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# Engineering exposed vertical nano-TiO<sub>2</sub> (001) facets/BiOI nanosheet heterojunction film for constructing a satisfactory PEC glucose oxidase biosensor†

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In the field of photoelectrochemical (PEC) enzyme biosensors, constructing efficient photoelectrodes, in which the recombination of photogenerated carriers is an important factor affecting the performance, is of great significance. Herein, to enhance the separation efficiency of photogenerated carriers, titanium dioxide (TiO<sub>2</sub>) nanosheet (NS)/bismuth oxyiodide (BiOI) NS/glucose oxidase (GOx) composites were prepared *via* hydrothermal and solvothermal methods. Single-crystal anatase TiO<sub>2</sub> NSs with a high percentage of (001) facets lead to better photocarrier separation due to heterojunctions between facets. After coupling with BiOI NSs, the photoelectrochemical performance of the electrode was greatly improved. The photogenerated electrons from TiO<sub>2</sub> and BiOI gathered at TiO<sub>2</sub> (101) and were exported through the fluorine-doped tin oxide (FTO) substrate to generate electrical signals. Photogenerated holes were transferred to TiO<sub>2</sub> (001) and BiOI to participate in the enzymatic reaction, showing the outstanding separation of electrons and holes. The prepared TiO<sub>2</sub> NS/BiOI NS/GOx glucose biosensor achieved satisfactory results, with sensitivity of 14.25 μA mM<sup>-1</sup> cm<sup>-2</sup>, a linear measurement range of 0–1 mM, and a limit of detection (3S/N) of 0.01 mM in phosphate buffered saline (PBS) at a pH of 7.4. The mechanism for the efficient separation of photogenerated carriers based on the facet heterojunctions introduced in this paper also provides new insights into other optoelectronic biosensors.

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## 1. Introduction

Photoelectrochemical (PEC) stands out for its advantages of low cost, high sensitivity, and strong anti-interference, and has broad application foregrounded in biological analysis.<sup>1,2</sup> The performance of photoelectrodes plays a crucial role in biosensors, and the recombination of photogenerated carriers in the bulk phase is one of the factors affecting performance. Therefore, constructing an electrode that could efficiently separate photogenerated carriers is important.

Two-dimensional titanium dioxide (TiO<sub>2</sub>) nanosheets (NSs) are splendid alternatives for PEC biosensors due to their high

specific surface area and chemical reaction facets. Compared with one-dimensional TiO<sub>2</sub> nanorods, anatase TiO<sub>2</sub> NSs with a high percentage of (001) facets are more competitive in solar cells, photonic and optoelectronic devices, sensors, and the photocatalysis field.<sup>3</sup> Previous reports showed that anatase TiO<sub>2</sub> (101) are enriched facets for electrons. The electrons generated by photoexcitation on (001) facets would spontaneously migrate to (101) facets.<sup>4</sup> This behavior could alleviate the problem of bulk recombination of photogenerated carriers for single crystal materials. However, the application of TiO<sub>2</sub> in the visible light range is very restricted due to its large band gap energy.<sup>5</sup> To improve the response of TiO<sub>2</sub> in the visible light range, common methods include deposition of noble metal quantum dots, element doping, and modification with narrow band gap semiconductors.<sup>6–8</sup> Bismuth oxyiodide (BiOI) is a p-type semiconductor that has a narrow band gap between 1.7 and 1.9 eV, and a visible light response.<sup>9</sup> After p-type BiOI has been assembled with n-type TiO<sub>2</sub>, the photogenerated electrons of BiOI quickly transfer to the conduction band of TiO<sub>2</sub>, and the photogenerated holes of TiO<sub>2</sub> move to the valence band of BiOI, thereby achieving separation of electrons and holes.<sup>10</sup>

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Herein, a PEC enzyme biosensor was developed based on BiOI NS modified  $\text{TiO}_2$  NSs for glucose detection.  $\text{TiO}_2$  NSs were uniformly grown on the surface of a fluorine-doped tin oxide (FTO) substrate by a hydrothermal method. The tight binding of  $\text{TiO}_2$  to the substrate was beneficial to the transport of electrons. Furthermore, the exposed (001) facets of  $\text{TiO}_2$  contributed to the separation of photogenerated carriers due to facet heterojunctions. In addition, BiOI NSs grown *in situ* on  $\text{TiO}_2$  (101) planes by the solvothermal method could form a heterojunction with  $\text{TiO}_2$  and improve the separation of photogenerated charges and the absorption of material in the visible light range. The (101) facet of  $\text{TiO}_2$  acted as an electron-rich facet to accept photogenerated electrons from the (001) facets and BiOI, and transported them to the external circuit through the FTO substrate, whereas the photogenerated holes were transferred to the (001) facets and BiOI to participate in the enzymatic reaction, so that the electrons and holes were separated in space. In conclusion, the synergistic effect of crystal-plane heterojunctions and narrow-band gap semiconductors enhances the separation efficiency of photogenerated carriers and the absorption of visible light, providing new insights for other optoelectronic biosensors.

