


 Cite this: *RSC Adv.*, 2022, **12**, 7446

## Effect of hydrophilic block end groups and block junction on block copolymer self-assembly in solution†

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Recent research suggests that the end groups of polymers can affect their self-assembly. However, the effect of end groups on the self-assembly of block copolymers in solution remains unclear, and thus far, only micelle–vesicle transformations have been achieved *via* end-group modification. Herein, we report that hydrophilic block end groups and the junction between two blocks can affect the solution self-assembly of block copolymers, leading to the formation of different morphologies, including vesicles, cubosomes, and hexosomes. Poly(ethylene glycol)-*b*-polystyrene (PEG-*b*-PS) with hydroxyl, methoxy, azido, or amino groups at the PEG chain ends was synthesized and self-assembled in solution *via* the cosolvent method. As a result, the morphology of the block copolymers transformed from vesicles to hexosomes upon increasing the end-group hydrophobicity. In addition, a morphological transition from cubosomes to vesicles was observed upon changing the junction from a triazole to an amide, and the interaction between the solvent and end groups significantly affected the self-assembly behavior.

Received 24th January 2022

Accepted 1st March 2022

DOI: 10.1039/d2ra00493c

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### 1. Introduction

Amphiphilic block copolymers (ABCPs) self-assemble in solution to form various morphologies such as micelles, vesicles,<sup>1–5</sup> cubosomes,<sup>6–19</sup> and hexosomes.<sup>10,11,20–28</sup> The chemical structure, block ratio, and molecular weight distribution of ABCPs influence their self-assembly.<sup>4,18,29,30</sup> In addition, recent studies have shown that the end groups of polymers, despite only accounting for small mass and volume fractions of an entire polymer, could affect or induce polymer self-assembly. The end group effect was first investigated in the self-assembly of hydrophilic homopolymers.<sup>31–33</sup> Xu *et al.* synthesized hydrophilic homo- and copolymers with two hydrophobic pyrene or cholesterol groups at the  $\omega$ -terminus.<sup>31</sup> The polymers self-assembled to form polymer vesicles, and it was demonstrated that the two rigid hydrophobic end groups were responsible for the vesicle formation. O'Reilly and co-workers observed unusual micelle formation by the self-assembly of hydrophilic homopolymers with hydrophobic end groups at both ends.<sup>32</sup> They synthesized a series of hydrophilic homopolymers with a pyrene and a dodecyl carbonotriithiocarbonate group at each end *via* reversible addition–fragmentation chain transfer (RAFT) polymerization. The homopolymers with the hydrophobic groups at both ends self-assembled in an acidic aqueous solution to form

well-defined aggregated micelles that behaved similarly to ABA' block copolymers. Liu *et al.* demonstrated that the terminal alkyne group of hydrophilic homopolymers could drive the polymer self-assembly into vesicles and flower-like complex particles.<sup>33</sup> They prepared a series of differently end-functionalized hydrophilic homopolymers *via* atom transfer radical polymerization (ATRP). During self-assembly in aqueous solution, only the homopolymers with terminal alkyne groups formed various morphologies including spherical micelles, multicomartment vesicles, and flower-like complex particles. The authors suggest that intermolecular hydrogen bonding between the terminal protons of the alkyne groups and oxygen atoms of the carbonyl groups drives the self-assembly.

The effect of end groups on block copolymer self-assembly has been investigated mostly based on bulk self-assembly<sup>34–37</sup> or polymerization-induced self-assembly (PISA).<sup>38,39</sup> Park and co-workers investigated the effect of the end groups of PEG blocks on the self-assembled morphology of PS-*b*-PEG along with other properties including conductivity and ion transport properties.<sup>34–37</sup> They observed the formation of a disordered phase from hydroxyl-terminated PEG-*b*-PS while sulfonate-terminated PEG-*b*-PS formed lamellar, hexagonal, and gyroid phases depending on the concentration of lithium salt added during the self-assembly.<sup>34</sup> Blackman *et al.* compared the photo- and thermally initiated PISA behaviors of poly(ethylene glycol)-*b*-(2-hydroxypropyl methacrylate).<sup>38</sup> They found that photo-initiated PISA produced higher-order structures owing to the loss of the end group during the process. In addition, extended irradiation of light or heat could induce a worm-to-vesicle morphological transition. Davis and co-workers reported that

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† Electronic supplementary information (ESI) available: Experimental details including synthetic procedures, characterization, and spectroscopy data. See DOI: 10.1039/d2ra00493c



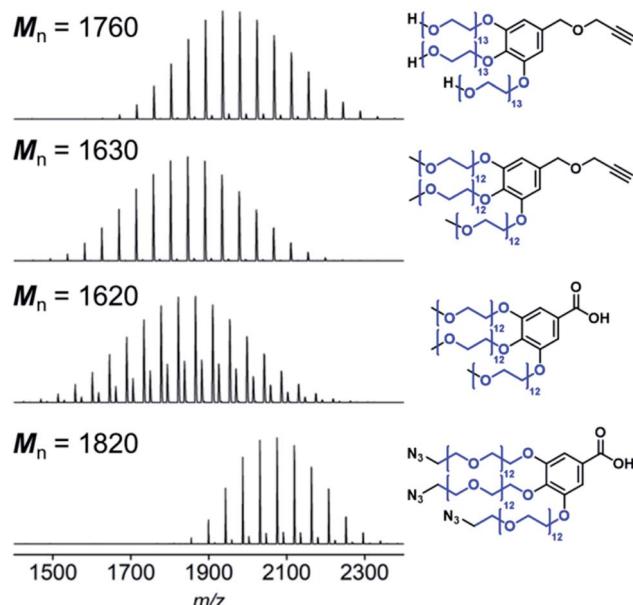


Fig. 1 MALDI-TOF spectra of the PEG blocks.  $M_n$  of the PEG chains (shown in blue) was calculated from the spectra (Table S1†).

