h. The formazan crystals were dissolved in 100 μL of DMSO, and the absorbance was measured at 490 nm with ELISA plate reader. The experiment was performed in triplicate.

3. Results and discussion

3.1 The structural identification

The chemical structure of the peak fractions was further identified by LC-HR/MS, ^1H NMR, ^{13}C NMR, and 2D NMR spectra. The HR-ESI mass spectrum of compound 1 showed an ion peak $[\text{M}-\text{H}]^-$ at m/z 593.1518 (calculated for $\text{C}_{27}\text{H}_{30}\text{O}_{15}$, 594.1585) (Fig. 1). Compared with vitexin, the compound 1 has an additional formula $\text{C}_6\text{H}_{10}\text{O}_5$, suggesting that 1 may be a vitexin monosaccharide derivative. The ^1H NMR spectra in low field showed six ethylenic signals [δ 6.77(1H, s), 6.27 (1H, s), 7.95 (2H, d, 8.8 Hz), 6.91 (2H, d, 8.8 Hz)] (Table 1). These were the characteristic vitexin signals. The high field peak δ 4.70 (d, 9.9 Hz) was the core vitexin glucose terminal hydrogen signal. The δ 73.9 ppm methylene may be β -sugar terminal carbon, which was connected with C–C bond. The δ 104.8 ppm quaternary carbon must be the other sugar terminal carbon. Data from HSQC spectra can classify all carbon signals except quaternary



carbon. The connections of each self-spinning segment can be inferred by HMBC spectra. From the ^{13}C NMR, ^1H NMR and 2D NMR spectra, we speculated that the monosaccharide was β -D-fructofuranose. A comparison analysis between the NMR data of compound 1 and vitexin revealed that the NMR data of two compounds were almost identical except for the signals due to a fructose moiety. The other diagnostic difference between 1 and vitexin was the carbon signal at C-6'' (δ_{C} 62.8) of the glucose moiety, which was about 1.1 ppm downfield from those of vitexin (δ_{C} 61.7) due to the known effects of *O*-glycosylation. Furthermore, the carbon signal of C-5'' (δ_{C} 75.8) was about 6.5 ppm upfield from those of vitexin (δ_{C} 82.3). The HMBC spectrum (Fig. S7†) showed long-range correlations between C-2''' (δ_{C} 104.8) and H-6'' of glucose moiety and H-1''' of fructose moiety, thus proving the correlation between C-2''' and H-6'' of compound 1. Therefore, the chemical structure of 1 was elucidated as β -D-fructofuranosyl-(2 \rightarrow 6)-vitexin.

The HR-ESI mass spectrum of compound 2 showed an ion peak $[\text{M}-\text{H}]^-$ at m/z 755.2054 (calculated for $\text{C}_{33}\text{H}_{40}\text{O}_{20}$, 756.2113) (Fig. 1). Compared with vitexin, the compound 1 has an additional formula $\text{C}_{12}\text{H}_{20}\text{O}_{10}$, suggesting that 1 may be a vitexin disaccharide derivative. The ^1H NMR spectra in low field showed six ethylenic signals [δ 6.77(1H, s), 6.28 (1H, s), 7.95 (2H, d, 8.5 Hz), 6.91 (2H, d, 8.0 Hz)] (Table 1). These were the characteristic vitexin signals. The high field peak δ 4.71 (d, 9.9 Hz) was the core vitexin glucose terminal hydrogen signal. The δ 73.9 ppm methylene should be β -sugar terminal carbon, which was connected with C-C bond. The δ 104.7 ppm and δ 104.6 ppm quaternary carbon should be the fructose terminal carbon. The difference between 2 and vitexin was the carbon signal at C-6'' (δ_{C} 63.0) of the glucose moiety, which was about

1.3 ppm downfield from those of vitexin (δ_{C} 61.7) due to the known effects of *O*-glycosylation. And other diagnostic difference between 2 and 1 were carbon signal at C-6''' (δ_{C} 62.9) and C-5''' (δ_{C} 76.2) of the fructose, which were about 0.9 ppm downfield and 6.3 ppm upfield from those of compound 1 (δ_{C} 62.0 and δ_{C} 82.5), respectively. The HMBC spectrum also showed long-range correlations between C-2''' (δ_{C} 104.6) and H-6''' of fructose moiety and H-1''' of fructose moiety (Fig. S14†), thus proving the correlation between C-2''' and H-6''' of compound 2. Therefore, the chemical structure of 2 was elucidated as β -D-difructofuranosyl-(2 \rightarrow 6)-vitexin. The spectroscopic data for ^1H and ^{13}C -NMR (600 MHz, $\text{DMSO}-d_6$) were summarized in Table 1.