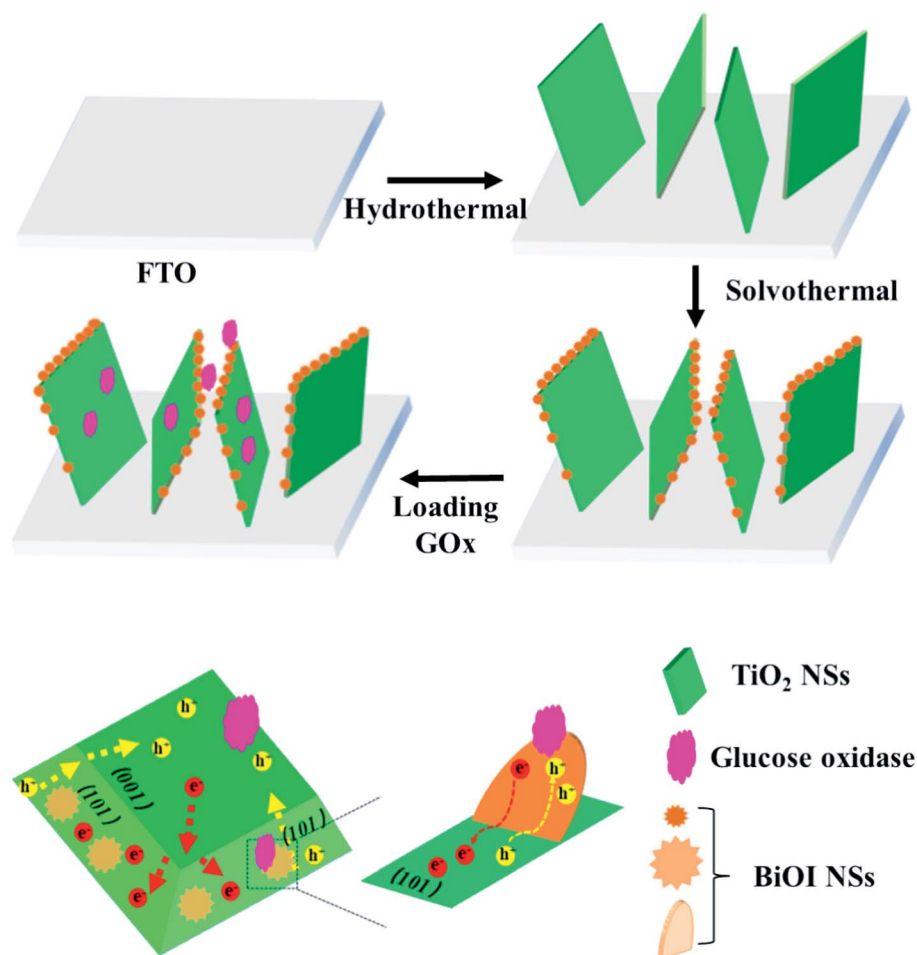
## 2. Experimental

### 2.1 Reagents and materials

FTO substrates were acquired from Outhwaite New Energy Company. Ammonium fluorotitanate ( $(\text{NH}_4)_2\text{TiF}_6$ ), tetrabutyl titanate ( $\text{C}_{16}\text{H}_{36}\text{O}_4\text{Ti}$ ), glucose oxidase (GOx), and bismuth nitrate pentahydrate ( $\text{Bi}(\text{NO}_3)_3 \cdot 5\text{H}_2\text{O}$ ) were purchased from Shanghai Maclin Biochemical Technology Co., Ltd. Ethylene glycol, ethanol, potassium iodide (KI), glucose, and hydrochloric acid (HCl) were acquired from the Guangzhou Chemical Reagent Factory. The water solvent was ultrapure water prepared in the laboratory. All reagents and solutions were not further processed.

### 2.2 Synthesis of $\text{TiO}_2$ NSs

The FTO substrates were thoroughly ultrasonically cleaned in isopropyl alcohol, acetone, and water for 20 min, and dried under pure argon flow gas for later use. Concentrated HCl was diluted with ultra-pure water at a ratio of 1 : 1 and stirred for 10 min. Then, 0.4 mL of tetrabutyl titanate was added drop by drop to 20 mL of diluted HCl and stirred vigorously for 30 min. Next, 0.15 g of ammonium fluorotitanate was added to the



Scheme 1 The fabrication of the  $\text{TiO}_2$  NS/BiOI NS electrode.



above solution and stirred for 30 min. Then, the mixture was transferred to a 30 mL inner lining of the autoclave, which contained the prepared FTO substrates. The conductive side of the FTO faced down, and the angle between it and the inner wall of the Teflon-lined autoclave was about 30°. After 12 h of hydrothermal reaction at 170 °C, when the autoclave was completely cooled down naturally, the samples were removed, thoroughly rinsed with deionized water, dried at 60 °C for 2 h in an oven, and annealed at 450 °C for 2 h in air.

### 2.3 Synthesis of TiO<sub>2</sub> NSs/BiOI NSs

The TiO<sub>2</sub> NS/BiOI NS electrode was synthesized by the secondary solvothermal method. 10 mM of KI and Bi(NO<sub>3</sub>)<sub>2</sub> ethylene glycol solution were prepared and fully stirred. 10 mL of Bi(NO<sub>3</sub>)<sub>2</sub> ethylene glycol solution was poured into a beaker, and kept under vigorous stirring; then 10 mL of KI ethylene glycol solution was added into the beaker dropwise, with continuous stirring for 30 min. Next, the resultant solution was transferred to a 30 mL Teflon-lined autoclave, which contained the FTO substrates with TiO<sub>2</sub> NSs. The temperature and time of the solvothermal reaction were set to 160 °C and 10 h, respectively. Finally, the samples were rinsed thoroughly with water and ethanol, and then dried in an oven at 60 °C for 2 h.

### 2.4 Fabrication of the PEC electrode

The prepared TiO<sub>2</sub> NS/BiOI NS electrodes were placed in a freezer at 4 °C for 30 min. The preparation of the electrode is shown in Scheme 1. Then, 15 μL of activated glucose oxidase 10 mg mL<sup>-1</sup>, Nafion solvent (1% w/w) solution, and 5 μL of chitosan solution were dropped onto the surface of the electrode and placed in an environment of 4 °C for 48 h until the surface liquid was completely dry.

