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Synthetic strategy and structure-activity relationship (SAR) studies of 3-(5'-hydroxymethyl-2'-furyl)-1-benzyl indazole (YC-1, Lificiguat): a review

Since 1994, YC-1 (Lificiguat, 3-(5'-hydroxymethyl-2'-furyl)-1-benzylindazole) has been synthesized, and

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many targets for special bioactivities have been explored, such as stimulation of platelet-soluble guanylate cyclase, indirect elevation of platelet cGMP levels, and inhibition of hypoxia-inducible factor-1 Received 5th November 2021 (HIF-1) and NF-κB. Recently, Riociguat®, the first soluble guanylate cyclase (sGC) stimulator drug used to treat pulmonary hypertension and pulmonary arterial hypertension, was derived from the YC-1 DOI: 10.1039/d1ra08120a structure. In this review, we aim to highlight the synthesis and structure-activity relationships in the development of YC-1 analogs and their possible indications.

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Introduction

YC-1 (Lificiguat, 3-(5'-hydroxymethyl-2'-furyl)-1-benzyl indazole) is an indazole derivative whose structure contains furyl and benzyl substituents at positions 1 and 3 (Fig. 1). In 1994, YC-1 was first synthesized and reported by Kuo's group and exhibited antiplatelet activity with a 70% inhibition of platelet aggregation induced at a concentration of 30 μ mol L⁻¹.^{1,2} YC-1 is a potential antiplatelet agent for prevention and treatment of vascular embolisms. The mechanisms of action of the antiplatelet activity of YC-1 include direct stimulation of plateletsoluble guanylate cyclase (sGC) and indirect elevation of platelet cyclic guanosine monophosphate (cGMP) levels. Since the discovery of YC-1 in 1994, many other molecular targets of YC-1 have been identified. YC-1 is a hypoxia-inducible factor-1 (HIF-1) inhibitor^{3,4} that exhibits anticancer activities,⁵⁻⁷ reverse chemoresistance, 8,9 retinal neovascularization inhibition, 10,11 neural protection12,13 and decreased allergic inflammatory gene expression.14 In addition, Carroll et al.15 found that YC-1 displays inhibitory activity against vascular endothelial growth factor (VEGF) for treating inflammation.¹⁶ Additionally, YC-1 was found to inhibit the activity of the nuclear factor kappa light chain enhancer of activated B cells (NF-κB),17 which is a critical signaling pathway for learning and memory, 18 as well as bone formation.19

The structure-activity relationship (SAR) of YC-1 and its derivatives have been studied in the past decade. However, in the past reviews^{20,21} of YC-1, only the relevant pharmacological activities were discussed, leaving open the synthesis strategy

and SAR. Some valuable YC-1 derivatives have even been developed into clinical drugs, such as Riociguat. Riociguat, whose structure is based on YC-1, is a clinical drug indicated for pulmonary artery hypertension (PAH) via stimulating soluble guanylate cyclase (sGC). Thus, we mainly focus on the chemical synthesis of YC-1, the synthesis of its derivatives, and discussion of the SAR in this review.

Based on an overview of all the chemical syntheses for YC-1 and its derivatives, the starting materials are easily available and, inexpensive raw materials.2,22-24 The derivatives are modified mainly on the functional groups in aromatic and furyl rings and the number of nitrogen atoms. The chemical modification reactions comprise reduction, oxidation, esterification, substitution reactions, Suzuki coupling, and direct arylation. In the following section of this review, we comprehensively describe the synthetic methods, biological activities, and the SAR of YC-1 and summarize the most effective structure for various biological activities (HIF-1, sGC, VEGF and NF-κB), which will be beneficial for future research and development of YC-1 derivatives with optimized bioactivity.

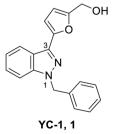


Fig. 1 Structure of YC-1 (1)

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Synthetic strategies of 3-(5'hydroxymethyl-2'-furyl)-1-benzyl indazole (YC-1) and YC-1 derivatives

The synthesis of YC-1 derivatives has attracted considerable attention in organic and medicinal chemistry due to their excellent bioactivities. Different synthetic strategies have been developed depending on the desired chemical structure. Here, we summarize several synthetic strategies used in the synthesis of YC-1 (Scheme 1).

The synthesis of YC-1 was first described by Teng et al. (2002).25,26 Their synthetic plan started from benzoyl chloride and ethyl furan-2-carboxylate catalyzed by ferric chloride to obtain carbonyl coupling compound 4 (route a). Then, methyl 5benzoylfuran-2-carboxylate (4) was converted to hydrazine compound 5 using benzyl hydrazine and acetic acid. Compound 5 underwent a cyclization reaction catalyzed by lead(iv) acetate [Pb(OAc)₄] and then boron trifluoride (BF₃) to obtain compound 6. Finally, the ester group was reduced to a hydroxyl group via calcium borohydride $[Ca(BH_4)_2]$ to yield the final compound 1 (YC-1), and the total yield was 4.3%.

In the original synthesis route of YC-1, the corresponding indazole from cyclization of the hydrazine compound had several drawbacks, including too many steps, use of organic metal reagents, and low yield. Collot et al.,27 Hering et al.28 and Takeuchi et al.22 modified the synthetic process of YC-1 starting

from indazole (9) (route b).29 First, indazole mixed with iodine (I₂), and potassium hydroxide (KOH) in dimethylformamide (DMF) was used to install the iodo group at position 3. Next, benzyl bromide (BnBr), potassium tert-butoxide (t-BuOK), and tetrabutylammonium iodide (Bu₄NI) in tetrahydrofuran (THF) were reacted with compound 10 to obtain benzyl substituent 11. Hering²⁸ and Takeuchi²² prepared YC-1 via stannane reaction using palladium catalysis from 1-benzyl-3-iodoindazole, and the yield increased to 74%. However, organostannanes are generally considered inappropriate for drug synthesis due to their toxicity. Collot avoided organostannane reagents. Instead, Suzuki coupling reaction of 11 with boronic acid substituted with aromatic compound (12) was carried out to obtain 13 by Vilsmeier–Haack reaction.²⁷ Reduction of the formyl group with sodium borohydride was applied to obtain YC-1, and the yield improved to 79%.

Even though the yield improved to above 70%, there were still doubts about the use of heavy metals in the total synthesis. Hence, in 2008, Carla et al. developed a synthesis method that did not involve the use of heavy metals.24,30 In this report, all of the reagents used were less toxic and less expensive than those used in previous studies (route d). The starting materials were 2aminobenzoic acid (20) and furan, which reacted with n-butyllithium in THF to obtain carbonyl compound 21. Compound 21 reacted with hydroxylamine hydrochloride to give oxime compound 22, which underwent a cyclization reaction using methanesulfonyl chloride (MsCl) to give compound 19. The

Scheme 1 Summary of synthesis of YC-1 (1). Route a: synthesis by Teng et al. (2002); route b: synthesis by Collot et al. and Hering et al. (1999, 2006); route c: synthesis by Meng et al. (2017); route d: synthesis by Carla et al. (2008). Reagents and conditions: (a) FeCl₃; (b) PhCH₂NHNH₂, HOAc; (c) Pb(OAc)₄; (d) BF₃Et₂O; (e) Ca(BH₄)₂; (f) NaBH₄, MeOH, rt; (g) Pd₂(dba)₃, AsPh₃, DMF; (h) POCl₃, DMF, 0 °C to 80 °C; (i) I₂, KOH, DMF; (j) BnBr, t-BuOK, Bu₄NI, THF; (k) Pd(PPh₃)₄, NaHCO₃, DME, reflux; (l) ArCH₂Br, t-BuOK, THF, 0 °C to rt; (m) THF, -80 °C; (n) MnO₂, CH₂Cl₂; (o) (i) TsNHNH₂, HCl, MeOH; (ii) Cu₂O, i-AmOH, reflux; (p) Mg, MeOH; (q) MsCl, Et₃N, CH₂Cl₂; (r) furan, n-BuLi, THF, -78 °C to 0 °C; (s) HO-NH₂ HCl, pyridine, MeOH, reflux.

final product, YC-1, was prepared following formylation and reduction of compound 19.²⁷ This synthetic route not only reduced heavy metal usage but also retained the total yield.

In addition, in 2004 and 2007, Inamoto *et al.*³¹ and Sakamoto *et al.*³² reported the first method for preparing indazole through intramolecular cyclization of Pd-catalyzed *ortho*-bromine tosylhydrazone under the action of a base and ligand. However, the reaction must be carried out under alkaline conditions, and compound **24** was formed by Bamford–Stevens reaction, where the *E*-form was the main product. The *E*-form isomer cannot obtain the corresponding product (**25**); in contrast, the *Z*-form can give better reaction results (Scheme 2). In 2017, Meng *et al.* pointed out for the first time that indazole (**25**) can be obtained by the reaction of an *E*-form or *Z*-form tosylhydrazone isomer with cuprous oxide (Cu₂O) by heating (Scheme 2), which is suitable for a variety of functional groups and has an extremely high yield.²³

Meng's group also reported another synthetic strategy (Scheme 1, route c) based on 2-bromobenzaldehyde (14). (2-Bromophenyl)(furan-2-yl)methanone (17) was prepared through affinity addition and oxidation, after which it was cyclized by tosylhydrazone to form indazole compound 18. Removing tosyl and adding benzyl groups was obtained compound 19. Subsequently, 19 was subjected to Vilsmeier–Haack reaction and reduced to obtain YC-1 (1).

