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Fluvirucins B₇–B₁₀, new antifungal macrolactams from a marine-derived *Nonomuraea* sp. MYH522†

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Marine rare actinomycetes are an important source of secondary metabolites. From a marine-derived actinomycete *Nonomuraea* sp. MYH522, four new macrolactams, fluvirucins B₇–B₁₀, together with known fluvirucin B₆ were isolated. Their structures were determined based on comprehensive analysis of HRESIMS and NMR spectroscopic data as well as by comparing ¹³C NMR resonances and optical rotation values with those for related congeners. Fluvirucins are characterized by a 14-membered macrolactam attached by an aminosugar moiety. The discovery of fluvirucins B₆–B₁₀ enriched the *N*-acetylated derivatives of fluvirucins. The diverse alkyl substituents at C-2 and C-6 implied substrate promiscuity in fluvirucin polyketide biosynthesis. These compounds didn't exhibit any antibacterial or antifungal activities when used alone, which suggested the importance of the free amino group in the antimicrobial activity of fluvirucins. However, fluvirucins B₆, B₉, and B₁₀ showed synergistic antifungal activity with fluconazole against fluconazole-resistant isolates of *Candida albicans*.

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Introduction

Actinomycetes are characterized as one of the most important microorganisms for producing therapeutic agents in treating infections, pathogens, and cancer.¹ Due to the decrease in discovering new chemical scaffolds, more attention has been drawn to rare actinomycetes or actinomycetes from previously underexplored environments. Marine actinomycetes are found to be a noteworthy source of new bioactive molecules.^{2–4} Moreover, rare actinomycetes, which are poorly assessed in contrast to the genus *Streptomyces*, are capable of producing a large number of new secondary metabolites.^{5–7}

Fluvirucins are a group of 14-membered macrolactams particularly produced by rare actinomycetes such as the genera of *Actinomadura*, *Nonomuraea*, and *Nocardioopsis*. It should be noted that numerous species of *Actinomadura* were reclassified as *Nonomuraea*.⁸ Since the first member Sch 38516 discovered in 1990, twenty fluvirucins have been reported.^{9–17} Fluvirucins are shown with various biological properties, *i.e.*, antifungal,^{9,14,15} antibacterial,^{18–20} antiviral,^{10,20,21} and anthelmintic^{16,17} activities. The unique structure and the bioactivity of fluvirucins have also attracted attentions of chemists for total synthesis.^{22–24}

During our ongoing investigation on new secondary metabolites from marine organisms,^{25–27} we encountered a rare

actinomycete *Nonomuraea* sp. MYH522, which was isolated from a marine sponge in South China Sea. Chemical investigation of this strain led to the discovery of known fluvirucin B₆ (1) together with four new analogs fluvirucins B₇–B₁₀ (2–5) (Fig. 1). Herein, we reported the isolation, structure elucidation, and biological evaluation of these compounds.

Results

16S rDNA gene sequence and phylogenetic analysis

The 16S rDNA gene of strain MYH522 was amplified by PCR to obtain a 1450 bp sequence with NCBI GenBank accession number of NR117924.1. According to the results of nucleotide BLAST, MYH522 showed the highest sequence similarity to *Nonomuraea jabiensis* A4036 (99.24%). The related reference strains were used to multiple sequence alignment and

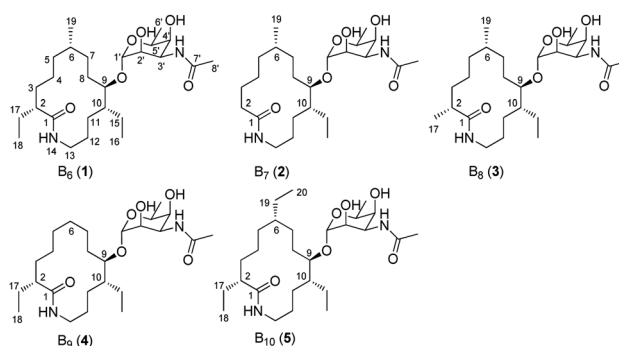


Fig. 1 Structures of fluvirucins B₆–B₁₀ (1–5).

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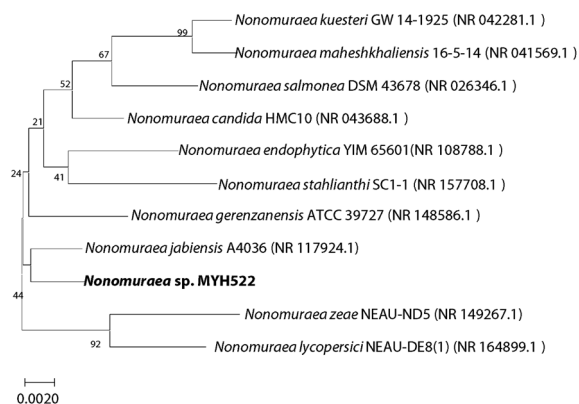


Fig. 2 Neighbor-joining phylogenetic tree of strain MYH522.

phylogenetic analysis based on a neighbor-joining method *via* MEGA7. In the neighbor-joining tree, strain MYH522 fell into the cluster of the genus *Nonomuraea* (Fig. 2). Thus, the strain was notated as *Nonomuraea* sp. MYH522.

Isolation and structural identification

The MYH522 strain was cultivated with 18 L fermentation broth and extracted with ethyl acetate (EtOAc). The extract was fractionated successively by column chromatography (CC) on silica gel, Sephadex LH-20, and RP-HPLC to produce a group of macrolactams. The major constituent was readily determined to be fluvirucin B₆ (**1**) by comparison of ¹H and ¹³C NMR data with those reported.¹⁸ Besides, four new analogs named fluvirucins B₇–B₁₀ (**2**–**5**) were co-isolated. The new compounds were subjected to structure elucidation by a set of spectroscopic experiments.

