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Recent progress in treatment of dyes wastewater using microbial-electro-Fenton technology

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Globally, textile dyeing and manufacturing are one of the largest industrial units releasing huge amount of wastewater (WW) with refractory compounds such as dyes and pigments. Currently, wastewater treatment has been viewed as an industrial opportunity for rejuvenating fresh water resources and it is highly required in water stressed countries. This comprehensive review highlights an overall concept and in-depth knowledge on integrated, cost-effective cross-disciplinary solutions for domestic and industrial (textile dyes) WW and for harnessing renewable energy. This basic concept entails parallel or sequential modes of treating two chemically different WW *i.e.*, domestic and industrial in the same system. In this case, contemporary advancement in MFC/MEC (METs) based systems towards Microbial-Electro-Fenton Technology (MEFT) revealed a substantial emerging scope and opportunity. Principally the said technology is based upon previously established anaerobic digestion and electro-chemical (photo/UV/Fenton) processes in the disciplines of microbial biotechnology and electro-chemistry. It holds an added advantage to all previously establish technologies in terms of treatment and energy efficiency, minimal toxicity and sludge waste, and environmental sustainable. This review typically described different dyes and their ultimate fate in environment and recently developed hierarchy of MEFS. It revealed detail mechanisms and degradation rate of dyes typically in cathodic Fenton system under batch and continuous modes of different MEF reactors. Moreover, it described cost-effectiveness of the said technology in terms of energy budget (production and consumption), and the limitations related to reactor fabrication cost and design for future upgradation to large scale application.

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1 Introduction

Textiles are included amongst the most pollution causing industrial sectors into air, water and terrestrial environments. Persistent organic pollutants (POPs) such as dyes and pigments are extensively manufactured and discharged ($5\text{--}1500\text{ mg L}^{-1}$) in textile industry effluents. Other industries including leather, foods, beverages, pharmaceuticals, cosmetics, paper and pulp are also releasing significant amounts of such pollutants in their effluent.^{1,2}

Developing countries are mostly devoid of any domestic or industrial wastewater treatment facilities, while those operating are often ineffective for refractory chemicals.³ Moreover, power shortages and cost of treatment facilities are one of the major drawbacks. Considering these facts the wastewater generally originating from industrial units is released in existing water channels (streams and rivers) after primary settlement tanks. The refractory contaminants stay in waster channels or settle in

soil and sediments depending upon their half-lives, causing enormous issue to freshwater ecology.

Dyes are highly diverse ($\sim 100\ 000$ types)⁴ xenobiotic and stable under ambient conditions of light and oxygen in natural ecosystems and conventional treatment plants. Such pollutants, due to their strong structural integrity and reactive nature, impose a varying degree of toxicity and mutagenicity to living systems, and create long term environmental issues.⁵ Physically, color due to dyes is aesthetically objectionable in water and more specifically red and purple as being unnatural create more concerns compared to blue, green and brown that are normally natural colors.⁶ Moreover, color prevents sunlight penetration in water,⁷ thereby limits gross ecological productivity (photosynthesis) of aquatic environment. Under these conditions, the natural processes can no longer compensate the bacterial consumption or decomposition of such and other contaminants resulting a stagnant aquatic environment.⁸ Chemically, these compounds affect the water chemistry and associated quality due to increasing load of pollutants indicators such as TDS, COD and BOD. Depleting oxygenic levels causes oxygenic stress that limits a great deal of respiratory physiology in living organisms. Whereas high levels of TDS results creates electric conductivity level that may add further toxicity. Such factors ultimately lead toward death and decay of aquatic flora and fauna.⁹

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Discharge of untreated dyes containing wastewater (WW) is highly objectionable and extensive treatments are required for environmental health and sustainability. Worldwide, various traditional and advance physico-chemical treatments¹⁰ methods including; adsorption,¹¹ ozonation, photo-catalysis (UV + H₂O₂), coagulation, electrochemical oxidation, and filtration have been used for textile dyes and WW from industries. Coagulation and adsorption have been the simplest and commonly used methods.¹¹ Irradiation is very efficient but unsuccessful against photo stable dyes.^{12,13} Membrane filtration is highly proficient though need sophisticated nano or ultra-filtration membranes.¹² Chemical or electro-chemical methods including ozonation Fenton process or Electro-Fenton process (EF), and advance oxidation process (AOPs) showed great prospects for degradation of both soluble and insoluble dyes, however, these treatment are either costly or facing issues of sludge waste management^{12,14–18}. Moreover, both chemical and physical methods have been reported for the removal of only 20–30% dyes from WW.¹⁹ Biological wastewater treatments include an array of living organisms (bacteria, fungi and plants) integrated into different suspended and attached growth reactors. They are considered ultimate due to diverse physiological capabilities of organisms to be operationalized both under aerobic and anaerobic conditions for complete decomposition of dyes. Still they are slow to evolve and functionalize until given additional technical or operational support.^{20,21} Smart application of biological process may also lead to re-innovating useful products from organic waste (renewable energy & industrial products). Moreover, they are environmental friendly, cost-effective and highly sustainable.

In the last two decades considerable advancements have been made on multifaceted microbial electrochemical technologies (METs). Typically, microbial fuel cell (MFC) and microbial electrochemical cell (MEC) have been mostly developed for treatment of variety of pollutants linked to domestic and industrial sectors.^{9,22,23} METs principally work on the anaerobic bio-catalysis of contaminants for harnessing renewable energy [electrical²⁴ or gaseous energy (H₂)].⁷ Recently, advancement on MXCs as Micro (Bio-)electro-Fenton process (MEFP) is taking lead relying on the previously established WWT technologies typically including EFP, AOP, biological processes and MES.^{15,18} MEFS using MFC or MECs have gone through a series of innovations and advancements.^{25–27} In 2000, Lin and Chang, reported a combine sequential approach of treating landfill leachate and refractory compounds using EF and biological processes²⁸ Later, in 2009, the 1st MEFS was developed for aerobically stable pollutant like dyes and it further expanded the scope of such innovative technologies.^{27,29–33} As compared to traditional EF technologies, the MEF technology is more efficiency and cost-effective (approx. 10 fold) for refractory compounds in WWT.²¹ Besides it creates almost negligible amount of sludge. This technology offer superior possibilities of treating persistent organic pollutants (POPs) of xenobiotic nature in WW^{34,35} including dyes,^{32,36,37} pesticides, dioxins, personal care products and pharmaceuticals^{24,38,39} lignin (paper and pulp), bagasse and molasses from different industries. Besides, it is equally effective for municipal and

agriculture waste treatment under varying physicochemical loads.^{32,40}

A basic MEFS include a similar METs set up²¹ for anaerobic degradation of biodegradable compounds in anodic chamber, however, it is also supplemented with a Fenton Process (FP) for simultaneous treatment of refractory compounds (dyes, pesticides or aromatic compounds) in a non-selective manner in cathodic chamber.³⁴ Cathodic oxidation of contaminants involves oxygen species as hydroxyl radicals that are highly reactive and generated through catalysis of H₂O₂ by Fe²⁺.^{10,41,42} By Fenton's reagent, oxidation of organic compounds is rapid and exothermic process that helps almost complete mineralization of contaminants into inorganics *i.e.*, carbon dioxide and water.²⁹ Additionally reports have also suggested possibilities of coupling photo-catalytic (UV/light) oxidation along with typical FP to enhance the overall efficacy of MEFS.²¹ An added advantage of this innovative technology is treatment of two different wastewaters (pollutants) in the same reactor. In the last few years, further improvement in architecture and scope of MEFS helped development of new reactors that are yet to be operationalized at pilot and large scales.

2 Dyes and pigments; characteristics, source and environmental fate

2.1 Characteristics of dyes and pigments

Worldwide, an annual production of various dyes and pigments is about 700 000 tons.⁴ Generally dyes and pigments are made up aromatic backbone structure of benzene with alternating double C bonds (delocalized electrons pairs) that helps in transmission and reflectance of natural white light resulting in creation of different hues.¹ From an industrial perspective, successful dyes have a strong structural integrity and high water solubility for possible sustainable application in different products.⁸ On the other hand, pigments are made hydrophobic with large molecular structures and intermolecular attractions due to hydrogen bonding (N–H and C=O groups).⁴³ Globally, 40% of colorants used comprise chlorine that is organically-bound.⁶ With their widespread applications and complicated chemistry, the utilization of colorants in textile industry could also be a major cause of dioxins and their precursors that typically relates to POPs. Being the most toxic compound, dioxins are associated to halogens homologues. The recalcitrant nature of dyes and pigments is typically associated with their benzoic structure that makes them highly objectionable as they avoid natural process of mineralization under normal oxygenic environment⁴⁴ thereby creating environmental issues of low biotic productivity. The reactive nature of such compounds due to presence (*e.g.*) chloro, nitro, and sulphonic groups make them toxic and mutagenic at varying degrees at different trophic levels.

The basic structure of dyes and related compounds comprises a chromophore group represented by “the azo group (–N=N–), ethylene group (=C=C=), methine group (–CH=), carbonyl group (=C=O), carbon-nitrogen (=C=NH; –CH=N–), carbon-sulphur (=C=S; ≡CS–S–C≡), nitro (–NO₂; –NO–OH),



nitroso ($-N=O$; $=N-OH$) or chinoid groups. The auxochrome groups are ionizable groups that confer to the dyes the binding capacity onto the textile or any material. The usual auxochrome groups are: $-NH_2$ (amino), $-COOH$ (carboxyl), $-SO_3H$ (sulphonate) and $-OH$ (hydroxyl).⁴⁵ Overall, the major classes include; azo or anthraquinone accounting for 65–75% in textile dyes. An about 2/3 of them are azoic in nature. The precursor compounds or intermediates are aniline, chloroanilines, naphthylamines, methylanilines, benzidines, phenyldiamines, and others.⁶ Azo dyes are characterized by the reactive groups that make covalent bonds with $HO-$, $HN-$, and $HS-$ groups present in fibers *e.g.* cotton, silk, nylon, wool. These dyes are frequently utilized for the yellow, red and orange colors.

2.2 Classification of the dyes

Environmental positioning of dyes is related to; C atoms number and the aromatic structure, number and nature of substituents, and overall backbone molecular structure.⁶ Typically, classification of dyes and pigments is either based upon their chemical structure or their mode of applications to different substrate products. Structure based classification is related to the nature and chromophore groups that helps predicting dyeing and associated oxidation – reduction. There are 8 different categories of dyes based upon their chemical structures. Dyes based upon application depends on chromophore grouping that help dyeing based upon dye solubility in dye bath and its affinity with different fibers and nature of fixation. On solubility basis, dyes and pigments are broadly characterized as anionic, nonionic and cationic in nature.⁴⁶ Among anionic dyes (direct, acid and reactive) reactive and acidic are most problematic due to their bright colors and water solubility in conventional wastewater treatments.⁶ Nonionic dyes such as disperse do not ionize in the aqueous system, while, cationic ones are anthraquinone, azoic, reactive and disperse dyes. Few disperse dyes are efficiently bioaccumulate and their azo and nitro groups are reductively cleaved (reduced) in soil sediments to toxic aromatic amines.⁶ On this basis, they are 9 different types partially distributed in the two categories of water soluble and insoluble. Some basic details on classification of dyes is mentioned in Table 1.

2.3 Source and environmental fate of dyes and pigment

Industrial units including textile, pulp/paper, leather tanning, dyestuff manufacturing, pharmaceutical and kraft bleaching industries have been reported as major point sources contaminating dyestuffs in water sources.⁶ Typically dyes are released in water ways from dyebath (dyeing unit) and dyes manufacturing units of industries.⁵⁷ The annual estimated load of pollutants in textile wastewater varies from 200 000–250 000 t salts. The exhaustion rate of dyes and pigments from dyebath unit is 10–60% comprising 10–50% reactive dyes, 5–20% vat dyes, 5–30% direct dyes and 10–40% sulphur dyes and that is 280 000 tons per year.¹ Release of raw or partially treated effluents adds a huge load of TOC, nitrites, nitrates, phosphates and heavy metal ions (zinc, iron, mercury, lead, chromium, cobalt and copper) thereby also creating an issue of eutrophication.

Typically, textile dyes are highly relevant in ecological perspective. Among them, azo are taken as model dyes due to their large production, consumption (60–70%) and discharge in the hydrosphere. Effluent of leather dyeing may contain greater than 250 ppm dyes causing an about 7000 ppm COD load. It is highly acidic (pH 4.5 approx)⁵⁸ although alkaline in the case of the textile industry. Effluents from these industries are also source of dioxins because of the presence of dioxazin and anthraquinone dyes and pigments made from chloranil (tetrachloro benzoquinones) and pentachlorophenol. Dioxins are generally released during manufacturing and treatment of dyes by photolysis/UV.⁵⁹

Long time persistence (*i.e.* half-life = several years/2 – 13 years) of dyes accumulate them in abiotic sediments or living or dead biomasses particularly in fish and other aquatic forms of life. Mostly they are soluble still they tend to biosorb or absorb and accumulation in suspended and settled abiotic and abiotic components of water bodies. With solubility greater than 2000 mg L⁻¹ in water, bioaccumulation is not estimated for dyes. Adsorption is high in basic and direct dyes while range is from high to medium in disperse dyes, and it depends on degree of sulphonation or ease of hydrolysis. In group 1, ability to adsorb on the biomass and water solubility is increased due to higher levels of sulphonation. In Group 2, dyes are also greatly sulphonated but indorsed good adsorption performance on sludge moderately. Additional information in bio-elimination of different reactive dyes stated that disazos, anthraquinones, triphenyldioxazines and phthalocyanines are usually better adsorbed than monoazos. It is crucial to highlight that toxic compounds *e.g.* benzidine, aromatic amines and its derivatives can be produced in environment *via* transformation of textile dye precursors such as by reduction or hydrolysis of azo dyes. Moreover, sequential bioaccumulation in entire food chain⁶⁰ and later biomagnified at higher trophic levels cause lethal effects.

As dyes are photo-chemically stable under ambient environmental temperature, so, they normal create problem in conventional industrial and residential treatments systems.⁶¹ Dyes are highly polar (log K_{ow} up to 3) and shows recalcitrance also depending on environmental variables such as redox milieu or pH. A compound like aniline (1st synthetic known dye) has been found easily degradable under aerobic condition; however, it is highly stable under anoxic condition.⁶² The recalcitrant and xenobiotic nature of dyes usually influence the structure and function of aquatic ecology.⁵⁷ Additionally these compounds also inflict a negative effect on microbial communities of the soil and germination of plants. Typically, living species at the higher trophic levels of the food web are exposed to a thousand times higher concentration of toxicants than those at lower levels.⁸ Ecological and Toxicological Association of Dyes (ETAD) reported that 98% of dyes have LC50 value higher than 1 mg L⁻¹ for fishes and 59% of dyes have LC50 greater than 100 mg L⁻¹ and 28% higher than 500 mg L⁻¹. LD₅₀ reported for most of the azo dyes varies between 100–2000 mg kg⁻¹ body weight.⁶² They affect physiological status of biotic community and in England, it was reported that inhibition of respiration rate in sewage bacteria is caused by 18% of 200 dyes.





Table 1 Classification and properties of Dyes

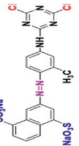
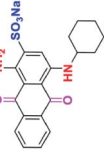
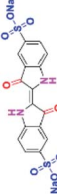
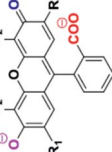
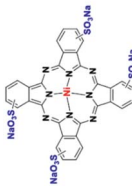
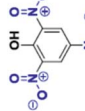
Category (structure and mode of application)	Sub-category (type/nature)	Chromophore	Auxochrome	Properties/examples	Discharge rate (%)	Fabric type	Structure and chemical formula	References
Chemical structure	Azo	-N=N-	NH ₂ , OH, SO ₃ , Cl	70% of synthetic dyes, mon, di, tri azo. Yellow reactive 4, black reactive 5	5–10	Cellulose	 Yellow reactive 4 C ₂₀ H ₁₂ -C ₁₂ N ₆ Na ₂ O ₆ S	47
	Anthraquinone	C=O, NH ₂ , and =C=C=, forming an anthraquinone complex (quinone nucleus)	NH ₂ , OH, SO ₃	Most important after azo, derived from anthracene. <i>e.g.</i> ; acid blue 62, reactive blue 19	2–10		 Acid blue 62 C ₂₀ H ₁₉ N ₂ NaO ₅ S	46
	Indigo	C=O, NH ₂ , C=C, C-O, C=N, C-C	SO ₃ , NH ₂ , C=O	Derived from indigo. <i>e.g.</i> ; blue acid 74, indigo blue			 Blue acid 74 C ₁₆ H ₈ N ₂ Na ₂ O ₈ S ₂	48
	Xanthene	Xanthylum or di-benzo-g-pyran nucleus	Amino or hydroxyl, COOH	Intense fluorescence, used as markers/tracers in maritime accidents or underground river. <i>e.g.</i> ; fluorescein, eosin, erythrosine			 Erythrosine C ₂₀ H ₆ Na ₂ O ₅	49
	Phthalocyanine	Phthalocyanine nucleus	SO ₃	Metal complex (Cu, Ni, Co, Pt <i>e.g.</i> ; copper phthalocyanine. <i>e.g.</i> ; pigment blue 15/3, nickel(II) tetrasulfonic acid			 Nickel (II) tetrasulfonic acid C ₃₂ H ₁₂ N ₈ Na ₄ NiO ₁₂ S ₄	32
Chemical structure	Nitrated & nitrosated	O-NH=O, O-NH=O, phenol, nitro	Nitro (-NO ₂), OH, NH ₂ , CH ₃ NH	Limited number, older <i>e.g.</i> ; picric acid (2, 4, 6-trinitrophenol), 2-nitrophenol, 2-amino-4-nitrophenol			 Picric acid (2, 4, 6-trinitrophenol) C ₆ H ₃ N ₃ O ₇	50



Table 1 (Contd.)

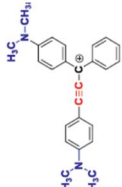
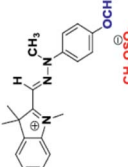
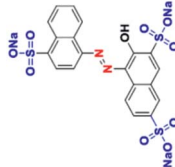
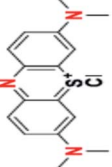
Category (structure and mode of application)	Sub-category (type/nature)	Chromophore	Auxochrome	Properties/examples	Discharge rate (%)	Fabric type	Structure and chemical formula	References
Application	Diphenylmethane & triphenylmethane		CH ₃ , NH, SO ₃	Oldest, derivative of auramine and triphenylmethane <i>e.g.</i> ; fuchsin and malachite green			 p -Dimethylamino-phenylethynyl C ₂₃ H ₂₅ C ₁ N ₂	51
	Polymethinic	=HC-HC=	OH, SO ₃ ⁻ , CH ₃ , HN-N-CH ₃	Also called cyanines, <i>e.g.</i> ; basic yellow 28, polymethine dye 2630			 Basic yellow 28 C ₂₁ H ₂₇ N ₃ O ₃ S	52
	Acid or anionic	Azo, anthraquinone or triaryl	SO ₃ ⁻ , NH ₂ , OH	For wool, polyamide, silk and acrylic, <i>e.g.</i> ; red acid 27, acid blue 90, acid blue 74	7–20	Wool & nylon	 Red acid 27 C ₂₀ H ₁₁ N ₂ Na ₃ O ₁₀ S	53
H ₂ O soluble	Basic or cationic	Diaryl, triaryl, anthraquinone, azo, phthalocyanine	NH ₂	<i>e.g.</i> ; basic blue 9, basic yellow 37, blue nile	2–3	Acrylic	 Basic blue 9 C ₁₆ H ₁₈ C ₁ N ₃ S ₃ H ₂ O	54
	Metalliferous	Azo, phthalocyanine		Acidic dye with metals (Cr, Cu, Ni and Co)	2–10	Leather, finishing, stationery, printing, inks, coloring for metals, plastic		32



Table 1 (Contd.)

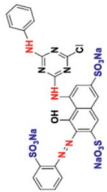
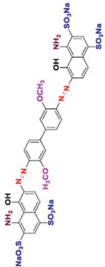
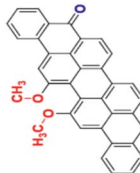
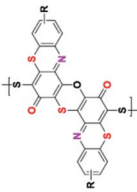
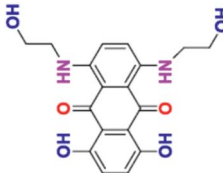
Category (structure and mode of application)	Sub-category (type/nature)	Chromophore	Auxochrome	Properties/examples	Discharge rate (%)	Fabric type	Structure and chemical formula	References
H ₂ O insoluble	Reactive	Azo, anthraquinone, and phthalocyanine	SO ₃ , NH ₂	<i>e.g.</i> ; reactive red 198	10–50	Wool and nylon	 Red cibacrom 3 C ₃₂ H ₂₃ C ₁ N ₈ O ₁₄ S ₄	55
	Direct or substantive	Azo, phthalocyanine	NH ₂ , SO ₃ , OH	Large molecules with positive and negative charges, affinity to cellulose, wide variety, easy to apply and low cost, <i>e.g.</i> ; direct blue 1	5–30	Cellulose	 Direct blue 1 C ₃₄ H ₂₄ N ₆ Na ₄ O ₁₆ S ₄	8
	Vat dyes	Anthraquinone	NH ₂ , C=O, CH ₃	Good resistant to degradation (soap and sunlight), good affinity to cotton, linen, wool, silk, rayon, use for dyeing jeans. <i>e.g.</i> ; indigo (vat blue 1), blue indanthrene RS (vat blue 4, vat green 1	5–20	Cellulose	 Vat green 1 C ₃₆ H ₂₀ O ₄	56
Sulfur	Sulfur	Thiazoles, thiazine, thianthrene, and phenothiazonethioanthrone	NH ₂ , OH, SO ₃	Like vat dyes, complex high MW, used for cotton, dark shades, <i>e.g.</i> ; sulfur black, sulfur blue 15	30–40	Cellulose	 Sulfur black C ₁₈ H ₈ N ₄ O ₅ S ₂	8
	Disperse or dispersible	Anthraquinone	NH ₂ , OH	Plastosolubles, stable at high temperature, used in polyester and polyamide, <i>e.g.</i> ; blue disperse 7, red disperse 60	2–20	Synthetic	 Blue disperse 7 C ₁₈ H ₁₈ N ₂ O ₆	8



Table 1 (Contd.)