3.2 Efficient glycosylation in hydrophobic organic solvents

The β -fructosidase from *Arthrobacter nicotianae* has been shown excellent synthesis of puerarin glycosides in hydrophilic organic solvent.²⁰ However, the solubility of vitexin is not obvious increased in hydrophilic organic solvent. In order to improve the solubility of vitexin, the enzymatic synthesis was investigated in hydrophobic organic solvents. The low yield (40%) of glycosylation was observed in aqueous solution because of the poor solubility of vitexin (Fig. 2A). Supplement with several hydrophobic organic solvents (50%, v/v) led to a significant improvement in the glycosylation of vitexin. The higher yield (99%) of vitexin glycosides was obtained in 50% ethyl acetate solvent system compared with hexane and *n*-butanol solvent systems (Fig. 2A). The effect of ethyl acetate proportion on the vitexin glycosylation was studied with the increase of the concentration from 0% to 80%. The glycosylation reaction system with addition of 10%, 30% and 50% of ethyl acetate led

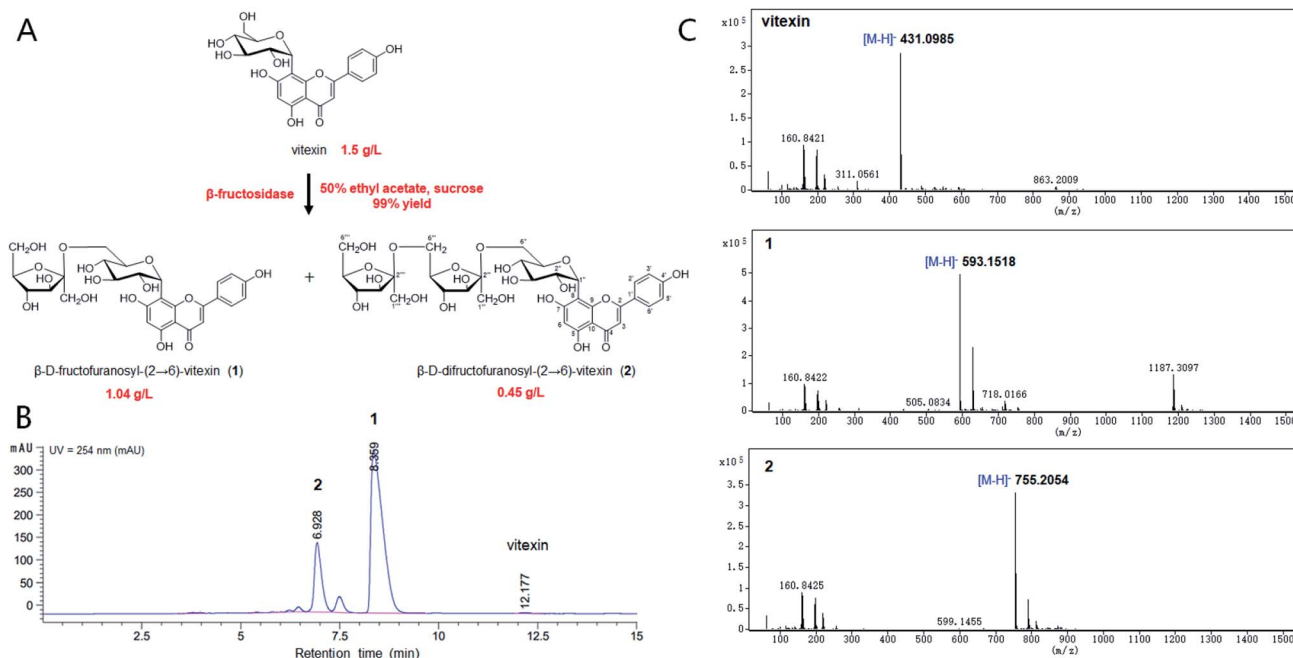


Fig. 1 Bioconversion of vitexin glycosides by β -fructosidase. (A) The chemical structures of vitexin and vitexin glycosides. (B) HPLC profiling of biotransformation for vitexin glycosides. (C) The LC-HR/MS analysis of the reaction products.



