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Catalytic, asymmetric azidations at carbonyls: achiral and *meso*-anhydride desymmetrisation affords enantioenriched γ -lactams \dagger

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An unprecedented organocatalytic process involving the asymmetric addition of azide to *meso*-anhydrides has been developed, promoted by novel sulfamide-substituted *Cinchona* alkaloid-based catalysts. Readily available glutaric anhydrides can be smoothly converted to enantioenriched hemi-acyl azides and from there to either γ -amino acids or γ -lactams.

Since the first preparation of phenyl azide by Griess in 1864,¹ the reactivity of organic azides (R-N₃) as [1,3]-dipoles, electrophiles, nucleophiles and radical acceptors have been widely exploited.^{2,3} In addition, the capacity of azide-containing compounds to liberate molecular nitrogen facilitates an array of reaction pathways with the capacity to yield complex products from relatively-simple precursors.

In nucleophilic substitution reactions, azide can either be a useful N1 synthon for the introduction of functional groups (primary amine, amide) or used to install a particular structural motif (1,2,3-triazoles, tetrazoles). Although the high reactivity of azide can be beneficial - with a Mayr nucleophilicity parameter⁴ exceeding that of some α -effect nucleophiles – the utilisation of organic azides in a catalytic asymmetric context is challenging. Both organometallic and organocatalytic approaches to asymmetric azidations have been explored.5 Jacobsen and co-workers⁶ used the privileged 'salen' ligand in the Cr-catalysed silylazidation of meso-epoxides with trimethylsilyl azide (TMSN₃, Fig. 1A) - later expanded to the kinetic resolution of epoxides, the desymmetrisation of meso-aziridines⁸ and the first asymmetric β -azidation of α,β -unsaturated imides with excess hydrazoic acid (HN₃).9 As the intrinsic properties of HN₃ (toxic, volatile and explosive) prevent its practical use, 10 the pursuit of organocatalytic, asymmetric strategies to obviate the direct use of HN₃ has been of interest. 11-16

The first organocatalytic, asymmetric β -azidation, reported by Miller and co-workers in 2000, ¹¹ relied on safer generation of HN₃ *in situ* through the use of TMSN₃ and an organic acid additive (Fig. 1B). Asymmetric β -azidations were further developed thereafter by other research groups, ^{16a} culminating recently in

the first organic acid-free β -azidation of α,β -unsaturated ketones. Recently, we reported the first organocatalytic reactions between cyclic anhydrides and TMSN₃, which allowed controlled access to a variety of pharmaceutically-active γ -amino acid and γ -lactam derivatives from anhydrides. γ -

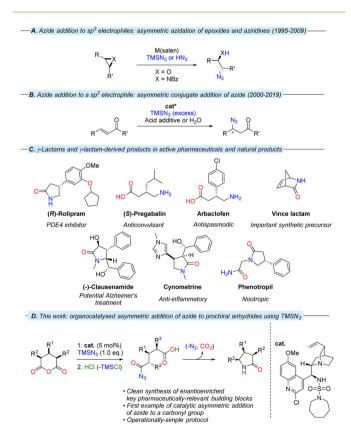


Fig. 1 Asymmetric nucleophilic azidations; the prevalence of chiral γ -lactams in the synthesis of biologically-active compounds and a summary of this work.

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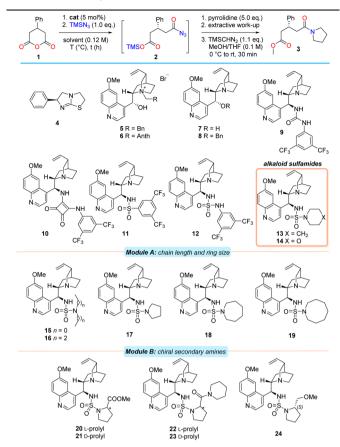
This bioactive class of compounds that contains the γ -aminobutyric acid (GABA) motif are potent central nervous system-active agents (Fig. 1C), $^{18-24}$ and are present in a wide variety of natural products and numerous APIs. The advantage of this reaction cascade lies in rapid access to valuable scaffolds from uncomplicated substrates in a robust manner. In a similar fashion, enantioselective desymmetrisations of achiral or *meso*-anhydrides via existing organocatalytic methodologies (alcoholysis, thiolysis, cycloaddition, *inter alia*) can offer a powerful strategy to access molecular complexity from inexpensive, accessible precursors. $^{25-28}$

Though great advances have been made in the catalytic enantioselective azidations;^{29,30} an analogous catalytic, asymmetric transformation involving the reaction of azide at a carbonyl centre has not yet been reported. In this report, we have expanded upon a study involving a racemic variant of this process¹⁷ and demonstrate the first examples of the enantioselective desymmetrisation of prochiral cyclic anhydrides *via* azidolysis (Fig. 1D).

At the outset, 1 was chosen as the model substrate. After considerable experimentation (see ESI†), suitable reaction conditions were developed in order to facilitate an initial catalyst screen (Table 1). As in the racemic process, ¹⁷ tertiary amines were effective promoters of the silylazidation of 1 with equimolar TMSN₃ to produce the intermediate acyl azide 2 in CHCl₃ at $-50~^{\circ}$ C. ³¹ In order to separate any confounding factors that could alter the enantioselectivity in the desymmetrisation step, the intermediate acyl azide 2 was efficiently quenched with excess pyrrolidine to provide amido ester 3 after desilylation, extraction of the acid and methylation with TMSCHN₂.

Commercial (S)-benzotetramisole³² (4), natural configuration Cinchona alkaloid-based phase-transfer agents 5-6 and bifunctional free-base alkaloid catalysts 7-8 were found to promote the reaction efficiently but with an almost complete absence of enantiocontrol (entries 1-6). Examination of the 9epi-quinine-derived urea, -squaramide and -sulfonamide catalysts 9-11 with superior hydrogen bond donor (HBD) units provided amido acid 3 in only modest ee and curiously, with a preference for the formation of the opposite enantiomer in the case of squaramide 10 (entries 7-9). 16 Incorporation of the sulfamide motif as the HBD into the Cinchona alkaloid scaffold (i.e., alkaloid 12) proved advantageous and provided 3 in marginally improved ee (entry 10). Further modest improvement in enantioselectivity was observed after exchange of the aniline moiety for an aliphatic, secondary amine (i.e., catalyst 13, entry 11). This was somewhat surprising in view of both literature precedent³³ and the loss of the catalyst's ability to participate in efficient bifurcated hydrogen bond donation. However, substitution of the piperidine unit for morpholine did little to influence the enantioselectivity of the process, suggesting that the electronic characteristics at the secondary amine substituent of the sulfamide (i.e., 14) are unimportant (entry 12). After establishing the class of HBD most suitable, further modifications of both the tertiary sulfamide unit and alkaloid core were undertaken.