Category (structure and mode of application)	Sub-category (type/nature)	Chromophore	Auxochrome	Properties/examples	Discharge rate (%)	Fabric type	Structure and chemical formula	References
Pigments				Organic (benzoic), inorganic (metals; Ti, Zn, Ba, Pb, Fe, Mo, antimony, zirconium Ca, Al, Mg, Cd, Cr), kept in suspension tanks to dispersants, film formation by heating, used in printing		Paint, ink, plastic, fabric, cosmetics, food, and other materials		43

Dyes generally composed of benzene moieties that makes them recalcitrant (xenobiotic nature) toxigenic and mutagenic.⁶³ Additionally, certain functional groups such as halogens (chloride), amide, amino, sulphides and metals may add further toxicity and mutagenicity at different trophic levels.^{61,64} They may also carry precursors of toxic compounds like dioxins that as by-products produced during the synthesis of chlorobenzenes, chlorophenols, polyvinyl chloride, chlorobiphenyls, pigments, dyes, and printing inks. They can even be toxic at 1 mg L⁻¹ concentration in effluents so that they are related to environmental deterioration and various diseases in living beings.^{61,65} Despite recalcitrance they are potentially degradable or transformable in anoxic environmental condition and soil sediments, releasing sometime toxic compounds. Acute toxicity to textile dyes relates to ingestion and inhalation,⁶⁶ causing bladder cancer,⁶⁷ dermatitis, nervous disorders⁶⁵ impaired enzymatic activities,⁶⁸ eliciting irritations to the eyes and skin.⁶⁷ The personnel handling and producing reactive dyes may have contact rhinitis, allergic conjunctivitis, dermatitis, asthma or other allergic reactions.^{67,69}

This persistence nature of dyes has also been closely related to their reactivity. It increases with the number and types of electron donating substituents typically halogens, nitro, sulphonic and alkyl groups.⁸ Besides, substituents in *ortho* and *para* position further upsurge the carcinogenic potential. Essentially, the toxicity lessened by the protonation of aminic groups.⁶ Highly reactive nature of dyes and aromatic products makes them potential candidates to transform biomolecules at very minor to major levels in the living bodies. Such changes in the living biomolecular structure impair the physiological status of living organism that becomes sever under hypoxic conditions in aquatic sediments or living biomass. Under extreme situations azo dyes may leads to DNA damage that can result in mutation and cancerous growth and sometime lethal effects like death. A substantial decline in tadpole survivorship was witnessed at 209 µg g⁻¹ while a major upsurge in malformations at two peak concentrations was tested in sediment.⁷⁰ Especially in the case of azo dyes, carcinogenicity can be produced by both the dye itself and its own metabolized compounds.⁶⁷ Few well-known azo dyes *i.e.* azodisalicylate, precursor for 4-phenylenediamine; Direct Black 38, precursor for benzidine and their derivatives are benzidine and its derivatives, and also higher number of anilines *i.e.* 4-chloroaniline, 2-nitroaniline, 4-phenylenediamine, 4,4'-dimethylendianiline, dimethylamines, nitrosamines *etc.* causing cancer in humans and animals.

At genotoxicity level, large concentrations of dye (DY7) in sediment prompted cellular stress-related gene transcription and influenced the genes associated with chromosome condensation, necrotic cell death, and mRNA processing.^{70,71} Dyes may⁶⁷ create chromosomal aberrations other times mutagenic.⁶⁹ Dyes like Azure B found to be affecting helical structure of DNA^{67,72} and duplex RNA.⁷³ Whereas at cytotoxic level, they inhibition enzymes (monoamine oxidase A) (MAO-A),⁷⁴ involved in nervous system⁷⁵ related to human behavior.⁷⁶ Similarly, inhibition of glutathione reductase⁷⁷ disturbed redox homeostasis cellular.⁷⁸ Disperse red 1 dye has been reported exhibiting

mutagenic potential⁷⁹ on human lymphocyte and hepatoma (HepG2) cells, hepatocyte imitative cells,⁸⁰ causing occurrence of micronuclei,⁸¹ indicated mutagenicity at chromosome level.⁸² Besides, it caused DNA adducts⁸³ a key cancer causing event in *Salmonella* spp.⁸⁴ Similarly, disperse orange 1 exhibited mutagenicity⁷⁹ causing DNA damage—a base pair substitution and frameshift mutations in *Salmonella* spp. Moreover, it caused cytotoxicity with apoptosis in HepG2 cells. Another dye, Sudan I (solvent yellow 14),⁸⁵ although illegal, used as a food supplement in paprika, has enzymatically transformed into carcinogenic aromatic amines by intestinal microbiota. In rats, the presence of Sudan I dye is confirmed by neoplastic liver nodules. Basic red 9 a commonly used dye in textile, leather, paper and ink industries⁸⁶ indicated carcinogenic potential in human and environmental⁸⁷ after partially degradation into carcinogenic aromatic amines in anaerobic conditions. Disposal of such compounds in water bodies showed potential for allergic reactions, skin irritation, mutations and cancer. Besides they comprise local sarcomas and tumors in the liver, bladder,⁸⁸ mammary glands and hematopoietic system. Crystal violet, a cationic dye (triphenylmethane group),⁸⁹ caused mitotic dysfunction and suspension at metaphases⁹⁰ and inducing disruption of chromosomes in Chinese hamster ovules. Crystal violet has also been reported promoting fish tumors, and hepatocarcinoma, reticular cell sarcoma in vagina, uterus, ovary and bladder, hardened gland adenoma and ovarian atrophy in rats. Besides, it caused cystitis, irritation of the skin and digestive tract, respiratory and renal failure.⁹⁰

3 Degradation mechanism of dyes and organic compounds in MEFS

MEFS is an integration of biological (microbial) degradation with that of electrochemical process *i.e.* Fenton process, for efficient removal of pollutants in less time. In a typical Microbial-electro-Fenton processes using a two chamber MFC, electrons originating from anaerobic bio-catalysis (oxidation) of organics and inorganics follows an anodic pathway reach the cathode to combine to reduce (2 electron) O_2 to form H_2O_2 (79

to 196 $mg\ L^{-1}$) with simultaneous electricity production in 1st stage. Then it follows the production of highly reactive (oxidative) OH radical through chemical catalysis (Fe) of H_2O_2 in the 2nd stage. The enhanced oxidation of the target compounds such as dyes through hydroxyl radical is the final 3rd stage. MEC, an extended version of MFCs, requires a minor input of electricity (0.2–0.8 V) in order to accelerate overall performance and H_2O_2 production (1300–1447 $mg\ L^{-1}$). A part of free energy formed is taken by bacteria for catabolic activities and rest can be employed to produce electricity for maintainable operations of the system.^{91–94} Such systems can equally be good to treat two different dyes and their byproducts at 1st and 3rd operational stages simultaneously. Another good possibility can be sequential treatment (back and forth) of same refractory dyes in the two chambers of reactor in order to achieve complete mineralization as shown in Fig. 1. Beside these systems creates a workable electric potential to facilitate reactor performance and make them cost-effective and sustainable. A biological catalyst is either an enzyme or bacteria immobilized on the anodic reactions. Although the cathodic reactions may involve a biological or chemical catalyst in a typical MFC.^{40,95,96} For advanced oxidation of refractory compounds in cathodic chamber, a chemical catalyst like Fe^{2+} is utilized to generate OH radical from H_2O_2 .

Microbial electrochemical systems depend on diverse bacterial communities that encompasses electricigens or ARB. In the anode compartment, a mutualistic activities of several anaerobic microorganisms (fermentative, syntrophic, acetogenic, and methanogenic) consequent in the degradation of complicated organic substances into simple and stabilized compounds, primarily, methane and CO_2 . Initially such systems often operated with glucose and acetate as a fermentable substrate to upkeep the growth of electricigens. The fermenters and other syntrophic bacteria perform hydrolytic monomerization and fermentation of complicated substrates^{97,98} from food, beverages, pulp and paper, leather, pharmaceutical and residential wastewater. The enrichment and catalysis role of fermenters helps improved oxidation rates of monomers by electricigens in the secondary stage.⁹⁹ It also work to ease the

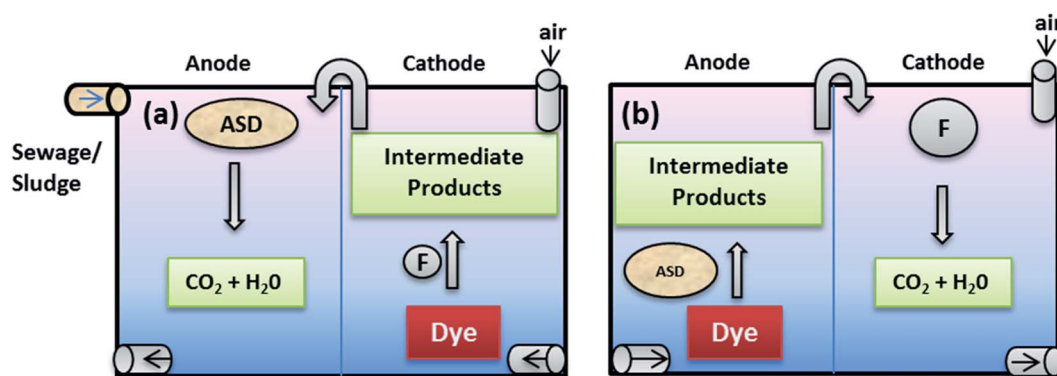


Fig. 1 Model concept of MXCs based MEFS: sequential batch or continuous modes; (a) aerobic degradation of dyes in cathode through Fenton reaction (FR) followed by anaerobic mineralization of dyes products and sewage/sludge in anode, (b) anaerobic degradation of dyes and sewage/sludge in anode followed by aerobic mineralization of products of dyes and sewage/sludge in cathode through FR (ASD = anaerobic sewage sludge digestion; F = Fenton agent 'OH).

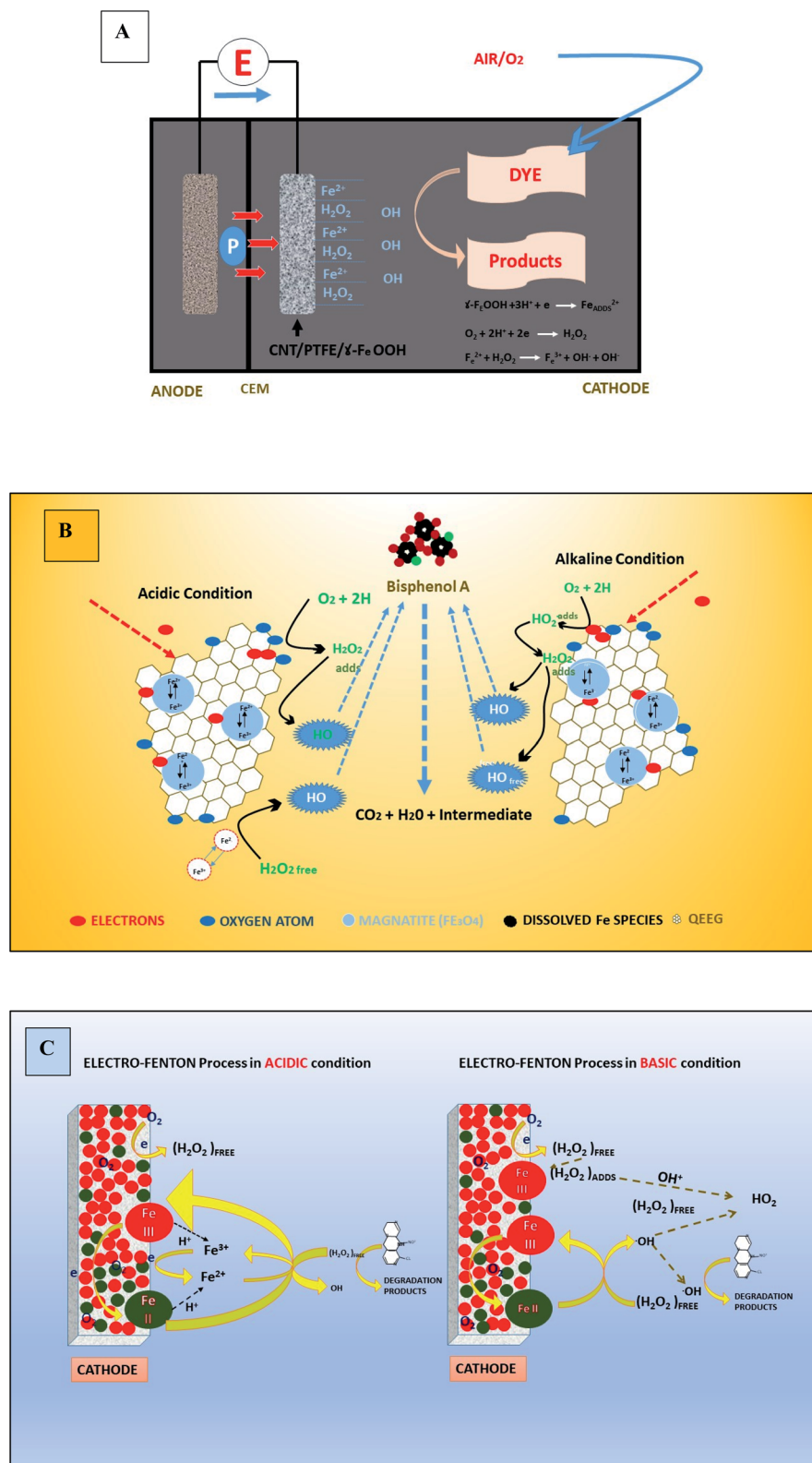


Fig. 2 Cathodic electrochemical Fenton reaction: (A) a typical MFC; mechanisms of reactive oxygen species formation under acid and alkaline conditions (B) the graphene/Fe₃O₄ composite electrode (refabricated from ref. 161) (C) Fe₃O₄@Fe₂O₃/ACA cathode (refabricated from ref. 162).

role of hydrogen scavengers.¹⁰⁰ Generally, anaerobic digestion (AD) involves liquefaction or hydrolysis of insoluble compounds,¹⁰¹ followed by acidogenesis/acetogenesis and

methanogenesis.¹⁰² Bacteria like *Streptococcus* and *Enterobacterium* release extracellular hydrolytic enzymes *e.g.* cellobiase, xylanase, cellulase, amylase, protease, and lipase to degrade



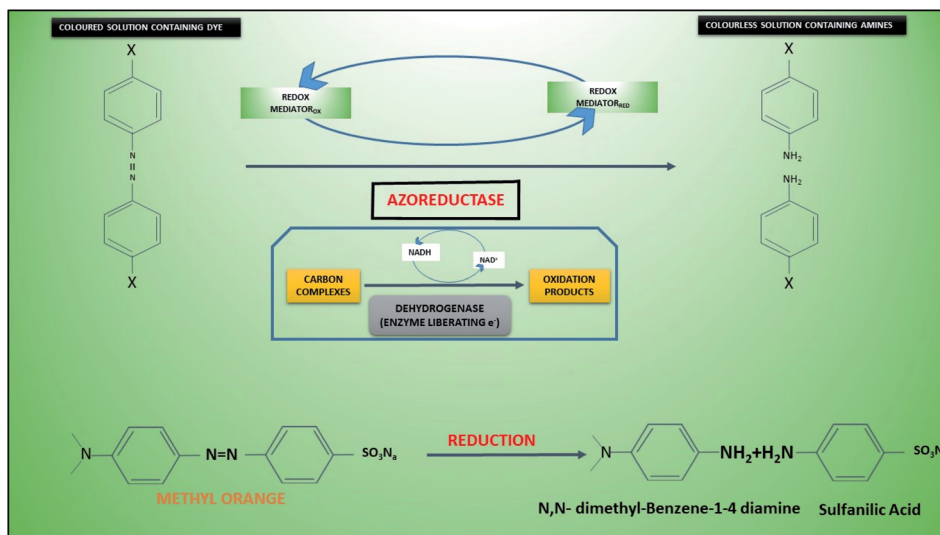


Fig. 3 Proposed scheme of partial anaerobic degradation (decolorization) of methyl orange 7 (refabricated from ref. 169).

biopolymers¹⁰³ into simple monomers and dimers *i.e.* monosaccharides, fatty acids and amino acids.^{104–107} In the 2nd stage, monomeric products are then transformed into “short-chain organic acids (formic, acetic, propionic, butyric, and pentanoic) alcohols (methanol, ethanol), aldehydes, carbon dioxide, and hydrogen by acidifying bacteria including *Pseudomonas*, *Bacillus*, *Clostridium*, *Micrococcus*, or *Flavobacterium*”.^{105,108,109} In the 3rd stage, acetogens including *Syntrophomonas* and *Syntrophobacter* transform the acid phase products into acetates and hydrogen that could be utilized by methanogens¹¹⁰ in syntrophic relation with acetogenic bacteria.^{110,111} *Methanosarcina* and *Methanosaeta*, acetogenic methanogens, generate methane that accounts an about 2/3 in AD in the 4th stage.^{112,113} Methylated compounds and Carbon dioxide are the alternate substrates for production of methane by *Methanosarcina*, *Methanothermobacter*, and hydrogenotrophic methanogens. Whereas, methylated compounds are appropriate substrates for *Methanospiraera*, *Methanosarcina* and methylotrophic methanogens.¹¹³ Overall, biogas production during AD comprises 65–70% methane and 30–35% carbon dioxide.¹¹⁴

Typically MEFS based reactors demonstrated enrichment of diverse ABR bacterial community in anodic biofilms.¹¹⁵ *Proteobacteria* has been reported most abundant phylum under these setups due to its versatile abilities for aromatic compounds (azo dyes) biodegradation, electricity generation, and fermentation.¹¹⁶ Functionality based analysis showed chemoheterotrophy the predominant followed by fermentation (6.6 ± 1.6%) and nitrate reduction (7.4 ± 2.3%). Two distinctive electricity producing bacteria *i.e.* *Shewanella* and *Geobacter*,¹¹⁷ were anticipated to be the key players in the improvement of iron respiration.^{118,119} Other bacteria with selective relevant function include *Serratia*, *Stenotrophomonas* for EET, *Stenotrophomonas Dysgonomonas*, *Brevundimonas*, and *Achromobacter* for reduction of azo-dye or degradation of aromatic compounds.^{120–122} The relative abundances of fermenters

Acinetobacter, *Cloacibacillus* and methanogens *Methanobacterium* were lowered under MES systems.^{123–125}

Compound like azo dyes from dyeing and production sectors of industries account for an about 60% of the total known dyes. Usually they are taken as model compounds to be treated in any biological, physical or chemical treatment. Azo dyes have shown partial to complete mineralization in most of the MEFS.²⁰ Still, most of the previous studies have reported microbial-electro-Fenton process achieving complete degradation of azo dyes including amaranth, orange G, methylene blue, crystal violet, Lissamine green B, and orange II by integrating effective electro-Fenton reaction in the cathode chamber.^{126–128} Generally, for azo dyes removal two different types of MEFS such as MFC or MEC and their modified version have been used. Azo dyes due to their charged large structures are expected to be reduced either extracellularly or with membrane bounded enzymes. Under anaerobic condition azo dyes cleavage at N=N to produce colorless aromatic amines. Anodic catalysis in MFC using microbial consortia broadens the horizon of substrates utilization due to diverse microbial metabolic apparatus in terms of enzymes, coenzymes and cofactors this helps improves the energy efficiency and less toxicity in the system.^{129–131}

During anodic biocatalysis of dyes the role of oxidoreductases (dehydrogenases (alcohol, formaldehyde, formate, fructose) oxidases, oxido-reductases), cofactors and coenzymes “(nicotinamide adenine dinucleotide (NAD⁺), nicotinamide adenine dinucleotide phosphate (NADP⁺), pyrroloquinoline quinine (PQQ), Flavin adenine dinucleotide (FAD) have always been vital (Fig. 3). In case of FAD cofactor dependent oxidoreductases, three FAD-dependent glucose dehydrogenase (FAD-GDH), cellobiose dehydrogenase (CDH) and fructose dehydrogenase (FDH) have been extensively studied”.¹³² Additionally, the role of natural and artificial mediators (methylene green, methylene blue, neutral red,¹³³ phenazines, methylene blue, alizarin yellow, methyl violet, thionine, Prussian blue, azure A, toluidine blue and azure C) that reacts with reduced enzymes or



