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Chiral fluorescent sensor based on H₈-BINOL for the high enantioselective recognition of D- and L-phenylalanine†

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Phenylalanine an essential aromatic amino acid for humans and animals, cannot be synthesized by humans and animals on their own. However, it synthesizes important neurotransmitters and hormones in the body and is involved in gluconeogenesis and lipid metabolism. Moreover, the two opposite configurations of phenylalanine have different activities. For example, L-phenylalanine is a biologically active optical isomer involved in crucial biological processes, the lack of which will lead to intellectual disability, while D-phenylalanine only acts as a chiral intermediate. In this research, an H₈-BINOL chiral fluorescent sensor modified with 1,2,3-triazole was synthesized in high yield (95%) by nucleophilic substitution and click reaction. The chiral fluorescent sensor showed high enantioselectivity toward phenylalanine. L-Phenylalanine enhanced the fluorescence response of the probe significantly, while D-phenylalanine had no obvious fluorescence response change. The enantioselective fluorescence enhancement ratio [ef = (I_L - I₀)/(I_D - I₀), where I₀ is the fluorescence of the sensor without amino acids] for the highest fluorescence intensity at 20.0 equivalents of amino acids was 104.48. In this way, the probe could be used to identify and differentiate different configurations of phenylalanine.

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1 Introduction

Chirality is a universal phenomenon and an important feature of nature, and is widely present in the generation of life and various evolutionary mechanisms.^{1–3} As most of the drugs people use in real life are chiral drugs, with further understanding and exploration of chiral issues, it is recognized that the enantiomers of some chiral compounds differ in their physiological effects and metabolic processes. Different isomers of the chiral drugs may exhibit different effects in physiological processes, so much that one enantiomer is effective in treating diseases while the other is harmful or fatal.^{4–6} Amino acids, which are essential for the maintenance of life, are also a class of chiral molecules.^{7–10} L-Phenylalanine is a physiologically active aromatic amino acid and is one of the essential amino acids that cannot be synthesized naturally by humans and animals.¹¹ In addition to L-phenylalanine, a variety of commercially valuable derivatives can be produced through the L-phenylalanine biosynthetic pathway. L-Phenylalanine deficiency leads to a number of disorders such as phenylketonuria (PKU), one of the most common inherited disorders of amino acid metabolism that occurs due to the destruction of

phenylalanine dehydrogenase (PAH)¹² and manifests specifically with symptoms such as intellectual disability and irreversible brain damage.^{13–16} Studies have shown that L-phenylalanine can also be used as a carrier of anti-cancer drugs, introducing drug molecules directly to the nidus and the effect is three to five times that of other amino acids.^{17,18} However, D-phenylalanine is largely absent in human metabolism due to its specific structure and activity. It is also used as a very significant chiral intermediate in organic synthesis, new drug development, and the synthesis of peptide compounds.^{19,20} Therefore, it is very important to distinguish the enantiomers and detect the optical purity.

There are a number of techniques for detecting the enantiomer composition of chiral compounds. The traditional identification techniques consist of electrochemistry, ligand exchange, high-throughput screening, and spectroscopy, such as high-performance liquid chromatography (HPLC)^{21,22} and circular dichroism (CD)²³ are commonly used. Colorimetry and fluorescent probes are excellent approaches to identify enantiomers on account of their high-pitched visual resolution, high sensitivity, ease of operation, convenient equipment, real-time imaging, and online monitoring.^{24–26} It has been used to differentiate the conformation of chiral molecules and determine enantiomeric composition and has led to numerous high quality and quantitative results in the enantioselective fluorescence identification of amino acids, carboxylic acids, and amines.

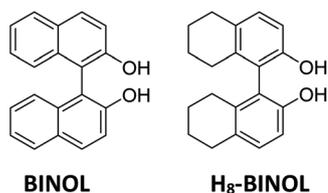
BINOL is a rigid structure, which is a very suitable substrate for fluorescent sensors.^{27–29} For example, Vivek Panyam