3. Structure—activity relationship (SAR) result of YC-1

3.1. Soluble guanylate cyclase (sGC)

YC-1 was first discovered as a novel sGC stimulator that is influenced by NO or NO donors.³³ Upon sGC stimulation, several physiological events occur, including inhibition of the contraction of vascular smooth muscles and inhibition of platelet adhesion and aggregation.^{2,34} YC-1 may also potentiate the action of endogenous and exogenous carbon monoxide (CO) on sGC and stimulate sGC with NO in a synergistic manner. In addition, administration of YC-1 may affect age-related learning

and memory dysfunction *via* the NO/cGMP signaling pathway, which sGC crucially contributes to. ^{18,35,36} For example, in Morris water maze (MWM) avoidance response tests, YC-1-treated group (1 mg kg⁻¹) improved rodent learning behavior and performed a higher avoidance response rate in which the mean avoidance response was 18.7% and 33.3% for the control and YC-1-treated rats, respectively, on day 11.³⁷

Moreover, YC-1 has been found to have an important regulatory effect on sGC and cGMP signaling in the cardiovascular system, where YC-1 directly stimulates sGC and then activates NO production that subsequently relaxes vascular smooth muscles. 38,39 Furthermore, YC-1 inhibits focal adhesion kinase and proliferative factor transforming growth factor β 1 (TCF- β 1) to reduce the growth of vascular smooth muscle. 40 In addition, YC-1 prevents oxidized LDL-mediated apoptosis and induces Hsp70 expression. 39

Due to many stimulate activities related to sGC, the SAR discussion in the next section is divided into two parts: relaxation of vascular smooth muscles and antiplatelet activities.

3.1.1. YC-1 derivatives as sGC stimulator. In all the SAR studies of YC-1, sGC has been the most investigated. Compound inhibition abilities (IC_{50}) were measured by the inhibition of the maximum constriction of phenylephrine-induced preconstricted rabbit aortic rings^{26,41} and the inhibition of sodium nitroprusside (SNP)-induced apoptosis of rat aortic smooth muscle cells.⁴² The following SAR research is discussed according to \mathbb{R}^1 , \mathbb{R}^2 , heterocycle A and heterocycle B functional groups (Fig. 2).

Fig. 2 Generic structure to represent SAR for YC-1.

Scheme 2 Synthesis of indazole (25), an intermediate of YC-1 synthesis.

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$$\begin{array}{c} \text{Relaxation of the preconstricted} \\ \text{PC-1 (1), R= benzyl} & \text{IC}_{50}=10 \ \mu\text{M} \\ \textbf{26a, R}^1=2\text{-fluorobenzyl} & \text{IC}_{50}=4.9 \ \mu\text{M} \\ \textbf{26b, R}^1=3\text{-fluorobenzyl} & \text{IC}_{50}=10 \ \mu\text{M} \\ \textbf{26c, R}^1=4\text{-fluorobenzyl} & \text{IC}_{50}=19 \ \mu\text{M} \\ \textbf{26d, R}^1=2,6\text{-difluorobenzyl} & \text{IC}_{50}=15 \ \mu\text{M} \\ \textbf{26e, R}^1=2\text{-cyanobenzyl} & \text{IC}_{50}=8.7 \ \mu\text{M} \\ \textbf{26e, R}^1=2\text{-cyanobenzyl} & \text{IC}_{50}=26 \ \mu\text{M} \\ \textbf{R}^1 & \textbf{26g, R}^1=2\text{-pyridyl} & \text{IC}_{50}=16 \ \mu\text{M} \\ \textbf{26 dh, R}^1=3\text{-pyridyl} & \text{IC}_{50}=9.6 \ \mu\text{M} \\ \textbf{26 de, R}^1=3\text{-pyrimldinyl} & \text{IC}_{50}=3.2 \ \mu\text{M} \\ \textbf{26 de, R}^1=3\text{-pyrimldinyl} & \text{IC}_{50}=3\text{-pyrimldinyl} \\ \textbf{26 de, R}^1=3\text{-pyrimldinyl} & \text{IC}_{50}=3\text{-pyr$$

Fig. 3 Effect of 1-*N*-substituted derivatives (26) on relaxation of preconstricted aortic rings.

According to the structure of YC-1, Straub et al. discussed the SAR based on 1-N-substituent whose synthesis followed route b in Scheme 1 and replaced the benzyl bromide functional group to obtain compound 26 (Fig. 3)26,41. Among all the derivatives, only fluoro or cyano substitution at the ortho position of the benzene ring led to better inhibitory activity, evidenced from 26a and 26e (IC₅₀ = 4.9 and 8.7 μ M, respectively). If the substitution was at the meta or para position, this inhibitory activity was reduced, such as in compounds 26b, 26c and 26f $(IC_{50} = 10, 19 \text{ and } 26 \mu M, \text{ respectively})$. The dual substitution of fluoro also reduced inhibition activity, for which the IC₅₀ value of 26d was 15 µM. However, only 5-pyrimidinyl (26i) had good inhibitory activity among all of the heterocyclic substitutions (26g-26i), where the IC₅₀ value was 3.2 μ M. According to the results shown in Fig. 3, the substituted position of fluoro or cyano plays an important role in the relaxation of preconstricted aortic rings.

In the core structure of YC-1 (heterocycle A), Straub *et al.* studied the impact of different cores on relaxation effects on preconstricted aortic rings (Fig. 4). 26,41 The synthesis of compound 27 followed route b in Scheme 1 and replaced the indazole starting material. The results showed that more nitrogen atoms in the heterocycles contributed to little increases in inhibitory activity (27f, IC₅₀ = 9.4 μ M). The most effective compound was pyrazolopyridinyl pyrimidine (27b), with an IC₅₀ value of 1.7 μ M. Unfortunately, other pyrazolopyridine derivatives exhibited inferior activities, such as 27c, 27d and 27e (IC₅₀ values of 6.9, 9.8, and 20.4 μ M, respectively). Therefore, the position of nitrogen has a close relationship with inhibitory activity. A compound with 1*H*-pyrazolo[4,3-c]pyridine (27b) was found to be more potent than one with indazole (27a) in the core structure.

Relaxation of the preconstricted
$$IC_{50} = 4.9 \mu M$$
 $IC_{50} = 1.7 \mu M$ $IC_{50} = 6.9 \mu M$ $IC_{50} = 9.8 \mu M$ $IC_{50} = 9.4 \mu M$

Fig. 4 $\,$ Effect of YC-1 derivatives (27) on relaxation of preconstricted aortic rings.

Fig. 5 Effect of YC-1 derivatives (28) on relaxation of preconstricted aortic rings.

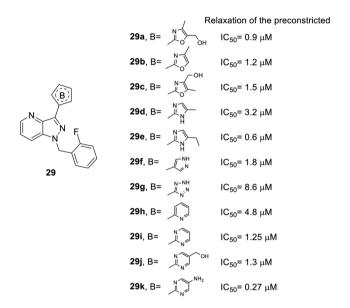


Fig. 6 Effect of heterocycle B-substituted YC-1 derivatives (29) on relaxation of preconstricted aortic rings.

Next, when R^2 was substituted as compound 28, whose synthesis followed Scheme 1, route b, only the hydroxymethyl side chain retained inhibitory activity (Fig. 5). For example, compound 27b with a hydroxymethyl side chain resulted in good inhibitory activity ($IC_{50}=1.7~\mu M$), but the inhibitory activity was reduced ($IC_{50}=9.3~and~8.0~\mu M$, respectively) when the hydroxymethyl group was replaced with hydrogen (28a) or sulfonamide (28b).

As for heterocycle B, the synthesis of the derivatives followed route b in Scheme 1 and replaced compound 12 to other

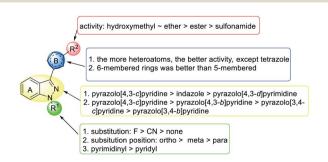


Fig. 7 SAR of YC-1 analogs on relaxation of preconstricted aortic rings.

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Structure of YC-1 (1), BAY 41-2272 (30) and Riociguat (31).

heterocycles. When the furan ring of compound 28 was replaced with isoxazole (29a-c), 1H-pyrazole (29d-e) or other heterocycles (29f-k), better inhibitory effects were observed (Fig. 6). For instance, the isoxazole of compound 29a-c ($IC_{50} = 0.9$, 1.2 and 1.5 μ M, respectively) and 1*H*-pyrazole of compound 29e (IC₅₀ = 0.6 µM) manifested better activity than the furan-substituted functional group (28b, $IC_{50} = 1.7 \mu M$). Furthermore, dinitrogen-containing heterocycles B [imidazole (29e-f) and pyrimidine (29i-k)] increased inhibitory performance (IC₅₀ = 0.6, 1.8, 1.25, 1.3 and 0.27 μM, respectively). However, tetrazolesubstituted 29g significantly reduced this activity ($IC_{50} = 8.6$ μM).