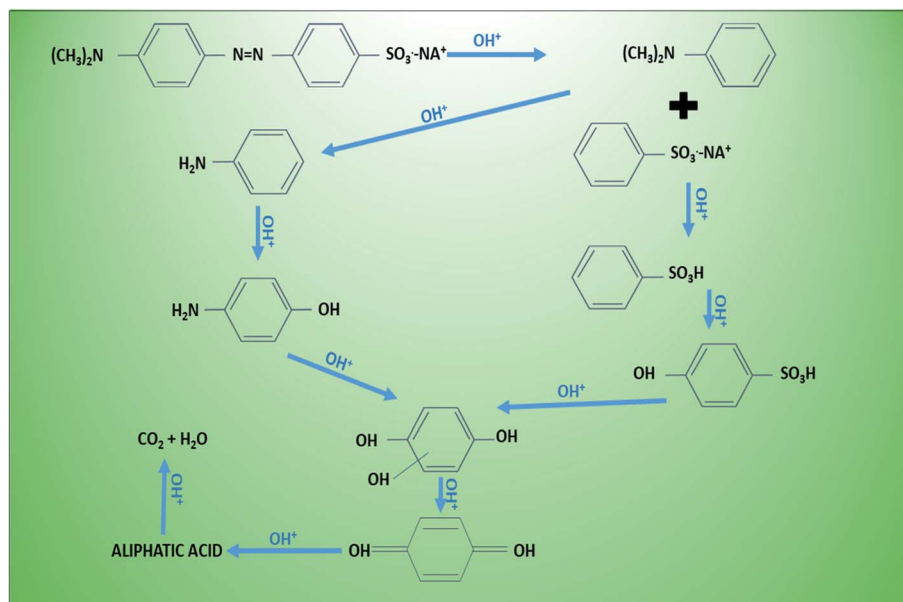


Fig. 4 Degradation of methyl orange using hybrid advanced oxidation (refabricated from ref. 170).

coenzyme in order to transfer electrons to electrode is highly effective.^{134–137} Moreover, the degradation products of dye (AO7) such as naphthol have been reported as an alternative for electron mediation.¹³⁸ On the other hand, the electrode can also reduce enzyme thereby enzymatic reduction of substrate *i.e.* dye (bioelectrocatalytic reduction). Direct enzymatic reduction of electrode proved to be more effective, however,^{139,140} if cofactors are involved then they should not be 20 Å (2 nm) away from electrode surface.^{141,142}

Metals and their alloys have also been reported playing a major role as catalyst due to their different oxidation state in Microbial-electrochemical Fenton system. These metals include Pt, Au, Ni, Co¹⁴³ Fe and Cr. Ti, V, Cr, Mg, Mo, and Re.¹⁴⁴ However, they are sometime expensive,¹⁴⁵ susceptible to adsorption by the impurities^{144,146} and show limited substrate specificities. “Commonly occurring metallocofactor motifs of metalloenzymes are: haem centres, Fe–S clusters ([Fe–S]), Fe centres, Cu centres, Mo centres (Mo-cofactors, Moco) and W, including various iterations and combinations. In many cases, only a single catalytic redox cofactor is found, although others can be found to be involved in internal ET (commonly the case of [4Fe–4S] containing proteins). Frequently, oxidoreductases that comprise non-metallocofactors (FAD) and PQQ-dependent enzymes also utilize metallocofactors to transfer electrons to and from their redox partners, or in other cases electrode surfaces. Additionally, metalloenzymes have been known to function as a transmembrane enzyme that functions on either side of a membrane or the enzyme contains a deeply hidden catalytic redox cofactor. One of the most fascinating examples of such an internal ET pathway can be found in prokaryotic nitrate reductase (Nar NR), such as that in *Escherichia coli* (NarGHI NR).¹⁴⁷ Modern studies have reported that Mtr respiratory pathway in *S. oneidensis* MR1 involve OmcA/MtrC playing role of “azo reductase”.^{148,149} Where flavins have been reported

to assist and improve the decolorization process.¹⁵⁰ Besides, removal of dye (AO-7) was largely associated with bio-reduction due to dehydrogenase activity (DHA) rather than electrochemical reduction.¹¹⁹ Greater DHA activity reflected greater transfer activity of intracellular electron *e.g.*, AO-7 reduction by anodic bacteria, but some of them showed EET capability.¹⁵¹

Different bioanodic degradation pathways and byproduct have been reported in case of dyes. Fig. 3 reveals a typical anaerobic degradation an azo dye methyl orange 7 (MO7) that possible occurs in anodic compartment generating by products including *N,N* dimethyl-benzene 1,4 diamine and sulfanilic acid. The reaction initiated through intra-cellular dehydrogenases producing NADH from oxidation of reduced C compounds. Azoreductase then reduced the azo bond using reduced mediator from NADH generating primary extra-cellular degradation product of dye. In a typical MFC anode using *S. oneidensis*, a symmetrically cleavage pathway of AO7 was predicted after reduction of the azo bond in the creation of aminobenzene sulfonic acid (sulfanilic acid), 1-amino-2-naphthalenol (1-A-2-N), and 2-amino-1-naphthalenol.¹⁵² Previously, the existence of aminobenzenes under anaerobic decolorization of AO7 was reported by.¹⁵³ In case of mordant azo dyes salicylic acid derivatives were numerically predominant byproducts, whereas, *N,N*-dimethylaniline, sulfanilic acid and sodium sulfanilate in case of methyl orange.¹⁵⁴ Likewise, microbial or electrochemical degradation of AO-7 produces 1-amino-2-naphthol (AN) and 4-aminobenzenesulfonic acid (AA) in anaerobic environments.²⁰ However, mineralized of a Congo red indicated less toxic and more degradable organic byproducts such as maleic acid and malonic acid in MEF-COR system.³⁵ Current density has been reported to increase with the higher concentration of azo dyes. It further helps in degradation of dyes.¹⁵⁵ Beside higher current efficiency puts a low energy demand in the system, nonetheless, required



extended time for achieving acceptable mineralization.¹⁷ Despite MFC being energy gaining rather consuming, it has CE losses due to dyes and their increasing concentrations of dyes (AO-7) and electron consumption.¹¹⁹ In some cases, degradation of azo-dyes displayed prompt electricity generation^{156,157} compared MFCs inoculated sludge. This has been linked with products of dye such as phenazine, aminophenol, and naphthol playing role as electron mediators in electricity generation.^{138,158} Mostly reports have indicated the production of toxic or mutagenic byproducts of partially degraded azo dyes in conventional anaerobic digestion (AD). Such compounds were preferred to be treated under aerobic conditions for complete mineralization. So, current approach of treating dyes in MEFS is considerably required. Sometimes, dyes or their products became toxic due to synergistic effects of several chemical or biological constituents in the effluents instead of one or certain contaminants.

Iron ($\text{Fe}^{2+}/\text{Fe}^{3+}$) is a key player in cathodic Fenton reaction. In a typical Fenton reaction ferrous salt (Fe^{2+} ions) in the catholytes catalyze the production of $\cdot\text{OH}$ from H_2O_2 .¹⁰ Usually Fe^{2+} (Fenton) is preferred over Fe^{3+} (Fenton-like), however, a combine blend of both (*e.g.*, magnetite and ilmenite) in pure homogenous or heterogeneous and in conjugation form with other cations⁵⁹ and composite electrodes can be effectively utilized. In case a typical Fenton reaction, powerful $\cdot\text{OH}$ is generated ($E^\circ = 2.80 \text{ V}$) ($k = 63 \text{ M}^{-1} \text{ s}^{-1}$). Whereas in Fenton like reaction, less powerful $\cdot\text{OOH}$ radical ($E^\circ = 1.65 \text{ V}$) is released and causing a rate limiting step in hydroxyl radical production. Moreover, Fe^{2+} is constantly renewed through cathodic reduction of Fe^{3+} and through chain of other Fenton reactions.¹⁵⁹ Reports have clearly validated better performance of Fenton reaction rather than Fenton-like reaction in pollutants (dyes) degradation.²³ For optimum MEFS, factors like type and concentration of Fe catalyst and pollutant, electrodes surface area and spacing, bio-electrochemical potential and water quality are vital. It is reported that rise in Fe^{2+} concentration improved the rate of degradation of pollutants (dyes), but the rate declines above a definite value because of acceleration of parasitic reactions between Fe^{2+} and hydroxyl radical thereby loss of the system efficiency.^{10,23} Beside efficiency of FR also depends upon the bio-anodic and cathodic potentials. Excess concentration of Fe^{2+} may result %OH loss and thereby gradual decline in degradation efficiency of the pollutant. Previously, the maximum degradation efficiency of azo dye was reported at 1.0 mM of Fe^{2+} in MEFS, however, further increase in Fe^{2+} concentration to 5 mM reduced degradation efficiency to 80.15% from 100%.⁹² Beside excessive Fe^{2+} addition may cause further operational costs¹⁶⁰ due to great volume of sludge generation and its successive treatment. Some solid iron oxides such as $\alpha\text{-FeOOH}$, FeO , Fe_3O_4 , Fe_2O_3 have been considered as iron catalysts in Electro-Fenton process. The removal efficiency of POPs was related to release Fe^{2+} from its oxides into the solution for Fenton's reaction (Fig. 2C). Moreover, synthetic iron-containing nanoparticles (*e.g.*, nano- Fe_3O_4 , $\alpha\text{-Fe}_2\text{O}_3$ and $\text{Pd/Fe}_3\text{O}_4$) have also been considered for the treatment of POPs based upon electro-Fenton process.⁵⁹

Degradation of methyl orange using hybrid advanced oxidation (Fig. 4) suggested involvement of hydroxyl radical

with amputation at electron-rich site of $\text{N}=\text{N}$ (azo bonding),¹⁶³ creating intermediates like dimethylaniline (a) and sodium benzenesulfonate (b) which then converted to aniline (c) and benzenesulfonic acid (d) respectively. Further hydroxylation of resulted conversion of aniline into hydroxylaniline (e) and benzenesulfonic acid into 4-hydroxybenzenesulfonic acid (f), and both these intermediates decomposed into hydroquinone (g) and then *p*-benzoquinone (h). The final aromatic end product is transformed into an aliphatic acid (oxalic acid or carboxylic acid) for complete mineralization into CO_2 and H_2O .¹⁶⁴ Another sequential (symmetrical and asymmetrical) degradation of dye MO using hydroxyl radical demonstrated aromatic products such as hydroquinone and *p*-benzoquinone. Whereas aliphatic byproducts like succinic acid, malic acid, acetic acid and isobutyric acid are easily assimilate into TCA and other respiratory pathways. To attain high degradation performance in MEFS, high H_2O_2 concentration and production rate are obligatory. In MXCs-integrated systems containing both biotic and abiotic phases, the relationship between current, ARB, and the electrode revealed^{123,165,166} voltage (Ohmic) losses in the electrodes due to resistance at the interface of biofilm, anolytes/catholytes and electrodes, CEM/PEM membrane, and the cathode over potential (for H_2 production or O_2 reduction).¹¹⁹ The process conversion efficiency can be lowered by 100% due to losses at the bio-anode. In cathode chamber, the reaction mechanism is comparable with classical electro-Fenton process. However, in both cases H_2O_2 reacts with Fe^{2+} to yield hydroxyl radical that could be used for oxidation of dyes.¹⁶² The cathode potential is one of the key players in H_2O_2 production and it varies with different types and composition of cathodes. For example, a maximum H_2O_2 concentration can be achieved at cathode potential of -0.4 V using graphite cathode.¹⁶⁷ Though, the maximum H_2O_2 concentration of 711.2 mg L^{-1} was witnessed under cathode potential of -0.85 V with 0.5 V in MEC using 3D electrodes as cathode.¹⁶⁸ Apart from cathodic potential, cathodic material should be resistant to corrosion and should have good electrical conductivity.

4 Materials used in MEFS

4.1 Electrode material

Electrodes play an integral role on the efficiency of any electrochemical or bio-electro-chemical cells. Electrodes types are characterized on the basis of their material, conductivity (electronic potential), reactivity, surface area, and their integration in the MEFS. Some of the electrodes have also been fabricated in laboratory.¹⁷¹ Typically in a MEFS, these electrodes are vital in terms of their compatibility with cellular organelles and excretions (extra-polymeric substances = EPS) in order to enrich biofilms of exo-electrogens on them.¹⁷² In many reactors, both electrodes *i.e.* cathode and anode were similar¹⁷³ but in others cases they may be different.¹⁷⁴ Carbon-based materials are preferred in different MES including carbon paper, glassy carbon, carbon felt, carbon cloth, graphite plate, granule graphite, granule active carbon, graphite brush carbon mesh, carbon aerogels and reticulated vitrified carbon (Fig. 5). Most of the carbon materials, such as carbon paper, glassy carbon,



carbon cloth, are composed of 2D structures that are plane with identical surface property and morphology. The electrochemical property of these pristine carbon materials can be improved effectively by proper surface treatment (acid, heat, amino gas, *etc.*) and modification with nanomaterials, conductive polymer, and immobilized electron shuttle that results insignificant improvement in performance of MES. Beside, these electrodes were modified with various types of materials like nanomaterials, activated carbon, metal (Pt catalyst) and *etc.* in order to improve their performance.¹⁷⁵

3D anodes are grabbing substantial consideration for the improvement of profoundly efficient systems.^{176,177} These anodes have demonstrated 80–90% hydrogen recovery and COD removal efficiency.¹⁵³ Additionally, layers of carbonized corrugated cardboard as anode material increased the current density by manifold.¹⁷⁸ Furthermore, pre-treatment of anode with ammonia, phosphate, trace metals proved to be increasing; porosity (biocatalytic sites increases), conductivity, and particles capable of supporting biofilm growth.^{179–181} Whereas modification of C anode with polypyrrole/anthraquinone-2,6-disulfonate cause the increased power density, redox properties and associated microbial-electrocatalysis of azo dye.¹⁷⁴ Modification of carbon fibers with carbon nanotubes and polypyrrole composite resulted in excellent biocompatibility and conductivity. Efficiency of modified anode in producing electricity is 2.63 times higher than that of unmodified ones, generating 1876.62 mW m⁻² power density, and enhanced mineralization rate of orange II dye.^{182,183}

4.2 Cathode and Fe forms and types

Cathode material should be porous and resistant to corrosion in order to support exchange of protons and electrons and interaction with oxidants. It is also usually carbon based due to less cost, good conductivity and stability.^{162,177,186} These include carbon brush, activated carbon, carbon fiber and modified CNT, carbon paper and cloth *etc.* that have been employed in MEFS.^{177,178,187,188} Production of H₂O₂ is vital in MEFS, and it then converted into highly responsive oxygen species such as [•]OH radicals to mineralize pollutants in cathode.^{153,162,177,186} Modified cathodes with C substrates with catalysts' layers and diffused (channeled) air supply proved to be highly effective in catalytic reduction of oxygen and H₂O₂ production.^{177,184,189} The iron source is one of the vital element to impact the performance of the MEFS. Usually, dissolve Fe²⁺ (Fenton) and sometimes Fe³⁺ (Fenton-like) salts have also been used in catholytes as catalysts for Fenton reaction in MEFS. Some researchers also have used iron alginate beads, scrap iron or iron plates as Fenton catalysts. Carbon-based modified cathodes with iron or transition metal oxides have also been preferred.¹⁸⁴ Wang *et al.* reported the use of composite cathodes in MEFS for dye wastewater treatment. The cathode was fabricated by generating CNTs on SS316 stainless steel mesh with addition of iron phthalocyanine as catalyst, which resulted in enhanced current density and power of the system by 937 and 2594 times respectively along with 84.6% of dye decolorization in 12 h.¹⁷¹

Combination of both divalent iron and trivalent iron (*e.g.*, magnetite and ilmenite) has found more effective for Fenton reactions. These catalysts of iron were also used in EF studies in pure forms, doped with other cations,⁵⁹ or mostly as composite electrodes.¹⁹⁰ The usage of self-designed electrodes like Fe@Fe₂O₃/CF,¹⁹¹ γ-FeOOH/CF,¹⁹² carbon nanotube (CNT)/γ-FeOOH/CF, PPy/AQDS/CF¹⁶² and FeVO₄/CF¹⁹¹ are providing iron catalyst as heterogeneous or homogeneous reactions has expanded the scope. Study showed that degradation rates of rhodamine B were 38%, 63% and 79% (15.3–30.1 a cm⁻²) with non-catalyzed CF (NCF), NCF with Fe²⁺ solution (Fe²⁺/NCF) and Fe@Fe₂O₃/NCF composite cathodes at closed circuit (1000 Ω), however, it improved to 49%, 64% and 95% (current density of 43.7–65.2 μA cm⁻²) at short circuit (0 Ω external resistor), respectively, in 12 h. Among homogenous form of iron(III) chloride hexahydrate (FeCl₃·6H₂O) and Iron(II) sulfate heptahydrate (FeSO₄·7H₂O) iron salts are mostly used and relatively cheaper than heterogeneous salts.⁴⁰ But they may cause Fe sludge generation and require adjustment of low pH for Fenton reaction thereby making system more costly. Contrarily, Fenton reaction with heterogeneous iron salts^{35,162} such as goethite (α-FeOOH), magnetite (Fe₃O₄), maghemite (γ-Fe₂O₃), clinopyrrhotite (Fe_{1-x}S) or hematite (α-Fe₂O₃) can be highly efficient for FR and catalysis of pollutants in mesophilic soil and wastewater.¹⁹³ Moreover, coating of C felt and graphite cathodes with γ-FeOOH composite,¹⁵⁷ pyrrhotite,³¹ and Fe@Fe₂O₃ (ref. 194) proved to be slowly leaching out and discharging ferric or ferrous ions in cathodic solution. Carbon aerogel with iron and copper proved to be another effective modification of cathode Fenton reaction enhancing WWT containing dyes (MB).¹⁹⁵ Such modification help control release of iron and decreases sludge generation and management cost.¹⁷⁵

5 Types and operation models of microbial-electro-Fenton systems

In the last two decades, developments in MEFS have provided considerable scope for innovative efficient solutions for wastewater treatment. Such green multifaceted technology has shown great prospects^{7,22,23,196–198} though they are still under stages of research and development for large scale application. Studies have reported treatment of dyes and pigments through different MEFS that has been implemented in different configurations, with different membranes, mostly proton exchange membrane (PEM), cation exchange membrane (CEM), and through their integration with biological degradation of dyes. In most of the studies, most commonly treated dyes in MEF systems include; Congo red, orange G, orange II, rhodamine B, methyl orange, reactive black 5 Lissamine green B and crystal violet. Comparatively, orange II (Acid orange 7) is the most studied azo dye in the said systems.

MEF technology typically evolved as an extended version of MFCs and MECs, where apart from anodic (anaerobic) oxidation of organic compounds additional cathodic oxidative detoxification and decomposition of certain pollutants is carried out through highly reactive hydroxyl radicals generated





Table 2 Removal of dyes in different (batch or sequential) MEFS

Reactor configuration	Anodic inoculum	Cathode material	Pollutants	Concentration	pH	Amendments	Power density	Removal efficiency	Operation time	Reference
MFC	<i>Shewanella decolorationis</i> S12	CNT/PTEF/ γ -FeOOH	Orange II	35 mg L ⁻¹	7	Synthesis of H ₂ O ₂ in MFC using cathode as spectrographically pure graphite (SPG) MEF was operated with effluents of constructed wetland	25.13 mW m ⁻²	100%	43 h	126
MFC	Anaerobic sludge	Graphite	Acid blue 113	300 mg L ⁻¹	3	Modified electrodes used to enhance performance	3.5 A m ⁻²	91.57%	43 h	94
Dual chamber MFC	<i>Shewanella decolorationis</i> S12	Ppy/AQDS/carbon	Orange II	10 mg L ⁻¹	7	Modified electrodes used in dual chamber MFC	823 mW cm ⁻²	97%	7 h	211
Dual chamber MFC	<i>Shewanella decolorationis</i> S12	CNT/ γ -FeOOH	Orange II	10 mg L ⁻¹	7	Modified electrodes used in dual chamber MFC	230 W m ⁻²	100%	14 h	92
MFC	Glucose	Spectrographic pure graphite (SPG)	OrangeII, Amaranth	75 mg L ⁻¹	7	Iron conc. Changes 1.14–3.43 mg L ⁻¹ & 0.1–1 mg L ⁻¹	25.13 mW m ⁻²	82.59%	1 h	92 and 126
MFC	Anaerobic sludge	Graphite	Amaranth	75 mg L ⁻¹	3	Optimal cathode conditions were applied n cell for H ₂ O ₂ production	42.6 Am	84.24%	2 h	167
MFC	Brewery wastewater	Fe@Fe2O3/NFC	Rhodamine B	15 mg L ⁻¹	3	The double chamber MFC was proposed by utilizing Fe@Fe ₂ O ₃ /carbon felt	307 mW m ⁻²	95%	12 h	195
MFC (H-type)	<i>T.versicolor</i> <i>S.oneidensis</i>	Graphite rod	Lissamine green B	10 mg L ⁻¹	2	Composite cathode Combination of fungus and bacterium used	1.2 W m ⁻³	94%	9 h	212
MFC (H-type)	<i>T.versicolor</i> <i>S.oneidensis</i>	Graphite rod	Crystal violet	10 mg L ⁻¹	2	Combination of fungus and bacterium used	1.2 W m ⁻³	83%	9 h	212
Hybrid reactor of benthic MFC and EF	Sewage sludge, marine sediments	Graphite sheet	Lissamine green B, reactive black 5, indigo carmine	10 mg L ⁻¹ , 20 mg L ⁻¹ , 50 mg L ⁻¹	7	Marine sediment microbial fuel cell used to drive external electrochemical and electro-Fenton processes	1033–1046 mV	100%	1 h	36
MFC (hybrid cell)	Sewage sludge	Graphite sheet	Reactive black 5	50 mg L ⁻¹	7.5	SMFC anode and electro-Fenton cathode connected with a salt bridge	1033 mV	88.2%	15 min	36



Table 2 (Contd.)

