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## A “turn-on” ESIPT fluorescence probe of 2-(aminocarbonyl)phenylboronic acid for the selective detection of Cu(II)†

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Herein, we report a highly selective fluorescent probe for the detection of Cu(II). The detection mechanism relies on the Cu(II)-catalyzed oxidative hydroxylation of 2-(aminocarbonyl)phenylboronic acid into salicylamide, thus recovering the excited-state intramolecular proton transfer (ESIPT) effect and inducing more than 35-fold fluorescence enhancement. The simple structure and readily available fluorescent probe give a novel method for quantitatively detecting Cu(II) in the linear range of 0–22 µM, with a limit of detection down to 68 nM, and exhibiting high selectivity for Cu(II) over 16 other metal ions.

### 1. Introduction

As one of the critical heavy metal elements, the level of copper ion content significantly impacts the ecological environment and human health. Especially after biological enrichment, excessive Cu(II) causes metabolic and neurological disorders, leading to rheumatoid arthritis, Wilson's syndrome, Parkinson's disease, and Alzheimer's disease.<sup>1–3</sup> Therefore, it is of great significance to develop an efficient detection assay for Cu(II) in clinical diagnosis and disease prevention.

In recent years, powerful assays for Cu(II) detection have been developed, including titrimetry,<sup>4</sup> UV-Vis spectrophotometry,<sup>5</sup> atomic absorption spectrophotometry (AAS),<sup>6</sup> inductively coupled plasma-atomic emission spectroscopy (ICP-AES),<sup>7</sup> ion selective electrode (ISE),<sup>8</sup> and chemiluminescence.<sup>9</sup> Among various analysis methods, the fluorescent probe is an appealing tool for Cu(II) detection owing to its simple operation, high sensitivity and *in situ* detection in living cells.<sup>10–12</sup>

Nowadays, several fluorescent probes for Cu(II) detection based on the sensing strategy of coordination or chemical reaction have been designed. However, based on the paramagnetic characteristics of Cu(II), the complex generated from the probe and Cu(II) ion usually leads to fluorescent quenching, which gives false-positive results due to the lower sensitivity and anti-interference property.<sup>13</sup> Alternatively, reaction-based fluorescence probes exhibit higher sensitivity and selectivity owing

to the fluorescent signal change related to the unique chemical reaction between analyte and probe molecular.<sup>14–16</sup> Gratifying examples of fluorescence enhancement probes for Cu(II) have been reported based on distinct reactions, such as the Cu(II)-induced hydrolysis,<sup>17,18</sup> ring-opening of the spirolactam,<sup>19,20</sup> C–O bond cleavage,<sup>21–23</sup> Cu(II)-mediated oxidation reaction of *o*-phenylenediamine,<sup>24–26</sup> Cu(II)-catalyzed Heck coupling and click reaction.<sup>27–29</sup> Among these probes, the boronic ester/acid-based fluorescence probe possesses excellent biocompatibility, which facilitates the *in situ* detection of intracellular Cu(II), and has been favored in recent years.<sup>30</sup> However, the borate groups mostly bind with the fluorophore, such as coumarin,<sup>31</sup> fluorescein,<sup>32</sup> resorufin,<sup>33</sup> benzotriazole<sup>34</sup> by chemical modification, leading to challenges in synthetic procedure and high cost. Therefore, it is urgent to develop a new fluorescence probe of Cu(II) using arylboronic acid with a simple structure and low cost.

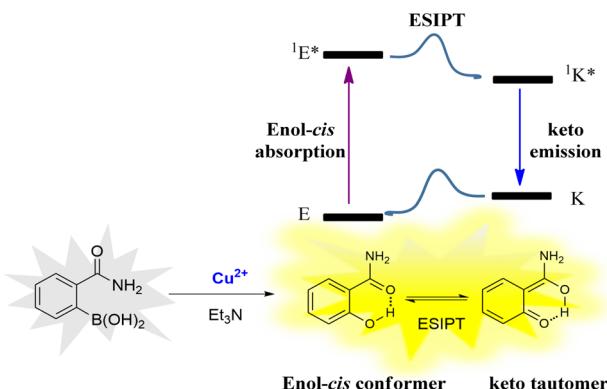
Salicylamide, the simplest derivative of salicylic acid, has long been used as an anti-bacterial and anti-viral drug.<sup>35,36</sup> Notably, salicylamide has attracted considerable attention because of its excited-state intramolecular proton transfer (ESIPT) fluorescent property, which was first reported by Weller in the 1950s for salicylic acid.<sup>37</sup> In general, molecules can exhibit ESIPT fluorescence if their structures incorporate an intramolecular hydrogen bonding interaction between a hydrogen bond donor (–OH and NH<sub>2</sub>) and a hydrogen bond acceptor (=N– and C=O), and the molecular structure of salicylamide fits this feature.<sup>38,39</sup> Therefore, we envision that 2-(aminocarbonyl)phenylboronic acid could be utilized as a “turn-on” fluorescent probe for the detection of Cu(II), in which the boronic acid group acts as the recognition group of Cu(II) and hinders the ESIPT effect of the probe molecule. By employing the Cu(II) catalytic oxidative hydroxylation of the boronic acid group to the hydroxyl group, the ESIPT effect can be recovered