### 2.5 Detection and instruments

The electrochemical tests used equipment from Chenhua Electrochemical Equipment, Shanghai, China, and the model was CHI660E. The light source was a simulated daylight xenon light source provided by Beijing CEALight Technology Co., Ltd, and the product model was CEL-S500. The X-ray diffraction (XRD) analysis used an instrument from the Rigaku Co., Ltd of Japan. The equipment model was D/max 2550 V, diffracted with monochromatized Cu K $\alpha$  radiation ( $\lambda = 0.15418$  nm). In the experimental test, the rotation angle went from 10° to 80°, the scanning speed was 10° min<sup>-1</sup>, and each step was 0.01°. The UV-visible light absorption spectrum was taken with a Japanese Jasco UV-Vis absorption spectrophotometer; the model was UV-550, and the wavelength range set in this experiment went from

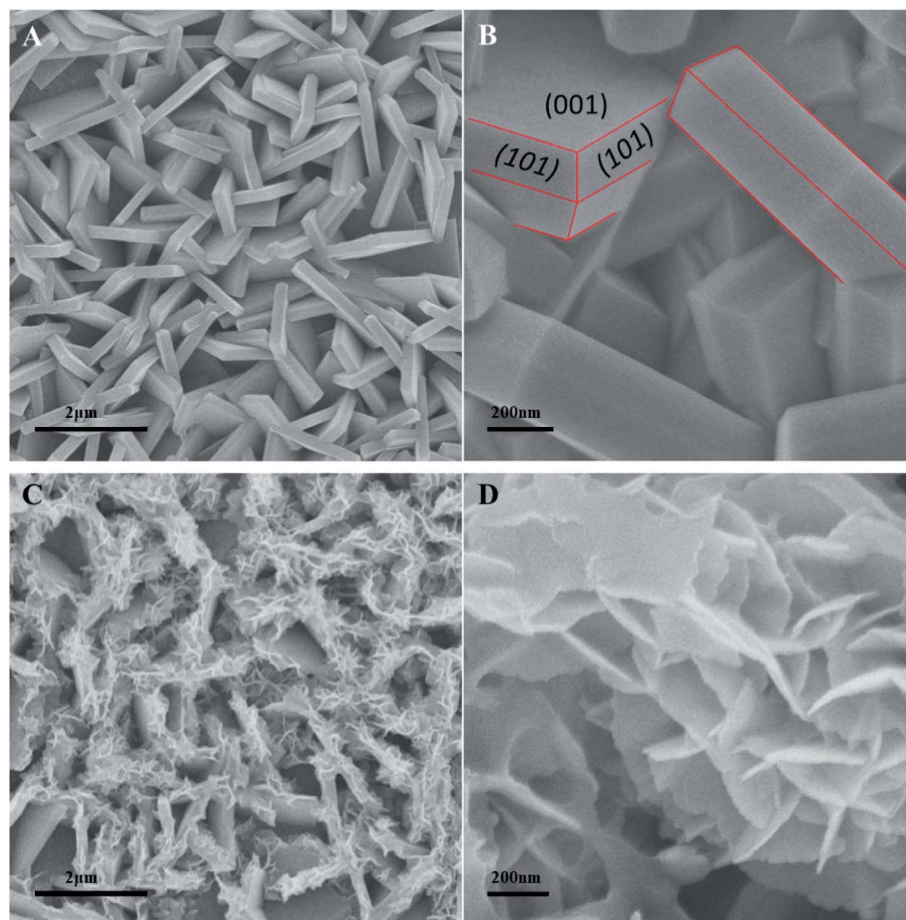


Fig. 1 SEM images of (A and B) TiO<sub>2</sub> NSs and (C and D) TiO<sub>2</sub> NSs/BiOI NSs.



200 nm to 800 nm. The scanning electron microscope (SEM) was supplied by Hitachi, Japan, and the model was Hitachi SU 8010. The transmission electron microscope (TEM) was manufactured by JEOL, Japan, and the model was JEOL JEM 2100. The chemical composition of the sample was measured by an X-ray electron spectroscopy (XPS) instrument produced by Kratos; the model was Axis Ultra DLD, and the excitation source was an Al- $K\alpha$  monochromator.

### 3. Results and discussion

To characterize the morphology of the electrode materials, the materials prepared in this work were observed by SEM. Fig. 1A shows that the FTO substrates are densely and uniformly covered with NSs, which cross one another and are closely arranged. The intimate, homogeneous combination between  $\text{TiO}_2$  NSs and FTO substrate could ensure fast electron transfer.

Fig. S1† shows that  $\text{MnO}_2$  is photo-deposited on the  $\text{TiO}_2$  NSs. The results show that  $\text{MnO}_2$  has the characteristics of selective growth in space, indicating that the exposed facets are hole-enriched (001) facets. Fig. 1B shows that the upper and lower sides of the NSs are square (001) facets, the sides are composed of symmetrical isosceles trapezoids corresponding to the (101) facets, the average thickness of the NS is about 190 nm, and the average side length of the NS is about 1.2  $\mu\text{m}$ .

Fig. 1C and D present the  $\text{TiO}_2$  NS/ $\text{BiOI}$  NS images synthesized by a secondary solvothermal method. A cluster of NS structures grows on top of the  $\text{TiO}_2$  NSs. In addition, Fig. 1C shows that  $\text{BiOI}$  NSs selectively grow on the (101) facets of  $\text{TiO}_2$ . Combined with the results of XPS,  $\text{TiO}_2$  NSs still contain a small amount of  $\text{F}^+$  after annealing, which is closely combined with  $\text{TiO}_2$  (001) facets as a morphology regulator. The reason could be the repulsion between halogen atoms, which is not conducive to  $\text{BiOI}$  NS growth on  $\text{TiO}_2$  (001) facets. In Fig. 1D, the