Reactor configuration	Anodic inoculum	Cathode material	Pollutants	Concentration	pH	Amendments	Power density	Removal efficiency	Operation time	Reference
MFC (hybrid cell)	Sewage sludge	Graphite sheet	Lissamine green B	10 mg L ⁻¹	7.5	SMFC anode and electro-Fenton cathode connected with a salt bridge	1034 mV	98.2%	15 min	36
MFC (hybrid cell)	Sewage sludge	Graphite sheet	Crystal violet	5 mg L ⁻¹	7.5	SMFC anode and electro-Fenton cathode connected with a salt bridge	1046 mV	96.2%	15 min	36
MFC (hybrid cell)	Sewage sludge	Graphite sheet	Indigo carmine	20 mg L ⁻¹	7.5	SMFC anode and electro-Fenton cathode connected with a salt bridge	1045 mV	97.2%	15 min	36
MFC (hybrid cell)	Sewage sludge	Graphite sheet	Poly R-478	80 mg L ⁻¹	7.5	SMFC anode and electro-Fenton cathode connected with a salt bridge	1035 mV	19.1%	15 min	36
MFC	<i>Shewanella decolorationis</i> S12	Ppy/AQDS/carbon	Orange II	70 mg L ⁻¹	7	Modified electrodes used in dual chamber MFC	823 mW cm ⁻²	100%	50 h	174
MFC powered advanced FS	Anaerobic sludge	Carbon felt	Acid orange 7	50 mg L ⁻¹	3	MFC was combined with Fenton-like technology to simultaneously generate electricity and degrade dye	15.9 W m ⁻³	89%	60 h	204
MFC	Anaerobic sludge	Fe ₂ O ₃ /ACF	Methyl orange	5 mg L ⁻¹	2	Composite Fe ₂ O ₃ /ACF electrode was used	268.10 mW m ⁻³	86.7%	12 h	213
MFC	Anaerobic sludge	Graphite rod	Congo red	100 mg L ⁻¹	7	MFC and a COR reactor were integrated together	808.3 mW m ⁻³	90%	72 h	35
MFC	Domestic wastewater	FePc/CNT/SS316	Reactive black 5	50 mg L ⁻¹	7	Composite cathode is used	726.55 mW m ⁻²	80%	12 h	171
MFC	Anaerobic sludge	Graphite rod	Acid blue113	100 mg L ⁻¹	3	Graphite cathode was treated with nitric acid	36.438 mW m ⁻²	71.36%	12 h	17
MREC	Domestic wastewater	Graphite	Orange G	400 mg L ⁻¹	2	Salinity gradient energy drove the microbial-electro-Fenton process	2 A m ⁻²	100%	10 h	32
3D-EF-MFC	Anaerobic sludge	Activated carbon	Methyl orange	100 mg L ⁻¹	3	MFC coupled with 3D electro Fenton technique	566 mW m ⁻³	84%	72 h	183



Table 2 (Contd.)

Reactor configuration	Anodic inoculum	Cathode material	Pollutants	Concentration	pH	Amendments	Power density	Removal efficiency	Operation time	Reference
MFC-MEC	Domestic wastewater	Graphite	Methylene blue	50 mg L ⁻¹	3	MFC as renewable power source used to power MEC-electro-Fenton process	50.1 mW m ⁻²	97%	16 h	27
MFC power	Anaerobic sludge	Graphite plate	Acid orange 7	16 mg L ⁻¹	6	A novel heterogeneous EAFL system driven by MFC used	54.02 mW m ⁻²	96.4%	2 h	214
MEF-COR	Glucose	Graphite rod	Congo red	20 mg L ⁻¹	7	Integration of MEF with catalytic oxidation system	808.3 mW m ⁻³	90%	72 h	35
MFC-AFT	Anaerobic sludge	Iron plate with carbon paper	Acid orange 7	400 mL (0.16 M NaCl)	3	External addition of 2 mM H ₂ O ₂ and 0.3 mW power output	0.27 mW	85%	—	173
Automatic MEFs	Electroactive biofilm	Carbon impregnated iron oxide	Acid orange 7	50 mg L ⁻¹	7	Microchannel-structured carbon enlivened with iron oxides utilized as electro-Fenton cathode	—	93–96%	24 h	202



Fig. 5 Different anode material used in MEFS (adopted from ref. 184 and 185).

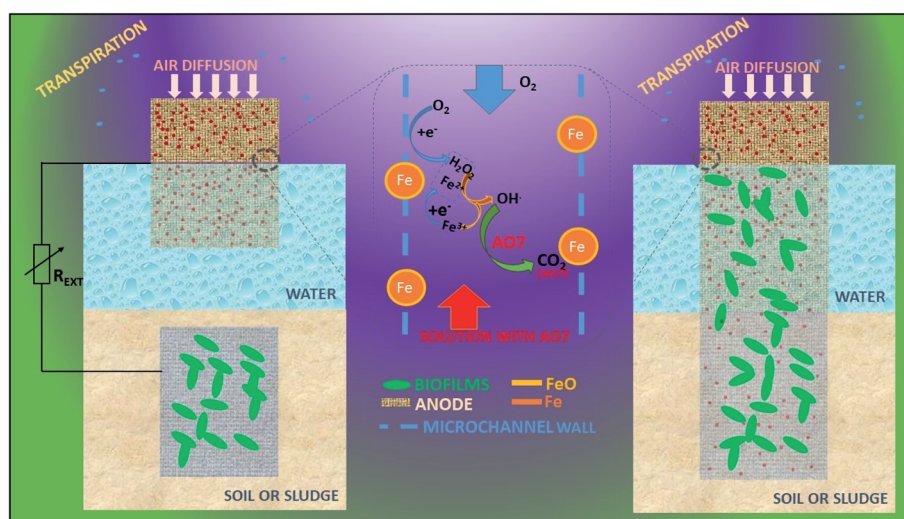


Fig. 6 Automatic microbial electro-Fenton system (AMEFS) (refabricated from ref. 202).

in Fenton reaction. However, previous WWT processes including batch, sequential or continuous flow reactors also helped in innovating MEF technology. Extensive studies of MEFS are underway toward development of new designs for achieving ideal system performance. MFC-Fenton system is though less efficient but cost effective in terms of electrical and H_2O_2 outputs associated Fenton reaction for dyes treatment.^{30,154,199} Development of single chamber rather than double chamber considerably improved the energy out of the MFC system.²⁰⁰ MFC powered EFS has also gained advancement through integration of super-capacitor for bio-electricity storage for subsequent use in Fenton reaction.^{21,201} Considering treatment efficiency on priority compared to electrical energy outputs, MEC based Fenton process has shown greater prospects. Such system is highly efficient for treating high load of contaminants and is equally good to treat domestic and industrial wastewater at the same time. Still this system need an

additional input of minor energy (0.2–0.8 V) although, it is about 100 times lower than conventional EFS.³² But the possibility that lies in usage of single chamber MFC to empower a two-chamber MEC-electro-Fenton system for treatment of persistent pollutants is another good option to avoid any external input electrical energy for self-sustainable.²⁷ Before seeking industrial scale applications there are several other challenges that need to be addressed.

Microbial Reverse Electro-dialysis Cells (MRECs), Automated Microbial Fenton Cell (AMFC) are the most innovative in MEFS.^{32,183,202} The development of continuous or semi-continuous mode of reactors is highly required in order to further enhance these technologies at large commercial scales relying on previously developed anaerobic and MFC/MEC based reactors. MEFS technology have offered considerable opportunity of treating two different substrates (pollutants) or wastewater in two separate chambers of the same reactor.¹⁷⁵ They



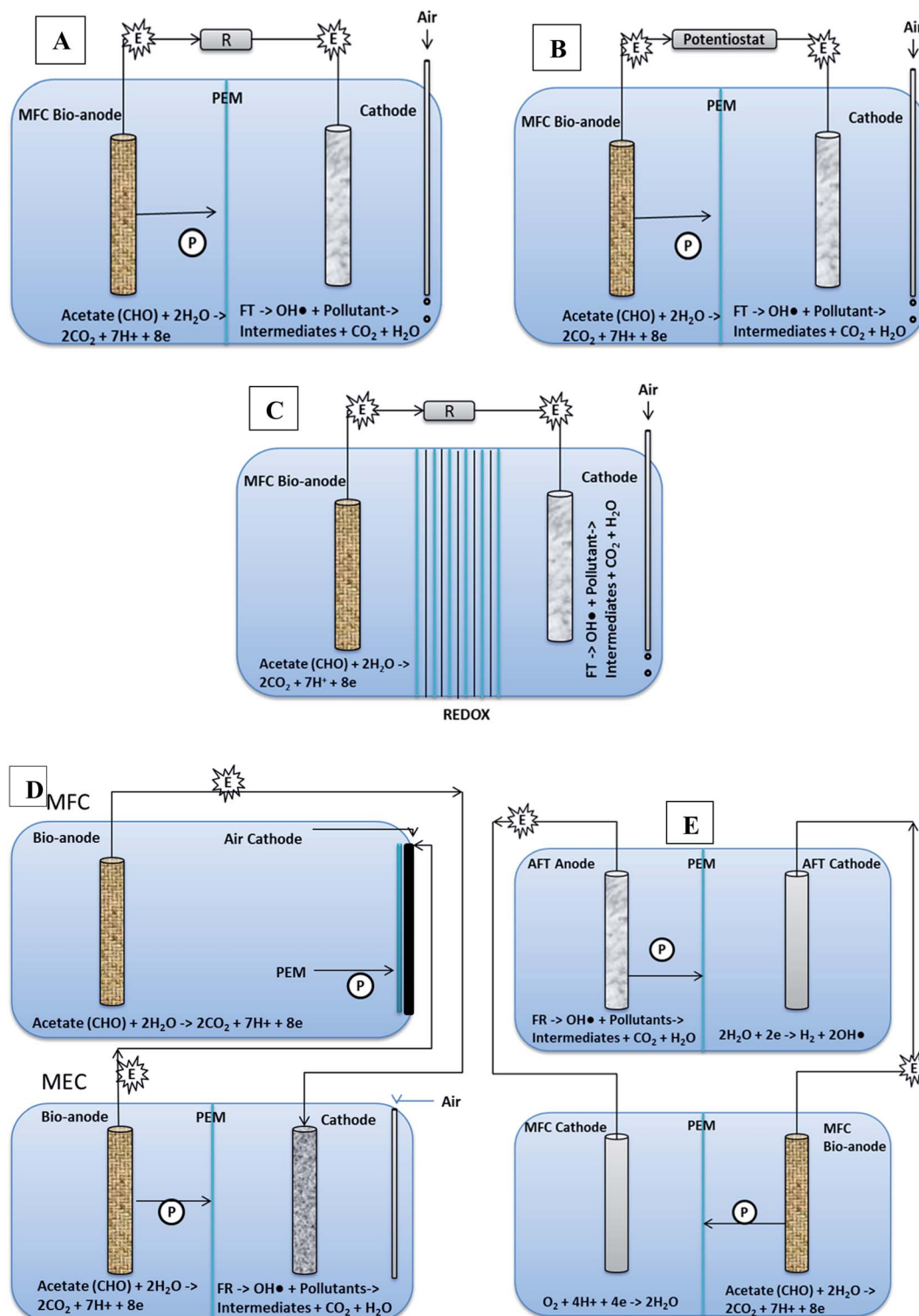


Fig. 7 Removal of dyes in different batch MEFS (A) MFC-electro Fenton system (B) MEC-electro Fenton system (C) MREC-electro Fenton system (A–C single cells) (D) MFC-MEC-electro Fenton system (double cell) (E) MFC assisted AFT (double cell).

have been studied for the removal of natural (biopolymers) and synthetic/recalcitrant compounds including azo dyes (Congo red,³⁵ methylene blue,²⁷ acid orange,¹⁷³ orange II, rhodamine B), pesticides, heavy metals, phenol, trimethoprim, caffeine,

sulfamethazine, and ranitidine.^{203,204} The efficiency of systems above varies with electrode material, microorganisms, pH, current density, cathode potential, and concentration of ferrous/ferric ions their homogenous and heterogeneous forms.



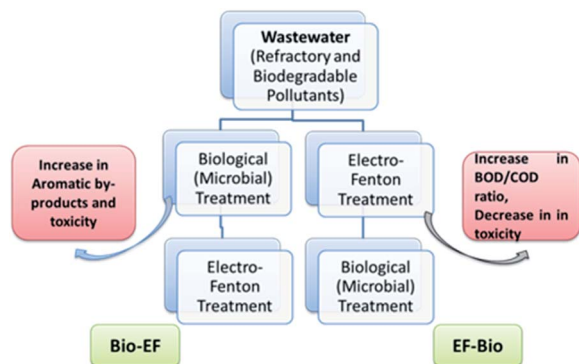


Fig. 8 Sequential treatment of refractory compounds (dyes) through biological and EF/EAOP.

5.1 Classification of microbial-electro-Fenton reactors

MEFS can be classified based upon their configurations and mode of operations. There are operated as (i) single cell [e.g. MFC or MEC or MREC], (ii) double cell [MFC-AFT],²⁹ and (iii) hybrid or sequential [EF and degradation of microbes].²⁰³ Most of them run under batch mode and only few under continuous modes.^{205,206} Besides, there are mostly laboratory scale reactors, and few pilot scale with none large scale.⁴⁴ Considering MFC batch reactors as the bench mark, several modifications have been done in its configuration and fabricating material converting it into more efficient wastewater treatment and energy generating technology.²⁰⁷ For instance, Modification of C anode with polypyrrole/anthraquinone-2,6-disulfonate cause the power density to increase, redox properties and associated bio-electrocatalysis of azo dye.¹⁷⁴ Modification of carbon fibers with carbon nanotubes and polypyrrole composite resulted in excellent biocompatibility and conductivity. Efficiency of modified anode in producing electricity is 2.63 times higher than that of unmodified ones, generating $1876.62 \text{ mW m}^{-2}$ power density, and enhanced mineralization rate of orange II dye.¹⁸³

5.1.1 Batch mode of operation. In batch mode, specified amount of waste liquors as anolytes and catholytes are treated for a stipulated period of time and then drawn out to revive the reactor.²⁰⁸ MFC or MEC reactors have been mostly used under batch mode reactors in various studies. Agitation is provided in the system using magnetic stirrer for the anolytes and catholytes.²³ Aeration is maintained in a cathodic chamber by using commercial air pumps.²⁰⁹ Time span of treatment usually varies 2–10 days; however, it may exceed to 30 days. It purely depends upon the efficiency of the system with reference to specific waste type. For simple substrates generally it usually takes less time whereas for complex substrate longer time is required.²⁰⁷ However, major limitation of batch reactor is organic compound's mass transfer rate mass to microbes on anodic surface for biocatalysis and harnessing of electrons and protons to harvest cathodic H_2O_2 (ref. 50) for FR. However, depletion of nutrients in medium under batch mode may cause potential losses and inefficiencies in the systems.²¹⁰ Table 2 shows details on different batch or sequential mode MEF reactors with their

operational conditions and efficiencies for dyes wastewater treatments.

5.1.1.1 One step (*in situ*) integrated process. One step integrated process involves a single MFC/MEC or MREC reactor, generating electrical potential for cathodic Fenton process. It is classified into following types based upon their configurations or designs.

5.1.1.1.1 MFC based electro Fenton system (MFC-EFS). MFC-EFS is basically a two chamber MEFS with *in situ* FR system in cathode. It can mineralize biodegradable organics in anodic chamber, and recalcitrant organic pollutants (dyes, pesticides) in cathodic chamber through Fenton reaction (FR) or electrochemical Fenton (EF) treatment shown in Fig. 7A.¹⁷⁴ MFC-EFS reactors are cost-effective than Electro Fenton (EF) processes because they are energy generating rather than consuming. Besides they also generate quantifiable amount of hydroxyl radical from FR for decomposing pollutants.¹⁷

5.1.1.1.2 Automatic microbial electro-Fenton system (AMEFS). A three-dimensional micro and macro-porous geometrical electrode provides large ratio of surface area to volume for biofilm development and electrolyte or substrate diffusion for efficient MEFS shown in Fig. 6.^{215,216} AMEFS is highly innovative and workable development in MEFS for *on situ* remediation of dyes and other related pollutants in water, soil or sludge. It is fabricated with microchannel-structured carbon impregnated iron oxides cathode for EF reaction and with an additional but similar anode for biofilm bio-catalysis in a two-electrode configuration connected by an external circuit. The second configuration of AMEFS includes a similar but single-electrode configuration serving both as bioanode and cathode. AMEFS is a spontaneously driven system similar to natural transpiration system in plants. The highest degradation efficiency (93–96%) at 50 ppm is generated at the condition of short-circuit with two-electrodes and that was comparable to single-electrode configuration. Such a system can be easily installed for sludge and soil purification by directly inserting microchannel structured carbon electrodes in the contaminated site or reactor.²⁰² Furthermore AMEF is equally good to operate under batch or continuous mode operations.

5.1.1.1.3 Other novel designs. MECs an extended version of MFC with small input of charge (0.2–0.8 V) (Fig. 7B) has proved to be yielding H_2O_2 several time higher than MFCs. Therefore, MEC electro Fenton reactor can be suitable for treating wastewater that contains comparatively excessive loads of pollutants in both anodic and cathodic chambers with remarkably lower energy consumption than classical electro Fenton process.²¹⁷ This system was efficiently used for azo dye WWT.³² Besides, it uses (approx low cost $25.93 \text{ kW h kg}^{-1}\text{-TOC}$) an average of 75% (17 fold) less energy as compared to the regular electro-Fenton process ($45.8\text{--}865 \text{ kW h kg}^{-1}\text{ TOC}$).^{218,219} The degradation efficiency of TOC was found to be 93.1%. The larger removal at the short circuit leads to larger current density of $43.7\text{--}65.2 \mu\text{A cm}^{-2}$.¹⁹⁵

MFC-3D EF is originally an MFC reactor but filled with granular AC either in both anode and cathode chambers or in



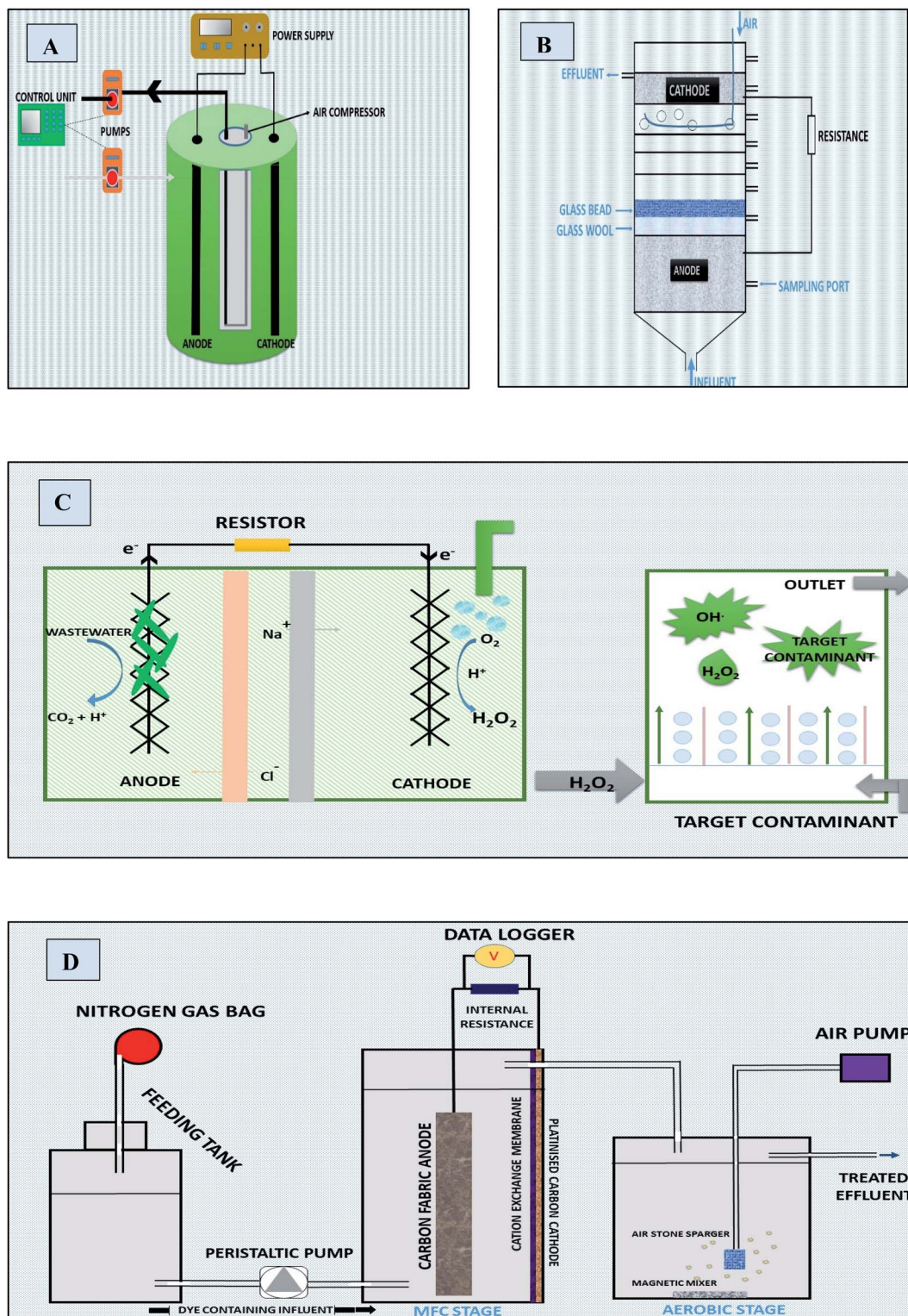


Fig. 9 Continuous operated reactors (A) schematic diagram of Airlift continuous reactor with electro Fenton setup (refabricated from ref. 229) (B) schematics diagram of mediator- and membrane-less MFC column reactor (refabricated from ref. 240) (C) microbial desalination reactor (refabricated from ref. 175) (D) schematic of integrated MFC-aerobic system (refabricated from ref. 153).

