



Cite this: *Org. Biomol. Chem.*, 2022, 20, 1150

Received 26th September 2021,
Accepted 21st December 2021

DOI: 10.1039/d1ob01905h

rsc.li/obc

Synthetic and biosynthetic methods for selective cyclisations of 4,5-epoxy alcohols to tetrahydropyrans

James I. Bowen,^{†a} Luoyi Wang,^{†b} Matthew P. Crump^a and Christine L. Willis^{†*a}

Tetrahydropyrans (THPs) are common structural motifs found in natural products and synthetic therapeutic molecules. In Nature these 6-membered oxygen heterocycles are often assembled *via* intramolecular reactions involving either oxy-Michael additions or ring opening of epoxy-alcohols. Indeed, the polyether natural products have been particularly widely studied due to their fascinating structures and important biological properties; these are commonly formed *via endo*-selective epoxide-opening cascades. In this review we outline synthetic approaches for *endo*-selective intramolecular epoxide ring opening (IERO) of 4,5-epoxy-alcohols and their applications in natural product synthesis. In addition, the biosynthesis of THP-containing natural products which utilise IERO reactions are reviewed.

1. Introduction

Tetrahydropyrans (THPs) are common structural features in many classes of natural products and biologically active molecules such as the potent anti-cancer macrolide lactone bryosta-

tin 1, and various marine polycyclic polyethers including the brevetoxins (Fig. 1). Furthermore, these rings are regularly employed as scaffolds in medicinal chemistry programs and indeed they are the sixth most prevalent ring system amongst all FDA approved small molecule drugs.¹

Many methods have been developed for the synthesis of THPs as described in previous informative reviews.^{2–6} Common approaches include Prins cyclisations, hetero-Diels–Alder reactions and ring closing metatheses (Fig. 2).^{7–9} In addition, reviews dedicated to the synthesis of natural products with structures incorporating THP rings have been published.^{10–13} As well as synthetic studies, the biosynthesis of

^aSchool of Chemistry, University of Bristol, Cantock's Close, Bristol, BS8 1TS, UK.
E-mail: chris.willis@bristol.ac.uk

^bCAS Key Laboratory of Microbial Physiological and Metabolic Engineering, State Key Laboratory of Microbial Resources, Institute of Microbiology, Chinese Academy of Sciences, Beijing, China

[†]These authors contributed equally.



James I. Bowen

James graduated from the University of Birmingham in 2018 with a MSci in Chemistry with Industrial Experience, having spent one year working at GSK in Stevenage. In his final year he investigated gold catalysed polycyclisation cascades of ynamides under the supervision of Dr Paul Davies, for which he was awarded the Alfred Bader Prize. He then moved to the University of Bristol to join the Chemical Synthesis CDT and is currently in the final year of his PhD studies with Professors Chris Willis and Matt Crump, investigating the biosynthesis of polyketide natural products.



Luoyi Wang

Luoyi Wang received his PhD in natural products chemistry from Shanghai Institute of Materia Medica, Chinese Academy of Sciences in 2013. After one year of postdoctoral research at Utah State University and Ohio State University in the US, he joined Professor Chris Willis and her group at the University of Bristol as a PDRA within BrisSynBio working in the field of polyketide biosynthesis. In 2020, he moved to Beijing and established his own research group at the Institute of Microbiology, Chinese Academy of Sciences. His research focuses on mechanistic enzymology in natural products biosynthesis and enzyme engineering for biocatalytic applications.



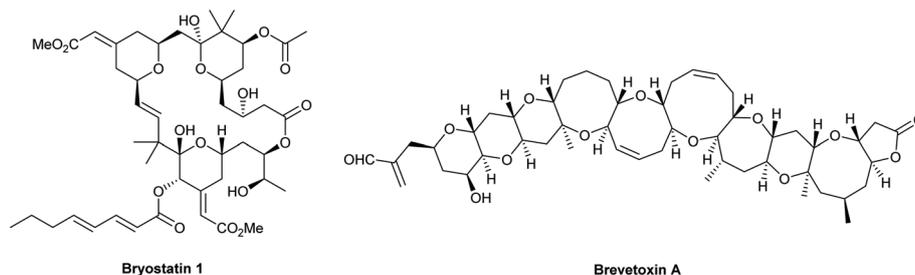


Fig. 1 Bryostatin 1 and brevetoxin A.