Table 1 Initial catalyst screen



Entry	Catalyst	t (h)	Conversion ^a (azide 2, %)	ee ^b (%)
1	_	24	<5	
2	4	16	67	-3
3	5	16	99	rac
4	6	16	99	16
5	7	16	90	8
6	8	16	90	rac
7	9	16	81	14
8	10	16	86	-28
9	11	16	72	24
10	12	16	72	34
11	13	16	90	55
12	14	16	90	54
13	15	24	90	47
14	16	18	89	40
15	17	18	90	48
16	18	18	99	58
17	19	18	99	40
18	20	24	80	56
19	21	18	90	33
20	22	24	99	29
21	23	24	90	23
22	24	18	99	57

 $[^]a$ Determined by $^1{\rm H}$ NMR spectroscopic analysis. b Determined by CSP-UHPLC, see ESI.†

A modular catalyst design strategy was adopted (Table 1). Module A examined the effect of either incorporating acyclic amines or modifying amine ring size on enantioselectivity. Module B involved an additional peripheral chirality element.

Module C represents the later combination of the optimal structural features from modules A and B and the finalisation of catalyst development (Table 2).

Alteration of the secondary amine to either an acyclic amine or a reduction in ring size from 6 to 5 resulted in poorer enantioselectivities relative to 13 (i.e., 15-17, entries 13-15). While increasing the heterocycle ring size from 6 to 7 atoms was beneficial (i.e., catalyst 18, entry 16), further expansion to an azocane system resulted in substantially poorer ee (i.e., 19, entry 17).

Evaluation of 20 and 21, prepared from proline methyl ester antipodes, revealed stark differences between the diastereomers with respect to reactivity and selectivity. In the case where the stereochemistry on the prolyl unit matched that at C9 of the alkaloid core (i.e., 'matched' centres), the reaction required marginally extended reaction times, but provided the product in significantly higher ee compared to the 'mismatched' case (entries 18 and 19). The same (albeit less pronounced) effect was also observed in the case of the diastereomeric prolinamide-derived sulfamides 22 and 23 (entries 20 and 21). Separately, evaluation of the 'matched' case of methyl ether 24 provided the amido acid 3 in a slightly more selective process than obtained using 20 (entry 22).

Examination of model systems (see ESI†) based on fragments of catalyst 13 revealed that the quinoline endocyclic nitrogen atom (located far from the catalyst's stereochemical information) could independently participate in the activation of TMSN₃, thereby competing with catalysis at the quinuclidine moiety. This could be obviated in the model system through the installation of a chlorine atom at C2. In a similar vein, methoxy-quinolines were expected to be more active catalysts than quinoline itself.

To test the hypothesis that the quinoline moiety negatively contributes to enantioselective reaction in bifunctional systems, the cinchonidine-derived piperidine sulfamide 27, along with C2'-substituted analogues of 13 and 18 (i.e., 28-29 and 30-31 respectively) were prepared and evaluated (Table 2). Anhydride 1 was subjected to the azidolysis conditions promoted by core-modified sulfamide catalysts 27-31, and the intermediate silvl ester 2 then cleaved with anhydrous HCl to isolate the acyl azide 25. Facile Curtius rearrangement and subsequent lactamisation provided the more potent enantiomer of (R)-phenibut lactam (26).

Gratifyingly, the cinchonidine-derived sulfamide 27 outperformed the analogous quinine-derived catalyst 13 (entry 1). A further increase in selectivity was observed on substitution of the C2' position of the quinoline unit of the catalyst to incorporate either a phenyl group or a chlorine atom (entries 2 and 3). A consistent trend in enantioselectivity was observed upon examination of both the C2'-phenyl azepane sulfamide 30 (entry 4), and its C2'-chloro derivative 31; the latter proved a marginally more selective promoter of the desymmetrisation process (entry 5, 70% ee).

With conditions in hand for the enantioselective azidolysis, a range of cyclic anhydrides 32 were subjected to the azidative desymmetrisation procedure to provide acyl azides 33, promoted by sulfamide 31. These intermediates could be telescoped into Curtius rearrangement and ring-contractive lactamisation steps (vide supra) to provide enantioenriched γ-lactams 34 in one-pot (Table 3).

Both electron-donating and electron-withdrawing substituents on the aromatic ring were well-tolerated; providing access to β-aryl-γ-lactams 26 and 35-40 in uniformly high yields and good ee, most notably arbaclofen lactam (36) and the PDE4 inhibitor rolipram (37). Regarding aliphatic substitution patterns: while methyl and isopropyl substituents were compatible when placed at the 3-position (i.e., lactams 42 and 43), the presence of larger silyl ether and isobutyl groups led to a small

Table 2 Further catalyst development and optimisation

Ph 1. cat (5 mol%) 2. TMSN ₃ (1.0 o CHCl ₃ (0.12 -50 °C, 16 h 3. HCl (1.6 eq.)	eq.) M) HO O	1. CHCl ₃ (0.01 M) 60 °C, 3 h 2. DMAP (5 mol%) 25 °C, 2 h
	Module C: Alkaloid core mod	dification
NH NH O=\$-N	OMe H N N O=S-N O	OMe NH NO = S-N R
27	28 R = C ₆ H ₅ 29 R = CI	30 R = C_6H_5 31 R = CI

Entry	Catalyst	Conversion ^a (azide 25 , %)	25 ,% Yield ^b (%)	
1	27	91	81	61
2	28	>99	90	64
3	29	>99	90	66
4	30	>99	90	65
5	31	>99	91	70

^a Determined by ¹H NMR spectroscopic analysis. ^b Isolated yield. ^c Determined by CSP-UHPLC, see ESI.†

Table 3 Substrate scope

loss in enantioselectivity, although reactivity in the subsequent lactamisation process was maintained (*i.e.*, lactam 41 and pregabalin lactam (44), respectively). Interestingly, comparable enantioselectivities were obtained when conformationally-locked norcamphoric anhydride was examined, providing access to the Vince lactam derivative 45.

As is the case in the analogous sulfonamides,³⁴ *Cinchona* alkaloid-derived sulfamides are found to exist as a pair of two rotational isomers (rotamers) in a *ca.* 2:1 ratio at room temperature on the ¹H NMR spectroscopic timescale, which interconvert by rotation about the C9–C4′ bond axis (Fig. 2). Although it could be postulated that one rotamer could be contributing negatively to stereoselection; variable temperature-NMR spectroscopy of piperidine sulfamide 13 revealed temperature-dependent convergence of rotamer populations, with the major rotamer at room temperature (*i.e.*, rotamer A) present almost exclusively at low temperature when observed *in situ* during catalysis (see ESI†).

