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# A benzothiazole-based new fluorogenic chemosensor for the detection of $\text{CN}^-$ and its real-time application in environmental water samples and living cells†

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Since the cyanide ion is used in a wide range of industries and is harmful to both human health and the environment, a number of research efforts are dedicated to creating fluorescence sensors for the detection of cyanide ( $\text{CN}^-$ ). Herein, for the fluorescence detection of  $\text{CN}^-$ , a new highly selective and sensitive sensor 2-(3-(benzo[d]thiazol-2-yl)-4-hydroxybenzylidene)-1*H*-indene-1,3(2*H*)-dione (**BID**) was created by conjugating a benzothiazole moiety with 1*H*-indene-1,3(2*H*)-dione. The donor and acceptor components of this hybrid receptor were covalently connected through a double bond. The nucleophilic addition of a cyanide anion to the **BID** inhibits the intramolecular charge transfer (ICT) transition, resulting in spectral and colour alterations in the receptor. When the solvent polarity was increased from *n*-hexane to methanol, this molecule exhibited a bathochromic shift in the emission wavelength (610 to 632 nm), suggesting the presence of a solvatochromic action. The sensor **BID** has shown strong specificity towards  $\text{CN}^-$  by interrupting its internal charge transfer (ICT), resulting in a significant change in the UV-vis spectrum and a notable blue shift in the fluorescence emission spectrum. The cyanide anion ( $\text{CN}^-$ ) is responsible for the optical alterations observed by **BID**, as opposed to the other anions examined. The detection limit was 5.97 nM, significantly less than the WHO's permitted amount of  $\text{CN}^-$  in drinking water. The experimental findings indicate that **BID**'s fluorescence response to  $\text{CN}^-$  is pH insensitive throughout a wide pH range of 6.0 to 12.0. The interaction mechanism between the **BID** and  $\text{CN}^-$  ions has been studied by HRMS, <sup>1</sup>H-NMR titration experiments, FT-IR, and DFT, which confirmed the nucleophilic addition of  $\text{CN}^-$  on vinylidene and subsequent disturbance of ICT. Additionally, we demonstrated the real-time detection application of  $\text{CN}^-$  in environmental water samples and live-cell imaging.

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## Introduction

Ions have a variety of important functions in chemical and biological processes.<sup>1–9</sup> Among them, many anions are naturally occurring, and they have significant roles in essential disciplines, including life science, chemistry, medicine, catalysis, and the environment.<sup>10–14</sup> To this end, for the past few years, anion recognition has become an area of growing concern in the field of environmental and biological sciences. Many types of anions are considered to be extremely dangerous, but one in particular, cyanide ( $\text{CN}^-$ ), has always been of interest all over the world because of its potency and universal presence in many processes

such as manufacturing of plastic and fiber products, tanning, metallurgy, the petrochemical industry, herbicide production, chelators in water treatment, and silver or gold extraction. Despite its overwhelming application in various fields,  $\text{CN}^-$  is still harmful to humans even at a dosage of only 0.5 mg per kilogram.<sup>15</sup> Any unintentional discharge of cyanide ions will result in significant environmental risks. The hydrolysis of cyanogenic glycosides in plants such as bitter almonds and bitter cassava as well as cherry, apple, and peach seeds can also create toxic HCN besides industrial disposal. The health effects of excessive exposure to  $\text{CN}^-$  include paralysis of the central nervous system, poisoning of the respiratory enzymes, and hemoglobin poisoning. As a result, breathing becomes difficult, and the body's cells begin to die from hypoxia (low oxygen levels). Therefore, a more sensitive and selective approach for detecting hazardous cyanide anions is still greatly needed.<sup>16</sup>

Various analytical techniques have been applied to determine cyanide ion concentration, including voltammetry, electrochemistry, polarography, and gas chromatography.<sup>17–23</sup>

