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Three-component assembly of stabilized fluorescent isoindoles†

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The tandem addition of an amine and a thiol to an aromatic dialdehyde engages a selective three-component assembly of a fluorescent isoindole. While an attractive approach for diversity-based fluorophore discovery, isoindoles are typically unstable and present considerable challenges for their practical utility. We found that introduction of electron-withdrawing substituents into the dialdehyde component affords stable isoindole products in one step with acceptable yields and high purity.

Since the preparation of the first isoindole (**1**, Fig. 1) in 1951 (ref. 1) and the isolation of the parent unsubstituted isoindole (**2**) in 1972,² the relative instability of this heterocyclic ring system has been an important impediment to the discovery of new chemical transformations and biological applications. The position of the equilibrium between the two tautomeric forms **2** and **3**, respectively *2H*- and *1H*-isoindoles, could be invoked to assess the stability of the *2H*-form. It has been found that substituents on the isoindole ring system play a key role. For example, it appears that electron-donating groups, such as methyl groups in **4**, destabilize the *2H*-isoindole,³ whereas electron acceptors in **5**⁴ and **6**⁵ improve stability.

Multicomponent reactions^{6,7} that enable three or more discrete molecules to combine into one product not only curtail synthetic operations but also advance the complexity viable within a single operation. To date, many of our fluorescent probes are prepared by two-component processes wherein moiety A is attached to moiety B to generate a fluorescent probe. Conventionally, this is achieved by adding dye A to a biological molecule B, however methods have been established that generate the probe motif as part of the coupling process.^{8,9} The latter, referred to as turn-on labeling, advantageously removes potential non-fluorescent impurities as only the desired product displays the proper fluorescence. While rare, advance of multicomponent turn-on labeling strategies offers a robust ability to improve the selectivity of labeling as well as to further expand the diversity possible within a labeling reaction. Here, we turn our attention to explore a three-component strategy to prepare isoindoles with improved stability.

An early report of the formation of fluorescent species when *o*-diacetylbenzene was exposed to proteins¹⁰ led to the discovery of a multicomponent reaction between *o*-phthalaldehyde (**7**), amines **8** and thiols **9**, which yields highly fluorescent isoindole **10** (Fig. 2).^{11–16} This reaction now forms the basis for the quantitative determination of amino acids and is used in commercial amino acid analyzers.¹⁷ The method is characterized by high sensitivity, although the lack of mechanistic understanding has led researchers to optimize the conditions primarily empirically.

An important drawback of the method is the lack of stability of the product isoindoles **10** (Fig. 2), which, once formed, undergo further conversions introducing inaccuracies due to unstable fluorescence. In general, the highest fluorescence must be achieved within 5–25 min and remain time-independent for another 20–30 min, the conditions which are hard to fulfill as the rate of isoindole formation and its stability depend on an individual amino acid.^{18,19} Naphthalene-1,2-dicarboxaldehyde (**11**) was recommended as an alternative reagent with claims that the product isoindoles **12** would be more stable, but the proposal seemingly has not received acceptance from the scientific community as the increases in stability are probably not significant to justify the cost of this reagent.¹⁷

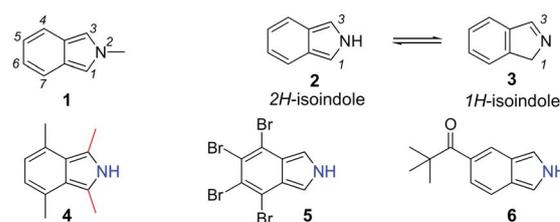


Fig. 1 *N*-methylisoindole (**1**) and a selection of the first isoindoles prepared **4–6** along with a depiction of the substituent-based tautomeric preferences.

