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Nanocomposites based on the graphene family for food packaging: historical perspective, preparation methods, and properties

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Nanotechnology experienced a great technological advance after the discovery of the graphene family (graphene – Gr, graphene oxide – GO, and reduced graphene oxide-rGO). Based on the excellent properties of these materials, it is possible to develop novel polymeric nanocomposites for several applications in our daily routine. One of the most prominent applications is for food packaging, offering nanocomposites with improved thermal, mechanical, anti-microbial, and barrier properties against gas and water vapor. This paper reviewed food packaging from its inception to the present day, with the development of more resistant and intelligent packaging. Herein, the most common combinations of polymeric matrices (derived from non-renewable and renewable sources) with Gr, GO, and rGO and their typical preparation methods are presented. Besides, the interactions present in these nanocomposites will be discussed in detail, and their final properties will be thoroughly analyzed as a function of the preparation technique and graphene family-matrix combinations.

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1. Introduction

The nanotechnology field has been expanding with nanomaterials' development regarding their different structural, morphological, and chemical characteristics, guaranteeing enormous versatile applications. One of these features includes the ability to link molecules of different natures and functionalities that can be used for diverse applications for various products: pharmaceutical nanoformulations, ointments/creams for therapeutic or aesthetic purposes, curative films, active packaging, food additives, food packaging, and others.^{1–3} Considering these cutting-edge nanotechnology applications, food packaging is of great interest since several physical and chemical properties can be tuned through the formulation of nanocomposites.^{1–3} The application of nanomaterials in the food industry – mainly in the packaging market – increased its market value from \$20.4 billion in 2006 to more than

\$3 trillion in 2020.⁴ Moreover, nanotechnology contributes significantly to reducing food lost amount and thus offers an alternative for the increasing food demand as the worldwide population increases. Food packaging acts as a physical barrier against the interaction of food and the environment and thus preventing its contamination with microorganisms, dust, or even spoiling as a consequence of moisture, light, and shock. Considering that, several strategies for food packaging have been developed over the centuries, most notably in the last century, in which materials chemistry exerted a pivotal role in the search for smart packaging. The combination with conventional polymers used for packaging with other materials offered a wide range of new materials (composites) with improved properties.⁴

In this way, graphene (Gr), graphene oxide (GO), and reduced graphene oxide (rGO)^{5–23} are examples of these nanomaterials which are used in composites formulations with polymers, such as polypropylene (PP),²⁴ polystyrene (PS),²⁵ polyethylene terephthalate (PET),²⁶ polyvinyl alcohol (PVA),²⁷ poly lactic acid (PLA),²⁸ polyhydroxybutanoate (PHB),²⁹ chitosan (CS),³⁰ starch (ST),³¹ cellulose³² and others polymeric matrix.^{33–36} The interaction between Gr or GO with these polymers occurs through chemical bonds or molecular interactions and thus improves their chemical,³⁷ mechanical, thermal, optical, anti-microbiological, and barrier properties.^{16,19,38–45}

In light of the considerations mentioned above, this review will focus on the recent developments of the graphene family (graphene, graphene oxide, and reduced graphene oxide)-based

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nanocomposites used as food packaging systems. We will provide an overview of the historical use of food packaging materials and present the most common preparation methods of graphene-based nanocomposites – with several fossils and renewable-derived polymeric matrices – their physicochemical interactions, final properties as a food packaging material, and most common applications found in the market.

2. Food packaging

2.1. Historical perspective

The first types of food packaging arose between prehistory and ancient history. People stored and protected their food in plant leaves, clay artifacts, seashells, baskets, gourds, skin bags, and animal organs (stomach and bladder), coats, hooves, and

horns.⁴⁶ Years later, in A.D. 751, paper became known and made throughout Europe, but its use for food packaging became famous many years later. The development of food and beverage storage techniques was essential to transition from nomadic activities to more complex societies.⁴⁶

The discovery of novel technologies applied to food packaging is of great importance for preserving and maintaining the shelf life of foods in general. Many techniques have been employed to manufacture packaging and have continued to change over the centuries. The correct selection of the type of packaging for each food is essential for maintaining the integrity and quality of the food, as they will keep the food protected, fresh and healthy since they leave the industries until they reach the consumer's table. Several types of material have been used

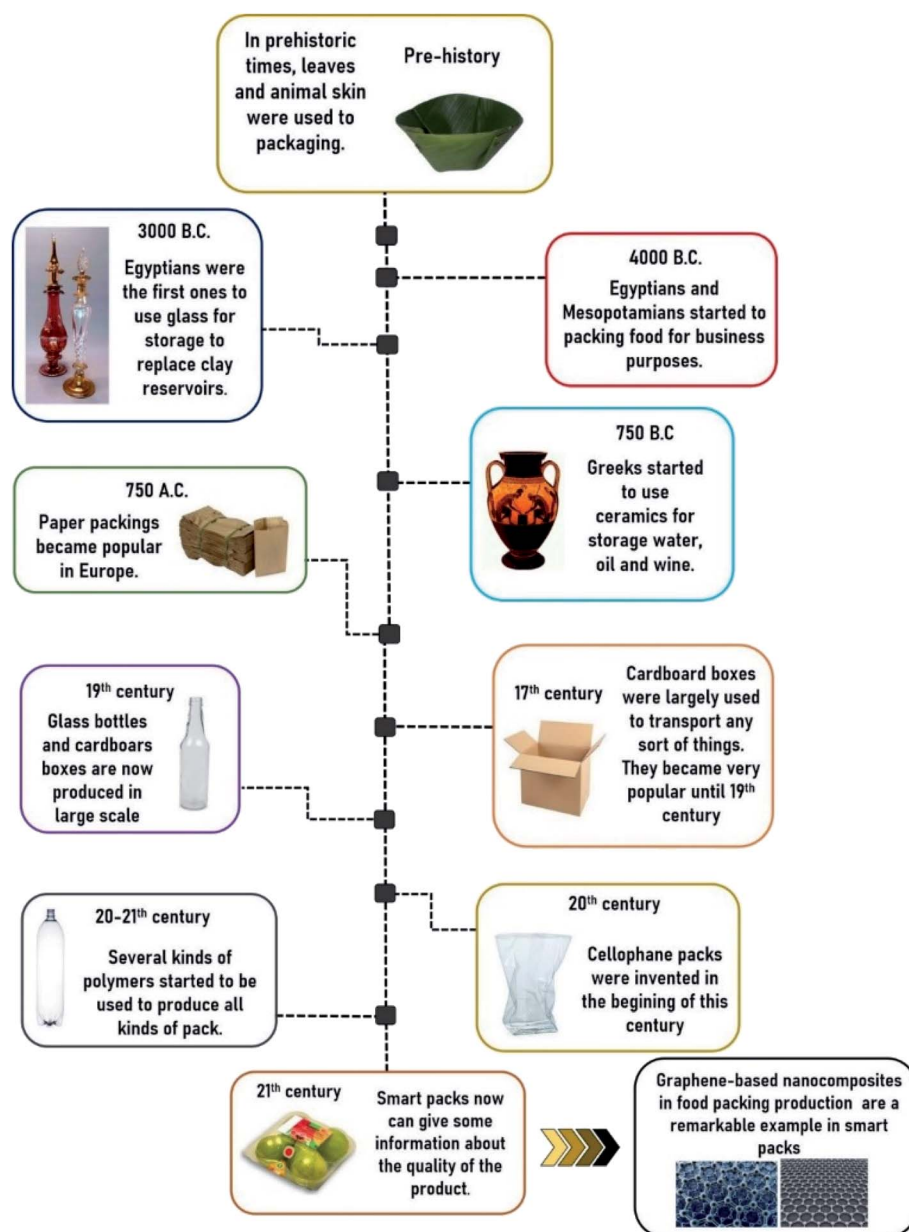


Fig. 1 Historical perspective of food packaging.

to pack food, such as glass, aluminum, tin-free steel, paper, cardboard, and rigid and flexible plastics.⁴⁷

The use of glass for food storage dates back to 3000 B.C. The glass containers have characteristics to be impermeable to gases and vapors, keeping food and drink fresh, and offer the possibility of storage for long periods without interfering with the food taste. The main disadvantage of using glass containers is their difficulty transporting because they are fragile and bulky. The glass package may break if there are impacts, thermal shocks or if the internal pressure increases.^{47,48}

Metal packaging is generally produced from aluminum, tin-free steel, and tinplate. These materials are versatile and have good protection and barrier properties, and also possess a high decorative potential and more resistance, being lighter and more malleable than glass packaging.^{47–49}

In that sense, aluminum packaging is resistant to corrosion because it is naturally coated with aluminum oxide, creating an effective barrier to gases and vapors, chemical attacks, variations in humidity and temperature, odors, light, and microorganisms.^{47–49} Steel packages without tin or chrome steel coated with oxide or electrolytic chromium require the coating with organic material to be completely resistant to corrosion.^{47–49}

Paper-based packaging is also important for the food industry and is widely applied. Paper and cardboard use as food packaging began in the 17th century and expanded in the late 19th century.^{47,50} Kraft paper is the most resistant paper packaging used to pack vegetables, dried fruits, sugar, and flour. Also, glassine is a paper that undergoes a more extreme hydration process than the previously mentioned and produces a denser, smoother, and shiny paper used to pack fried foods, baked goods, and fast foods. Currently, laminated papers are coated or not with cellulose and kraft sulfite. They can also be laminated with plastic or aluminum to improve their gas and moisture barrier properties, substantially increasing the cost of paper.^{47,48,50}

The rise of modern food packaging began in the 19th century after the industrial revolution, with the studies of can-food by

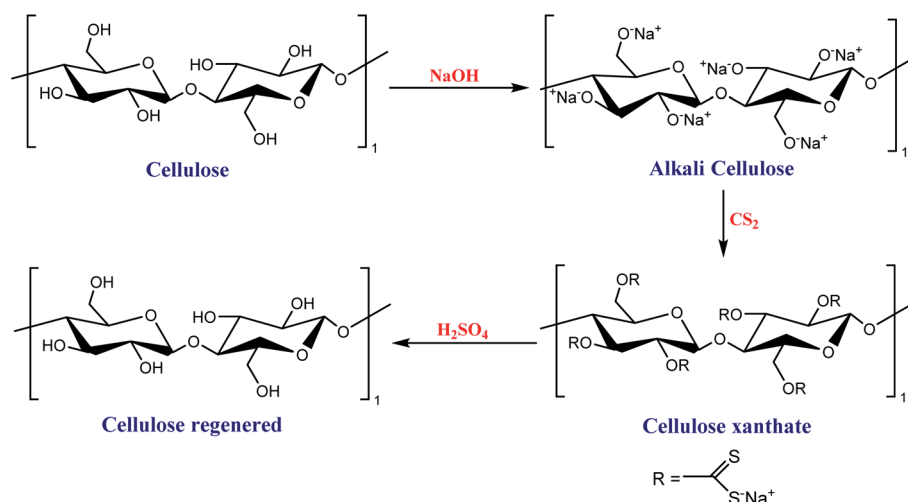
Appert, based on the food microbiology of Pasteur, Prescott, and Underwood.⁵¹ In this way, Fig. 1 shows the types of food packaging used by humanity over the centuries.

In the early 20th century, some food packaging appeared from unexpected sources due to unsuccessful attempts, such as the transparent tablecloths by Jacques Brandenberger, a Swiss textile engineer. However, cellophane on regenerated cellulose kind was created through this unsuccessful attempt.⁵¹

For cellophane production, cellulose is first dissolved in sodium hydroxide solution. It is treated with carbon sulfide producing viscose (cellulose xanthate), which is extruded in a sulfuric acid and sodium sulfate solution to convert the viscose into regenerate cellulose. The regenerated cellulose (cellophane) goes through several baths: sulfur removal, bleaching, and glycerin to avoid fragility,^{52,53} as shown in Scheme 1. Cellophane is transparent, with a high permeability barrier to oils, greases, and air; it is used to pack sweets, generally.⁵⁴

During the first (1914–1918) and the second (1939–1945) World wars, there was a lot of innovation in food packaging, such as oil products and wax for dry cereals and cookies. In this way, plastics like polyethylene (PE) and polyvinylidene chloride (PVDC) emerged as aseptic packaging and flexible metal cans for bottling beer. The development of polymers such as polyethylene (PE), polyethylene terephthalate (PET), polypropylene (PP), and polyvinyl alcohol allowed a partial replacement of glass, metal, and rigid plastic packaging by these new flexible plastic packaging.^{48,50,51,55} Since the end of the 20th century, many polymers produced from microorganisms, plants, and animals have been studied. Furthermore, in the 20th century, intelligent packaging capable of controlling oxygen diffusion, being a breath mediator, scent controller, and antimicrobial was developed.^{48,50,51,54–56}

In 2007 Kalaitzidou and co-workers²⁴ were the first group of researchers to create a polymer of polypropylene and graphene. They observed that the addition of graphene to the polymer matrix improved the physical, thermal, and mechanical



Scheme 1 Cellophane production route.



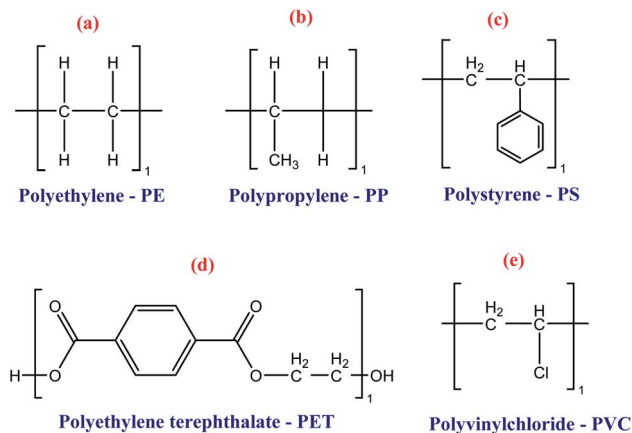


Fig. 2 Structures of: (a) polyethylene – PE, (b) polypropylene – PP, (c) polystyrene – PS, (d) polyethylene terephthalate – PET and (e) polyvinyl chloride – PVC.

properties and that the polymeric nanocomposite could be used to produce food packaging. Item 3.4 presents more details about graphene-based nanocomposites applied to food packaging.

Considering the pivotal role that polymers-based packaging possesses in our modern society, the following sections will cover the non-renewable and renewable polymers most used in food packaging.

2.2. Non-renewable-based polymers applied to food packaging

Since the 20th century, the main polymers used in food packaging have been polyethylene, polyethylene terephthalate, polystyrene, polypropylene, and polyvinyl because of their high applicability and availability low molecular weight, and low cost.⁵¹ Most of these materials have some drawbacks to be used as food packaging. In most cases, they are not biodegradable and take several years to completely degrade by nature.^{33,48,50}

2.2.1. Polyethylene. Polyethylene (PE) is one of the most used polyolefins-based polymers in the manufacture of food packaging.^{47,56} PE, Fig. 2(a), is produced from the polymerization reaction of ethylene monomers. Two polyethylene types called low and high density can be made, LDPE and HDPE, respectively. LDPE is widely used in food packaging due to its easy processing and extensive ability to coat various substrates; it has light color, lightness, flexibility, and low cost.⁵⁷ Moreover, LDPE has low permeability to water vapor, although it does not present barriers to protect gases, volatile compounds, and other gases. It is used to produce grocery bags, food packaging for frozen food, papers and laminates coating, bread bags, *etc.*^{47,50,57} On the other hand, HDPE has high resistance, a higher gas permeability barrier, and thermal resistance.^{47,50}

2.2.2. Polypropylene. Polypropylene (PP) is also one of the most used olefin-based in food packaging manufactured through the polymerization reaction of propylene monomers Fig. 2(b). PP has a strong barrier to protecting against humidity, water vapor, and gases and is resistant to chemicals. In this sense, it is resistant to impacts and ruptures, making it suitable food packaging to be used even in a microwave oven.^{47,50,57}

2.2.3. Polystyrene. Polystyrene (PS) is formed by the polymerization reaction between the styrene monomers, Fig. 2(c).^{47,58} PS is used to manufacture cups, cutlery, disposable plates, food trays, and bottles. The expanded PS is very light, rigid, and opaque, thus acquiring excellent barrier and impact properties, making it suitable to protect fragile products.^{50,53,60}

2.2.4. Polyethylene terephthalate. Polyethylene terephthalate (PET) is a polyester formed from the polymerization of terephthalic acid (TA) and ethylene glycol (EG), Fig. 2(d). PET is thermally resistant to temperature and impact, also owing good barrier of protection against O₂ and CO₂.⁵⁸ PET is a light, transparent material with an amorphous structure and, therefore, widely used in food packaging and soft drink bottles since it retains CO₂.^{47,50,57,59}

2.2.5. Polyvinyl chloride. The polymerization among vinyl chloride monomers produces polyvinyl chloride (PVC),

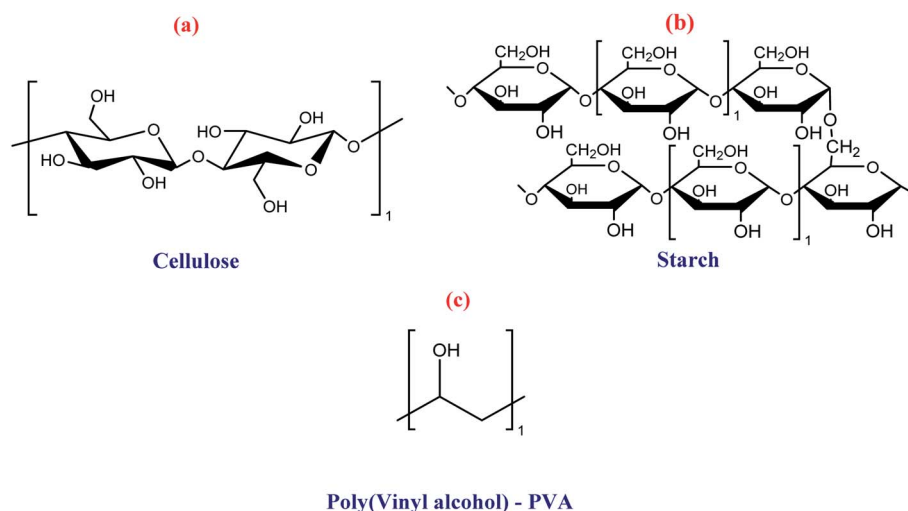


Fig. 3 Structures of: (a) cellulose, (b) starch and (c) poly(vinyl alcohol) – PVA.