Fluvirucin B₇ (**2**) was obtained as an amorphous powder. The molecular formula was determined as C₂₄H₄₄N₂O₆ by the HRESIMS ion at *m/z* 457.3287 [M + H]⁺ (calcd 457.3278 for C₂₄H₄₅N₂O₆), which required four degrees of unsaturations. The IR spectrum of **2** displayed amide absorptions at 3302 cm⁻¹ and 1643 cm⁻¹. The structure of **2** was fully assigned by comparison with those of **1** and by further analyses of the 1D and 2D NMR data.¹⁸ The ¹H spectrum displayed four methyl groups shown as one methyl singlet, one methyl triplet, and two methyl doublets, which were one methyl triplet less than those of **1**. The ¹³C NMR and DEPT spectra indicated 24 signals that corresponded to 2 sp² (2 C=O) and 22 sp³ carbon atoms (4 CH₃, 10 CH₂, 3 CH, 4 OCH, and 1 OCO), accounting for two double bond equivalents (DBEs) (Tables 1 and 2). The remaining unsaturations were attributed to two cycles in the structure. The *O*-bearing methine protons resonating between δ_H 3.56 and δ_H 4.86 in conjunction with an acetal carbon (δ_C 99.1, C-1', CH) suggested the presence of a sugar unit. Analysis of COSY spectrum delineated a long proton spin system of H₂-2/H₂-3/H₂-4/H₂-5/H₂-6/H₂-7/H₂-8/H₂-9/H₂-10/H₂-11/H₂-12/H₂-13, H-6/H₃-19, and H-10/H₂-15/H₃-16 (Fig. 3). The HMBC correlations from H₂-2 (δ_H 2.18, 2.27) and H₂-13 (δ_H 3.20, 3.35) to C-1 (δ_C 176.1, C) connected the long COSY fragment and permitted the establishment of a 14-membered macrolactam skeleton. The HMBC cross-peaks from

Table 1 ¹³C NMR spectroscopic data for **2**–**5** (in CD₃OD, 150 MHz)

No.	2	3	4	5
1	176.1, C	179.7, C	178.9, C	178.9, C
2	36.2, CH ₂	43.0, CH	50.9, CH	51.2, CH
3	26.5, CH ₂	36.2, CH ₂	34.1, CH ₂	34.6, CH ₂
4	26.3, CH ₂	26.1, CH ₂	26.5, CH ₂	26.3, CH ₂
5	34.2, CH ₂	35.2, CH ₂	27.9, CH ₂	33.4, CH ₂
6	32.4, CH	32.4, CH	27.8, CH ₂	39.8, CH
7	25.7, CH ₂	26.1, CH ₂	27.2, CH ₂	23.2, CH ₂
8	21.9, CH ₂	22.8, CH ₂	20.0, CH ₂	22.6, CH ₂
9	78.5, CH	78.7, CH	78.4, CH	78.5, CH
10	41.8, CH	42.3, CH	42.3, CH	42.0, CH
11	26.9, CH ₂	26.3, CH ₂	26.2, CH ₂	26.3, CH ₂
12	25.8, CH ₂	28.5, CH ₂	28.6, CH ₂	28.7, CH ₂
13	39.7, CH ₂	39.8, CH ₂	39.7, CH ₂	39.7, CH ₂
15	22.8, CH ₂	22.3, CH ₂	22.2, CH ₂	22.0, CH ₂
16	10.3, CH ₃	9.4, CH ₃	9.3, CH ₃	9.2, CH ₃
17		18.9, CH ₃	27.7, CH ₂	27.5, CH ₂
18			12.4, CH ₃	12.4, CH ₃
19	20.9, CH ₃	20.9, CH ₃		28.2, CH ₂
20				12.8, CH ₃
1'	99.1, CH	99.4, CH	99.2, CH	99.3, CH
2'	71.1, CH	71.2, CH	71.1, CH	71.2, CH
3'	49.6, CH	49.6, CH	48.8, CH	48.8, CH
4'	72.2, CH	72.3, CH	72.2, CH	72.2, CH
5'	68.8, CH	68.8, CH	68.8, CH	68.8, CH
6'	17.0, CH ₃	17.0, CH ₃	17.1, CH ₃	17.0, CH ₃
7'	173.0, C	173.0, C	173.0, C	173.0, C
8'	22.6, CH ₃	22.7, CH ₃	22.7, CH ₃	22.7, CH ₃

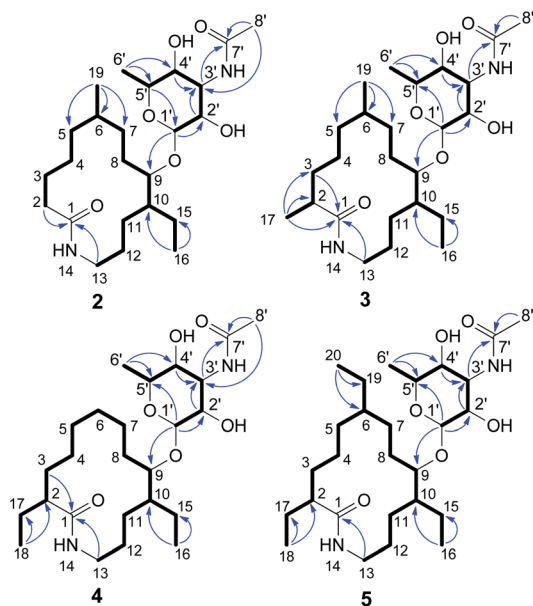
a methyl doublet (δ_H 0.92, H₃-19) to C-5 (δ_C 34.2, CH₂), C-6 (δ_C 32.4, CH), and C-7 (δ_C 25.7, CH₂) allowed the assignment of a methyl group at C-6. The methyl triplet (δ_H 0.90, H₃-16) suggested the presence of an ethyl group, which was positioned at C-10 based on the HMBC correlations from H₃-16 to C-15 (δ_C 22.8, CH₂) and C-10 (δ_C 41.8, CH).