THP-containing natural products is of widespread interest and recent work has revealed three main processes found in Nature: oxa-Michael conjugate addition, modification of hemiacetals and intramolecular nucleophilic opening of epoxides (Fig. 2).^{14–16} Indeed, intramolecular epoxide ring opening (IERO) of 4,5-epoxy alcohols has also been widely used in organic synthesis to create oxygen heterocycles giving either 5-membered tetrahydrofurans (THFs) or 6-membered THPs depending on the site of attack of the alcohol onto the epoxide (Fig. 3). This review focuses on the synthesis and biosynthesis of THP rings *via* IERO of 4,5-epoxy alcohols, highlighting how regioselectivity of epoxide ring opening may be achieved.

In the literature the terms *5-exo* and *6-endo* have often been used to describe formation of THFs and THPs respectively from 4,5-epoxy alcohols.¹⁷ This nomenclature is adopted in this review and refers to the process of intramolecular epoxide ring opening by a numerical prefix, describing the size of the ring formed, followed by *exo/endo* to indicate whether the C–C bond of the initial epoxide is located outside or inside the newly formed ring respectively. It is important to note that this nomenclature does not relate to Baldwin's rules, a set of guidelines for predicting the outcome of ring-forming reactions whereby the numerical prefix defines the ring size formed, *exo* and *endo* refer to whether the bond broken during cyclisation

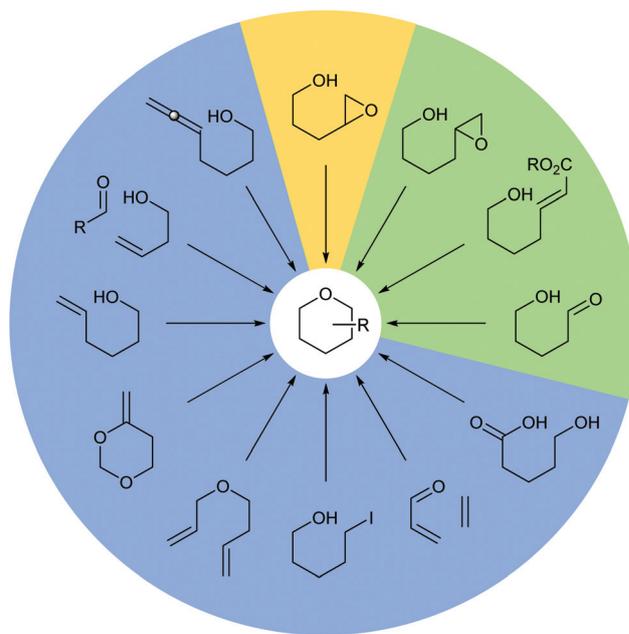


Fig. 2 Common methods for the synthesis and biosynthesis of THP rings. Synthetic methods (blue), synthetic and biosynthetic methods (green) and the focus of this review (yellow).



Matthew P. Crump

Matt Crump is Professor of NMR and Structural Biology at the University of Bristol where he established biological NMR across a broad range of chemical and biological research areas. His major interests are on structural and mechanistic aspects of natural product biosynthesis with a focus on NMR and X-ray crystallography of natural product enzymes and their interactions with biosynthetic intermediates.



Christine L. Willis

Chris Willis is Professor of Organic Chemistry and Head of Organic and Biological Chemistry at the University of Bristol. Her research focuses on natural product biosynthesis including the application of total synthesis, isotopic labelling, pathway engineering and mechanistic studies to produce biocatalysts and new bioactive molecules. She was the recipient of the Natural Product Chemistry Award of the Royal Society of Chemistry in 2020.