Fig. 2(e),⁶⁰ generating an amorphous, transparent, ductile, striated, dense, and brittle material that can be heat sealed.^{47,50,57}

2.3. Renewable-based polymers applied to food packages

Several polymeric materials from renewable feedstocks are already accessible, representing an evolution towards a more sustainable society, allowing at least a partial replacement of fossil-based polymers. These polymers can be classified into four groups: polymers from vegetal sources (cellulose and starch); polymers from animal sources (chitin and chitosan); polymers from microbiological sources (polyhydroxyalkanoates – PHAs); and polymers chemically synthesized from agricultural raw materials (poly(vinyl alcohol – PVA; polylactic acid – PLA; biopolyethylene – bioPE; biopolyethylene terephthalate – bioPET)).^{54,56}

2.3.1. Cellulose. Cellulose is one of the most abundant fractions present in lignocellulosic biomass and can be used as starting material for several food packaging purposes. The chemical structure of cellulose, Fig. 3(a), is responsible for its remarkable properties. An equatorial position of the hydroxyls of its structure allows H bonds to occur – allowing a high degree of organization and crystallinity.^{34,35,54,56,61}

Cellulose is highly hydrophilic, insoluble in most solvents, and non-thermoplastic, *i.e.*, it does not melt and cannot be moldable at a specific temperature. Besides, several functional groups, such as carbonyl and carboxyl, can be intercalated in its structure.^{34,35,54,56,61} The changes made in the cellulose structure usually involve the intrinsic hydroxyl groups through esterification and etherification reactions, thus changing their physicochemical properties.

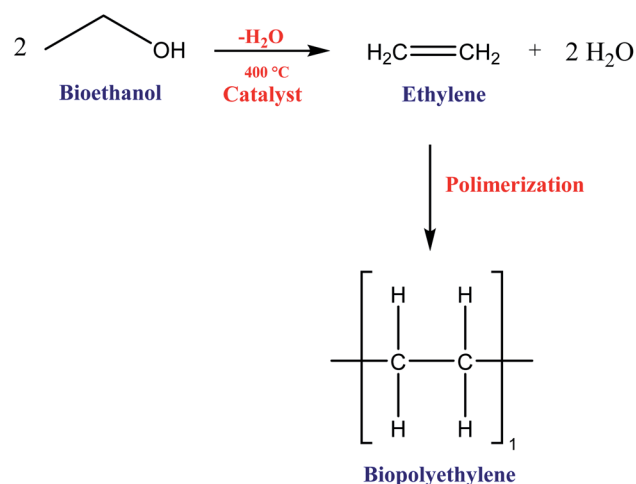
Superhydrophobic cellulose-based materials are on the micrometric scale and have low surface energy. These materials are in evidence because they are abundant on Earth. In addition to being biodegradable, non-toxic, and renewable, they have chemical, physical and mechanical properties superior to non-renewable materials. In other words, they are a more sustainable and environmentally friendly alternative to polymers based on fossil fuels. Traditional hydrophobicity treatments do not insert the functionalities found in superhydrophobic cellulose materials. One of the methods used to manufacture cellulose-based superhydrophobic materials is wet chemistry, which produces more durable coatings (important for materials with abrasion resistance and wash cycles). The other method is the dry method, which has the main advantage of simplicity by using one-step processes without using organic solvents. Its promising features have potential application in self-cleaning, self-curing, oil and water separation, shielding against electromagnetic interference, *etc.* The perspective is that the subsequent studies aim to seek these materials for more practical and suitable applications in different areas.^{62,63}

The most known cellulose-derived materials are carboxymethyl cellulose, hydroxyethyl cellulose, hydroxypropyl cellulose, ethylcellulose, and cellulose acetate. Their uses are being investigated for numerous purposes in food packaging, such as fruit, vegetables, fast food, bread, and others.^{54,56,64}

2.3.2. Starch. Starch (Fig. 3(b)) is a compound used by vegetables to store their energy, being present in plants' roots, seeds, and tubers. It is produced from wheat, rice, potatoes, cassava, and others. This polymer is comprised of two types of polysaccharides: amylose and amylopectin. The former is a linear polymer with α -1-4 bonds among D-glucose monomers, and the latter is formed by branched α -1-6 linkages.⁵⁴ Moreover, it is a promising alternative raw material in the manufacture of food packaging due to its high availability, low cost, and sustainability, which can be added to the formulation of fossil polymers, increasing the biodegradability of the copolymer formed.⁶⁴ The properties of starch polymers can be varied and often unstable in the presence of water, as it is a hydrophobic material; also, microbial action might create pores through their structure and accelerate its disintegration, conferring to them a biodegradable feature. Furthermore, the expansion of starch carried out by steam consists of a promising alternative material and finds applications in the manufacture of cups, plates, and food packaging as green alternatives to replace polystyrene, a non-renewable material.^{34,35}

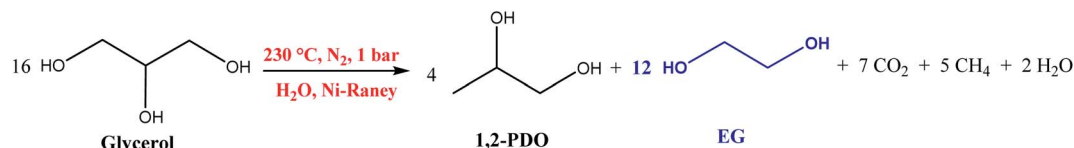
2.3.3. Biopolyethylene. As afore mentioned, PE is mainly produced from petrochemicals, *i.e.*, non-renewable feedstock. However, biopolyethylene (bioPE) can also be made through the dehydration reaction of bioethanol that produces ethylene, as shown in Scheme 2. The ethylene produced by bioethanol dehydration is polymerized to produce biopolyethylene,⁶⁵ which has the same characteristics and applications for food packaging as polyethylene from fossil sources, as discussed in Section 2.2.1.^{54,56,65}

2.3.4. Biopolyethylene terephthalate. Biopolyethylene terephthalate (bio-PET) can be produced partially or totally from renewable sources. Considering a partially renewable protocol, one non-renewable precursor, TA or EG, is derived from renewable sources. In a fully renewable approach, both molecules are produced from renewable sources. In this case, TA is usually made by the oxidation of *p*-xylene – from the fermentation of lignocellulosic materials – and then condensed with



Scheme 2 Biopolyethylene production by bioethanol dehydration route.





Scheme 3 General route glycerol APR-hydrogenolysis reactions to produce ethylene glycol in a single process.

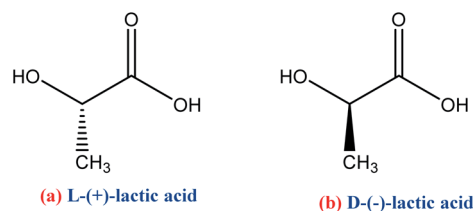


Fig. 4 (a) L-(+)-lactic acid and (b) D-(-)-lactic acid structures.

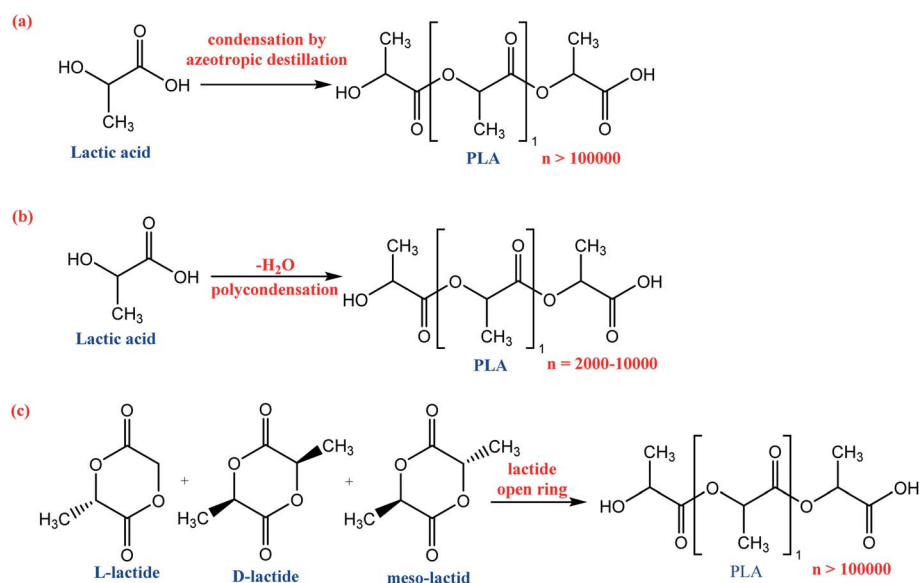
EG to form monomers.^{54,61} Furthermore, EG can be produced through the hydrogenolysis reaction of glycerol^{66,67} and by reforming the aqueous phase of glycerol (APR),^{68,69} as shown in Scheme 3. In this way, it is worth mentioning that bio-PET possesses the same properties and applications for food packaging purposes as those mentioned in non-renewable PET Section 2.2.4.^{54,61}

2.3.5. Poly(vinyl alcohol). Poly(vinyl alcohol) (PVA), Fig. 3(c), is produced by the polymerization of vinyl acetate and then hydrolyzed to PVA. The -OH groups in the PVA have intermolecular forces such as hydrogen bonds, leading to a high melting point, $T_m = 200\text{ }^{\circ}\text{C}$, in which its structure is generally organized in a zigzag and presents a semi-crystalline feature. PVA is widely used due to its biocompatibility, biodegradability, water solubility, and low toxicity. Despite its hydrophilicity, PVA

has good gas barrier properties and tear resistance. Also, as a result of this feature, it is essential in its manufacture to add crosslinking agents to block its hydrophobicity and hence be able to apply in food packaging. In this way, PVA is generally used as a packaging film for fruits and vegetables.^{33,58,61}

2.3.6. Poly(lactic acid). Poly(lactic acid) (PLA) is a polymer belonging to the group of aliphatic polyesters and synthesized from lactic acid monomers, obtained from biological sources mediated by biocatalysis and chemocatalysis (homo- and heterogeneous). In this context, lactic acid (LA) production is mainly prepared using biocatalysis, which occurs through the fermentation of sugar (sucrose, glucose, fructose, and others) that produces two stereoisomers of lactic acid, L(+), and D(-), Fig. 4.^{35,54,59,61} Chemocatalysis can also furnish LA through catalysis to convert sugars derived from non-edible lignocellulosic biomass and glycerol. In the fermentation process, a homo or heterofermentative bacteria is used as an enzymatic catalyst.^{35,54,59,61}

PLA can be synthesized by different polymerization routes,⁷⁰ Scheme 4, (a) LA condensation by azeotropic distillation; (b) LA polycondensation and dehydration; (c) lactide ring-opening polymerization. The main properties of PLA are good protection barrier, high mechanical and chemical resistance, low toxicity, transparency, and insolubility in water. However, it presents lower thermal stability compared to PET. Besides, the PLA barrier property to gases such as O_2 and CO_2 is lower than



Scheme 4 PLA production routes: (a) LA condensation by azeotropic distillation; (a) LA polycondensation, dehydration; (c) lactide ring-opening polymerization.



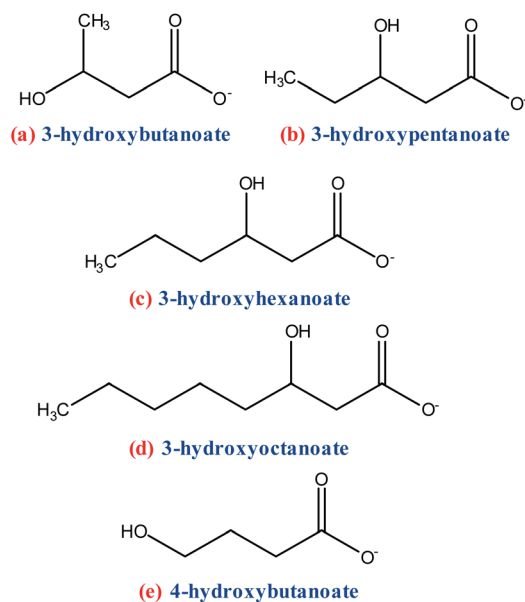


Fig. 5 Different monomers used for PHAs production.

PS and approximately equal to PET polymers. PLA mechanical properties are superior to those observed in PP, HDPE, and PS due to their chemical structure and hardness, and T_m between 173–178 °C. The commercial PLA consists of a copolymer formed between the stereoisomers D(–) and L(+) of the lactic acid, as shown in Scheme 4c. Considering its applications, PLA is used to manufacture food packaging such as meat and soft drinks, among other purposes. Furthermore, packages composed of PLA/cellulose and PLA/natural fiber copolymers gain strength in several packaging and material sectors as they are biodegradable.^{35,54,59,61}

2.3.7. Poly(hydroxyalkanoates). Poly(hydroxyalkanoates) (PHAs) are ester polymers with optically active carbon atoms. PHAs are thermoplastics and elastomers possessing a melting point ranging from 40 to 180 °C according to the monomer (substrate or reagent) used in the polymerization reaction. These polymers are usually produced by bacterial fermentation of large amounts of carbon, such as sugars and lipids. These bacteria can also consume the polymer when there are few sources of carbon to feed them.^{35,54,59,61}

Fig. 5 shows the different monomers used in PHAs production.⁷¹ The polymers shown in Fig. 5 also be chemically synthesized with biodegradability similar to those produced by a bacterial pathway. The most common PHAs is polyhydroxybutanoate (PHB), the polymerization product of 3-

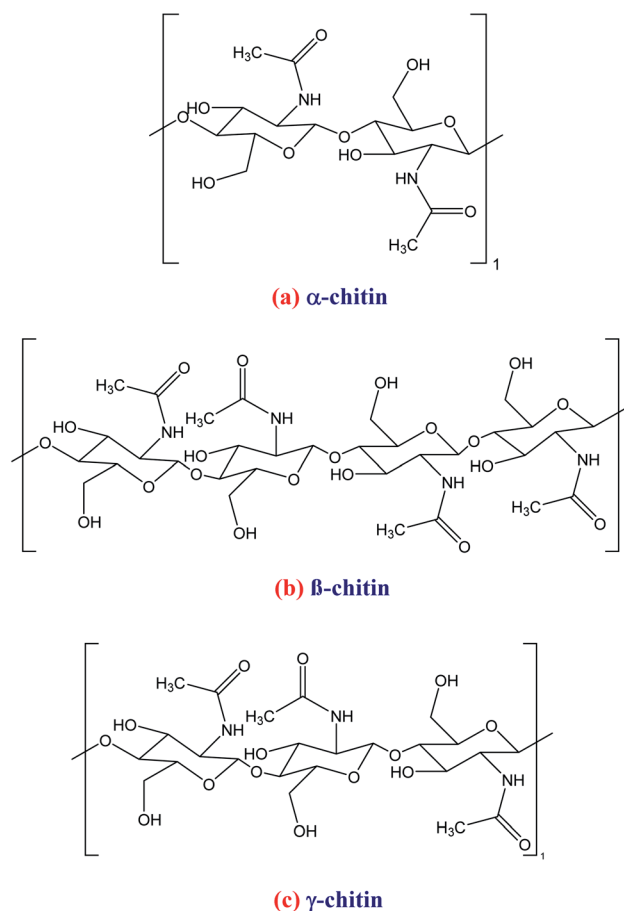
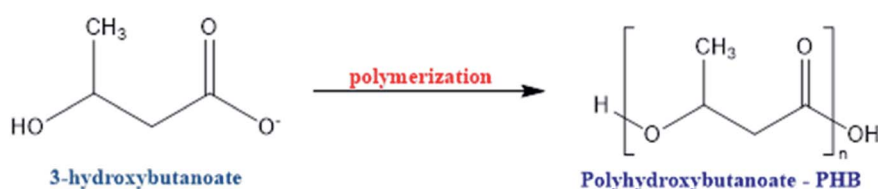