The second fragment was interpreted starting from another proton sequence of H-1'/H-2'/H-3'/H-4'/H-5'/H₃-6' deduced by COSY spectrum. The HMBC correlations from H-1' to C-5', from H-2' and H-4' to C-3' (δ_C 49.6, CH), and from H₃-6' to C-4' and C-5' suggested an amino sugar. An acetyl group (δ_H 2.03 H₃-8'; δ_C 22.6 C-8' CH₃, 173.0 C-7', C) was substituted at C-3' NH based on the HMBC correlations from H-3' (δ_H 4.14) and H₃-8' to C-7' and the diagnostic long-range HMBC correlation from H₃-8' to C-3'. The relative configuration of amino sugar was determined by analysis of coupling constants and NOESY experiment. The obvious NOE correlation of H-3'/H-5' indicated they were axial protons and in *syn* relationship (Fig. S1†). The anomeric proton (δ_H 4.86, d, *J* = 1.5) and carbon are resonated at deshielding region indicating an α-pyranoside configuration and an equatorial orientation of H-1',¹⁰ which was suggested by the absence of NOE correlations of either H-1'/H-3' or H-1'/H-5'. The small coupling constants of ³J_{H-2',H-3'} (3.0) and ³J_{H-3',H-4'} (3.0) indicated that both H-2' and H-4' were equatorially orientated. The amino sugar was then determined to be *N*-acetyl-4-*epi*-mycosamine. Finally, a HMBC correlation from H-1' to C-9 (δ_C 78.5, CH) suggested the connectivity of two fragments, completing the structure assignment of **2**.



Table 2 ^1H NMR spectroscopic data for 2–5 (in CD_3OD , 600 MHz)

No.	2	3	4	5
2	2.27, m; 2.18, m	2.28, ddt (12.9, 9.1, 4.4)	2.10, m	2.08, ddt (13.7, 8.9, 3.9)
3	1.81, m; 1.53, m	1.49, m; 1.42, m	1.56, m; 1.46, m	1.57, m; 1.48, m
4	1.38, m; 1.24, m	1.41, m; 1.16, m	1.55, m; 1.18, m	1.47, m; 1.14, m
5	1.26, m; 1.24, m	1.43, m; 1.06, m	1.46, m; 1.21, m	1.46, m; 1.02, m
6	1.70, m	1.68, m	1.48, m; 1.48, m	1.37, m
7	1.53, m; 1.47, m	1.42, m; 1.41, m	1.36, m; 1.36, m	1.47, m; 1.31, m
8	1.68, m; 1.37, m	1.56, m; 1.49, m	1.36, m; 1.36, m	1.55, m; 1.46, m
9	3.63, m	3.62, m	3.62, m	3.62, m
10	1.54, m	1.53, m	1.55, m	1.55, m
11	1.47, m; 1.25, m	1.40, m; 1.31, m	1.56, m; 1.48, m	1.47, m; 1.41, m
12	1.47, m; 1.26, m	1.62, m; 1.35, m	1.62, m; 1.36, m	1.64, m; 1.34, m
13	3.35, m; 3.20, m	3.57, m; 2.95, ddd (13.4, 5.5, 2.9)	3.59, m; 2.98, ddd (13.4, 5.5, 2.8)	3.60, m; 2.96, ddd (13.6, 5.4, 2.8)
14	8.04, dd (5.9, 5.9)		8.05, dd (8.0, 2.8)	
15	1.62, m; 1.47, m	1.57, m; 1.53, m	1.61, m; 1.39, m	1.63, m; 1.33, m
16	0.90, t (7.5)	0.87, t (7.3)	0.87, t (7.2)	0.87, overlapped
17		1.08, d (6.9)	1.53, m; 1.39, m	1.57, m; 1.38, m
18			0.87, t (7.2)	0.87, overlapped
19	0.92, d (7.0)	0.91, d (6.7)		1.26, m; 1.26, m
20				0.87, overlapped
1'	4.86, d (1.5)	4.87, d (1.6)	4.87, s	4.87, d (0.9)
2'	3.56, m	3.55, m	3.55, m	3.57, m
3'	4.14, dd (3.0, 3.0)	4.15, dd (3.0, 3.0)	4.14, dd (3.0, 3.0)	4.15, dd (3.0, 3.0)
4'	3.56, m	3.55, m	3.55, m	3.57, m
5'	4.02, q (6.5)	4.03, q (6.5)	4.03, q (6.5)	4.03, q (6.5)
6'	1.21, d (6.5)	1.21, d (6.5)	1.21, d (6.5)	1.21, d (6.5)
8'	2.03, s	2.03, s	2.03, s	2.03, s

Fig. 3 Key ^1H – ^1H COSY correlations (bold) and HMBC correlations (blue arrows) of 2–5.