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Despite their low detection limits, these techniques need sophisticated sample processing, costly equipment, and trained operators. Due to the increased popularity of fluorescent  $\text{CN}^-$  sensors, which benefit from excellent selectivity, fast reaction time, and less complex sample preparation, these sensors have advanced significantly in the past several years.<sup>24–37</sup> The reported cyanide fluorescence sensors involved the following mechanistic concepts: coordination, hydrogen bonding interaction, anion-metal affinity, nucleophilic addition reactions, and based on nanotechnology.<sup>32,38–52</sup> Hydrogen bonding, anion metal affinity, and coordination sensors all have significant drawbacks, including a lack of selectivity and interference from competing analytes such as fluoride, phosphate, and acetate ions. However, among the numerous chemosensors described, reaction-based sensors (chemodosimeters) offer the benefits of high specificity, outstanding sensitivity, and rapid response time. Due to the nucleophilic nature of cyanide ions, reaction-based approaches often exhibit high selectivity for cyanide detection. These factors provide the groundwork for the development of reaction-based sensors for the detection of cyanide ions. For example, some of the already reported reaction-based cyanide sensors include imines,<sup>53,54</sup> amide,<sup>55,56</sup> hydrazones,<sup>57</sup> deprotonation mechanism of C–H,<sup>58</sup> acridinium salts,<sup>59</sup> cationic boranes,<sup>60</sup> oxazines.<sup>61</sup> However, slow response time, requirement of high temperature, need of basic medium, high detection limit, less selectivity, inefficiency in aqueous medium hamper the sensing ability of these chemodosimeters. Nucleophilic-based cyanide fluorescence probes have been created and used widely because of their excellent selectivity and increased sensitivity. In recent times, sono-aided chemical precipitation methods have been developed for the synthesis of ZnS, ZnO and quantum dots (QDs).<sup>74,72</sup> It exhibits sensational photocatalytic as well as biological activities.<sup>73,74</sup> Moreover, the biological toxicity levels of ZnS QDs were characterised by UV-visible absorption and fluorescence emission spectrum.<sup>75</sup>

With our interest in  $\text{CN}^-$  recognition,<sup>34,62,63</sup> in this work, we employ benzothiazole as a fluorophore to develop and manufacture a turn-on cyanide fluorescence probe based on the nucleophilic addition process. A benzothiazole unit was introduced because of its rigid and planar shape and exceptional photophysical characteristics. The donor and acceptor units were covalently bonded together in this hybrid structure by the use of a double bond. Nucleophilic addition to the vinyl double bond causes a disruption of the intramolecular charge transfer (ICT) transition, which results in alterations in both spectral and colour properties. In the probe (**BID**), the benzothiazole moiety functions as an electron donor, while dione acts as an electron receiver, generating fluorescence through Intramolecular Charge Transfer (ICT). Benzothiazole and 1*H*-indene-1,3(2*H*)-dione are linked by Knoevenagel condensation reaction, which renders increased conjugation and nucleophilic addition site for recognizing cyanide ions. When  $\text{CN}^-$  ion was added to an ACN/ $\text{H}_2\text{O}$  solution containing **BID**, the fluorescence spectra indicated that the substantial fluorescence amplification occurred at 512 nm and the solution colour changed to yellow under a 365 nm UV light. At the same time, **BID**'s absorption spectra showed a definite peak at 593 nm with a high intensity.

**BID**'s determined detection limit for CN was 5.97 nM, which is quite low. The fast response to  $\text{CN}^-$ , strong detecting properties, and ease of setup make this new sensor very promising. The sensor **BID** was used to demonstrate the applicability by detecting  $\text{CN}^-$  in water, agricultural goods, using test strips, and live cells imaging. It provides a dependable method for detecting  $\text{CN}^-$  in environmental and biological systems.

## Experimental section

### Material and apparatus

The required chemicals were obtained from Sigma Aldrich and Alfa Aesar. Chemicals were utilized in the reaction without further purification. Solvents used for synthesis were LR-grade solvents and were used without further purification. Solvents used in spectroscopy were of HPLC grade. Silica gel 100–200 mesh was used for column chromatography.  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectra were taken in BrukerAscent-400 spectrometer at 25 °C.  $^1\text{H}$  NMR and  $^{13}\text{C}$  were recorded in  $\text{CDCl}_3$  and  $\text{DMSO}-d_6$ . Chemical shifts ( $\delta$ ) are given in ppm relative to TMS internal standard. Absorption spectra were recorded on a Shimadzu 3600 spectrophotometer. Emission spectra were recorded on a PerkinElmer LS-55 luminescence spectrometer. Samples for absorption and emission were taken in a quartz cuvette (4 ml volume).