In summary (Fig. 7), in terms of the core structure, the best structure is pyrazolopyridinyl pyrimidine, which further develops into Riociguat. At R¹, the best substituent is the *ortho* fluoro benzyl group. The best substituent for the R² position is a hydroxymethyl or ether group, and the worst is sulfonamide. The presence of pyrimidine ring as A ring as well as R1 substitution gave considerable IC₅₀ values in compounds, 27b, 26i, 29e, and 29k. More nitrogen-substituted rings have shown enhancement in activity for relaxation effects on preconstricted aortic rings. In the ring B position, heterocycles are generally effective except for tetrazole, and 6-membered rings are better than 5-membered rings (Fig. 7).

Scheme 3 Synthesis of YC-1 analogs (42-44). Reagents and conditions: (a) TFA, 1,4-dioxane, reflux, 8 h; (b) TFA, 1,4-dioxane, reflux, 3 days; (c) NH₃/MeOH, rt, 2 days; (d) TFAA/pyridine, pyridine, rt, 8 h; (e) MeONa/MeOH, rt, 2 h; (f) and (i) NH₄Cl/HOAc, MeOH, reflux, 8 h; (ii) Na₂CO₃/H₂O, rt; (g) HOAc, 100 °C, 2 h; (h) toluene, reflux, overnight; (j) t-BuOK, [Pd₂dba₃], (rac)-BINAP, toluene, 70 °C, overnight; (i) neat, 105 °C, slight vacuum, 3-12 h.

			Relax	ation of the preconstricted
	42, R ¹ = N-morpholino;	$R^2=H$;	R ³ =H	IC ₅₀ = 0.93 μM
	43a, R1= Et;	$R^2=NH_2$;	R ³ =H	IC ₅₀ = 0.27 μM
	43b, R1= N-morpholino;	$R^2=NH_2$;	R ³ =H	IC ₅₀ = 0.26 μM
∕~ F	43c, R ¹ = <i>n</i> -propyl;	$R^2=NH_2$;	R ³ =H	IC ₅₀ = 0.25 μM
	43d, R1= n-hexyl;	$R^2=NH_2$;	R ³ =H	IC ₅₀ = 1.04 μM
	43e, R1= t-butyl;	$R^2=NH_2$;	R ³ =H	IC ₅₀ = 0.71 μM
N- N.	43f, R1= cyclopropyl;	$R^2=NH_2$;	R ³ =H	IC ₅₀ = 0.30 μM
N N	43g, R1= cyclopentyl;	$R^2=NH_2$;	R ³ =H	IC_{50} = 0.90 μ M
N	43h, R1= CN;	$R^2=NH_2$;	R ³ =H	IC ₅₀ = 0.39 μM
N	43i, R1= 6-cyanoheptyl;		R ³ =H	IC ₅₀ = 1.29 μM
·->=<	43j, R1= OCH3;	$R^2=NH_2$;	R ³ =H	IC ₅₀ = 0.34 μM
R ³ R¹	43k, R1= F;	$R^2=NH_2$;	R ³ =H	IC ₅₀ = 0.48 μM
	44a , R ¹ = Et;	$R^2 = NH_2$;	$R^3=NH_2$	IC ₅₀ = 0.11 μM
	44b, R1= N-morpholino;		$R^3=NH_2$	IC ₅₀ = 0.10 μM
	44c, R1= cyclopropyl;	$R^2 = NH_2$;	$R^3=NH_2$	IC ₅₀ = 0.23 μM

Fig. 9 Effects of pyrazolopyridinyl pyrimidine derivatives on relaxation of preconstricted aortic rings.

On the other hand, one of the YC-1 derivatives (Bay 41-2272, **30**,) Fig. 8 has been developed into a clinically used drug by Bayer Pharmaceuticals as a new pyrazolopyridinyl pyrimidine derivative (Riociguat, **31**). This drug acts on the NO-sGC-cGMP pathway to effectively dilate blood vessels and inhibit smooth muscle hyperplasia. The synthesis of Riociguat-like, pyrazolopyridinyl pyrimidine derivatives was published in a report in 2009 by Mittendorf *et al.* (Scheme 3).⁴³

Mittendorf et al. started from ethyl cyanopyruvate (33) and 2fluorobenzylhydrazine (32) to generate aminopyrazole (34). The subsequent 3-dimethylaminoacrolein (35) underwent cyclocondensation to give pyrazolopyridine 36 with a 50% overall yield. Then, the ester group of 39 was transformed into an amide using ammonia in methanol and reacted with trifluoroacetic anhydride (TFAA) to give cyano compound 38. Compound 38 underwent a pinner reaction with sodium methoxide, followed by substitution with ammonium chloride to give 40. Condensation of amidine compound 40 and 2-bromomalondialdehyde in acetic acid resulted in 4,6-unsubstituted 41; then, Buchwald-Hartwig reaction was applied to obtain morpholine substituted compound 42. In addition, amidine compound 40 reacted with alkyl-substituted malonic dinitrile under neat conditions to obtain diaminopyrimidines (43) with a yield of 40-70%. The nitrile compounds were transformed into the corresponding enol ether. Condensation of the enol ether and amidine 40 yielded 44. The structure-activity relationship of the pyrazolopyridinyl pyrimidine derivatives (42-44) are discussed later in this work.

The development of Bay 41-2272 and Riociguat were based on the SAR results mentioned above, and sGC stimulator **29k** with pyrazolopyridinyl pyrimidine structure was identified and found to be suitable for further research. The results for additional pyrazolopyridinyl pyrimidine derivatives on the relaxation of preconstricted aortic rings are shown in Fig. 9.⁴³ Overall, pyrazolopyridinyl pyrimidine derivatives (42–44) are more active than YC-1. In most cases, the diamino analogs (44) are slightly more potent than their monoamino (43) counterparts. For example, the inhibitory effect of 44a (IC₅₀ = 0.11 μ M), a diamino analog, is better than that of monoamino compound 43a (IC₅₀ = 0.27 μ M). The same pattern was observed for compounds 43b/44b and 43f/44c. At the R¹ position of 42–44, a smaller steric group led to better inhibitory activity. *n*-Propyl substituted 43c

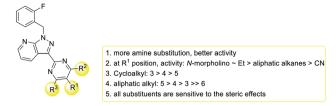


Fig. 10 SAR of pyrazolopyridinyl pyrimidine derivatives on relaxation of preconstricted aortic rings.

(IC₅₀ = 0.25 μM) exhibited better inhibitory activity than *n*-hexyl substituted **43d** (IC₅₀ = 1.04 μM). The bulky *t*-butyl substituted **43e** reduced inhibitory activity (IC₅₀ = 0.71 μM). In addition, cyclic substitution, such as cyclopropyl **43f** (IC₅₀ = 0.30 μM) and cyclopentyl **43g** (IC₅₀ = 0.90 μM) also decreased potency. However, conversion of an alkyl group to cyano substitution (**43h**) slightly reduced the potency (IC₅₀ = 0.39 μM). When installing a linker into **43h** as compound **43i** with a 6-cyanoheptyl substituent, the inhibitory activity was greatly reduced (IC₅₀ = 1.29 μM). Furthermore, the electron-donating groups exhibited better activity than the electron-withdrawing substitution. For instance, methoxy substituted **43j** (IC₅₀ = 0.34 μM) was superior to the fluoro substituted **43k** (IC₅₀ = 0.48 μM).

In summary, the SAR of pyrazolopyridinyl pyrimidine derivatives is shown in Fig. 10. Generally, more amino substituents result in better inhibitory activity. Furthermore, all substituents are sensitive to steric effects. Thus, a smaller cycloalkane or a shorter alkyl led to better activity. At the R¹ position, the best substituents are *N*-morpholino and ethyl, and the worst is cyano substitution.

In addition to the inhibitory activity on preconstricted aortic rings, Lien $\it et~al.$ also studied the effects of the derivatives on SNP-induced apoptosis of rat aortic smooth muscle cells (Fig. 11). 42 YC-1 almost completely reversed SNP-induced apoptosis of VSMCs (cell survival: 94.7%) at 30 $\mu M.^{42}$ Conversion of the hydroxymethyl (YC-1, 1) into ether (45a) at the R^2 position retained the activity. Conversion of the hydroxymethyl to amino (45b) and diethylamino (45c) substitution significantly reduced the activity, while conversion into a methyl group (45d) produced a cytotoxic effect in the VSMCs, resulting in a significant reduction in the anti-apoptotic effects. Finally, replacement of the hydroxymethyl by an ester (45e) only slightly reduced the activity. 26,41,42

Insertion of any atom into the YC-1 indazole structure will reduce the cell survival rate.⁴¹ For example, conversion of the R³

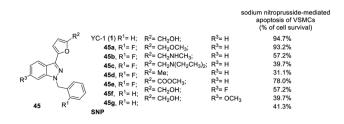


Fig. 11 Anti-apoptotic effect of YC-1 derivatives (45) on the sodium nitroprusside-mediated apoptosis of VSMCs.

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Fig. 12 Effects of YC-1 derivatives (46a-e, 45f-g) on platelet aggregation induced by thrombin, arachidonic acid (AA), collagen, and platelet-activating factor (PAF).

positions with a bridging dioxomethylene group led to even lower activity (cell survival: 39.7%). Introduction of fluoro (45f) or methoxy (45g) groups into the R³ position slightly reduced the potency of the inhibitory VEGF.

3.1.2. YC-1 derivatives as antiplatelet agents. In 2001, Lee et al. reported the SAR of YC-1 analogs on antiplatelet activities, as summarized in Fig. 12 and 13.2

Conversion of the 1-benzyl group of YC-1 to hydrogen significantly reduced the antiplatelet activity. Thus, aromatic rings are needed at the R1 position in a broad spectrum of antiplatelet activities.