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Scheme 1 Detection mechanism of the probe and the ESIPT process of salicylamide.

owing to keto–enol tautomerism. The ESIPT process starts with the absorption of radiation from an enol *cis* conformer ( $^1\text{E}$ ) and into the excited state ( $^1\text{E}^*$ ), this conformer undergoes ESIPT to form a keto tautomer ( $^1\text{K}^*$ ) giving rise to an emission, leading to the fluorescent “turn on” (Scheme 1). Using widely distributed and cheap 2-(Aminocarbonyl)phenylboronic acid as a fluorescence probe, we provide a concise and efficient assay for quantitatively determining Cu(II).

## 2. Experimental section

### 2.1. Materials and methods

2-(Aminocarbonyl)phenylboronic acid, Et<sub>3</sub>N, Cu(OAc)<sub>2</sub>·H<sub>2</sub>O, EtOH, ACN, DMSO, DMF, KCl, NaCl, CaCl<sub>2</sub>, MgCl<sub>2</sub>·6H<sub>2</sub>O, Sr(NO<sub>3</sub>)<sub>2</sub>, AgNO<sub>3</sub>, ZnCl<sub>2</sub>, MnCl<sub>2</sub>·4H<sub>2</sub>O, CoCl<sub>2</sub>·6H<sub>2</sub>O, Pd(OAc)<sub>2</sub>, HgCl<sub>2</sub>, FeCl<sub>3</sub>, CrCl<sub>3</sub>, AlCl<sub>3</sub>, Pr(NO<sub>3</sub>)<sub>3</sub>·6H<sub>2</sub>O, and ceric ammonium nitrate were purchased from Shanghai Aladdin Bio-Chem Technology Co, Ltd. and used as received. The testing solvents were prepared with deionized water.

### 2.2. General instrumentation

Emission spectra were performed on a fluorescence spectrometer (F-4600). Error limits were estimated:  $\lambda$  ( $\pm 1$  nm),  $\tau$  ( $\pm 10\%$ ), and  $\phi$  ( $\pm 10\%$ ).  $^1\text{H}$  NMR (400 MHz) and  $^{13}\text{C}$  NMR (100 MHz) spectra were recorded on a Bruker Avance AV400 spectrometer (Bruker, Billerica, MA, USA) unless otherwise noted. The chemical shifts ( $\delta$ ) were quoted in parts per million from tetramethylsilane for  $^1\text{H}$  and DMSO-*d*<sub>6</sub> for  $^{13}\text{C}$  spectroscopy. The pH values of the real water samples were recorded by applying B6337-02 benchtop pH/conductivity meters.

### 2.3. Fluorescence detection of Cu(II)

The experimental details of the fluorescent detection condition optimization are provided in the ESI.† As for sensitivity testing, 2-(aminocarbonyl)phenylboronic acid (100  $\mu\text{M}$ ), Et<sub>3</sub>N (150  $\mu\text{M}$ ) and the various concentrations of Cu(II) (0–96  $\mu\text{M}$ ) were added in a cuvette sequentially. After the above-mixed solutions reacted for 2.5 h, the fluorescence spectra were measured and recorded using a fluorescence spectrophotometer.

The limit of detection (LOD) is calculated using the following formula:

$$\text{LOD} = 3\sigma/k,$$

where  $\sigma$  is the standard deviation of the blank signals ( $n = 3$ ).

### 2.4. Selectivity and anti-interference experiments

Various metal ion interferents (2 mM K<sup>+</sup> and Na<sup>+</sup>, 1 mM Ca<sup>2+</sup> and Mg<sup>2+</sup>, 50  $\mu\text{M}$  Sr<sup>2+</sup>, Ag<sup>+</sup>, Zn<sup>2+</sup>, Mn<sup>2+</sup>, Co<sup>2+</sup>, Pd<sup>2+</sup>, Hg<sup>2+</sup>, Fe<sup>3+</sup>, Cr<sup>3+</sup>, Al<sup>3+</sup>, Pr<sup>3+</sup>, Ce<sup>4+</sup>), instead of Cu<sup>2+</sup>, were added to the fluorescent probe solution, and then 50  $\mu\text{M}$  Cu<sup>2+</sup> was mixed with this solution to explore the anti-interference capacity of the probe.