**Synthesis of 2-(3-(benzo[*d*]thiazol-2-yl)-4-hydroxybenzylidene)-1*H*-indene-1,3(2*H*)-dione (**BID**).** The compound 3-(benzo[*d*]thiazol-2-yl)-4 hydroxybenzaldehyde was dissolved in analytical grade EtOH (15 ml). To this crude solution, indane-1,3-dione (1 mmol) and a catalytic amount of piperidine was added. Then, the reaction mixture was allowed to reflux for about three hours. TLC confirmed the completion of the reaction, and the pure compound had been isolated by column chromatography (in 50% ethyl acetate in *n*-hexane). The obtained pure **BID** is orange colour solid. The yield was found to be 32%.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  6.968 (s, 1H), 7.425–7.443 (t, 1H,  $J = 7.2$  Hz), 7.533–7.552 (t, 1H,  $J = 7.6$  Hz), 7.796 (s, 1H), 7.890–7.896 (t, 5H,  $J = 2.4$  Hz), 8.068–8.087 (d, 1H,  $J = 7.6$  Hz), 8.152–8.171 (d, 1H,  $J = 7.6$  Hz), 8.676 (s, 1H), 9.448 (s, 1H).  $^{13}\text{C}$  NMR ( $\text{DMSO}-d_6$ , 100 MHz):  $\delta$  100.98, 117.07, 121.49, 125.16, 129.17, 129.42, 131.43, 135.80, 140.89, 152.47, 166.48, 188.14.

**Optical selectivity and titration experiments.** A stock solution of **BID** ( $2 \times 10^{-5}\text{M}$ ) was prepared in ACN/ $\text{H}_2\text{O}$  (7 : 3) system. A quartz optical cell of a 1 cm optical path was used to record absorption and emission spectra. For each addition of  $\text{CN}^-$ , the concentration of **BID** was maintained at  $2 \times 10^{-5}\text{M}$ . The excitation wavelength used for fluorescence analysis was 320 nm, and we got emissions at 393 nm and 408 nm. Stock solutions of various anions such as  $\text{F}^-$ ,  $\text{Cl}^-$ ,  $\text{Br}^-$ ,  $\text{I}^-$ ,  $\text{NO}_3^-$ ,  $\text{SCN}^-$ ,  $\text{N}_3^-$ ,  $\text{ClO}_4^-$ ,  $\text{H}_2\text{PO}_4^-$ ,  $\text{AcO}^-$ ,  $\text{HCO}_3^-$ ,  $\text{SO}_4^{2-}$ , and  $\text{HSO}_4^-$  were prepared by dissolving their respective salts in double-distilled water.

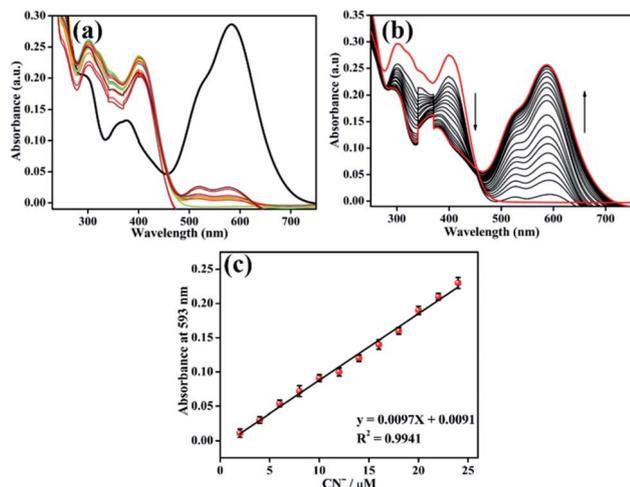
## Results and discussion

### Molecular design, synthesis, and structural characterization

With the dione-vinyl group of **BID**, we intended to use the nucleophilic addition reaction of  $\text{CN}^-$  to develop a sensor

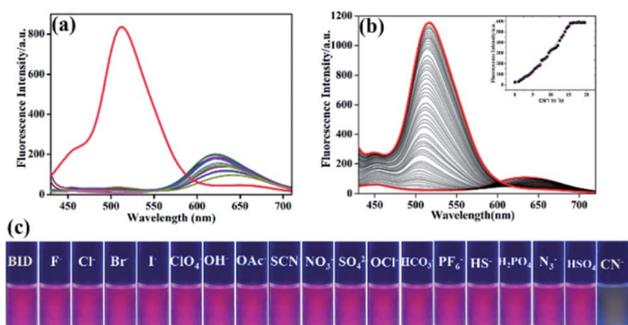






**Fig. 2** (a) UV-vis absorption spectrum of sensor **BID** ( $1 \times 10^{-5}$  M) in the presence of 10.0 equiv. of different anions in an ACN/H<sub>2</sub>O (7 : 3, v/v) solution; (b) sensor **BID** ( $1 \times 10^{-5}$  M) absorption spectra in an ACN/H<sub>2</sub>O (7 : 3, v/v) solution after addition of different concentrations of CN<sup>-</sup> anions (0–2.6 equiv.); (c) linear relationship between  $A_{593}$  nm and the cyanide concentration (0–2.6 equiv.).