Although the isolated catalyst does not display this temperature-dependent behaviour as the free base form, the monoprotic acetate salt of **13** exhibited similar behaviour to that observed *in situ* (Fig. 3). Attempts to isolate the analogous hydrogen azide salt by several methods were unsuccessful. This can be adequately rationalised in the context of similar

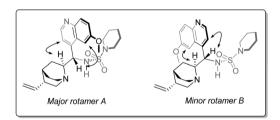


Fig. 2 The two rotamers associated with 13, showing key NOE interactions involved in corroborating *in silico*-derived data.

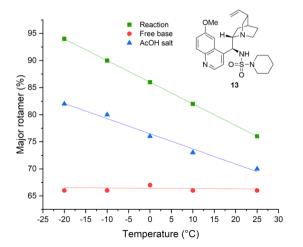


Fig. 3 Temperature-dependent populations of rotameric states of catalyst **13** as the free base, the acetic acid salt and the catalyst observed *in situ* by variable-temperature ¹H NMR spectroscopy.

studies;³⁵ poor room temperature association has been observed in other amine complexes with HN_3 , resulting in its dissociation on irreversible loss of $HN_{3(g)}$ by evaporation. However, as the pK_a (AcOH) $\cong pK_a$ (HN₃) at 25 °C, and given the similarities regarding the temperature-dependent behaviour (with respect to rotamer ratios and ¹H NMR spectroscopic chemical shifts) displayed by the catalyst species *in situ* and the isolated AcOH salt of 13 were found, the evidence suggests that the catalytically-active species in solution is the structurally-related HN_3 complex with 13.

Furthermore, it can be proposed that the free base form of the model sulfamide catalyst **13** is first converted to the active hydrazoate complex **13a** by trapping of adventitious HN₃, which is present in small amounts in commercial samples of TMSN₃ (Fig. 4). This nucleophilic species then facilitates transfer of azide to the anhydride *via* a stereodetermining additionelimination reaction at the prochiral carbonyl centre of **1**. The resulting carboxylate **13b** is then silylated by TMSN₃ to liberate the product **25** and regenerate the active catalyst **13a**.

A DFT conformational analysis exploring the low-energy chemical space associated with **13** was performed. Two predominant conformers differing by the rotation of the C9–C4′ bond were identified (Fig. 5). The Boltzmann population ratio (66:34) predicted by the calculation in CHCl₃ is in good agreement with those obtained from ¹H NMR spectroscopic analysis. In addition, a repeat of the calculations at 223 K yielded a very similar population ratio of 61:39.

A characterisation of the different intramolecular noncovalent interactions was also performed by means of the quantum theory of atoms in molecules (QTAIM) methodology (Fig. 5). An intramolecular hydrogen-bond between the quinuclidine N-atom and the sulfamide unit is a discernible rigidifying feature of both conformations. The other interactions identified appear to be weak in nature.

As is the case in the analogous sulfonamides,³⁴ the *Cinchona* alkaloid-derived sulfamide catalysts exist as a pair of rotamers in a *ca.* 2:1 ratio at 25 °C on the ¹H NMR spectro-

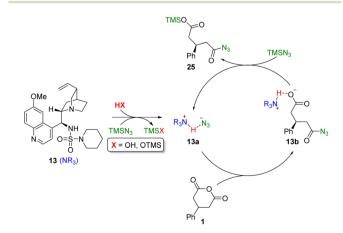


Fig. 4 Proposed catalytic cycle for the desymmetrisation of cyclic anhydrides with equimolar TMSN₃ promoted by *Cinchona* alkaloid sulfamide organocatalysts.

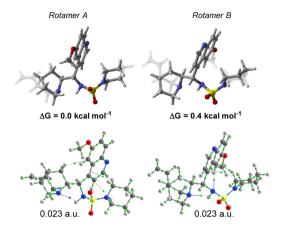
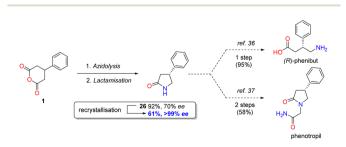


Fig. 5 DFT calculations: structures, relative stabilities and QTAIM analysis of the major rotamers of catalyst 13.

scopic timescale, which interconvert by rotation about the C9-C4' bond axis. Variable temperature-NMR spectroscopy revealed a convergence of rotamer populations, with the major rotamer at room temperature present almost exclusively at low temperature when observed in situ during catalysis (see ESI†). Although the isolated catalyst does not display this temperature-dependent behaviour as the free base form, the AcOH salt of 13 exhibited similar behaviour to that observed in situ. For spectroscopic evidence supporting a hydrazoic acid salt of 13 catalyst resting state, DFT calculations on the catalyst rotamers and a proposed reaction mechanism, see the ESI.†

In a demonstration of the potential synthetic utility of the desymmetrisation, lactam 26 could be prepared on a larger scale under the developed conditions and then converted to enantiopure form in 61% overall yield in a single recrystallisation, which can be transformed to either the anxiolytic phenibut³⁵ or the nootropic phenotropil³⁶ (Scheme 1).

In summary, the first catalytic asymmetric addition of azide to a carbonyl electrophile has been developed. In the presence of novel bifunctional Cinchona alkaloid-derived sulfamide catalysts, prochiral glutaric anhydride derivatives undergo desymmetrisation via addition of TMSN3. The resulting enantioenriched acyl azide derivatives can be readily converted to either γ -amino acid derivatives or a wide range of γ -lactams of considerable medicinal/pharmaceutical interest. studies on the scope, utility and mechanism are underway.



Scheme 1 Recrystallisation of (R)-phenibut lactam 26

Experimental section

NMR spectral data were obtained from a Bruker DPX (400 MHz) or Bruker Avance II (600 MHz) using CDCl₃, DMSO-d₆ or D₂O with chemical shift data referenced relative to residual protic resonances of the deuterated solvent, ($\delta_{\rm H}$ = 7.26, 2.50, and 4.79 ppm respectively). ¹³C (100.9 or 150.9 MHz) spectra were recorded on the same instruments with total proton decoupling. Additional 2D spectral acquisitions (HSQC-ME, HMBC, TOCSY, NOESY/EXSY) were obtained in order to assist in the assignment of resonances where required. Conventional abbreviations for describing peak morphologies in NMR spectroscopic analysis are observed (i.e. s, singlet; br s, broad singlet; d, doublet; dd, doublet of doublets, etc.). All coupling constants (J) are reported in hertz (Hz). Infrared spectra were obtained as neat solids or liquids unless otherwise stated on a Perkin-Elmer Spectrum100 FT-IR instrument fitted with an attenuated-total reflectance (ATR) accessory. Abbreviations used for descriptions of transmission band intensities are as follows: w, weak; m, medium; s, strong; vs, very strong; br., broad.