Table 1 NMR (600 MHz) spectral data of vitexin and glucosides in DMSO- d_6

| Position | Vitexin | | β -D-Fructofuranosyl-(2 \rightarrow 6)-vitexin | | β -D-Difructofuranosyl-(2 \rightarrow 6)-vitexin | |
|----------|------------|------------------------|--------------------------------------------------------|------------------------|----------------------------------------------------------|------------------------|
| | δ_C | δ_H | δ_C | δ_H | δ_C | δ_H |
| 2 | 164.4 | | 164.3 | | 164.4 | |
| 3 | 102.9 | 6.79 (s) | 103.0 | 6.77 (s) | 102.9 | 6.77 (s) |
| 4 | 182.5 | | 182.6 | | 182.6 | |
| 5 | 160.8 | | 161.0 | | 161.0 | |
| 6 | 98.6 | 6.28 (s) | 98.7 | 6.27 (s) | 98.7 | 6.28 (s) |
| 7 | 163.1 | | 163.3 | | 163.2 | |
| 8 | 105.1 | | 104.8 | | 104.6 | |
| 9 | 156.4 | | 156.5 | | 156.5 | |
| 10 | 104.5 | | 104.5 | | 104.5 | |
| 1' | 122.1 | | 122.1 | | 122.1 | |
| 2' | 129.4 | 8.03 (d, J = 8.7 Hz) | 129.1 | 7.95 (d, J = 8.8 Hz) | 129.2 | 7.95 (d, J = 8.5 Hz) |
| 3' | 116.3 | 6.90 (d, J = 8.7 Hz) | 116.3 | 6.91 (d, J = 8.8 Hz) | 116.3 | 6.91 (d, J = 8.0 Hz) |
| 4' | 161.6 | | 161.6 | | 161.6 | |
| 5' | 116.3 | 6.90 (d, J = 8.7 Hz) | 116.3 | 6.91 (d, J = 8.8 Hz) | 116.3 | 6.91 (d, J = 8.0 Hz) |
| 6' | 129.1 | 8.03 (d, J = 8.7 Hz) | 129.1 | 7.95 (d, J = 8.8 Hz) | 129.2 | 7.95 (d, J = 8.5 Hz) |
| 1'' | 73.8 | 4.69 (d, J = 9.9 Hz) | 73.9 | 4.70 (d, J = 9.9 Hz) | 73.9 | 4.71 (d, J = 9.9 Hz) |
| 2'' | 71.3 | 3.84 (t) | 71.7 | | 71.4 | |
| 3'' | 79.1 | 3.26 (m) | 79.0 | | 78.9 | |
| 4'' | 71.0 | 3.39 (m) | 71.2 | | 71.2 | |
| 5'' | 82.3 | 3.77 (m) | 75.8 | | 75.6 | |
| 6'' | 61.7 | 3.53 (m) | 62.8 | | 63.0 | |
| 1''' | | | 63.0 | | 63.1 | |
| 2''' | | | 104.8 | | 104.7 | |
| 3''' | | | 80.4 | | 80.7 | |
| 4''' | | | 77.4 | | 77.0 | |
| 5''' | | | 82.5 | | 76.2 | |
| 6''' | | | 62.0 | | 62.9 | |
| 1''' | | | | | 61.4 | |
| 2''' | | | | | 104.6 | |
| 3''' | | | | | 80.3 | |
| 4''' | | | | | 76.9 | |
| 5''' | | | | | 82.5 | |
| 6''' | | | | | 61.9 | |
| 13-OH | | 13.18 (s) | | 13.17 (s) | | 13.17 (s) |

to 1.8, 2.3 and 2.4 times yield than in aqueous solution (Fig. 2B). Then the inhibition effect on glycosylation was appeared when the concentration of ethyl acetate up to 80%. Furthermore, the optimization of vitexin concentration was conducted shown in Fig. 2C. The efficient glycosylation was achieved in 1.5 g L^{-1}

substrate concentration with 99% yield of vitexin glycosides. The β -D-fructofuranosyl-(2 \rightarrow 6)-vitexin (1.04 g L^{-1}) and β -D-difructofuranosyl-(2 \rightarrow 6)-vitexin (0.45 g L^{-1}) was attained in 50% (v/v) ethyl acetate solvent after 48 h of bioconversion.