vesicle formation could be induced from a micelle-forming PISA system by increasing the RAFT end-group hydrophobicity of the macromolecular chain transfer agent (macro-CTA).<sup>39</sup> They prepared two hydrophilic macro-CTAs with a carboxylic acid or a methyl ester end group and compared the PISA morphologies after the RAFT aqueous emulsion polymerization of styrene. Consequently, the methyl-ester-terminated block copolymer formed vesicles, whereas the carboxyl-terminated one formed micelles. With regard to self-assembly using the film hydration method, Grzelakowski and Kita-Tokarczyk synthesized a series of ABA or BA block copolymers comprising poly(2-methyloxazoline) (PMOXA) and poly(dimethylsiloxane) (PDMS) with different functional groups at both ends.<sup>40</sup> Notably, while the ABA block copolymer with hydroxyl groups at both ends formed micelles, the block copolymer self-assembled into vesicles after methacrylation of the hydroxyl groups despite a low conversion (*ca.* 5%) of the methacrylation. According to the authors, the morphological transition could be attributed to

the difference between the hydroxyl and methacryl groups in terms of size, dipole moment, and polarizability.

Recently, Che *et al.* reported a morphological transition of azide-modified PEG-*b*-PS ( $N_3$ -PEG-*b*-PS) vesicles into hexagonally packed hollow hoops (HHHs) induced by osmotic pressure during dialysis against NaCl solution.<sup>41</sup> In contrast, vesicles of PEG-*b*-PS with a methoxy end group (mPEG-*b*-PS) deformed into stomatocytes. The authors suggest that azide coordination to water molecules increases the hydrodynamic volume of the PEG at the internal side of the vesicles, compared to the PEG volume at the external side, leading to the formation of HHHs.

Most studies on the effect of end groups on block copolymer self-assembly have focused on the morphological transition between micelles and vesicles. The effect of end groups on inverse-phase-forming block copolymers has rarely been studied. Herein, we investigated the influence of the end groups of the hydrophilic block on inverse-phase-forming block copolymer self-assembly in solution. Additionally, we report that the junction between two blocks can affect the solution self-assembly of block copolymers. First, branched PEG hydrophilic blocks with methoxy, hydroxyl, or azido groups at the PEG chain ends were synthesized and coupled with PSSs *via* copper-catalyzed azide–alkyne cycloadditions (CuAAC) or amidation. PEG-*b*-PS with amino end groups was obtained *via* reduction of the PEG-PE with azido groups. Block copolymers with different

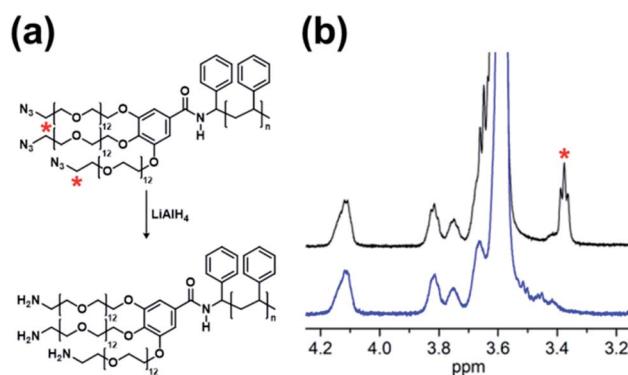


Fig. 3 (a) Reduction scheme of  $(N_3\text{-PEG})_3\text{-amd-PS1}$ . (b) NMR spectra (right) of  $(N_3\text{-PEG})_3\text{-amd-PS1}$  (black) and  $(H_2N\text{-PEG})_3\text{-amd-PS1}$  (blue).

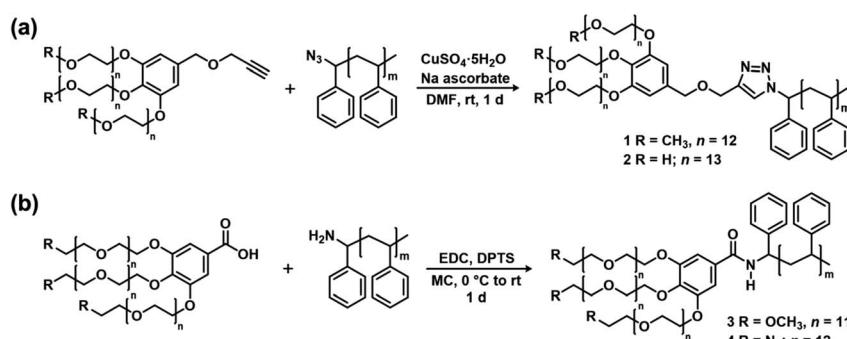


Fig. 2 Synthesis of the block copolymers with different end groups and junctions *via* (a) CuAAC and (b) amidation.