Thin-layer chromatography (TLC) analyses were performed using Merck-F254 silica gel plates and were visualised under ultraviolet (UV) irradiation, potassium permanganate, ninhydrin, ammonium molybdate or bromocresol green staining methods. Column and flash chromatography was performed using Sigma-Aldrich 60 Å, 230-400 mesh particle silica gel. Melting point data were recorded on a Griffin Melting Point Apparatus; readings were obtained in triplicate and are reported uncorrected. High-resolution mass spectrometry experiments were carried out in the Mass Spectrometry Unit, School of Chemistry, TCD.

Anhydrous CHCl3 (amylene-stabilised) and HCl (as 2 M solution in Et2O) were obtained from Sigma-Aldrich Ireland and transferred to reaction vessels using Schlenk techniques. Hünig's base on polystyrene (DIPEA@PS, product ID: 38343) was purchased from Sigma-Aldrich Ireland and all other chemicals were of regent-grade, obtained from commercial suppliers and used without further purification unless otherwise noted.

Safety considerations

While we have not experienced any issues surrounding the use of TMSN₃ in these studies, it is imperative that the appropriate safety precautions are taken, especially when working on reaction scales >1 mmol. In the following preparations, TMSN₃ has the potential to liberate toxic and explosive HN₃ on contact with H₂O or in acidic media. Any volatiles removed should be carried out in a well-ventilated fume hood and reactions performed with a blast shield in large-scale preparations. It is advised that all azide-containing waste should be quenched cautiously, and in an appropriate manner.³⁷

General procedure A: preparation of 9-epi-9-amino Cinchona alkaloids from native configuration alkaloids

To an oven-dried 250 mL round bottomed flask containing a stirrer bar, Cinchona alkaloid derivative (18.50 mmol) and PPh₃ (5.82 g, 22.19 mmol) under Ar atmosphere, anhydrous THF (125 mL, 0.15 M) was added via syringe. The solution was cooled to 0 °C before DIAD (4.40 mL, 22.19 mmol) and DPPA (4.77 mL, 22.19 mmol), were added sequentially dropwise via syringe. The resulting yellow solution was warmed to room temperature and stirred at 20 °C for 24 h. The flask was fitted with a reflux condenser and the solution stirred at 50 °C for a further 2 h. PPh₃ (5.82 g, 22.19 mmol) was added portionwise with stirring and the solution heated at 50 °C for 2 h or until nitrogen evolution had ceased. H2O (26.4 mL, 0.7 M) was added and the solution stirred at room temperature for 16 h. The resulting mixture was concentrated as far as possible in vacuo and the residue partitioned between 2 M HCl and CH₂Cl₂ (100 mL each). The aqueous phase was removed and the organic layer extracted with 2 M HCl (3 × 50 mL). The combined aqueous extracts were washed with CH₂Cl₂ (5 × 50 mL) and concentrated as far as possible. The viscous residue was stirred in EtOH and the resulting precipitate filtered and dried in vacuo. The precipitate can be purified by reprecipitation from boiling MeOH using EtOAc as antisolvent to give the alkaloid hydrochloride as a powder.

General procedure B: sulfamoyl chloride synthesis

SO₂Cl₂ (1.5 eq.) in anhydrous CH₂Cl₂ (1.00 M) was cooled to -20 °C under Ar atmosphere before a solution of NEt₃ (1.5 eq.) and the appropriate secondary amine (1.0 eq.) in anhydrous CH₂Cl₂ (2 M with respect to amine) was added dropwise via syringe (>30 min, exothermic). The resulting solution was stirred at -20 °C for 30 min before warming to room temperature over 1.5 h. The resulting yellow mixture was slowly poured into ice-H₂O using CH₂Cl₂ to effect the transfer. The biphasic mixture was partitioned and the organic layer washed with H₂O and brine before being dried over anhydrous MgSO₄, filtered and the filtrate concentrated in vacuo. The residue was dissolved in the minimum CH₂Cl₂ and passed through a short plug of silica, eluted with CH2Cl2 to provide the analyticallypure sulfamoyl chloride product after drying in vacuo.

General procedure C: Cinchona alkaloid sulfamide preparation from sulfamoyl chlorides

To a 25 mL round bottomed flask containing a magnetic stirrer bar and the appropriate alkaloid hydrochloride salt (1.00 mmol) was added anhydrous CH₂Cl₂ (5.00 mL, 0.20 M).

To the resulting suspension, NEt₃ (4.20 mmol) was added dropwise at −5 °C and the resulting suspension stirred vigorously for 30 min before sulfamoyl chloride (1.20 mmol) was added dropwise via syringe. The resulting solution was stirred at room temperature for 24-48 h until consumption of the sulfamoyl chloride was observed by TLC analysis. The solution was diluted with CH2Cl2 (25 mL) and washed sequentially with half-saturated NaHCO_{3(aq.)}, H_2O and brine (2 × 10 mL each). The solution was dried over anhydrous MgSO₄, filtered and concentrated in vacuo to give a yellow oil which was purified by flash chromatography as appropriate.

Azepane-1-sulfonyl chloride

Prepared according to general procedure B using azepane (376 µL, 3.33 mmol) and purified by passing through a short plug of silica, eluting with CH₂Cl₂ to give the product as a colourless oil (394.4 mg, 60%). TLC (CH₂Cl₂, ninhydrin): $R_f = 0.83$. δ_H (400 MHz, CDCl₃): 3.46-3.53 (4H, m, H-1), 1.80–1.86 (4H, m, H-2) and 1.63–1.69 (4H, m, H-3) ppm. $\delta_{\rm C}$ (101 MHz, CDCl₃): 50.1 (C-1), 27.5 (C-3) and 27.0 (C-2) ppm. $\nu_{\rm max}$ (neat)/cm⁻¹: 2932 (m), 2860 (m), 1462 (w), 1386 (S=O, s), 1367 (s), 1172 (s), 1144 (m), 1042 (m), 888 (m) and 693 (s) cm⁻¹.