The fluorescence sensing capability of **BID** towards CN<sup>-</sup> in a solution was then investigated. Excitation of probe **BID** at 398 nm resulted in a maximal emission band with low intensity at 625 nm, as illustrated in Fig. 3a. The addition of different anions such as (10 equiv.) F<sup>-</sup>, Cl<sup>-</sup>, Br<sup>-</sup>, I<sup>-</sup>, OH<sup>-</sup>, AcO<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, SCN<sup>-</sup>, OCl<sup>-</sup>, HCO<sub>3</sub><sup>-</sup>, HS<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, HSO<sub>4</sub><sup>-</sup>, PF<sub>6</sub><sup>-</sup> and N<sub>3</sub><sup>-</sup> to the sensor solution did not affect **BID**'s colour and emission spectra remained nearly unchanged. The only significant change occurred when CN<sup>-</sup> was introduced, resulting in a blue-shifted fluorescence at 512 nm, was the complete disappearance of the emission band at 625 nm (Fig. 3a). To further examine **BID**'s CN<sup>-</sup> fluorescence sensing characteristics, a series of fluorescence titration studies with CN<sup>-</sup> were performed (Fig. 3b). The cyanide ion inhibited the ICT transition inside the molecule,

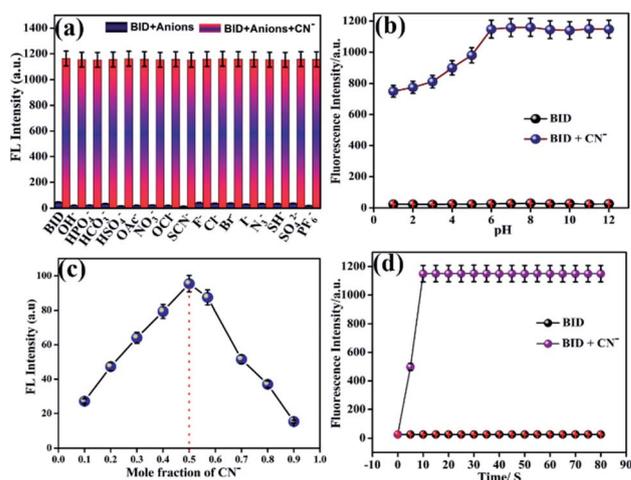


**Fig. 3** (a) Fluorescence spectra of **BID** ( $1 \times 10^{-6}$  M) in the presence of different analytes (10.0 equiv.  $1 \times 10^{-6}$  M); (b) fluorescence emission of probe **BID** ( $1 \times 10^{-6}$  M) in ACN/H<sub>2</sub>O (7 : 3, v/v) solution in the presence of a varying concentration of CN<sup>-</sup> from 0 to 1.5 μM, inset shows the plot of  $I_{525}$  vs. the concentration of CN<sup>-</sup>; (c) changes in the solution fluorescence color of **BID** with different analytes under a UV light of 365 nm.

raising the sensor's emission peak at 512 progressively when CN<sup>-</sup> ions were gradually added to the sensor solution. Fluorescence increase at 525 nm of **BID** was related to CN<sup>-</sup> concentrations throughout an extensive range from 0 to 1.5 μM, as indicated by the correlation coefficient of 0.9947, suggesting that **BID** was an excellent sensor for the quantitative detection of CN<sup>-</sup>. This whole event corresponds to a fluorescence colour change of **BID** solution from pink to pale yellow. The sensitivity of **BID** was also investigated fluorometrically. Various solutions of anions were added individually to the **BID** solutions. Meanwhile, the visual colour changes were observed as shown in Fig. (3c). The detection limit (LOD) of sensor **BID** for CN<sup>-</sup> was determined to be 5.97 nM with a linear range of concentration to be 0–0.5 μM (Fig. S6†), which is far below the allowable content of CN<sup>-</sup> in drinking water and the results were compared with previously reported chemosensors for CN<sup>-</sup> (Table S3†). The limit of quantitation (LOQ) was estimated to be 1.99 nM, which is calculated based on the equation of  $10\sigma$ /slope.