Conversion of the alcohol group into an ether (46b) at the R² position resulted in considerably increased activity. In addition, amine-substituted 46d enhanced the inhibitory activity toward thrombin- and PAF-induced aggregation but was inactive against collagen- and AA-induced aggregation. Introduction of an ester resulted in weak inhibitory antiplatelet activities as high as 300 μM. Finally, the acid derivatives were also less potent than their hydroxymethyl counterparts (YC-1, 1). When the furan was converted into a benzene ring, the activity was significantly reduced.

However, introduction of an F atom at the R³ position (45f) dramatically enhanced the inhibition of collagen-, AA- and PAFinduced aggregation but reduced the inhibitory effect on thrombin-induced aggregation. Methyl (46e) and methoxy (45g) substituted derivatives did not alter the platelet aggregation inhibition activity. The derivatives with bridging dioxomethylene groups significantly diminished the activity.

3.2. Hypoxia-inducible factor (HIF)

Hypoxia-inducible factor (HIF) is a transcription factor that induces the expression of various genes to resist tissue hypoxia.4

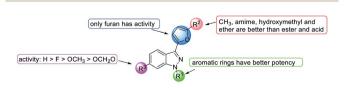


Fig. 13 Effects of the SAR of YC-1 analogs on antiplatelet activities.