Fluivirucin B₈ (3) was isolated as an optically active powder. The HRESIMS gave a molecular formula of $\text{C}_{25}\text{H}_{46}\text{N}_2\text{O}_6$ (m/z 471.3444 $[\text{M} + \text{H}]^+$, calcd for 471.3434), which are 14 atomic mass units more than that of 2. The ^1H and ^{13}C NMR spectroscopic data of 3 closely resembled those of 2. The obvious difference was observed for the presence of one additional

methyl (δ_{H} 1.08, d, $J = 6.9$ Hz, H_3 -17; δ_{C} 18.9, C-17) in 3. The COSY cross-peak of H_3 -17/ H -2 and HMBC correlations from H_3 -17 to C-1 (δ_{C} 179.7, C), C-2 (δ_{C} 43.0, CH), and C-3 (δ_{C} 36.2, CH_2) allowed the assignment of the methyl group at C-2. Extensive analysis of 1D and 2D NMR spectra revealed that the rest part of 3 was the same as that of 2. Therefore, the structure of 3 was determined as depicted.

Fluivirucin B₉ (4) was assigned a molecular formula as same as that of 3 on the basis of HRESIMS data. Comparison of ^1H and ^{13}C NMR spectra of 4 and 3 revealed a high similarity. The obvious difference was observed for the disappearance of one methyl and the presence of one more ethyl group in the ^1H spectrum of 4. The methyl group of C-19 in 2 and 3 was replaced by a hydrogen, which led to a C-6 methylene. The ethyl group (δ_{H} 1.39, 1.53, H_2 -17 and 0.87, t, $J = 7.2$ Hz, H_3 -18; δ_{C} 27.7, CH_2 , C-17 and 12.4, CH_3 , C-18) was located at C-2 by interpretation of COSY cross-peaks of H_3 -18/ H_2 -17/ H -2 and HMBC correlations from H_3 -18 to C-17 and C-2 (δ_{C} 50.9, CH). The remaining part was determined to be as same as that of 3 on the basis of comprehensive analysis of 2D NMR data. The structure of 4 was then deduced as shown.

Fluivirucin B₁₀ (5), an optically active powder, possessed a molecular formula of $\text{C}_{27}\text{H}_{50}\text{N}_2\text{O}_6$ on the basis of HRESIMS data at m/z 499.3763 $[\text{M} + \text{H}]^+$ (calcd for $\text{C}_{27}\text{H}_{51}\text{N}_2\text{O}_6$, 499.3747). The ^1H and ^{13}C NMR spectra of 5 was close to those of 4 with the exception of one more ethyl group (δ_{H} 1.26, H_2 -19 and 0.87, H_3 -20; δ_{C} 28.2, CH_2 , C-19 and 12.8, CH_3 , C-20). This is in consistent with the fact that molecular weight of 5 was 28 mass units more than that of 4. The ethyl group was positioned at C-6 by COSY cross-peaks of H_3 -20/ H_2 -19/ H -6 and by HMBC correlations from



Table 3 *In vitro* antifungal activities of 1–5 (MIC₈₀, μg mL⁻¹)

Compounds	Used alone			With 8 μg mL ⁻¹ of fluconazole	
	SC5314	901	904	901	904
1	>64	>64	>64	0.125	0.125
2	>64	>64	>64	>64	>64
3	>64	>64	>64	>64	>64
4	>64	>64	>64	0.125	0.125
5	>64	>64	>64	0.125	0.125
Fluconazole	0.125	>64	>64	>64	>64

H₃-20 to C-19 and C-6 (δ_{C} 39.8, CH). Cumulative analyses of the 1D and 2D NMR spectroscopic data allowed the structure assignment of 5.

The relative configurations at C-2, C-6, C-9, and C-10 of aglycons in fluvirucins B₇–B₁₀ could not be assigned independently on the basis of NMR spectroscopic data. Fluvirucins B₆–B₁₀ are featured with a *N*-acetylated amino sugar in contrast to other fluvirucins. Specifically, fluvirucins B₆, B₉, and B₁₀ are *N*-acetylated derivatives of B₁, B₀, and B₃/Sch 39185, respectively.^{10,14,16,18} The NMR spectroscopic data of B₉ and B₁₀ was compared with those of B₀ and B₃/Sch 39185. The ¹³C NMR resonances of B₉ and B₁₀ showed a high similarity in aglycone part (Table S1†). Moreover, the negative optical rotation (OR) value of fluvirucin B₉ ($[\alpha]_{\text{D}}^{28.6} -29$, *c* 1.0, MeOH) was close to that of B₀ ($[\alpha]_{\text{D}}^{28.6} -38$, *c* 0.5, 1 : 1 MeOH/CHCl₃).¹⁶ Meanwhile, the NMR and OR data of fluvirucin B₁₀ ($[\alpha]_{\text{D}}^{30.9} -9.7$, *c* 0.2, MeOH) were highly similar to those of fluvirucin B₃/Sch 39185 ($[\alpha]_{\text{D}}^{26} -5.8$, *c* 0.5, MeOH).^{10,14} All of fluvirucins have identical absolute configurations at C-2, C-6, C-9, and C-10, which were determined by X-ray crystallography for fluvirucins A₁ and B₁.^{9,28} Fluvirucins B₇–B₁₀ are likely to have the same configurations as known fluvirucins according to high similarity in NMR data of aglycon and OR values, and the assumed similar biogenetic pathway.

Antimicrobial activity

The isolated compounds were screened for antimicrobial activities against pathogenic bacteria including *Staphylococcus aureus* ATCC25923 and *Escherichia coli* ATCC25922, and *Candida albicans* (the standard clinical isolated strain SC5314, the fluconazole-resistant clinical isolates of 901 and 904).²⁹ Compounds 1–5 did not exhibit any activities against *S. aureus* and *E. coli* up to 128 μg mL⁻¹ (Table S2†). Moreover, 1–5 were not active against *C. albicans* SC5314, 901, or 904 at 64 μg mL⁻¹ (Table 3). However, when combined with 8 μg mL⁻¹ of fluconazole, compounds 1, 4, 5 showed synergistic antifungal activities against fluconazole-resistant isolates of *C. albicans* 901 and 904 during 24 hours.