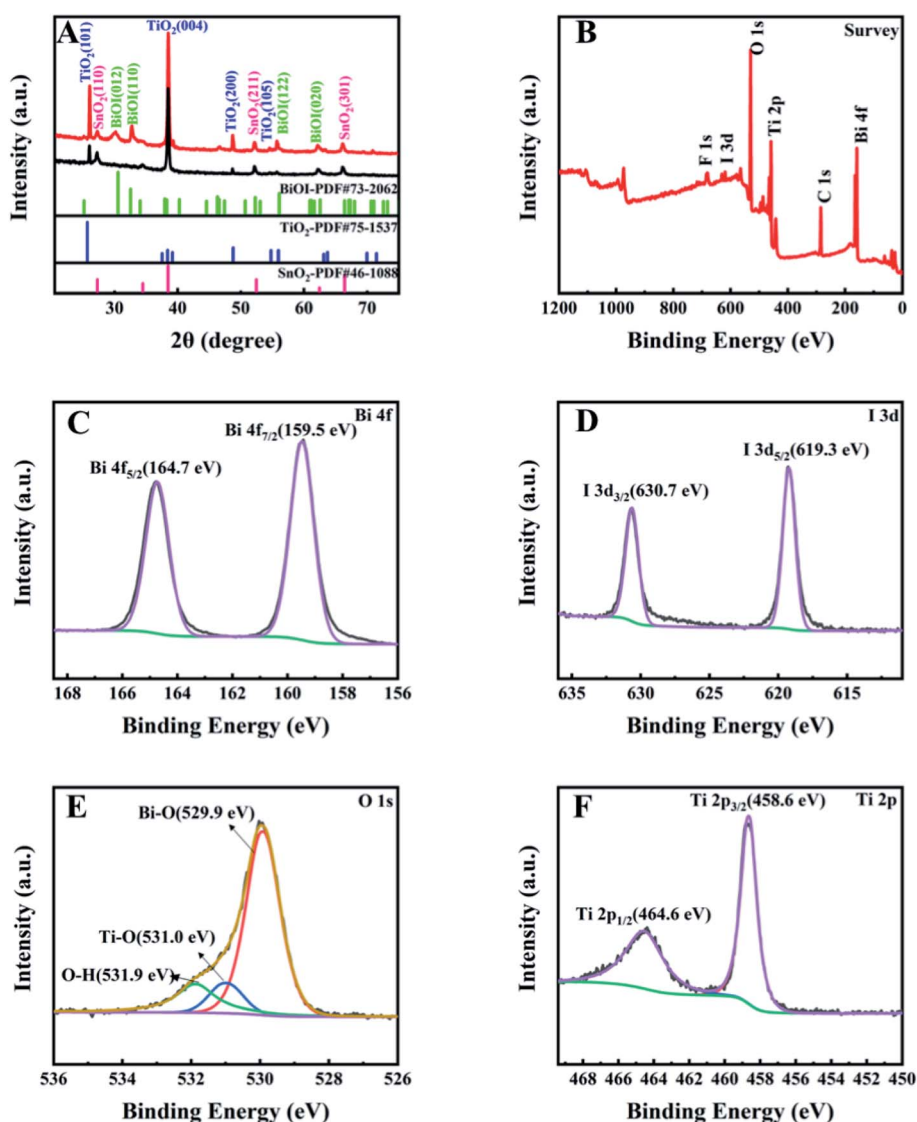


Fig. 2 (A) XRD patterns of  $\text{TiO}_2$  NSs (black curve) and  $\text{TiO}_2$  NSs/ $\text{BiOI}$  NSs (red curve). XPS patterns: (B) survey spectra, (C) Bi 4f, (D) I 3d, (E) O 1s, and (F) Ti 2p.



thickness of the BiOI nano structure in the magnified image is about 20 nm.

To investigate the crystal structure of the electrode materials, the materials synthesized in this work were analyzed by XRD. The black and red curves in Fig. 2A are the XRD patterns of TiO<sub>2</sub> NSs and TiO<sub>2</sub> NSs/BiOI NSs, respectively, and the crystal planes of the substances corresponding to the diffraction peaks are marked. The prepared TiO<sub>2</sub> NSs are in the anatase phase. Diffraction peaks located at 25.9°, 38.4°, and 48.6° correspond to the (101), (004), and (200) facets of TiO<sub>2</sub>, respectively, consistent with the PDF standard card JCPDS Number 75-1537. The diffraction peaks (red curve) at 29.7°, 31.7°, 45.5°, and 51.4° correspond to the (012), (110), (020), and (114) facets of BiOI, respectively, indicating that the prepared BiOI NSs have a tetragonal chloroformate structure<sup>11</sup> and correspond to the standard PDF card JCPDS Number 73-2062. The diffraction peaks are very sharp, verifying the high crystallinity of the samples. Thus, a binary composite composed of TiO<sub>2</sub> NSs and BiOI NSs was successfully synthesized. XPS was used to explore the elemental composition and valence state of the TiO<sub>2</sub> NS/BiOI NS electrode. According to the survey XPS spectrum of Fig. 2B, the main constituent elements of the electrode are Bi, I, O, and Ti (the spectrum was calibrated with the banding energy

of C 1s). The high-resolution spectrum of each element is collated in Fig. 2C–F. Fig. 2C shows two Bi 4f peaks for the sample at 159.5 and 164.7 eV, which are attributed to Bi 4f<sub>7/2</sub> and Bi 4f<sub>5/2</sub>, respectively.<sup>12</sup> Fig. 2D shows two I 3d peaks at 619.3 and 630.7 eV that belong to I 3d<sub>5/2</sub> and I 3d<sub>3/2</sub>, respectively, meaning that the valence of I is negative monovalent.<sup>13</sup> Fig. 2E shows that the O 1s peaks consist of three individual binding energies of 529.9, 531.0, and 531.9 eV that could belong to the Bi–O bond, Ti–O bond, and O–H bond, respectively.<sup>14,15</sup> In addition, the appearance of an O–H bond may be due to the hydroxyl group in the water adsorbed by the sample.<sup>16</sup> Fig. 2F shows that the two peaks at 458.6 and 464.6 eV are assigned to Ti 2p<sub>3/2</sub> and Ti 2p<sub>1/2</sub>, indicating that the valence of Ti is +4.<sup>17</sup>