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HIF-1 transcriptional activity
YC-1 (1), R1= benzyl
                                                            IC_{50}= 1.20 ± 0.50 \mu M
     47a, R1= Me
                                                             IC<sub>50</sub>> 30.0 μM
     47b, R1= H
                                                            IC_{50} = 5.0 \mu M
     47c, R1= SO<sub>2</sub>Ph
                                                             IC<sub>50</sub>= 16.9 μM
     47d, R1= biphenly
                                                             IC<sub>50</sub>= 3.3 μM
     47e, R1= 4-azidophenyl
                                                             IC<sub>50</sub>= 1.24 \pm 0.34 \mu M
     47f. R1= 2-(trifluoromethyl)benzyl
                                                             IC_{50}= 13.9 \pm 0.39 \mu M
     47a R1= 4-(trifluoromethyl)benzyl
                                                             IC<sub>50</sub>= 9.40 ± 1.76 µM
     47h. R1= 4-cyanobenzyl
                                                             IC_{50}= 0.27 ± 0.01 \muM
     47i. R1= 4-methoxybenzyl
                                                             IC_{50} = 0.97 \pm 0.01 \, \mu M
     26a. R1= 2-fluorobenzyl
                                                             IC<sub>50</sub>= 0.26 ± 0.01 μM
     26b. R1= 3-fluorobenzyl
                                                             IC_{50}= 0.48 \pm 0.09 \muM
                                                             IC<sub>50</sub>= 0.53 ± 0.09 μM
     26c, R1= 4-fluorobenzyl
     47j, R1= 2,4-difluorobenzy
                                                             IC_{50} = 0.31 \pm 0.01 \mu M
     47k, R<sup>1</sup>= 2,6-difluorobenzyl
                                                             IC_{50}= 0.10 \pm 0.01 \muM
     47I, R1= 2,3,5,6-tetrafluorobenzyl
                                                             IC_{50}= 0.07 ± 0.01 \muM
                                                             IC_{50}= 0.18 \pm 0.05 \mu M
     47m.R1= 2.3.4.5.6-pentafluorobenzy
                                                             IC<sub>50</sub>= 0.65 ± 0.06 µM
     47n, R1= 2-chlorobenzyl
                                                             IC_{50}= 2.80 \pm 0.25 \muM
     47o. R1= 4-chlorobenzyl
     26g, R<sup>1</sup>= 2-pyridyl
                                                             IC_{50}= 0.30 ± 0.06 µM
     26h, R1= 3-pyridyl
                                                             IC_{50}= 0.46 \pm 0.03 \mu M
     47p, R1= 4-pyridyl
                                                             IC_{50}= 0.83 ± 0.11 \muM
     47q, R1= 4-azidomethylbenzyl
                                                             IC_{50} = 0.70 \pm 0.08 \mu M
                                                             IC_{50}= 0.61 \pm 0.09 \mu M
     47r, R1= 4-N-(prop-2-yn-1-yl)aminobenzyl
```

Fig. 14 The effect of YC-1 derivatives (47) on HIF-1 transcriptional activity.

Cellular adaptability to hypoxia is an important strategy for tumor survival.44 In many studies, hypoxic conditions have been associated with malignancy in various cancers. Chemicals and drugs targeting HIF-1α have been discovered, and YC-1 is one of them. Moreover, under hypoxia conditions, YC-1 can inhibit HIF-1α activity and achieve anticancer effects.³ As an inhibitor of HIF-1, YC-1 has manifested excellent effects in many different cancer cells,45 including lung cancer,45 bladder cancer,46 breast cancer,6 canine lymphoma,47 chemoresistance,8,9 colorectal cancer, gastric carcinoma, 48,49 hepatocellular carcinoma cancer (HCC),50-53 preeclampsia,54 ovarian cancer,55 and non-small cell lung cancer (NSCLC).50,56-60

In addition, HIF-1α is upregulated in lung parenchyma and alveolar macrophages in asthma and is also viewed as a major regulator of inflammation. YC-1 can inhibit the increase in HIF-1α expression controlling allergen-induced airway diseases in (AIAD) mice.¹⁴ YC-1 can also decrease blood eosinophilia, the aryl hydrocarbon receptor (AHR), and allergic inflammatory gene expression: IL-13, IL-5, myeloperoxidase, and nitric oxide synthase (iNOS).14

However, YC-1 downregulated HIF-1α expression, a key component in osteogenesis, results in significant inhibition of regenerative bone formation. 61-63 In addition, YC-1 partially eliminates the protection ability of FG-4592, which stabilizes HIF-1α expression in spinal cord and PC-12 cells.⁶⁴

Furthermore, under hypoxic/ischemic conditions in the brain, YC-1 can induce the regulatable subunit of HIF-1α, a potent therapeutic target in cerebral ischemia.⁶⁵ In 2011, Yan et al. found that inhibition of HIF-1 expression improves the ischemia-induced blood-brain barrier (BBB) disruption but does not influence brain edema.66 Moreover, YC-1 suppressed the release of neutrophil, inflammatory factor and the activation of the HMGB1/TLR4/NF-NF-κB signaling pathway to protect BBB integrity.⁶⁷ Therefore, YC-1 has a protective effect on the cerebral vascular system. 66,67

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Fig. 15 The effect of YC-1 derivatives (48) on HIF-1 transcriptional activity.

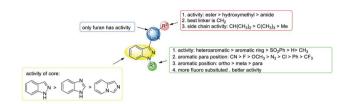


Fig. 16 SAR of YC-1 analogs for HIF-1 transcriptional activity.

HIF- 1α also leads to an imbalance in the expression of the angiogenic protein that is considered to be the primary factor in hypoxic conditions prevalent in preeclampsia. ^{54,68} In 2015, Brownfoot *et al.* discovered that YC-1 significantly decreases soluble fms-like tyrosine kinase-1 (sFlt1), soluble endoglin (sENG) secretion and endothelial dysfunction in primary human tissues, which is a novel drug target for treating preeclampsia. ⁶⁹

The first structure–activity relationship study of YC-1 on HIF- 1α inhibition in 2011 discussed the aromatic ring and the R¹ and R² functional groups of YC-1, ⁷⁰ and in 2012, Takeuchi *et al.* further researched SAR studies of the N-terminal aromatic ring. ²²

In order to test the effects of YC-1 derivatives (47-65) on the inhibition of HIF-1 transcriptional activity, a cell-based HRE reporter gene assay was applied in HRE (X5)-Luc transfected

HeLa cells. The main chemical modifications of the derivatives are located on the different substituents of heterocycles A, B, R¹, and R² (Fig. 2), including different heterocycles, electron donor or withdrawal groups, halide substitution, and other structures, which follow synthetic route a in Scheme 1 to give YC-1 derivatives (47–48). The SAR discussions are summarized in Fig. 14–16 and Schemes 5–8.

Firstly, the activity of pyridyl substitution was better than that of YC-1 (benzyl, 1). Introduction of trifluoromethyl substitution (47f–g) at both the *para* and *ortho* positions resulted in reduced activity. In addition, fluorination was also an effective modification for inhibiting HIF-1 transcriptional activity. Among the monofluoro-substituted compounds (26a–c), *ortho* derivative 26a showed better inhibitory activity than *meta* (26b) and *para* (26c) derivatives. In addition, *ortho*-difluoro-substituted derivative 47k exhibited significant inhibition (IC $_{50} = 0.10 \, \mu$ M) compared with *para*, *ortho*-difluoro-substituted derivative 47j (IC $_{50} = 0.31 \, \mu$ M). However, pentafluoro-substituted derivative 47m also showed marked inhibition but was less potent than 47l. These results indicated that fluoro substitution at the *ortho* position is crucial for inhibiting the transcriptional activity of HIF-1.

At the heterocycle B position, only furan substitution led to inhibitory activity. Other heterocycles, such as isoxazole, oxazole, thiazole, *etc.*, lost their inhibitory activity.

Almost all the R^2 altered substitutions displayed reduced activity, except for the esters. The ester derivatives (48b, 48c and 48f) had better inhibitory ability (IC₅₀: 0.74, 0.89 and 0.9 μ M, respectively).

In addition, from 2014 to 2015, Chen *et al.*⁷¹ and Masoud *et al.*⁷² was based on YC-1 structure molecular modeling and synthesized two series of *N*-substituted indole derivatives 53 and 58 by using either a mixture of 1H-benzo[d]imidazol-2-amine (49) or 2-(azidomethyl)-1H-benzo[d]imidazole (55) and various benzyl bromide (50) in ethanol at room temperature to obtain intermediates 51 and 56. Subsequently, the

HIF-1 transcriptional activity YC-1 (1), IC_{50} = 1.20 \pm 0.50 μM 53a, R¹= CI, R²=4-fluorobenzyl IC₅₀= 1.58 $^\pm$ 0.75 μM 53b, R¹= Cl, R²=4-chlorobenzyl IC $_{50}$ = 2.77 \pm 1.39 μM 53c, R¹= Cl. R²=4-bromobenzyl IC₅₀= -53d, R¹= Br, R²=4-bromobenzyl IC₅₀= $1.78\pm0.75~\mu M$ 53e, R¹= Me, R²=4-bromobenzyl IC₅₀= 53f, R¹= Me, R²=4-(trifluoromethyl)benzyl IC₅₀= -53g, R¹= Me, R²=4-methoxybenzyl IC₅₀= - IC_{50} = 0.60 ± 0.10 μM **53h**, R^1 = H, R^2 =3-methylphenyl