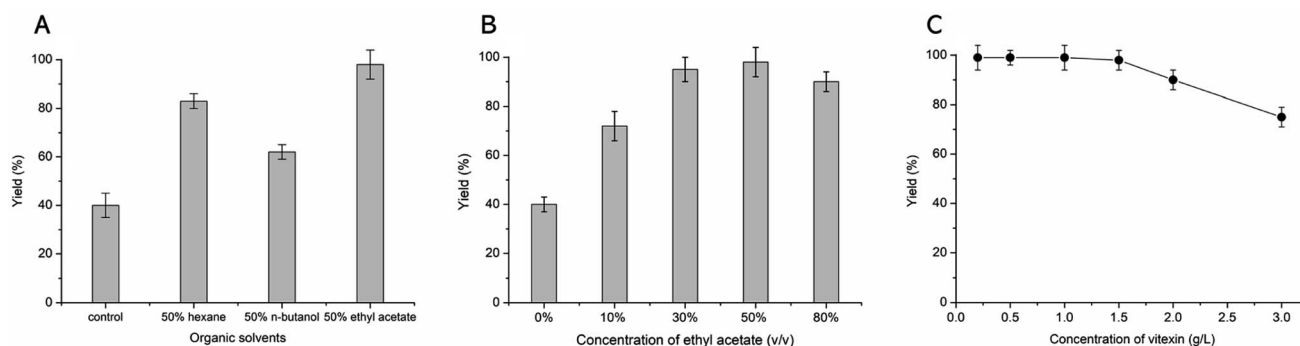


Fig. 2 Optimized bioconversion conditions for the glycosylation of vitexin. The effect of organic solvents (A), concentration of ethyl acetate (B) and concentration of substrate (C) on the vitexin glycosides yield.



Table 2 Kinetic parameters of β -fructosidase in different media

| Solvent | V_{\max}^a [$\mu\text{M s}^{-1}$] | K_m [μM] | K_{cat}^b [s^{-1}] | K_{cat}/K_m [$\text{s}^{-1} \text{mM}^{-1}$] |
|-------------------|---------------------------------------|-------------------------|----------------------------------------|---------------------------------------------------------|
| Water | 5.6 | 115.8 | 64.3 | 555.3 |
| 50% hexane | 4.9 | 112.7 | 58.1 | 515.5 |
| 50% n-butanol | 3.8 | 82.6 | 41.4 | 501.2 |
| 50% ethyl acetate | 5.5 | 114.3 | 62.8 | 549.4 |

^a Sucrose fixed at 50 mM, vitexin varied from 20 μM to 200 μM . ^b K_{cat} values were calculated using a molecular mass of 55 KDa for the β -fructosidase.

Table 3 Aqueous solubility of vitexin and vitexin glycosides

| Compound | Aqueous solubility ^a (mg L^{-1}) |
|----------------------------------------------------------|--------------------------------------------------------|
| Vitexin | 28.2 ± 2.5 |
| β -D-Fructofuranosyl-(2 \rightarrow 6)-vitexin | 2961.0 ± 98.5 |
| β -D-Difructofuranosyl-(2 \rightarrow 6)-vitexin | 8911.2 ± 637.4 |

^a Deionized H_2O at pH 7.0.

Table 4 Cytotoxicity of vitexin and vitexin glycosides

| Compound | IC_{50} (Mean \pm SD, $\mu\text{g mL}^{-1}$) | |
|----------------------------------------------------------|----------------------------------------------------------|----------------|
| | MCF-7 | MDA-MB-231 |
| Vitexin | 98.5 ± 6.2 | 65.8 ± 3.4 |
| β -D-Fructofuranosyl-(2 \rightarrow 6)-vitexin | 81.9 ± 5.6 | 52.4 ± 2.5 |
| β -D-Difructofuranosyl-(2 \rightarrow 6)-vitexin | 81.3 ± 6.7 | 52.1 ± 3.8 |