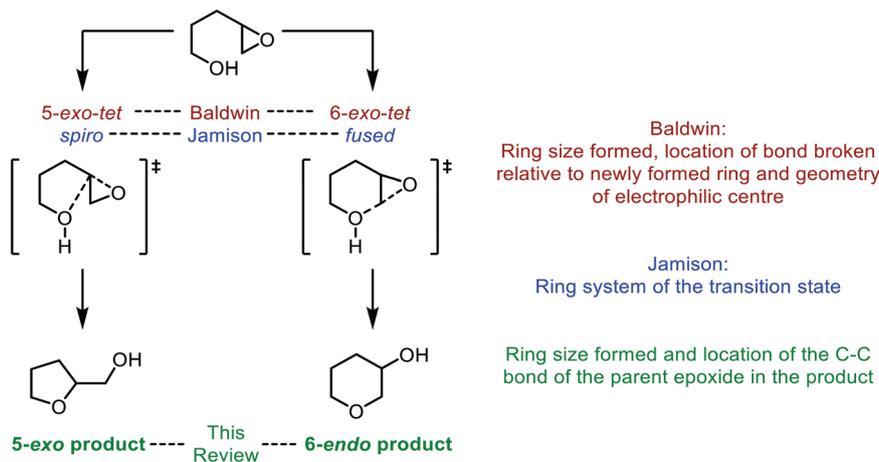


Fig. 3 Intramolecular epoxide ring opening of 4,5-epoxy alcohols and corresponding nomenclature as described by Baldwin (red), Jamison (blue) and in this review (green).

is inside or outside of the newly formed ring, and the suffix indicates the geometry of the electrophile. Using these rules both modes of cyclisation of 4,5-epoxy alcohols are *exo* processes as the C–O bond broken during ring formation is outside of the newly formed heterocycle (Fig. 3).^{18,19} According to Baldwin's rules, both 5-*exo-tet* and 6-*exo-tet* are favoured processes, whilst 6-*endo-tet* cyclisations are disfavoured. Jamison *et al.* have suggested an alternative nomenclature which refers to the transition state of the cyclisation as either spiro or fused for THF and THP formation respectively (Fig. 3).⁵

Whilst both THF and THP rings can be generated from 4,5-epoxy alcohols, the THF is often the major product as noted by Baldwin.¹⁷ This selectivity originates from improved orbital overlap and a lower overall energetic barrier for formation in a 5-*exo* process.²⁰ Furthermore, formation of 5-membered rings is often preferred kinetically over the corresponding 6-membered rings. Therefore, to form THP rings through intramolecular epoxide ring opening of 4,5-epoxy alcohols, this inherent preference to form the 5-membered ring needs to be overcome. Many synthetic methods have been developed to favour THP formation using substrates and reaction conditions which either stabilise the 6-*endo* transition state or destabilise the 5-*exo* transition state. To achieve selectivity in natural product biosynthesis, enzymes are often used. In this review, we will outline synthetic strategies which have been developed to favour THP formation alongside examples of their use in total synthesis. This topic will be divided into substrate controlled and reagent controlled cyclisations with examples of their use in natural product synthesis. Furthermore, examples of THP and THF formation in natural product biosynthesis will be described, highlighting how nature employs enzymes to promote and control IERO.

2. Synthetic methods

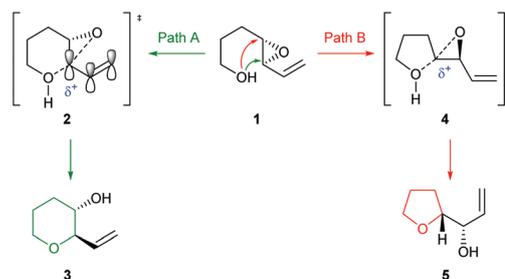
Vilotijevic and Jamison have published reviews on the use of epoxide-opening cascades to create ladder polyethers and oxas-

qualenoid natural products.^{5,21} In this section of our review, we build upon these publications providing an overview of synthetic approaches to achieve 6-*endo*-selective epoxide ring opening of 4,5-epoxy alcohols alongside their use in natural product synthesis.