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Muralidharan has designed and synthesized chiral fluorescent BINOL bearing benzoisoquino-line-1,3-dione on the minor grooves. It was found to be a sensor for the identification of *R*-amino alcohols and to determine the enantiomeric composition of α -amino alcohols.²⁷ BINOL is also easily modified and has multiple modification sites that can modulate its fluorescence by introducing various moieties at different positions. Because of the unique chirality, BINOL and its derivatives can also provide interesting fluorescence signals and good chiral recognition. In the past few years, BINOL and its derivatives have gained global visibility in the field of enantioselective fluorescent sensor, which was used in the recognition of chiral molecules.^{30–34} H₈-BINOL is the 5,5',6,6' site reduction of BINOL. Similar to BINOL, due to the electron giving action of the alkyl ring and the sp³ hybridization of the carbon, the inductive effect causes the electron density of the benzene ring of H₈-BINOL to increase and higher reaction activity. Most previous reports on H₈-BINOL focused on asymmetric catalysis.^{35–40} However, few reports paid attention to the enantioselective fluorescent sensors to date.⁴¹ In 2011, Pu *et al.* reported H₈-BINOL-diamine compounds that displayed a good enantioselective fluorescent response toward mandelic acid, and the enantioselective fluorescence enhancement ratio was up to 3.5.⁴² Therefore, a novel H₈-BINOL-based triazole-modified fluorescent probe was designed and synthesized to investigate the enantioselective recognition of phenylalanine in this study.



2 Experimental

2.1 Materials and methods

All experimental reagents used in the experiments were purchased from Energy Chemicals and Shanghai Aladdin and were not sonically purified. All analytical grade solvents were distilled prior to use. The solvents used for the probe synthesis experiments were purchased from Innochem. ¹H NMR and ¹³C NMR were measured on a Bruker AM-400WB spectrometer using CDCl₃. Fluorescence spectra data were measured on a Hitachi F-7100 fluorescence spectrophotometer with slit widths of 2.5 nm and 2.5 nm, respectively ($\lambda_{\text{ex}} = 260$ nm). Melting points were carried out using an X-4 melting point tester. The optical rotation was measured on a Rudolph AUTOPOL IV automatic polarimeter. Elemental analyses were performed on a Vario EL/MACRO elemental analyzer using polyethylene as a blank control.

2.1.1 Synthesis of propargyl derivatives a and b. *R*-5,5',6,6',7,7',8,8'-Octahydrobinol (1.00 g, 3.40 mmol) and K₂CO₃ (1.08 g, 7.81 mmol) were put in a 100 mL single-necked flask and 30 mL of acetone was added to dissolve them. Then, 3-bromopropyne (0.73 mL, 8.48 mmol) was added slowly

dropwise to the reaction system. After the mixture was stirred for 20 min, the reaction was heated to about 55 °C for refluxing and stirred overnight. The reaction was intercepted, while thin layer chromatography detection proved the vanishing point of the start material and a new blot was generated. The temperature of the system was then reduced to ambient temperature, and the reaction solution was recovered and filtered, washed three times with acetone to collect the light yellow liquid, and dried with anhydrous MgSO₄. Then, the crude product was obtained by rotary concentration on a rotary evaporator and later separated by column chromatography (silica gel 200–300 mesh, eluting solvent petroleum ether: ethyl acetate = 15 : 1, v/v) to get 0.48 g (derivatives **a**) of white solid in 38% yield, together with 0.45 g (derivatives **b**) of a white solid with a yield of 40%. [derivatives **a**, M.p. 97–99 °C. $[\alpha]_{\text{D}}^{25} +0.50$ (c 0.2, CH₃OH). ¹H NMR (400 MHz, chloroform-*d*) δ 7.05 (d, *J* = 8.5 Hz, 1H), 6.90 (dd, *J* = 17.7, 8.4 Hz, 2H), 6.70 (s, 1H), 4.51 (t, *J* = 2.2 Hz, 2H), 4.31 (s, 1H), 2.67 (q, *J* = 8.7, 7.5 Hz, 4H), 2.36–2.11 (m, 3H), 2.10–1.83 (m, 2H), 1.80–1.40 (m, 10H)]. Elemental analysis results: C (experimental value: 83.01%, calculated value: 83.10%), H (experimental value: 7.09%, calculated value: 7.28%), O (experimental value: 9.90%, calculated value: 9.63%). [derivatives **b**, M.p. 86–87 °C. $[\alpha]_{\text{D}}^{25} +0.35$ (c 0.2, CH₃OH). ¹H NMR (400 MHz, chloroform-*d*) δ 7.09 (d, *J* = 8.4 Hz, 2H), 6.94 (d, *J* = 8.4 Hz, 2H), 4.56 (d, *J* = 2.4 Hz, 4H), 2.79 (td, *J* = 6.4, 2.8 Hz, 4H), 2.39 (t, *J* = 2.3 Hz, 2H), 2.08 (q, *J* = 5.8, 5.0 Hz, 1H), 1.79–1.66 (m, 6H), 1.58 (s, 1H)]. Elemental analysis results: C (experimental value: 84.20%, calculated value: 84.29%), H (experimental value: 6.99%, calculated value: 7.07%), O (experimental value: 8.81%, calculated value: 8.64%).