Discussion

Fluvirucins are characterized by 14-membered macrolactam attached with an aminosugar moiety. The structures of fluvirucins differ in the alkyl substituents and the sugar units. The alkyl substituents are usually located at C-2, C-6, and C-10,

respectively, varying among methyl, ethyl, and hydroxyethyl groups (Fig. 4). The aminosugar, normally mycosamine or 4-*epi*-mycosamine, is appended to the macrocyclic skeleton through a glycosidic linkage. According to the position of aminosugar attachment, fluvirucins are classified into “A” series having a sugar at C-3 (A₁, A₂) and “B” series with a sugar at C-9 (B₀–B₆).^{10,21} Rarely, the aminosugar is *N*-substituted by a 2-phenethylaminocarbonyl group (B₄, B₅) or an acetyl group (B₆).¹⁸ The aminosugar monosaccharide may also be replaced by disaccharide (Sch 42729) or trisaccharide (Sch 42282).^{11,12} Besides, fluvirucins without any sugar unit (C₁, C₂) have been naturally isolated.¹⁹ The discovery of fluvirucins B₇–B₁₀ enriched the *N*-acetylated derivatives of fluvirucin.

Biosynthetically, fluvirucins belong to an uncommon class of polyketides (PKS) using a specific β-alanine as starter unit. Fluvirucins B₆–B₁₀ share an ethyl group at C-10, but differ from each other in the alkyl substituents at C-2 and C-6. The alkyl substituents at both C-2 and C-6 vary between hydrogen atom, methyl, and ethyl groups which raises the question that the fluvirucin PKS may accept and process different substrates during chain extension. The biosynthetic gene clusters (BGC) for fluvirucins B₁ (*flu*) and B₂ (*flv*) have been completely identified and characterized.^{30,31} The *flu* and *flv* show a high similarity between each other containing three modular PKS genes (*flu* A–C and *flv* P1–P3) and β-amino acid forming genes. The Flu A–C as well as the homologous proteins of Flv P1–P3 are both divided into five modules (M1–M5). The substrate specificity of acyl transferase (AT) in each module is experimentally examined, showing M2_AT and M4_AT specific for malonyl-CoA, M3_AT for methylmalonyl-CoA, and M1_AT and M5_AT for ethylmalonyl-CoA. We re-analyzed the AT domains in Flu A–C and Flv P1–P3 using bioinformatic tools (Fig. 5).^{32,33} The M2_AT and M4_AT have the conserved “HAFH” motif (residues 194–197) that is specific to malonyl-CoA. Whereas, the M1_AT, M3_AT, and M5_AT have “VASH” or “YASH” motifs (residues 194–197), which are more specific for methylmalonyl-CoA (YASH motif) than ethylmalonyl-CoA (XAGH motif). The conserved motifs of AT domains could not explain the substrates promiscuity in fluvirucins. Probably, some residues dominating the canonical substrate specificity are not identified.

Fluvirucins are known to have antimicrobial and anthelmintic activities.^{9,15,17,21} Compounds 1–5 displayed almost no antibacterial or antifungal activities when used alone at the tested concentrations. Decreased antimicrobial activity have been observed for other *N*-substituted fluvirucins or fluvirucins without aminosugar unit.^{11,18–20} The current results implied that the aminosugar especially the free amino group in fluvirucins was essential for antimicrobial activity. The synergistic antifungal activities of 1, 4, and 5 against the fluconazole-resistant *C. albicans* suggested that fluvirucins may help fluconazole availability in the *C. albicans* cells by damaging cell wall or increasing intracellular fluconazole concentration, or targeting other proteins on a parallel pathway that converges on an essential process. Probably, the ethyl group at C-2 plays an important role in the synergistic antifungal activity.



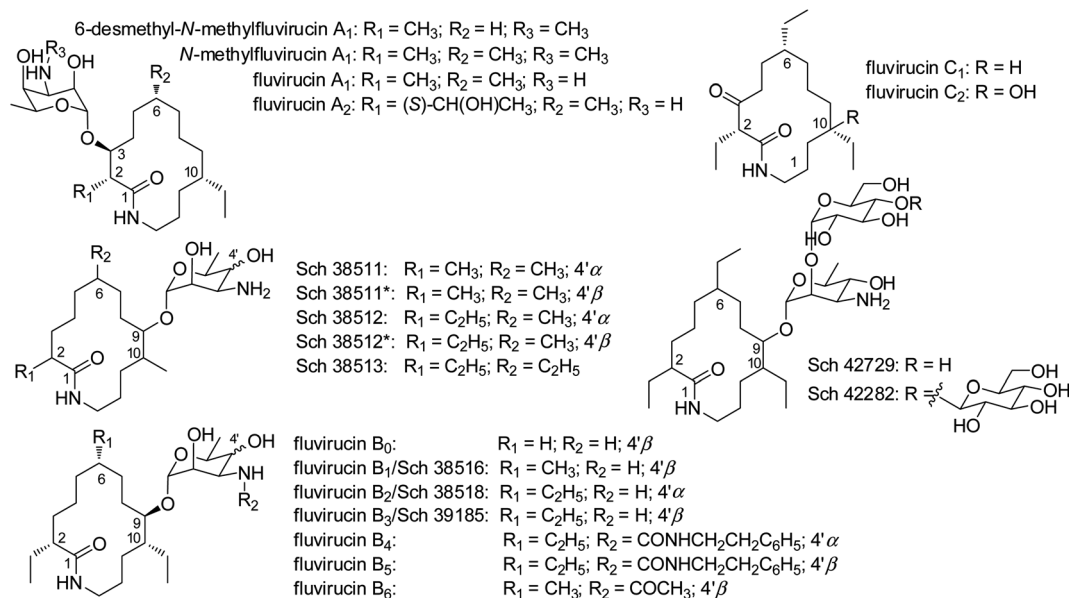
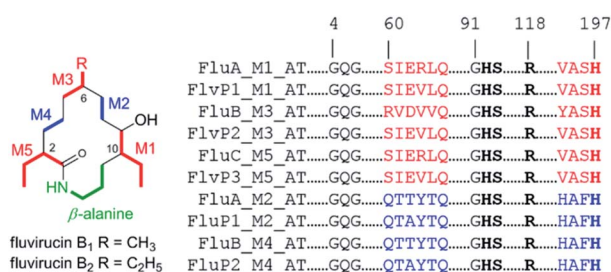


