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Development of an MRI contrast agent for both detection and inhibition of the amyloid- β fibrillation process[†]

 Rohmad Yudi Utomo,^a Satoshi Okada,^{ID *abc} Akira Sumiyoshi,^d Ichio Aoki^d and Hiroyuki Nakamura^{ID *ab}

A curcumin derivative conjugated with Gd-DO3A (Gd-DO3A-Comp.B) was synthesised as an MRI contrast agent for detecting the amyloid- β (A β) fibrillation process. Gd-DO3A-Comp.B inhibited A β aggregation significantly and detected the fibril growth at 20 μ M of A β with 10 μ M of probe concentration by T_1 -weighted MR imaging.

A significant increase of Alzheimer's disease (AD) patients urges the development of therapeutic and diagnostic technology.¹ As with the therapeutic development, diagnostic technology also faces several obstacles. To date, the definite diagnosis of AD relies on the histopathological data of post-mortem.^{2,3} The non-invasive imaging technology targeting AD biomarkers such as amyloid β (A β) could provide phenotypical diagnostics, although the development of A β probes still remains challenging. Several contrast agents for single photon emission computed tomography (SPECT) and positron emission tomography (PET) such as Flortetapir-¹⁸F and Pittsburgh compound B ([¹¹C]PiB) were developed as efficient tracers in mild cognitive impairment patients.^{4,5} However, PET- and SPECT-based diagnostics require injection of radioactive probes, which cannot be measured frequently due to radiation exposure and limited availability of facilities. They also provide limited information on the anatomic profile of biomarkers due to their low spatial resolution and imprecise microscopic localization.⁶ In contrast, magnetic resonance imaging (MRI) contrast agents could quantify the A β accumulation in the anatomic brain image.⁷

Several reported MRI contrast agents using gadolinium (Gd) complexes demonstrate potential use of A β detection. A clinically approved contrast agent, Gd(III) diethylenetriaminepenta-acetic acid (Gd-DTPA) complex accumulates in brain after opening the blood-brain barrier (BBB) by using mannitol and

detects A β deposits in the mice AD-model.⁸ To improve the selectivity, Gd complexes were conjugated with compounds binding to A β such as Pittsburgh compound B (Gd-DO3A-PiB) which also serves as an approach for increasing MRI sensitivity.^{9,10} An α, β -unsaturated ketone compound curcumin has been widely reported as an A β probe due to its ability to bind the hydrophobic site of A β .^{11,12} Allen *et al.* firstly reported the direct conjugation of curcumin with Gd-DTPA which binds to A β with four times higher relaxivity than free Gd-DTPA.¹³ Furthermore, a polymeric acid-based nanoparticle covalently linked with curcumin and Gd-DOTA could also detect A β in human brain specimen by MRI.¹⁴ These previous studies demonstrate that the curcumin structure has significant potential for the development of MRI contrast agents for AD diagnosis.

Previously, we reported a curcumin derivative, compound B, possesses 100-times stronger inhibitory activity of A β aggregation than curcumin on the basis of thioflavin T (ThT) competitive binding assay.^{15,16} According to this result, we designed curcumin-based Gd probes for the detection and inhibition of

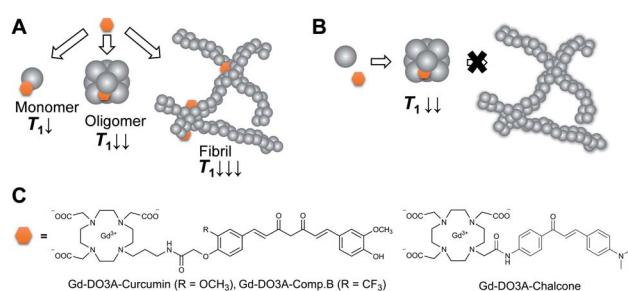


Fig. 1 (A) A probe concept that produces T_1 in a dependent manner of A β fibrillation process. (B) Inhibitor-based probes that cause moderate T_1 decreases due to inhibitory activity of fibrillation. (C) The chemical structures of the synthesized Gd probes for A β detection and inhibition.