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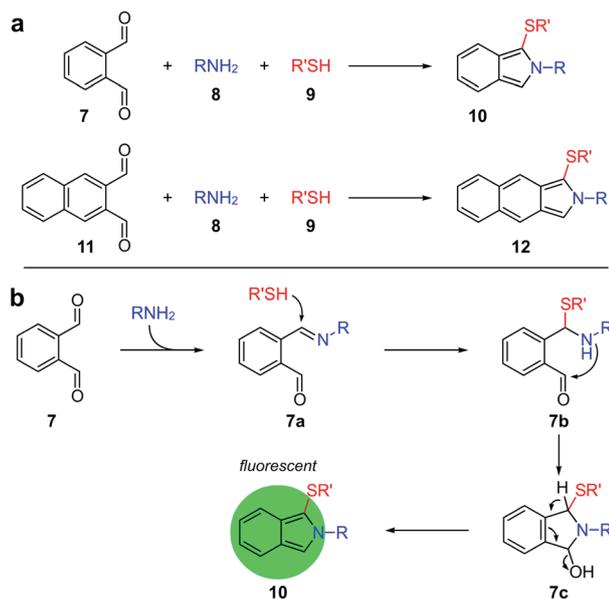


Fig. 2 The three-component isoindole reaction. (a) Reaction of phthalaldehyde (7) or 2,3-naphthalenedicarboxaldehyde (11) with amines (8) and thiols (9) to afford fluorescent isoindoles 10 and 12, respectively. (b) Proposed mechanism of the reaction with 7 where the initial formation of imine 7a is followed by an attack by a thiol 9 to form acetal-like species 7b. This undergoes cyclization to give hemiaminal 7c with a subsequent elimination of water to result in isoindole 10.

Our initial attempt to characterize the product of the three-component reaction between phthalaldehyde 7, amines and thiols (Fig. 3) met with significant synthetic challenges, as evident by the low stability of 10a. This compound is unstable at room temperature and rapidly decomposed when column chromatography purification was attempted. In order to obtain an analytical sample, 10a was repeatedly recrystallized from cold CH₃CN, dried in vacuum at 0 °C and immediately analyzed by NMR. Isoindole 12a, derived from 2,3-naphthalenedicarboxaldehyde (11), turned out to be only slightly more stable in comparison to product 10a and also rapidly decomposed when column chromatography purification was attempted.

Electron-deficient dialdehydes 13, 15, 17 and 19 (Fig. 3) on the other hand, all gave stable isoindoles 14a, 16a (structure assigned by NOESY analyses, see ESI[†]), 18a and 20a respectively (Fig. 3), when reacted with butyl amine and protected cysteine. These isoindoles could be purified by column chromatography and were considerably easier to handle.

Next, we turned our attention to evaluate if *n*-butyl- and *n*-octylphthalimidic phthalaldehydes 17 and 19 produced readily isolated products from a variety of aliphatic or aromatic amines and thiols. As shown in Fig. 4, isoindoles 18b–d and 20b–f can be obtained by adding a thiol and amine to the solution of 17 or 19 in CH₃CN at 0 °C. Subsequent simple removal of the volatiles on the rotary evaporator and purification of the product, facilitated by its fluorescence on TLC, gave the desired 18b–d and 20b–f in a straightforward manner. These reactions could be conducted with a 1 : 1 : 1 ratio of dialdehyde, amine, and thiol