Fig. 4 Structures of known fluvirucins.

Fig. 5 Alignment of partial amino acid sequences of the AT domains of Flv PKSs (fluvirucin B₁) and Flv PKSs (fluvirucin B₂). Key catalytic residues are highlighted in bold, red for methylmalonyl-CoA or ethylmalonyl-CoA and blue for malonyl-CoA.

the residual CD₃OD signals ($\delta_{\text{H}} = 3.31$ ppm; $\delta_{\text{C}} = 49.0$ ppm) as internal standard, and coupling constants (J) are in Hz; assignments are supported by COSY, HSQC, HMBC, and NOESY experiments. High-resolution mass spectral data were obtained using a Bruker APEXIII 7.0 T FT-MS spectrometer in m/z , resolution 5000; an isopropyl alcohol solution of sodium iodide (2 mg mL^{-1}) was used as a reference. Semi-preparative HPLC was performed on an Agilent 1100 system with a UV detector using a YMC ODS-A column ($5 \mu\text{m}$, $250 \times 10 \text{ mm}$). Silica gel ($200\text{--}300$ and $400\text{--}500$ mesh; Yantai, China) was used for column chromatography. Precoated silica gel plates (HSGF254; Yantai, China) were used for TLC. Compounds were detected on TLC under UV light or by heating after spraying with anisaldehyde-sulfuric acid reagent. Antimicrobial activity assay was measured by a microplate reader SpectraMax M5.

Conclusions

In summary, we have discovered fluvirucins B₇–B₁₀ as new *N*-acetylated derivatives from a marine rare actinomycete. The results not only enrich the family of fluvirucin but also imply the substrates promiscuity of fluvirucin PKS. The biological results indicate the importance of free amino group of fluvirucins in the antimicrobial activity. The synergistic antifungal activity of fluvirucins with fluconazole is worthy of further investigation.

Experimental

General experimental procedures

Optical rotations were measured in MeOH on an Autopol-IV polarimeter at the sodium D line (589 nm). Infrared spectra were recorded in thin polymer film on a Nexus-470 FT-IR spectrophotometer (Nicolet, USA). The NMR spectra were recorded at 300 K on a Bruker AVANCE 600 NMR spectrometer. Chemical shifts are reported in parts per million (δ), with use of

Strain isolation and identification

Strain MYH522 was isolated by ISP2 medium (yeast extract 4.0 g, malt extract 10.0 g, dextrose 4.0 g, agar 20.0 g, distilled water 1.0 L, pH 7.2) from a marine sponge, which was collected off Xisha Island in South China Sea.

For phylogenetic study, strain MYH522 was grown in tryptic soy broth medium for 3 days at 28 °C. The 16S rDNA gene sequence was amplified by universal primers 27F (5'-AGTTT-GATCMTGGCTCAG-3') and 1492R (5'-GGTTACCTTGTTAC-GACTT-3'). PCR amplification reactions were prepared in a 25 μL reaction volume containing 12.5 μL of PCR 2 \times T5 Mix, 8.5 μL of distilled H₂O, 1 μL of 27F primer, 1 μL of 1492R primer, 1 μL DMSO, and 1 μL of DNA template, and was performed under reference conditions. PCR products were sent to the Sangon Biotech for sequencing. The 16S rDNA sequence was submitted to GenBank and blasted on NCBI. The multiple sequences were aligned by Clustal W. Phylogenetic trees were constructed with the MEGA7 software using neighbor-joining method. Bootstrap



analysis was used to evaluate the trees topology. Kimura two-parameter model was used for phylogeny construction and evolutionary distances analysis.

Fermentation, isolation, and structure characterization

The ISP2 medium was used for sporulation. Strain MYH522 was cultured with fermentation medium (starch soluble 10.0 g, yeast extract 4.0 g, peptone 2.0 g, CaCO₃ 1.0 g, Fe₂(SO₄)₃·4H₂O (8 g L⁻¹) 5.0 mL, KBr (20 g L⁻¹) 5.0 mL, distilled water 1.0 L, pH 7.0) at 28 °C for 7 days. An 18 L of fermentation broth was extracted with EtOAc for three times. The combined extracts were concentrated to afford a residue (1.54 g), which was subjected to ODS column chromatography, eluted by gradient MeOH–H₂O solution (30–100%, v/v) to yield 6 fractions (Fr. 1–6). Fr. 3 was separated by Sephadex LH-20 (MeOH/CH₂Cl₂, 1 : 2) to give 5 fractions (Fr. 3.1–3.5). Fr. 3.2 was subjected to semipreparative RP-HPLC (70% MeOH, 2.0 mL min⁻¹) to afford 2 (2.3 mg, *t*_R 15.6 min), 4 (14.9 mg, *t*_R 16.4 min), 3 (2.0 mg, *t*_R 18.5 min), 1 (65.5 mg, *t*_R 20.7 min), and 5 (6.5 mg, *t*_R 27.2 min), respectively.