The microstructure and morphology of TiO<sub>2</sub> NSs and TiO<sub>2</sub> NSs/BiOI NSs were further characterized by TEM. Fig. 3A shows the TEM image of the prepared TiO<sub>2</sub> NSs. The morphology is matched with that observed by SEM, showing a rectangular outline. Fig. 3B shows the crystalline lattice fringes of TiO<sub>2</sub> NSs in the enlarged image are 0.35 nm, which correspond to anatase (101) facets. The selected area electron diffraction (SAED) pattern is taken from the NS part, according to the bright, periodically arranged diffraction spots, indicating that the prepared TiO<sub>2</sub> NSs are single crystals and show excellent

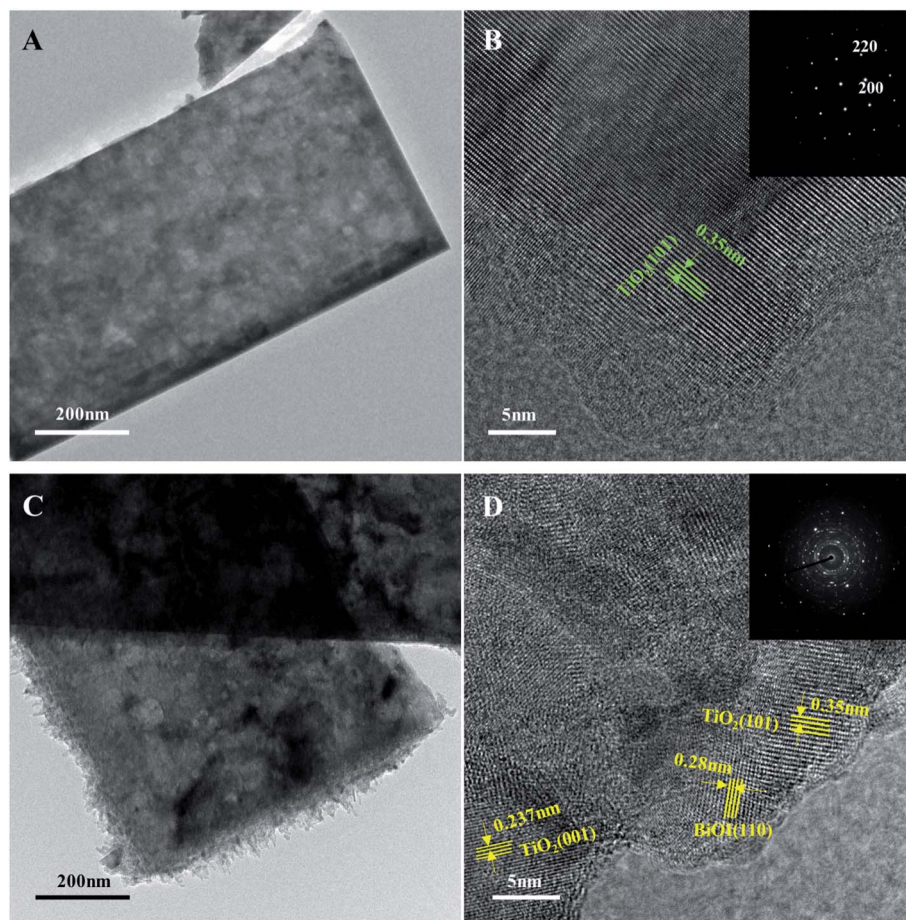


Fig. 3 (A) TEM and (B) HRTEM images of TiO<sub>2</sub> NSs and (inset) the SAED pattern. (C) TEM and (D) HRTEM images of TiO<sub>2</sub> NSs/BiOI NSs and (inset) the SAED pattern.



crystallinity.<sup>18</sup> The distance between the diffraction spots and the center point was measured and proofread with standard PDF cards, suggesting that the TiO<sub>2</sub> NSs are taken along the [001] zone axis.<sup>19</sup> Fig. 3C is the TEM image of a single TiO<sub>2</sub> NS grown with BiOI NSs. Nanosized lamellar structures selectively grow on the edge of the NS, and the results are consistent with the SEM observations. Fig. 3D is a high-resolution TEM image showing the clear lattice fringes of the TiO<sub>2</sub> NS/BiOI NS structure. The lattice spacings of 0.237 and 0.35 nm correspond to the (001) and (101) facets of anatase phase TiO<sub>2</sub>, respectively. In addition, the lattice fringes exhibit a lattice spacing of 0.28 nm, corresponding to the (110) facets of BiOI. The inset SAED of the area of Fig. 3D shows a ring-like pattern and has a polycrystalline feature. The EDS spectra and EDX mapping images (Fig. S2†) of the TiO<sub>2</sub> NSs/BiOI NSs further confirm the successful preparation of TiO<sub>2</sub> and BiOI binary composite electrodes.

The light absorption properties of TiO<sub>2</sub> NSs and TiO<sub>2</sub> NSs/BiOI NSs in the range of 250–800 nm were studied with UV-vis diffuse reflectance spectra. Fig. 4A shows that, after BiOI NSs were modified on TiO<sub>2</sub> NSs, the absorption peaks of the samples shifted significantly to the visible region, and the absorption edge increased from 391 nm to 544 nm because BiOI is a narrow band gap semiconductor and could be excited by longer wavelength light, which is beneficial to keeping the bioactivity of the enzyme immobilized on the TiO<sub>2</sub> NS/BiOI NS electrode<sup>11</sup> and reducing the occurrence of side reactions in the detection system.<sup>20</sup>

The band gap values could be calculated with the following formula:<sup>21</sup>

$$(\alpha h\nu)^n = B(h\nu - E_g) \quad (1)$$

where  $\alpha$ ,  $h$ ,  $\nu$ ,  $B$ , and  $E_g$  represent absorption coefficients, the Planck constant, light frequency, a proportionality constant,