Scheme 4 Synthesis and HIF-1 transcriptional activity effect of YC-1 analogs (53). Reagents and conditions: (a) KOH, EtOH, rt, 18 h; (b) EDCA, HOBt, DMF, rt, 24 h.

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Scheme 5 Synthesis of YC-1 analogs (58). Reagents and conditions: (a) NaN₃, DMSO, rt, 5 h; (b) KOH, EtOH, rt, 18 h; (c) CuSO₄·5H₂O, sodium ascorbate, DCM/water/t-BuOH, rt, 24 h.

intermediates were reacted with substituted acetyl halide (52) or substituted with ethynylbenzene (57) to produce 53 and 58 derivatives, respectively (Schemes 4 and 5).

Among the compounds (53) in this series, the most potent was 53h, with an IC_{50} value of 0.6 μM (Scheme 4). For R^1 substituents, halogen substitution was more potent than alkyl substitution. For example, bromo substituted 53d ($IC_{50}=1.78$ μM) exhibited better inhibitory activity than methyl substituted derivatives (53e-g). In the R^2 position, fluoro-substituted 53a exhibited better inhibitory activity ($IC_{50}=1.58$ μM) than bromoor chloro-substituted derivatives (53b and 53c). Bioisosteric replacement of an amide linker to triazole ring (58) tended to extend conformational flexibility. However, none of the triazole series compounds (58) showed more than a 50% inhibition of HIF-1 α expression. 72

In 2016, Fuse *et al.* synthesized another series of imidazo-pyridine derivatives, **63** and **64**. In this series, 1-iodoimidazo [1,5-a] pyridine (**59**) was used as the starting material, followed by Suzuki reaction to obtain compound **60**. Direct acylation catalyzed by Pd(OAc)₂ was applied for the formation of compounds **61** and **62**. Finally, sodium borohydride reduction was used to achieve final products **63** and **64** (Scheme 6).

In the SAR study (Scheme 6), derivatives **63b–63d** ($IC_{50} = 39$ –75 μ M) at the R¹ position containing electron-donating groups, such as methoxyphenyl and methylphenyl, had similar levels of

63, X= furan HIF-1 transcriptional activity YC-1 (1). IC_{50} = 1.2 ± 0.5 μM 63a, X= furan; R1= phenyl $IC_{50} = 70 \pm 6 \mu M$ 63b, X= furan; R¹= 4-methoxyphenyl IC₅₀= 61 ± 6 μM 63c, X= furan; R1= 2-methoxypheny IC₅₀= 75 ± 5 μM 63d, X= furan; R1= 4-methylphenyl IC_{50} = 39 ± 5 μ M 63e. X= furan: R1= 4-(trifluoromethyl)phenyl IC_{50} = 17 ± 5 μ M **63f**, X= furan; R¹= 3,5-bis(trifluoromethyl)phenyl IC_{50} = 25 ± 12 μ M 63g, X= furan; R1= 4-cyanophenyl IC_{50} = 7.4 ± 0.6 μ M 64, X= thiophene; R1= phenyl

Scheme 6 Synthesis and HIF-1 transcriptional activity effect of YC-1 analogs (63–64). Reagents and conditions: (a) 5 mol% Pd(PPh₃)₄, arylboronic acids, K₂CO₃, 1,4-dioxane/H₂O, 110 °C, microwave, 1 h; (b) 5 mol% Pd(OAc)₂, 5-iodofuran-2-carbaldehyde (for **61**) or 5-bromothiophene-2-carbaldehyde (for **62**), CsOAc, 100 °C, microwave, 1.5 h; (c) NaBH₄, MeOH, rt, 5 min.

inhibitory activity to 63a (IC $_{50}=70~\mu M$). Derivatives 63e–63g containing electron-withdrawing compounds, such as (trifluoromethyl)phenyl and cyanophenyl, exerted higher levels of inhibitory activity. Despite the similar steric bulkiness of the substituents at the R 1 position, the inhibitory activity was 63d (4-methoxyphenyl) < 63e (4-(trifluoromethyl)phenyl) < 63g (4-cyanophenyl) in increasing order. In the case of heterocycle X substitution, thiophene substituted 64 (IC $_{50}=3.7~\mu M$) exerted a better effect than furan substituted 63a (IC $_{50}=70~\mu M$). However, these derivatives were all less potent than YC-1.

In conclusion, in terms of the core derivatives, indazole has better inhibitory activity than benzoimidazole and imidazopyridine. For heterocycle B substitution, only furan has HIF-1 inhibitory activity. Among the R¹ functional groups, the most effective is heterocyclic substitution. Furthermore, the best substitution position is the *ortho* position of the aromatic ring with fluoro substitution. Also, at the R² position, a hydroxymethyl group is a more effective substituent than the hydroxymethyl group. Finally, the most suitable side chain structure is isobutene (Fig. 16).

Recently, Fuse *et al.* synthesized indeno[2,1-*c*]pyrazolone compounds to test their HIF-1 transcriptional inhibitory activity *via* the use of a dual luciferase assay under hypoxic condition, as well as to test their antiproliferative activity using an MTT assay ((3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide)) in HeLa cells under normoxic conditions.⁷⁴

Pyrazole (65) reacted with 66 to give 1-*N*-substituted pyrazole 67. Compound 69 was obtained using nucleophilic addition from 67 and 68, followed by cyclization to give compound 70 (Scheme 7).

Introduction of a nitrogen atom to the X position deteriorated HIF-1 inhibitory activity, for example, as in **70b**. In the R¹ position, the ring activating groups, such as methyl (**70c**), methoxy (**70d**) and hydroxy (**70e**), at the *para* position improved HIF-1 inhibition activity. These indeno[2,1-c]pyrazolones derivatives (**70**) didn't show strong toxicity, with the exception of compound **70e** (cytotoxicity IC₅₀ = 2.2 μ M). Compound **70e** showed impressive HIF-1 inhibitory activity and cytotoxicity under normoxic conditions, suggesting that other biological processes also contribute to antiproliferative effects.

3.3. Vascular endothelial growth factor (VEGF)

Vascular endothelial growth factor (VEGF) is a signaling protein that stimulates blood vessel formation.⁷⁵ Without the nourishment of blood vessels, the size of a tumor mass cannot exceed 2

$$N = 0$$
 $N = 0$
 $N =$

		Cytotoxicity	HIF-1	
YC-1 (1)	IC ₅₀	56 ± 0.93	2.4 ± 0.51	μΜ
70a , X= C, R ¹ = Ph	IC ₅₀	>10	1.4 ± 0.83	μΜ
70b , X= N, R ¹ = Ph	IC ₅₀	52 ± 5.7	10 ± 0.46	μΜ
70c , X= C, R ¹ = 4-MePh	IC ₅₀	>10	0.79 ± 0.16	μΜ
70d , X= C, R ¹ = 4-MeOPh	IC ₅₀	>10	0.61 ± 0.10	μΜ
70e , X= C, R ¹ = 4-OHPh	IC ₅₀	2.2 ± 0.067	0.39 ± 0.088	μΜ
70f , $X = C$, $R^1 = H$	IC ₅₀	>30	>30	μΜ

Scheme 7 Synthesis, cytotoxicity of HeLa cells and HIF-1 transcriptional activity of indeno[2,1-c] pyrazolones (70). Reagents and conditions: (a) Cu₂O, CsCO₃, DMF, 100 °C, 22 h; (b) n-BuLi, THF; (c) (i) K_2 CO₃, ICI, CHCl₃, rt, 16 h; (ii) Pd(PPh₃)₂Cl₂, NaOAc, DMA, 130 °C, 20 h.

mm³; therefore, angiogenesis is essential for tumor progression.^{76,77} Additionally, the exudation process requires vascularization during metastasis.

Since VEGF is overexpressed in various human cancers and is released by a variety of tumor cells, it has been identified as the most important antiangiogenic factor contributing to tumor progression. When a drug can inhibit VEGF production or block its receptor signaling, it also leads to a significant inhibition of tumor growth, such as is the case with YC-1.

YC-1 blocks the induction of erythropoietin (EPO) and VEGF mRNA by inhibiting the hypoxia accumulation of HIF-1 α under hypoxic conditions. According to studies by Yan *et al.*, in experimental central retinal venous occlusion (CRVO) in rhesus monkeys, the macular edema thickness was significantly decreased after YC-1 injection (90 μ l, 200 μ M) at 1 week. Meanwhile, the concentration of IL-6, IL-8, VEGF, the inflammation markers, were significantly decreased in YC-1 injection group. YC-1 showed anti-inflammatory and anti-angiogenesis-related expression and significantly alleviated macular edema. ¹⁶

Temirak *et al.*, Abdullaziz *et al.* synthesized a series of YC-1 analogs 73–78 (Scheme 8) and discussed the SAR with cytotoxicity and VEGF levels. ^{78–80}

1,2-Phenylenediamine (71) was condensed with furfural compound 72 in p-toluenesulfonic acid and DMF to obtain 5-

methyl derivatives 73. Then, compound 73 was reacted with various chloro compounds with K_2CO_3 in acetone at room temperature to give *N*-substituted product 74. Under the same reaction conditions, treatment of 1*H*-benzimidazoles 73 with ethyl bromoacetate can provide *N*-alkylated products 75. Next, compound 76 was obtained from compound 75 with hydrazine hydrate in ethanol. Compound 76 was reacted with potassium hydroxide (KOH) and ethanol to obtain compound 77. Hydrazide compound 76 was cyclized with carbon disulfide in potassium hydroxide and ethanol to obtain oxadiazole ring compound 78 (Scheme 8).

Benzimidazole derivatives (73–78) were tested for cytotoxicity in MCF-7 human breast cancer cells. An ELISA kit was used to measure the level of human VEGF in MCF-7 cells, and the inhibition percentage was calculated based on the level of untreated cells. ^{81,82} The SAR was discussed based on the R¹, R², R³ positions and aromatic substituents, as shown in Fig. 17–19. ^{78–80}

The results showed that steric effect has an extreme impact on the R¹ position. Conversion to methyl at the R¹ position significantly enhanced VEGF inhibition activity. For example, R¹ methyl compounds **74c** significantly enhanced VEGF inhibition (VEGF inhibition was 97%), and VEGF inhibition of hydrogen compounds **74b** and **76b** ranged from 14–23%.