2.1.2 Synthesis of *R*-1 probe. *R*-2-Alkynylmethoxy-5,5',6,6',7,7',8,8'-octahydrobinol (200 mg, 0.81 mmol) was put in a 100 mL aubergine flask, evacuated several times, and protected by argon, and then 6 mL of tetrahydrofuran was added and stirred thoroughly to dissolve completely. Then 2-azidoacetate (0.09 mL, 0.97 mmol) was added to the mixture. Sodium ascorbate (129 mg, 1.70 mmol) and copper sulfate pentahydrate (337 mg, 0.81 mmol) were then accurately weighed, dissolved in 5 mL of water after a few minutes, and appended to the system. When the reaction was completed, it was monitored by thin layer chromatography and quenched by adding 15 mL of ice water to the reaction flask. It was then extracted three times with dichloromethane, collected, washed with prepared saturated brine, dried with anhydrous sodium sulphate for 30 min, and separated by column chromatography (silica gel 200–300 mesh, eluting solvent CH₃OH : CH₂Cl₂ = 1 : 2, v/v). A white solid of 0.26 g was obtained with a yield of 95%. M.p. 93–95 °C. $[\alpha]_{\text{D}}^{25} +0.75$ (c 0.2, CH₃OH). Elemental analysis results: C (experimental value: 69.54%, calculated value: 69.79%), H (experimental value: 6.29%, calculated value: 6.49%), N (experimental value: 9.31%, calculated value: 9.40%), O (experimental value: 14.96%, calculated value: 14.32%). ¹H NMR (400 MHz, chloroform-*d*) δ 7.24 (s, 1H), 7.09 (d, *J* = 8.4 Hz, 1H), 7.01 (d, *J* = 8.2 Hz, 1H), 6.93 (d, *J* = 8.4 Hz, 1H), 6.77 (d, *J* = 8.2 Hz, 1H), 5.48–4.92 (m, 4H), 3.79 (s, 3H), 3.11–2.70 (m, 4H), 2.36–2.06 (m, 4H), 1.85–1.43 (m, 11H), 1.26 (td, *J* = 7.1, 2.2 Hz, 2H). ¹³C NMR (101 MHz, chloroform-*d*) δ 153.26, 149.73, 145.06, 137.58,



135.84, 130.94, 129.77, 128.76, 123.23, 122.89, 111.76, 111.05, 62.45, 52.49, 50.20, 28.83, 26.81, 26.57, 22.64, 22.44 ppm. HRMS (ESI⁻): calcd for [C₂₆H₂₉N₃O₄⁺H⁺]⁻ 448.2158; found 448.1774.

2.1.3 Preparation of fluorescent probe solutions. 44.7 mg of probes *R-1*, 44.7 mg of probes *S-1*, or 60.0 mg of probe *R-2* were put in three 10 mL volumetric flasks, each; then, CH₃OH (chromatographic grade) was added to dissolve and fix the volume to the scale line, was shaken well to dissolve completely and mixed well to obtain the required concentration of 0.001 mol L⁻¹ of the master solution. Then, the above mixture was diluted with the same grade of methanol solution to the 2 × 10⁻⁵ mol L⁻¹ test solution for testing (need to be ready to use).

2.1.4 Preparation of solutions required for the test. Common amino acids (D/L-alanine, D/L-aspartic acid, D/L-glutamine, D/L-glutamic acid, D/L-leucine, D/L-lysine, D/L-methionine, D/L-phenylalanine, D/L-proline, D/L-serine, D/L-valine, D/L-arginine, D/L-aspartic acid, D/L-tyrosine, D/L-cysteine, D/L-glycine) accurately weighed in a 5 mL volumetric flask, and then, 2000 μL of deionized water was pipetted dropwise to obtain a solution with a concentration of 0.1 mol L⁻¹.