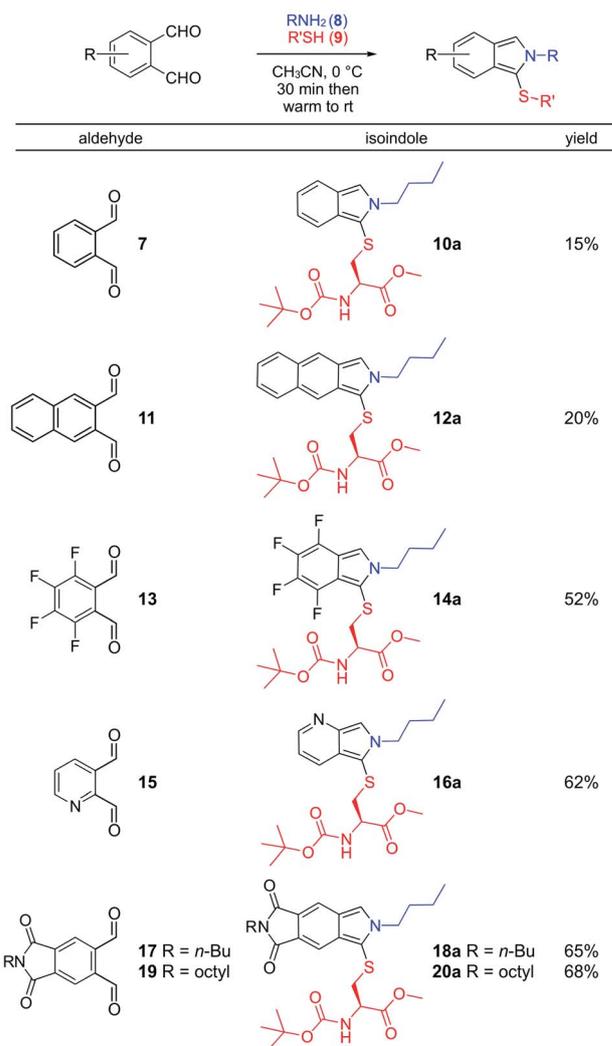


Fig. 3 One-pot synthesis of fluorescent 1-thio-2H-isoindoles and related structures from aromatic dialdehydes, butylamine and *N*-(*tert*-butoxycarbonyl)-L-cysteine methyl ester (Boc-Cys-OMe).

in acceptable isolated yields (48–66%, Fig. 4). The yields did not seem to be affected by the electronic properties of the reacting amines or thiols and were similar for reactions involving electron-rich (18b, c, 20c–f) vs. electron-deficient (18d, 20b) amines or electron-rich (18c, 20d) vs. electron-deficient (18b, d, 20b, c, e, f) thiols.

While an attractive analytical tool and fluorophore, practical applications of isoindoles suffer due to their rapid degradation. Proposed early on by Simons and Johnson,²⁰ the problem arises from nucleophilic attack by ROH (water or alcohols) at C1 (Fig. 5) resulting in the loss of the thiol and formation of the corresponding γ -lactam 21b.²¹ This is further complicated by the potential for self-dimerization by a Diels–Alder reaction as well as cycloaddition with oxygen, the latter of which results in the incipient formation of a rapidly degraded endoperoxide. In 1981, a team led by Simmons and Ammon reported the first stable isoindole by the reaction with dimethylene acetylenedicarboxylate.²² Here, the product of the Diels–Alder cycloaddition opened to deliver a stabilized isoindole by indirect