Table 1 Gel permeation chromatography (GPC) profiles of PS and block copolymers (BCPs)

BCP	$M_n$ (PEG) <sup>a</sup> (g mol <sup>-1</sup> )	$M_n$ (PS) <sup>b</sup> (g mol <sup>-1</sup> )	$M_n$ (BCP) <sup>b</sup> (g mol <sup>-1</sup> )	$D$ <sup>b</sup> (BCP)	$w_{PEG}$ (wt%)
(HO-PEG) <sub>3</sub> -trz-PS1	1760	16 900	20 200	1.09	10.4
(MeO-PEG) <sub>3</sub> -trz-PS2	1630	15 300	18 700	1.09	10.7
(MeO-PEG) <sub>3</sub> -amd-PS2	1620	15 300	18 900	1.05	10.6
(N <sub>3</sub> -PEG) <sub>3</sub> -amd-PS1	1820	16 900	20 000	1.05	10.8
(H <sub>2</sub> N-PEG) <sub>3</sub> -amd-PS1	1820	16 900	19 700	1.07	10.8

<sup>a</sup>  $M_n$  (PEG) was calculated from the MALDI-TOF spectra of each PEG block. <sup>b</sup>  $M_n$  (PS) ( $M_n$  of PS),  $M_n$  (BCP) ( $M_n$  of a block copolymer), and  $D$  (BCP) (dispersity of a block copolymer) were determined by GPC (DMF, 35 °C, 1 mL min<sup>-1</sup>) using PS standards.

end groups were self-assembled in solution *via* the cosolvent method to demonstrate the effect of the end groups on the self-assembled morphologies.<sup>4</sup> In addition, the variation in morphological changes in response to a solvent change from dioxane to acetone was studied based on the block copolymers with different end groups.

## 2. Results and discussion

### Synthesis of PEG-*b*-PS with different end groups

Starting from PEG monomethyl ether or PEG diol, we synthesized hydrophilic blocks with three PEG branches end-functionalized with methoxy, hydroxyl, and azido groups (Fig. S1†). Hydroxyl PEG block with an alkyne group at the other end was synthesized according to our previous study.<sup>13</sup> Methoxy PEG blocks with an alkyne group or a carboxyl group were synthesized by tethering  $\alpha$ -methoxy- $\omega$ -tosyl PEGs to a methyl gallate, which was followed by hydrolysis or reduction and subsequent etherification with a propargyl bromide to introduce carboxyl or alkyne group for conjugation with PS. Azido PEG block with a carboxyl group was prepared from  $\alpha$ -azido- $\omega$ -tosyl PEG in the same way. Synthesis of the hydrophilic blocks was confirmed by MALDI-TOF mass spectrometry and NMR spectroscopy (Table S1,† Fig. 1 and S2–S5†).

Two different PSs, PS1 ( $M_n = 16 900$  g mol<sup>-1</sup>) and PS2 ( $M_n = 15 300$  g mol<sup>-1</sup>), were obtained by ATRP. Their molecular weights were adjusted such that the block copolymers had a similar block ratio ( $w_{PEG}$ ), which was calculated as the ratio of the  $M_n$  of the PEG block to that of the PS block (Table 1 and S1†). The terminal bromo group of the PSs was converted into an azido group or an amino group for conjugation with the PEG blocks according to a previous study.<sup>42</sup>

The PEG blocks and the PS were coupled *via* CuAAC (Fig. 2a) or amidation (Fig. 2b) to obtain hydroxyl, methoxy, and azido PEG-*b*-PSs with a triazole (denoted “trz”) or an amide bond (denoted “amd”) linking the two blocks. The azido PEG-*b*-PS was further reduced to obtain the amino PEG-*b*-PS, (H<sub>2</sub>N-PEG)<sub>3</sub>-amd-PS1 and the conversion was confirmed by the disappearance of the NMR proton peak for the carbon adjacent to the azido group (Fig. 3). As a result, we prepared five block copolymers with different end groups and junctions but similar block ratios, as confirmed by GPC and NMR analysis (Table 1, Fig. 4 and S6–S10†).

### Self-assembly of the block copolymers in solution

The block copolymers were self-assembled *via* the cosolvent method using acetone or dioxane, which are common solvents, as previously described.<sup>13</sup> A block copolymer was dissolved in a common solvent and self-assembly was induced by slowly adding the same volume of water. We observed a wide range of morphologies from vesicles to hexosomes depending on the different end groups and junctions.

(HO-PEG)<sub>3</sub>-trz-PS1 formed vesicles in dioxane (Fig. 5b), whereas a mixture of spongesomes and cubosomes was formed in acetone (Fig. 5a and S11c†). As previously reported, the PS chain is assumed to have a reduced critical chain length ( $l_c$ ) owing to its lower solubility in acetone than in dioxane, leading to the formation of morphologies with a higher packing parameter ( $p = V/a_0 l_c$ ).<sup>13</sup> Under the same self-assembly conditions, (MeO-PEG)<sub>3</sub>-trz-PS2 self-assembled into cubosomes in both solvents (Fig. 5c, d, S11a and b†). Considering that the  $l_c$  in dioxane is greater than in acetone, the hydrophilic headgroup area ( $a_0$ ) of (MeO-PEG)<sub>3</sub>-trz-PS2 should be smaller in dioxane than in acetone to compensate for the difference in  $l_c$ .