Fluvirucin B₇ (2). White amorphous powder; *R*_f 0.28 (CH₂Cl₂/MeOH 15 : 1); [α]_D^{27.8} –76 (*c* 0.2, MeOH); IR (film) ν_{max} 3302, 2935, 2873, 1712, 1643, 1444, 1258, 981 cm⁻¹; ¹H and ¹³C NMR spectroscopic data, see Tables 1 and 2; HRESIMS *m/z*: 457.3287 [M + H]⁺ (calcd for C₂₄H₄₅N₂O₆, 457.3278).

Fluvirucin B₈ (3). White amorphous powder; *R*_f 0.30 (CH₂Cl₂/MeOH 15 : 1); [α]_D^{28.2} –29 (*c* 0.2, MeOH); IR (film) ν_{max} 3284, 2923, 2873, 1722, 1641, 1462, 1103, 979 cm⁻¹; ¹H and ¹³C NMR spectroscopic data, see Tables 1 and 2; HRESIMS *m/z*: 471.3444 [M + H]⁺ (calcd for C₂₅H₄₇N₂O₆, 471.3434).

Fluvirucin B₉ (4). White amorphous powder; *R*_f 0.30 (CH₂Cl₂/MeOH 15 : 1); [α]_D^{28.6} –29 (*c* 1.0, MeOH); IR (film) ν_{max} 3294, 2932, 1638, 1443, 1281, 1102, 1017, 827 cm⁻¹; ¹H and ¹³C NMR spectroscopic data, see Tables 1 and 2; HRESIMS *m/z*: 471.3449 [M + H]⁺ (calcd for C₂₅H₄₇N₂O₆, 471.3434).

Fluvirucin B₁₀ (5). White amorphous powder; *R*_f 0.33 (CH₂Cl₂/MeOH 15 : 1); [α]_D^{28.9} –9.7 (*c* 0.2, MeOH); IR (film) ν_{max} 3251, 2922, 1665, 1600, 1495, 1453, 1202, 702 cm⁻¹; ¹H and ¹³C NMR spectroscopic data, see Tables 1 and 2; HRESIMS *m/z*: 499.3763 [M + H]⁺ (calcd for C₂₇H₅₁N₂O₆, 499.3747).

Antibacterial activity assays

The antibacterial activity assays were performed against Gram-positive bacteria *S. aureus* and Gram-negative bacteria *E. coli* according to the Clinical and Laboratory Standards Institute (CLSI) methods.³⁴ The bacteria were grown in LB medium at 37 °C, 200 rpm for overnight, and were diluted to a concentration of 10⁵ colony forming unit (CFU) mL⁻¹. Tested compounds and kanamycin in dimethyl sulfoxide (DMSO) were 2-fold serially diluted with LB medium to prepare 100 μL of solutions ranging from 128 to 4 μg mL⁻¹, and were added to 96-well microtiter plates with 100 μL of bacterial suspensions in each well. The plates were incubated at 37 °C for 24 h. Optical density was recorded at 630 nm using microplate reader. IC₅₀ values refer the lowest concentration of compounds that cause 50% inhibition of bacterial growth.

Antifungal activity assays²⁹

The antifungal activity assays was performed using a broth microdilution methodology adjusted from CLSI M38-A2 and M27-A3 standard. The fungal suspension was applied to 96-well plates at a concentration of 10³ CFU mL⁻¹ in RPMI 1640 medium. The tested compounds and fluconazole were serially diluted with final concentrations ranging from 64 to 0.125 μg mL⁻¹. To evaluate the synthetic effect, fluconazole was added into the fungal suspension at a concentration of 8 μg mL⁻¹ before applied to 96-well plates, while the compounds were serially diluted. Plates were incubated for 24 hours at 30 °C. Optical density was measured at 630 nm with microplate reader, and background optical density was removed. The minimum inhibitory concentration (MIC₈₀) is that at which the growth of 80% tested strains is inhibited.

Author contributions

Conceptualization and supervision – P. S.; writing – H. Y. and P. S.; investigation and methodology – H. Y., S. C., H. L., and R. W.; review & editing – Y. J. and L. Y.; all authors have given approval to the final version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 J. Bérdy, *J. Antibiot.*, 2005, **58**, 1–26.
- 2 W. Fenical and P. R. Jensen, *Nat. Chem. Biol.*, 2006, **2**, 666–673.
- 3 P. R. Jensen, B. S. Moore and W. Fenical, *Nat. Prod. Rep.*, 2015, **32**, 738–751.
- 4 P. Manivasagan, K.-H. Kang, K. Sivakumar, E. C. Y. Li-Chan, H.-M. Oh and S.-K. Kim, *Environ. Toxicol. Pharmacol.*, 2014, **38**, 172–188.
- 5 S. Qi, M. Gui, H. Li, C. Yu, H. Li, Z. Zeng and P. Sun, *Chem. Biodiversity*, 2020, **17**, e2000024.
- 6 R. Subramani and D. Sipkema, *Mar. Drugs*, 2019, **17**, 249.
- 7 K. Tiwari and R. K. Gupta, *Crit. Rev. Biotechnol.*, 2012, **32**, 108–132.
- 8 Z. Zhang, Y. Wang and J. Ruan, *Int. J. Syst. Bacteriol.*, 1998, **48**, 411–422.