3 Results and discussion

3.1 Synthesis procedure

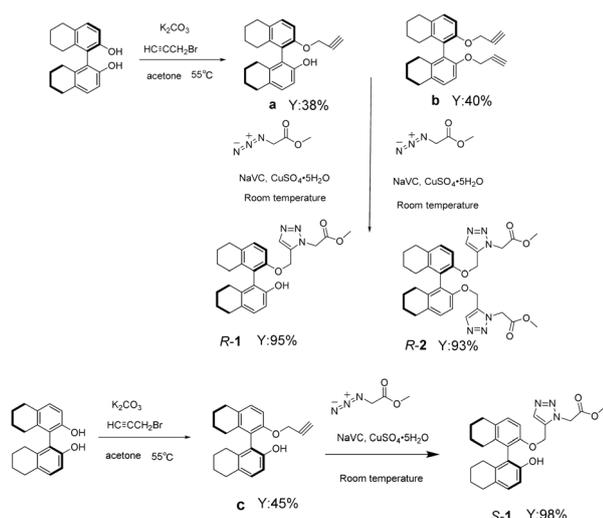
The synthesis process of sensors *R-1*, *R-2* and *S-1* is outlined in Scheme 1. *R-5,5',6,6',7,7',8,8'*-Octahydrobiol (H₈-BINOL) was used as the starting material, the fluorescent sensors *R-1* and *R-2* based on H₈-BINOL derivatives modified with 2- and 2,2'-triazoles were obtained by a click reaction. According to the previous literature,⁴³ *R*-H₈-BINOL and 3-bromopropyne were synthesized as propargyl derivatives **a** and **b**. Propargyl derivative **a** and methyl azidoacetate were dissolved in tetrahydrofuran at room temperature and reacted with anhydrous copper sulphate and sodium ascorbate to afford 1,2,3-triazole modified H₈-BINOL derivative *R-1*, which was further purified by column chromatography in a high yield of 95%. Similar probes *R-2* and

S-1 were synthesized by the same synthetic route with high yields (93% and 98%). The specific structures of the experimentally synthesized compounds were all validated by ¹H NMR, ¹³C NMR, and ESI-MS.⁴⁴

3.2 Fluorescence study

The fluorescence responses of the *R-1* probe toward two enantiomers of sixteen kinds of common amino acids without the involvement of metal ions were explored first. In this experiment, various amino acid (0.1 M) test solutions were prepared in distilled-deionized water or DMSO. Methanol was used to dissolve the *R-1* probe (2.0 × 10⁻⁵ M). A pair of enantiomers (20.0 equivalents) of each amino acid was mixed into the solution at room temperature for fluorescence measurements. As shown in Fig. 1, L-phenylalanine and D-aspartic acid tremendously increased the fluorescence response of the *R-1* probe, with *I*/*I*₀ values of 2.04 and 1.43, respectively. However, D-phenylalanine and L-aspartic acid, as well as other amino acids, had little effect on the probes, with almost no fluorescence change. Thus, the novel chiral fluorescence *R-1* exhibited a high degree of enantioselectivity and chemoselectivity for phenylalanine and aspartic acid.

The fluorescence responses of the *R-1* probe toward D-phenylalanine and L-phenylalanine were then studied in detail. The fluorescence spectra of the *R-1* probe after the treatment with 20 equivalents of D-phenylalanine and L-phenylalanine are provided in Fig. 2a. For these measurements, the *R-1* probe was blended in methanol (2.0 × 10⁻⁵ M) with either L-phenylalanine or D-phenylalanine (20 equivalents). The fluorescence spectrum of the *R-1* probe at L-phenylalanine spiked from 5 to 100 equivalents, as given in Fig. 2c. As revealed in Fig. 2c, when the concentration of L-phenylalanine was raised to 20 equivalents, the fluorescence strength of the *R-1* probe at 315 nm rose to 2.04 times its original value. However, when D-phenylalanine was used, the fluorescence of the *R-1* probe did not change significantly over this concentration range (as shown in Fig. 2b). At 20.0 equivalents of amino acids, the maximum enantioselective fluorescence enhancement ratio⁴⁵ [ef = (*I*_L - *I*₀)/(*I*_D - *I*₀), where *I*₀ is the fluorescence of the sensor in the absence of amino acid]



Scheme 1 Synthetic procedure of fluorescent sensors *R-1*, *R-2*, and *S-1*.