2.1 Substrate controlled formation of THPs from 4,5-epoxy alcohols

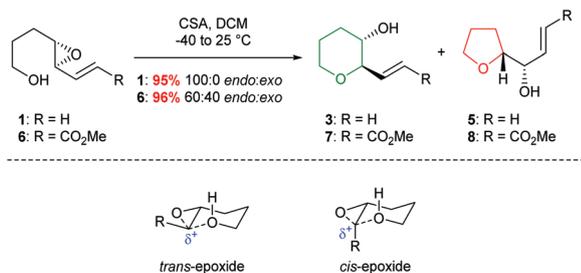
2.1.1 Alkenic epoxides. Due to widespread interest in marine polycyclic polyether natural products, such as the ladder polyether gambierol and the oxasqualenoid thyriferol, many groups have developed methods to achieve selective 6-*endo* ring closure of 4,5-epoxy alcohols. In 1985 Nicolaou reported studies towards the total synthesis of brevetoxin B in which selective cyclisation could be achieved by stabilisation of the incipient positive charge in the 6-*endo* transition state through the incorporation of a π -system adjacent to the epoxide (Scheme 1).²²

Indeed, acid catalysed cyclisation of unsaturated epoxide **1** proceeded with complete regioselectivity, affording 6-*endo* product **3** in excellent yield (Scheme 2).²³ Various π -systems have been investigated including, vinyl halides and unsaturated esters, where the greater electron withdrawing nature of the conjugated functional group decreases the observed



Scheme 1 Stabilisation of 6-*endo* transition state by conjugation.





Scheme 2 Acid catalysed cyclisation of unsaturated epoxides **1** and **6** (top) and transition states for the cyclisation of *cis* and *trans*-epoxides (bottom).

selectivity. Whilst *trans*-epoxides often give good yields of THPs under these conditions, poor regioselectivity was obtained with the corresponding *cis*-epoxides. Nicolaou proposed that *cis*-epoxides are unable to adopt the required planar geometry to achieve stabilisation in the transition state. Indeed, earlier studies by Coxon *et al.* illustrated that cyclisation of *cis*-4,5-epoxy alcohols gave preferential 5-*exo* ring closure compared to the corresponding *trans*-epoxides.²⁴ The 6-*endo* transition state for *cis*-epoxides requires a group to be placed axially, resulting in steric clashes and an overall increased barrier for THP formation (Scheme 2).

Unsaturated 4,5-epoxy alcohols have been widely used as intermediates in natural product synthesis.^{25,26} Examples include the preparation of mucocin, amphidinol 3 and the formation of the E ring in maitotoxin, as well as the botcinins, and more recently in the total synthesis of meayamycin B, illustrating the widespread value of the method (Scheme 3).^{27–32}

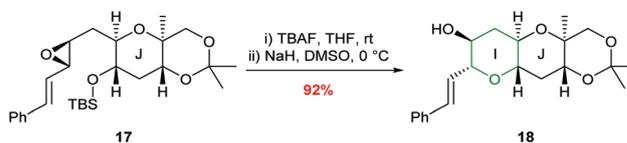
In 1997, Nakata and co-workers reported that introduction of a styryl group adjacent to the epoxide provided greater stabilisation in the 6-*endo* transition state compared with the alkenic epoxides.^{33,34} Treatment of *trans*-epoxides containing a styryl group with either acid or base generated THP products in excellent yield, but improved stereocontrol was achieved using NaH (entries 1 and 2, Table 1). Whilst Nicolaou reported that unsaturated *cis*-epoxides gave poor 6-*endo* selectivity under acidic conditions,²³ Nakata showed that styryl epoxides may be converted solely to THP products under acidic conditions, with no evidence for the formation of the corresponding THF (entries 3 and 4, Table 1). In contrast, under basic conditions *cis*-epoxides gave a mixture of 5- and 6-membered rings (entry 5). Isomerisation of the styryl double bond may occur in acid but in general this is not a problem as this