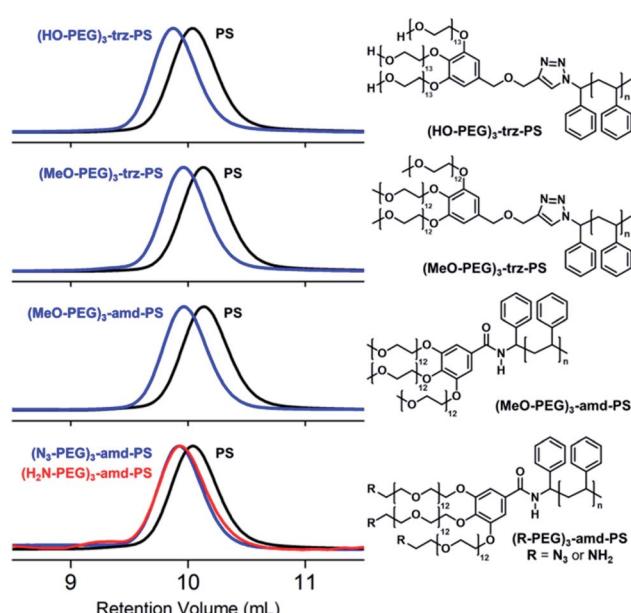


Fig. 4 GPC chromatograms (left) and chemical structures (right) of the BCPs. In the GPC chromatograms, PSs are shown in black and BCPs are shown in blue or red.



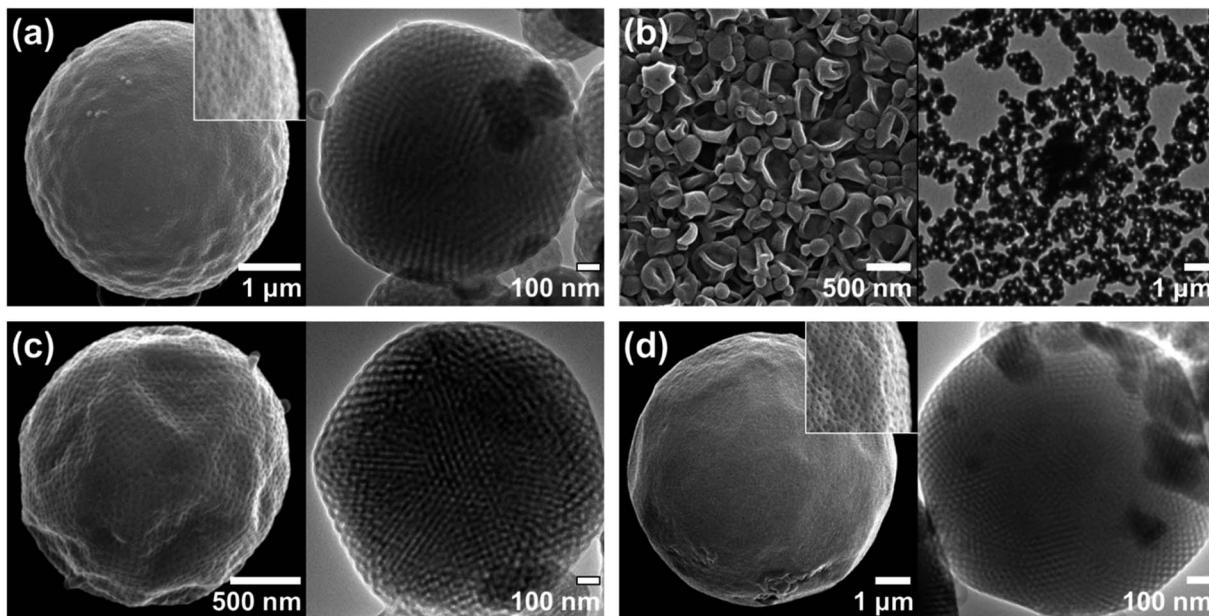


Fig. 5 Representative SEM and TEM images of (a and b)  $(HO-PEG)_3$ -trz-PS1 self-assembled in (a) acetone (cubosomes and spongesomes) and (b) dioxane (vesicles), and (c and d)  $(MeO-PEG)_3$ -trz-PS2 in (c) acetone (cubosomes) and (d) dioxane (cubosomes).

Moreover, it can be inferred that the  $a_0$  value is larger for  $(HO-PEG)_3$ -trz-PS1 than for  $(MeO-PEG)_3$ -trz-PS2 in both solvents, possibly owing to the higher hydrophilicity of the hydroxyl

group compared to that of the methoxy group. These results are consistent with those of the previous study by Davis and co-workers that revealed micelle–vesicle transformations with an