cathode chamber as electrodes. The bed electrodes played an arbiter job in cathode and anode chambers, elevating the output voltage and power density, and lost cause internal

impedance to get lowered. Dye evacuation efficiency is observed to be 84%.¹⁸³

Microbial reverse electro-dialysis cells (MREC) is a novel hybrid bio-electro-chemical system for efficient H_2O_2



Table 3 Characteristics of continuous mode reactors

Reactor configurations	Inoculum	Pollutants	Operational parameters	Hydraulic Retention time	Applied-voltage/current	Efficiency	References
Two chambers having 10 L each, anode: Carbon fiber brush, cathode: graphite plate SMFC, electrodes: Graphite sheet	Domestic waste water	Methylene blue	Aeration rate of 350 mL min ⁻¹ , 20 mg L ⁻¹ , pH 2, Fe ²⁺ of 0.2 mM, applied voltage 0.4 V	28 h	0.2 V	95% dye decolorization 89% TOC removal	44
	Marine sediment	Lissamine green	Na ₂ SO ₄ 0.01 mol L ⁻¹ , iron concentration 150 mg L ⁻¹	1 h	—	97–99% decolorization	36
Cylindrical reactor containing two electrodes at the center, cathode: modified graphite with carbon nanotubes, anode: Graphite	Dye solution	C.I. acid red 14 (a), C.I. acid blue 92 (b)	NaCl: 1 g L ⁻¹ , 0.05 mM Fe ³⁺ , pH: 3, aeration 10 mg L ⁻¹ dye, effective volume 1 L, flow rate: 0.33 L h ⁻¹	—	0.14 A	86.78% COD removal in 60 min, pollutant degrading efficiency 91.22% (a) 93.45% (b)	235
Microbial fuel cell MFC (H type)	<i>T. versicolor</i> and <i>S. onetensis MR-1</i>	Lissamine green B, crystal violet	Graphite rod used as anode, dye conc. 10 mg L ⁻¹ , pH 2, iron concentration 150 mg L ⁻¹	—	—	94% Lissamine green B, 83% crystal violet after 9 h treatment time	212
Fluidized bed reactor, cathode: nickel foam layered with iron-chitosan, anode: graphite sheet	Dye solution	Lissamine green B	pH 2, 0.15 L working volume, 0.01 M, Na ₂ SO ₄ , 100 mg L ⁻¹ dye, mixing by continuous air flow at 0.15 vvm ^a	45 min and 90 min	5 V	75% TOC removal, dye degradation efficiency 95%	236
Cylindrical glass reactor, electrodes: graphite bar	Dye solution	Lissamine green B (a) reactive black 5 (b)	pH = 2, 22 °C, catalyst 115 g Fe alginate gel beads made of sodium alginate, BaCl ₂ , Fe ₂ (SO ₄) ₃ , air bubbling near cathode at 1.5 L min ⁻¹ , working volume 1.5 L	6 h (a), 12.5 h (b)	3 V	TOC removal 81% (a), % (a), 87% (b)	229
Cylindrical glass reactor. Electrodes: graphite sheet	Dye solution	Lissamine green B (a) azure B (b)	Catalyst: 8.69 g Fe alginate beads, air bubbling near the cathode at 1 L min ⁻¹ , working volume 0.15 L, pH=2	30 min	14.19 V	TOC removal 93% (a) 89% (b)	237

Table 3 (Contd.)

Reactor configurations	Inoculum	Pollutants	Operational parameters	Hydraulic Retention time	Applied-voltage/current	Efficiency	References
Bubble reactor formed of cylindrical glass, electrode: graphite	Dye solution	Lissamine green B, methyl orange, reactive Black 5, Fuchsin acid	0.04 M Na ₂ SO ₄ , 8.5 mg L ⁻¹ (LGB), 1.5 mg L ⁻¹ (MO), 70 mg L ⁻¹ (RB5), 15 mg L ⁻¹ (FA), pH=2, bubbling compressed air at L min ⁻¹ , 0.675 L working volume	21 hour	15 V	47% TOC removal, 43% dye removal	238
Bubble glass column reactor, electrodes: graphite	Textile wastewater	Rhodamine B	pH 3, catalyst: FeCl ₃ at 5 mg L ⁻¹ , 3 L of electrolyte at 10 mL min ⁻¹	8 h	3.5 V	98% dye - removal	239

production through *in situ* electric energy generation by the microbial oxidation of biodegradable matters and salinity-slope that is between different fresh and salt channels and does not require power input (Fig. 7C). The cathodic and anodic chambers are separated with RED stacks and the reaction of cathode is not affected by the continuous flow of wastewater in anode chamber.³² It proved to be highly efficient in comparison with MFC and MEC with dye removal efficiency up to 99%. Moreover, the cost of the treatment proved to be lower (25.93 kW h kg⁻¹ TOC) by $\geq 45\%$ than other Fenton processes (45.8–865 kW h kg⁻¹ TOC).^{218,219}

Using MFC as renewable energy source to power MEC Fenton process (Fig. 7D), electricity can be saved. But removal efficiency of pollutants and power density is found to be low as compared to traditional Fenton processes.²⁷

MFC-AFT is a two cells system; where anodic Fenton reaction based treated of pollutants occurs in chemical cell supported by electronic energy harnessed from bio-anodic compartment of another MFC (Fig. 7E). Kinetic studies indicated that this system had higher pseudo-first order rate constant than other conventional Fenton processes and 87% of the dye was degraded in AFT system within 15 minutes. According to electrochemical analysis, the corrosion of iron was not hindered by dye. It was also revealed that MFC power density was improved by increasing dissolved oxygen in cathode compartment, which resulted in increased degradation of dye.¹⁷³

5.1.1.2 Sequential or hybrid process. Sequential MEFS is a two-step sequential treatment of refractory compounds through pre or post EF oxidation with biological method in order to achieve a comprehensive efficiency shown in Fig. 8. The span of EF is minimized in order to keep the system cost effective. However, the type of effluent and its priority pollutant helps in deciding whether EF should be used as pre-treatment or post-treatment in conjugation with biological methods. EF as pre-treatment, involves hydroxyl radicals for oxidative mineralization of refractory compounds or into aliphatic by-products that may be sometimes toxic.²²⁰ Later biological process completely removes any remaining short chain compounds.^{159,221,222} EF-microbial oxidation proved to be effective treatment strategy for real textile wastewater. If EF is applied as post treatment then it helps complete decomposition of toxic and mutagenic aromatic by-products of compounds like azo dyes such as alpha-naphthol or sulfanilic acid that are stable specifically under anaerobic biological condition.^{21,223} Selection of treatment strategy still greatly depends upon the nature of dyes. Previously, an EF-biological (aerobic) (yeast) process helped to degrade reactive black 5 dye 91%.²²⁴ EF and aerobic-microaerophilic process when combined removed almost 85% color and COD with 56% TOC of textile wastewater. Whereas, 52.7% color, 82.5% COD, and 41% TOC were observed when EF-aerobic microbial process was used. Moreover, dyes removal rates improved to >95% with 85% of TOC in a bubble column MFC reactor coupled *ex situ* EF process.^{36,153} The dyes wastewater usually contain many organic compounds beside of dyes. Some pretreatment *e.g.* coagulation, precipitation may be essential to improve dyes removal. Therefore, integrated process based on MEFT would be useful for dye wastewater



treatment. Afanga *et al.* reported the sequential treatment of dye wastewater employing the integrated coagulation-electro-Fenton process. The system achieved remarkable removal of organic matter along with dye degradation. The results revealed the removal of COD, TOC, and TSS up to 97%, 98% and 98% respectively.²²⁵

5.1.2 Continuous mode of operation. In continuous mode reactors, the substrate (anolytes or catholytes) is continuously charged along with discharge of products simultaneously during upstream and downstream processing. Such systems are more dynamic and overcome the issues of mass transfer rates (coefficient) and nutrients depletion^{226,227} through constant flow and agitation provided.²³ Current density obtained by continuous MFC was 2 times more than that obtained from MFC operated under batch mode.²⁰⁷ Continuous mode has higher efficiency in mass transfer and heat transfer and are less spacious than batch mode.²²⁸ In addition, continuous operation allows the usage of extreme conditions that aids to improve yield, continuous can allow up to 300° and 30 bar of temperature and pressure. Moreover, continuous process is easier, flexible, and faster to scale up than that of batch process.

Previously, different continuous flow reactors *viz.* Airlift²²⁹ Fluidized bed reactor, bubble reactor, Tubular,²³⁰ Plug flow,^{226,231} MFC, MDC and MEC [MFC-MEC, MFC-membrane²³² were used based upon either microbial or electro-chemical processes for treatment of different pollutants (Table 3). Such reactors provided a solid platform for development of any continuous MEF reactors. Fig. 9 shows the various continuous flow reactors.

5.1.2.1 MFC and MEC. Earlier MES studies reported both partial and almost complete azo dyes degradation (*e.g.* acid orange-7, *etc.*)²³³ in batch and continuous flow integrated MFC (-aerobic system) and MEC (-membrane).¹⁵³ Single-chamber MFCs or membrane-less MFCs are favored over double chamber even for large scale WWT.²³⁴ Moreover, single chambered MFC tubular or inner circular electrodes proved to be superior for efficient treatment of azo dyes. Integration of Fenton process in MFC holds the greater potential of efficient degradation and detoxification of the dyes through maintaining maximum cathode half-cell potential for higher production of H₂O₂. Recently, a pilot scale (20 L) two chamber rectangular MEF reactor was developed to treat dyes WWT under continuous mode.⁴⁴

5.1.2.2 Microbial desalination cell (MDC). A microbial desalination cell (MDC) is one of the most innovative modified version of MFC containing a desalination compartment sandwiched between CEM and AEM for separating it from cathode and anode chambers respectively. MDC is capable of simultaneous WWT and desalination with minor input of energy.¹⁷³ Charge balance is maintained by salt ions migration from desalination chamber to cathode and anode by ion exchange membranes.¹¹⁷ MDC-MEF hybrid system, comparatively a novel approach, could be employed for the efficient removal of dyes from wastewater. Huang *et al.* reported application of MEF integrated MDC for increased degradation of methylene blue. The system reached maximum; power density of 566 mW m⁻³ and H₂O₂ of 24.07 g per m³ per day with degradation efficiency

up to the 82.8% under the influence of Fenton reaction. Without which the system was capable to attain only 53% efficiency.¹⁸³ Comparison in performance of a MEFS was investigated by switching MDC batch mode to continuous. System demonstrated a maximum power density of 15.9 W m⁻³ in continuous and 13.9 W m⁻³ in batch mode. Additionally, 83.7% of COD was eliminated in the continuously fed system at two days hydraulic retention time, and it was 13.8% more than that obtained under a two days batch system. It was demonstrated that enrichment of ARB and high mass transfer under continuous flow and feeding condition might have improved the system performance.^{207,228}

5.1.2.3 Hybrid cell: sediment MFC (SMFC) or benthic MFC (BMFC). Benthic or Sediment MFC (BMFC or SMFC) assisted FR and BMFC-FR hybrid processes also highlights importance of continuous or semi-continuous cost effective wastewater treatments.

6 Feasibility of scaling up of MEFS and cost benefit analysis

Current developments on MEFS propose innovative energy efficient solutions for dyes and industrial WWT in near future.²¹ Most of the MEF studies that are conducted so far are at lab scale, relying upon MFC and MEC configurations.⁴⁴ The greatest driver in up-scaling of MEFS would be MFC/MEC based multifaceted reactors that work on the principles of harnessing renewable energy from WWT to minimise the capital cost.⁵⁹ Initial advancement in MFC or MEC at pilot scale integrating EF reactors by addressing previously reported drawbacks could be the benchmark for further developments. Nevertheless, some reports outlined the practicality of the said technology for direct/or indirect power generation from wastewater at volumes exceeding those in lab scales MEC²⁴¹ and MFC.^{176,242–245} Comparatively, WWT capacity of MEC based MEF system proved to be much greater than that of MFC based system.²⁷ A very recent study mentioned up-scaling of MEC (0.4 V) based MEFS of 20 L for the treatment of dye WW using C fibre brushes as anode and graphite plate and silver/silver chloride as cathode with Fe²⁺ 0.2 mM and Na₂SO₄ 50 mM. Almost 100% removal of methylene blue dye was observed after 8 hours treatment. Likewise the dyes and TOC removal rates were 100%, 70–90% in case of Meldola's blue (MeB), Toluidine blue (TB), Orange G (OG) and Rhodamine 6G (Rh6G) and Rhodamine B (RhB) after 28 hours respectively.⁴⁴ Overall, complete mineralisation of dyes was significantly less compared to their decolorization and it has been related to their complex aromatic structure and high molecular weight. Other studies on lab scale MEFS reported that for MB removal, the K_{app} was 0.43 h⁻¹ and K_{TOC} was 0.22 h⁻¹, while compared to Orange II, K_{app} was 0.212 h⁻¹ and K_{TOC} was 0.0827 h⁻¹.¹⁷⁴ Thus, the results explained that scaled-up MEFS has higher efficiency than that of MEFS operated at lab scale for the treatment of dyes containing wastewater.

Prior studies have revealed that MEF technology is a cost-effective substitute to traditional EF technology for the treatment of dye wastewater with energy consumption of only 25.93



$\text{kW h (kg TOC)}^{-1}$.^{32,39} But for large scale application, cost benefit analysis of MEF systems is vital for its feasibility in dyes and industrial WWT.⁴⁴ MFC based MEFS are facing technical and cost challenges for commercial viability at large scale. Moreover, improving the performance and reducing the capital costs of reactor and its operation need to be addressed. System scaling-up requires extensive studies on the reactors' designs and material and operational variables, typically related to continuous *in situ* regeneration of catalyst *i.e.* Fe^{2+} , and H_2O_2 in order to prevent sludge formation.²¹ So, primarily, the design of bio-reactor is extremely important for optimum functioning and then up-scaling. Initial modeling through computer simulated program of MEFS is highly important as an effective tool for reactor configuration.²⁴⁶ The major cost based variables include electrodes and membranes (CEM/PEM). Specifically, the cost of the membranes sometimes cost an about 38% of the total cost of the reactor. For instant, Nafion membrane cost around 2500 \$ per m^2 .²⁴⁷ And is 37 fold higher than sulfonated biochar (SBC)-600 with PVA membrane costing 77 \$ per m^2 . Contrarily, per unit cost of SBC-600 membrane for proton conductivity was 0.42 S per cm per \$ and that proved to be 32 times greater than Nafion membrane (1.31×10^{-2} S per cm per \$). Likewise, the estimated standardized energy reclamation for SBC based MFC was much lower *i.e.*, $0.014 \text{ kW h m}^{-3}$ ($6.356 \text{ kW h kg COD}^{-1}$) than Nafion based MFC ($0.024 \text{ kW h m}^{-3}$ ($9.59 \text{ kW h kg COD}^{-1}$) system.²⁴⁸ Additionally, GORE-TEX membrane has been reported quite cheaper and costing an about 21 \$ per m^2 and besides, the PVA modified membranes has been reported equally cost-effective.²⁴⁷ The electrodes do a crucial job in electricity generation and a main factor in deciding the implementation of the MFC technology at a large level. Different materials ranging from non-corrosive stainless steel to versatile carbons have been investigated as anodes in different structures and shapes.²⁴⁹ The cost of C clothes varies from \$ 100–1000/ m^2 . Still, cheap carbon mesh based electrodes costing an about \$10–40/ m^2 can also be another good alternative to of carbon cloth.²⁵⁰ Moreover, the cost of cathode increases when it is modified metal catalysts [*e.g.*, FeSO_4 cost = \$13782/kg and FeOOH cost = \$ 826/kg]²⁵¹ to apply in MEFS. In this case possibility of using scrap iron waste and the wastewater originating from steel and mining industry could be a good source of iron in cathode for Fenton reaction.^{251,252} In the similar situation, usage of non-polished graphite over carbon might increase the microbial adhesion over anode and will be cost-effective for huge-scale operation. Recently cheaper such as activated carbon and metal mesh showed oxygen reduction performance.²⁴⁷ Usage of graphite coated with metal as cheaper electrode material and carbon felt results in better performance and electron transfer because of their low cost, good conductivity and huge surface area in contrast to other expensive materials.^{253–255} Bio-electricity production in system (*e.g.*, MEC-MEF) through microbial bio-catalysis of organic compounds cut down energy budget by almost 40–50%.²⁵² When the MEF process run without external energy than the normal electrical energy budget was 11 526.6 $\text{kW h (kg TOC)}^{-1}$ with respect to TOC removal costing US\$ 2420.59. The higher cost relates to higher energy consumption and lower efficiency in MFC based

MEF process requiring more reaction time and aeration costs. However, MEC based system requires a small input (0.2–0.8 V) of electricity in order to generate H_2O_2 in cathode at much high rate as compared to the MFCs. The production of H_2O_2 requires much lower energy *i.e.*, $0.93 \text{ kW h kg-H}_2\text{O}^{-1}$ with a cost price of US\$ 0.195 in MECs compared to traditional electrochemical process^{20,27} Still only small amount of electrical energy was directly applied *i.e.* $5.96 \text{ kW h (kg TOC)}^{-1}$ with a cost of US\$ 1.25,⁴⁴ and it was considerably low in comparison to the traditional EF processes.^{256,257} Besides, the energy budget of MEC based MEF system is highly economical considering the *in situ* bioelectricity generation by microbes that reduces the electrical energy consumption [$0.728 \text{ kW per h per kg}$ for aniline] may be hundred times lower than conventional electro Fenton process ($74 \text{ kW h kg}_{\text{aniline}}^{-1}$).²¹⁷ The approximate capital cost of MES (MFC and MEC) reactors were reported to be \$100,000/ton COD \times day and \$1220/ m^3 .^{258,259} MREC-Fenton system has been reported to treat azo dyes (Orange G) containing wastewater with a cost of US\$ 5.44 for $25.93 \text{ kW h (kg TOC)}^{-1}$ which is very less as compared to traditional EF processes costing US\$ 9.6–179.76 for $45.8\text{--}865 \text{ kW h (kg TOC)}^{-1}$.^{21,218,219} The substitution of electrical energy from power grid to the renewable salinity gradient for direct driving of Fenton process is the main advantage of MREC-Fenton system.

Previously MEFS were mostly tested with artificial WW, so there is a need to operationalize the reactors with real wastewaters. At laboratory stage it was 0.25 kW h m^{-3} or $0.40 \text{ kW per h per kg}$ of COD/ 0.18 kW h m^{-3} and $0.12 \text{ kW per h per kg}$ of COD in terms of energy consumption for the reactors with pure substrates such as acetate and glucose respectively and was significantly higher compared to domestic [0.04 kW h m^{-3} or $0.17 \text{ kW per h per kg}$ of COD] or industrial [0.01 kW h m^{-3} or $0.04 \text{ kW per h per kg}$ of COD] wastewaters.²⁴³ But systems performance was still lower in power density that could be insufficient enough for large scale industrial plants.²⁶⁰ So there is a need to optimize the MEFS with real WW to apply it at large scale. Moreover, life cycle assessment of the most efficient microbial-electron-Fenton systems is vital to operationalize them at commercial level.²⁶¹

7 Conclusions

Microbial electro Fenton technology for dye wastewater treatment have emerged as scientific inquisitiveness and are proved to be innovative, multidimensional approach that can be operated under batch and continuous mode where chances of treating domestic and dye wastewater can go side by side. Comparatively, the system is preferable to other methods due to its higher efficiency in terms of time and treatment. Characteristics, classification, environmental fate, toxicity and degradation mechanism of dyes have been assessed critically. Various types and operational models of MEFS for dyes wastewater treatment have also been reviewed. MFC being a platform technology has been greatly explored. In past, MFC based MEFS studies have shown to reduce the operational time from days to hours, even minutes, along with mineralization rate up to 90% and even above in case of dyes. Being an innovative technology,



dual advantages of electricity production along with dye decolorization simultaneously have also been reported by *in situ*-designed MFC based Fenton process and generated electricity has been utilized to run *ex situ* Fenton process in continuous and batch operations. The scope of technology has broadened as it is heading toward bio-monitoring, paving way for upcoming competitive approaches. Despite promising, it still faces several challenges of reactor configuration, high electrode/membrane cost, iron toxicity and information on large scale application. Therefore, extensive research and investigation is indispensable for future exploration and upgradation.

In future studies, there could be an opportunity of replacing chemical catalyst by biological catalysts produced by microorganisms in cathode chamber to accelerate and improve the pollutant treatment. This can mitigate the predicament of iron toxicity along with the provision of other beneficial opportunities.

Conflicts of interest

There are no conflicts to declare.