Table 1 Cyclisation of hydroxy styryl epoxides³³

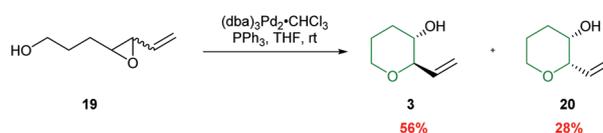
Entry	Epoxide	Conditions	Yield	14 : 15 : 16
1	12	CSA (0.2 eq.), DCM, -78 °C, 1 h	100%	90 : 10 : 0
2	12	NaH (10 eq.), DMSO, rt, 2.5 h	97%	100 : 0 : 0
3	13	CSA (0.2 eq.), DCM, rt, 10 min	85%	30 : 70 : 0
4	13	AcOH-H ₂ O (10 : 1), rt, 19 h	62%	15 : 85 : 0
5	13	NaH (20 eq.), DMSO, rt, 19 h	77%	0 : 66 : 34

directing group is often removed after cyclisation. Hence, although both *cis* and *trans*-epoxides with a styryl group can be cleanly cyclised, the need to remove the styryl directing group can be a disadvantage and as such, these substrates are only employed when the stabilisation of alkenic epoxides is insufficient to achieve the desired selectivity. For example, Nakata and co-workers used styryl-containing epoxides in their synthesis of the IJK ring system of brevetoxin B and the total synthesis of hemibrevetoxin B (Scheme 4).^{35,36}

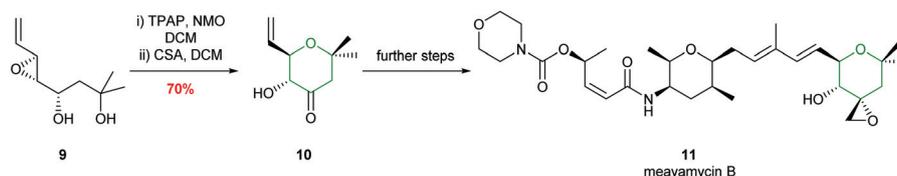
Both palladium and rhodium catalysts have been successfully employed in selective 6-*endo* ring closures of 4,5-epoxy alcohols. Pioneering work by Trost demonstrated that activation of vinyl epoxides could lead to exclusively the 6-*endo* products, albeit with poor stereoselectivity (Scheme 5).³⁷ The reaction proceeds *via* initial formation of a π -allylpalladium intermediate, which in turn is trapped by the appended alcohol. The formation of the π -allylpalladium intermediate ensures only the THP is formed.



Scheme 4 Synthesis of the I ring of brevetoxin B.³⁵

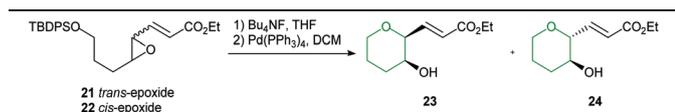


Scheme 5 Palladium catalysed cyclisation of 4,5-epoxy alcohols.³⁷



Scheme 3 Total synthesis of meayamycin B by Koide *et al.*³²



Table 2 Improved palladium catalysed cyclisation of 4,5-epoxy alcohols³⁸

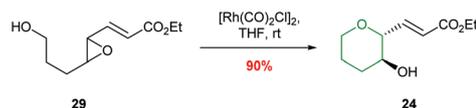
Entry	Substrate	Reaction time	Yield	23 : 24
1	21	5 min	90%	>99 : 1
2	22	6 min	86%	2 : 98

Through modification of the substrate, Hirama and co-workers converted both *cis* and *trans*-epoxides to the corresponding *anti* and *syn*-products in excellent yields and stereoselectivities (Table 2).³⁸ *In situ* formation of an ammonium alkoxide nucleophile alongside the use of an α,β -unsaturated ester to replace the alkene were vital for achieving the desired transformation.

Hirama employed a palladium-mediated cyclisation in the synthesis of the HIJ ring system of ciguatoxin and the AB ring fragment of gambiertoxin 4B (Scheme 6).^{39,40} The key cyclisations proceeded smoothly in 93% and 74% yields respectively to afford single diastereoisomers.

Ha and co-workers expanded the scope of transition metals by using rhodium catalysis to promote selective THP formation (Scheme 7).⁴¹ Stirring *trans*-epoxide **29** with $[\text{Rh}(\text{CO})_2\text{Cl}]_2$ in THF at room temperature gave *anti*-THP **24** with overall retention of stereochemistry *via* a double inversion mechanism. Interestingly, *cis*-epoxides were unreactive under these conditions. This was proposed to be due to a steric clash preventing the formation of the required π -allylrhodium species.