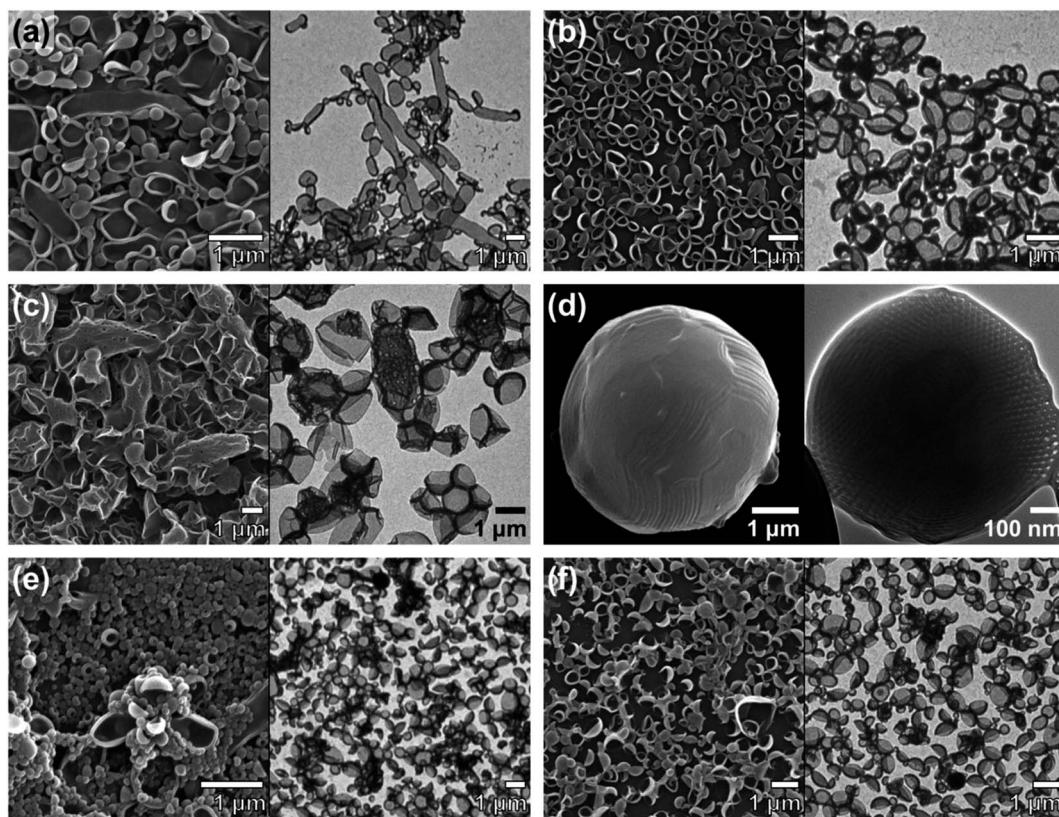


Fig. 6 Representative SEM and TEM images of (a and b)  $(MeO-PEG)_3$ -amd-PS2 self-assembled in (a) acetone (vesicles and lamellae) and (b) dioxane (vesicles), (c and d)  $(N_3-PEG)_3$ -amd-PS1 in (c) acetone (vesicles and sponges) and (d) dioxane (hexosomes), and (e and f)  $(H_2N-PEG)_3$ -amd-PS1 in (e) acetone (vesicles) and (d) dioxane (vesicles).

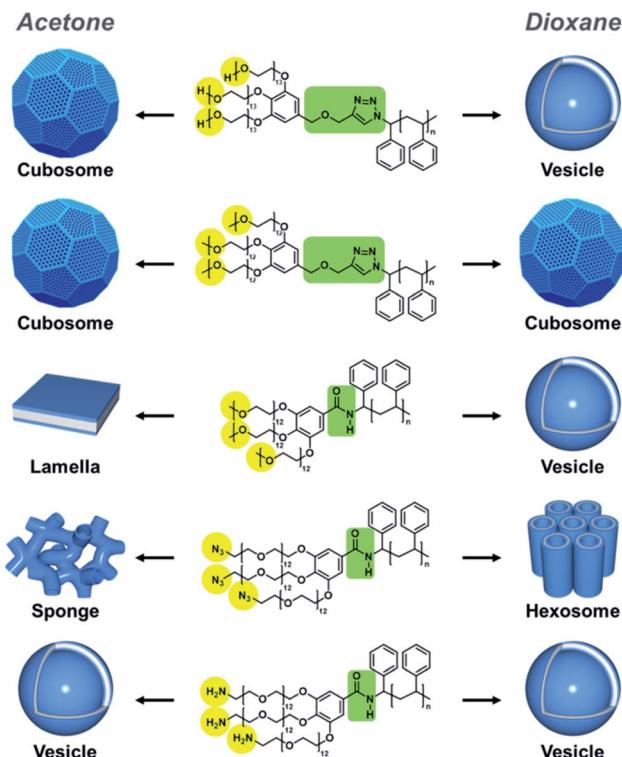


Fig. 7 Schematic representation of BCP self-assembly showing major morphologies self-assembled in acetone (left) and dioxane (right).

increasing packing parameter upon increasing the end-group hydrophobicity.<sup>39</sup> The hydroxyl and methoxy end groups directly affect the BCP self-assembly by varying the  $a_0$  due to their different interactions with water and the common solvent.

To further investigate the effect of azido and amino end groups, we compared the self-assembly of (MeO-PEG)<sub>3</sub>-amd-PS2, (N<sub>3</sub>-PEG)<sub>3</sub>-amd-PS1, and (H<sub>2</sub>N-PEG)<sub>3</sub>-amd-PS1 (Fig. 6 and 7). (MeO-PEG)<sub>3</sub>-amd-PS2 and (N<sub>3</sub>-PEG)<sub>3</sub>-amd-PS1 self-assembled in acetone to form a mixture of lamellae and vesicles or a mixture of sponges and vesicles, respectively (Fig. 6a and c). Interestingly, (N<sub>3</sub>-PEG)<sub>3</sub>-amd-PS1 self-assembled into hexosomes in acetone (Fig. 6d and S11d†) while (MeO-PEG)<sub>3</sub>-amd-PS2 only formed vesicles (Fig. 6b). This substantial difference in morphologies in terms of the packing parameter may arise from the higher hydrophobicity of azido PEGs. Hartwig and co-workers reported that the conversion of a hydrogen to an azide in cycloheximide and digoxigenin