2'-Chloro-9-amino-(9-deoxy)-epi-quininium trihydrochloride (precursor to 31)

Prepared according to general procedure A using C2'-chloroquinine38 (1.39 g, 3.88 mmol) and precipitated after co-evaporation of residual H2O with EtOH to give the product as a bright yellow powder (1.09 g, 80%), m.p. 198-204 °C (decomp.); $[\alpha]_D^{24} = +2.5$ (c = 0.20, H_2O). 1H , ^{13}C NMR and EXSY spectroscopic analyses in DMSO- d_6 revealed rotameric species in the ratio 93:7 at 25 °C. 13C resonances are clearly observable for the major rotamer only. Major rotamer: $\delta_{\rm H}$ (400 MHz, DMSO-d₆): 11.12, 9.48 (3H, br s), 8.15 (1H, s), 7.97 (1H, d, J 9.2), 7.83 (1H, d, J 1.9), 7.58 (1H, dd, J 9.2, 1.9), 5.87-5.96 (1H, m), 5.82 (1H, d, J 10.4), 5.26 (1H, d, J 17.3), 5.16 (1H, d, J 10.5), 4.61-4.68 (1H, app. q), 4.09-4.18 (1H, m), 4.01 (3H, s), 3.70–3.76 (1H, m), 3.28–3.38 (2H, m), 2.76 (1H, br s), 1.80–1.92 (3H, m), 1.57-1.63 (1H, m) and 0.87 (1H, dd, J 13.3, 8.4) ppm. $\delta_{\rm C}$ (151 MHz, DMSO- $d_{\rm 6}$): 158.7, 146.9, 143.6, 141.6, 138.3, 130.4, 126.7, 123.7, 122.0, 116.7, 103.0, 58.7, 56.4, 52.1, 47.7, 41.6, 35.9, 25.5, 23.6 and 23.4 ppm. Minor rotamer: $\delta_{\rm H}$ (400 MHz, DMSO-d₆): 11.12, 9.48 (3H, br s), 8.11 (1H, s), 7.98 (1H, d, J 9.0), 7.54 (1H, dd, J 9.0, 2.0), 7.49 (1H, d, J 2.0), 5.77-5.86 (1H, m), 5.41 (1H, d, J 17.5), 5.22-5.25 (1H, m, J 10.4), 5.16 (1H, d, J 10.5), 4.96 (1H, app. q.), 4.06 (3H, s), 3.93-3.95 (1H, m), 3.70-3.76 (1H, m), 3.28-3.38 (2H, m), 2.76 (1H, br s), 1.99 (1H, br s), 1.80–1.92 (2H, m), 1.21–1.29 (1H, m) and 1.06–1.15 (1H, dd, J 13.3, 8.4) ppm. ν_{max} (neat)/cm⁻¹: 3478 (m, NH st.), 2560 (w), 1617 (s), 1510 (m), 1460 (m), 1395 (m), 1320 (w), 1279 (m), 1235 (s), 1140 (s), 1019 (m), 920 (s), 831 (s), 774 (s), 728 (w) and 681 (s) cm⁻¹. HRMS (APCI⁺) m/z: Found: 358.1685 ($[M + H]^+$ C₂₀H₂₅ClN₃O; requires 358.1680).

Paper

C2'-Chloroquinine azepane sulfamide 31

Prepared according to general procedure C using 2'-chloro-9amino-(9-deoxy)-epi-quininium trihydrochloride (573 1.33 mmol) and azepane-1-sulfonyl chloride (291.3 mg, 1.47 mmol), purified by flash chromatography (7:3 CH₂Cl₂/ EtOAc) to give the title product as a white, crystalline powder (271 mg, 39%), m.p. 58-60 °C. TLC (98:2 $CH_2Cl_2/MeOH$): $R_f =$ 0.44. $[\alpha]_D^{22} = +2.1$ (c = 0.13, CHCl₃). ¹H, ¹³C NMR and EXSY spectroscopic analyses in CDCl3 revealed rotameric species in the ratio 70:30 at 25 °C. Major rotamer: $\delta_{\rm H}$ (600 MHz, CDCl₃): 7.95 (1H, d, J 9.2), 7.56 (1H, s), 7.48 (1H, d, J 2.7), 7.42 (1H, dd, J 9.2, 2.7), 6.12 (1H, br. s), 5.68-5.74 (1H, m), 5.03 (1H), 4.94-4.99 (2H, m), 3.98 (3H, s), 3.19-3.26 (2H, m), 2.75-2.83 (2H, m), 2.55-2.72 (5H, m), 2.28-2.33 (1H, m), 1.57-1.68 (3H, m), 1.37-1.41 (1H, m), 1.27-1.38 (6H, m), 0.86-0.92 (1H, m) ppm. $\delta_{\rm C}$ (151 MHz, CDCl₃): 158.4, 148.5, 148.0, 144.1, 141.0, 130.9, 127.5, 122.7, 121.2, 114.9, 101.5, 61.3, 55.8, 55.75, 53.0, 48.2, 40.4, 39.4, 28.6, 27.9, 27.4, 26.7, 25.2 ppm. Minor rotamer: $\delta_{\rm H}$ (600 MHz, CDCl₃): 7.96 (1H, d, J 9.2), 7.87 (1H, d, J 2.8), 7.40 (1H, dd, J 9.2, 2.8), 7.30 (1H, s), 6.29 (1H, br. s), 5.60-5.66 (1H, m), 4.89-4.95 (2H, m), 4.34 (1H, d, J 10.9), 3.93 (3H, s), 3.35-3.40 (1H, m), 3.19-3.25 (1H, m), 3.05-3.12 (1H, m), 2.72-2.76 (1H, m), 2.55-2.72 (5H, m), 2.28-2.33 (1H, m), 1.73-1.76 (1H, m), 1.58-1.61 (2H, m), 1.27-1.38 (7H, m) and 0.94-0.99 (1H, m) ppm. $\delta_{\rm C}$ (151 MHz, CDCl₃): 157.3, 147.5, 145.1, 144.7, 141.0, 131.0, 125.9, 124.0, 122.5, 114.8, 104.0, 62.6, 56.0, 55.7, 48.3, 40.0, 39.6, 28.6, 27.6, 27.4, 26.6, 26.5 ppm. $\nu_{\rm max}$ (neat)/cm⁻¹: 3189 (w, br, N-H st.), 3073 (w, N-H st.), 2926 (m, C-H st.), 2862 (w), 1620 (s), 1581 (m), 1505 (s), 1455 (s), 1394 (m), 1234 (m), 1228 (m), 1143 (vs, br), 1101 (w), 1044 (w), 1030 (w), 987 (m), 941 (s), 880 (m), 828 (m), 768 (w), 692 (vs), 669 (m), 617 (w) and 576 (s) cm⁻¹. HRMS (APCI⁺) m/z: Found: 519.2195 ($[M + H]^+ C_{26}H_{36}ClN_4O_3S$; requires 519.2192).