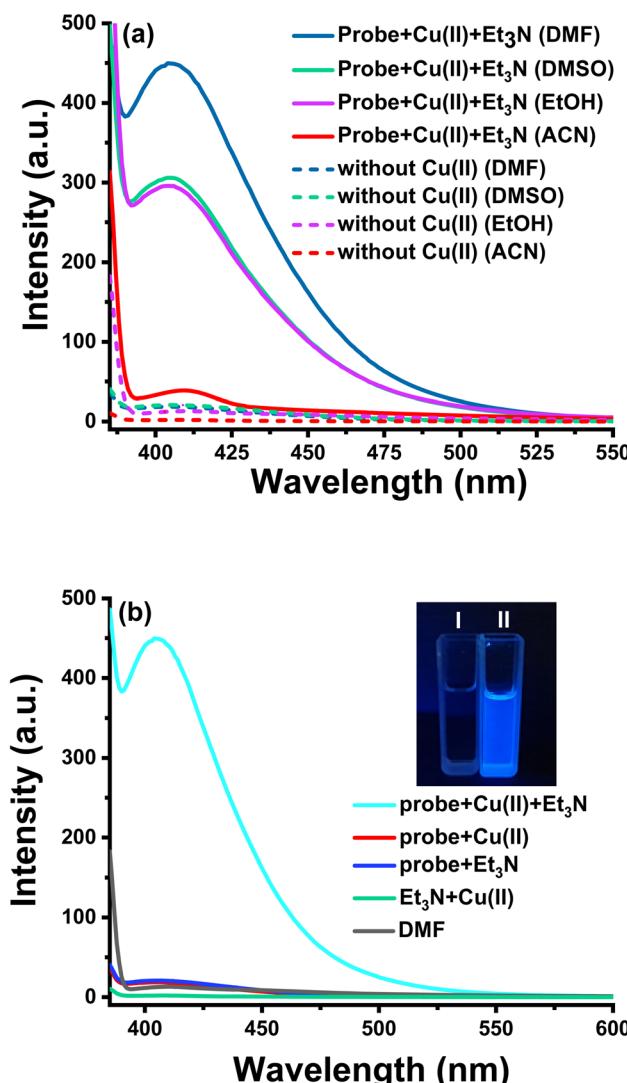


Fig. 1 (a) Fluorescence emission spectra ( $\lambda_{\text{ex}} = 365$  nm) of the sensing system containing 100  $\mu\text{M}$  2-(aminocarbonyl)phenylboronic acid, 50  $\mu\text{M}$  Et<sub>3</sub>N, with or without 50  $\mu\text{M}$  Cu(II) in indicated solvents and (b) the emission spectra of the control experiments carried out in DMF under various conditions. Inset: photographs of 1 mM of 2-(aminocarbonyl)phenylboronic acid without Cu<sup>2+</sup> (I) and with 0.5 mM of Cu<sup>2+</sup> (II) under 365 nm UV illumination.



## 2.5. Fluorescence detection of Cu(II) in real water samples

Ganjiang River water was collected from the Ganjiang River, tap water was collected from a chemistry laboratory, and rainwater was collected from Ganzhou. Cu(II) was spiked into various water samples to obtain sample stock solutions with 5 mM of Cu(II). Real water samples were added to a sensing system containing 2-(aminocarbonyl)phenylboronic acid (100  $\mu$ M) and Et<sub>3</sub>N (150  $\mu$ M), resulting in a final solution with 20  $\mu$ M of Cu(II) in the DMF solution. Then, their fluorescent spectra were measured and recorded by applying a fluorescence spectrometer. All average values were obtained from three parallel experiments ( $n = 3$ ).