derivatives increased their distribution coefficient values.<sup>43</sup> Therefore, the azido PEG should have higher hydrophobicity than that of the methoxy PEG.<sup>44–46</sup> Specifically, (N<sub>3</sub>-PEG)<sub>3</sub>-amd-PS1 should possess a significantly smaller  $a_0$  value than that of vesicle-forming (MeO-PEG)<sub>3</sub>-amd-PS2, leading to high-packing-parameter hexosome formation in dioxane. Moreover, it can be inferred that (N<sub>3</sub>-PEG)<sub>3</sub>-amd-PS1 should possess a much smaller  $a_0$  value in dioxane than that in acetone to increase the packing parameter while the  $l_c$  of PS decreases. Meanwhile, only vesicles were observed from the self-assembly of (H<sub>2</sub>N-PEG)<sub>3</sub>-amd-PS1 in both solvents owing to the high hydrophilicity of the amino groups.<sup>43</sup>

Remarkably, we found that the junction between the two blocks could also affect the BCP self-assembly. In both solvents, (MeO-PEG)<sub>3</sub>-trz-PS2 and (MeO-PEG)<sub>3</sub>-amd-PS2 self-assembled into cubosomes and mainly vesicles, respectively (Fig. 5c, d, S11a, b, S6a, and b†). Owing to the methoxy end groups, both BCPs exhibited small morphological changes upon varying the solvent from acetone to dioxane; this is due to the decrease in  $a_0$  offsetting the increase in  $l_c$ . An amide unit can act both as a strong hydrogen-bonding donor and an acceptor, whereas a 1,2,3-triazole unit is a weak hydrogen bonding acceptor.<sup>47,48</sup> Therefore, the hydrogen bonding between the amide bond in (MeO-PEG)<sub>3</sub>-amd-PS2 and other amide junctions or water molecules could affect the BCP conformation. The difference in the length of the junction between (MeO-PEG)<sub>3</sub>-trz-PS2 and (MeO-PEG)<sub>3</sub>-amd-PS2 could be another factor.

Based on the aforementioned reasons, (MeO-PEG)<sub>3</sub>-amd-PS2 may possess a larger  $a_0$  value than that of (MeO-PEG)<sub>3</sub>-trz-PS2, leading to vesicle formation, whereas (MeO-PEG)<sub>3</sub>-trz-PS2 forms cubosomes under the same self-assembly conditions. Moreover, the effect of hydrogen-bonding-forming units on the self-assembly process has been studied based on the supramolecular self-assembly of low-molecular-weight  $\pi$ -amphiphiles.<sup>49–53</sup> Considering the differences in the self-assembly behaviors of (MeO-PEG)<sub>3</sub>-amd-PS2 and (MeO-PEG)<sub>3</sub>-trz-PS2, we suggest that a small difference in the junction could affect the self-assembly of high-molecular-weight BCPs. Regarding the effect of the junction, Hawker and co-workers previously demonstrated that the phase separation strength of a BCP could be increased by ionization of its triazole junction, which leads to electrostatic interactions between counterions of adjacent domain junctions.<sup>54</sup> The self-assembly behavior with respect to the end groups and the junction is summarized in Table 2 and Fig. 7 for each common solvent.

Table 2 Summary of self-assembled morphologies in acetone and dioxane

BCP	Self-assembled morphology		End group hydrophilicity	Junction
	Acetone	Dioxane		
(HO-PEG) <sub>3</sub> -trz-PS1	Cubosome	Vesicle	Hydrophilic	Triazole
(MeO-PEG) <sub>3</sub> -trz-PS2	Cubosome	Cubosome	Hydrophobic	Triazole
(MeO-PEG) <sub>3</sub> -amd-PS2	Lamella, vesicle	Vesicle	Hydrophobic	Amide
(N <sub>3</sub> -PEG) <sub>3</sub> -amd-PS1	Sponge, vesicle	Hexosome	Hydrophobic	Amide
(H <sub>2</sub> N-PEG) <sub>3</sub> -amd-PS1	Vesicle	Vesicle	Hydrophilic	Amide



From the thermodynamic aspect, BCP self-assembly in solution relies on interfacial energy, which is determined by solvent affinity and block composition of the BCP.<sup>46,55</sup> In the cosolvent method, the self-assembled morphology of a BCP depends on the affinity of the blocks to a solvent mixture of water and the common solvent. Considering the effect of the core-block hydrophobicity on BCP self-assembly,<sup>44–46</sup> hydrophilicity of the corona block should also influence the self-assembled morphologies of the BCP. According to our findings, different end groups strongly affect the hydrophilicity of the corona block, thus leading to significant changes in self-assembled morphologies.

### 3. Conclusion

We have investigated the effect of different end groups on BCP self-assembly in solution. BCPs with hydroxyl, methoxy, azido, and amino groups at the end of the hydrophilic PEG block self-assembled into different morphologies ranging from vesicles to hexosomes under the same self-assembly conditions. Based on the packing parameter ( $p$ ) analogy, we speculate that the interactions between the end groups and a common solvent (acetone and dioxane) or water could determine the headgroup area ( $a_0$ ) of the BCPs in solution, leading to the different self-assembly behaviors. Interestingly, different junctions between two blocks such as amide and triazole units also resulted in the self-assembly of different morphologies. This was attributed to the hydrogen bonding capability or the different lengths of the junctions. In summary, we have demonstrated that the end groups and junction, which account for a very small portion of an entire BCP, can significantly influence the BCP self-assembly. We envision that our findings will provide new insights to control the self-assembly of block copolymers.