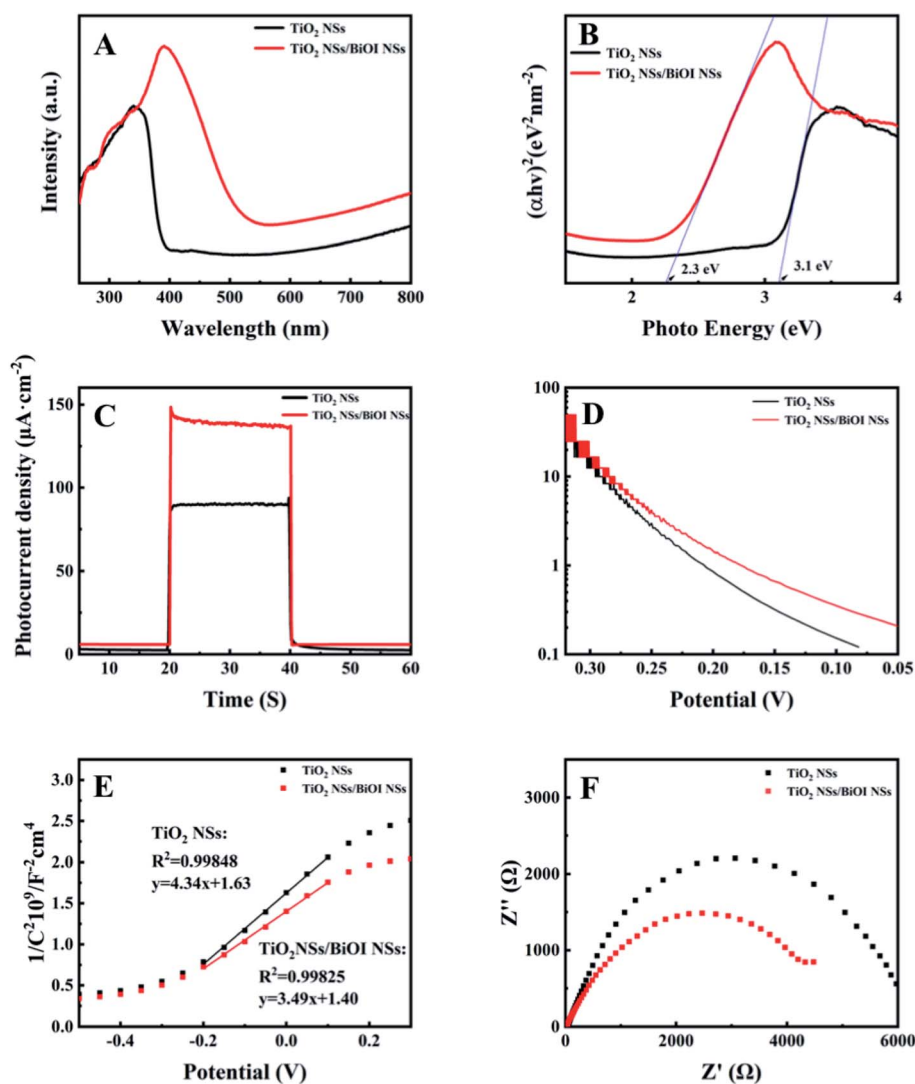


Fig. 4 (A) UV-vis diffuse reflectance spectra of TiO<sub>2</sub> NSs (black curve) and TiO<sub>2</sub> NSs/BiOI NSs (red curve). (B) Plots of  $(F(R)h\nu)^{1/2}$  versus  $E$  (eV). (C) Photocurrent responses of TiO<sub>2</sub> NSs and TiO<sub>2</sub> NSs/BiOI NSs electrodes. (D) Electronic lifetimes in 0.1 M PBS (pH = 7.4). (E) Mott–Schottky plots. (F) EIS Nyquist plots.



and the energy band gap, respectively.  $\text{TiO}_2$  and BiOI are indirect semiconductors, so the value of  $n$  is  $1/2$ . The data were brought in, and the band gap values of  $\text{TiO}_2$  NSs and  $\text{TiO}_2$  NSs/BiOI NSs were 3.1 and 2.3 eV, respectively, as shown in Fig. 4B.

Fig. 4C shows the photoelectrical response of  $\text{TiO}_2$  NSs and  $\text{TiO}_2$  NS/BiOI NS electrodes under simulated sunlight irradiation. The electrode was significantly improved after the BiOI NSs were modified. To explore the PEC performance of the electrode further, a three-electrode system was constructed to detect the intrinsic electronic characteristics of the electrode, where the  $\text{TiO}_2$ /NS/BiOI NS electrode was the working electrode, a Pt electrode was the counter electrode, and an Ag/AgCl electrode was the reference electrode (saturated KCl).

The electrode generated a photovoltage as photogenerated electron-hole pairs accumulated on the cathode and anode under illumination.<sup>22</sup> When the light was canceled, the photogenerated holes collected on the anode and the photogenerated electrons collected on the cathode quickly recombined, and then the open circuit potential was recorded. The lifetime of the photogenerated carriers could be calculated with formula (2).<sup>23</sup>

$$\tau = \frac{k_B T}{e} \left( \frac{dV_\alpha}{dt} \right)^{-1}, \quad (2)$$

where  $\tau$  is the electron lifetime;  $V_\alpha$  is the open-circuit voltage at time  $t$ ;  $k_B$ ,  $e$ , and  $T$  are the Boltzmann constant, absolute temperature, and charge of a single electron, respectively. The

electron lifetime of the corresponding samples could be calculated from known, measured data, and the result is shown in Fig. 4D. Clearly, the carrier lifetime of the  $\text{TiO}_2$  NS/BiOI NS electrode is greater than that of  $\text{TiO}_2$  NSs. Consequently, by forming a heterojunction with BiOI NSs, the carrier lifetime of the  $\text{TiO}_2$  NS electrode is significantly prolonged, and the separation of carriers is promoted.