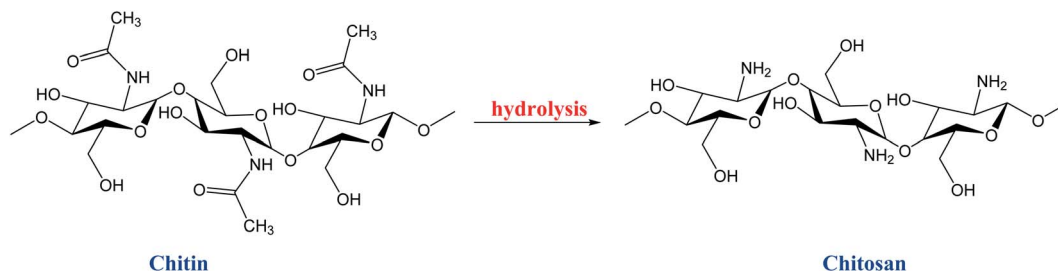
Fig. 6 Different chitin molecular structures, (a) α- (b) β- and (c) γ-chitin.

hydroxybutanoate monomers,^{35,61,71,72} Scheme 5. PHB has an orthorhombic crystalline structure, with a flat amorphous phase between the lamellae; it is a thermoplastic with high crystallinity (80%) and possesses higher rigidity and fragility compared to PP. It has good gas barrier properties, high hydrophobicity, resistance to organic solvents, and is highly biodegradable. The disadvantages of industrial PHB processing are low deformability and thermal stability, the difficulty of being processed industrially, and being expensive. However, some studies have been reported to modify PHAs, such as the use of their combination with plasticizers, acrylic acid, epoxidized soybean oil, and cellulose acetate to improve properties and facilitate the processing for future applications in food packaging, paper coating, and other purposes.^{35,54,59,61,64,71,73,74}



Scheme 5 Different monomers used for PHB production polymerization of 3-hydroxybutanoate monomers.





Scheme 6 Chitin deacetylation by hydrolysis route to produce chitosan.

2.3.8. Chitin and chitosan. Chitin (CT) or *N*-acetyl-D-glucosamine is a polysaccharide found in three different structures α , β and γ ,^{54,73,74} Fig. 6.

The α -chitin structure has higher physical and chemical resistance because it has an antiparallel arrangement between its oligomers, causing higher strength in the hydrogen bonds of its structure. β -Chitin is characterized by the parallel arrangement of chitin oligomers for the formation of its structure. The *N*-acetyl groups are structural spacers, facilitating the hydration of this macromolecule. The γ -chitin structure is formed by two parallel and one antiparallel chitin sheets, and the *N*-acetyl groups play the same role as in β -chitin. The strength and resistance of β - and γ -chitin are similar and weaker compared to α -chitin. Amorphous chitin structures have also been reported and were found in fungi.^{54,73,74}

Nature produces billions of tons of crystalline chitin present as the exoskeleton of some insects, arthropods, and mollusks, and in the cell walls of fungi, yeasts, and plants. Chitin is highly hydrophobic, non-toxic, biodegradable, and easily compacted. It is generally soluble only in mixtures of hexafluoroacetone, chloroalcohols, and hexafluoroalcohols with aqueous solutions of strong inorganic acids. Due to its poor chemical reactivity, cellulose is industrially preferred to chitin. However, its derivative chitosan (CS) possesses higher solubility in a wide range of solvents and has gained considerable attention from researchers and industries, resurging the interest in nature chitin. Amines' presence in the chitin and chitosan structure can be chemically modified to improve their properties and hence furnishing several applications in technological and industrial fields.^{54,74} In this sense, CS is produced by the hot leaching process of chitin, which causes the removal of the acetate group from the chitin molecule by hydrolysis,^{54,74} Scheme 6.

After this process, chitosan is washed, dried, ground, and ready to be commercialized.^{34,35,61} The chitosan application for food packaging formulation tests showed excellent mechanical and barrier (O_2 and moisture) properties, suitable antimicrobial and antioxidant activities, and good biodegradability, making chitosan a promising polymer in the packaging industry.^{41–44,75–78}

Although biopolymers based on renewable sources consist a step forward for food packaging to achieve a more sustainable society, we are still highly dependent on polymers derived from non-renewable sources for such purposes. In addition, the

current packaging technologies lack several points, such as higher resistance barrier properties, hence the research of new technologies for food packaging. In this way, nanocomposites (mixture of two different nanomaterials with distinct properties) endure a promising pathway for high-tech packaging. They might improve mechanical, thermal, barrier properties, cytotoxicity, microbiology, and biodegradable properties.^{41–44,75}

2.4. Nanocomposites-based packaging

The first example of composites for food packaging was reported at the beginning of this century (21st). Metal cans were covered with a multilayer plastic barrier preventing the contact of the food with the metal and thus increasing its shelf life. Also, other technological changes in food packaging were reported, such as the change in the internal atmosphere, the resistance of the packaging to the microwave oven, trays for gas and electric ovens, and retractable film labels, among other examples.⁵¹

Due to the several combinations of polymer matrix/dispersed nanomaterials, polymer-based nanocomposites can provide different properties for a wide range of applications.^{33,79}

Graphene-based nanocomposites have been utilized to improve UV resistance as a barrier against gases and still have good thermal, mechanical, and electrical properties compared to their polymeric matrices.^{80–83} Furthermore, graphene-based nanomaterials combined with biodegradable polymers offer high potentials to be used as antimicrobial and antioxidant in active packaging technology resulting in more quality, safety, extended shelf-life, and added value.^{84–86}

Some examples of nanocomposites based on polymer matrix/nanomaterials can be found in the literature. For example, polyvinyl alcohol (PVA)/graphene-based nanocomposites are just one example of polymeric nanocomposites

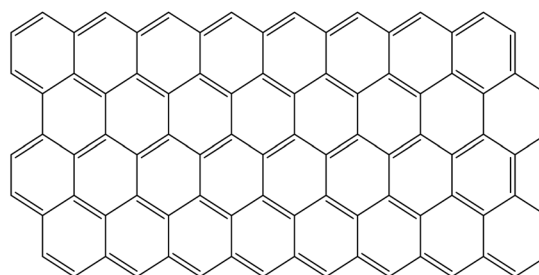


Fig. 7 Graphene structure (Gr).

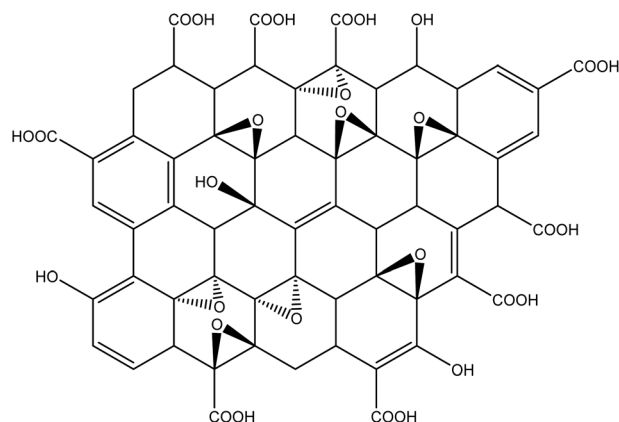


Fig. 8 Graphene oxide (GO) structure.

with considerable potential in the food packaging industry. They have excellent gas and moisture barrier properties and good thermal resistance. Besides protecting the product, the new concept of packaging technologies brings technological innovations to adding novel features for barrier performance. An example is the active packaging technologies and modified atmosphere packaging for fresh vegetables and fruits.⁸⁷

The interaction between the packaging, the product, and the environment through partial gas permeability increases shelf-life providing oxygen to cellular respiration.³³

Biobased polymer packaging generally are films made from biopolymers and biodegradable polymers, such as starch,⁸⁸ pullulan,⁸¹ PVA,^{85,86} and PLA.⁸⁹ These nanocomposites can be used in food packaging as nanocoating films to control moisture transfer and gas exchange, provide security, and preserve nutrition and sensory features.⁷⁹

Furthermore, in smart packaging based on polymer nanocomposites, graphene can be used as a sensor for biochemical or microbial changes in the product packed to detect specific food-borne pathogens or gases. Thus, this type of packaging can serve as oxygen and spoilage indicators for food safety and quality monitoring.⁹⁰ Due to the advanced features required for the food packaging industry, we highlight in this review graphene-based nanocomposites. Specially graphene oxide is a promising packaging material due to its unique characteristics, such as high barrier properties; good mechanical, electric, and thermal properties; large specific surface area; high electrical conductivity; carrier mobility; antibacterial, antifungal antioxidant, and biocompatibility.^{84,87,89,90,92–101}

The electrical properties mean that the food in the package can be quickly heated and provide a package that “speaks to the consumer”. The gas barrier means that the food can be preserved for longer, preventing oxidation and external contamination by microorganisms.^{80,83,91–95}

Since all physicochemical properties of food packaging materials can be improved using graphene-based composites, a closer look at the graphene family exerts a pivotal role in better understanding the final product. The following section will present a detailed discussion regarding the graphene family and their nanocomposites.

3. Graphene-based nanocomposites applied to food packaging

3.1. Graphene, graphene oxide, and reduced graphene oxide

Graphene (Gr) was introduced in 1986 by Bohen's group, aiming to understand a two-dimensional atomic sheet of graphite. However, in the 21st century, discoveries concerning graphene were made, and it was observed that graphene was thermodynamically unstable under ambient conditions. In 2004, Novoselov's group isolated and characterized a graphene monolayer obtained by mechanical exfoliation under ambient conditions. IUPAC defines graphene as a single layer of aromatic polycyclic carbon of almost infinite size, a single flat layer of carbon atoms with sp^2 hybridization, compacted together in a dense shape-shaped grid in two ordered dimensions (2D),^{11,12,39,40,99,100} Fig. 7.

The graphene unit cell has a hexagonal shape and comprises two equivalent sub-networks of carbon atoms, connected by sigma (σ) bonds, where the length of this C–C bond is 0.142 nm. Each carbon has a pi (π) orbital in this network that generates a delocalized electronic effect. These bonds and structures provide excellent properties that are highly superior compared to other nanomaterials such as clays, zeolites, zinc oxide, and silicate-based.^{11,12,39,40,99,100}

Graphene presents several interesting electronic properties due to its two-dimensional framework.^{101–103} High carrier mobility is observed, reaching $15\,000\text{ cm}^2\text{ V}^{-1}\text{ s}^{-1}$ at room temperature (or $200\,000\text{ cm}^2\text{ V}^{-1}\text{ s}^{-1}$ when impurity scattering is minimized). The high carrier is explained by promoting electrons in the valence band to the conduction band, forming a positively charged space that allows the current formation under an external electric field.^{101–103}

The structure of graphene can be modified to generate different structures derived from graphene (graphene-based materials), thus presenting other physicochemical properties.^{7–13,19–23}

GO (Fig. 8) possesses reduced thermal and electrical properties than Gr. However, the formation of polar groups in the GO structure favors its application in composites and polymeric nanocomposites.¹¹ The structure of the GO is still the target of several discussions in the literature,^{7–13,19–23} in which physicochemical studies indicate that epoxy and hydroxyl groups are created – usually in the basal planes of the GO leaves – and carboxyl groups commonly in the leaf edges or the pores of the

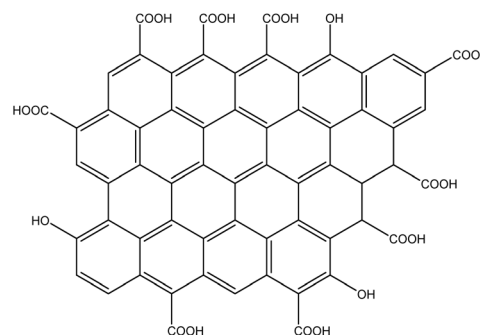


Fig. 9 Structure of reduced graphene oxide (rGO).



GO structure. Also, it a variation in the amount, location, and functional available in the GO structure is observed, which can be controlled by the synthesis conditions and methods as a function of the specific application and thus reflected the desired properties.¹¹

Graphene oxide may present insulating or semiconducting behavior, possessing sp^2 and sp^3 -bonded carbon atoms in its honeycomb structure. Oxygenated groups such as epoxy and hydroxyl groups are usually covalently bonded with sp^3 hybridized carbons, hindering their conductivity and leading to higher hydrophilicity.^{103–106} Recent works stated that depending on the oxygen content, GO might present a bandgap from 1.7 eV (semiconductor) to 2.1 eV (insulator), having a higher electrical resistance than graphene.¹⁰⁷

Generally, graphene oxide (GO) synthesis uses well-known methods described by Staudenmaier,¹⁰⁸ Brudie,¹⁰⁹ and Hummers.¹¹⁰ In this manner, Staudenmaier,¹⁰⁸ and Brodie,¹⁰⁹ in a pivotal study, oxidized graphite with a combination of HNO_3 and $KClO_3$. Furthermore, H_2SO_4 , HNO_3 , and $HClO_4$ have also been used to promote graphite oxidation, as described by Staudenmaier,¹⁰⁸ Brudie,¹⁰⁹ and Hummers' method.^{110,111}

However, the most used GO preparation methods considering biomedical and food packing purposes are the Hummers' method¹¹⁰ and the modified Hummers' method.^{108,112} Classical Hummers' process involves the treatment of graphite with H_2SO_4 , $NaNO_3$, $KMnO_4$ and H_2O_2 .¹¹⁰ The modified Hummers' method uses H_2SO_4 , $KMnO_4$, and H_2O_2 without the addition of $NaNO_3$, which generates less toxic gases during the procedure.¹¹² $KMnO_4$ is one of the most potent oxidizing agents in an acidic medium. In concentrated H_2SO_4 , this oxidizing agent provides complete graphite intercalation, forming graphite bisulfate intercalated in all the graphene layers. This intercalation favors the diffusion of the $KMnO_4$ and thus leads to effective graphene oxidation to graphene oxide without the need for $NaNO_3$.^{113,114} Furthermore, other variations of the Hummers' method are reported, by the replacement of H_2SO_4 with a mixture of H_2SO_4/H_3PO_4 , causing a less exothermic process without the formation of hazardous gases and also leading to more considerable oxidation in graphite, higher hydrophilicity and a more regular structure due to the cyclic phosphates creation.¹¹⁵

From a sustainable point of view, greener methods for synthesizing graphene and graphene oxide are reported, such as the synthesis using pyrolysis biochar from the lignocellulosic biomass leaves of *Cinnamomum Camphora*, in which the π - π interaction of the biochar suspension with D-tyrosine solution promotes the separation of Gr layers by centrifugation.²² Other graphene-like carbons from biomass pyrolysis methods found in the literature are the salt-based method, chemical blowing technique, model-based confinement, coupling to hydrothermal carbonization treatment, and post-exfoliation.²³

Notwithstanding, changes in GO structure can also be carried out by a reduction reaction and generating rGO. This reduction consists of partially removing oxygenated groups, thus conferring excellent structural stability compared to GO,¹⁰ Fig. 9.