### Conflicts of interest

There are no conflicts of interest to declare.

### Acknowledgements

We acknowledge supports from the National Research Foundation (NRF) of Korea (2019R1A2C3007541), the Technology Innovation Program (or Industrial Strategic Technology Development Program-Alchemist Project) (20012390, 4D Molecular-Nano-Addressable Lithographic Self-Assembly (4D MONALISA)) funded by the Ministry of Trade, Industry & Energy (MOTIE, Korea), and Creative-Pioneering Researchers Programs (30520190050) by Seoul National University (SNU).

### Notes and references

- 1 J. C. M. van Hest, D. A. P. Delnoye, M. W. P. L. Baars, M. H. P. van Genderen and E. W. Meijer, *Science*, 1995, **268**, 1592–1595.
- 2 D. E. Discher and A. Eisenberg, *Science*, 2002, **297**, 967–973.
- 3 A. Blanazs, S. P. Armes and A. J. Ryan, *Macromol. Rapid Commun.*, 2009, **30**, 267–277.
- 4 Y. Mai and A. Eisenberg, *Chem. Soc. Rev.*, 2012, **41**, 5969–5985.
- 5 U. Tritschler, S. Pearce, J. Gwyther, G. R. Whittell and I. Manners, *Macromolecules*, 2017, **50**, 3439–3463.
- 6 Y. La, C. Park, T. J. Shin, S. H. Joo, S. Kang and K. T. Kim, *Nat. Chem.*, 2014, **6**, 534–541.
- 7 H. Yu, X. Qiu, S. P. Nunes and K.-V. Peinemann, *Nat. Commun.*, 2014, **5**, 4110.
- 8 T. H. An, Y. La, A. Cho, M. G. Jeong, T. J. Shin, C. Park and K. T. Kim, *ACS Nano*, 2015, **9**, 3084–3096.
- 9 Y. La, T. H. An, T. J. Shin, C. Park and K. T. Kim, *Angew. Chem., Int. Ed.*, 2015, **54**, 10483–10487.
- 10 Z. Lin, S. Liu, W. Mao, H. Tian, N. Wang, N. Zhang, F. Tian, L. Han, X. Feng and Y. Mai, *Angew. Chem., Int. Ed.*, 2017, **56**, 7135–7140.
- 11 X. Lyu, A. Xiao, W. Zhang, P. Hou, K. Gu, Z. Tang, H. Pan, F. Wu, Z. Shen and X.-H. Fan, *Angew. Chem., Int. Ed.*, 2018, **57**, 10132–10136.
- 12 Y. La, J. Song, M. G. Jeong, A. Cho, S.-M. Jin, E. Lee and K. T. Kim, *Nat. Commun.*, 2018, **9**, 5327.
- 13 S. Ha and K. T. Kim, *Polym. Chem.*, 2019, **10**, 5805–5813.
- 14 J. Kim, M. Yoon, S.-M. Jin, J. Lee, Y. La, E. Lee and K. T. Kim, *Polym. Chem.*, 2019, **10**, 3778–3785.
- 15 H.-K. Liu, L.-J. Ren, H. Wu, Y.-L. Ma, S. Richter, M. Godehardt, C. Kübel and W. Wang, *J. Am. Chem. Soc.*, 2019, **141**, 831–839.
- 16 X. Liu and I. Gitsov, *Macromolecules*, 2019, **52**, 5563–5573.
- 17 Y. Sun, J. Kim and K. T. Kim, *RSC Adv.*, 2019, **9**, 25423–25428.
- 18 S. Ha, Y. La and K. T. Kim, *Acc. Chem. Res.*, 2020, **53**, 620–631.
- 19 H. Ma and K. T. Kim, *Macromolecules*, 2020, **53**, 711–718.
- 20 L. Zhang, C. Bartels, Y. Yu, H. Shen and A. Eisenberg, *Phys. Rev. Lett.*, 1997, **79**, 5034–5037.
- 21 A. L. Parry, P. H. H. Bomans, S. J. Holder, N. A. J. M. Sommerdijk and S. C. G. Biagini, *Angew. Chem., Int. Ed.*, 2008, **47**, 8859–8862.
- 22 B. E. McKenzie, F. Nudelman, P. H. H. Bomans, S. J. Holder and N. A. J. M. Sommerdijk, *J. Am. Chem. Soc.*, 2010, **132**, 10256–10259.
- 23 S. J. Holder and N. A. J. M. Sommerdijk, *Polym. Chem.*, 2011, **2**, 1018–1028.
- 24 B. E. McKenzie, S. J. Holder and N. A. J. M. Sommerdijk, *Curr. Opin. Colloid Interface Sci.*, 2012, **17**, 343–349.
- 25 B. E. McKenzie, J. F. de Visser, H. Friedrich, M. J. M. Wirix, P. H. H. Bomans, G. de With, S. J. Holder and N. A. J. M. Sommerdijk, *Macromolecules*, 2013, **46**, 9845–9848.
- 26 B. E. McKenzie, H. Friedrich, M. J. M. Wirix, J. F. de Visser, O. R. Monaghan, P. H. H. Bomans, F. Nudelman, S. J. Holder and N. A. J. M. Sommerdijk, *Angew. Chem., Int. Ed.*, 2015, **54**, 2457–2461.
- 27 B. E. McKenzie, J. F. de Visser, G. Portale, D. Hermida-Merino, H. Friedrich, P. H. H. Bomans, W. Bras, O. R. Monaghan, S. J. Holder and N. A. J. M. Sommerdijk, *Soft Matter*, 2016, **12**, 4113–4122.
- 28 H. He, K. Rahimi, M. Zhong, A. Mourran, D. R. Luebke, H. B. Nulwala, M. Möller and K. Matyjaszewski, *Nat. Commun.*, 2017, **8**, 14057.