3.3 Kinetic analysis of β -fructosidase in organic solvent systems

The kinetic parameters were determined using the purified β -fructosidase in various systems are listed in Table 2. Compared to the buffer media, V_{\max} of the β -fructosidase to vitexin slight slowed from $5.6 \mu\text{M s}^{-1}$ to 4.9, 3.8 and $5.5 \mu\text{M s}^{-1}$ in 50% (v/v) of hexane, n-butanol and ethyl acetate solvent, respectively. This observation suggests that β -fructosidase possesses great solvent tolerance, and especially shows excellent activity in 50% (v/v) of ethyl acetate. The K_m values in organic solvents (Table 2) suggest that β -fructosidase showed almost equal affinity for vitexin in water and 50% (v/v) of ethyl acetate solvent. The catalytic turnover constant (K_{cat} , 64.3 s^{-1} and 62.8 s^{-1}) and the catalytic efficiency (K_{cat}/K_m , $555.3 \text{ s}^{-1} \text{mM}^{-1}$ and $549.4 \text{ s}^{-1} \text{mM}^{-1}$) were found to be nearly the same in water and 50% (v/v) of ethyl acetate. The excellent values of K_{cat} and K_{cat}/K_m in 50% (v/v) of ethyl acetate suggest that the β -fructosidase may be beneficial for improving the yield of vitexin glycosides in the solvent system.

3.4 Solubility of vitexin glucosides

The aqueous solubility of vitexin glucosides were obviously improved through glycosylation (Table 3). The solubility of β -D-fructofuranosyl-(2 \rightarrow 6)-vitexin and β -D-difructofuranosyl-(2 \rightarrow 6)-vitexin was over 100 times and 300 times than vitexin, respectively. The increase aqueous solubility of vitexin glucosides could be better improved its clinical application.

3.5 Cytotoxicity assay

MTT assay was performed to compare the cell cytotoxicity induced between vitexin and its glycosides. As shown in Table 4, vitexin was showed effective cytotoxicity on MCF-7 cells (IC_{50} $98.5 \mu\text{g mL}^{-1}$) and MDA-MB-231 cells (IC_{50} $65.8 \mu\text{g mL}^{-1}$) after 48 h of incubation. The cytotoxicity of vitexin glycosides were better than that of vitexin, β -D-fructofuranosyl-(2 \rightarrow 6)-vitexin presented IC_{50} values of 81.9 and $52.4 \mu\text{g mL}^{-1}$ for MCF-7 and MDA-MB-231 cell lines, and β -D-difructofuranosyl-(2 \rightarrow 6)-vitexin presented IC_{50} values of 81.3 and $52.1 \mu\text{g mL}^{-1}$, respectively. However, there no obvious difference of antitumor activity between β -D-fructofuranosyl-(2 \rightarrow 6)-vitexin and β -D-difructofuranosyl-(2 \rightarrow 6)-vitexin. This suggests that glycosylation of vitexin can enhances the antitumor activity.

4. Conclusions

The aim of this study was to investigate the applicability of hydrophobic organic solvents for enzymatic glycosylation of vitexin. The yield of vitexin glycosides was significantly improved by using the 50% (v/v) ethyl acetate solvent system compared to the aqueous solution. Moreover, the β -fructosidase showed high stability and activity in 50% (v/v) ethyl acetate solvent system with 99% yield of glycoside products. Two vitexin glycosides exhibited excellent aqueous solubility and better anti-tumor activity.

Author contributions

X.M. Wu conceived and supervised the project. T.T. Xu designed and performed experiments, and wrote the manuscript. All authors contributed to the data analysis and manuscript preparation.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The work was financed by National Natural Science Foundation of China (82174089 and 81991524).