### Effect of competitive anions, pH, reaction time and salt concentrations

A competitive experiment was conducted to illustrate the unique selectivity of **BID** for CN<sup>-</sup>. Anion CN<sup>-</sup> (2.0 equiv.) was applied to sensor **BID** in the presence of other competing analytes (also 2.0 equiv.) evaluated in the optical experiments. Anions coexisting with CN<sup>-</sup> did not affect its fluorescence change, as demonstrated in Fig. 4a. These results indicate that sensor **BID** has good selectivity for the fluorescence detection of cyanide anion in the presence of another competing anion, making it particularly relevant in practical applications for cyanide detection. In addition, the impact of pH on cyanide anion sensing was investigated. Fig. 4b depicts the changes in



**Fig. 4** (a) Competitive experiments of the **BID** + CN<sup>-</sup> system with interfering anions ( $\lambda_{\text{ex}} = 398$  nm) in ACN/H<sub>2</sub>O (7 : 3); (b) the fluorescence intensity of **BID** (1 μM) in the absence and presence of CN<sup>-</sup> ( $\lambda_{\text{ex}} = 398$  nm) as a function of pH (1.0–12.0); (c) Job's plot for determining the stoichiometry of **BID** and CN<sup>-</sup> ions; (d) the time-dependent fluorescence spectrum of **BID** toward CN<sup>-</sup> in ACN/H<sub>2</sub>O (7 : 3).



sensor **BID**'s fluorescence at 525 nm as a function of pH, with and without the addition of  $\text{CN}^-$ . It was found that the fluorescence signal of the free sensor **BID** in the pH range of 1.0 to 12.0 was very weak and was only marginally influenced by  $\text{H}^+$  or  $\text{OH}^-$ . When  $\text{CN}^-$  is present in the pH range of 1.0–12.0, the fluorescence at 525 nm of the sensor **BID** increases considerably up to pH = 6, and the **BID** fluorescence response to  $\text{CN}^-$  between 6.0 and 12.0 was almost pH-independent. Because  $\text{CN}^-$  anions are quickly converted to weak acid, HCN and due to the protonation of **BID** (the guest), the host-guest contact is reduced, and the fluorescence emission of **BID** is reduced or diminished. The binding mode of **BID**– $\text{CN}^-$  was examined by Job's plot experiments. Thereby, the complex (**BID**– $\text{CN}^-$ ) demonstrates a maximum absorbance in the 0.5 mole fraction. Hence, 1 : 1 stoichiometry of complexation has been achieved (Fig. 1c). To determine the probe **BID**'s capability for fast cyanide detection, we studied the time courses of the fluorescence intensity increase of probe **BID** ( $1 \times 10^{-6}$  M) in the presence of  $\text{CN}^-$  (2 equiv.) in a mixture of ACN/ $\text{H}_2\text{O}$  (7 : 3, v/v) at room temperature (Fig. 4d). Fig. S7† illustrates the fluorescence changes in **BID** and **BID**– $\text{CN}^-$  at various salt concentrations demonstrate that the changes in the exhibited fluorescence curve are minor in NaCl solution, showing that the synthesised **BID** and **BID**– $\text{CN}^-$  are highly stable.

### Investigation of sensing mechanism

Further, we decided to investigate the **BID**– $\text{CN}^-$  reaction product to learn more about the sensing mechanism of sensor **BID** for  $\text{CN}^-$  exposure. The  $^1\text{H}$  NMR spectra data in  $\text{DMSO}-d_6$  at room temperature before and after cyanide anion addition are shown in Fig. 5a. A peak at 8.75 ppm corresponds to the vinyl proton ( $\text{H}_a$ ), which was eliminated following the addition of 1.0 equiv.  $\text{CN}^-$ , whereas a new signal at 5.18 ppm corresponds to the  $\alpha$ -proton  $\text{H}_b$  appeared. Additionally, the phenolic proton, which appeared at 9.44 ppm, presented an up-field shift to 8.48 ppm. Meanwhile, the aromatic proton signals from 1*H*-indene-1,3(2*H*)-dione exhibited a significant up-field shift compared to those from the **BID**. These outcomes may be