## 3. Results and discussion

To verify the design hypothesis, the fluorescence response of 2-(aminocarbonyl)phenylboronic acid to Cu(II) in various solvents was tested, as shown in Fig. 1a. The solutions of 2-(aminocarbonyl)phenylboronic acid alone exhibit weak fluorescence emission above 400 nm, while the maximum fluorescence enhancement could be observed in DMF upon the addition of Cu(II), indicating the relatively significant solvent effect in the sensing system. The effect of each component in the sensing system was investigated *via* a series of control experiments. As shown in Fig. 1b, we found that no fluorescence enhancement was produced regardless of discarding any of the components. Moreover, under ultraviolet irradiation at 365 nm, blue fluorescence could be observed with the naked eye in the complete detection system (inset of Fig. 1b). To gain insight into the proposed reaction mechanism, we confirmed the structure of the final fluorescent product in the millimolar level detection reaction as salicylamide by employing <sup>1</sup>H NMR and <sup>13</sup>C NMR (Fig. S1 and S2<sup>†</sup>). The aforementioned results clearly demonstrate that the reaction between Cu(II) and 2-(aminocarbonyl)

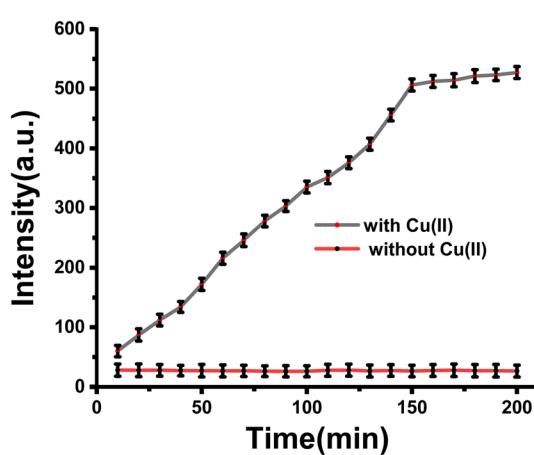


Fig. 2 The fluorescence intensity of the system in the presence and absence of Cu(II) at the indicated time. The detection concentration is 100  $\mu$ M for 2-(aminocarbonyl)phenyl boronic acid, 150  $\mu$ M for triethylamine and 50  $\mu$ M for Cu(II) in DMF. The fluorescence was monitored at 10 min intervals and collected at a wavelength of 410 nm. Each data point represents the average of the three trials. Error bars were calculated as standard deviations.

phenylboronic acid leads to the oxidative hydroxylation of the boronic acid moiety in the presence of Et<sub>3</sub>N, resulting in ESIPT fluorescence.

Alkaline conditions proved to be favorable for the Cu(II)-catalyzed nucleophilic substitution of boronic acid.<sup>40–42</sup> The effect of the triethylamine concentration on the assay system was then examined. As shown in Fig. S3,<sup>†</sup> the fluorescence intensity at 410 nm dramatically increased along with the increasing concentration of Et<sub>3</sub>N (0–210  $\mu$ M), and fluorescent intensity enhanced up to 13 times. Thus, the suitable concentration of triethylamine was chosen as 150  $\mu$ M. For further investigation, we optimized detection conditions, such as the H<sub>2</sub>O/DMF ratio and incubation time. It can be found that the fluorescence intensity decreased when elevating the ratio of

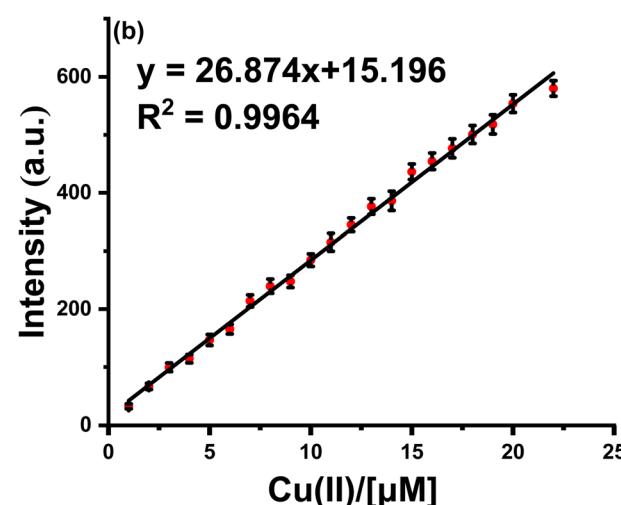
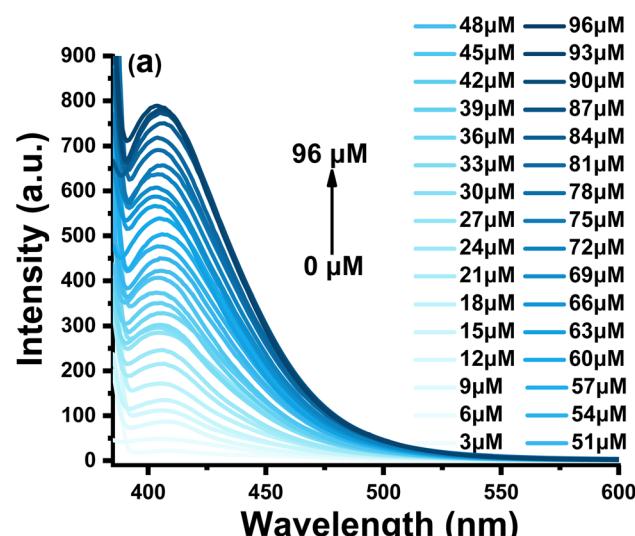