References

- 1 K. Masike, A. de Villiers, D. de Beer, E. Joubert and M. A. Stander, *J. Food Compos. Anal.*, 2022, **106**, 104308.
- 2 T. T. Phan, L. T. M. Do, T. M. Lien, T. V. Phung, T. T. H. Nguyen, V. N. Huynh, D. T. T. Ngo, K. P. P. Nguyen and T. T. A. Nguyen, *Nat. Prod. Res.*, 2022, **36**, 171–176.
- 3 M. He, J. W. Min, W. L. Kong, X. H. He, J. X. Li and B. W. Peng, *Fitoterapia*, 2016, **115**, 74–85.
- 4 Y. J. Fu, Y. G. Zu, W. Liu, L. Zhang, M. H. Tong, T. Efferth, Y. Kong, C. L. Hou and L. Y. Chen, *J. Sep. Sci.*, 2008, **31**, 268–275.
- 5 E. Gaitan, R. H. Lindsay, R. D. Reichert, S. H. Ingbar, R. C. Cooksey, J. Legan, E. F. Meydrech, J. Hill and K. Kubota, *J. Clin. Endocrinol. Metab.*, 1989, **68**, 707–714.
- 6 K. K. Khoja, R. M. J. Howes, R. Hider, P. A. Sharp, I. W. Farrell and G. O. Latunde-Dada, *Nutrients*, 2022, **14**, 784.
- 7 J. Huang, Y. Zhou, X. Zhong, F. Su and L. Xu, *Oxid. Med. Cell. Longevity*, 2022, 3129155.
- 8 P. Q. D. Nguyen, H. T. Nguyen, L. T. K. Nguyen, H. Q. Vo, A. T. Le, T. T. Do and D. V. Ho, *Trop. J. Nat. Prod. Res.*, 2020, **4**, 703–707.
- 9 N. Yan, H. Zhang, Z. Zhang, H. Zhang, L. Zhou, T. Chen, S. Feng, C. Ding and M. Yuan, *Arabian J. Chem.*, 2022, **15**, 103707.
- 10 J. Tang, B. Dong, M. Liu, S. Liu, Xi. Niu, C. Gaughan, A. Asthana, H. Zhou, Z. Xu, G. Zhang, R. H. Silverman and H. Huang, *J. Med. Chem.*, 2022, **65**, 1445–1457.
- 11 K. K. Noor, M. U. Ijaz, N. Ehsan, A. Tahir, D. K. Yeni, S. M. N. K. Zihad, S. J. Uddin, A. Ashraf and J. Simal-Gandara, *Biomed. Pharmacother.*, 2022, **150**, 112934.
- 12 A. Farooq, A. Iqbal, N. F. Rana, M. Fatima, T. Maryam, F. Batool, Z. Rehman, F. Menaa, S. Azhar, A. Nawaz, F. Amin, Z. M. Mohammedsaleh and S. S. Alrdahe, *Int. J. Mol. Sci.*, 2022, **23**, 3131.
- 13 G. Huang, M. Lv, J. Hu, K. Huang and H. Xu, *Mini-Rev. Med. Chem.*, 2016, **16**, 1013–1016.
- 14 J. Zhao, J. Yang and Y. Xie, *Int. J. Pharm.*, 2019, **570**, 118642.
- 15 D. Piedrabuena, A. Rumbero, E. Pires, A. Leal-Duaso, C. Civera, M. Fernandez-Lobato and M. Hernaiz, *RSC Adv.*, 2021, **11**, 24312–24319.
- 16 P. Nekvasilová, M. Hovorková, Z. Mészáros, L. Petrášková, H. Pelantová, V. Křen, K. Slámová and P. Bojarová, *Int. J. Mol. Sci.*, 2022, **23**, 4106.
- 17 L. Mestrom, M. Przypis, D. Kowalczykiewicz, A. Pollender, A. Kumpf, S. R. Marsden, I. Bento, A. B. Jarzebski, K. Szymanska, A. Chrusciel, D. Tischler, R. Schoevaart, U. Hanefeld and P. L. Hagedoorn, *Int. J. Mol. Sci.*, 2019, **20**, 5263.
- 18 C. Moulis, D. Guieysse, S. Morel, E. Séverac and M. Remaud-Siméon, *Curr. Opin. Chem. Biol.*, 2021, **61**, 96–106.
- 19 J. Y. Wu, T. Y. Wang, H. Y. Ding, Y. R. Zhang and T. T. Chang, *Molecules*, 2021, **26**, 6274.
- 20 X. M. Wu, J. L. Chu, B. Wu, S. Zhang and B. F. He, *Bioresour. Technol.*, 2013, **129**, 659–662.
- 21 T. T. Xu, C. Wang, S. J. Jiang, T. T. Yang and X. M. Wu, *RSC Adv.*, 2022, **12**, 18232–18237.
- 22 L. Zhang, X. La, J. Tian, H. Li, A. Li, Y. Liu, C. Wu and Z. Li, *J. Funct. Foods*, 2021, **85**, 104620.