General procedure D: organocatalytic, enantioselective synthesis of chiral γ-lactams from prochiral anhydrides

To a 5 mL round bottomed flask containing a magnetic stirrer bar, sulfamide 31 (6.4 mg, 0.012 mmol) and achiral or mesoanhydride 32 (0.246 mmol) under Ar atmosphere, anhydrous CHCl₃ (2.00 mL, 0.12 M) was added via syringe before the solution was cooled to -50 °C for 30 min. TMSN₃ (32.4 μ L, 0.246 mmol) was then added in one portion and the resulting solution stirred at -50 °C for 16 h. HCl in Et₂O (200 μL, 0.400 mmol) was added in one portion and the resulting solution stirred for 15 min at -50 °C. The solution was filtered, using anhydrous CHCl₃ (1 mL) to effect the transfer and volatiles removed expediently in vacuo to give the analytically-pure acyl azide 33. The solid was placed under Ar atmosphere and anhydrous CHCl₃ (25.0 mL, 0.01 M) added via syringe. The resulting solution was heated gently (vigorous gas evolution observed at ca. 40 °C) to 60 °C for 3 h under Ar atmosphere. The solution of isocyanate was then cooled to 25 °C before DMAP (1.5 mg, 0.012 mmol) was added in one portion and the resulting solution stirred vigorously at 25 °C for 2 h. The solution was concentrated *in vacuo* and the residue purified by flash column chromatography to give the γ -lactam product.

(R)-Phenibut lactam (26). Prepared according to general procedure D using anhydride 1 (46.8 mg, 0.246 mmol) and purified by flash column chromatography (98:2 CH₂Cl₂/MeOH) to give the product as a white powder (35.7 mg, 90%, 69% ee), m.p. 75–76 °C (lit.,³⁹ m.p. 73–75 °C). TLC (CH₂Cl₂/MeOH, 98:2): $R_f = 0.42$. A larger scale preparation using anhydride 1 (190.2 mg, 1.00 mmol) afforded the title product by the same method (148.3 mg, 92%, 70% ee) which was recrystallised from hot Hex/EtOAc to provide large, colourless plate crystals (100.6 mg, 62%, >99% ee) with $\left[\alpha\right]_{D}^{22} = -39.6$ (c = 0.91, CHCl₃), (lit., $\alpha_{\rm D}^{40} = -39.4$ (c = 0.90, CHCl₃) for 99% ee of the (R)enantiomer). Spectroscopic data correlates well to that in the literature. CSP-SFC analysis: step 3 was employed with UV detection at 254 nm; R_T : 3.45 min (minor enantiomer) and 3.56 min (major enantiomer). $\delta_{\rm H}$ (400 MHz, CDCl₃): 7.33–7.36 (2H, m), 7.25-7.28 (3H, m), 6.09 (1H, br s), 3.79 (1H, dd, J 9.4, 8.3), 3.71 (1H, app. quin.), 3.43 (1H, dd, J 9.4, 7.3), 2.75 (1H, dd, J 17.0, 9.0) and 2.52 (1H, dd, J 17.0, 8.9) ppm. $\delta_{\rm C}$ (100 MHz, CDCl₃): 177.7, 142.1, 129.1, 127.4, 126.9, 49.7, 40.5 and 38.1 ppm. HRMS (APCI⁺) m/z: Found: 162.0912 ([M + H]⁺; $C_{10}H_{12}NO$ requires: 162.0913).

(*R*)-Tolibut lactam (35). Prepared according to general procedure D using S2 (50.2 mg, 0.246 mmol, see ESI†) and purified by flash column chromatography (98:2 CH₂Cl₂/MeOH) to give the product as a white powder (40.5 mg, 94%, 65% ee), m.p. 110–112 °C (lit., ⁴¹ m.p. 108–110 °C). TLC (EtOAc): $R_{\rm f} = 0.40$. [α]_D²² = -9.6 (c = 0.15, CHCl₃), (lit., ⁴⁰ [α]_D²⁰ = -33.7 (c = 0.95, CHCl₃) for 99% ee). CSP-SFC analysis (see ESI†); $R_{\rm T}$: 5.50 min (minor enantiomer) and 5.88 min (major enantiomer). $\delta_{\rm H}$ (400 MHz, CDCl₃): 7.14 (4H, app. s), 6.71 (1H, br. s), 3.77 (1H, dd, J 9.4, 8.3), 3.61–3.70 (1H, m), 3.40 (1H, dd, J 9.4, 7.4), 2.77 (1H, dd, J 16.9, 8.8), 2.49 (1H, dd, J 16.9, 8.9) and 2.33 (3H, s) ppm. $\delta_{\rm C}$ (100 MHz, CDCl₃): 177.9, 139.0, 136.8, 129.5, 126.7, 49.8, 40.0, 38.2 and 21.0 ppm.

(*R*)-Baclofen lactam (36). Prepared according to general procedure D using S3 (55.3 mg, 0.246 mmol, see ESI†) and purified by flash chromatography (EtOAc) to give the product as a white powder (46.2 mg, 96%, 64% ee), m.p. 110–112 °C (lit., 42 m.p. (from Hex/EtOAc) 108–110 °C). TLC (98:2 CH₂Cl₂/MeOH): $R_{\rm f} = 0.31$. $[\alpha]_{\rm D}^{\rm 22} = -16.5$ (c = 0.15, CHCl₃), (lit., 39 $[\alpha]_{\rm D}^{\rm 20} = -39.0$ (c = 1.00, CHCl₃) for 99% ee). CSP-SFC analysis (see ESI†): $R_{\rm T}$: 3.47 min (major enantiomer) and 3.70 min (minor enantiomer). $\delta_{\rm H}$ (600 MHz, CDCl₃): 7.30–7.32 (2H, m), 7.17–7.20 (2H, app. d.), 6.14 (1H, br. s, H-1), 3.78 (1H, dd, J 9.5, 8.3), 3.65–3.70 (1H, m), 3.38 (1H, dd, J 9.5, 7.1), 2.74 (1H, dd, J 16.9, 9.0) and 2.46 (1H, dd, J 16.9, 8.6) ppm. $\delta_{\rm C}$ (151 MHz, CDCl₃): 177.2, 140.6, 133.0, 129.0, 128.1, 49.3, 39.7 and 37.7 ppm.