- 9 V. R. Hegde, M. G. Patel, V. P. Gullo, A. K. Ganguly, O. Sarre, M. S. Puar and A. T. McPhail, *J. Am. Chem. Soc.*, 1990, **112**, 6403–6405.
- 10 N. Naruse, T. Tsuno, Y. Sawada, M. Konishi and T. Oki, *J. Antibiot.*, 1991, **44**, 741–755.
- 11 V. R. Hegde, M. G. Patel, V. P. Gullo, A. C. Horan, A. H. King, F. Gentile, G. H. Wagman, M. S. Puar and D. Loebenberg, *J. Antibiot.*, 1993, **46**, 1109–1115.
- 12 V. R. Hegde, M. G. Patel, A. C. Horan, A. H. King, F. Gentile, M. S. Puar and D. Loebenberg, *J. Antibiot.*, 1998, **51**, 464–670.
- 13 R. Cooper, I. Truumees, R. Yarborough, D. Loebenberg, J. Marquez, A. Horan, M. Patel, V. Gullo, M. Puar and B. Pramanik, *J. Antibiot.*, 1992, **45**, 633–638.
- 14 V. Hegde, M. Patel, A. Horan, V. Gullo, J. Marquez, I. Gunnarsson, F. Gentile, D. Loebenberg, A. King, M. Puar and B. Pramanik, *J. Antibiot.*, 1992, **45**, 624–632.
- 15 V. R. Hegde, M. G. Patel, V. P. Gullo and M. S. Puar, *J. Chem. Soc., Chem. Commun.*, 1991, 810–812.
- 16 S. Ayers, D. L. Zink, J. S. Powell, C. M. Brown, A. Grund, O. Genilloud, O. Salazar, D. Thompson and S. B. Singh, *J. Antibiot.*, 2008, **61**, 59–62.
- 17 S. Ayers, D. L. Zink, K. Mohn, J. S. Powell, C. M. Brown, T. Murphy, A. Grund, O. Genilloud, O. Salazar, D. Thompson and S. B. Singh, *J. Nat. Prod.*, 2007, **70**, 1371–1373.
- 18 A. S. Leutou, I. Yang, T. C. Le, D. Hahn, K. M. Lim, S. J. Nam and W. Fenical, *J. Antibiot.*, 2018, **71**, 609–612.
- 19 M. Costa, P. Zúñiga, A. M. Peñalver, M. Thorsteinsdottir, M. Pérez, L. M. Cañedo and C. Cuevas, *Nat. Prod. Commun.*, 2017, **12**, 679–682.
- 20 N. Naruse, O. Tenmyo, K. Kawano, K. Tomita, N. Ohgusa, T. Miyaki, M. Konishi and T. Oki, *J. Antibiot.*, 1991, **44**, 733–740.
- 21 K. Tomita, N. Oda, Y. Hoshino, N. Ohkusa and H. Chikazawa, *J. Antibiot.*, 1991, **44**, 940–948.
- 22 E. Llàcer, F. Urpí and J. Vilarrasa, *Org. Lett.*, 2009, **11**, 3198–3201.
- 23 Y. G. Suh, S. A. Kim, J. K. Jung, D. Y. Shin, K. H. Min, B. A. Koo and H. S. Kim, *Angew. Chem., Int. Ed. Engl.*, 1999, **38**, 3545–3547.
- 24 M. Martín, G. Mas, F. Urpí and J. Vilarrasa, *Angew. Chem., Int. Ed. Engl.*, 1999, **38**, 3086–3089.
- 25 P. Sun, F.-Y. Cai, G. Lauro, H. Tang, L. Su, H.-L. Wang, H. H. Li, A. Mándi, T. Kurtán, R. Riccio, G. Bifulco and W. Zhang, *J. Nat. Prod.*, 2019, **82**, 1264–1273.
- 26 J. Gong, P. Sun, N. Jiang, R. Riccio, G. Lauro, G. Bifulco, T.-J. Li, W. H. Gerwick and W. Zhang, *Org. Lett.*, 2014, **16**, 2224–2227.
- 27 P. Sun, D.-X. Xu, A. Mándi, T. Kurtán, T.-J. Li, B. Schulz and W. Zhang, *J. Org. Chem.*, 2013, **78**, 7030–7047.
- 28 N. Naruse, M. Konishi, T. Oki, Y. Inouye and H. Kakisawa, *J. Antibiot.*, 1991, **44**, 756–761.
- 29 H. Quan, Y. Y. Cao, Z. Xu, J. X. Zhao, P. H. Gao, X. F. Qin and Y. Y. Jiang, *Antimicrob. Agents Chemother.*, 2006, **50**, 1096–1099.
- 30 T. Y. Lin, L. S. Borketey, G. Prasad, S. A. Waters and N. A. Schnarr, *ACS Synth. Biol.*, 2013, **2**, 635–642.
- 31 A. Miyanaga, Y. Hayakawa, M. Numakura, J. Hashimoto, K. Teruya, T. Hirano, K. Shin-Ya, F. Kudo and T. Eguchi, *Biosci., Biotechnol., Biochem.*, 2016, **80**, 935–941.
- 32 F. Del Vecchio, H. Petkovic, S. G. Kendrew, L. Low, B. Wilkinson, R. Lill, J. Cortes, B. A. Rudd, J. Staunton and P. F. Leadlay, *J. Ind. Microbiol. Biotechnol.*, 2003, **30**, 489–494.
- 33 C. D. Reeves, S. Murli, G. W. Ashley, M. Piagentini, C. R. Hutchinson and R. McDaniel, *Biochemistry*, 2001, **40**, 15464–15470.
- 34 J. H. Rex and M. J. Ferraro, in *Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically; Approved Standard—Ninth Edition*, Clinical and Laboratory Standards Institute, Wayne, PA, 2012, vol. 32, 07–A0907–A0907–A09.