GO reduction to rGO can be carried out by chemical^{17,18} or thermal^{19,116} routes. Several reducing agents are used in the

chemical reduction, such as hydrazine, sulfur compounds, sodium borohydride, ascorbic acid, tyrosine, dimethylhydrazine, ethylenediamine, and others reducing agents and reducing atmosphere.^{17,18} The first publication regarding GO reduction by chemical route was in 2006, in which it was used hydrazine and dimethylhydrazine as reducing agents. The nitrogen atom from hydrazine possesses a nucleophilic character, essential to reducing GO. In addition, other examples of reducing nucleophilic compounds are amines, hydrazine derivatives, and sodium borohydride. The reducing agents can also be electrophilic, such as borane in THF solution, one of the most common electrophiles for GO reduction.¹⁷ Furthermore, another way to reduce GO is to treat the suspension in an acid solution under reflux. The number of oxygenated species to be reduced can be easily controlled by the reflux time.¹⁷ Still, considering the chemical methods for rGO preparation, most of them use a lot of solvents and hazardous reductants, thus making the process expensive and non-sustainable due to the large amount of waste produced. In this way, other processes of chemical reduction of GO that use more sustainable reducers, such as tryptophan and ascorbic acid, have been reported. However, in these reduction processes, the agglomeration of rGO particles can occur, reducing their surface area.¹⁹ The thermal reduction process involves submitting GO to rapid heating under an inert atmosphere (Ar , N_2). Firstly, GO is dried, placed in a quartz tube and subjected to high temperatures (in the range of 700 to 1000 °C) for a short time, usually less than 1 h. During this thermal decomposition, H_2O , CO_2 and CO are released, which increases the internal pressure, causing the separation of the leaves, forming the rGO.¹¹ The amount of oxygenated groups remaining in the rGO depends on the initial oxygen concentration in the GO, the proportion of epoxy and hydroxyl groups in the GO, and the temperature used to reduce the GO.¹¹⁷

In this way, chemical and thermal reduction methods generate rGO with similar electronic and structural properties. However, the thermal protocol does not require a purification step or solvents since it only uses temperature as a reducing agent, making it a faster and cheaper protocol.^{17,18,116} Notwithstanding, decomposition gases (CO_2 and CO) can be collected and used as reagents in several green chemical processes.^{118–121}

Thus, graphene-based materials have a high potential for application in the production of new energy storage batteries, magnetic sensors, materials, superconductors, medication administration, ultra-sensitive biosensors, tissue engineering, catalysis, nanocomposites and polymeric nanocomposites, and food packing.²¹ Considering the encouraging features of GO and rGO, such as chemical, mechanical and antimicrobiological properties, these materials are highly promising to be used in food packing composites.

3.2. Graphene-based nanocomposites – properties

It is essential to select the most suitable food packaging for storage and transportation to maintain food quality, *i.e.*, from the moment they leave the industry until reaching consumer usage. Also, proper packaging provides food with safety,



lifetime extension, and protection against insects, microorganisms, light, and chemical contaminants. Furthermore, appropriate packaging can control the humidity, oxidation, diffusion of gases, and taste-odor maintenance.^{57,70}

The molecular structure of the polymeric materials used for food packaging directly influences the polarity, rigidity, malleability, crystallinity, and intermolecular forces present in the polymeric chain, controlling the diffusion of the external molecules into the packaging. The chemical polymer composition can alter the barrier properties, impacting the characteristics of resistance, rigidity, cost, efficiency, ease of processing, and increasing its useful life.^{57,70}

Polar polymers have good gas barrier properties. However, they are hydrophilic, and they allow water vapor permeation. On the other hand, nonpolar polymers do not allow the water vapor passage but have weak barrier properties, facilitating gas permeation independently of the polymer thickness. Copolymerization or additives incorporation in the polymeric matrix for food packaging formulations may affect these barrier properties as they fill and reinforce the polymeric matrix-free spaces. However, this will depend on the degree of adhesion and polymer and additive compatibility.^{57,70}

Knowing the polymers' thermal properties is essential to understanding the physical and chemical transformation in nanocomposites structure with the temperature variation. The glass transition temperature, crystalline melting temperature, and crystallization temperature are the most critical transformations in these materials.⁷⁰

3.3. Graphene-based preparation for nanocomposite production

In this way, the encouraging features of GO and rGO, such as chemical, mechanical and antimicrobiological properties, these

materials are highly promising to be used in food packaging composites.²¹ Generally, before producing graphene-based nanocomposites, the Gr should be in the GO or exfoliated forms (eGO). Therefore, incorporating the polymeric matrix into the structure of the eGO can be carried out, generating nanocomposites of commercial interest. GO exfoliation can be performed by two main routes: (i) thermal route; (ii) using polar solvents (water, ethanol, dimethylformamide – DFM, methyl pyrrolidone – NMP, propylene carbonate – PC) under stirring or ultrasonication.^{19,37} In the thermal process, the powdered GO is subjected to high temperatures (400–1000 °C) and a high heating rate for a few seconds. Then, the decomposition of the –OH and –COOH groups on the GO surface into CO, CO₂, and H₂O occurs. This process is advantageous since it produces a thermally exfoliated GO (TeGO) with a high surface area (700–1500 m² g^{−1}), low apparent density, and platelets similar to the eGO platelets produced using the sonication process.¹⁹

The suspension is electrostatically stabilized in the solvent and ultrasound-assisted method by the carboxylated groups present in the GO lamellae peripheries.¹⁹ The GO exfoliation performed by the sonication process in aqueous suspension furnishes GO monolayer platelet with high yield and hence forming eGO monolayers. However, the exfoliation process with agitation formed eGO monolayers with larger platelets, and this process is time-consuming and presents a low yield.^{19,37} Fig. 10 shows the graphene oxide (GO) and exfoliated graphene oxide (eGO) structures, layers, and platelets.¹⁹

The exfoliated platelets by stirring or ultrasound can be later reduced by chemical processes. These processes involve the formation of a suspension between eGO and a reducing agent, such as sodium borohydride, hydrazine monohydrate, or dimethylhydrazine, in a polar solvent (water, NMP, DMF, DMSO) or a mixture of these solvents.¹⁹

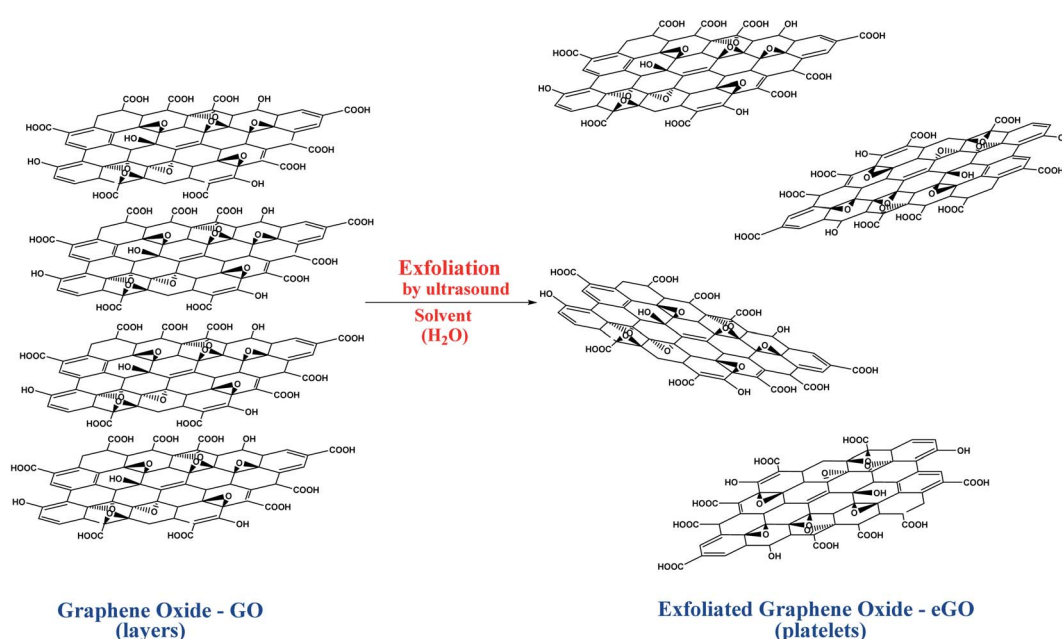


Fig. 10 Exfoliation of GO layers to eGO platelets using solvent and ultrasound.



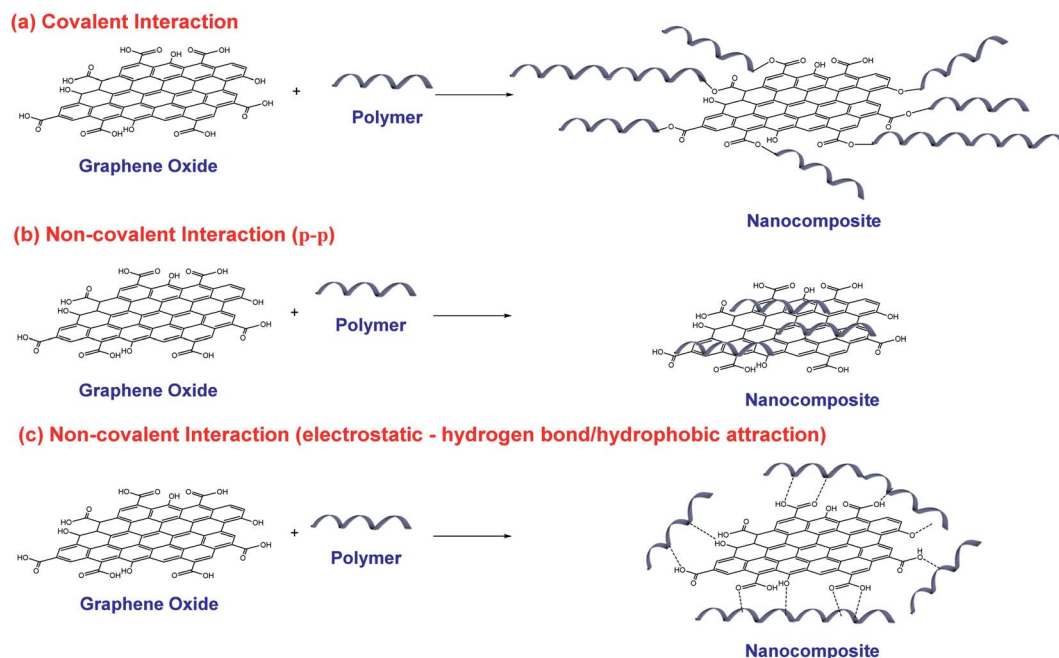


Fig. 11 Types of functionalization of GO with polymers for nanocomposites synthesis: (a) covalent functionalization; (b) π - π non-covalent functionalization and (c) electrostatic non-covalent functionalization.

Generally, chemically reduced eGO platelets result in materials (erGO) with a C : O ratio = 10 : 1, in which the oxygen content is related to the -OH and -COOH non-reduced groups in the eGO structure.^{19,37} Biochemical reducing agents such as vitamin C (ascorbic acid) and tryptophan have also been tested but have resulted in the aggregation and reduction of the eGO platelet surface area.^{19,122,123}

GO, rGO, eGO, and erGO structures have interesting functional groups, such as hydroxyl and epoxy, carboxyl, carbonyl, ketone, phenol, and lactone groups. These moieties allow the insertion of new atoms, molecules, or macromolecules (thermoplastic, thermosetting and biodegradable polymers) and thus creating the graphene-based nanocomposites.^{19,37,124}

The functionalization of graphene-based nanomaterials can be performed by covalent or non-covalent pathways. The covalent functionalization occurs from creating the covalent bonds between functional groups on the eGO platelets surface with the polymeric matrix,^{16,19,37,38} (Fig. 11(a)). This pathway can be carried out by nucleophilic substitution reactions, electrophilic substitution, condensation, cycloaddition, esterification, and hydrolytic condensation. In this manner, covalent functionalization is a propitious procedure for adjusting the charge transfer interface but impairs the electrical properties of the formed nanocomposite. On the other hand, non-covalent functionalization involves π - π interactions (Fig. 11(b)), hydrogen bonding (Fig. 11(c)), and hydrophobic attraction.^{16,19,37,38}

Covalent and non-covalent functionalization generates stable dispersions in organic solvents and can improve the compatibility of GO platelets with the polymers to be inserted in the nanocomposite structure.^{16,19,37,38}

Graphene-based/polymer nanocomposites can be prepared by different methods such as: mixing (MX), melt-mixing (MM),

in situ polymerization (ISP), and layer-by-layer (LBL) assembling. Here, it is worth mentioning that the chosen method will influence the properties of the final nanocomposite, such as surface area, molecular interaction, dispersion, and other properties.^{16,19,37,38}

The MX method consists of adding a polymeric matrix solution to a graphene-based suspension. The resulting mixture is subjected to sonication, mechanical stirring and solvent evaporation. This method effectively obtains a fine dispersion of the polymeric material, but it uses non-benign solvents, and removing the solvent is difficult. As an example, the synthesis of graphene-based/poly(vinyl)alcohol nanocomposite.^{16,19,37,38}

The MM technique is the most efficient and environmentally benign since solvents are not used. This method is commonly applied when *in situ* polymerization is not indicated, generally for thermoplastic polymers. The synthesis consists of melting the polymer and dispersing the graphene-based material through an extrusion and injection molding. However, the polymer can undergo thermal degradation, and the structure of the graphene-based nanocomposite can be disrupted by the high shear force involved during synthesis. For example, MM is applied to synthesize the graphene-based/polypropylene, graphene-based/poly(lactic acid), and graphene-based/polyethylene terephthalate nanocomposites.^{16,19,37,38}

ISP protocol is used to disperse the graphene-based material homogeneously in the polymeric matrix. In this technique, the graphene-based material is dispersed in the monomer followed by the polymerization reaction, with subsequent polymer grafting over the graphene-based surface. *In situ* polymerization uses heat or microwave radiation to form nanocomposites. Using ISP, a more homogeneous and fine dispersion occurs between graphene-based and the polymeric matrix, being very



Table 1 Non-renewable polymeric matrix/graphene-based group^a

Nanocomposite types	Matrix : graphene-based ratio and others	Synthesis method	Polymers properties modifications and improvements	Important comments and applications	Ref.
PP-Gr	-Gr-PP 3 vol%; extrusion at 180 °C for 3 min; injection molding at $T_{\text{barrel}} = 180\text{ °C}$ and $T_{\text{mold}} = 80\text{ °C}$, injection pressure at 1.1 MPa	MM/IM	-↓ Thermal conductivity -↓ O ₂ barrier	-Viscoelastic properties allow the fabrication of composites with desired properties at high loadings -Improvement of physical, thermal, and mechanical properties -Can be applied to food packaging -Gr uniformly dispersed in the matrix	Kalaizidou and co-workers (2007) ²⁴
PET-Gr	-Gr 1.5 wt%, sandwiched between two PET films, hot pressed, and chopped to granules -Melt compound at 260 °C for 5 min -Molded into thin films (0.17 mm thick) -Dried under vacuum at 120 °C for 4 h	MM	-↓ Thermal stability -↓ Young's modulus, 1.16 to 1.40 GPa -↓ Elongation at break, 40% -↓ Tensile strength, 56%	-Enhanced crystallinity -The formed films present Gr exfoliated morphology -Gr addition enhanced brittleness	Harel and co-workers (2012) ²⁶
PS-GO	-2 g of PS, GO 20 wt%, and 4-vinyl benzyl chloride (VBC) -Stirring -Dried for 24 h on a glass plate	ISPs	-↓ O ₂ barrier -↓ Crystallinity -↓ Hydrophobicity -↓ Thermic stability -↓ T_g -↓ Young modulus, 60% -↓ Impact resistance -↓ Water vapor transmission rate, 50 to 16.6 wt	-Can be applied to food packaging The nanocomposite showed high barrier and antimicrobial properties and could be utilized in yogurt containers and bottles for medicine capsules -Applied to food packaging	Ghanem and co-workers (2020) ²⁵

^a Gr: graphene; GO: graphene oxide; PP: polypropylene; PS: polystyrene; PET: polyethylene terephthalate; MM: melt-mixing; ISPs: *in situ* polymerization with solvent; IM: injection molding. ↑: increase; ↓: decrease.



Table 2 Renewable polymeric matrix/graphene-based group^a

Nanocomposite types	Matrix : graphene-based ratio and others	Synthesis method	Polymers properties modifications and improvements	Important comments and applications	Ref
CS-GO	-GO + water, sonication for 45 min -CS + acetic acid solution 0.5% v/v (water) -GO : CS ratio = 1 : 1, GO suspension was added on CS solution and stirring for 24 h -Dried at 40 °C, in glass plate until weight equilibration	MX	-↑Young's modulus, 64% -↑Tensile strength, 88% -↑ <i>T_g</i> , 175.4 to 180.4 °C	-Good GO dispersion on CS matrix -The CS-GO nanocomposite formation occurred by H bonding between CS molecules with GO oxygenated groups and by electrostatic interaction between polycationic CS and the negative charge of GO surface -Can be applied to food packaging	Li and co-workers (2010) ¹²⁹
PVA/GO and PVA/rGO	-PVA/GO and PVA/rGO: 0.3 wt% -PET: a substrate	MX	-↓O ₂ permeability, 17–87 times bigger -73% light transmittance at 550 nm	-Promising for the development of transparent high-gas-barrier films -The H-bonding interaction between GO and PVA is not very large to change the PVA thermal properties -Can be applied to food packaging and bottles -Environmentally benign	Lee and co-workers (2011) ²⁷
PLA/GO	-Hybrids solutions were cast onto PET film at 90 °C -GO 0.4 wt%, dispersed in acetone, sonicated for 5 h -GO + PLA/chloroform (plasticizer) solution, sonicated for 15 min -PLA/GO films formed in polytetrafluoroethylene, coated plate -Dried at room temperature for 7 days	SC	-↑Young's modulus -↑Tensile strength -↑O ₂ and N ₂ barrier	-Possible biomedical application -Formation of transparent films	Magalhães and co-workers (2013) ¹³⁰
PVA-rGO	-GOR 0.8 wt% -PVA aqueous solution (1.0 g mL ⁻¹), 90.2 wt% -Glycerol (plasticizer), 9 wt% -Stirred at 95 °C for 24 h -Dried 2 days, 45 °C, in a glass plate, after 80 °C for 24 h	MM	-↓Elongation at break, 451.9 to 229.1% -↑Tensile strength, 35.7 to 39.9 Mpa -↑Thermal stability -↑O ₂ barrier	-No detectable effects in the elongation at break -Can be applied to food packaging -GO reduction process result in a good dispersion in PVA matrix -Can be applied to food packaging	Hu and co-workers (2013) ¹³¹



Table 2 (Contd.)