(*R*)-Rolipram (37). Prepared according to general procedure D using anhydride S5 (74.9 mg, 0.246 mmol, see ESI†) and the crude residue purified by flash column chromatography (98 : 2 CH₂Cl₂/MeOH) to give the title product as an off-white crystalline powder (64.3 mg, 95%, 70% ee), m.p. 132–133 °C (lit., 43 m.p. 131–133 °C). TLC (98 : 2 CH₂Cl₂/MeOH): $R_f = 0.30$. [a]_D² =

-12.1 (c=0.15, MeOH), (lit., 39 [α] $_{\rm D}^{27}=-33.0$ (c=1.00, MeOH) for 99.3% ee). CSP-SFC analysis: $R_{\rm T}$: 4.01 min (minor enantiomer) and 4.22 min (major enantiomer). $\delta_{\rm H}$ (400 MHz, CDCl₃): 6.83–6.84 (1H, m), 6.76–6.79 (2H, m), 6.06 (1H, br s.), 4.74–4.79 (1H, m), 3.83 (3H, s), 3.75 (1H, dd, J 9.3, 8.2), 3.38 (1H, dd, J 9.3, 7.4), 2.71 (1H, dd, J 16.9, 8.8), 2.47 (1H, dd, J 16.9, 8.9)1.78–1.97 (6H, m) and 1.56–1.66 (2H, m) ppm. $\delta_{\rm C}$ (100 MHz, CDCl₃): 177.6, 149.3, 148.0, 134.6, 118.9, 113.9, 112.3, 80.8, 56.2, 49.8, 40.1, 38.1, 32.9 and 24.1 ppm.

(*R*)-4-(Thiophen-3-yl)pyrrolidin-2-one (38). Prepared according to general procedure D using anhydride S6 (48.3 mg, 0.246 mmol, see ESI†) and the crude residue purified by flash column chromatography (98 : 2 CH₂Cl₂/MeOH) to give the title product as a white crystalline powder (37.4 mg, 91%, 65% ee), m.p. 86–88 °C. TLC (98 : 2 CH₂Cl₂/MeOH): $R_{\rm f} = 0.28$. [α]_D²² = -14.0 (c = 0.15, CHCl₃). CSP-SFC analysis (see ESI†): $R_{\rm T}$: 3.35 min (major enantiomer) and 3.52 min (minor enantiomer). $\delta_{\rm H}$ (400 MHz, CDCl₃): 7.32 (1H, dd, J 5.0, 2.9), 7.02 (1H, dd, J 2.9, 1.3), 6.99 (1H, dd, J 5.0, 1.3), 6.58 (1H, br. s), 3.73–3.82 (2H, m), 3.39–3.45 (1H, m), 2.70–2.77 (1H, m,) and 2.44–2.54 (1H, m, H-2b) ppm. $\delta_{\rm C}$ (100 MHz, CDCl₃): 177.8, 142.7, 126.7, 126.2, 120.4, 49.2, 37.9 and 35.9 ppm.

(*R*)-Fluoribut lactam (39). Prepared according to general procedure D using S4 (51.2 mg, 0.246 mmol, see ESI†) and purified by flash column chromatography (4:1 EtOAc/CH₂Cl₂) to give the product as a white, crystalline powder (39.5 mg, 90%, 66% ee), m.p. 97–99 °C (lit., 44 m.p. 98–99 °C). TLC (1:1 EtOAc/CH₂Cl₂): $R_f = 0.15$. $[\alpha]_D^{22} = -7.8$ (c = 0.15, MeOH), (lit., 45 $[\alpha]_D^{25} = -26.2$ (c = 1.00, MeOH) for 96% ee). CSP-SFC analysis (see ESI†): R_T : 3.01 min (major enantiomer) and 3.14 min (minor enantiomer). δ_H (400 MHz, CDCl₃): 7.18–7.23 (2H, m), 6.99–7.05 (2H, m), 6.70 (1H, br s), 3.77 (1H, dd, J 9.3, 8.3), 6.62–3.71 (1H, m), 3.37 (1H, dd, J 9.3, 7.2), 2.72 (1H, dd, J 16.9, 8.9) and 2.44 (1H, dd, J 16.9, 8.7) ppm. δ_F (376 MHz, CDCl₃): –115.53 (s) ppm. δ_C (100 MHz, CDCl₃): 177.8, 162.0 (d, ${}^1J_{C-F}$ 245.5), 138.0 (d, ${}^4J_{C-F}$ 3.1), 128.4 (d, ${}^2J_{C-F}$ 8.0), 115.8 (d, ${}^3J_{C-F}$ 21.2), 49.8, 39.8 and 38.2 ppm.

(R)-4-(2-Chlorophenyl)pyrrolidine-2-one (40).Prepared according to general procedure D using S7 (55.3 mg, 0.246 mmol, see ESI†) and purified by flash column chromatography (98:2 CH₂Cl₂/MeOH) to give the product as a white, crystalline powder (45.2 mg, 94%, 65% ee), m.p. 112-114 °C (lit., 41 m.p. 112–115 °C). TLC (EtOAc): $R_f = 0.49$. $[\alpha]_D^{22} = -9.4$ (c = 0.10, CHCl₃). CSP-SFC analysis (see ESI†): R_T : 6.77 min (minor enantiomer) and 7.13 min (major enantiomer). $\delta_{\rm H}$ (400 MHz, CDCl₃): 7.39 (1H, dd, J 7.8, 1.4), 7.33 (1H, dd, J 7.7, 1.6), 7.25-7.29 (1H, m), 7.18-7.23 (1H, m), 6.45 (1H, br. s), 4.12-4.20 (1H, m), 3.86 (1H, dd, J 9.7, 8.2), 3.42 (1H, dd, J 9.7, 6.0), 2.79 (1H, dd, J 17.0, 9.1) and 2.53 (1H, dd, J 17.0, 7.3) ppm. $\delta_{\rm C}$ (101 MHz, CDCl₃): 177.5, 139.3, 133.8, 130.0, 128.4, 127.4, 127.2, 48.3, 36.68 and 36.66 (C-3) ppm.