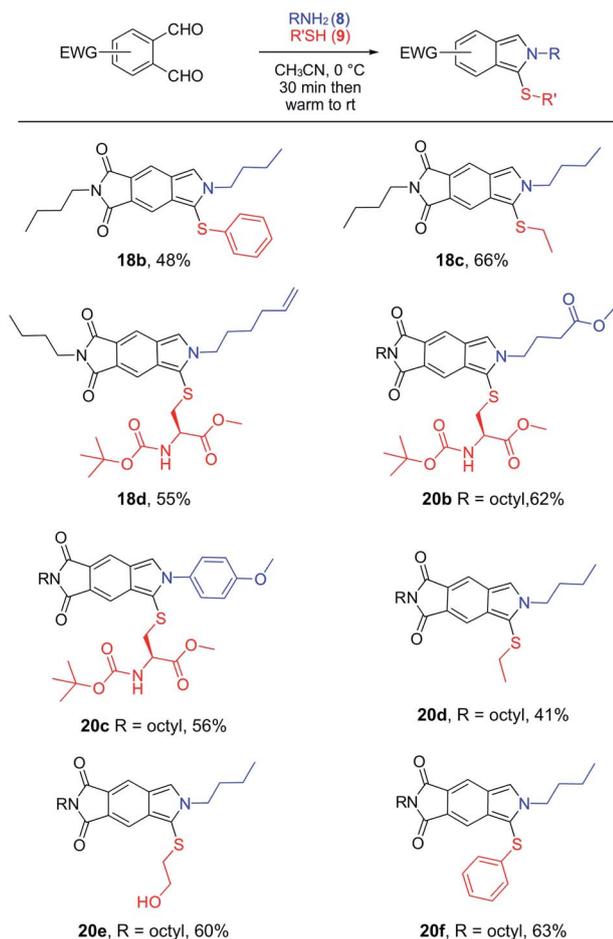


Fig. 4 One-pot synthesis of fluorescent 1-thio-2H-isoindoles from *n*-butyl- and *n*-octylphthalimidic phthalaldehydes **17** and **19**, aliphatic and aromatic amines, and aliphatic and aromatic thiols.

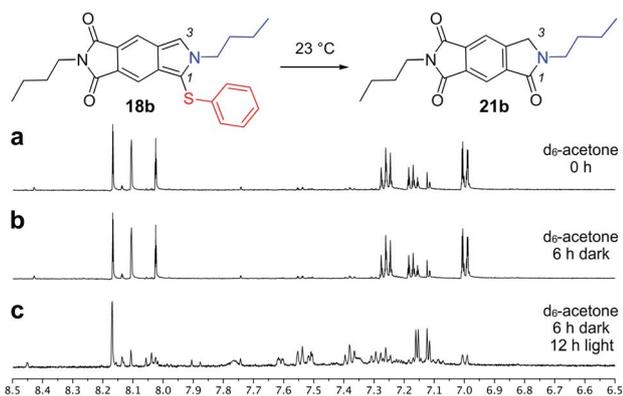


Fig. 5 Stability analyses on isoindole **18b** were evaluated using time course NMR studies. Selected spectra are shown with expansion of the aromatic region from 6.50–8.50 ppm. The predominant product of this degradation step found to be **21b**. Expansions and spectra from treating **18a**, **18c** and **18d** under the same conditions are provided in the ESI.†

functionalization at C3. While a practical discovery, the overall analytical utility of isoindoles would benefit from discovery of a material with bench stability.

Overall, we were able to prepare and collect spectral data on 13 new isoindoles (Fig. 3 and 4). These materials were sufficiently stable for purification and spectral analyses. While we were able to prepare and isolate **10a** and **12a**, spectra on these materials needed to be collected immediately after preparation. NMR analyses were most challenging due to the fact that trace acidic or basic materials in the solvents led to rapid decomposition ultimately leading to the use of acetone-*d*₆ for NMR data collection.

To further evaluate their utility, samples of **18a–d** were monitored for their stability neat and in solution. Subjecting these samples to the same protocol (see ESI†), isoindoles **18c** and **18d** were not sufficiently stable in neat form to endure >30 days at –20 °C followed by 48 h at 23 °C (conditions that modelled compound storage). Under the same conditions, **18a** and **18b** (Fig. 5a) were stable when stored (>30 days at –20 °C followed by 48 h at 23 °C) and when dissolved acetone-*d*₆ (Fig. 5a) and kept in the dark (Fig. 5b). Exposure to light, however, led to decomposition of both **18a** and **18b** (Fig. 5c). These observations indicated that steric bulk within the amine and thiol components contributed to the products stability. Additionally, the presence of alkene functionality within the amine was not tolerated, as given by the comparison of unstable **18d** to stable **18a**.

Synthesis of isoindoles²³ through a three-component coupling provides a robust tool to rapidly access diverse fluorescent materials as recently demonstrated by adaptation for a Click-like processes²⁴ or crosslinking,²⁵ called Flick. Here, we describe how the addition of electron withdrawing groups effectively stabilizes the materials as demonstrated by **18a** and **18b**. While this data suggests that stability can be achieved, the light sensitivity of these agents suggests a future potential as photochemical sensors or modifiers. Efforts are now underway to explore this application.