Nanocomposite types	Matrix : graphene-based ratio and others	Synthesis method	Polymers properties modifications and improvements	Important comments and applications	Ref
PVA-CS-rGO	-rGO 0.8 wt%	MM	-↓ Elongation at break, 102.3 to 76.5%	-GO reduction process result in a good dispersion in CS-PVA matrix	Hu and co-workers (2013) ¹³¹
	-PVA 63.1 wt%		-↑ Tensile strength, 46.2 to 64.5 Mpa	-PVA/CS/rGO showed improvement in mechanical and thermal properties	
	-CS 27.1 wt%		-↑ Thermal stability	-Can be applied to food packaging	
	-Glycerol (plasticizer), 9 wt% -Stirred at 95 °C for 24 h -Dried 2 days at 45 °C, in glass plate, after 80 °C for 24 h -PLA: 99.84 wt%		-↑ O ₂ barrier		
PLA/PEI-GO	-PEI: 0.1 wt% in deionized water	LBL	-↓ O ₂ permeability, ≈ 99%	-Use on biodegradable packaging materials	He and co-workers (2014) ¹³²
			-↑ Tensile strength 93.2 to 120.2 MPa	-Increasing GO concentration in the nanocomposite structure can reduce oxygen permeation	
	-GO: 0.06 wt%		-↑ Elongation at break, 57.5 to 63.3%	-Can be applied to food packaging	
	-XGO is GO functionalized with oxygen groups	MM	-↑ Young's modulus, 27.6 to 37.8 MPa	-The PVA-XGO mechanical and barrier properties depend on the dispersion and the alignment of graphene-based in PVA matrix	
PVA-XGO	-XGO 0.3 wt%		-↑ Tensile strength, 25.4 to 37.9 MPa	-Can be applied to food packaging	Loryuenyog and co-workers (2015) ¹³³
	-PVA solution aqueous 10wt%, 20 g		-↑ Elongation at break, 260 to 317%		
			-↑ O ₂ barrier		
			-↑ Water vapor barrier		
CH-PAA-rGO	Solvent: deionized water	LBL	-↑ Thermic stability	-Different pH's may affect the thickness of the film and also the permeability of O ₂	Grunlan and co-workers (2015) ¹³⁴
	-CS: 0.1 wt% (pH = 3.5; 4.5; 5.5)		-↑ T _g : 51.8 to 53.5 °C	-In comparison with PET, the O ₂ barrier of this nanocomposite is 20 times higher	
	-PAA: 0.2 wt% (pH = 3, 4 and 5)		-↑ T _m : 175.7 to 178.2 °C	-The authors performed a thermal reduction of GO to increase the barrier and gas selectivity of this material by increasing the hydrophobicity of the film	
			-↑ Crystallinity: 24.8 to 28.0%	-Can be applied to food packaging	



Table 2 (Contd.)

Nanocomposite types	Matrix : graphene-based ratio and others	Synthesis method	Polymers properties modifications and improvements	Important comments and applications	Ref
PLA-rGO	-Exfoliated GO: 0.1 wt% (by sonication) -A support was used to deposit a thin layer of each polymer and GO. -Thermal reduction was performed at 175 °C for 90 min -The assembling process comprised a freestanding rGO film between two pieces of PLA film with heat pressing compression: 2000 pounds-force at 65 °C for 30 min	LBL	-↑ Gas selectivity (H ₂ and CO ₂)	-Promising solution to food packaging with outstanding environmental sustainability	Chen and co-workers (2016) ^{13,5}
	-50 mL GO solution (by sonication): obtain a homogeneous and stable dispersion solution -CS 1 wt%: dissolving CS in 0.5% (v/v) aqueous acetic acid solution -GO was dropwise into the CS solution		-↑ Hydrophobicity -↑ Resistance towards moisture penetration of 87.6% -↑ O ₂ barrier, 99%		
	-Stirring for 24 h -Ratio GO : CS : TiO ₂ (1 : 20 : 4); NPs ₂		-The spectrum of GO-CS nanocomposites exhibited neither a peak at 1596 cm ⁻¹ related to -NH ₂ absorbance vibration nor a peak at 1730 cm ⁻¹ related to the C=O stretch of the carboxylic group		
GO-CS		MX		-Can be applied to food packaging	Xu & Liu (2017) ^{9,3}
GO-CS-TiO ₂	-50 mL GO solution (by sonication, 60 W): obtain a homogeneous and stable dispersion solution -CS 1 wt%: dissolving CS in 0.5% (v/v) aqueous acetic acid solution	MX	-Absorption bands of oxygen-containing functional groups were dramatically reduced -C-OH and carbonyl C=O bands were observed at 1200 and 1729 cm ⁻¹ , respectively -Strong absorption bands at 450 and 670 cm ⁻¹ , indicating the presence of the Ti-O-Ti bond in TiO ₂	-Non-toxic -Exhibited significant antimicrobial effects against <i>B. subtilis</i> and <i>A. niger</i> biofilm formation -Can be employed as a cling film that effectively delays the loss of moisture in fruits and vegetables	Xu & Liu (2017) ^{9,3}
	-GO was dropwise into the CS solution -Stirring for 24 h -NaOH solution: adjusted pH to 6.5 -200 µL 25% glutaraldehyde solution (magnetic stirring overnight) -Centrifuged at 6000 rpm for 1 h -Washed three times			-Can be applied to food packaging	



Table 2 (Contd.)

Nanocomposite types	Matrix : graphene-based ratio and others	Synthesis method	Polymers properties modifications and improvements	Important comments and applications	Ref
Cellulose-GO	-The supernatant was discarded, and the deposit was freeze-dried for 12 h in freeze dryer	MX			Huang and co-workers (2016) ³²
	-Cellulose/16 wt% GO and cellulose/4 wt% GO		-↓ O ₂ permeability, 99.85%	-CPAM prevents the GO self-aggregation in the fiber matrix	
	-Cationic polyacrylamide (CPAM)		-↓ Burst strength 81.4% (4 wt% GO)	-Potential packaging applications	
CS _{0.5} CA-Gr (6 wt%)	-CPAM/GO: 0.04 : 1 wt%	MX		-Permit the large scale preparation of GO/cellulose paper with high doping amount of GO	Demetri and co-workers (2016) ¹³⁶
	-2 g CS was suspending in acetic acid solution (0.1 M)		-↓ Elastic modulus, 2.5 to 3.8 GPa	-Can be applied to food packaging	
	-Immersion in thermostatic bath: 25 °C		-↓ Fracture strength, 50 to 90 MPa	-Presence of expanded graphite led to higher values of mechanical properties, likely due to the increase in chain entanglements	
	-Mechanical stirrer: 2 h		-↓ Elongation at break, 15 to 9%	-Small concentrations of CA are sufficient to functionalize chitosan and are effective against the natural proliferation of mold	
	-Gr: 6 wt% -CA: 0.5 wt% -Under controlled temperature: 25 °C		-↓ Efficiency against mold	-Can be applied to food packaging	
CS-GO	-Chitosan (CS): water solution 1 v/v% mixed with acetic acid solution 2 wt%	MM	-↑ Young's modulus, 22.7 to 5843.7 MPa	-Potential application for food packaging	Advincula and co-workers (2017) ¹³⁷
	-GO 0.6 wt%		-↑ Tensile strength, 32.4 to 43.27 MPa	-CS-GO showed higher antimicrobial properties than CS	
	-Sonication at 60 °C for 1 h, dried at 120 °C overnight		-↓ Thermal stability	-CS does not show toxicity for bacteria	
PLA-GO	-PLA + GO, 1 wt%	MX	-↑ Tensile strength 32.4 to 40.6 Mpa	-Can be applied to food packaging	Ahmed and co-workers (2017) ⁸⁹
	-GO and PLA mixed and sonicated for 30 min (was added polyethylene glycol as plasticizer)		-↓ Elongation at break, 41 to 31.6%	-Demonstrated antibacterial activity in food packaging	
			-↑ T _g , +5 °C -↑ T _m , 81.5 to 84.8 °C -↓ O ₂ barrier, 40%	-Can be applied to food packaging	



Table 2 (Contd.)

Nanocomposite types	Matrix : graphene-based ratio and others	Synthesis method	Polymers properties modifications and improvements	Important comments and applications	Ref
Gly-nanocellulose- GO (Glycerol/ Nanocellulose/ GO = GGN)	- GO 0.5% wt - GO aqueous solution sonicated for 3 h	MM	-↑ Tensile strength, 18.4 to 25.0 MPa -↓ Thermal stability	-Glycerol and GO in the nanocellulose matrix led to synergistic effects -GGN films show potential to be used in the food package since they can bear heavier products than a zip-lock bag -Can be applied to food packaging	Arcoot and co-workers (2017) ¹³⁸
	-Glycerol 40 wt% (plasticizer) -GO, glycerol, and cellulose were mixed -Films formed in Petri dishes; dried at 40 °C for 2 days		-↑ Elasticity -↑ Elongation at break, 1.9 to 9.4% -↑ Contact angle of the films -↑ Moisture sorption -↑ Water vapor permeability -↑ O ₂ barrier -↑ Tensile strength 58.5 to 17.2 MPa		
OS-rGO (OS: oxidized starch)	-rGO: 1.0 wt% -Plasticizing the OS: mixture and stirring (OS: rGO) at 90 °C, 300 rpm for 60 h	MM	-↓ O ₂ permeability	-The nanocomposite mechanical property can be controlled by the rGO reduction time -Can be applied to food packaging	Jiang and co-workers (2017) ¹³⁹
PLA-CNC/Gr	-PLA/CNC/Gr: 95/0.5/0.5 wt%, prepared by melt compounding -Stirring 70 rpm at 185 °C for 10 min	MC	-↑ Young's modulus, 8% -↑ Tensile strength, 11% -↑ O ₂ barrier, 23% -↑ Hydrophobicity -↑ Thermic stability -↑ <i>T_g</i> : 51.8 to 53.5 °C -↑ <i>T_m</i> : 175.7 to 178.2 °C -↑ Crystallinity: 24.8 to 28.0%	-Good GO/matrix dispersion -Significant improvement in the antifungal activity by Gr -↑ Transparency loss on materials -Potential applications for food packaging	Montes and co-workers (2018) ¹⁴⁰
EVOH-GO	-GO: 0.5 wt% -EVOH: 99.5 wt%	MM	-↑ Electrical conductivity -↑ <i>T_g</i> -↑ <i>T_c</i>	-Excellent GO dispersion on polymeric matrix -Can be used in smart food packaging	Lagaron and co-workers (2018) ¹⁴¹
GO-CS	-GO/EVOH sonication 15 min -Electrospinning process -50 mL GO solution (by sonication): obtain a homogeneous and stable dispersion solution	MX	-The spectrum of GO-CS nanocomposites exhibited neither a peak at 1596 cm ⁻¹ related to -NH ₂ absorbance	-Can be applied to food packaging	Xu & Liu (2017) ⁹³



Table 2 (Contd.)

Nanocomposite types	Matrix : graphene-based ratio and others	Synthesis method	Polymers properties modifications and improvements	Important comments and applications	Ref
PLA/starch-Gr	-CS 1 wt%: dissolving CS in 0.5% (v/v) aqueous acetic acid solution -GO was dropwise into the CS solution -Stirring for 24 h -PLA/starch-Gr, 0.1 wt% of Gr	MM	vibration nor a peak at 1730 cm ⁻¹ associated with the C=O stretch of the carboxylic group	-Suitable for food packaging application -Can be applied to food packaging	Auras and co-workers (2018) ¹⁴²
	-Vacuum oven overnight -The extrusion was pelletized at 50 °C for 4 h		-↑Elongation at break, 103.4% -↑Toughness, 500–900% -↓Young's modulus, 1.2 to 0.8 GPa		
	-Starch/gelatin-GO matrix, 99.15 wt% -Starch/gelatin ratio 1 : 3		-↓O ₂ permeability, 50% -↓Tensile strength 57.97 to 76.09 MPa -↑Young's modulus, 20.59 to 35.91 MPa		
	-GO 0.85 wt% -Sonication for 30 min		-↓Elongation at break, 6.6 to 3.13% -↑Thermal stability -↓Water vapor permeability -↑Zone of inhibition -↑Antibacterial activity under visible light		Baniasadi and co-workers (2018) ³¹
PVA/Cu ₂ O/TiO ₂ -rGO	-Cu ₂ O-TiO ₂ /rGO -Synthesized by ultrasonic reduction and wet impregnation method using TiO ₂ NPs, Cu(NO ₃) ₂ 3H ₂ O, and GO -PLA-Cu ₂ O-TiO ₂ /rGO -PVA 3 wt%	SC	-↓Antimicrobial activity	-Environmentally benign -Uniform distribution of the particles in the PVA films -Effective antimicrobial activity under visible light against four different microorganisms -Potential for ambient light food packaging	Venkatapras and co-workers (2018) ¹⁴³
PLA-GO	-Cu ₂ O-TiO ₂ /rGO 12.5 mg mL ⁻¹ -PLA + Cu ₂ O-TiO ₂ /rGO, solutions were stirred -Dried in Petri plates at room temperature for 48 h -PLA/0.5 wt% GO (10 : 90)	MM	-↓Water vapor permeability -↑T _m , 165.9 to 170.2 °C -↓Young's modulus, 30% -↑Thermal stability	-Lightweight and strong packaging materials for food and industrial applications -Homogeneous dispersion of fGO in PLA-ST matrix	Peijs and co-workers (2018) ²⁸
PLA-ST-fGO	-fGO: GO functionalized with maleic anhydride and dodecyl amine -4 g of PLA + 0.45 g of starch dissolved in chloroform -5 wt% tributyl citrate	SC	-↑Crystallization capacity	-Potential versatile nanohybrids for food packaging and pharmaceutical industries	Sheng & Xiong (2019) ¹⁴⁴



Table 2 (Contd.)

Nanocomposite types	Matrix : graphene-based ratio and others	Synthesis method	Polymers properties modifications and improvements	Important comments and applications	Ref
CS-rGO and CS-GO	-Stirred for 1 h -0.4 wt% fGO, added and stirred for 1 h -Dried at 40 °C for 4 days in a polytetrafluorethylene frame	MX	-↓ Plasticizer migration rates -↑ UV shielding capacity -↑ Hydrophobicity -↑ Aging resistance -↑ Storage modulus -↓ Electric conductivity, graphite 1200 to GO 3.1 S cm ⁻¹ and to rGO 60 S cm ⁻¹ -↓ GO and rGO dispersion on CS matrix	-The addition of rGO did not affect the CS matrix hydrophilicity	Malmonge & Basso (2019) ³⁰
	-GO or rGO + acetic acid solution 1% v/v (water), 20 mL sonication for 9 min -CS (0.4 mg) + acetic acid solution 1% v/v (water), 20 mL			-The GO and rGO addition on the CS matrix did not accelerate the composite degradation process -Can be applied to food packaging	
	-GO or rGO suspension was added to CS solution, stirring for 5 min and sonication for 9 min -Solvent removal and dried -Starch/5 wt%, PVA-GO				
Starch/PVA-GO	-GO: 2 mg mL -All films should be adjusted before being tested for temperature ($T \approx 24^\circ\text{C}$) and relative humidity ($\approx 50\%$) -22.5 mg GO in water, 2.71 mL, sonication 1 h -GO/water suspension was added in PVA-GA matrix and stirred at 6000 rpm 15 min -Dried overnight, at room temperature, on a glass plate	MX	-↑ Tensile strength 18.40 to 25.28 MPa -↓ Elongation at break, 184.03 to 143.78% - ↑ thermal stability	-Starch/PVA-GO composite film can resist the water molecules migration -Can be applied to food packaging	Lin & Pu (2019) ¹⁴⁵
PVA-GA-GO		MX	-↑ Young's modulus 0.66 to 1.55 MPa -↑ Tensile strength, 0.66 to 1.51 MPa -↑ Thermic stability	-PVA-GA-GO demonstrated antibacterial activity and can be applied to food packaging	Chowdhury & Mah (2020) ¹⁴⁶
PHB-Gr	-0.7 wt% Gr on PHB -Solvent: chloroform -Gr nanoparticles were dispersed in chloroform (1 mL) by	MX	-↓ Water vapor transmission rate, 38.49 to 32.13% -↓ O ₂ permeability, 1.53 to 0.4 mm per m ² per day ² per atm	-Was found to be environmentally safe and highly biodegradable -Applications in packaging of light-sensitive food products -High strength is desired for the packaging of heavy food items	Pakshirajan & Pugazhenthil (2020) ²⁹

Table 2 (Contd.)