Fig. 3 (a) Fluorescence responses of the sensing system to serial concentrations of Cu(II). The detection concentration is 100  $\mu$ M for 2-(aminocarbonyl)phenyl boronic acid, 150  $\mu$ M for triethylamine and the concentration of Cu(II) is in the range of 0–96  $\mu$ M. (b) The linear correlations between fluorescence intensity and the concentration of Cu(II) (0–22  $\mu$ M),  $\lambda_{ex} = 365$  nm.



Table 1 Comparison of the proposed method for Cu(II) detection with other previously reported assays

Entry	Analytical method	Reagents/detection technique	Linear range (μM)	LOD (μM)	Ref.
1	FAAS	HNO <sub>3</sub> /deionized water	—	0.016	6
2	ICP-AES	8-Hydroxyquinoline/C18 column	—	0.16	7
3	UV, NIR	Carbon dots	0.3–1.6	0.09	44
4	UV-vis absorption spectrum	PNPs	0–8	0.062	45
5	“Turn off” fluorescence probe	NaClO <sub>4</sub> /reverse pulse polarography	0–1	0.0467	46
6	Fluorescence probe	AIE dots	0.025–6	0.1	47
7	“Turn-on” fluorescent probe	2-(Aminocarbonyl)phenylboronic acid	0–22	<b>0.068</b>	<b>This work</b>

Table 2 Study of the detection of Cu(II) in real water samples<sup>a</sup>

Samples	pH	Added amount (μM)	Found amount (μM)	Std dev. (μM)	RSD (%, n = 3)	Recovery (%, n = 3)
Tap water	6.42	20.00	18.90	0.36	0.93	94.50
Ganjiang river water	7.82	20.00	17.80	0.57	1.08	89.00
Rainwater	5.83	20.00	19.10	0.33	0.76	95.50

<sup>a</sup> Experimental condition: Cu(II) (20 μM), the concentrations of 2-(aminocarbonyl)phenyl boronic acid (100 μM) and triethylamine (150 μM); incubation time: 2.5 h. The pH values of the water samples were determined using a pH meter.

H<sub>2</sub>O/DMF and achieved the highest enhancement with the diluting solution as pure DMF at room temperature (25 °C) (Fig. S4†). Because the stock solution of Cu(II) was prepared in deionized water, the test solution contained a smidgen of water.

Incubation time is an important indicator of the response capability of the probe, so we examined the alteration of fluorescence at various incubation times. As shown in Fig. 2, no obvious fluorescence enhancement occurred in the absence of Cu(II), exhibiting the excellent stability of the probe in this system. After adding Cu(II), the fluorescence intensity of the sensing system increased gradually with an extended incubation time, reaching a steady state at 2.5 h. Consequently, we chose 2.5 h as the best incubation time for subsequent experiments.

To test sensitivity, fluorescence spectra with different concentrations of Cu(II) were studied. When the concentration of Cu(II) was increased from 0 to 96 μM, the fluorescence emission intensity increased significantly and obtained the highest 35-fold fluorescence enhancement (Fig. 3a). A linear equation of  $F_{410 \text{ nm}} = 15.196 + 26.874 [\text{Cu}^{2+}]$  ( $R^2 = 0.9964$ ) was obtained at a concentration range of 0–22 μM Cu(II), with a limit of detection (LOD) calculated as low as 68 nM based on  $\text{LOD} = 3\sigma/k$  (where  $\sigma$  is the standard deviation of blank measurement and  $k$  is the slope between the fluorescence intensity *versus* Cu(II) concentration) (Fig. 3b). As shown in Table S1,† the proposed assay is comparable to other previously reported fluorescence probes for monitoring Cu(II) (see Table S1† for details) and is much lower than the limit of Cu in drinking water (~20 μM) set by the U.S. Environmental Protection Agency.<sup>43</sup>

Furthermore, we summarized the studies of Cu(II) sensors on applicability and analytical methods in LOD and linear range, as illustrated in Table 1. Compared with the reported results, our study exhibits higher sensitivity, lower detection limit, and wider linear range. Particularly, our analytical method showed

some impressive advantages, such as operational simplicity, convenience and low cost.