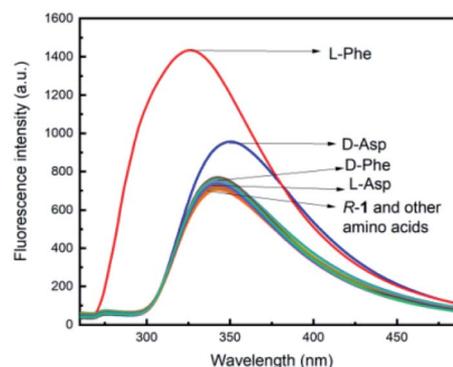


Fig. 1 Fluorescence spectra of *R-1* (2.0 × 10⁻⁵ M in CH₃OH) with different enantiomers of sixteen ordinary amino acids (20.0 equivalents) in the absence of metal ions (λ_{ex} = 260 nm, slits = 2.5/2.5 nm).



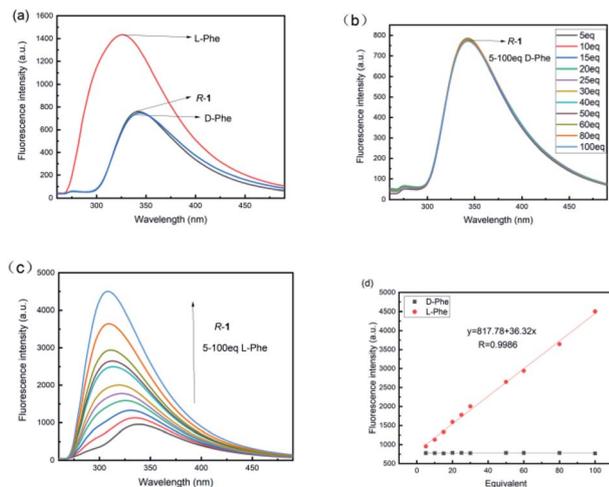


Fig. 2 (A) Fluorescence spectra of *R*-1 (2.0×10^{-5} M in CH_3OH) toward *D*- and *L*-phenylalanine (20.0 equiv.). (b) Fluorescence titration of *R*-1 with *D*-phenylalanine in CH_3OH . (c) Fluorescence titration of *R*-1 with *L*-phenylalanine in CH_3OH . (d) Fluorescence intensities at $\lambda = 315$ nm versus the equivalents of phenylalanine ($\lambda_{\text{ex}} = 260$ nm, slits = 2.5/2.5 nm).

was 104.28. The fluorescence intensity of *D*-phenylalanine remained relatively constant with increasing equivalents (5–100 equivalents), while the fluorescence of *L*-phenylalanine then gradually increased and showed a linear relationship $R = 0.9986$, as shown in Fig. 2d.

To investigate the enantiomeric composition of amino acids, the fluorescence response of *R*-1 with phenylalanine at different ee [ee, ⁴⁶ enantiomeric excess, = $(l - d)/(l + d)$] values were also studied. In the experiment, the *R*-1 probe was mixed with phenylalanine at different ee values and its fluorescence responses were then measured (Fig. 3). It was possible to measure the enantiomeric composition of amino acids when using *R*-1 with an excess of *L*-phenylalanine in methanol solution. To demonstrate the enantiomeric composition of amino

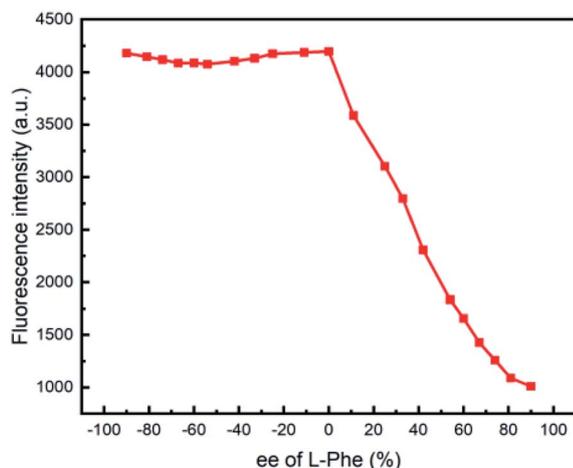


Fig. 3 Fluorescence response at $\lambda_{\text{em}} = 315$ nm for the mixtures of *R*-1 toward *L*-phenylalanine (100.0 equiv.) at various ee values.

acids, the fluorescence response of *R*-1 could be significantly enhanced by *L*-phenylalanine. As the excess of *L*-phenylalanine increased, the fluorescence intensity progressively enhanced.