Nanocomposite types	Matrix : graphene-based ratio and others	Synthesis method	Polymers properties modifications and improvements	Important comments and applications	Ref
	sonication for 45 min (1.5 s on cycle: 45 s off/cycle)		-↓ Water vapor permeability, 9.26 to 4 mm per m ² per day ² per atm		
	-Stirring at 100 rpm for 60 min		-↑ Thermal stability -↑ T _m , 172.8 to 182.5 °C -↑ Tensile strength, 4.5 to 9.0 MPa -↓ Elongation at break, 15 to 12.2% -↓ Transparency to UV and visible light		

^a Gr: graphene; GO: graphene oxide; rGO: reduced graphene oxide; GS: chitosan; PLA: poly lactic acid; PVA: poly(vinyl alcohol); PEI: polyethylenimine; ST: starch; GA: glutaraldehyde; CNC: cellulose nanocrystals; EVOH: poly(ethylene-co-vinyl alcohol); PAA: poly(acrylic acid); OS: oxidized starch; PHB: poly(hydroxybutanoate); MX: mixing; MM: melt-mixing; MC: melt compounding; PEP: pickering emulsion polymerization; ISPs: *in situ* polymerization with solvent; LBL: layer-by-layer; PS: plasticized-starch; SP: solution processing; SC: solution casting method. ↑: increase; ↓: decrease.

effective for both heating techniques. For example, this protocol is used to synthesize the graphene-based/polystyrene nanocomposite.^{16,19,37,38}

The LBL technique offers the possibility of building nanocomposite alternating layers between the graphene-based material and polymeric matrix. This synthesis method creates a nanomaterial with good dispersion and graphene-based orientation. Also, it allows the possibility of controlling the amount of graphene added between the polymer layers and the morphology of the generated nanocomposite. For example, the GO/poly(vinyl) alcohol nanocomposite construction can be mentioned.^{16,19,37,38}

Graphene-based nanocomposites have several applications in biosensors, energy-related devices, food packaging, and biomedical, among others. However, many challenges are still encountered, especially for materials used in health and food. However, many challenges are still faced, especially for health and food packaging materials. There are already successes in using graphene nanocomposites with polymeric matrices of renewable sources in drug delivery. The low solubility of graphene in an aqueous solution makes its implementation difficult. The aqueous stability and the solubility of graphene can be modified by the induction of π - π interactions with drugs, aromatic molecules, or other organic molecules. However, it is possible to circumvent it by replacing graphene with graphene oxide in the nanocomposite assembly. The advantages of using GO and rGO to construct polymeric nanocomposites consist of oxygenated groups (hydroxyl, epoxide, carboxyl, and carbonyl) on both GO and rGO nanosheets that favor the assembly of new nanometric structures and the new polymeric nanocomposite is formed. This assembly occurs through the stacking system between the nanomaterial and the polymeric matrix and between the hydrogen bonds and π - π interactions that will significantly improve the properties of the polymeric nanocomposite formed as biocompatibility, low toxicity, thermal detachment, and mechanical stability.^{125,126}

3.4 Graphene-based nanocomposites – types

Gr, GO, and rGO are used as polymeric matrices additives for several nanocomposites' formulations. In this sense, these polymeric nanocomposites formulations often present improved properties for food packaging.^{127,128}

Several research groups have been developing new materials for the manufacture of packaging based on nanocomposites, such as clays, tourmaline, zeolites, oxides, and carbon-based materials, such as graphene-based, to improve and control food packaging properties.^{25,33–36,51,61} In this sense, no polymeric matrix has all the necessary/desired properties in food packaging production. However, with the nanotechnology advent, polymeric nanocomposites (polymer matrix + nanomaterial) arise, improving their thermal, mechanical, electrical, optical, and barrier properties of food packaging.⁷⁰

The matrix/graphene-based nanocomposites will be divided into two groups in the next section: (a) non-renewable polymeric matrix/graphene-based group (petroleum-derived polymers)



and (b) renewable polymeric matrix/graphene-based group (biomass-derived polymers).

3.4.1. Non-renewable polymeric matrix/graphene-based group. Table 1 shows the main nanocomposites types comprised of the non-renewable polymeric matrix and graphene-based material, the experimental conditions, synthesis methods, and properties.

Table 1 shows that melt-mixing is the most used synthesis method for constructing polymeric nanocomposites (non-renewable origin). The addition of graphene-based nanomaterials increases the nanocomposite's thermal, barrier, and other properties.

3.4.2. Polymeric matrix renewable/graphene-based group. Table 2 shows the main nanocomposites based on renewable polymeric matrices, experimental conditions, synthesis methods, and properties modifications.

In this sense, Table 2 shows that MX and MM are the methods for synthesizing polymeric nanocomposites (from renewable sources) based on graphene most used by most research groups. It is due to their ease and simplicity of handling on a small scale. It is indisputable that the addition of graphene-based nanomaterials (Gr, GO, and rGO) generates a significant improvement in the various properties of these nanomaterials when compared to their pure polymeric matrices.

Improvements in polymeric nanocomposites occurred in their thermal, mechanical (high tensile strength, low elongation at break, increase in Young's modulus), and barrier properties (low permeability to water vapor; low permeability to O₂). Generally, there is also an excellent dispersion of Gr, GO, or rGO in the polymer matrix, generating environmentally safe and biodegradable nanocomposites. However, the addition of GO or rGO in the polymeric matrices did not accelerate the degradation process of the composite. Many graphene-based polymeric nanocomposites have demonstrated antimicrobial activity and can apply to food packaging.

A comparison between Tables 1 and 2 shows that several studies have been devoted to using renewable polymeric matrix/graphene compared to non-renewable-based composites. This perspective shows the great scientific concern for environmental causes. The creation of new packaging from nanocomposites is one of the most impressive and promising packaging technology results for the near future challenges.¹²⁴

4. Microbiological properties of graphene-based nanocomposites

Fungal and bacterial transmission in contaminated objects and surfaces is one of the vectors of disease transmission by these microorganisms, although it is not the main transmission route. They adhere to surfaces and colonize the plants, insects, animals, fabrics, food, medical apparatus, packaging, and hard materials. This transmission by adhesion to solid organic and inorganic surfaces is a relevant challenge to public health, epidemiological significance, and economic losses.¹⁴⁷ Depending on the surface type and its humidity condition, the microorganisms can survive and grow for extended storage periods, sometimes even months. Removing them is necessary to disinfect and destroy microorganisms by using organic substances and chemical disinfectants that can effectively combat these pathogenic microorganisms.¹⁴⁸

Another strategy is to use organic and inorganic compounds with fungal and bactericidal activities incorporated in polymeric matrices (fabrics, paper, paints, *etc.*) to eliminate microorganisms by contact. In this way, graphene and graphene-based materials emerge as a practical pathway to minimize the proliferation of these microorganisms.

Graphene is a biocompatible material with low or no cytotoxicity and promising for food packaging composites, and its oxidized form has different properties and antimicrobial effects in solution and when deposited on surfaces. In that perspective, several studies reported in the literature involving the graphene family and reporting its antibacterial activity^{148–157} (*e.g.*, fungi

Table 3 Selected recent applications of graphene-based nanocomposites as antimicrobials

Material	Microorganism	Ref
Reduced graphene oxide/Ag/Ag ₂ S	<i>E. coli</i>	165
Reduced graphene oxide/ZnO	<i>E. coli</i>	166
Graphene oxide	<i>S. aureus</i>	159
Graphene oxide-catechol	<i>E. coli/S. aureus</i>	165
Reduced graphene oxide/CuO	<i>E. coli/S. aureus</i>	166
Graphene oxide cellulose/CuO	<i>S. aureus, B. subtilis, E. coli/P. aeruginosa</i>	167
Graphene oxides NiS–MoO ₃	<i>E. coli/S. pyogenes</i>	168
Reduced graphene oxide/Ag	<i>E. coli/K. pneumoniae</i>	169
Reduced graphene oxide/Ag	<i>S. aureus/E. coli/P. aeruginosa</i>	170
Graphene oxide	<i>E. coli K12</i>	171
Graphene oxide/ <i>p</i> -aminophenol	<i>E. coli/S. aureus</i>	172
Graphene oxide/modified sodium anthraquinone-2-sulfonate	<i>E. coli</i>	173
Graphene oxide chloramine bromosuccinimide/FeCl ₃ /KIO ₃	<i>P. pneumoniae/S. aureus</i>	174
Graphene Oxide-chitosan/Ag	<i>Pseudomonas</i> sp	175
Graphene oxide/Ag	<i>E. coli/S. aureus</i>	176
Graphene oxide/ampicillin, chloramphenicol, or tetracycline	<i>P. aeruginosa</i>	177
Graphene oxide/NiS–In ₂ O ₃	<i>E. coli/S. aureus</i>	178



and bacteria) – making these materials highly promising for food packaging nanocomposites.^{158–161}

Regarding graphene-family composites, their antibacterial activity can be related to several intrinsic properties emerging from geometric and electronic structures.¹⁶² The most common mechanism accepted for the antibacterial activities of graphene is oxidative stress caused in the microorganism by transferring electrons to their membrane and deactivating its protein functions and lipids through the reactive oxygen species (ROS). Furthermore, also can be found studies in the literature reporting that antibacterial activity can occur when phospholipids are extracted by supramolecular interactions with graphene and damage in the permeability of the bacterial cell wall and/or membrane.^{163,164}

Moreover, the antimicrobial properties of the graphene family might be improved by the insertion of molecules by covalent bonds and the deposition of metal ions on its surface. These graphene-based nanocomposites materials may contain nanoparticles of metal ions/oxides/sulfides (manganese disulfide, cadmium sulfide), titanium dioxide, zinc oxide, copper oxide, polymers, antibiotics, among others.^{163,164}

Nanotechnology involving the graphene family and its composites can be a solution for coping with bacteria in various situations. Table 3 shows some selected most recent examples of graphene-based-nanomaterials with high efficiency against bacteria, such as *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas pneumonia*, and *P. aeruginosa*, *Staphylococcus aureus*, *Staphylococcus pyogenes*, and *Bacillus subtilis*.

Despite the importance of graphene-based nanocomposites for safer packaging with resistance against microorganisms, it is noteworthy to mention that the specialized literature lacks in filling the gaps regarding the risks of nanomaterials used. In this way, information regarding their interaction with cellular components, migration to the food, tolerance dosage levels, toxicity to the human organism, long-term exposure, and toxicity to ecologically organisms when disposed of incorrectly.

5. Graphene-based nanocomposites – toxicity and (bio)degradability

Graphene and its derivatives (GD) have chemical, mechanical, electrical, and optical properties of interest for different applications such as biotechnology, biosensors, biomedicine, nanocomposites, catalysis, and drug delivery systems, metal detection, and removal, among others. Because of this, many studies are being developed to assess the effects of GD toxicity on the environment and the human body.²⁹ GD toxicity is influenced by the shape, size, concentration, synthesis methods, and type of functional groups. For example, the PHB/graphene nanocomposite showed promising results in improving the food packaging life and high biodegradability. However, graphene in polymer nanocomposites can be released into the air and groundwater depending on size, shape, and concentration.^{155,179,180}

In the formulation of nanocomposite PVA/gelatin was added to cellulose nanocrystals to make food packaging. The addition of cellulose nanocrystals in the nanocomposite showed improved properties (mechanical, thermal, and barrier). The results indicate that PVA-gelatin films reinforced with cellulose nanocrystals can be considered a potential biodegradable packaging material, especially for food packaging.^{63,181} They can be tested in the construction of graphene-based polymeric nanocomposites.

Although the excellent adsorption capacity of graphene allows the capture of organic contaminants in aqueous media, DGs can promote the spread of these contaminants in groundwater, where DGs are more unstable and have greater mobility. Because of this, some research has been developed to evaluate the effects of these materials on aquatic organisms. It has been observed that the cell division of photosynthetic microorganisms is inhibited as the concentration of GD is high. On the other hand, invertebrates present several changes: changes in biochemical performances, the survival rate and inhibition of the swimming behavior of crustaceans, and

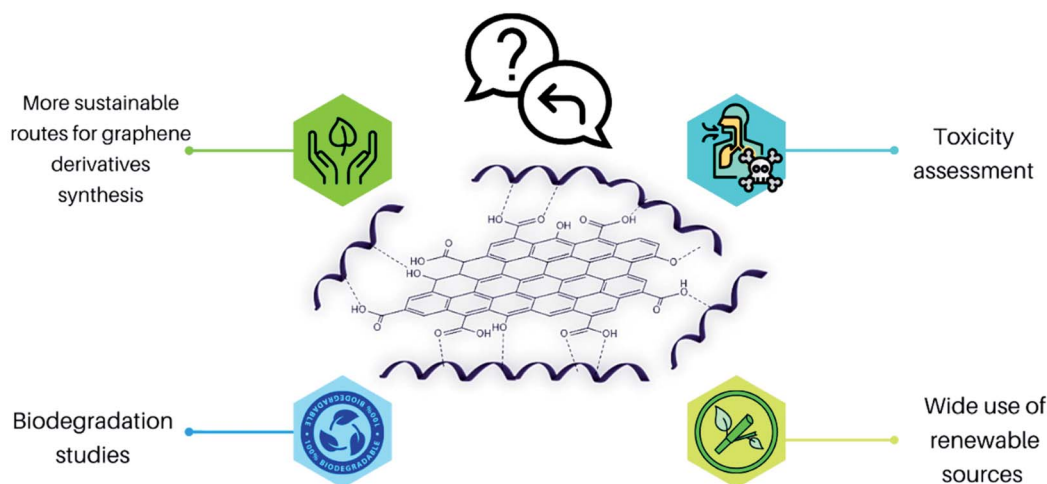


Fig. 12 Challenges in the field of graphene-based polymeric nanocomposites.



alterations in membranes. The impact observed in zebrafish was from inhibition of embryo growth to cardiac development.¹⁸²

Graphene and its derivatives have excellent properties such as good dispersibility and stability in human physiological environments, making them promising for application in biomedicine, drug delivery systems, and food packaging. However, studies of GD toxicity in human biological organisms are not conclusive. Yang's group (2013) performed bio-distribution and toxicology nanographene oxide analyses in mice, by oral and intraperitoneal administration. The researchers noted that the GDs were not adsorbed by the organs but were easily excreted. Despite this, they emphasize that toxicity will depend on the surface coating, size, and routes of administration.¹⁸³ Liu *et al.* (2012) conducted a study of size and dose on the biodistribution of GO in mice. The results showed that sizes between 1 and 5 μm accumulate in the lungs and between 110 and 500 nm in the liver. Therefore, they concluded that GO is not suitable for human use.^{184,185}

On the other hand, some research shows that GD effectively fights cancer cells and is safe for healthy cells. The administered dose influences the cytotoxicity and apoptosis of human cells; that is, the use of small doses ($<50 \text{ mg L}^{-1}$) of these materials proved to be safe.^{186–188} In addition, Manikandan *et al.* (2020) considered that PHB/graphene nanocomposites have a negligible cytotoxic effect.¹ Thus, the toxicity of GD in human cells still needs thorough research, as there are few publications on genotoxicity tests and lots of contradictory information.

One of the biggest challenges in this area is producing Gr, GO and rGO by greener processes and on an industrial scale to formulate polymeric nanocomposites that are increasingly less toxic and aggressive to the environment through greener and more sustainable routes, Fig. 12.

Final considerations

The development of novel packaging from polymeric nanocomposites based on graphene opens several possibilities for the food packaging industry and reduces production costs. It also brings lightness and higher resistance to these novel packaging resulting in numerous applications. The Gr, GO, and rGO application in nanocomposites significantly improves the packaging's thermal, mechanical, and barrier properties, reducing waste and increasing food safety. The more significant interest in using renewable polymers in the novel polymeric nanocomposites creation with the use of origin natural and biodegradable polymeric matrices and a smaller number of investigations using polymers is also noticeable of fossil origin, showing the concern with the environment. In general, studies are still lacking on the application of graphene nanocomposite packaging in wet foods and also on its toxicity and biodegradability, suggesting that further research in this regard should be carried out to ensure the safety and efficiency application of graphene and graphene-based in the formulation of new food packaging.