Conflicts of interest

There are no conflicts to declare.

Notes and references

- 1 G. Wittig, H. Tenhaeff, W. Schoch, G. Koenig and G. Einige, *Justus Liebig's Ann. Chem.*, 1951, **572**, 1–22.
- 2 R. Bonnett and R. F. C. Brown, *J. Chem. Soc., Chem. Commun.*, 1972, **7**, 393–395.
- 3 C. O. Bender and R. Bonnett, *J. Chem. Soc., Chem. Commun.*, 1966, **7**, 198–199.
- 4 R. Kreher and K. J. Herd, *Angew. Chem., Int. Ed. Engl.*, 1974, **13**, 739–740.
- 5 R. Kreher, N. Kohl and G. Use, *Angew. Chem., Int. Ed. Engl.*, 1982, **21**, 621.
- 6 P. S. G. Nunes, H. D. Vidal and A. G. Correa, *Org. Biomol. Chem.*, 2020, **18**, 7751–7773.



- 7 J. Fairsoosa, S. Saranya, S. Radhika and G. Anillkumar, *ChemistrySelect*, 2020, **5**, 5180–5197.
- 8 V. Jamsheena, R. K. Mishra, K. S. Veena, S. Sini, P. Jayamurthy and R. S. Lankalapalli, *ACS Omega*, 2018, **3**, 856–862.
- 9 S. B. Wagh, V. A. Maslivets, J. J. La Clair and A. Kornienko, *ChemBioChem*, 2021, **22**, 3109–3139.
- 10 G. Hillmann, *Hoppe-Seyler's Z. für Physiol. Chem.*, 1943, **277**, 222–232.
- 11 R. Roth, *Anal. Chem.*, 1971, **43**, 880–882.
- 12 S. S. Simons Jr and D. F. Johnson, *J. Org. Chem.*, 1978, **43**, 2886–2891.
- 13 S. S. Simons Jr and D. F. Johnson, *J. Am. Chem. Soc.*, 1976, **98**, 7098–7099.
- 14 Z. Yao, X. Feng, C. Li and G. Shi, *Chem. Commun.*, 2009, 5886–5888.
- 15 R. G. Carlson, K. Srinivasachar, R. S. Givens and B. K. Matuszewski, *J. Org. Chem.*, 1986, **51**, 3978–3983.
- 16 O. S. Wong, L. A. Sternson and R. N. Schowen, *J. Am. Chem. Soc.*, 1985, **107**, 6421–6422.
- 17 P. Zuman, *Anal. Lett.*, 2005, **38**, 1213–1220.
- 18 E. Trepman and R. F. Chen, *Arch. Biochem. Biophys.*, 1980, **204**, 524–532.
- 19 J. R. Cronin, S. Pizzarello and W. E. Gandy, *Anal. Biochem.*, 1979, **93**, 174–179.
- 20 S. S. Simons Jr and D. F. Johnson, *Anal. Biochem.*, 1978, **90**, 705–725.
- 21 M. C. G. Alvarez-Coque, M. J. M. Hernández, R. M. V. Camañas and C. M. Fernández, *Anal. Biochem.*, 1989, **178**, 1–7.
- 22 S. S. Simons Jr, H. H. Ammon, R. Doherty and D. F. Johnson, *J. Org. Chem.*, 1981, **46**, 4739–4744.
- 23 D. Grosheva and N. Cramer, *Angew. Chem., Int. Ed. Engl.*, 2018, **57**, 13644–13647.
- 24 M. Todorovic and D. M. Methods, *Enzymol*, 2020, **639**, 313–332.
- 25 M. Todorovic, K. D. Schwab, J. Zeisler, C. Zhang, F. Bénard and D. M. Perrin, *Angew. Chem., Int. Ed. Engl.*, 2019, **58**, 14120–14124.