The large non-linear plot is shown in Fig. 3. It also demonstrated that the conformation of the amino acid did not improve the fluorescence, which could greatly curtail the fluorescence enhancement by its enantiomer. The most simple-minded approach to describe this phenomenon was that the two configurations of amino acids may bind to each other and slowly form a structure between them.⁴⁷ This structure prevented them from binding to the probe for reaction. A surplus of the enantiomeric amino acid of the configuration that enhanced fluorescence reacted with the probe and turned on the fluorescence of the sensor, inducing it to enhance fluorescence.

The fluorescence reaction of the *R*-1 enantiomer *S*-1 toward *D*- and *L*-phenylalanine was also studied under the same conditions. As displayed in Fig. 4-a, the fluorescence responses of *S*-1 toward the phenylalanine enantiomer were identical to that of the *R*-1 probe, both recognizing only *L*-phenylalanine. We also investigated the fluorescence response of *R*-2, the double substitution product of *R*-1, toward *D*- and *L*-phenylalanine, as displayed in Fig. 4b. As shown, the fluorescence responses of *R*-2 to the phenylalanine enantiomer were identical to those of the *R*-1 probe. All three probes exhibited that *L*-phenylalanine could display a significant fluorescence enhancement, while *D*-

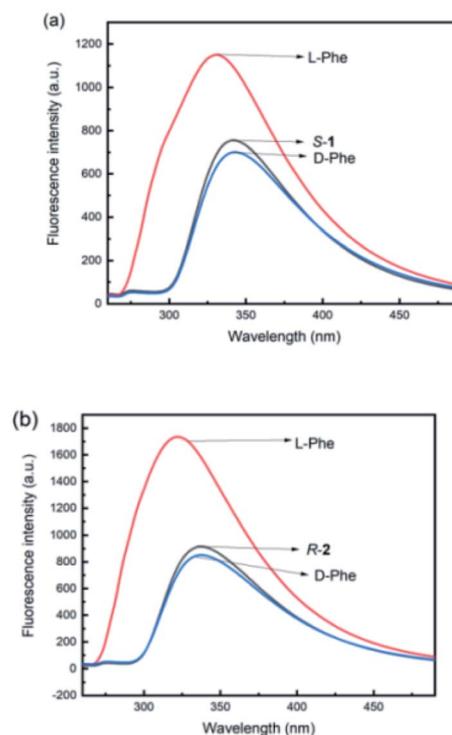


Fig. 4 (a) Fluorescence spectra of *R*-2 (2.0×10^{-5} M in CH_3OH) toward *D*- and *L*-phenylalanine (20.0 equiv.) in the absence of metal ions. (b) Fluorescence spectra of *S*-1 (2.0×10^{-5} M in CH_3OH) toward *D*- and *L*-phenylalanine (20.0 equiv.) in the absence of metal ions ($\lambda_{\text{ex}} = 260$ nm. Excitation/emission slit widths: 2.5/2.5 nm).



phenylalanine exhibited no obvious change. Hence, it was clear that the recognition of phenylalanine was independent of the configuration of the probes and the degree of triazole modification. Further study was focused on the exact mechanism of the recognition.

4 Conclusions

In summary, a novel fluorescent sensor of 1,2,3-triazole-modified H₈-BINOL derivative with high correspondence selectivity was synthesized by nucleophilic substitution and click reaction with a high yield. It could be used as a novel chiral fluorescence sensor to identify and differentiate different configurations of phenylalanine as it exhibited high enantioselectivity and sensitivity to L-phenylalanine in the fluorescence experiments. The maximum enantioselective fluorescence enhancement ratio was 104.48 at 20.0 amino acid equivalents. The results showed that the novel synthesized sensor had the great possibility to be used as a fluorescent sensor to identify the different configurations of phenylalanine as well as to determine the enantiomeric composition of amino acids. Moreover, in contrast to previously reported amino acid fluorescent probes, the newly synthesized sensor also has the advantage of convenient use in observing fluorescence enhancement, because when many previously reported amino acid fluorescent probes are used in observing fluorescence enhancement, additional metals have to be used.

Conflicts of interest

There are no conflicts to declare.

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