^aSchool of Life Science and Technology, Tokyo Institute of Technology, 4259 Nagatsuta, Midori, Yokohama, Kanagawa, 226-8503, Japan

^bLaboratory for Chemistry and Life Science, Institute of Innovative Research, Tokyo Institute of Technology, 4259 Nagatsuta, Midori, Yokohama, Kanagawa 226-8503, Japan. E-mail: sokada@res.titech.ac.jp; hiro@res.titech.ac.jp

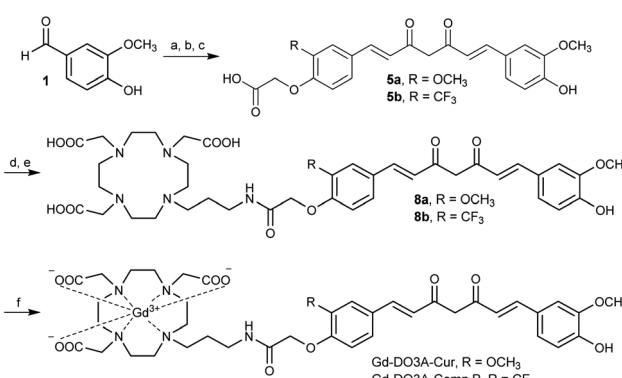
^cJST, PRESTO, 4259 Nagatsuta, Midori, Yokohama, Kanagawa 226-8503, Japan

^dInstitute for Quantum Medical Science, National Institutes for Quantum Science and Technology, 4-9-1 Anagawa, Inage, Chiba 263-8555, Japan

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$\text{A}\beta$ (Fig. 1A–C). We hypothesized that these probes could accelerate proton longitudinal relaxation depending on the fibrillation stage of $\text{A}\beta$, because molecular tumbling rate of the Gd complexes becomes slower (Fig. 1A).¹⁷ As a result, the probes permit the detection of $\text{A}\beta$ by longitudinal relaxation time (T_1)-weighted imaging. This mechanism could also be utilized to estimate the inhibitory activity of the probes by T_1 -based analysis (Fig. 1B). The curcumin and compound B were directly conjugated with the macrocyclic DO3A ligand through the propylamine linker to obtain Gd-DO3A-Cur and Gd-DO3A-Comp.B, respectively (Fig. 1C).

Gd-DO3A-Cur and Gd-DO3A-Comp.B were synthesized according to Scheme 1 (detail in Scheme S1, ESI†). The compound **5a** and **5b**, which have asymmetric curcumin derivatives containing carboxylic acid group, were synthesized by three step reactions. Amide bond formation with DO3A(*t*Bu)₃-propylamine ligand¹⁸ by condensation reaction afforded compound **7a** and **7b**. The *tert*-butyl groups were deprotected by trifluoroacetic acid producing compound **8a** and **8b**. The complexation was performed with GdCl₃·6H₂O by adjusting the reaction pH to 7, giving 43 and 41% yields of Gd-DO3A-Cur and Gd-DO3A-Comp.B, respectively. The T_1 relaxivities (r_1) of the curcumin-based Gd probes were estimated by T_1 measurement using a 1 tesla NMR relaxometry (Fig. S1, ESI†). For the comparison, we synthesized Gd-DO3A-Chal which is a reported probe for $\text{A}\beta$.¹⁹ The r_1 of Gd-DO3A-Comp.B, Gd-DO3A-Cur, and Gd-DO3A-Chal were 7.1, 6.1 and 5.3 mM⁻¹ s⁻¹, respectively. These r_1 values are higher than that of clinically approved Gd-DOTA (3.9 mM⁻¹ s⁻¹).²⁰ The molecular weight of Gd-DO3A-Comp.B and Gd-DO3A-Cur is almost two times larger than that of Gd-DOTA. Because the r_1 increases approximately linearly with molecular weight in low magnetic field,¹⁷ the high r_1 values of Gd-DO3A-Comp.B and Gd-DO3A-Cur might be mainly attributed to their high rotational correlation time, rather than the high number of coordinated water molecules. The r_1 of Gd-DO3A-Chal was comparable to the value reported previously.¹⁹



Scheme 1 Synthetic scheme of Gd-DO3A-Cur and Gd-DO3A-Comp.B. (a) $\text{B}(\text{OH})_3$, morpholine, DMF, 100 °C, 10 min. (b) **3a/3b**, $\text{B}(\text{OH})_3$, morpholine, DMF, 100 °C, 10 min. (c) TFA, DCM. (d) DO3A(*t*Bu)₃-propylamine ligand, PyBOP, HOBT, Et₃N, DMF. (e) **7a/7b**, TFA, DCM. (f) $\text{GdCl}_3 \cdot 6\text{H}_2\text{O}$, NaOH, H_2O .