Scheme 8 Synthesis of benzimidazole derivatives (73–78). Reagents and conditions: (a) p-TsOH, DMF, 100 °C; (b) ethyl bromoacetate, K_2CO_3 , acetone, rt; (c) chloro compounds, K_2CO_3 , acetone, rt; (d) NH_2NH_2 , ethanol, 90 °C; (e) EtOAc, KOH, ethanol, reflux; (f) CS_2 , KOH, ice followed by reflux.

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Fig. 17 Effects of the compounds 74-75 on the MCF-7 breast cancer cell line and the VEGF level (pg mL $^{-1}$) in the MCF-7 breast cancer cell line.

Morpholinomethyl substitution 74**f** led to a moderate inhibition effect. At the R² position, the conversion of hydrogen to benzyl significantly increased the activity (74**a**, VEGF inhibition = 98%). The cyanomethyl compound (74**d**) slightly decreased VEGF inhibition activity. Conversion into ethoxy methanoneyl (75) diminished the activity, where VEGF inhibition ranged from 28–62% (Fig. 17).

When the N-terminal functional group was converted to hydrazineyl, benzoylhydrazineyl, benzylhydrazineyl and methylcarbamothioylhydrazineyl substitutions, the derivatives had weak anticancer activity, such as compounds **76a–d**. On the other hand, phenyl at the R³ position (**77a**, VEGF inhibition = 5%) had no VEGF inhibition activity, but phenyls with substitutes were potent functional groups for VEGF inhibition (**77b–d**, VEGF inhibition was above 96%). Conversion to a heterocycle at the R³ position decreased activity, including pyridine (**77e**, VEGF inhibition 58%) and furan (**77f**, VEGF inhibition 30%). Side chains at the R³ position played important roles in inhibitory activity. For instance, methyl side chains of compounds **77g** and **77h** led to worse VEGF inhibition (VEGF inhibition = 31% and 19%, respectively) (Fig. 18).

Finally, replacement of the R² position with a heterocycle dramatically reduced the inhibitory potency of VEGF, where VEGF inhibition was less than 41%, including oxadiazole (78b, 78c, 78d), thiadiazole (78e) and imidazole (78f). However, some

			MCF-7 IC ₅₀ (μg/ml)	VEGF level (Pg/ml)	Inhibition (%)
	76a , R ¹ = H;	R ³ = H	20.20	4010.45	(23%)
	76b, R ¹ = CH ₃		22.40	760.40	(85%)
N O L DI	76c, R ¹ =H;	R ³ = CONHPh	16.00	2300.14	(56%)
н)	76d, R ¹ =H;	R ³ = COPh	22.10	4800.00	(9%)
R ³ ·N·N·	76e, R ¹ =H;	R3= CH2Ph	21.80	4670.23	(11%)
H 0 76	76f, R1=H;	R3= COCH3	18.70	4050.45	(23%)
	76g , R ¹ =H;	R3= CSNHCH3	17.50	2600.40	(51%)
N	77a , R ¹ =H;	R3= CH(Ph)	37.90	5000.40	(5%)
	77b , R ¹ =H;	R^3 = CH(4-CH ₃ Ph)	11.80	196.80	(96%)
N O D	77c, R ¹ =H;	R^3 = CH(4-OCH ₃ Ph)	8.63	196.76	(96%)
н)	77d, R ¹ =H;	R3= CH(4-CIPh)	11.20	184.30	(97%)
R3 N	77e, R ¹ =H;	R ³ = CH(3-pyridine)	15.40	2190.66	(58%)
Ö 77	77f, R ¹ =H;	R ³ = CH(2-furan)	16.60	3700.00	(30%)
	77g , R ¹ =H;	R^3 = CCH ₃ (Ph)	16.00	3600.90	(31%)
	77h, R ¹ =H;	R^3 = CCH ₃ (4-OHPh)	21.80	4230.50	(19%)
	77i, R ¹ =H;	R3= CCH3(4-CH3Ph)	7.80	104.30	(98%)
	DMSO			5250.00	(-)

Fig. 18 Effect of the compounds 76-77 on the breast cancer cell line MCF-7 and the VEGF level (pg mL $^{-1}$) in the breast cancer cell line MCF-7.

N_{R^2}								
	78							
		H₂N,	~	MCF-7 IC ₅₀ (μg/ml)	VEGF level (Pg/ml)	Inhibitior (%)		
	78a , R ¹ = H;	R ² = s≠	N,N	20.50	480.45	(91%)		
	78b , R ¹ = H;	R ² = s=	N H	16.90	3112.78	(41%)		
	78c , R ¹ = CH ₃	₃ ; R²= s≈	, , ,	16.60	3200.80	(39%)		
	78d , R ¹ = H;	R ² = `s	N N N	19.30	4420.70	(16%)		
	78e , R ¹ = H;	R ² = s=	N N	21.30	4195.46	(14%)		
	78f , R ¹ = H;	R ² = нn≈	N N	24.52	3790.38	(22%)		
	78g , R ¹ = H;	R²= ∘≠	S NH	9.39	535.90	(86%)		
	78h , R ¹ = H;	R ² = E10	s kn	11.41	362.80	(92%)		
	78i , R ¹ = H;	R ² = H ₂ NHN	S N	8.29	210.60	(96%)		
	DMSO				5250.00	(-)		

Fig. 19 Effects of the compounds 78 on the MCF-7 breast cancer cell line and the VEGF level (pg mL^{-1}) in the breast cancer MCF-7 cell line.

of heterocycles in the R² position retained inhibitory effects, such as a 91% and 86% inhibition on triazole (78a) and thiazolidine (78g), respectively. In addition, heterocycles with a side chain can improve activity. For example, 2-ethoxy-2-oxoethyl (78h) and 2-hydrazineyl-2-oxoethyl (78i) displayed excellent inhibitory activity on VEGF levels (92% and 96%, respectively).

The SAR of benzimidazole derivatives on VEGF inhibition is summarized in Fig. 20. At the R¹ position, methyl was best for VEGF inhibitory activity. However, the R² position needs to maintain an aromatic ring or heterocycle with a side chain to have good activity.

3.4. Nuclear factor kappa light chain enhancer of activated B cells (NF- κ B)

In 2014, Kuo *et al.* synthesized a series of YC-1 analogs **79** and discussed the SAR with osteoclast inhibition.⁸³ To examine the

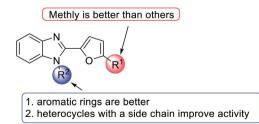


Fig. 20 Effects of the SAR of benzimidazole derivatives on VEGF inhibition.

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Fig. 21 Structure of YC-1 analogs (48, and 79).

effects of the inhibitory activity of each derivative on osteoclast formation, each derivative was added to bone marrow-derived osteoclasts (from male rats) for testing. In the presence of RANKL (50 ng mL⁻¹) and M-CSF (20 ng mL⁻¹), 10 μM of different test compounds were added for 5 days. After 5 days, the formation of osteoclasts was confirmed using tartrateresistant acid phosphatase staining (TRAP staining). Among these derivatives, only 79f, with a furan ring and 2-(dimethylamino)ethyl, inhibited osteoclastogenesis by over 90% (Fig. 21).

4. Conclusions

Since YC-1 was discovered in 1994, there have been discussions on many related physiological activities as well as SAR. Riociguat was even developed based on YC-1 activity. The 83 articles, including 4 patent literatures, selected and discussed in this article confirm its wide use.

In this review, we summarized the synthetic strategies of YC-1 derivatives based on various substituents of YC-1, systematically reviewed their biological activities, including sGC, HIF-1, VEGF, and NF-κB, and described the SAR in detail. Many chemists have modified the compound structures associated with these pharmacological activities to obtain more derivatives with greater biological activity in areas including anticancer, reverse chemoresistance, neovascularization, neural protection, asthma, learning and memory, and bone formation. However, most of the studies were on HIF-1 and sGC. The latest research indicated that YC-1 and NF-κB inhibition are significantly related, but there are few relevant studies. We believe that the inhibition of NF-κB by YC-1 derivatives is worth focusing on.

From a future perspective, to promote the innovation of YC-1 analog drugs, green, combinatorial, and diversity-oriented synthetic methods are essential for developing large-scale analog libraries for high-throughput drug screening and SAR research. In addition, although YC-1 has a wide range of applications, no studies have focused on molecular docking methods. Relevant studies in the future will help to explain the related multipharmacology and will also lead to the discovery of relevant multitarget drugs for clinical use.

Author contributions

Data curation, K.-H. Y., investigation, K.-H. Y., H.-Y. H. Project administration, H.-Y. H.; resources, H.-Y. H.; supervision, H.-Y.

H.; writing – original draft, K.-H. Y.; writing – review and editing, H.-Y. H. All authors have read and agreed to the published version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

Notes and references

- 1 C. C. Wu, F. N. Ko, S. C. Kuo, F. U. Lee and C. M. Teng, *Blood*, 1994, **84**, 4226–4233.
- 2 J. C. Lien, F. Y. Lee, L. J. Huang, T. M. Huang, S. C. Tsai, C. M. Teng, C. C. Wu, F. C. Cheng and S. C. Kuo, *J. Med. Chem.*, 2001, 44, 3746–3749.
- 3 E. J. Yeo, Y. S. Chun, Y. S. Cho, J. Kim, J. C. Lee, M. S. Kim and J. W. Park, *J. Natl. Cancer Inst.*, 2003, **95**, 516–525.
- 4 T. Bollinger, S. Gies, J. Naujoks, L. Feldhoff, A. Bollinger, W. Solbach and J. Rupp, *J. Leukocyte Biol.*, 2014, **96**, 305–312.
- 5 T. C. Hsia, W. H. Liu, W. W. Qiu, J. Luo and M. C. Yin, Molecules, 2014, 19, 19892–19906.
- 6 L. Bergandi, S. Canosa, G. Pittatore, F. Silvagno, S. Doublier, G. Gennarelli and C. Benedetto, *Biol. Reprod.*, 2019, 100, 1521–1535.
- 7 X. Li, J. Xing, H. Wang and E. Yu, *Biosci. Rep.*, 2019, 39(5), BSR20180268.
- 8 W. L. Chen, C. C. Wang, Y. J. Lin, C. P. Wu and C. H. Hsieh, *J. Transl. Med.*, 2015, **13**, 389–402.
- S. Kaowinn, I. R. Cho, J. Moon, J. Soh, H. Y. Kang, C. R. Jung,
 S. Oh, H. Song, S. S. Koh and Y. H. Chung, *Int. J. Oncol.*, 2015,
 46, 2076–2082.
- 10 J. Wu, X. Ke, N. Ma, W. Wang, W. Fu, H. Zhang, M. Zhao, X. Gao, X. Hao and Z. Zhang, *Drug Des.*, *Dev. Ther.*, 2016, 10, 3071–3081.