Author contributions

The authors confirm contribution to the paper as follows: conceptualization: Vinicius Rossa and Thiago M. Lima. investigation: Vinicius Rossa, Luanne E. Monteiro, Sancler C. Vasconcelos and Vinicius G. Madriaga. Data curation: Vinicius Rossa, Luanne E. Monteiro, Sancler da Costa Vasconcelos, Vinicius G. Madriaga, Eric Thomas Tai Shimabukuro, Anna Paula Carvalho, Vitor Francisco Ferreira and Thiago M. Lima. Writing – original draft preparation: Vinicius Rossa, Luanne E. Monteiro, Sancler C. Vasconcelos, Anna Paula Carvalho, Sibebe Berenice Castellã Pergher and Thiago M. Lima. Visualization: Thiago M. Lima, Vinicius Rossa, Vinicius G. Madriaga, Sancler C. Vasconcelos, Sibebe Berenice Castellã Pergher and Thiago M. Lima. Writing – review & editing: Thiago M. Lima, Vinicius Rossa, Sibebe Berenice Castellã Pergher, Fernando de Carvalho da Silva, Vitor Francisco Ferreira, Anna Paula Carvalho and Carlos Adam Conte Junior. Supervision: Thiago M. Lima and Vinicius Rossa.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 H. N. Lim, N. M. Huang and C. H. Loo, *J. Non-Cryst. Solids*, 2012, **358**, 525–530.
- 2 D. Han, L. Yan, W. Chen and W. Li, *Carbohydr. Polym.*, 2011, **83**, 653–658.
- 3 J. Jia, Y. Gai, W. Wang and Y. Zhao, *Ultrason. Sonochem.*, 2016, **32**, 300–306.
- 4 *Ecosustainable Polymer Nanomaterials for Food Packaging*, ed. C. Silvestre and S. Cimmino, CRC Press, 2013.
- 5 V. G. L. Souza and A. L. Fernando, *Food Packag. Shelf Life*, 2016, **8**, 63–70.
- 6 H. Bouwmeester, S. Dekkers, M. Y. Noordam, W. I. Hagens, A. S. Bulder, C. de Heer, S. E. C. G. ten Voorde, S. W. P. Wijnhoven, H. J. P. Marvin and A. J. A. M. Sips, *Regul. Toxicol. Pharmacol.*, 2009, **53**, 52–62.
- 7 R. Mahfouz, F. J. Cadete Santos Aires, A. Brenier, E. Ehret, M. Roumié, B. Nsouli, B. Jacquier and J. C. Bertolini, *J. Nanopart. Res.*, 2010, **12**, 3123–3136.
- 8 D. Cade, E. Ramus, M. Rinaudo, R. Auzély-Velty, T. Delair and T. Hamaide, *Biomacromolecules*, 2004, **5**, 922–927.
- 9 A. Haase, A. Mantion, P. Graf, J. Plendl, A. F. Thuenemann, W. Meier, A. Taubert and A. Luch, *Arch. Toxicol.*, 2012, **86**, 1089–1098.



- 10 P. R. Sivashankari and M. Prabakaran, in *Biopolymer-Based Composites*, Elsevier, 2017, pp. 381–397.
- 11 C. Botas, P. Álvarez, P. Blanco, M. Granda, C. Blanco, R. Santamaría, L. J. Romasanta, R. Verdejo, M. A. López-Manchado and R. Menéndez, *Carbon*, 2013, **65**, 156–164.
- 12 S. S. Nanda, G. C. Papaefthymiou and D. K. Yi, *Crit. Rev. Solid State Mater. Sci.*, 2015, **40**, 291–315.
- 13 H. Ahmad, M. Fan and D. Hui, *Composites, Part B*, 2018, **145**, 270–280.
- 14 X. Wang, M. Su, C. Liu, C. Shen and X. Liu, *J. Renewable Mater.*, 2020, **8**, 89–99.
- 15 M. J. Nine, M. A. Cole, D. N. H. Tran and D. Losic, *J. Mater. Chem. A*, 2015, **3**, 12580–12602.
- 16 T. Gatti, N. Vicentini, M. Mba and E. Menna, *Eur. J. Org. Chem.*, 2016, **2016**, 1071–1090.
- 17 S. Park and R. S. Ruoff, *Curr. Opin. Colloid Interface Sci.*, 2015, **20**, 322–328.
- 18 C. K. Chua and M. Pumera, *Chem. Soc. Rev.*, 2014, **43**, 291–312.
- 19 J. R. Potts, D. R. Dreyer, C. W. Bielawski and R. S. Ruoff, *Polymer*, 2011, **52**, 5–25.
- 20 S. Stankovich, D. A. Dikin, G. H. B. Dommett, K. M. Kohlhaas, E. J. Zimney, E. A. Stach, R. D. Piner, S. T. Nguyen and R. S. Ruoff, *Nature*, 2006, **442**, 282–286.
- 21 T. Sattar, *Top. Curr. Chem.*, 2019, **377**, 10.
- 22 S. S. Shams, L. S. Zhang, R. Hu, R. Zhang and J. Zhu, *Mater. Lett.*, 2015, **161**, 476–479.
- 23 X. Kong, Y. Zhu, H. Lei, C. Wang, Y. Zhao, E. Huo, X. Lin, Q. Zhang, M. Qian, W. Mateo, R. Zou, Z. Fang and R. Ruan, *Chem. Eng. J.*, 2020, **399**, 125808.
- 24 K. Kalaitzidou, H. Fukushima and L. T. Drzal, *Carbon*, 2007, **45**, 1446–1452.
- 25 A. F. Ghanem, A. M. Youssef and M. H. Abdel Rehim, *J. Mater. Sci.*, 2020, **55**, 4685–4700.
- 26 A. Al-Jabareen, H. Al-Bustami, H. Harel and G. Marom, *J. Appl. Polym. Sci.*, 2012.
- 27 H. M. Kim, J. K. Lee and H. S. Lee, *Thin Solid Films*, 2011, **519**, 7766–7771.
- 28 Y. Gao, O. T. Picot, W. Tu, E. Bilotti and T. Peijs, *J. Appl. Polym. Sci.*, 2018, **135**, 46041.
- 29 N. A. Manikandan, K. Pakshirajan and G. Pugazhenth, *Int. J. Biol. Macromol.*, 2020, **154**, 866–877.
- 30 T. G. Maraschin, R. da S. Correa, L. F. Rodrigues, N. M. Balzaretti, J. A. Malmonge, G. B. Galland and N. R. de S. Basso, *Mater. Res.*, 2019, 221–10.
- 31 S. Afshar and H. Baniyadi, *Int. J. Biol. Macromol.*, 2018, **109**, 1019–1028.
- 32 Q. Huang, M. Xu, R. Sun and X. Wang, *Ind. Crops Prod.*, 2016, **85**, 198–203.
- 33 A. Allahbakhsh, in *Food Packaging*, Elsevier, 2017, pp. 699–737.
- 34 T. B. Rouf and J. L. Kokini, *J. Mater. Sci.*, 2016, **51**, 9915–9945.
- 35 T. B. Rouf and J. L. Kokini, in *Bionanocomposites for Packaging Applications*, Springer International Publishing, Cham, 2018, pp. 149–177.
- 36 S. S. Sablani, *Polymer Nanocomposites for Food Packaging Applications*, in *Food Nanoscience and Nanotechnology, Food Engineering Series*, ed. H. Hernández-Sánchez and G. Gutiérrez-López, Springer, Cham, 2015, pp. 205–212.
- 37 *Graphene Properties, Preparation, Characterization and Applications*, N. C. Chhetri, S. Kuila and T. Murmu, Wiley-VCH Verlag GmbH & Co. KGaA, Published 2016 by Wiley-VCH Verlag GmbH & Co. KGaA., 1st edn, 2016, pp. 63–111.
- 38 M. Bera and P. K. Maji, *MOJ Polym. Sci.*, 2017, **1**, 94–97.
- 39 Z. Terzopoulou, G. Kyzas and D. Bikiaris, *Materials*, 2015, **8**, 652–683.
- 40 J. Byun, *J. Microbiol. Biotechnol.*, 2015, **25**, 145–151.
- 41 S. Cesur, C. Köroğlu and H. T. Yalçın, *J. Vinyl Addit. Technol.*, 2018, **24**, 376–387.
- 42 S. Tripathi, G. K. Mehrotra and P. K. Dutta, *e-Polymers*, 2008, **93**, 1–7.
- 43 V. Manigandan, R. Karthik, S. Ramachandran and S. Rajagopal, in *Biopolymers for Food Design*, Elsevier, 2018, pp. 469–491.
- 44 M. Aider, *LWT-Food Sci. Technol.*, 2010, **43**, 837–842.
- 45 S. Omid and A. Kakanejadifard, *RSC Adv.*, 2018, **8**, 12179–12189.
- 46 C. Cavalcanti and C. Chagas, *História Da Embalagem No Brasil*, 1st edn, 2006.
- 47 K. Marsh and B. Bugusu, *J. Food Sci.*, 2007, **72**, R39–R55.
- 48 A. Ojha, A. Sharma, M. Sihag and S. Ojha, *Agric. Rev.*, 2015, **36**, 241.
- 49 B. Page, in *Packaging Technology*, Elsevier, 2012, pp. 122–162.
- 50 D. Raheem, *Emir. J. Food Agric.*, 2013, **25**, 177.
- 51 A. L. Brody, B. Bugusu, J. H. Han, C. K. Sand and T. H. McHugh, *J. Food Sci.*, 2008, **73**, R107–R116.
- 52 L. McKeen, in *The Effect of Sterilization Methods on Plastics and Elastomers*, Elsevier, 2018, pp. 417–435.
- 53 B. Rodgers and W. Waddell, in *The Science and Technology of Rubber*, Elsevier, 2013, pp. 653–695.
- 54 A. Sharif and M. E. Hoque, in *Bio-based Polymers and Nanocomposites*, Springer International Publishing, Cham, 2019, pp. 1–28.
- 55 A. P. M. Landim, C. O. Bernardo, I. B. A. Martins, M. R. Francisco, M. B. Santos and N. R. de Melo, *Polímeros*, 2016, **26**, 82–92.
- 56 H. Moustafa, A. M. Youssef, N. A. Darwish and A. I. Abou-Kandil, *Composites, Part B*, 2019, **172**, 16–25.
- 57 A. Akelah, *Functionalized Polymeric Materials in Agriculture and the Food Industry*, Springer US, Boston, MA, 2013, vol. 9781461470.
- 58 F. Ciardelli, M. Bertoldo, S. Bronco and E. Passaglia, *Polymers from Fossil and Renewable Resources*, Springer International Publishing, Cham, 2019.
- 59 V. Siracusa, P. Rocculi, S. Romani and M. D. Rosa, *Trends Food Sci. Technol.*, 2008, **19**, 634–643.
- 60 F. A. MEDEIROS and H. WIEBECK, *Polim.: Cienc. Tecnol.*, 2013, **23**, 636–643.
- 61 D. Turan, G. Gunes and A. Kilic, in *Bionanocomposites for Packaging Applications*, Springer International Publishing, Cham, 2018, pp. 1–32.



- 62 D. W. Wei, H. Wei, A. C. Gauthier, J. Song, Y. Jin and H. Xiao, *J. Bioresour. Bioprod.*, 2020, **5**, 1–15.
- 63 L. Zhao, G. Duan, G. Zhang, H. Yang, S. Jiang and S. He, *Nanomaterials*, 2020, **10**, 150.
- 64 N. Kumar, P. Kaur and S. Bhatia, *Nutr. Food Sci.*, 2017, **47**, 591–606.
- 65 A. Morschbacker, *Polym. Rev.*, 2009, **49**, 79–84.
- 66 M. Akiyama, S. Sato, R. Takahashi, K. Inui and M. Yokota, *Appl. Catal., A*, 2009, **371**, 60–66.
- 67 J. Chaminand, L. auren. Djakovitch, P. Gallezot, P. Marion, C. Pinel and C. Rosier, *Green Chem.*, 2004, **6**, 359.
- 68 D. G. Lahr and B. H. Shanks, *Ind. Eng. Chem. Res.*, 2003, **42**, 5467–5472.
- 69 A.-Y. Yin, X.-Y. Guo, W.-L. Dai and K.-N. Fan, *Green Chem.*, 2009, **11**, 1514.
- 70 *Nanotechnologies in Food and Agriculture*, ed. M. Rai, C. Ribeiro, L. Mattoso and N. Duran, Springer International Publishing, Cham, 2015.
- 71 U. Bhardwaj, P. Dhar, A. Kumar and V. Katiyar, *Food Additives and Packaging*, 2014, ch. 19, pp. 275–314.
- 72 R. Sindhu, P. Binod and A. Pandey, in *Industrial Biorefineries & White Biotechnology*, Elsevier, 2015, pp. 575–605.
- 73 K. B. Rufato, J. P. Galdino, K. S. Ody, A. G. B. Pereira, E. Corradini, A. F. Martins, A. T. Paulino, A. R. Fajardo, F. A. Aouada, F. A. La Porta, A. F. Rubira and E. C. Muniz, in *Hydrogels - Smart Materials for Biomedical Applications*, IntechOpen, 2019.
- 74 S. Talegaonkar, H. Sharma, S. Pandey, P. K. Mishra and R. Wimmer, in *Food Packaging*, Elsevier, 2017, pp. 79–110.
- 75 H. Wang, J. Qian and F. Ding, *J. Agric. Food Chem.*, 2018, **66**, 395–413.
- 76 M. Sabzevari, D. E. Cree and L. D. Wilson, *ACS Omega*, 2018, **3**, 13045–13054.
- 77 U. Siripatrawan and W. Vitchayakitti, *Food Hydrocolloids*, 2016, **61**, 695–702.
- 78 Y. Peng, Y. Wu and Y. Li, *Int. J. Biol. Macromol.*, 2013, **59**, 282–289.
- 79 B. Kuswandi, *Nanotechnology in Food Packaging*, in *Nanoscience in Food and Agriculture 1. Sustainable Agriculture Reviews*, ed. S. Ranjan, N. Dasgupta and E. Lichtfouse, Springer, Cham, 2016, vol. 20, pp. 151–183.
- 80 Y. Huang, T. Wang, X. Zhao, X. Wang, L. Zhou, Y. Yang, F. Liao and Y. Ju, *J. Chem. Technol. Biotechnol.*, 2015, **90**, 1677–1684.
- 81 I. U. Unalan, C. Wan, L. Figiel, R. T. Olsson, S. Trabattini and S. Farris, *Nanotechnology*, 2015, **26**, 275703.
- 82 S. F. Abdellah Ali, *IOP Conf. Ser.: Mater. Sci. Eng.*, 2016, **137**, 012035.
- 83 S. Kashi, R. K. Gupta, N. Kao, S. A. Hadigheh and S. N. Bhattacharya, *J. Mater. Sci. Technol.*, 2018, **34**, 1026–1034.
- 84 A. P. A. de Carvalho and C. A. Conte Junior, *Trends Food Sci. Technol.*, 2020, **103**, 130–143.
- 85 J. Wang, X. Wang, C. Xu, M. Zhang and X. Shang, *Polym. Int.*, 2011, **60**, 816–822.
- 86 R. Zhang, Y. Wang, D. Ma, S. Ahmed, W. Qin and Y. Liu, *Ultrason. Sonochem.*, 2019, **59**, 104731.
- 87 M. A. Cerqueira, A. A. Vicente and L. M. Pastrana, in *Nanomaterials for Food Packaging*, Elsevier, 2018, pp. 1–11.
- 88 A. Usman, Z. Hussain, A. Riaz and A. N. Khan, *Carbohydr. Polym.*, 2016, **153**, 592–599.
- 89 Y. A. Arfat, J. Ahmed, M. Ejaz and M. Mullah, *Int. J. Biol. Macromol.*, 2018, **107**, 194–203.
- 90 A. K. Sundramoorthy, T. H. Vignesh Kumar and S. Gunasekaran, in *Graphene Bioelectronics*, Elsevier, 2018, pp. 267–306.
- 91 A. Barra, N. M. Ferreira, M. A. Martins, O. Lazar, A. Pantazi, A. A. Jderu, S. M. Neumayer, B. J. Rodriguez, M. Enăchescu, P. Ferreira and C. Nunes, *Compos. Sci. Technol.*, 2019, **173**, 53–60.
- 92 A. L. Rivera-Briso, F. L. Aachmann, V. Moreno-Manzano and Á. Serrano-Aroca, *Int. J. Biol. Macromol.*, 2020, **143**, 1000–1008.
- 93 W. Xu, W. Xie, X. Huang, X. Chen, N. Huang, X. Wang and J. Liu, *Food Chem.*, 2017, **221**, 267–277.
- 94 N. Pal, S. Banerjee, P. Roy and K. Pal, *Mater. Sci. Eng., C*, 2019, **104**, 109956.
- 95 D. Olteanu, A. Filip, C. Socaci, A. R. Biris, X. Filip, M. Coros, M. C. Rosu, F. Pogacean, C. Alb, I. Baldea, P. Bolfa and S. Pruneanu, *Colloids Surf., B*, 2015, **136**, 791–798.
- 96 J. Du and H.-M. Cheng, *Macromol. Chem. Phys.*, 2012, **213**, 1060–1077.
- 97 J. Jose, M. A. Al-Harhi, M. A.-A. AlMa'adeed, J. Bhadra Dakua and S. K. De, *J. Appl. Polym. Sci.*, 2015, **132**, 41827.
- 98 F. Li, H.-Y. Yu, Y.-Y. Wang, Y. Zhou, H. Zhang, J.-M. Yao, S. Y. H. Abdalkarim and K. C. Tam, *J. Agric. Food Chem.*, 2019, **67**, 10954–10967.
- 99 E. C. Romani, D. G. Larrude, L. Nachez, C. Vilani, J. B. de Campos, S. B. Peripolli and F. L. Freire, *Tribol. Lett.*, 2017, **65**, 96.
- 100 A. P. Kauling, A. T. Seefeldt, D. P. Pisoni, R. C. Pradeep, R. Bentini, R. V. B. Oliveira, K. S. Novoselov and A. H. Castro Neto, *Adv. Mater.*, 2018, **30**, 1803784.
- 101 J. Wang, F. Ma, W. Liang and M. Sun, *Mater. Today Phys.*, 2017, **2**, 6–34.
- 102 Y. Zhu, S. Murali, W. Cai, X. Li, J. W. Suk, J. R. Potts and R. S. Ruoff, *Adv. Mater.*, 2010, **22**, 3906–3924.
- 103 Z. U. Khan, A. Kausar, H. Ullah, A. Badshah and W. U. Khan, *J. Plast. Film Sheeting*, 2016, **32**, 336–379.
- 104 G. Venugopal, K. Krishnamoorthy, R. Mohan and S.-J. Kim, *Mater. Chem. Phys.*, 2012, **132**, 29–33.
- 105 Y. Yao, X. Chen, J. Zhu, B. Zeng, Z. Wu and X. Li, *Nanoscale Res. Lett.*, 2012, **7**, 363.
- 106 M. Jin, H.-K. Jeong, W. J. Yu, D. J. Bae, B. R. Kang and Y. H. Lee, *J. Phys. D. Appl. Phys.*, 2009, **42**, 135109.
- 107 A. Haque, M. A.-A. Mamun, M. F. N. Taufique, P. Karnati and K. Ghosh, *IEEE Trans. Semicond. Manuf.*, 2018, **31**, 535–544.
- 108 L. Staudenmaier, *Eur. J. Inorg. Chem.*, 1898, **31**, 1481–1487.
- 109 B. C. Brodie, *Proc. R. Soc. London*, 1860, **10**, 11–12.
- 110 W. S. Hummers and R. E. Offeman, *J. Am. Chem. Soc.*, 1958, **80**, 1339.
- 111 D. Li, M. B. Müller, S. Gilje, R. B. Kaner and G. G. Wallace, *Nat. Nanotechnol.*, 2008, **3**, 101–105.