High selectivity to detect objects over potentially competing species is an important indicator of the anti-interference capability of the probe. Competitive experiments on Cu(II) were carried out to evaluate the selectivity of the probe for Cu(II). As demonstrated in Fig. 4, only Cu(II) resulted in a remarkable fluorescence enhancement at 410 nm, whereas a range of common metal ions, including alkali and alkali-earth metal ions (K<sup>+</sup>, Na<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, Sr<sup>2+</sup>), transition metal ions (Ag<sup>+</sup>, Zn<sup>2+</sup>,

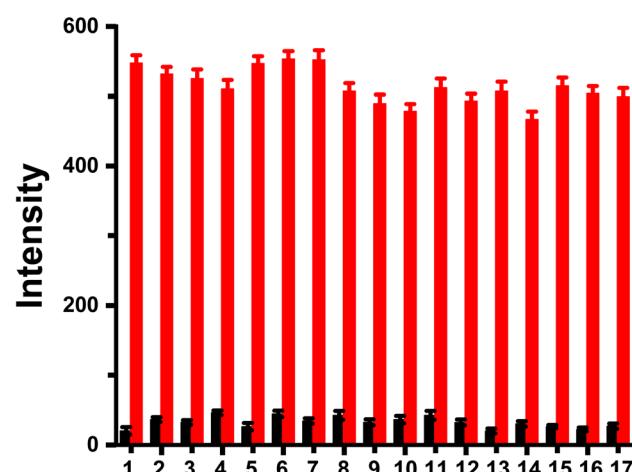


Fig. 4 Fluorescence intensity changes of probe (100 μM) upon the addition of common metal ions: 1, none; 2, K<sup>+</sup> (2 mM); 3, Na<sup>+</sup> (2 mM); 4, Ca<sup>2+</sup> (1 mM); 5, Mg<sup>2+</sup> (1 mM); 6, Sr<sup>2+</sup> (50 μM); 7, Ag<sup>+</sup> (50 μM); 8, Zn<sup>2+</sup> (50 μM); 9, Mn<sup>2+</sup> (50 μM); 10, Co<sup>2+</sup> (50 μM); 11, Pd<sup>2+</sup> (50 μM); 12, Hg<sup>2+</sup> (50 μM); 13, Fe<sup>3+</sup> (50 μM); 14, Cr<sup>3+</sup> (50 μM); 15, Al<sup>3+</sup> (50 μM); 16, Pr<sup>3+</sup> (50 μM); and 17, Ce<sup>4+</sup> (50 μM). Black bars: probe treated with marked metal ions in the absence of Cu(II). Red bars: probe treated with the marked metal ions followed by Cu(II) (50 μM).



$Mn^{2+}$ ,  $Co^{2+}$ ,  $Pd^{2+}$ ,  $Hg^{2+}$ ,  $Fe^{3+}$ ,  $Cr^{3+}$ ,  $Al^{3+}$ ) and rare-earth metal ions ( $Pr^{3+}$ ,  $Ce^{4+}$ ) merely caused slight enhancement in the fluorescence, even when  $K^+$ ,  $Na^+$ ,  $Mg^{2+}$  and  $Ca^{2+}$  were added at micromolar levels (Fig. 4). The results demonstrate the specific oxidative hydroxylation of the arylboronic acid catalyzed by  $Cu^{(II)}$  and indicate the capability of the probe in the selective detection of  $Cu^{(II)}$ .

Finally, a sensing system was applied to detect  $Cu^{(II)}$  in real water samples. Tap water, Ganjiang river water and rainwater were collected and pretreated based on previous studies.<sup>48,49</sup> When exogenous  $Cu^{(II)}$  (20  $\mu M$ ) was added to these samples, the fluorescence intensity increased significantly. As shown in Table 2, the recovery rates are 94.50% and 95.50% with relative standard deviations of 0.93% and 0.76% for tap water and rainwater, respectively. These results suggest that the fluorescence probe has potential applicability for  $Cu^{(II)}$  monitoring in real water samples.

## 4. Conclusions

In this study, we reported a simple structure of arylboronic acid as a “turn-on” fluorescence probe for the detection of  $Cu^{(II)}$ . The recognition mechanism is based on the  $Cu^{(II)}$ -catalyzed oxidative hydroxylation of the arylboronic acid moiety to phenol, leading to the recovery of the ESIPT effect and switching on the fluorescent. The probe exhibited excellent sensitivity (detection limit of 68 nM) and high selectivity for  $Cu^{(II)}$  over 16 other metal ions. The probe is also reliable for determining exogenous  $Cu^{(II)}$  in real water samples from Ganzhou, demonstrating its potential applicability for the detection of  $Cu^{(II)}$  in real-world scenarios.

## Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have influenced the work reported in this paper.

## Acknowledgements

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## References

- 1 K. J. Barnham and A. I. Bush, *Chem. Soc. Rev.*, 2014, **43**, 6727.
- 2 E. Gaggelli, H. Kozlowski, D. Valensin and G. Valensin, *Chem. Rev.*, 2006, **106**, 1995.
- 3 A. Lamboux, E. Couchonnal-Bedoya, O. Guillaud, C. Laurencin, L. Lion-François, A. Belmalih, E. Mintz, V. Brun, M. Bost, A. Lachaux and V. Balter, *Metallomics*, 2020, **12**, 1781.
- 4 C. S. Yang, W. L. Lu and K. H. Lu, *Chin. J. Anal. Chem.*, 2000, **28**, 68.
- 5 H. B. Zengin and R. Gürkan, *Talanta*, 2021, **224**, 121789.
- 6 Y. R. Hou, J. L. Lu, Y. C. Fan, K. Abudusalamu, X.-D. Tang, Q. Q. Wei, J.-K. Guo and W. Zhao, *Spectrosc. Spectral Anal.*, 2022, **42**, 2102.
- 7 G. Abbasse, B. Ouddane and J. C. Fischer, *J. Anal. At. Spectrom.*, 2002, **17**, 1354.
- 8 T. A. Ali, A. A. Abd-Elaal and G. G. Mohamed, *Microchem. J.*, 2021, **160**, 105693.
- 9 M. Amjadi and Z. Abolghasemi-Fakhri, *Sens. Actuators, B*, 2018, **257**, 629.
- 10 D. Udhayakumari, S. Naha and S. Velmathi, *Anal. Methods*, 2017, **9**, 552.
- 11 J. Zhao, Y. Y. Wang, W. L. Chen, G. S. Hao, J. P. Sun, Q. F. Shi, F. Tian and R. T. Ma, *RSC Adv.*, 2022, **12**, 3073.
- 12 J. G. Huang, M. Liu, X. Q. Ma, Q. Dong, B. Ye, W. Wang and W. B. Zeng, *RSC Adv.*, 2014, **4**, 22964.
- 13 Z. P. Li, Y. W. Zhang, H. Xia, Y. Mu and X. M. Liu, *Chem. Commun.*, 2016, **52**, 6613.
- 14 L. Q. Li, M. H. Zheng, X. Y. Yan, H. Huang, S. X. Cao, K. M. Liu and J.-B. Liu, *J. Photochem. Photobiol. A*, 2022, **432**, 114096.
- 15 Q. X. Ye, S. F. Ren, H. Huang, G. G. Duan, K. M. Liu and J. B. Liu, *ACS Omega*, 2020, **5**, 20698.
- 16 W. H. Wang, T.-L. Yung, S. S. Cheng, F. Chen, J. B. Liu, C.-H. Leung and D.-L. Ma, *Sens. Actuators, B*, 2020, **132**, 128486.
- 17 D. J. Zhu, Y. H. Luo, X. W. Yan, W. Xie, W. Cai and X. Zhong, *RSC Adv.*, 2016, **6**, 87110.
- 18 R. B. An, D. T. Zhang, Y. Chen and Y. Z. Cui, *Sens. Actuators, B*, 2016, **222**, 48.
- 19 M. Tian, H. He, B. B. Wang, X. Wang, Y. Liu and F. L. Jiang, *Dyes Pigm.*, 2019, **165**, 383.
- 20 K. Y. Liu, H. M. Shang, F. F. Meng, Y. Liu and W. Y. Lin, *Talanta*, 2016, **147**, 193.
- 21 M. Taki, S. Iyoshi, A. Ojida, I. Hamachi and Y. Yamamoto, *J. Am. Chem. Soc.*, 2010, **132**, 5938.
- 22 Z. H. Shi, X. L. Tang, X. Y. Zhou, J. Cheng, Q. X. Han, J. A. Zhou, B. Wang, Y. F. Yang, W. S. Liu and D. C. Bai, *Inorg. Chem.*, 2013, **52**, 12668.
- 23 K. K. Yu, K. Li, J. T. Hou and X. Q. Yu, *Tetrahedron Lett.*, 2013, **54**, 5771.
- 24 X. M. Wu, Z. Q. Guo, Y. Z. Wu, S. Q. Zhu, T. D. James and W. H. Zhu, *ACS Appl. Mater. Interfaces*, 2013, **5**, 12215.
- 25 Y. K. Jang, U. C. Nam, H. L. Kwon, I. H. Hwang and C. Kim, *Dyes Pigm.*, 2013, **99**, 6.
- 26 J. B. Li, Y. Zeng, Q. H. Hu, X. L. Yu, J. Guo and Z. Q. Pan, *Dalton Trans.*, 2012, **41**, 3623.
- 27 Q. Y. Wu and E. V. Anslyn, *J. Am. Chem. Soc.*, 2004, **126**, 14682.
- 28 Z. J. Hu, J. W. Hu, Y. Cui, G. N. Wang, X. J. Zhang, K. Uvdalb and H. W. Gao, *J. Mater. Chem. B*, 2014, **2**, 4467.
- 29 Z. Zhang, Y. Zou and C. Q. Deng, *RSC Adv.*, 2017, **7**, 14742.
- 30 K. M. K. Swamy, S.-K. Ko, S. K. Kwon, H. N. Lee, C. Mao, J.-M. Kim, K.-H. Lee, J. Kim, I. Shin and J. Yoon, *Chem. Commun.*, 2008, 5915.