References

- 1 M. Berradi, R. Hsissou, M. Khudhair, M. Assouag, O. Cherkaoui, A. El Bachiri, *et al.*, Textile finishing dyes and their impact on aquatic environs, *Heliyon*, 2019, 5(11), e02711.
- 2 V. San, V. Spoann and J. Schmidt, Industrial pollution load assessment in Phnom Penh, Cambodia using an industrial pollution projection system, *Sci. Total Environ.*, 2018, **615**, 990–999.
- 3 P. Nidheesh, M. Zhou and M. A. Oturan, An overview on the removal of synthetic dyes from water by electrochemical advanced oxidation processes, *Chemosphere*, 2018, **197**, 210–227.
- 4 P. R. Sahoo, K. Prakash and S. Kumar, Light controlled receptors for heavy metal ions, *Coord. Chem. Rev.*, 2018, **357**, 18–49.
- 5 B. Lellis, C. Z. Fávaro-Polonio, J. A. Pamphile and J. C. Polonio, Effects of textile dyes on health and the environment and bioremediation potential of living organisms, *Biotechnol. Res. Int.*, 2019, 3(2), 275–290.
- 6 Z. Carmen and S. Daniela, *Textile organic dyes-characteristics, polluting effects and separation/elimination procedures from industrial effluents-a critical overview*, IntechOpen Rijeka, 2012.
- 7 M. Hassan, A. S. Fernandez, I. San Martin, B. Xie and A. Moran, Hydrogen evolution in microbial electrolysis cells treating landfill leachate: Dynamics of anodic biofilm, *Int. J. Hydrogen Energy*, 2018, **43**(29), 13051–13063.
- 8 S.-L. Lim, W.-L. Chu and S.-M. Phang, Use of *Chlorella vulgaris* for bioremediation of textile wastewater, *Bioresour. Technol.*, 2010, **101**(19), 7314–7322.
- 9 M. Hassan, H. Wei, H. Qiu, Y. Su, S. W. H. Jaafry, L. Zhan, *et al.*, Power generation and pollutants removal from landfill leachate in microbial fuel cell: variation and influence of anodic microbiomes, *Bioresour. Technol.*, 2018, **247**, 434–442.
- 10 E. Brillas, I. Sirés and M. A. Oturan, Electro-Fenton process and related electrochemical technologies based on Fenton's reaction chemistry, *Chem. Rev.*, 2009, **109**(12), 6570–6631.
- 11 N. Chaukura, W. Gwenzi, N. Tavengwa and M. M. Manyuchi, Biosorbents for the removal of synthetic organics and emerging pollutants: opportunities and challenges for developing countries, *Environ. Dev.*, 2016, **19**, 84–89.
- 12 D. Bhatia, N. R. Sharma, J. Singh and R. S. Kanwar, Biological methods for textile dye removal from wastewater: A review, *Crit. Rev. Environ. Sci. Technol.*, 2017, **47**(19), 1836–1876.
- 13 S. N. Banu, Comparative Study Of Photocatalytic Degradation Of Methylene Blue And Methyl Orange In Presence Of Solar Light Using Green Synthesized CuO nanocatalyst, *International Journal of Engineering Applied Sciences and Technology*, 2019, **4**(8), 365–370.
- 14 J. J. Pignatello, E. Oliveros and A. MacKay, Advanced oxidation processes for organic contaminant destruction based on the Fenton reaction and related chemistry, *Crit. Rev. Environ. Sci. Technol.*, 2006, **36**(1), 1–84.
- 15 F. Sun, H. Liu, B. Liang, R. Song, Q. Yan and A. Wang, Reductive degradation of chloramphenicol using bioelectrochemical system (BES): a comparative study of abiotic cathode and biocathode, *Bioresour. Technol.*, 2013, **143**, 699–702.
- 16 M. T. Yagub, T. K. Sen, S. Afroze and H. M. Ang, Dye and its removal from aqueous solution by adsorption: a review, *Adv. Colloid Interface Sci.*, 2014, **209**, 172–184.
- 17 A. Asghar, A. A. A. Raman and W. M. A. W. Daud, Challenges and recommendations for using membranes in wastewater-based microbial fuel cells for in situ Fenton oxidation for textile wastewater treatment, *Rev. Chem. Eng.*, 2015, **31**(1), 45–67.
- 18 F. Yu, M. Zhou and X. Yu, Cost-effective electro-Fenton using modified graphite felt that dramatically enhanced on H₂O₂ electro-generation without external aeration, *Electrochim. Acta*, 2015, **163**, 182–189.
- 19 S. Saroj, K. Kumar, N. Pareek, R. Prasad and R. P. Singh, Biodegradation of azo dyes Acid Red 183, Direct Blue 15 and Direct Red 75 by the isolate *Penicillium oxalicum* SAR-3, *Chemosphere*, 2014, **107**, 240–248.
- 20 Y. Mu, K. Rabaey, R. A. Rozendal, Z. Yuan and J. Keller, Decolorization of azo dyes in bioelectrochemical systems, *Environ. Sci. Technol.*, 2009, **43**(13), 5137–5143.
- 21 X. Li, S. Chen, I. Angelidaki and Y. Zhang, Bio-electro-Fenton processes for wastewater treatment: Advances and prospects, *Chem. Eng. J.*, 2018, **354**, 492–506.
- 22 V. Kiran and B. Gaur, Microbial fuel cell: technology for harvesting energy from biomass, *Rev. Chem. Eng.*, 2013, **29**(4), 189–203.
- 23 M. Hassan, N. Pous, B. Xie, J. Colprim, M. D. Balaguer and S. Puig, Employing Microbial Electrochemical Technology-



- driven electro-Fenton oxidation for the removal of recalcitrant organics from sanitary landfill leachate, *Bioresour. Technol.*, 2017, **243**, 949–956.
- 24 W. Wang, Y. Lu, H. Luo, G. Liu, R. Zhang and S. Jin, A microbial electro-Fenton cell for removing carbamazepine in wastewater with electricity output, *Water Res.*, 2018, **139**, 58–65.
 - 25 H. Wang and Z. J. Ren, A comprehensive review of microbial electrochemical systems as a platform technology, *Biotechnol. Adv.*, 2013, **31**(8), 1796–1807.
 - 26 B. E. Logan, M. J. Wallack, K.-Y. Kim, W. He, Y. Feng and P. E. Saikaly, Assessment of microbial fuel cell configurations and power densities, *Environ. Sci. Technol. Lett.*, 2015, **2**(8), 206–214.
 - 27 Y. Zhang, Y. Wang and I. Angelidaki, Alternate switching between microbial fuel cell and microbial electrolysis cell operation as a new method to control H₂O₂ level in Bioelectro-Fenton system, *J. Power Sources*, 2015, **291**, 108–116.
 - 28 S. H. Lin and C. C. Chang, Treatment of landfill leachate by combined electro-Fenton oxidation and sequencing batch reactor method, *Water Res.*, 2000, **34**(17), 4243–4249.
 - 29 X. Zhu and J. Ni, Simultaneous processes of electricity generation and p-nitrophenol degradation in a microbial fuel cell, *Electrochem. Commun.*, 2009, **11**(2), 274–277.
 - 30 R. A. Rozendal, E. Leone, J. Keller and K. Rabaey, Efficient hydrogen peroxide generation from organic matter in a bioelectrochemical system, *Electrochem. Commun.*, 2009, **11**(9), 1752–1755.
 - 31 X. Zhu and B. E. Logan, Using single-chamber microbial fuel cells as renewable power sources of electro-Fenton reactors for organic pollutant treatment, *J. Hazard Mater.*, 2013, **252**, 198–203.
 - 32 X. Li, X. Jin, N. Zhao, I. Angelidaki and Y. Zhang, Novel bio-electro-Fenton technology for azo dye wastewater treatment using microbial reverse-electrodialysis electrolysis cell, *Bioresour. Technol.*, 2017, **228**, 322–329.
 - 33 X. Li, I. Angelidaki and Y. Zhang, Salinity-gradient energy driven microbial electrosynthesis of hydrogen peroxide, *J. Power Sources*, 2017, **341**, 357–365.
 - 34 A. Asghar, A. A. Abdul Raman and W. M. A. W. Daud, Recent advances, challenges and prospects of in situ production of hydrogen peroxide for textile wastewater treatment in microbial fuel cells, *J. Chem. Technol. Biotechnol.*, 2014, **89**(10), 1466–1480.
 - 35 G.-E. Yuan, Y. Li, J. Lv, G. Zhang and F. Yang, Integration of microbial fuel cell and catalytic oxidation reactor with iron phthalocyanine catalyst for Congo red degradation, *Biochem. Eng. J.*, 2017, **120**, 118–124.
 - 36 M. F. de Dios, O. Iglesias, E. Bocos, M. Pazos and M. Sanromán, Application of benthonic microbial fuel cells and electro-Fenton process to dye decolourisation, *J. Ind. Eng. Chem.*, 2014, **20**(5), 3754–3760.
 - 37 L. Zhang, X. Yin and S. F. Y. Li, Bio-electrochemical degradation of paracetamol in a microbial fuel cell-Fenton system, *Chem. Eng. J.*, 2015, **276**, 185–192.
 - 38 Y. Wang, C. Feng, Y. Li, J. Gao and C.-P. Yu, Enhancement of emerging contaminants removal using Fenton reaction driven by H₂O₂-producing microbial fuel cells, *Chem. Eng. J.*, 2017, **307**, 679–686.
 - 39 H. Nadais, X. Li, N. Alves, C. Couras, H. R. Andersen, I. Angelidaki, *et al.*, Bio-electro-Fenton process for the degradation of non-steroidal anti-inflammatory drugs in wastewater, *Chem. Eng. J.*, 2018, **338**, 401–410.
 - 40 M. Hassan, X. Wang, F. Wang, D. Wu, A. Hussain and B. Xie, Coupling ARB-based biological and photochemical (UV/TiO₂ and UV/S₂O₈²⁻) techniques to deal with sanitary landfill leachate, *Waste Manag.*, 2017, **63**, 292–298.
 - 41 N. Wang, T. Zheng, G. Zhang and P. Wang, A review on Fenton-like processes for organic wastewater treatment, *J. Chem. Environ. Eng.*, 2016, **4**(1), 762–787.
 - 42 S. Gligorovski, R. Strekowski, S. Barbati and D. Vione, Environmental implications of hydroxyl radicals ([•]OH), *Chem. Rev.*, 2015, **115**(24), 13051–13092.
 - 43 T. A. Nguyen, C.-C. Fu and R.-S. Juang, Effective removal of sulfur dyes from water by biosorption and subsequent immobilized laccase degradation on crosslinked chitosan beads, *Chem. Eng. J.*, 2016, **304**, 313–324.
 - 44 R. Zou, I. Angelidaki, B. Jin and Y. Zhang, Feasibility and applicability of the scaling-up of bio-electro-Fenton system for textile wastewater treatment, *Environ. Int.*, 2020, **134**, 105352.
 - 45 D. Suteu, C. Zaharia and T. Malutan, Removal of orange 16 reactive dye from aqueous solutions by waste sunflower seed shells, *J. Serb. Chem. Soc.*, 2011, **76**(4), 607–624.
 - 46 T. Robinson, G. McMullan, R. Marchant and P. Nigam, Remediation of dyes in textile effluent: a critical review on current treatment technologies with a proposed alternative, *Bioresour. Technol.*, 2001, **77**(3), 247–255.
 - 47 C. R. Holkar, A. J. Jadhav, D. V. Pinjari, N. M. Mahamuni and A. B. Pandit, A critical review on textile wastewater treatments: possible approaches, *J. Environ. Manag.*, 2016, **182**, 351–366.
 - 48 M. Alkan, S. Çelikçapa, Ö. Demirbaş and M. Doğan, Removal of reactive blue 221 and acid blue 62 anionic dyes from aqueous solutions by sepiolite, *Dyes Pigm.*, 2005, **65**(3), 251–259.
 - 49 A. Paz, J. Carballo, M. J. Pérez and J. M. Domínguez, Biological treatment of model dyes and textile wastewaters, *Chemosphere*, 2017, **181**, 168–177.
 - 50 Z. Ahmad, S. M. Abdullah and K. Sulaiman, Temperature-sensitive chemical cell based on Nickel (II) phthalocyanine-tetrasulfonic acid tetrasodium salt, *Sensor Actuator Phys.*, 2012, **179**, 146–150.
 - 51 X. Wang, H. Mao, W. Huang, W. Guan, X. Zou, J. Pan, *et al.*, Preparation of magnetic imprinted polymer particles via microwave heating initiated polymerization for selective enrichment of 2-amino-4-nitrophenol from aqueous solution, *Chem. Eng. J.*, 2011, **178**, 85–92.
 - 52 S. Akiyama, N. Si, K. Nakashima and S. Yamasaki, Diphenylmethane and triphenylmethane dye ethynovinologues with absorption bands in the near-infrared, *Dyes Pigments*, 1988, **9**(6), 459–466.



- 53 G. Moussavi and M. Mahmoudi, Removal of azo and anthraquinone reactive dyes from industrial wastewaters using MgO nanoparticles, *J. Hazard Mater.*, 2009, **168**(2–3), 806–812.
- 54 V. K. Puttaswamy and N. Gowda, Oxidation of CI Acid Red 27 by chloramine-T in perchloric acid medium: spectrophotometric, kinetic and mechanistic approaches, *Dyes Pigment.*, 2008, **78**(2), 131–138.
- 55 D. Ž. Mijin, M. L. A. Ivić, A. E. Onjia and B. N. Grgur, Decolorization of textile dye CI Basic Yellow 28 with electrochemically generated active chlorine, *Chem. Eng. J.*, 2012, **204**, 151–157.
- 56 M. A. Hassaan, A. El Nemr and F. F. Madkour, Testing the advanced oxidation processes on the degradation of Direct Blue 86 dye in wastewater, *Egypt. J. Aquat. Res.*, 2017, **43**(1), 11–19.
- 57 D. Rawat, R. S. Sharma, S. Karmakar, L. S. Arora and V. Mishra, Ecotoxic potential of a presumably non-toxic azo dye, *Ecotoxicol. Environ. Saf.*, 2018, **148**, 528–537.
- 58 B. Mella, B. SdC. Barcellos, D. E. da Silva Costa and M. Gutterres, Treatment of leather dyeing wastewater with associated process of coagulation-flocculation/adsorption/ozonation, *Ozone: Sci. Eng.*, 2018, **40**(2), 133–140.
- 59 S. O. Ganiyu, C. A. Martinez-Huitle and M. A. Rodrigo, Renewable energies driven electrochemical wastewater/soil decontamination technologies: a critical review of fundamental concepts and applications, *Appl. Catal., B*, 2020, **270**, 118857.
- 60 S. Sandhya, Biodegradation of azo dyes under anaerobic condition: role of azoreductase, *Biodegradation of azo dyes*, 2010:39–57.
- 61 C. Zaharia, D. Suteu and A. Muresan, Options and solutions for textile effluent decolorization using some specific physico-chemical treatment steps, *Environ. Eng. Manage. J.*, 2012, **11**(2), 493–509.
- 62 H. Börnick and T. C. Schmidt, *Amines. Organic pollutants in the water cycle: properties, occurrence, analysis and environmental relevance of polar compounds*, 2006, pp. 181–209.
- 63 J. Khatri, P. Nidheesh, T. A. Singh and M. S. Kumar, Advanced oxidation processes based on zero-valent aluminium for treating textile wastewater, *Chem. Eng. J.*, 2018, **348**, 67–73.
- 64 B. Sharma, A. K. Dangi and P. Shukla, Contemporary enzyme based technologies for bioremediation: a review, *J. Environ. Manag.*, 2018, **210**, 10–22.
- 65 S. Khan and A. Malik, Toxicity evaluation of textile effluents and role of native soil bacterium in biodegradation of a textile dye, *Environ. Sci. Pollut. Res.*, 2018, **25**(5), 4446–4458.
- 66 M. Clark, *Handbook of textile and industrial dyeing: principles, processes and types of dyes*, Elsevier, 2011.
- 67 R. Christie, *Environmental aspects of textile dyeing*, Elsevier, 2007.
- 68 F. Copaciu, O. Opriş, V. Coman, D. Ristoiu, Ü. Niinemets and L. Copolovici, Diffuse water pollution by anthraquinone and azo dyes in environment importantly alters foliage volatiles, carotenoids and physiology in wheat (*Triticum aestivum*), *Water, Air, Soil Pollut.*, 2013, **224**(3), 1–11.
- 69 K. Hunger, *Industrial dyes: chemistry, properties, applications*, John Wiley & Sons, 2007.
- 70 J. Mathieu-Denoncourt, C. J. Martyniuk, S. R. de Solla, V. K. Balakrishnan and L. VrS, Sediment contaminated with the azo dye disperse yellow 7 alters cellular stress- and androgen-related transcription in *Silurana tropicalis* larvae, *Environ. Sci. Technol.*, 2014, **48**(5), 2952–2961.
- 71 I. S. Thakur, *Environmental biotechnology: basic concepts and applications*, IK International, 2006.
- 72 I. Haq and A. Raj, Biodegradation of Azure-B dye by *Serratia liquefaciens* and its validation by phytotoxicity, genotoxicity and cytotoxicity studies, *Chemosphere*, 2018, **196**, 58–68.
- 73 A. Y. Khan and G. S. Kumar, Spectroscopic studies on the binding interaction of phenothiazinium dyes, azure A and azure B to double stranded RNA polynucleotides, *Spectrochim. Acta, Part A*, 2016, **152**, 417–425.
- 74 A. Petzer, B. H. Harvey, G. Wegener and J. P. Petzer, Azure B, a metabolite of methylene blue, is a high-potency, reversible inhibitor of monoamine oxidase, *Toxicol. Appl. Pharmacol.*, 2012, **258**(3), 403–409.
- 75 S. A. Factor and W. J. Weiner, *Parkinson's disease: diagnosis and clinical management*, 2007.
- 76 G. Di Giovanni, V. Di Matteo and E. Esposito, *Serotonin-dopamine interaction: experimental evidence and therapeutic relevance*, Elsevier, 2008.
- 77 P. Paul and G. S. Kumar, Thermodynamics of the DNA binding of phenothiazinium dyes toluidine blue O, azure A and azure B, *J. Chem. Thermodyn.*, 2013, **64**, 50–57.
- 78 N. Couto, J. Wood and J. Barber, The role of glutathione reductase and related enzymes on cellular redox homeostasis network, *Free Radicals Biol. Med.*, 2016, **95**, 27–42.
- 79 F. M. D. Chequer, J. P. F. Angeli, E. R. A. Ferraz, M. S. Tsuboy, J. C. Marcarini, M. S. Mantovani, *et al.*, The azo dyes Disperse Red 1 and Disperse Orange 1 increase the micronuclei frequencies in human lymphocytes and in HepG2 cells, *Mutat. Res., Genet. Toxicol. Environ. Mutagen.*, 2009, **676**(1–2), 83–86.
- 80 Y. Will, J. E. McDuffie, A. J. Olaharski and B. D. Jeffy, *Drug discovery toxicology: From target assessment to translational biomarkers*, John Wiley & Sons, 2016.
- 81 F. H. Fernandes, E. Bustos-Obregon and D. M. F. Salvadori, Disperse Red 1 (textile dye) induces cytotoxic and genotoxic effects in mouse germ cells, *Reprod. Toxicol.*, 2015, **53**, 75–81.
- 82 M. C. Duarte and M. Rai, *Therapeutic medicinal plants: from lab to the market*, CRC Press, 2015.
- 83 F. M. D. Chequer, T. M. Lizier, R. de Felício, M. V. B. Zanoni, H. M. Deboni, N. P. Lopes, *et al.*, Analyses of the genotoxic and mutagenic potential of the products formed after the biotransformation of the azo dye Disperse Red 1, *Toxicol. in Vitro*, 2011, **25**(8), 2054–2063.
- 84 F. I. Vacchi, P. C. Von der Ohe, A. F. de Albuquerque, J. A. de Souza Vendemiatti, C. C. J. Azevedo, J. G. Honório, *et al.*,