- 112 J. Chen, B. Yao, C. Li and G. Shi, *Carbon*, 2013, **64**, 225–229.
- 113 D. R. Dreyer, S. Park, C. W. Bielawski and R. S. Ruoff, *Chem. Soc. Rev.*, 2010, **39**, 228–240.
- 114 N. E. Sorokina, M. A. Khaskov, V. V. Avdeev and I. V. Nikol'skaya, *Russ. J. Gen. Chem.*, 2005, **75**, 162–168.
- 115 D. C. Marcano, D. V. Kosynkin, J. M. Berlin, A. Sinitskii, Z. Sun, A. Slesarev, L. B. Alemany, W. Lu and J. M. Tour, *ACS Nano*, 2010, **4**, 4806–4814.
- 116 M. J. McAllister, J. Li, D. H. Adamson, H. C. Schniepp, A. a Abdala, J. Liu, M. Herrera-Alonso, D. L. Milius, R. Car, R. K. Prud'homme and I. a Aksay, *Chem. Mater.*, 2007, **19**, 4396–4404.
- 117 A. Bagri, C. Mattevi, M. Acik, Y. J. Chabal, M. Chhowalla and V. B. Shenoy, *Nat. Chem.*, 2010, **2**, 581–587.
- 118 J. George, Y. Patel, S. M. Pillai and P. Munshi, *J. Mol. Catal. A: Chem.*, 2009, **304**, 1–7.
- 119 J. Li and T. Wang, *J. Chem. Thermodyn.*, 2011, **43**, 731–736.
- 120 C. K. Rofer-DePoorter, *Chem. Rev.*, 1981, **81**, 447–474.
- 121 A. P. Amrute, K. Jeske, Z. Łodziana, G. Prieto and F. Schüth, *Chem. Mater.*, 2020, **32**, 4369–4374.
- 122 J. Zhang, H. Yang, G. Shen, P. Cheng, J. Zhang and S. Guo, *Chem. Commun.*, 2010, **46**, 1112–1114.
- 123 J. Gao, F. Liu, Y. Liu, N. Ma, Z. Wang and X. Zhang, *Chem. Mater.*, 2010, **22**, 2213–2218.
- 124 I. Uysal Unalan, G. Cerri, E. Marcuzzo, C. A. Cozzolino and S. Farris, *RSC Adv.*, 2014, **4**, 29393–29428.
- 125 R. Miraftab and H. Xiao, *J. Bioresour. Bioprod.*, 2019, **4**, 200–201.
- 126 H. Qian, J. Wang and L. Yan, *J. Bioresour. Bioprod.*, 2020, **5**, 204–210.
- 127 X. Ji, Y. Xu, W. Zhang, L. Cui and J. Liu, *Composites, Part A*, 2016, **87**, 29–45.
- 128 Y. Joo, V. Agarkar, S. H. Sung, B. M. Savoie and B. W. Boudouris, *Science*, 2018, **359**, 1391–1395.
- 129 X. Yang, Y. Tu, L. Li, S. Shang and X. Tao, *ACS Appl. Mater. Interfaces*, 2010, **2**, 1707–1713.
- 130 A. M. Pinto, J. Cabral, D. A. P. Tanaka, A. M. Mendes and F. D. Magalhães, *Polym. Int.*, 2013, **62**, 33–40.
- 131 X. Feng, X. Wang, W. Xing, B. Yu, L. Song and Y. Hu, *Ind. Eng. Chem. Res.*, 2013, **52**, 12906–12914.
- 132 L.-L. Wu, J. Wang, X. He, T. Zhang and H. Sun, *Packag. Technol. Sci.*, 2014, **27**, 693–700.
- 133 R. A. Hurley, A. Ouzts, J. Fischer and T. Gomes, *Packag. Technol. Sci.*, 2013, **26**, 399–412.
- 134 P. Tzeng, B. Stevens, I. Devlaming and J. C. Grunlan, *Langmuir*, 2015, **31**, 5919–5927.
- 135 K. Goh, J. K. Heising, Y. Yuan, H. E. Karahan, L. Wei, S. Zhai, J.-X. Koh, N. M. Htin, F. Zhang, R. Wang, A. G. Fane, M. Dekker, F. Dehghani and Y. Chen, *ACS Appl. Mater. Interfaces*, 2016, **8**, 9994–10004.
- 136 C. Demitri, V. M. De Benedictis, M. Madaghiele, C. E. Corcione and A. Maffezzoli, *Measurement*, 2016, **90**, 418–423.
- 137 C. D. Grande, J. Mangadlao, J. Fan, A. De Leon, J. Delgado-Ospina, J. G. Rojas, D. F. Rodrigues and R. Advincula, *Macromol. Symp.*, 2017, **374**, 1600114.
- 138 R. H. F. Faradilla, G. Lee, J. Roberts, P. Martens, M. Stenzel and J. Arcot, *Cellulose*, 2018, **25**, 399–416.
- 139 X. Ge, H. Li, L. Wu, P. Li, X. Mu and Y. Jiang, *J. Appl. Polym. Sci.*, 2017, **134**, 44910.
- 140 S. Montes, A. Etxeberria, V. Mocholi, A. Rekondo, H. Grande and J. Labidi, *eXPRESS Polym. Lett.*, 2018, **12**, 543–555.
- 141 S. Torres-Giner, Y. Echegoyen, R. Teruel-Juanes, J. Badia, A. Ribes-Greus and J. Lagaron, *Nanomaterials*, 2018, **8**, 745.
- 142 A. Bher, I. Uysal Unalan, R. Auras, M. Rubino and C. Schvezov, *Polymers*, 2018, **10**, 95.
- 143 M. Dhanasekar, V. Jenefer, R. B. Nambiar, S. G. Babu, S. P. Selvam, B. Neppolian and S. V. Bhat, *Mater. Res. Bull.*, 2018, **97**, 238–243.
- 144 P. Wang, S. Tang, F. Sheng, J. Cai, P. Fei, A. Nawaz, N. Walayat, A. B. Javaid and H. Xiong, *Int. J. Biol. Macromol.*, 2019, **132**, 1208–1220.
- 145 Z. Wu, Y. Huang, L. Xiao, D. Lin, Y. Yang, H. Wang, Y. Yang, D. Wu, H. Chen, Q. Zhang, W. Qin and S. Pu, *Int. J. Biol. Macromol.*, 2019, **123**, 569–575.
- 146 S. Chowdhury, Y. L. Teoh, K. M. Ong, N. S. Rafflisman Zaidi and S.-K. Mah, *Food Packag. Shelf Life*, 2020, **24**, 100463.
- 147 M. Moritz and M. Geszke-Moritz, *Chem. Eng. J.*, 2013, **228**, 596–613.
- 148 Q. Yu, Z. Wu and H. Chen, *Acta Biomater.*, 2015, **16**, 1–13.
- 149 A. M. Pinto, I. C. Gonçalves and F. D. Magalhães, *Colloids Surf., B*, 2013, **111**, 188–202.
- 150 G. Wu and P. Xu, in *Multifunctional Nanocomposites for Energy and Environmental Applications*, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, Germany, 2018, pp. 203–230.
- 151 X. Ren, H. Guo, J. Feng, P. Si, L. Zhang and L. Ci, *Chemosphere*, 2018, **191**, 389–399.
- 152 S. C. Smith and D. F. Rodrigues, *Carbon*, 2015, **91**, 122–143.
- 153 K. Tadyszak, J. Wychowanec and J. Litowczenko, *Nanomaterials*, 2018, **8**, 944.
- 154 S. Asha, A. N. Ananth, S. P. Jose and M. A. J. Rajan, *Appl. Nanosci.*, 2018, **8**, 395–405.
- 155 Q. Zhang, Z. Wu, N. Li, Y. Pu, B. Wang, T. Zhang and J. Tao, *Mater. Sci. Eng., C*, 2017, **77**, 1363–1375.
- 156 S. Goenka, V. Sant and S. Sant, *J. Control. Release*, 2014, **173**, 75–88.
- 157 X. Zhu, A. F. Radovic-Moreno, J. Wu, R. Langer and J. Shi, *Nano Today*, 2014, **9**, 478–498.
- 158 G. Franci, A. Falanga, S. Galdiero, L. Palomba, M. Rai, G. Morelli and M. Galdiero, *Molecules*, 2015, **20**, 8856–8874.
- 159 I. Sengupta, P. Bhattacharya, M. Talukdar, S. Neogi, S. K. Pal and S. Chakraborty, *Colloid Interface Sci. Commun.*, 2019, **28**, 60–68.
- 160 S. Aliamradni, V. Abolmaali and S. S. Borandeh, *J. Nanostruct.*, 2019, **3**, 402–413.
- 161 P. Kumar, P. Huo, R. Zhang and B. Liu, *Nanomaterials*, 2019, **9**, 737.
- 162 S. Gurunathan, J. Woong Han, A. Abdal Daye, V. Eppakayala and J. Kim, *Int. J. Nanomed.*, 2012, 5901.
- 163 S. Romero-Vargas Castrillón, F. Perreault, A. F. de Faria and M. Elimelech, *Environ. Sci. Technol. Lett.*, 2015, **2**, 112–117.



- 164 Y. Tu, M. Lv, P. Xiu, T. Huynh, M. Zhang, M. Castelli, Z. Liu, Q. Huang, C. Fan, H. Fang and R. Zhou, *Nat. Nanotechnol.*, 2013, **8**, 594–601.
- 165 P. Huo, C. Liu, D. Wu, J. Guan, J. Li, H. Wang, Q. Tang, X. Li, Y. Yan and S. Yuan, *J. Ind. Eng. Chem.*, 2018, **57**, 125–133.
- 166 L. T. Trinh, L. A. B. Quynh and N. H. Hieu, *Int. J. Nanotechnol.*, 2018, **15**, 108.
- 167 Q. Tu, Q. Zhang, Y. Wang, Y. Jiao, J. Xiao, T. Peng and J. Wang, *Prog. Org. Coatings*, 2019, **129**, 247–253.
- 168 Z. Yang, X. Hao, S. Chen, Z. Ma, W. Wang, C. Wang, L. Yue, H. Sun, Q. Shao, V. Murugadoss and Z. Guo, *J. Colloid Interface Sci.*, 2019, **533**, 13–23.
- 169 Y.-Y. Xie, X.-H. Hu, Y.-W. Zhang, F. Wahid, L.-Q. Chu, S.-R. Jia and C. Zhong, *Carbohydr. Polym.*, 2020, **229**, 115456.
- 170 M. A. Ashraf, Y. Yang and A. Fakhri, *Ceram. Int.*, 2020, **46**, 8379–8384.
- 171 S. Tan, X. Wu, Y. Xing, S. Lilak, M. Wu and J. X. Zhao, *Colloids Surf., B*, 2020, **185**, 110616.
- 172 N. M. Dat, P. N. B. Long, D. C. U. Nhi, N. N. Minh, L. M. Duy, L. N. Quan, H. M. Nam, M. T. Phong and N. H. Hieu, *Synth. Met.*, 2020, **260**, 116260.
- 173 R. K. Matharu, T. A. Tabish, T. Trakoolwilaian, J. Mansfield, J. Moger, T. Wu, C. Lourenço, B. Chen, L. Ciric, I. P. Parkin and M. Edirisinghe, *J. Colloid Interface Sci.*, 2020, **571**, 239–252.
- 174 Y. Zhang, H. Ruan, C. Guo, J. Liao, J. Shen and C. Gao, *Sep. Purif. Technol.*, 2020, **234**, 116017.
- 175 L. Zhang, P. Chen, Y. Xu, W. Nie and Y. Zhou, *Appl. Catal., B*, 2020, **265**, 118572.
- 176 A. Hashmi, A. K. Singh, B. Jain and S. A. C. Carabineiro, *Nanomaterials*, 2020, **10**, 105.
- 177 G. Jena, B. Anandkumar, S. C. Vanithakumari, R. P. George, J. Philip and G. Amarendra, *Prog. Org. Coatings*, 2020, **139**, 105444.
- 178 T. Vi, S. Kumar, J.-H. Pang, Y.-K. Liu, D. Chen and S. Lue, *Nanomaterials*, 2020, **10**, 366.
- 179 H. Velichkova, S. Kotsilkov, E. Ivanov, R. Kotsilkova, S. Gyoshev, N. Stoimenov and N. K. Vitanov, *Food Addit. Contam., Part A: Chem., Anal., Control, Exposure Risk Assess.*, 2017, **34**, 1072–1085.
- 180 H. Velichkova, I. Petrova, S. Kotsilkov, E. Ivanov, N. K. Vitanov and R. Kotsilkova, *J. Appl. Polym. Sci.*, 2017, **134**, 1–12.
- 181 H. C. Oyeoka, C. M. Ewulonu, I. C. Nwuzor, C. M. Obele and J. T. Nwabanne, *J. Bioresour. Bioprod.*, 2021, **6**, 168–185.
- 182 L. De Marchi, C. Pretti, B. Gabriel, P. A. A. P. Marques, R. Freitas and V. Neto, *Sci. Total Environ.*, 2018, **631–632**, 1440–1456.
- 183 K. Yang, H. Gong, X. Shi, J. Wan, Y. Zhang and Z. Liu, *Biomaterials*, 2013, **34**, 2787–2795.
- 184 X. Guo and N. Mei, *J. Food Drug Anal.*, 2014, **22**, 105–115.
- 185 J. H. Liu, S. T. Yang, H. Wang, Y. Chang, A. Cao and Y. Liu, *Nanomedicine*, 2012, **7**, 1801–1812.
- 186 L. Q. Chen, P. P. Hu, L. Zhang, S. Z. Huang, L. F. Luo and C. Z. Huang, *Sci. China Chem.*, 2012, **55**, 2209–2216.
- 187 Z. Ding, Z. Zhang, H. Ma and Y. Chen, *ACS Appl. Mater. Interfaces*, 2014, **6**, 19797–19807.
- 188 Z. Singh, *Nanotechnol. Sci. Appl.*, 2016, **9**, 15–28.