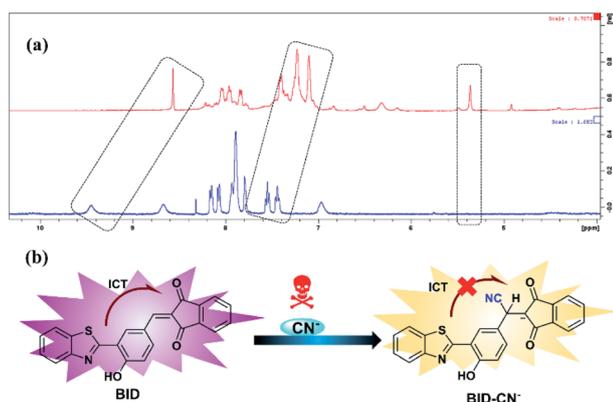


Fig. 5 (a)  $^1\text{H}$  NMR spectra (400 Hz,  $\text{DMSO}-d_6$ ) of sensor **BID** in the absence and presence of  $\text{CN}^-$  (1.0 equiv.). (b) The suggested response mechanism for the sensor **BID** in conjunction with  $\text{CN}^-$ .

explained by **BID**'s nucleophilic addition to  $\text{CN}^-$ , which disrupts conjugation between the benzothiazole framework and the 1*H*-indene-1,3(2*H*)-dione group and decreases dione's electron-withdrawing capacity. Furthermore, HRMS analysis verified the formation of the newly created adduct (**BID**– $\text{CN}^-$ ) (Fig. S3†). According to mass spectral analysis, the peak of **BID** was found to be 383.0618  $m/z$ . After the one equiv. of  $\text{CN}^-$  was added into the **BID** solution, the **BID**– $\text{CN}^-$  peak with a mass of 408.0566  $m/z$  appeared, showing conclusively that **BID** :  $\text{CN}^-$  is a 1 : 1 stoichiometric ratio. FT-IR spectra were also used to validate the mechanism of interaction between **BID** and  $\text{CN}^-$ . As depicted in Fig. S8,† it was noticed that in the presence of  $\text{CN}^-$  (1.0 equiv.), the typical stretching bands for the saturated hydrocarbon (CH) group emerged at 2972 and 2858  $\text{cm}^{-1}$ , while the distinctive stretching vibration of (C=C) at 1581  $\text{cm}^{-1}$  entirely vanished. Meanwhile, the absorption peak of keto (C=O) group stretching vibrations was moved from 1683 to 1726  $\text{cm}^{-1}$ .

### Theoretical calculations

The complex formation between the  $\text{CN}^-$  ion and **BID** was studied further at the molecular level utilizing a density functional theory-based computational method. The density functional theory at the B3LYP/631 G(d) level was utilized effectively to predict photochromism and reaction-based photochromic molecules with high accuracy.<sup>29,64–70</sup> The entire geometry optimizations of the probe **BID** and its interaction with  $\text{CN}^-$  were performed using the aforementioned technique, and its structure is shown in Fig. S9.† The transition energies and relative oscillator strengths of the probable important transitions for the **BID** and **BID** +  $\text{CN}^-$ , have been found (Table S1†). As shown in Fig. 6, HOMO of **BID** has electron densities that are restricted to the 1*H*-indene-1,3(2*H*)-dione group alone, and it has been calculated that there is 3.43 eV of energy difference ( $\Delta E$ ) between the HOMO and LUMO ( $E_{\text{HOMO}} = -6.4120$  eV and  $E_{\text{LUMO}} = -2.9777$  eV). As a result of the ICT that happens in the

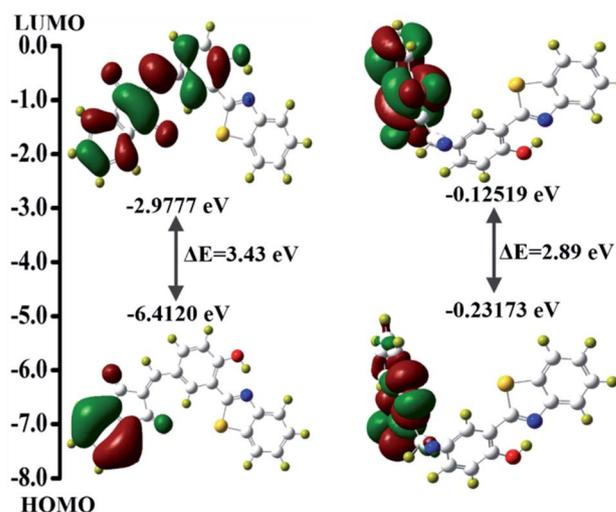


Fig. 6 The optimal structures of **BID** and **BID** +  $\text{CN}^-$  determined by DFT/B3LYP methods.