- Occurrence and risk assessment of an azo dye—The case of Disperse Red 1, *Chemosphere*, 2016, **156**, 95–100.
- 85 E. A. Petrakis, L. R. Cagliani, P. A. Tarantilis, M. G. Polissiou and R. Consonni, Sudan dyes in adulterated saffron (*Crocus sativus* L.): Identification and quantification by ¹H NMR, *Food Chem.*, 2017, **217**, 418–424.
 - 86 O. Duman, S. Tunc and T. G. Polat, Adsorptive removal of triarylmethane dye (Basic Red 9) from aqueous solution by sepiolite as effective and low-cost adsorbent, *Microporous Mesoporous Mater.*, 2015, **210**, 176–184.
 - 87 M. V. Foguel, X.-A. Ton, M. V. Zanoni, T. S. Maria Del Pilar, K. Haupt and B. T. S. Bui, A molecularly imprinted polymer-based evanescent wave fiber optic sensor for the detection of basic red 9 dye, *Sens. Actuators, B*, 2015, **218**, 222–228.
 - 88 R. P. Pohanish, *Sittig's handbook of toxic and hazardous chemicals and carcinogens*, William Andrew, 2017.
 - 89 H. Ali, S. Shehata and K. Ramadan, Microbial decolorization and degradation of crystal violet dye by *Aspergillus niger*, *Int. J. Environ. Sci. Technol.*, 2016, **13**(12), 2917–2926.
 - 90 S. Mani and R. N. Bharagava, Exposure to crystal violet, its toxic, genotoxic and carcinogenic effects on environment and its degradation and detoxification for environmental safety, *Rev. Environ. Contam. Toxicol.*, 2016, **237**, 71–104.
 - 91 R. A. Rozendal, H. V. Hamelers, K. Rabaey, J. Keller and C. J. Buisman, Towards practical implementation of bioelectrochemical wastewater treatment, *Trends Biotechnol.*, 2008, **26**(8), 450–459.
 - 92 L. Fu, S.-J. You, G.-q Zhang, F.-L. Yang and X.-h Fang, Degradation of azo dyes using in situ Fenton reaction incorporated into H₂O₂-producing microbial fuel cell, *Chem. Eng. J.*, 2010, **160**(1), 164–169.
 - 93 J.-y Chen, N. Li and L. Zhao, Three-dimensional electrode microbial fuel cell for hydrogen peroxide synthesis coupled to wastewater treatment, *J. Power Sources*, 2014, **254**, 316–322.
 - 94 J. B. Arends, S. Van Denhouwe, W. Verstraete, N. Boon and K. Rabaey, Enhanced disinfection of wastewater by combining wetland treatment with bioelectrochemical H₂O₂ production, *Bioresour. Technol.*, 2014, **155**, 352–358.
 - 95 D. E. Meerooff, F. Bloetscher, D. Reddy, F. Gasnier, S. Jain, A. McBarnette, *et al.*, Application of photochemical technologies for treatment of landfill leachate, *J. Hazard Mater.*, 2012, **209**, 299–307.
 - 96 M. Hassan, Y. Zhao and B. Xie, Employing TiO₂ photocatalysis to deal with landfill leachate: Current status and development, *Chem. Eng. J.*, 2016, **285**, 264–275.
 - 97 S. Freguia, K. Rabaey, Z. Yuan and J. Keller, Syntrophic processes drive the conversion of glucose in microbial fuel cell anodes, *Environ. Sci. Technol.*, 2008, **42**(21), 7937–7943.
 - 98 A. P. Borole and J. R. Mielenz, Estimating hydrogen production potential in biorefineries using microbial electrolysis cell technology, *Int. J. Hydrogen Energy*, 2011, **36**(22), 14787–14795.
 - 99 J. F. Miceli III, I. Garcia-Peña, P. Parameswaran, C. I. Torres and R. Krajmalnik-Brown, Combining microbial cultures for efficient production of electricity from butyrate in a microbial electrochemical cell, *Bioresour. Technol.*, 2014, **169**, 169–174.
 - 100 P. Parameswaran, H. Zhang, C. I. Torres, B. E. Rittmann and R. Krajmalnik-Brown, Microbial community structure in a biofilm anode fed with a fermentable substrate: the significance of hydrogen scavengers, *Biotechnol. Bioeng.*, 2010, **105**(1), 69–78.
 - 101 G. Lyberatos and I. Skiadas, Modelling of anaerobic digestion—a review, *Global NEST J.*, 1999, **1**(2), 63–76.
 - 102 G. Merlino, A. Rizzi, A. Schievano, A. Tenca, B. Scaglia, R. Oberti, *et al.*, Microbial community structure and dynamics in two-stage vs. single-stage thermophilic anaerobic digestion of mixed swine slurry and market bio-waste, *Water Res.*, 2013, **47**(6), 1983–1995.
 - 103 P. Weiland, Biogas production: current state and perspectives, *Appl. Microbiol. Biotechnol.*, 2010, **85**(4), 849–860.
 - 104 W. Parawira, J. S. Read, B. Mattiasson and L. Björnsson, Energy production from agricultural residues: high methane yields in pilot-scale two-stage anaerobic digestion, *Biomass Bioenergy*, 2008, **32**(1), 44–50.
 - 105 R. Conrad, Contribution of hydrogen to methane production and control of hydrogen concentrations in methanogenic soils and sediments, *FEMS Microbiol. Ecol.*, 1999, **28**(3), 193–202.
 - 106 M. Bryant, Microbial methane production—theoretical aspects, *J. Anim. Sci.*, 1979, **48**(1), 193–201.
 - 107 P. Smith, The microbial ecology of sludge methanogenesis, *Dev. Ind. Microbiol.*, 1966, **7**, 156–161.
 - 108 P. Claassen, J. Van Lier, A. Lopez Contreras, E. Van Niel, L. Sijtsma, A. Stams, *et al.*, Utilisation of biomass for the supply of energy carriers, *Appl. Microbiol. Biotechnol.*, 1999, **52**(6), 741–755.
 - 109 I. Lyberatos, G. Ntaikou and G. Antonopoulou, Biohydrogen Production from Biomass and Wastes via Dark Fermentation: A Review, *Waste Biomass Valorization*, 2010, 21–39.
 - 110 B. Schink, Energetics of syntrophic cooperation in methanogenic degradation, *Microbiol. Mol. Biol. Rev.*, 1997, **61**(2), 262–280.
 - 111 F. A. De Bok, H. J. Harmsen, C. M. Plugge, M. C. de Vries, A. D. Akkermans, W. M. de Vos, *et al.*, The first true obligately syntrophic propionate-oxidizing bacterium, *Pelotomaculum schinkii* sp. nov., co-cultured with *Methanospirillum hungatei*, and emended description of the genus *Pelotomaculum*, *Int. J. Syst. Evol. Microbiol.*, 2005, **55**(4), 1697–1703.
 - 112 S. H. Zinder, *Physiological ecology of methanogens*, Springer. Methanogenesis, 1993. pp. 128–206.
 - 113 Y. Liu and W. B. Whitman, Metabolic, phylogenetic, and ecological diversity of the methanogenic archaea, *Ann. N. Y. Acad. Sci.*, 2008, **1125**(1), 171–189.
 - 114 L. Appels, J. Lauwers, J. Degreè, L. Helsen, B. Lievens, K. Willems, *et al.*, Anaerobic digestion in global bio-



- energy production: potential and research challenges, *Renewable Sustainable Energy Rev.*, 2011, **15**(9), 4295–4301.
- 115 Y. Xiao, Y. Zheng, S. Wu, E.-H. Zhang, Z. Chen, P. Liang, *et al.*, Pyrosequencing reveals a core community of anodic bacterial biofilms in bioelectrochemical systems from China, *Front. Microbiol.*, 2015, **6**, 1410.
 - 116 S. Galai, Pérez De Los Ríos A, Hernández-Fernández FJ, Mateo Ramírez F, Haj Kacem S and Quesada-Medina J, Application of microbial fuel cell for azoic dye decolorization with simultaneous bioenergy production using single bacterial strain, *Chem. Eng. Technol.*, 2015, **38**, 1511–1518.
 - 117 L. Shi, D. J. Richardson, Z. Wang, S. N. Kerisit, K. M. Rosso, J. M. Zachara, *et al.*, The roles of outer membrane cytochromes of *Shewanella* and *Geobacter* in extracellular electron transfer, *Environ. Microbiol. Rep.*, 2009, **1**(4), 220–227.
 - 118 Y. Yang, M. Xu, Z. He, J. Guo, G. Sun and J. Zhou, Microbial electricity generation enhances decabromodiphenyl ether (BDE-209) degradation, *PLoS One*, 2013, **8**(8), e70686.
 - 119 Y. Yang, O. Luo, G. Kong, B. Wang, X. Li, E. Li, *et al.*, Deciphering the anode-enhanced azo dye degradation in anaerobic baffled reactors integrating with microbial fuel cells, *Front. Microbiol.*, 2018, **9**, 2117.
 - 120 D. Jin, X. Kong, Y. Li, Z. Bai, G. Zhuang, X. Zhuang, *et al.*, Biodegradation of di-n-butyl phthalate by *Achromobacter* sp. isolated from rural domestic wastewater, *Int. J. Environ. Res. Public Health*, 2015, **12**(10), 13510–13522.
 - 121 D. Paul, S. K. Kazy, T. D. Banerjee, A. K. Gupta, T. Pal and P. Sar, Arsenic biotransformation and release by bacteria indigenous to arsenic contaminated groundwater, *Bioresour. Technol.*, 2015, **188**, 14–23.
 - 122 L. Hao, B. Zhang, M. Cheng and C. Feng, Effects of various organic carbon sources on simultaneous V (V) reduction and bioelectricity generation in single chamber microbial fuel cells, *Bioresour. Technol.*, 2016, **201**, 105–110.
 - 123 D. R. Lovley, The microbe electric: conversion of organic matter to electricity, *Curr. Opin. Biotechnol.*, 2008, **19**(6), 564–571.
 - 124 K. V. Krivushin, V. A. Shcherbakova, L. E. Petrovskaya and E. M. Rivkina, *Methanobacterium veterum* sp. nov., from ancient Siberian permafrost, *Int. J. Syst. Evol. Microbiol.*, 2010, **60**(2), 455–459.
 - 125 T. Looft, U. Levine and T. Stanton, *Cloacibacillus porcorum* sp. nov., a mucin-degrading bacterium from the swine intestinal tract and emended description of the genus *Cloacibacillus*, *Int. J. Syst. Evol. Microbiol.*, 2013, **63**(Pt 6), 1960.
 - 126 L. Fu, S. J. You, Fl Yang, Mm Gao, Xh Fang and Gq Zhang, Synthesis of hydrogen peroxide in microbial fuel cell, *J. Chem. Technol. Biotechnol.*, 2010, **85**(5), 715–719.
 - 127 K. Solanki, S. Subramanian and S. Basu, Microbial fuel cells for azo dye treatment with electricity generation: a review, *Bioresour. Technol.*, 2013, **131**, 564–571.
 - 128 J. Svojitka, L. Dvořák, M. Studer, J. O. Straub, H. Frömel and T. Wintgens, Performance of an anaerobic membrane bioreactor for pharmaceutical wastewater treatment, *Bioresour. Technol.*, 2017, **229**, 180–189.
 - 129 M. Katz, T. Frejd, B. Hahn-Hägerdal and M. F. Gorwa-Grauslund, Efficient anaerobic whole cell stereoselective bioreduction with recombinant *Saccharomyces cerevisiae*, *Biotechnol. Bioeng.*, 2003, **84**(5), 573–582.
 - 130 S. D. Minteer, B. Y. Liaw and M. J. Cooney, Enzyme-based biofuel cells, *Curr. Opin. Biotechnol.*, 2007, **18**(3), 228–234.
 - 131 F. Davis and S. P. Higson, Biofuel cells—recent advances and applications, *Biosens. Bioelectron.*, 2007, **22**(7), 1224–1235.
 - 132 N. Mano, F. Mao and A. Heller, A miniature biofuel cell operating in a physiological buffer, *J. Am. Chem. Soc.*, 2002, **124**(44), 12962–12963.
 - 133 A. Chaubey and B. Malhotra, Mediated biosensors, *Biosens. Bioelectron.*, 2002, **17**(6–7), 441–456.
 - 134 A. Dubinin, F. Li, Y. Li and J. Yu, A solid-state immobilized enzyme polymer membrane microelectrode for measuring lactate-ion concentration, *J. Electroanal. Chem. Interfacial Electrochem.*, 1991, **320**(1), 131–135.
 - 135 A. A. Karyakin, O. V. Gitelmacher and E. E. Karyakina, Prussian blue-based first-generation biosensor. A sensitive amperometric electrode for glucose, *Anal. Chem.*, 1995, **67**(14), 2419–2423.
 - 136 T. Aoyagi, A. Nakamura, H. Ikeda, T. Ikeda, H. Mihara and A. Ueno, Alizarin yellow-modified β -cyclodextrin as a guest-responsive absorption change sensor, *Anal. Chem.*, 1997, **69**(4), 659–663.
 - 137 B. Brunetti, P. Ugo, L. M. Moretto and C. R. Martin, Electrochemistry of phenothiazine and methylviologen biosensor electron-transfer mediators at nanoelectrode ensembles, *J. Electroanal. Chem.*, 2000, **491**(1–2), 166–174.
 - 138 B.-Y. Chen, C.-C. Hsueh, S.-Q. Liu, I.-S. Ng and Y.-M. Wang, Deciphering mediating characteristics of decolorized intermediates for reductive decolorization and bioelectricity generation, *Bioresour. Technol.*, 2013, **145**, 321–325.
 - 139 V. Razumas, J. Jasaitis and J. Kulys, Electrocatalysis on enzyme-modified carbon materials, *Bioelectrochem. Bioenerg.*, 1984, **12**(3–4), 297–322.
 - 140 B. P. Tarasevich and E. V. Kuznetsov, Progress in the polymer chemistry of borates, *Russ. Chem. Rev.*, 1987, **56**(3), 203.
 - 141 J. Celis, *2-D Proteome analysis protocols*, ed. A. J. Link, Humana Press, Totowa, New Jersey, 1999, p. 704, \$89.50 (hardcover), *Protein Science*, 1999, **8**, (10), 2218.
 - 142 E. Kjeang, D. Sinton and D. Harrington, Strategic enzyme patterning for microfluidic biofuel cells, *J. Power Sources*, 2006, **158**(1), 1–12.
 - 143 V. R. Stamenkovic, B. S. Mun, M. Arenz, K. J. Mayrhofer, C. A. Lucas, G. Wang, *et al.*, Trends in electrocatalysis on extended and nanoscale Pt-bimetallic alloy surfaces, *Nat. Mater.*, 2007, **6**(3), 241–247.
 - 144 J. L. Anderson, E. F. Bowden and P. G. Pickup, Dynamic electrochemistry: methodology and application, *Anal. Chem.*, 1996, **68**(12), 379–444.



- 145 S. Perkins, Electrode enhancements: New materials may boost fuel cell performance, *Sci. News*, 2007, **171**(2), 21–22.
- 146 R. H. Petrucci, F. G. Herring and J. D. Madura. *General chemistry: principles and modern applications*, Pearson Prentice Hall, 2010.
- 147 L. Gorton, E. Csöregi, E. Dominguez, J. Emnéus, G. Jönsson-Pettersson, G. Marko-Varga, *et al.*, Selective detection in flow analysis based on the combination of immobilized enzymes and chemically modified electrodes, *Anal. Chim. Acta*, 1991, **250**, 203–248.
- 148 A. Brigé, B. Motte, J. Borloo, G. Buyschaert, B. Devreese and J. J. Van Beeumen, Bacterial decolorization of textile dyes is an extracellular process requiring a multicomponent electron transfer pathway, *Microb. Biotechnol.*, 2008, **1**(1), 40–52.
- 149 Y.-G. Hong and J.-D. Gu, Physiology and biochemistry of reduction of azo compounds by *Shewanella* strains relevant to electron transport chain, *Appl. Microbiol. Biotechnol.*, 2010, **88**(3), 637–643.
- 150 P.-J. Cai, X. Xiao, Y.-R. He, W.-W. Li, J. Chu, C. Wu, *et al.*, Anaerobic biodecolorization mechanism of methyl orange by *Shewanella oneidensis* MR-1, *Appl. Microbiol. Biotechnol.*, 2012, **93**(4), 1769–1776.
- 151 C. Koch and F. Harnisch, What is the essence of microbial electroactivity?, *Front. Microbiol.*, 2016, **7**, 1890.
- 152 P. Mani, V. Fidal, K. Bowman, M. Breheny, T. Chandra, T. Keshavarz, *et al.*, Degradation of azo dye (acid orange 7) in a Microbial fuel cell: comparison between anodic microbial-mediated reduction and cathodic laccase-mediated oxidation, *Front. Energy Res.*, 2019, **7**, 101.
- 153 E. Fernando, T. Keshavarz and G. Kyazze, Complete degradation of the azo dye Acid Orange-7 and bioelectricity generation in an integrated microbial fuel cell, aerobic two-stage bioreactor system in continuous flow mode at ambient temperature, *Bioresour. Technol.*, 2014, **156**, 155–162.
- 154 N. Li, J. An, L. Zhou, T. Li, J. Li, C. Feng, *et al.*, A novel carbon black graphite hybrid air-cathode for efficient hydrogen peroxide production in bioelectrochemical systems, *J. Power Sources*, 2016, **306**, 495–502.
- 155 S. Li, H. Zhang, R. Lu and A. Yu, Interaction between triethanolamine and singlet or triplet excited state of xanthene dyes in aqueous solution, *Spectrochim. Acta, Part A*, 2017, **184**, 204–210.
- 156 H. Gao, Y. D. Scherson and G. F. Wells, Towards energy neutral wastewater treatment: methodology and state of the art, *Environ. Sci.: Processes Impacts*, 2014, **16**(6), 1223–1246.
- 157 D. Wang, H. Hou, J. Hu, J. Xu, L. Huang, S. Hu, *et al.*, A bio-electro-Fenton system with a facile anti-biofouling air cathode for efficient degradation of landfill leachate, *Chemosphere*, 2019, **215**, 173–181.
- 158 F. Van der Zee, G. Lettinga and J. Field, The role of (auto) catalysis in the mechanism of an anaerobic azo reduction, *Water Sci. Technol.*, 2000, **42**(5–6), 301–308.
- 159 M. A. Oturan, M. Pimentel, N. Oturan and I. Sirés, Reaction sequence for the mineralization of the short-chain carboxylic acids usually formed upon cleavage of aromatics during electrochemical Fenton treatment, *Electrochim. Acta*, 2008, **54**(2), 173–182.
- 160 H.-C. Tao, X.-Y. Wei, L.-J. Zhang, T. Lei and N. Xu, Degradation of p-nitrophenol in a BES-Fenton system based on limonite, *J. Hazard Mater.*, 2013, **254**, 236–241.
- 161 G. Divyapriya, I. M. Nambi and J. Senthilnathan, An innate quinone functionalized electrochemically exfoliated graphene/Fe₃O₄ composite electrode for the continuous generation of reactive oxygen species, *Chem. Eng. J.*, 2017, **316**, 964–977.
- 162 C. Feng, F. Li, H. Liu, X. Lang and S. Fan, A dual-chamber microbial fuel cell with conductive film-modified anode and cathode and its application for the neutral electro-Fenton process, *Electrochim. Acta*, 2010, **55**(6), 2048–2054.
- 163 C. Galindo, P. Jacques and A. Kalt, Photodegradation of the aminoazobenzene acid orange 52 by three advanced oxidation processes: UV/H₂O₂, UV/TiO₂ and VIS/TiO₂: comparative mechanistic and kinetic investigations, *J. Photochem. Photobiol., A*, 2000, **130**(1), 35–47.
- 164 I. K. Konstantinou and T. A. Albanis, TiO₂-assisted photocatalytic degradation of azo dyes in aqueous solution: kinetic and mechanistic investigations: a review, *Appl. Catal., B*, 2004, **49**(1), 1–14.
- 165 G. Reguera, K. D. McCarthy, T. Mehta, J. S. Nicoll, M. T. Tuominen and D. R. Lovley, Extracellular electron transfer via microbial nanowires, *Nature*, 2005, **435**(7045), 1098–1101.
- 166 H. Richter, K. P. Nevin, H. Jia, D. A. Lowy, D. R. Lovley and L. M. Tender, Cyclic voltammetry of biofilms of wild type and mutant *Geobacter sulfurreducens* on fuel cell anodes indicates possible roles of OmcB, OmcZ, type IV pili, and protons in extracellular electron transfer, *Energy Environ. Sci.*, 2009, **2**(5), 506–516.
- 167 J. Sim, J. An, E. Elbeshbishy, H. Ryu and H.-S. Lee, Characterization and optimization of cathodic conditions for H₂O₂ synthesis in microbial electrochemical cells, *Bioresour. Technol.*, 2015, **195**, 31–36.
- 168 J.-y Chen, L. Zhao, N. Li and H. Liu, A microbial fuel cell with the three-dimensional electrode applied an external voltage for synthesis of hydrogen peroxide from organic matter, *J. Power Sources*, 2015, **287**, 291–296.
- 169 S. Batool, S. Akib, M. Ahmad, K. S. Balkhair and M. A. Ashraf, Study of modern nano enhanced techniques for removal of dyes and metals, *J. Nanomaterial.*, 2014, **2014**, 20.
- 170 H. Lee, Y.-K. Park, S.-J. Kim, B.-H. Kim, H.-S. Yoon and S.-C. Jung, Rapid degradation of methyl orange using hybrid advanced oxidation process and its synergistic effect, *J. Ind. Eng. Chem.*, 2016, **35**, 205–210.
- 171 Y.-T. Wang and R.-S. Wang, A bio-electro-Fenton system employing the composite fepc/cnt/ss316 cathode, *Materials*, 2017, **10**(2), 169.
- 172 B. E. Logan, B. Hamelers, R. Rozendal, U. Schröder, J. Keller, S. Freguia, *et al.*, Microbial fuel cells: methodology and technology, *Environ. Sci. Technol.*, 2006, **40**(17), 5181–5192.