- 29 K. E. B. Doncom, L. D. Blackman, D. B. Wright, M. I. Gibson and R. K. O'Reilly, *Chem. Soc. Rev.*, 2017, **46**, 4119–4134.
- 30 C. K. Wong, X. Qiang, A. H. E. Müller and A. H. Gröschel, *Prog. Polym. Sci.*, 2020, **102**, 101211.
- 31 J. Xu, L. Tao, C. Boyer, A. B. Lowe and T. P. Davis, *Macromolecules*, 2011, **44**, 299–312.
- 32 J. Du, H. Willcock, J. P. Patterson, I. Portman and R. K. O'Reilly, *Small*, 2011, **7**, 2070–2080.
- 33 T. Liu, W. Tian, Y. Zhu, Y. Bai, H. Yan and J. Du, *Polym. Chem.*, 2014, **5**, 5077–5088.
- 34 G. Jo, H. Ahn and M. J. Park, *ACS Macro Lett.*, 2013, **2**, 990–995.
- 35 G. Jo, O. Kim, H. Kim, U. Hyeok Choi, S.-B. Lee and M. Jeong Park, *Polym. J.*, 2016, **48**, 465–472.
- 36 H. Y. Jung, P. Mandal, G. Jo, O. Kim, M. Kim, K. Kwak and M. J. Park, *Macromolecules*, 2017, **50**, 3224–3233.
- 37 J. Kim, H. Y. Jung and M. J. Park, *Macromolecules*, 2020, **53**, 746–763.
- 38 L. D. Blackman, K. E. B. Doncom, M. I. Gibson and R. K. O'Reilly, *Polym. Chem.*, 2017, **8**, 2860–2871.
- 39 S. Y. Khor, N. P. Truong, J. F. Quinn, M. R. Whittaker and T. P. Davis, *ACS Macro Lett.*, 2017, **6**, 1013–1019.
- 40 M. Grzelakowski and K. Kita-Tokarczyk, *Nanoscale*, 2016, **8**, 6674–6683.
- 41 H. Che, L. N. J. de Windt, J. Zhu, I. A. B. Pijpers, A. F. Mason, L. K. E. A. Abdelmohsen and J. C. M. van Hest, *Chem. Commun.*, 2020, **56**, 2127–2130.
- 42 K. Matyjaszewski, Y. Nakagawa and S. G. Gaynor, *Macromol. Rapid Commun.*, 1997, **18**, 1057–1066.
- 43 R. R. Karimov, A. Sharma and J. F. Hartwig, *ACS Cent. Sci.*, 2016, **2**, 715–724.
- 44 J. C. Foster, S. Varlas, B. Couturaud, J. R. Jones, R. Keogh, R. T. Mathers and R. K. O'Reilly, *Angew. Chem., Int. Ed.*, 2018, **57**, 15733–15737.
- 45 S. Varlas, J. C. Foster, L. A. Arkinstall, J. R. Jones, R. Keogh, R. T. Mathers and R. K. O'Reilly, *ACS Macro Lett.*, 2019, **8**, 466–472.
- 46 J. C. Foster, I. Akar, M. C. Grocott, A. K. Pearce, R. T. Mathers and R. K. O'Reilly, *ACS Macro Lett.*, 2020, **9**, 1700–1707.
- 47 Y. Hua and A. H. Flood, *Chem. Soc. Rev.*, 2010, **39**, 1262–1271.
- 48 D.-Y. Wang, L.-Y. You, J.-L. Wang, H. Wang, D.-W. Zhang and Z.-T. Li, *Tetrahedron Lett.*, 2013, **54**, 6967–6970.
- 49 M. R. Molla and S. Ghosh, *Chem.-Eur. J.*, 2012, **18**, 9860–9869.
- 50 P. Rajdev, M. R. Molla and S. Ghosh, *Langmuir*, 2014, **30**, 1969–1976.
- 51 A. Sikder, A. Das and S. Ghosh, *Angew. Chem., Int. Ed.*, 2015, **54**, 6755–6760.
- 52 A. Sikder, D. Ray, V. K. Aswal and S. Ghosh, *Angew. Chem., Int. Ed.*, 2019, **58**, 1606–1611.
- 53 A. Sikder, Y. Xie, M. Thomas, M. J. Derry and R. K. O'Reilly, *Nanoscale*, 2021, **13**, 20111–20118.
- 54 Y. Luo, D. Montarnal, N. J. Treat, P. D. Hustad, M. D. Christianson, E. J. Kramer, G. H. Fredrickson and C. J. Hawker, *ACS Macro Lett.*, 2015, **4**, 1332–1336.
- 55 Q. Zhang, J. Lin, L. Wang and Z. Xu, *Prog. Polym. Sci.*, 2017, **75**, 1–30.