(*S*)-4-((*tert*-Butyldimethylsilyl)oxy)pyrrolidin-2-one (41). Prepared according to general procedure D using S8 (60.0 mg, 0.246 mmol, see ESI†), to give a crude residue, which was purified by flash column chromatography (1:1 Hex/EtOAc) to give the title product as a white powder (42.4 mg, 80%, 56% ee),

m.p. 78–80 °C (lit., 46 m.p. (from PE/EtOAc) 84–86 °C). TLC (1:1 Hex/EtOAc, ninhydrin): $R_{\rm f}=0.19$. [α] $_{\rm D}^{22}=-2.4$ (c=0.15, CHCl $_{\rm 3}$), (lit., 47 [α] $_{\rm D}^{22}=-7.4$ (c=1.30, CHCl $_{\rm 3}$)). CSP-SFC analysis (see ESI †): $R_{\rm T}$: 2.61 min (minor enantiomer) and 2.73 min (major enantiomer). $\delta_{\rm H}$ (400 MHz, CDCl $_{\rm 3}$): 5.98 (1H, br s), 4.53–4.58 (1H, m), 3.58 (1H, dd, J 10.0, 6.0), 3.24 (1H, dd, J 10.0, 3.4), 2.54 (1H, dd, J 17.0, 6.8) and 2.26 (1H, dd, J 17.0, 4.2) ppm. $\delta_{\rm C}$ (100 MHz, CDCl $_{\rm 3}$): 176.2, 68.0, 51.6, 40.5, 25.8, 18.0, –4.7, and –4.8 ppm.

(*S*)-4-Methylpyrrolidin-2-one (42). Prepared according to general procedure D using S9 (31.5 mg, 0.246 mmol, see ESI†) and the crude residue purified by flash column chromatography (98:2 CH₂Cl₂/MeOH) to give the product as a white powder (22.2 mg, 91%, 70% ee), m.p. 54–55 °C (lit., 48 m.p. (from Hex) 53–55 °C). TLC (95:5 CH₂Cl₂/MeOH, KMnO₄): $R_{\rm f}$ = 0.50. $[\alpha]_{\rm D}^{22}$ = -4.0 (c = 0.10, CHCl₃), (lit., 49 $[\alpha]_{\rm D}^{25}$ = -20.3 (c = 1.20, CHCl₃) for 99% ee). CSP-SFC analysis (see ESI†): $R_{\rm T}$: 5.26 min (minor enantiomer) and 5.43 min (major enantiomer). $\delta_{\rm H}$ (400 MHz, CDCl₃): 6.25 (1H, br s), 3.53 (1H, dd, J 9.4, 7.6), 2.99 (1H, dd, J 9.4), 2.51–2.64 (1H, m), 2.48 (1H, dd, J 16.5, 8.5), 1.97 (1H, dd, J 16.5, 7.1) and 1.16 (3H, d, J 6.7) ppm. $\delta_{\rm C}$ (100 MHz, CDCl₃): 178.6, 49.6, 38.5, 29.6 and 19.7 ppm.

(*R*)-4-Isopropylpyrrolidin-2-one (43). Prepared according to general procedure D using S10 (38.4 mg, 0.246 mmol, see ESI†) and the crude residue purified by flash column chromatography (98:2 CH₂Cl₂/MeOH) to give the product as a white powder (29.0 mg, 92%, 70% ee), m.p. 90–92 °C (lit., 50 m.p. 96–97 °C). TLC (97:3 CH₂Cl₂/MeOH, ninhydrin): $R_f = 0.25$. [α]_D²² = +1.8 (c = 0.10, CHCl₃), (lit., 50 [α]_D²⁵ = +16.9 (c = 1.05, CHCl₃) for 99% ee). CSP-SFC analysis (see ESI†): R_T : 5.12 min (minor enantiomer) and 5.30 min (major enantiomer). δ_H (600 MHz, CDCl₃): 5.89 (1H br. s), 3.46 (1H, dd, J 9.3, 8.3), 3.09 (1H, dd, J 9.2, 8.3), 2.39 (1H, dd, J 16.7, 8.7), 2.17–2.26 (1H, m), 2.07 (1H, dd, J 16.7, 9.6), 1.56–1.64 (1H, m), 0.93 (3H, d, J 6.7) and 0.90 (3H, d, J 6.6) ppm. δ_C (151 MHz, CDCl₃): 178.3, 46.2, 42.3, 35.2, 32.5, 20.6 and 20.0 ppm.

(*S*)-Pregabalin lactam (44). Prepared according to general procedure D using S11 (41.8 mg, 0.246 mmol, see ESI†) and purified by flash chromatography (Et₂O) to give the product as a colourless oil (32.6 mg, 94%, 64% ee). TLC (95:5 CH₂Cl₂/MeOH): $R_{\rm f} = 0.8$. [α]_D²² = -0.81 (c = 0.16, CHCl₃), (lit., ⁵¹ [α]_D²⁰ = -2.42 (c = 1.00, CHCl₃) for 99% ee). CSP-SFC analysis (see ESI†): $R_{\rm T}$: 1.88 min (minor enantiomer) and 2.00 min (major enantiomer). $\delta_{\rm H}$ (600 MHz, CDCl₃): 6.28 (1H, br s), 3.47 (1H, dd, J 9.3, 7.9), 2.98 (1H, dd, J 9.3, 7.1), 2.53 (1H, app. sept.), 2.40 (1H, dd, J 16.7, 8.6), 1.97 (1H, dd, J 16.7, 8.5), 1.52–1.61 (1H, m), 1.30–1.37 (2H, m) and 0.89 (6H, app. t, J 6.5) ppm. $\delta_{\rm C}$ (151 MHz, CDCl₃): 178.5, 48.3, 43.9, 37.1, 33.0, 26.2, 22.7 and 22.5 ppm. HRMS (ESI†) m/z: Found: 164.1047 ([M + Na]†; C_8H_{15} NNaO requires: 164.1046).

(15,4R)-2-Azabicyclo[2.2.1]heptan-3-one (45). Prepared according to general procedure D using S12 (34.5 mg, 0.246 mmol, see ESI†) to give a crude residue which was purified by flash column chromatography (98:2 CH₂Cl₂/MeOH) to give title compound as a white powder (21.8 mg, 80%, 72% ee), m.p. 79–81 °C (lit., 52 m.p. (from PrOH) 78–81 °C. TLC

(98:2 CH₂Cl₂/MeOH, ninhydrin): $R_f = 0.26$. $[\alpha]_D^{22} = -48.6$ (c = 0.15, CHCl₃), (lit., 53 [α]²² = -160.0 (c = 1.00, CHCl₃)). CSP-SFC analysis (see ESI†): R_T: 2.67 min (major enantiomer) and 2.78 min (minor enantiomer). $\delta_{\rm H}$ (400 MHz, CDCl₃): 6.02 (1H, br s), 3.86-3.89 (1H, m), 2.71-2.74 (1H, m), 1.77-1.93 (3H, m), 1.54–1.66 (2H, m) and 1.41 (1H, dt, I 9.3, 1.4) ppm. $\delta_{\rm C}$ (100 MHz, CDCl₃): 181.2, 55.4, 45.1, 41.3, 30.2 and 23.7 ppm.

Conflicts of interest

There are no conflicts to declare.

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