Formula (3)<sup>24</sup> was used to calculate capacitance, and the carrier density ( $N_D$ ) could be obtained through its conversion form.

$$\frac{1}{C^2} = \frac{2}{N_D e \epsilon_0 \epsilon} \left[ (U_S - U_{FB}) - \frac{k_B T}{e} \right], \quad (3)$$

where  $C$  is the space charge capacitance;  $e$  is the element charge value;  $\epsilon_0$  and  $\epsilon$  are vacuum permittivity and relative permittivity, respectively;  $U_S$  and  $U_{FB}$  are the applied potential and Fermi energy in the semiconductor, respectively;  $k_B$  is the Boltzmann constant;  $T$  is the temperature.

After conversion, the formula for the calculation of  $N_D$  was as follows:

$$N_D = - \left( \frac{2}{e \epsilon_0 \epsilon} \right) \left( \frac{d \left( \frac{1}{C^2} \right)}{d(U_S)} \right)^{-1}, \quad (4)$$

where  $N_D$  is inversely proportional to the first derivative of  $1/C^2$ . Hence, the slope value linearly fitted by the samples could be

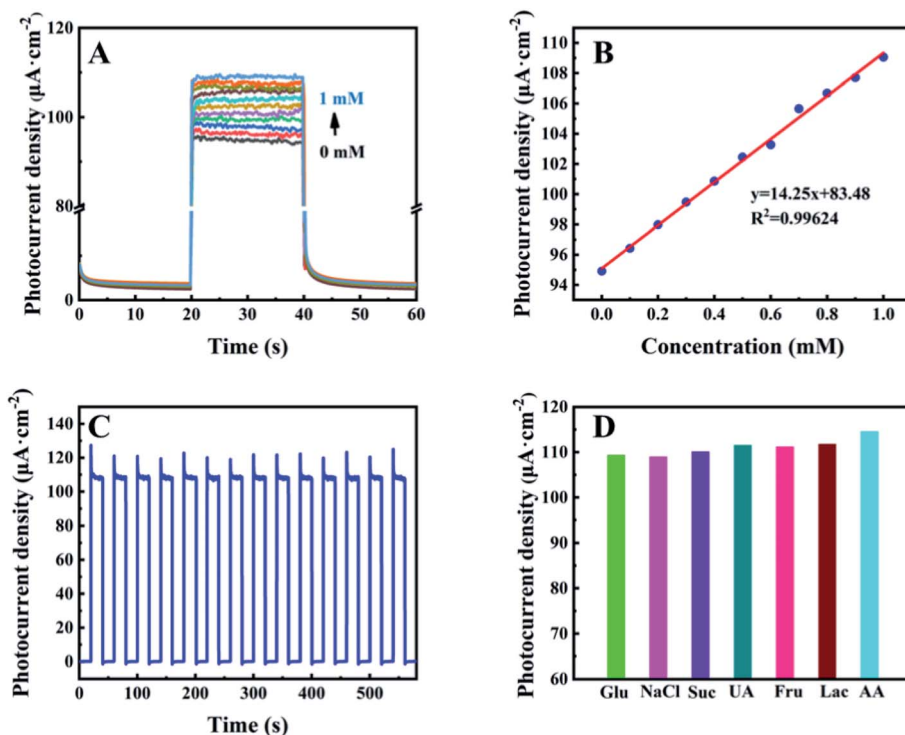


Fig. 5 (A) Photocurrent responses of the  $\text{TiO}_2$  NSs/BiOI NSs/GOx biosensor toward different concentrations of glucose from 0 mM to 1 mM in 0.1 M PBS (pH 7.4) electrolyte. (B) Linear calibration between glucose concentration and photocurrent density. (C) Stability under light conditions in the presence of 1 mM glucose. (D) Detection of the anti-interference performance of the electrode. The experiments were conducted in 0.1 M PBS (pH 7.4) at concentrations of 0.1 mM NaCl, sucrose (Suc), uric acid (UA), fructose (Fru), lactose (Lac), and ascorbic acid (AA) in the presence of 1 mM glucose.



Table 1 A comparison of the analytical characteristics of this sensor and other sensors used to detect glucose

GOx PEC biosensor	Linear range (mM)	Sensitivity ( $\mu\text{A mM}^{-1} \text{cm}^{-2}$ )	Ref.
GOx/SnS <sub>2</sub> /Nafion/GCE	0.025–1.10	7.6	25
Nafion/GOx/Ag-Pdop@CNT/GCE	0.050–1.10	3.1	26
GCE/GOx/Nafion/Br/TNT-AuNP	0.01–1.2	5.1	27
GOx/CNx-MWCNT/GCE	0.02–1.02	13	28
Nafion/GOx/PANI@GO/GCE	0.07–1.1	10.81	29
TiO <sub>2</sub> NSs/BiOI NSs/GOx	0.01–1.00	14.25	This work

used to compare the carrier density qualitatively. The measured data were brought into the formula and a linear fit applied, and the result is shown in Fig. 4E. The carrier density of the TiO<sub>2</sub> NS/BiOI NS electrode improved compared with the TiO<sub>2</sub> NS electrode. Moreover, the positive slope of the MS curve indicates that the prepared TiO<sub>2</sub> NS/BiOI NS electrode has n-type characteristics.

Electrochemical impedance spectroscopy (EIS) was performed to study the interface characteristics of the photoelectrodes and the dynamics of charge transfer. Fig. 4F shows that in the same frequency range, the prepared TiO<sub>2</sub> NS/BiOI NS composite electrode has a smaller radius, indicating that it has higher conductivity and photoelectric conversion efficiency.

Based on the good optoelectronic properties of the electrodes, the electrodes were constructed and tested for PEC enzymatic sensing under simulated sunlight and visible light (420 nm). Fig. 5A shows the glucose concentration gradient measured on the prepared TiO<sub>2</sub> NS/BiOI NS/GOx electrode. As the glucose concentration increased, the value of the photocurrent also rose, indicating a sensitive reaction to glucose concentration. The relationship between the glucose concentration and photocurrent density in simulated sunlight satisfies the equation  $J$  (photocurrent density) ( $\mu\text{A cm}^{-2}$ ) = 14.25[Glucose] (mM) + 83.48 (mM), and the statistically significant correlation coefficient is 0.99624 (Fig. 5B). Thus, the sensitivity of the biosensor under simulated sunlight is 14.25  $\mu\text{A mM}^{-1} \text{cm}^{-2}$ , the detection range for glucose is 0–1 mM, and the LOD (3S/N) is

0.01 mM. Fig. S3† shows that in the 0–0.1 mM range of glucose, the sensitivity of detection under visible light is 0.571  $\mu\text{A mM}^{-1} \text{cm}^{-2}$ .