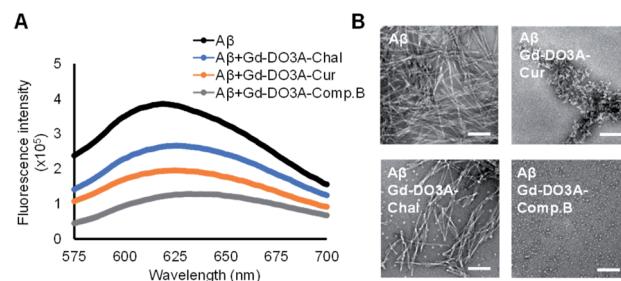


Fig. 2 Inhibitory effect of the Gd probes toward $\text{A}\beta$ aggregation measured by Congo red assay (A) and negative staining TEM images (B). The Gd probes were co-incubated with monomeric $\text{A}\beta$ for 24 h in PBS at pH 7.4. [Gd] = 10 μM , $[\text{A}\beta]$ = 20 μM . Scale bars = 100 nm.

We evaluated the inhibitory effect of three probes toward $\text{A}\beta$ aggregation by Congo red assay.²¹ After 24 h incubation of 20 μM $\text{A}\beta$ with 10 μM probe, Gd-DO3A-Comp.B showed the lowest fluorescence intensity, indicating the strongest inhibitory activity followed by Gd-DO3A-Cur (Fig. 2A). As the comparison, the reported MRI agents, Gd-DO3A-Chal showed slight inhibitory activity. The inhibitory effect was further evaluated by transmission electron microscopy (TEM) with negative staining (Fig. 2B). In the absence of the probes, $\text{A}\beta$ formed huge and massive fibril similar to the typical morphology of $\text{A}\beta$ fibril.²² The TEM images of $\text{A}\beta$ with Gd-DO3A-Comp.B showed the presence of white spheres below 10 nm, demonstrating that Gd-DO3A-Comp.B strongly inhibits $\text{A}\beta$ aggregation. In fact, the fibril growth stopped at a stage of oligomer formation. Lower inhibitory activity of Gd-DO3A-Cur was also found to provide a shortened worm-like fibril, which is the typical morphology of $\text{A}\beta$ exposed to curcumin.²³ In contrast, the small amount of white spheres and partial fibril disruption were found in the image of $\text{A}\beta$ with Gd-DO3A-Chal. In comparison with a reported Gd-DTPA-curcumin possessing inhibitory activity starting at 50 μM , Gd-DO3A-Comp.B possessed stronger inhibition of $\text{A}\beta$ aggregation at 10 μM .²⁴ The MTT assay using Neuro 2a cells showed that IC_{50} of Gd-DO3A-Cur and Gd-DO3A-Comp.B. were more than 500 μM , indicating that these compounds did not possess significant cytotoxicity (Fig. S2, ESI†).

To detect fibrillation process by NMR relaxometry, we measured T_1 of the probe mixture with $\text{A}\beta$ which were pre-

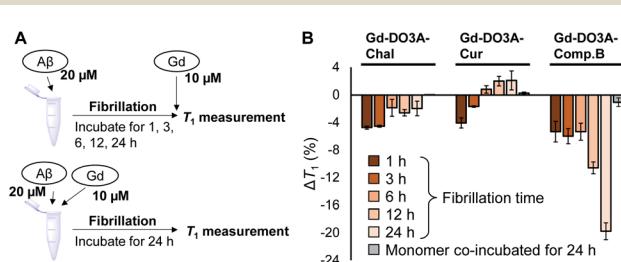


Fig. 3 (A) Experimental design of T_1 -based detection of $\text{A}\beta$ fibrillation and inhibition by using the Gd probes. (B) T_1 changes of the Gd probe solutions with pre-incubated fibrils and monomers in PBS at pH 7.4 (mean \pm SEM, n = 3). [Gd] = 10 μM , $[\text{A}\beta]$ = 20 μM .



incubated for 1, 3, 6, 12, and 24 h to make it form the fibrils of different growth stages (Fig. 3A and B). The T_1 of Gd-DO3A-Comp.B solution decreased with pre-incubation time of A β , demonstrating that the Gd-DO3A-Comp.B can detect A β fibril depending on the growth stage (Fig. 3B). Lower T_1 involved with A β growth could be caused by the reduction in tumbling rate of the Gd complex.²⁵ We also co-incubated the probes with the A β monomer and monitored T_1 changes over the incubation time (Fig. 3A, B and S3, ESI \dagger). Interestingly, the Gd-DO3A-Comp.B did not cause significant T_1 decreases even after 24 hours co-incubation with A β monomers, demonstrating that Gd-DO3A-Comp.B has a strong inhibitory effect on fibril formation and the inhibition can be monitored by T_1 measurement (Fig. 3B). The inhibitory effect was consistent with the results of Congo red assay and TEM (Fig. 2). On the other hand, the time-dependent increases of T_1 were observed in Gd-DO3A-Chal and Gd-DO3A-Cur. This might be because these two probes were buried in the hydrophobic pocket as A β fibril grew up and fewer water molecules permitted access to the Gd ions. It is also possible that these probes have lower binding affinity, especially for matured fibril, and require higher concentrations to produce significant T_1 changes.²⁶ These probe did not produce the significant ΔT_1 between monomer and fibril samples (Fig. 3B and S3, ESI \dagger), although they showed little inhibition in Congo red assay and TEM (Fig. 2).