- 11 X. Ke, J. Wu, W. Wang, H. Zhang, N. Ma, W. Fu, M. Zhao, X. Gao, X. Hao and Z. Zhang, *Int. J. Biol. Sci.*, 2016, **12**, 1363–1371.
- 12 X. Li, N. Lu, R. Tan, J. An, Z. Cai, X. Hu, F. Wang, H. Wang, C. Lu and H. Lu, *J. Mol. Neurosci.*, 2018, **66**, 238–250.
- 13 W. T. Lee, S. H. Tai, A. C. Lee, Y. W. Lin, H. Y. Hung, S. Y. Huang, Y. S. Wu and E. J. Lee, *Mol. Med. Rep.*, 2018, 17, 6490–6496.
- 14 E. McEachern, C. Dewitz, S. Shin, K. Akong, D. G. Nagle, D. H. Broide, P. Akuthota and L. E. Crotty Alexander, *J. Clin. Immunol.*, 2017, 176, 94–99.
- 15 Y. Liang, C. E. Carroll, I. Benakanakere, C. Besch-Williford and S. M. Hyder, *Int. J. Oncol.*, 2013, 4, 179–187.
- 16 J. An, Z. Yan, Q. Shang, N. Zhou and J. Ma, Curr. Eye Res., 2018, 43, 526–533.
- 17 W. S. Lee, M. S. Chang, B. C. Chen, J. R. Sheu and C. H. Lin, *Mol. Pharmacol.*, 2004, **66**, 561–571.
- 18 K. C. Liang, W. L. Chien, C. M. Teng, S. C. Kuo, F. Y. Lee and W. M. Fu, Mol. Pharmacol., 2003, 63, 1322–1328.
- 19 C. B. Yeh, J. W. Wang, S. J. Chou, K. C. Lu, T. H. Chu, W. Y. Chen, J. L. Chien, M. H. Yen, T. H. Chen and J. F. Shyu, *J. Bone Miner. Metab.*, 2018, 36, 508–518.

Review

- 20 Y. S. Chun, E. J. Yeo and J. W. Park, Cancer Lett., 2004, 207, 1-7.
- 21 C. H. Wu, C. H. Pan and M. J. Sheu, Vasc. Biol., 2019, 5, DOI: 10.5772/intechopen.84572.
- 22 M. Hori, A. Takeuchi, S. Sato, H. S. Ban, T. Kuchimaru, S. Kizaka-Kondoh, T. Yamori and H. Nakamura, Med. Chem. Commun., 2012, 3, 1455-1461.
- 23 T. Meng, K. Yuanfang and C. Bingje, China Patent, 106316958A, 2015.
- 24 C. M. Counceller, C. C. Eichman, B. C. Wray, C. Brenda and J. P. Stambuli, Org. Lett., 2008, 10, 1021–1023.
- 25 J. C. Lien, F. Y. Lee, L. J. Huang, T. M. Huang, S. C. Tsai, C. M. Teng, C. C. Wu, F. C. Cheng and S. C. Kuo, J. Med. Chem., 2001, 44, 3746-3749.
- 26 A. Straub, C. Furstner, U. Niewohner, T. Jaetsch, A. Feurer, R. Kast, J. P. Stasch, E. Perzborn, J. Hutter and K. Dembowsky, European Patent, EP0932403A1, 1996.
- 27 V. Collot, P. Dallemagne, B. Patrick, R. Philippe and S. Rault, Tetrahedron, 1999, 55, 6917-6922.
- 28 K. W. Hering, J. D. Artz, W. H. Pearson and M. A. Marletta, Bioorg. Med. Chem. Lett., 2006, 16, 618-621.
- 29 X. Yang, J. Yu, C. Wu and W. Su, J. Org. Chem., 2020, 85, 1009-1021.
- 30 C. M. Teng, F. Y. Lee and S. C. Kuo, US Pat., US 7345079B2,
- 31 M. Katsuno, K. Inamoto, T. Yoshino, L. Suzuki, K. Hiroya and T. Sakamoto, Chem. Lett., 2004, 33(8), 1026-1027.
- 32 K. Inamoto, M. Katsuno, T. Yoshino, Y. Arai, K. Hiroya and T. Sakamoto, Tetrahedron, 2007, 63, 2695-2711.
- 33 O. V. Evgenov and J. P. Stasch, Handb. Exp. Pharmacol., 2013, 218, 279-313.
- 34 S. C. Kuo, F. Y. Lee and C. M. Teng, US Pat, US5574168A, 1996.
- 35 T. Utkan, I. Komsuoglu Celikyurt, C. Ozer, N. Gacar and F. Aricioglu, Med. Sci. Monit. Basic Res., 2014, 20, 130-137.
- 36 K. C. Liang, W. L. Chien, C. M. Teng, S. C. Kuo, F. Y. Lee and W. M. Fu, Eur. J. Neurosci., 2005, 21, 1679-1688.
- 37 K. C. Liang, W. L. Chien and W. M. Fu, Eur. J. Pharmacol., 2008, 590, 233-240.
- 38 D. A. Tulis, Curr. Med. Chem., 2004, 2, 343-359.
- 39 K. J. Peyton, X. M. Liu, N. N. Mendelev, H. Wang, D. A. Tulis and W. Durante, Mol. Pharmacol., 2009, 75, 208-217.
- 40 W. C. Chang, C. H. Wu, G. Y. Chang, S. C. Kuo and C. M. Teng, J. Pharmacol. Sci., 2004, 94, 252-260.
- 41 A. Straub, J. P. Stasch, A. A. Cristina, B. B. Jordi, B. Ducke, A. Feurer and C. Fürstner, Bioorg. Med. Chem. Lett., 2001, **11**, 781-784.
- 42 F. Y. Lee, J. C. Lien, L. J. Huang, S. L. Pan, J. H. Guh, C. M. Teng and C. C. Kuo, J. Med. Chem., 2002, 45, 4947-
- 43 S. Weigand, J. Mittendorf, C. Alonso-Alija, E. Bischoff, A. Feurer, M. Gerisch, A. Kern, D. Knorr, D. Lang, K. Muenter, M. Radtke, H. Schirok, K. H. Schlemmer, E. Stahl, A. Straub, F. Wunder and J. P. Stasch, ChemMedChem, 2009, 4, 853-865.
- 44 P. Vaupel and M. Höckel, J. Natl. Cancer Inst., 2001, 93, 266-276.

- 45 T. Harada, K. Hirose, Y. Wada, M. Sato, K. Ichise, M. Aoki, T. Kato, K. Takeda and Y. Takai, J. Radiat. Res., 2020, 61, 524-534.
- 46 C. L. Shui, C. Liu, Q. Wang, H. Luo and C. G. Gu, Braz. J. Med. Biol. Res., 2017, 51, e6768-e6775.
- 47 M. Igase, S. Kambayashi, K. Kobayashi, A. Kimura, T. Shimokawa Miyama, K. Baba, S. Noguchi, T. Mizuno and M. Okuda, J. Vet. Med. Sci., 2015, 77, 1405-1412.
- 48 R. X. Wang, X. W. Ou, M. E. Kang and J. Q. Shi, Eur. Rev. Med. Pharmacol. Sci., 2018, 22, 8237-8247.
- 49 Y. Kitajima, K. Wakiyama, T. Tanaka, M. Kaneki, K. Yanagihara, S. Aishima, J. Nakamura and H. Noshiro, Sci. Rep., 2017, 7, 12653-12665.
- 50 J. Wu, P. Li, L. Zhao and X. W. Feng, Sleep Breath., 2016, 20,
- 51 J. Kong, X. Ding, W. Xu, S. Dong, Y. Du, C. Yao, J. Gao, S. Ke, S. Wang and W. Sun, Oncol. Lett., 2018, 16, 5230-5236.
- 52 Y. Liu, X. Wan, Y. Zhao, X. Sun, D. Fan and L. Guo, *PLoS One*, 2017, 12, e0184213.
- 53 X. Li, W. Zhao and Z. Li, *Pharmazie*, 2017, 71, 524-529.
- 54 Y. Zhang, H. J. Zhao, X. R. Xia, F. Y. Diao, X. Ma, J. Wang, L. Gao, J. Liu, C. Gao, Y. G. Cui and J. Y. Liu, Clin. Chim. Acta, 2019, 489, 203-211.
- 55 P. Lin, L. Sun, Z. Qin, Y. Liu, L. L. Deng and C. Lu, Exp. Biol. Med., 2015, 240, 1434-1445.
- 56 H. Hu, X. K. Miao, J. Y. Li, X. W. Zhang, J. J. Xu, J. Y. Zhang, T. X. Zhou, M. N. Hu, W. L. Yang and L. Y. Mou, Eur. J. Pharmacol., 2020, 874, 172961-172977.
- 57 J. Sakakibara-Konishi, Y. Ikezawa, H. Mizugaki, S. Oizumi and M. Nishimura, Int. J. Clin. Oncol., 2017, 22, 59-69.
- 58 F. Takahashi, F. Nurwidya, I. Kobayashi, A. Murakami, M. Kato, K. Minakata, T. Nara, M. Hashimoto, S. Yagishita, H. Baskoro, M. Hidayat, N. Shimada and K. Takahashi, Biochem. Biophys. Res. Commun., 2014, 455, 332-338.
- 59 J. Zhou, Q. Jin, X. Xu, F. Huang and W. Xu, Oncol. Lett., 2019, 1, 4034-4043.
- 60 W. Wu, J. Wan, Y. Huang, W. Ge and S. Liu, Oncol. Rep., 2016, 36, 659-668.
- 61 L. Ye, C. Zhou, C. Jiang, J. Bai, Y. Chi and H. Zhang, Tumor Biol., 2015, 36, 9179-9188.
- 62 H. Furusho, H. Sasaki, D. B. Rider, J. M. Dobeck, W. P. Kuo, A. Fujimura, S. Yoganathan, K. Hirai, S. Xu, K. Sasaki and P. Stashenko, J. Endod., 2019, 45, 181-188.
- 63 J. Shao, C. Wan, S. R. Gilbert, R. C. Riddle, F. Long, R. S. Johnson, E. Schipani and T. L. Clemens, Ann. N. Y. Acad. Sci., 2010, 1192, 322-326.
- 64 K. Zhou, K. Wu, Y. Wang, Y. Zhou, N. Tian, Y. Wu, D. Chen, D. Zhang, X. Wang, H. Xu and X. Zhang, Brain Res., 2016, **163**, 19-26.
- 65 Y. Tang, M. Bernaudin, M. Reilly, E. Petit and F. R. Sharp, J. Biol. Chem., 2002, 277, 39728-39738.
- 66 J. Yan, B. Zhou, S. Taheri and H. Shi, PLoS One, 2011, 6, e27798-e27808.
- 67 L. Kong, Y. Ma, Z. Wang, N. Liu, G. Ma, C. Liu, R. Shi and G. Du, Int. Immunopharmacol., 2021, 94, 107507-107519.
- 68 H. M. Brandon, A. Rajakumar, A. Daftary, R. Ness and K. P. Conrad, Placenta, 2004, 25, 763-769.

69 S. Tong, F. C. Brownfoot, N. J. Hannan, R. Hastie, P. Cannon, L. Tuohey and T. J. Kaitu'u-Lino, *Mol. Cell. Endocrinol.*, 2015, **41**, 202–208.

RSC Advances

- 70 N. J. Kim, H. An, J. W. Jung, H. Jang, J. W. Park and Y. G. Suh, *Bioorg. Med. Chem. Lett.*, 2011, 21, 6297–6300.
- 71 J. Wang, J. Chen, P. Schwab, K. T. Park, T. N. Seagroves, L. K. Jennings, D. D. Miller and W. Li, *Anticancer Res.*, 2014, 34, 3891–3904.
- 72 J. Wang, G. N. Masoud, J. Chen, D. Miller and W. Li, *Anticancer Res.*, 2015, 35, 3849–3859.
- 73 S. Fuse, T. Ohuchi, Y. Asawa, S. Sato and H. Nakamura, *Bioorg. Med. Chem. Lett.*, 2016, 26, 5887–5890.
- 74 S. Fuse, K. Suzuki, T. Kuchimaru, T. Kadonosono, H. Ueda, S. Sato, S. Kizaka-Kondoh and H. Nakamura, *Bioorg. Med. Chem.*, 2020, 28, 115207–115213.
- 75 S. J. Galli, D. R. Senger, A. M. Dvorak, C. A. Perruzzi, V. S. Harvey and H. F. Dvorak, *Science*, 1983, 219, 983–985.
- 76 M. O'Reilly, S. Parangi, G. Christofori, L. Holmgren, J. Grosfeld, J. Folkman and D. Hanahan, *Proc. Natl. Acad. Sci. U.S.A.*, 1996, 93, 2002–2007.

- 77 L. Holmgren, M. S. O'Reilly and J. Folkman, *Nat. Med.*, 1995, 1, 149–153.
- 78 A. Temirak, Y. M. Shaker, F. A. F. Ragab, M. M. Ali, S. M. Soliman, J. Mortier, G. Wolber, H. I. Ali and H. I. El Diwani, *Arch. Pharm.*, 2014, 347, 291–304.
- 79 Y. M. Shaker, A. Temirak, F. A. Ragab, M. M. Ali, H. I. Ali and H. I. El Diwani, *Eur. J. Med. Chem.*, 2014, 87, 868–880.
- 80 M. A. Abdullaziz, H. T. Abdel-Mohsen, A. M. El Kerdawy, F. A. F. Ragab, M. M. Ali, S. M. Abu-bakr, A. S. Girgis and H. I. El Diwani, *Eur. J. Med. Chem.*, 2017, **136**, 315–329.
- 81 P. Skehan, R. Storeng, D. Scudiero, A. Monks, J. McMahon, D. Vistica, J. T. Warren, H. Bokesch, S. Kenney and M. R. Boyd, J. Natl. Cancer Inst., 1990, 82, 1107–1112.
- 82 A. Monks, D. Scudiero, P. Skehan, R. Shoemaker, K. Paull, D. Vistica, C. Hose, J. Langley, P. Cronise, A. Vaigro-Wolff, M. Gray-Goodrich, H. Campbell, J. Mayo and M. Boyd, J. Natl. Cancer Inst., 1991, 83, 757–766.
- 83 T. H. Lin, T. H. Kuo, R. S. Yang, S. C. Kuo, W. M. Fu and H. Y. Hung, *J. Med. Chem.*, 2015, **58**, 4954–4963.