excited state, it is discovered that a considerable amount of electron transfer occurs between the benzothiazole unit and the 1*H*-indene-1,3(2*H*)-dione functionality (LUMO). Despite this, when the electron allocation in **BID** + CN<sup>-</sup> is taken into consideration, both the HOMO and LUMO localized entirely inside the 1*H*-indene-1,3(2*H*)-dione functionality, resulting in the electronic excitation occurring in the same chromophore, which severely limits the ICT effect.

### BID loaded paper-based indicators for CN<sup>-</sup> ion

To further evaluate the chemosensor's utility as a quick approach for detecting CN<sup>-</sup> levels in water samples, paper test strips were impregnated with the chemosensor **BID** ( $2 \times 10^{-3}$  M) using a concentrated acetonitrile solution. After dropping different anion solutions (2 mM) onto a "fluorescence paper-based indicator," the colour change was observed while exposed to a 365 nm UV light. Fig. 7 exhibited that the paper-based fluorescent indicator loaded with **BID** presented dark pink emission. The fluorescence emission remained constant even after adding various anions as tested in the selectivity study. As expected, it could be observed that the fluorescence was blue after CN<sup>-</sup> was dropped. As predicted, it was apparent that the fluorescence had changed from pink to blue once the CN<sup>-</sup> was dropped. **BID** paper-based indicator can thus be used for the visual inspection of CN<sup>-</sup> in an easy and straightforward manner.

### Application in real samples analysis

As a result of worldwide industrial operations and improper waste disposal, many water sources have become severely polluted, endangering aquatic life and the ecosystem. Thus, the development of an effective analytical technique for quantifying CN<sup>-</sup> in environmental samples is imperative. The sensitivity of the **BID** to the detection of CN<sup>-</sup> in real water samples such as tap water, distilled water, and pond water from Vellore Institute of Technology (VIT), Vellore campus, was assessed using the conventional addition approach. The collected water samples were filtered three times with a 0.45 mm membrane filter, then spiked with a particular amount of CN<sup>-</sup>, diluted ten times, and were put to the test using standard procedures (Table 2).

The standard addition technique was used to calculate the CN<sup>-</sup> concentration, which involved the addition of 2, 4, and 6 μM of CN<sup>-</sup> to the above-mentioned water samples. It can be seen from Table 1 that, with satisfactory recovery (97.4–100.2%) and low RSD (0.024–1.37%), the **BID** sensor was able to detect the spiked CN<sup>-</sup>, demonstrating the feasibility and reliability of the sensor **BID** for the determination of CN<sup>-</sup> in a variety of

BID	F <sup>-</sup>	Cl <sup>-</sup>	Br <sup>-</sup>	I <sup>-</sup>	OH <sup>-</sup>	SCN <sup>-</sup>	ClO <sub>2</sub> <sup>-</sup>	NO <sub>2</sub> <sup>-</sup>	OAc <sup>-</sup>
SO <sub>4</sub> <sup>2-</sup>	HCO <sub>3</sub> <sup>-</sup>	PF <sub>6</sub> <sup>-</sup>	SH <sup>-</sup>	N <sub>3</sub> <sup>-</sup>	H <sub>2</sub> SO <sub>4</sub>	H <sub>2</sub> PO <sub>4</sub> <sup>-</sup>	OCi	CN <sup>-</sup>	

Fig. 7 Photographs of **BID** in the presence of various anions and CN<sup>-</sup> under a UV lamp at 365 nm.