The feasibility of the Gd probes was further evaluated by *in vitro* MRI measurement using a 1 tesla scanner. The T_1 -weighted images showed that Gd-DO3A-Comp.B produced slight T_1 signal increases with A β monomers for 2 and 24 h (Fig. 4A and B). More significant signal increases were observed in the Gd-DO3A-Comp.B with A β fibril pre-incubated for 24 h (Fig. 4C). In contrast, Gd-DO3A-Chal and Gd-DO3A-Cur did not show significant signal changes in the presence of A β monomers or fibrils (Fig. 4A–C). These results were mostly consistent with the T_1 profile measured by NMR (Fig. 3). Compared to the previously reported Gd-DO3A-Chal that required 100 μ M of the probe concentration to detect the equimolar A β ,¹⁹ Gd-DO3A-Comp.B could detect five-times lower concentration of A β (20 μ M) with ten-times lower probe concentration (10 μ M). Therefore, Gd-DO3A-Comp.B could be promising to further develop highly sensitive diagnostic MRI contrast agents of AD.

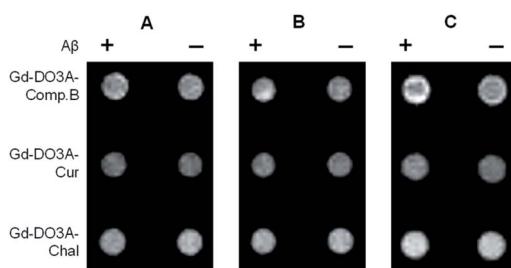


Fig. 4 T_1 -weighted images of the Gd probe solutions in the presence of monomeric A β at 2 h incubation (A), monomeric A β at 24 h incubation (B), and A β fibrils pre-incubated for 24 h (C). Incubation was conducted in PBS at pH 7.4.

In conclusion, we synthesized the curcumin-based Gd probes which enabled the detection and inhibition of A β fibril formation. Gd-DO3A-Comp.B allowed for the highly sensitive detection of A β fibril by the T_1 measurement. Moreover, the inhibitory activity could be estimated by T_1 measurement, because Gd-DO3A-Comp.B decreased T_1 depending on the growth stage of A β fibril formation. Such unique modality would be useful not only for the diagnostics but also for the direct evaluation of the therapeutic efficacy *in vivo*. For the future application, it would be important to combine with BBB penetration methods targeting the brain such as transient opening of the BBB using focused ultrasound or mannitol injection.^{27,28}

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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Notes and references

- 1 N. L. Batsch and M. S. Mittelman, *World Alzheimer Report 2012: Overcoming the stigma of dementia*, London, 2012.
- 2 G. McKhann, D. Drachman, M. Folstein, R. Katzman, D. Price and E. M. Stadlan, *Neurology*, 1984, **34**, 939.
- 3 B. Dubois, H. H. Feldman, C. Jacova, S. T. DeKosky, P. Barberger-Gateau, J. Cummings, A. Delacourte, D. Galasko, S. Gauthier, G. Jicha, K. Meguro, J. O'Brien, F. Pasquier, P. Robert, M. Rossor, S. Salloway, Y. Stern, P. J. Visser and P. Scheltens, *Lancet Neurol.*, 2007, **6**, 734–746.
- 4 A. Forsberg, H. Engler, O. Almkvist, G. Blomquist, G. Hagman, A. Wall, A. Ringheim, B. Långström and A. Nordberg, *Neurobiol. Aging*, 2008, **29**, 1456–1465.
- 5 A. Okello, J. Koivunen, P. Edison, H. A. Archer, F. E. Turkheimer, K. Någren, R. Bullock, Z. Walker, A. Kennedy, N. C. Fox, M. N. Rossor, J. O. Rinne and D. J. Brooks, *Neurology*, 2009, **73**, 754–760.
- 6 D. E. Huddleston and S. A. Small, *Nat. Clin. Pract. Neurol.*, 2005, **1**, 96–105.
- 7 A. C. Sedgwick, J. T. Brewster, P. Harvey, D. A. Iovan, G. Smith, X.-P. He, H. Tian, J. L. Sessler and T. D. James, *Chem. Soc. Rev.*, 2020, **49**, 2886–2915.
- 8 Y. Z. Wadghiri, E. M. Sigurdsson, M. Sadowski, J. I. Elliott, Y. Li, H. Scholtzova, C. Y. Tang, G. Aguinaldo, M. Pappolla, K. Duff, T. Wisniewski and D. H. Turnbull, *Magn. Reson. Med.*, 2003, **50**, 293–302.
- 9 A. F. Martins, J.-F. Morfin, A. Kubíčková, V. Kubíček, F. Buron, F. Suzenet, M. Salerno, A. N. Lazar,



C. Duyckaerts, N. Arlicot, D. Guilloteau, C. F. G. C. Geraldes and É. Tóth, *ACS Med. Chem. Lett.*, 2013, **4**, 436–440.