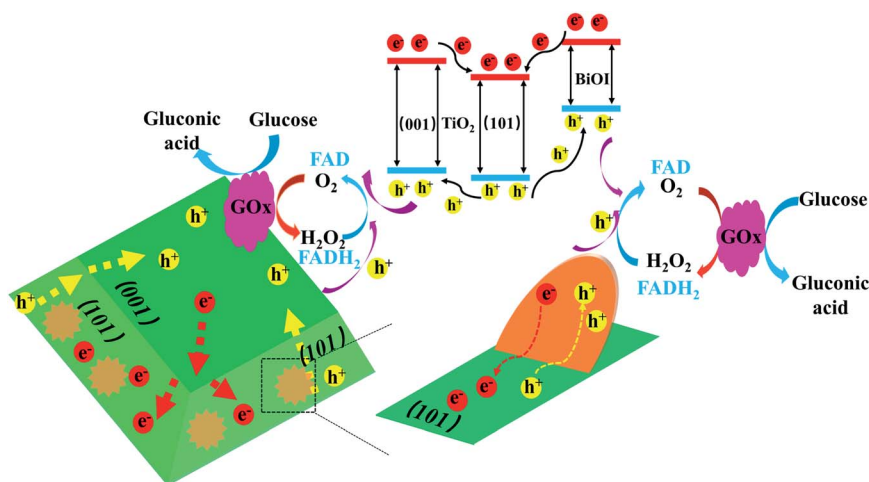
The stability of the TiO<sub>2</sub> NSs/BiOI NSs/GOx electrode was tested at 1 mM glucose by brief illumination for 600 s. Fig. 5C displays that the examined outcome shows that the photocurrent of the TiO<sub>2</sub> NSs/BiOI NSs/GOx electrode absolutely does not change with time.

Specificity and selectivity are important indices to judge the capability of a sensor. Possible coexisting and common molecules in the serum were selected as interfering substances, such as sucrose (Suc), fructose (Fru), uric acid (UA), ascorbic acid (AA), lactose (Lac), and NaCl to test the TiO<sub>2</sub> NSs/BiOI NSs/GOx biosensor. The results are shown in Fig. 5D. All kinds of interferers show no significant effect, indicating that the TiO<sub>2</sub> NSs/BiOI NSs/GOx biosensor has excellent anti-interference performance due to the selective catalysis of GOx fixed on TiO<sub>2</sub> NSs/BiOI NSs.

The performance of the PEC biosensor designed in this work was compared with some published glucose sensors, as listed in Table 1. The performance of the biosensor in this work on the two parameters of sensitivity and detection range is acceptable.

The satisfactory behavior of the PEC biosensor is attributed to the excellent photogenerated carrier separation mechanism shown in Scheme 2.

The TiO<sub>2</sub> NSs prepared in this paper expose a large proportion of (001) facets. The (101) and (001) facets of TiO<sub>2</sub> are identified as

Scheme 2 The manufacturing of the TiO<sub>2</sub> NS/BiOI NS/GOx biosensor and the PEC-sensing strategy for glucose detection.

electron-rich and hole-rich planes,<sup>30</sup> respectively. The photo-generated electrons generated on the (001) facets would migrate autonomously to the (101) facets. On the NSs, the (101) and (001) crystal facets were adjacent to each other and should have equal Fermi levels, so the two parts could form crystal facet heterojunctions,<sup>31,32</sup> which gives the TiO<sub>2</sub> NS single crystal material better photoelectric self-separation performance.<sup>33,34</sup> When BiOI NSs form a p–n heterogeneous structure with TiO<sub>2</sub> NSs, the Fermi level of BiOI and TiO<sub>2</sub> shifts until a new equilibrium is reached.<sup>35,36</sup> Under irradiation, the photogenerated electrons from TiO<sub>2</sub> and BiOI are gathered to TiO<sub>2</sub> (101) and exported through the FTO substrates to generate electrical signals. The photogenerated holes are transferred to TiO<sub>2</sub> (001) and BiOI to participate in the enzymatic reaction that oxidizes H<sub>2</sub>O<sub>2</sub> to O<sub>2</sub>. Thus, the TiO<sub>2</sub> NS/BiOI NS electrode could effectively promote the separation of the photogenerated electron–hole pairs and improve the PEC performance, which is crucial for achieving the outstanding analytical performance of the sensing platform.

## 4. Conclusions

A TiO<sub>2</sub> NS/BiOI NS photoelectrode was manufactured *via* hydrothermal and solvothermal methods on an FTO substrate, where the anatase TiO<sub>2</sub> film had a high percentage of (001) facets and BiOI NSs selectively grew on the TiO<sub>2</sub> (101) facets. Coupling with BiOI NSs broadened the absorption in the visible light region, extended the carrier lifetime, increased the photocurrent density, and significantly enhanced the PEC performance. The prepared TiO<sub>2</sub> NSs/BiOI NSs/GOx glucose biosensor under simulated sunlight achieved satisfactory results: the sensitivity was 14.25  $\mu\text{A mM}^{-1} \text{cm}^{-2}$ , the linear measurement range was 0–1 mM, and the LOD (3S/N) was 0.01 mM in PBS buffer at pH 7.4. In addition, the mechanism for the efficient separation of photogenerated carriers based on the crystal facet heterojunctions introduced in this paper could be used as a reference for the detection of other substances, such as nucleic acids, oxidases, and proteins.

## Conflicts of interest

The authors declare no competing financial interests.

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