Table 2 Results of CN<sup>-</sup> readings in water samples from the environment

Sample	Spiked (μM)	Found		
		( $X^a \pm SD^b$ ) (μM)	Recovery (%)	RSD <sup>c</sup> (%)
Tap water	2.0	1.992 ± 0.007	99.6	0.18
	4.0	3.998 ± 0.007	99.9	0.22
	6.0	6.009 ± 0.003	100.2	0.36
Distilled water	2.0	2.003 ± 0.002	100.2	0.34
	4.0	3.992 ± 0.007	99.8	0.44
	6.0	6.011 ± 0.008	100.2	0.62
Pond water	2.0	1.948 ± 0.004	97.4	0.60
	4.0	3.921 ± 0.006	98.0	0.51
	6.0	5.933 ± 0.005	98.9	0.78

<sup>a</sup> Mean of three determinations. <sup>b</sup> SD: standard deviation. <sup>c</sup> RSD: relative standard deviation.

environmental water samples. In addition, the obtained results were cross verified with <sup>1</sup>H-NMR based analysis. In this method, peak intensity gradually increases at 5.18 ppm and is used to derive <sup>1</sup>H-NMR calibration plot (Fig S10 and Table S2†).

### Bio-imaging application

Additionally, *Escherichia coli* (*E. coli*) cells were employed as a model system for further use of as-synthesized **BID** on bacterium imaging. As a result, preliminary cytotoxicity testing was performed. As shown in Fig. S11,† there was no discernible reduction in *E. coli* viability when **BID** concentrations were increased to 20 μM, supporting **BID**'s extremely minimal impact on *E. coli*. Additionally, it was discovered that *E. coli* had a distinct fluorescent red short dot form and that no apparent restriction of *E. coli* growth was seen under the fluorescence microscope. In the other group, **BID** was incubated with *E. coli*

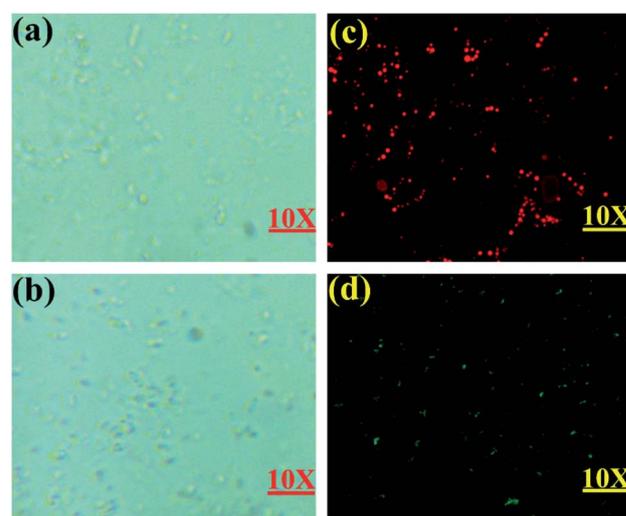


Fig. 8 Fluorescent cell imaging: (a) and (b) bright-field image of *E. coli* cells using the green filter of 510 to 560 nm; (c) fluorescent field image of **BID** treated *E. coli* cells; (d) fluorescent field image of **BID** treated *E. coli* cells incubated with CN<sup>-</sup> ions using the yellow filter of 532 to 587 nm.



bacteria for 20 minutes at 37 °C before being incubated with 20 μM of CN<sup>-</sup> for another 20 minutes, resulting in the development of green fluorescence (Fig. 8d). The results showed that the probe was capable of detecting the CN<sup>-</sup> ion in live cells.

## Conclusions

In summary, a new fluorescence sensor **BID** based on benzo-thiazole and a 1*H*-indene-1,3(2*H*)-dione derivative for detecting CN<sup>-</sup> has been developed. Optical studies revealed that **BID** has excellent sensitivity and selectivity toward CN<sup>-</sup> while other anions examined have no effect. Under UV light, a dramatic fluorescent color shift from pink to pale yellow was noticed, as well, as a substantial fluorescence increase was observed when CN<sup>-</sup> was added to the sensor **BID** solution. The sensing mechanism was built on the nucleophilic addition reaction of the sensor with CN<sup>-</sup>. This was also verified by optical studies, <sup>1</sup>H NMR titration, FT-IR spectra, HRMS, and DFT/TD-DFT calculations. The detection limit was determined to be as low as 5.97 nM, and the quick reaction time following the addition of CN<sup>-</sup>, indicates that sensor **BID** may be utilized as an effective sensor for CN<sup>-</sup> detection. Additionally, the sensor **BID** was successfully utilized to detect CN<sup>-</sup> in environmental water samples, enabling the sensor's expansion into food industries. Furthermore, the sensor **BID** responds to CN<sup>-</sup> with fluorescence, allowing image detection and real-time monitoring of CN<sup>-</sup> in live *E. coli* cells.

## Conflicts of interest

There are no conflicts to declare.

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