- 173 X.-W. Liu, X.-F. Sun, D.-B. Li, W.-W. Li, Y.-X. Huang, G.-P. Sheng, *et al.*, Anodic Fenton process assisted by a microbial fuel cell for enhanced degradation of organic pollutants, *Water Res.*, 2012, **46**(14), 4371–4378.
- 174 C.-H. Feng, F.-B. Li, H.-J. Mai and X.-Z. Li, Bio-electro-Fenton process driven by microbial fuel cell for wastewater treatment, *Environ. Sci. Technol.*, 2010, **44**(5), 1875–1880.
- 175 S. Li, T. Hua, F. Li and Q. Zhou, Bio-electro-Fenton systems for sustainable wastewater treatment: mechanisms, novel configurations, recent advances, LCA and challenges. An updated review, *J. Chem. Technol. Biotechnol.*, 2020, **95**(8), 2083–2097.
- 176 D. Jiang, M. Curtis, E. Troop, K. Scheible, J. McGrath, B. Hu, *et al.*, A pilot-scale study on utilizing multi-anode/cathode microbial fuel cells (MAC MFCs) to enhance the power production in wastewater treatment, *Int. J. Hydrogen Energy*, 2011, **36**(1), 876–884.
- 177 Z. Lv, D. Xie, X. Yue, C. Feng and C. Wei, Ruthenium oxide-coated carbon felt electrode: a highly active anode for microbial fuel cell applications, *J. Power Sources*, 2012, **210**, 26–31.
- 178 S. Chen, G. He, X. Hu, M. Xie, S. Wang, D. Zeng, *et al.*, A three-dimensionally ordered macroporous carbon derived from a natural resource as anode for microbial bioelectrochemical systems, *ChemSusChem*, 2012, **5**(6), 1059–1063.
- 179 S. Cheng and B. E. Logan, Ammonia treatment of carbon cloth anodes to enhance power generation of microbial fuel cells, *Electrochem. Commun.*, 2007, **9**(3), 492–496.
- 180 N. G. Yasri and G. Nakhla, Electrochemical behavior of anode-respiring bacteria on doped carbon electrodes, *ACS Appl. Mater. Interfaces*, 2016, **8**(51), 35150–35162.
- 181 D. Paul, M. Noori, P. Rajesh, M. Ghangrekar and A. Mitra, Modification of carbon felt anode with graphene oxide-zeolite composite for enhancing the performance of microbial fuel cell, *Sustain. Energy Technol. Assessments*, 2018, **26**, 77–82.
- 182 G. Huang, H. Wang, H. Zhao, P. Wu and Q. Yan, Application of polypyrrole modified cathode in bio-electro-Fenton coupled with microbial desalination cell (MDC) for enhanced degradation of methylene blue, *J. Power Sources*, 2018, **400**, 350–359.
- 183 T. Huang, L. Liu, J. Tao, L. Zhou and S. Zhang, Microbial fuel cells coupling with the three-dimensional electro-Fenton technique enhances the degradation of methyl orange in the wastewater, *Environ. Sci. Pollut. Res.*, 2018, **25**(18), 17989–18000.
- 184 C. Santoro, C. Arbizzani, B. Erable and I. Ieropoulos, Microbial fuel cells: From fundamentals to applications. A review, *J. Power Sources*, 2017, **356**, 225–244.
- 185 N. Yasri, E. P. Roberts and S. Gunasekaran, The electrochemical perspective of bioelectrocatalytic activities in microbial electrolysis and microbial fuel cells, *Energy Reports*, 2019, **5**, 1116–1136.
- 186 E. Guerrini, M. Grattieri, S. P. Trasatti, M. Bestetti and P. Cristiani, Performance explorations of single chamber microbial fuel cells by using various microelectrodes applied to biocathodes, *Int. J. Hydrogen Energy*, 2014, **39**(36), 21837–21846.
- 187 B. Logan, S. Cheng, V. Watson and G. Estadt, Graphite fiber brush anodes for increased power production in air-cathode microbial fuel cells, *Environ. Sci. Technol.*, 2007, **41**(9), 3341–3346.
- 188 L. Zhao, J. Li, F. Battaglia and Z. He, Computational investigation of the flow field contribution to improve electricity generation in granular activated carbon-assisted microbial fuel cells, *J. Power Sources*, 2016, **333**, 83–87.
- 189 M. Grattieri, S. Babanova, C. Santoro, E. Guerrini, S. P. Trasatti, P. Cristiani, *et al.*, Enzymatic oxygen microsensor based on bilirubin oxidase applied to microbial fuel cells analysis, *Electroanalysis*, 2015, **27**(2), 327–335.
- 190 P. Nidheesh and R. Rajan, Removal of rhodamine B from a water medium using hydroxyl and sulphate radicals generated by iron loaded activated carbon, *RSC advances*, 2016, **6**(7), 5330–5340.
- 191 P. Xu, H. Xu and Z. Shi, A novel bio-electro-Fenton process with FeVO₄/CF cathode on advanced treatment of coal gasification wastewater, *Sep. Purif. Technol.*, 2018, **194**, 457–461.
- 192 X.-Q. Wang, C.-P. Liu, Y. Yuan and F.-b Li, Arsenite oxidation and removal driven by a bio-electro-Fenton process under neutral pH conditions, *J. Hazard Mater.*, 2014, **275**, 200–209.
- 193 C. Li, L. Zhang, L. Ding, H. Ren and H. Cui, Effect of conductive polymers coated anode on the performance of microbial fuel cells (MFCs) and its biodiversity analysis, *Biosensors and Bioelectronics*, 2011, **26**(10), 4169–4176.
- 194 S. Hidayat, Y.-H. Song and J.-Y. Park, Performance of a continuous flow microbial reverse-electrodialysis electrolysis cell using a non-buffered substrate and catholyte effluent addition, *Bioresour. Technol.*, 2017, **240**, 77–83.
- 195 L. Zhuang, S. Zhou, Y. Yuan, M. Liu and Y. Wang, A novel bioelectro-Fenton system for coupling anodic COD removal with cathodic dye degradation, *Chem. Eng. J.*, 2010, **163**(1–2), 160–163.
- 196 S. B. Velasquez-Orta, T. P. Curtis and B. E. Logan, Energy from algae using microbial fuel cells, *Biotechnol. Bioeng.*, 2009, **103**(6), 1068–1076.
- 197 J. M. Pisciotto, Z. Zaybak, D. F. Call, J.-Y. Nam and B. E. Logan, Enrichment of microbial electrolysis cell biocathodes from sediment microbial fuel cell bioanodes, *Appl. Environ. Microbiol.*, 2012, **78**(15), 5212–5219.
- 198 H. Liu, R. Ramnarayanan and B. E. Logan, Production of electricity during wastewater treatment using a single chamber microbial fuel cell, *Environ. Sci. Technol.*, 2004, **38**(7), 2281–2285.
- 199 W. Miran, M. Nawaz, J. Jang and D. S. Lee, Chlorinated phenol treatment and in situ hydrogen peroxide production in a sulfate-reducing bacteria enriched bioelectrochemical system, *Water Res.*, 2017, **117**, 198–206.



- 200 G. Hernández-Flores, H. Poggi-Varaldo, O. Solorza-Feria, M. Ponce-Noyola, T. Romero-Castañón, N. Rinderknecht-Seijas, *et al.*, Characteristics of a single chamber microbial fuel cell equipped with a low cost membrane, *Int. J. Hydrogen Energy*, 2015, **40**(48), 17380–17387.
- 201 M. C. Hatzell, Y. Kim and B. E. Logan, Powering microbial electrolysis cells by capacitor circuits charged using microbial fuel cell, *J. Power Sources*, 2013, **229**, 198–202.
- 202 X. Yu, W. Fu, M. Jiang, G. Liu, Y. Zou and S. Chen, Automatic microbial electro-Fenton system driven by transpiration for degradation of acid orange 7, *Sci. Total Environ.*, 2020, **725**, 138508.
- 203 H. Olvera-Vargas, C. Trellu, N. Oturan and M. A. Oturan, *Bio-electro-Fenton: a new combined process—principles and applications*, Electro-Fenton Process, 2017, pp. 29–56.
- 204 Y. Luo, R. Zhang, G. Liu, J. Li, B. Qin, M. Li, *et al.*, Simultaneous degradation of refractory contaminants in both the anode and cathode chambers of the microbial fuel cell, *Bioresour. Technol.*, 2011, **102**(4), 3827–3832.
- 205 K. Zuo, L. Yuan, J. Wei, P. Liang and X. Huang, Competitive migration behaviors of multiple ions and their impacts on ion-exchange resin packed microbial desalination cell, *Bioresour. Technol.*, 2013, **146**, 637–642.
- 206 Y. Kim and B. E. Logan, Microbial desalination cells for energy production and desalination, *Desalination*, 2013, **308**, 122–130.
- 207 A. Ebrahimi, G. Najafpour and D. Kebria, Effect of batch vs. continuous mode of operation on microbial desalination cell performance treating municipal wastewater, *Iran. J. Hydrogen Fuel Cell*, 2016, **3**(4), 281–290.
- 208 H. Monteil, Y. Pechaud, N. Oturan and M. A. Oturan, A review on efficiency and cost effectiveness of electro-and bio-electro-Fenton processes: application to the treatment of pharmaceutical pollutants in water, *Chem. Eng. J.*, 2019, **376**, 119577.
- 209 P. Nidheesh and R. Gandhimathi, Textile wastewater treatment by electro-Fenton process in batch and continuous modes, *J. Hazard., Toxic Radioact. Waste*, 2015, **19**(3), 04014038.
- 210 M. Rahimnejad, G. Najafpour and A. A. Ghoreyshi, *Effect of mass transfer on performance of microbial fuel cell*, Intech. 2011, vol. 5, pp. 233–50.
- 211 M. Xu, J. Guo, X. Kong, X. Chen and G. Sun, Fe (III)-enhanced azo reduction by *Shewanella decolorationis* S12, *Appl. Microbiol. Biotechnol.*, 2007, **74**(6), 1342–1349.
- 212 M. Á. F. de Dios, A. G. del Campo, F. J. Fernández, M. Rodrigo, M. Pazos and M. Á. Sanromán, Bacterial-fungal interactions enhance power generation in microbial fuel cells and drive dye decolourisation by an ex situ and in situ electro-Fenton process, *Bioresour. Technol.*, 2013, **148**, 39–46.
- 213 T. Ling, B. Huang, M. Zhao, Q. Yan and W. Shen, Repeated oxidative degradation of methyl orange through bio-electro-Fenton in bioelectrochemical system (BES), *Bioresour. Technol.*, 2016, **203**, 89–95.
- 214 S. Yan, J. Geng, R. Guo, Y. Du and H. Zhang, Hydronium jarosite activation of peroxymonosulfate for the oxidation of organic contaminant in an electrochemical reactor driven by microbial fuel cell, *J. Hazard Mater.*, 2017, **333**, 358–368.
- 215 K. Rabaey, N. Boon, M. Höfte and W. Verstraete, Microbial phenazine production enhances electron transfer in biofuel cells, *Environ. Sci. Technol.*, 2005, **39**(9), 3401–3408.
- 216 W. He, M. J. Wallack, K.-Y. Kim, X. Zhang, W. Yang, X. Zhu, *et al.*, The effect of flow modes and electrode combinations on the performance of a multiple module microbial fuel cell installed at wastewater treatment plant, *Water Res.*, 2016, **105**, 351–360.
- 217 X. Li, X. Jin, N. Zhao, I. Angelidaki and Y. Zhang, Efficient treatment of aniline containing wastewater in bipolar membrane microbial electrolysis cell-Fenton system, *Water Res.*, 2017, **119**, 67–72.
- 218 G. Gao, Q. Zhang, Z. Hao and C. D. Vecitis, Carbon nanotube membrane stack for flow-through sequential regenerative electro-Fenton, *Environ. Sci. Technol.*, 2015, **49**(4), 2375–2383.
- 219 L. Xu, H. Zhao, S. Shi, G. Zhang and J. Ni, Electrolytic treatment of CI Acid Orange 7 in aqueous solution using a three-dimensional electrode reactor, *Dyes Pigm.*, 2008, **77**(1), 158–164.
- 220 A. Dirany, I. Sirés, N. Oturan, A. Özcan and M. A. Oturan, Electrochemical treatment of the antibiotic sulfachloropyridazine: kinetics, reaction pathways, and toxicity evolution, *Environ. Sci. Technol.*, 2012, **46**(7), 4074–4082.
- 221 I. Oller, S. Malato and J. Sánchez-Pérez, Combination of advanced oxidation processes and biological treatments for wastewater decontamination—a review, *Sci. Total Environ.*, 2011, **409**(20), 4141–4166.
- 222 C. Trellu, O. Ganzenko, S. Papirio, Y. Pechaud, N. Oturan, D. Huguenot, *et al.*, Combination of anodic oxidation and biological treatment for the removal of phenanthrene and Tween 80 from soil washing solution, *Chem. Eng. J.*, 2016, **306**, 588–596.
- 223 J. Kang, S. Irmak and M. Wilkins, Conversion of lignin into renewable carboxylic acid compounds by advanced oxidation processes, *Renewable Energy*, 2019, **135**, 951–962.
- 224 S. Li, Y. Liu, R. Ge, S. Yang, Y. Zhai, T. Hua, *et al.*, Microbial electro-Fenton: A promising system for antibiotics resistance genes degradation and energy generation, *Sci. Total Environ.*, 2020, **699**, 134160.
- 225 H. Afanga, H. Zazou, F. E. Titchou, Y. Rakhila, R. A. Akbour, A. Elmchaouri, *et al.*, Integrated electrochemical processes for textile industry wastewater treatment: system performances and sludge settling characteristics, *Sustainable Environ. Res.*, 2020, **30**(1), 1–11.
- 226 Y. Ling, H. Xu and X. Chen, Continuous multi-cell electrochemical reactor for pollutant oxidation, *Chem. Eng. Sci.*, 2015, **122**, 630–636.
- 227 I. M. S. Pillai and A. K. Gupta, Performance analysis of a continuous serpentine flow reactor for electrochemical oxidation of synthetic and real textile wastewater: energy consumption, mass transfer coefficient and economic analysis, *J. Environ. Manag.*, 2017, **193**, 524–531.



- 228 V. Hessel, I. V. Guersel, Q. Wang, T. Noel and J. Lang, Potential analysis of smart flow processing and micro process technology for fastening process development—use of chemistry and process design as intensification fields, *Chem. Ing. Tech.*, 2012, **84**(5), 660–684.
- 229 O. Iglesias, E. Rosales, M. Pazos and M. Sanromán, Electro-Fenton decolourisation of dyes in an airlift continuous reactor using iron alginate beads, *Environ. Sci. Pollut. Res.*, 2013, **20**(4), 2252–2261.
- 230 B. K. Körbahti, Response surface optimization of electrochemical treatment of textile dye wastewater, *J. Hazard Mater.*, 2007, **145**(1–2), 277–286.
- 231 R. Zhu, C. Yang, M. Zhou and J. Wang, Industrial park wastewater deeply treated and reused by a novel electrochemical oxidation reactor, *Chem. Eng. J.*, 2015, **260**, 427–433.
- 232 K. Pan and P. Zhou, Performance enhancement with a hydrophilic self-immobilized redox mediator modified anode in *Chlorella vulgaris*-based microbial solar cell, *ACS Sustainable Chem. Eng.*, 2015, **3**(9), 1974–1981.
- 233 E. Fernando, *Treatment of azo dyes in industrial wastewater using microbial fuel cells*, University of Westminster, 2014.
- 234 Z. He, S. D. Minter and L. T. Angenent, Electricity generation from artificial wastewater using an upflow microbial fuel cell, *Environ. Sci. Technol.*, 2005, **39**(14), 5262–5267.
- 235 E. Pajootan, M. Arami and M. Rahimdokht, Discoloration of wastewater in a continuous electro-Fenton process using modified graphite electrode with multi-walled carbon nanotubes/surfactant, *Sep. Purif. Technol.*, 2014, **130**, 34–44.
- 236 E. Bocos, D. Pérez-Álvarez, M. Pazos, M. C. Rodríguez-Argüelles and M. Á. Sanromán, Coated nickel foam electrode for the implementation of continuous electro-Fenton treatment, *J. Chem. Technol. Biotechnol.*, 2016, **91**(3), 685–692.
- 237 E. Rosales, O. Iglesias, M. Pazos and M. Sanromán, Decolourisation of dyes under electro-Fenton process using Fe alginate gel beads, *J. Hazard Mater.*, 2012, **213**, 369–377.
- 238 E. Rosales, M. Pazos, M. Longo and M. Sanromán, Electro-Fenton decoloration of dyes in a continuous reactor: a promising technology in colored wastewater treatment, *Chem. Eng. J.*, 2009, **155**(1–2), 62–67.
- 239 P. Nidheesh and R. Gandhimathi, Electro Fenton oxidation for the removal of Rhodamine B from aqueous solution in a bubble column reactor under continuous mode, *Desalin. Water Treat.*, 2015, **55**(1), 263–271.
- 240 J. K. Jang, T. H. Pham, I. S. Chang, K. H. Kang, H. Moon, K. S. Cho, *et al.*, Construction and operation of a novel mediator-and membrane-less microbial fuel cell, *Process Biochem.*, 2004, **39**(8), 1007–1012.
- 241 B. E. Logan, Exoelectrogenic bacteria that power microbial fuel cells, *Nat. Rev. Microbiol.*, 2009, **7**(5), 375–381.
- 242 H. Zhang, X. Ran and X. Wu, Electro-Fenton treatment of mature landfill leachate in a continuous flow reactor, *J. Hazard Mater.*, 2012, **241**, 259–266.
- 243 Z. Ge, J. Li, L. Xiao, Y. Tong and Z. He, Recovery of electrical energy in microbial fuel cells: brief review, *Environ. Sci. Technol. Lett.*, 2014, **1**(2), 137–141.
- 244 Y. Dong, Y. Qu, W. He, Y. Du, J. Liu, X. Han, *et al.*, A 90-liter stackable baffled microbial fuel cell for brewery wastewater treatment based on energy self-sufficient mode, *Bioresour. Technol.*, 2015, **195**, 66–72.
- 245 Z. Ge and Z. He, Long-term performance of a 200 liter modularized microbial fuel cell system treating municipal wastewater: treatment, energy, and cost, *Environ. Sci.: Water Res. Technol.*, 2016, **2**(2), 274–281.
- 246 M. Hassan, H. Olvera-Vargas, X. Zhu, B. Zhang and Y. He, Microbial electro-Fenton: an emerging and energy-efficient platform for environmental remediation, *J. Power Sources*, 2019, **424**, 220–244.
- 247 B. Tiwari, M. T. Noori and M. Ghangrekar, A novel low cost polyvinyl alcohol-Nafion-borosilicate membrane separator for microbial fuel cell, *Mater. Chem. Phys.*, 2016, **182**, 86–93.
- 248 I. Chakraborty, S. Das, B. Dubey and M. Ghangrekar, Novel low cost proton exchange membrane made from sulphonated biochar for application in microbial fuel cells, *Mater. Chem. Phys.*, 2020, **239**, 122025.
- 249 B. Logan and M. Regan, Microbial challenges and fuel cell applications, *Environ. Sci. Technol.*, 2006, **40**(17), 5172–5180.
- 250 P. Choudhury, U. S. Prasad Uday, T. K. Bandyopadhyay, R. N. Ray and B. Bhunia, Performance improvement of microbial fuel cell (MFC) using suitable electrode and Bioengineered organisms: A review, *Bioengineered*, 2017, **8**(5), 471–487.
- 251 A. Rubio-Clemente, E. Chica and G. A. Peñuela, Petrochemical wastewater treatment by photo-Fenton process, *Water, Air, Soil Pollut.*, 2015, **226**(3), 1–18.
- 252 M. Kahoush, N. Behary, A. Cayla and V. Nierstrasz, Bio-Fenton and Bio-electro-Fenton as sustainable methods for degrading organic pollutants in wastewater, *Process Biochem.*, 2018, **64**, 237–247.
- 253 C. Moreno-Castilla and F. Maldonado-Hódar, Carbon aerogels for catalysis applications: An overview, *Carbon*, 2005, **43**(3), 455–465.
- 254 H. Zhao, Y. Wang, Y. Wang, T. Cao and G. Zhao, Electro-Fenton oxidation of pesticides with a novel Fe₃O₄@Fe₂O₃/activated carbon aerogel cathode: high activity, wide pH range and catalytic mechanism, *Appl. Catal., B*, 2012, **125**, 120–127.
- 255 W. Chen, X. Yang, J. Huang, Y. Zhu, Y. Zhou, Y. Yao, *et al.*, Iron oxide containing graphene/carbon nanotube based carbon aerogel as an efficient E-Fenton cathode for the degradation of methyl blue, *Electrochim. Acta*, 2016, **200**, 75–83.
- 256 W. R. Barros, P. C. Franco, J. R. Steter, R. S. Rocha and M. R. Lanza, Electro-Fenton degradation of the food dye amaranth using a gas diffusion electrode modified with cobalt (II) phthalocyanine, *J. Electroanal. Chem.*, 2014, **722**, 46–53.
- 257 A. Thiam, I. Sirés, J. A. Garrido, R. M. Rodríguez and E. Brillas, Effect of anions on electrochemical degradation



- of azo dye Carmoisine (Acid Red 14) using a BDD anode and air-diffusion cathode, *Sep. Purif. Technol.*, 2015, **140**, 43–52.
- 258 A. Escapa, X. Gómez, B. Tartakovsky and A. Morán, Estimating microbial electrolysis cell (MEC) investment costs in wastewater treatment plants: Case study, *Int. J. Hydrogen Energy*, 2012, **37**(24), 18641–18653.
- 259 J. Beegle and A. Borole, Wastewater Energy Recovery Using Bioelectrochemical Systems and Anaerobic Digestion, *Proc. Water Environ. Fed.*, 2016, **2016**(11), 459–465.
- 260 V. Chaturvedi and P. Verma, Microbial fuel cell: a green approach for the utilization of waste for the generation of bioelectricity, *Bioresour. Bioprocess.*, 2016, **3**(1), 1–14.
- 261 M. Xu, C. Wu and Y. Zhou, Advancements in the Fenton process for wastewater treatment, *Adv. Oxid. Processes*, 2020, **61**, 61–77.