10 G. Bort, S. Catoen, H. Borderies, A. Kebsi, S. Ballet, G. Louin, M. Port and C. Ferroud, *Eur. J. Med. Chem.*, 2014, **87**, 843–861.

11 P. P. N. Rao, T. Mohamed, K. Teckwani and G. Tin, *Chem. Biol. Drug Des.*, 2015, **86**, 813–820.

12 M. García-Alloza, L. A. Borrelli, A. Rozkalne, B. T. Hyman and B. J. Bacska, *J. Neurochem.*, 2007, **102**, 1095–1104.

13 S. M. Vithanarachchi and M. J. Allen, *Chem. Commun.*, 2013, **49**, 4148–4150.

14 R. Patil, P. R. Gangalum, S. Wagner, J. Portilla-Arias, H. Ding, A. Rekechenetskiy, B. Konda, S. Inoue, K. L. Black, J. Y. Ljubimova and E. Holler, *Macromol. Biosci.*, 2015, **15**, 1212–1217.

15 R. Y. Utomo, Y. Asawa, S. Okada, H. S. Ban, A. Yoshimori, J. Bajorath and H. Nakamura, *Bioorg. Med. Chem.*, 2021, **46**, 116357.

16 D. Yanagisawa, N. Shirai, T. Amatsubo, H. Taguchi, K. Hirao, M. Urushitani, S. Morikawa, T. Inubushi, M. Kato, F. Kato, K. Morino, H. Kimura, I. Nakano, C. Yoshida, T. Okada, M. Sano, Y. Wada, K. Wada, A. Yamamoto and I. Tooyama, *Biomaterials*, 2010, **31**, 4179–4185.

17 J. Wahsner, E. M. Gale, A. Rodríguez-Rodríguez and P. Caravan, *Chem. Rev.*, 2019, **119**, 957–1057.

18 E. Pazos, M. Goličnik, J. L. Mascareñas and M. E. Vázquez, *Chem. Commun.*, 2012, **48**, 9534–9536.

19 G. Choi, H.-K. Kim, A. R. Baek, S. Kim, M. J. Kim, M. Kim, A. E. Cho, G.-H. Lee, H. Jung, J. Yang, T. Lee and Y. Chang, *J. Ind. Eng. Chem.*, 2020, **83**, 214–223.

20 Y. Shen, F. L. Goerner, C. Snyder, J. N. Morelli, D. Hao, D. Hu, X. Li and V. M. Runge, *Invest. Radiol.*, 2015, **50**, 330–338.

21 S. A. Hudson, H. Ecroyd, T. W. Kee and J. A. Carver, *FEBS J.*, 2009, **276**, 5960–5972.

22 Y. Xiao, I. Matsuda, M. Inoue, T. Sasahara, M. Hoshi and Y. Ishii, *J. Biol. Chem.*, 2020, **295**, 458–467.

23 A. Thapa, S. D. Jett and E. Y. Chi, *ACS Chem. Neurosci.*, 2016, **7**, 56–68.

24 A. Kochi, H. Lee, S. Vithanarachchi, V. Padmini, M. Allen and M. Lim, *Curr. Alzheimer Res.*, 2015, **12**, 415–423.

25 R. B. Lauffer, *Chem. Rev.*, 1987, **87**, 901–927.

26 S. Majdoub, Z. Garda, A. C. Oliveira, I. Relich, A. Pallier, S. Lacerda, C. Hureau, C. F. G. C. Geraldès, J.-F. Morfin and É. Tóth, *Chem.-Eur. J.*, 2021, **27**, 2009–2020.

27 M. Kinoshita, N. McDannold, F. A. Jolesz and K. Hynynen, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**, 11719–11723.

28 J. Yang, Y. Zaim Wadghiri, D. Minh Hoang, W. Tsui, Y. Sun, E. Chung, Y. Li, A. Wang, M. de Leon and T. Wisniewski, *NeuroImage*, 2011, **55**, 1600–1609.