31 J. M. V. Ngororabanga, C. Moyo, E. Hosten, N. Mama and Z. R. Tshentu, *Anal. Methods*, 2019, **11**, 3857.

32 M. Taki, K. Akaoka, K. Mitsui and Y. Yamamoto, *Org. Biomol. Chem.*, 2014, **12**, 4999.

33 D. Maity, A. Raj, D. Karthigeyan, T. K. Kundu and T. Govindaraju, *RSC Adv.*, 2013, **3**, 16788.

34 M. Y. Pei, H. H. Kong, A. Q. Tian, X. Liu, K. B. Zheng, Z. L. Ren and L. Wang, *J. Mol. Struct.*, 2022, **1250**, 131806.

35 R. A. Coburn, A. J. Batista, R. T. Evans and R. J. Genco, *J. Med. Chem.*, 1981, **24**, 1245.

36 J. M. Xu, J. Berastegui-Cabrera, H. Y. Chen, J. Pachón, J. Zhou and J. Sánchez-Céspedes, *J. Med. Chem.*, 2020, **63**, 3142.

37 A. Weller, *Naturwissenschaften*, 1955, **42**, 175.

38 T. Nishiya, S. Yamauchi, N. Hirota, M. Baba and I. Hanazaki, *J. Phys. Chem.*, 1986, **90**, 5730.

39 J. Catalan, F. Toribio and A. U. Acuna, *J. Phys. Chem.*, 1982, **86**, 303.

40 D. M. T. Chan, K. L. Monaco, R. H. Li, D. Bonne, C. G. Clark and P. Y. S. Lam, *Tetrahedron Lett.*, 2003, **44**, 3863.

41 J. M. Xu, X. Y. Wang, C. W. Shao, D. Y. Su, G. L. Cheng and Y. F. Hu, *Org. Lett.*, 2010, **12**, 1964.

42 A. A. Atia and M. Kimura, *Catalysts*, 2020, **10**, 1262.

43 Office of Water U.S. Environmental Protection Agency, *Edition of the Drinking Water Standards and Health Advisories*, U.S. EPA Office of Science and Technology, Washington, D.C., 2012, EPA 822-S-12 001.

44 A. Salinas-Castillo, M. Ariza-Avidad, C. Pritz, M. Camprubí-Robles, B. Fernández, M. J. Ruedas-Rama, A. Megia-Fernández, A. Lapresta-Fernández, F. Santoyo-Gonzalez, A. Schrott-Fischer and L. F. Capitan-Vallvey, *Chem. Commun.*, 2013, **49**, 1103.

45 S. G. Liu, N. Li, Y. Z. Fan, N. B. Li and H. Q. Luo, *Sens. Actuators, B*, 2017, **243**, 634.

46 G. Z. Huang, C. Li, X. T. Han, S. O. Aderinto, K. S. Shen, S. S. Mao and H. L. Wu, *Luminescence*, 2018, **33**, 660.

47 R. Jiang, N. Liu, F. Li, W. S. Fu, Y. Zhou and Y. Zhang, *Polymers*, 2018, **10**, 786.

48 A. R. Jacobson, S. Dousset, F. Andreux and P. C. Baveye, *Environ. Sci. Technol.*, 2007, **41**, 6343.

49 Y. D. Wang, J. H. Li, Y. Gao, Y. Yang, Y. Gao and Z. F. Xu, *Hydrometallurgy*, 2020, **191**, 105220.