The value of this methodology was demonstrated during the synthesis of the ABCDE ring fragment of ciguatoxin C. Hirama reported that cyclisation of acetal **30** under acidic conditions led to some deprotection of the acetal resulting in the required product **31** being isolated in only 41% yield (Scheme 8).⁴² In contrast using mild rhodium catalysed conditions, the required product **31** was isolated in 84% yield. Furthermore, in 2015 Jamison reported the creation of the B and C rings of (-)-brevisin through a cascade process under

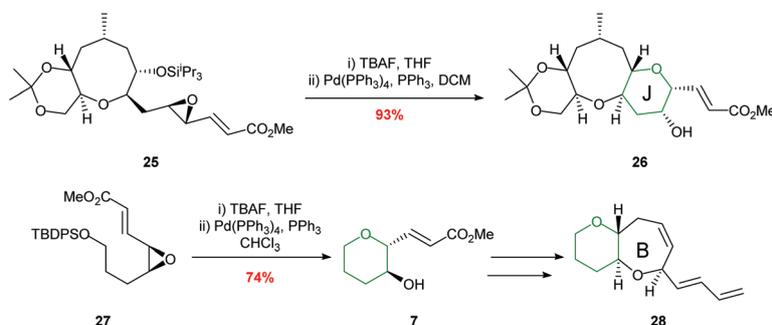
**Scheme 7** Rhodium catalysed intramolecular epoxide ring opening.⁴¹

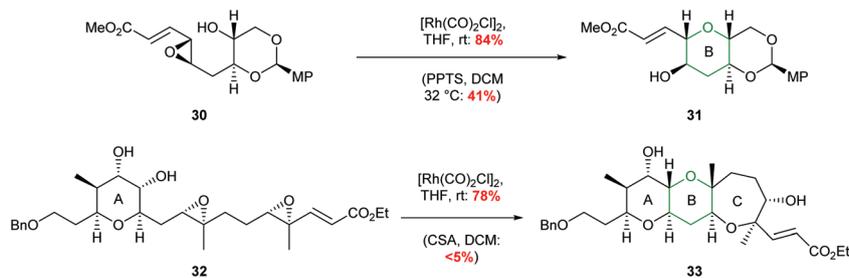
rhodium catalysis, whereas acidic conditions gave the required product **33** in low yield (Scheme 8).⁴³

2.1.2 Acetylenic epoxides. Another class of substrates which have been used to control cyclisation through cationic stabilisation are acetylenic epoxides. Hanaoka and co-workers reported that complexing acetylenic epoxide **34** with dicobalt octacarbonyl, followed by treatment with $\text{BF}_3 \cdot \text{Et}_2\text{O}$ afforded exclusively THP products (Scheme 9).⁴⁴ The alkynes can subsequently be restored by reaction with cerium ammonium nitrate (CAN). This adaptation of the Nicholas reaction proceeds with retention of configuration at both epoxide carbons *via* a double inversion of the propargylic centre (Scheme 10).⁴⁵ Both *cis* and *trans*-epoxides gave THPs in excellent yields and stereoselectivity, affording *anti* and *syn* products respectively. A variety of substituents on the alkyne can be used including silyl, alkyl, and aryl. It was proposed that the 6-*endo* product (THP) is formed in preference to the 5-*exo* product (THF) due to neighbouring group assistance of the cobalt complex (Scheme 10).

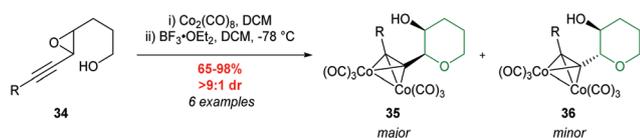
Hanaoka and co-workers adopted this chemistry to complete the first total synthesis of (-)-ichthyothereol, where cobalt complexation followed by 6-membered ring formation proceeded in 87% yield and with excellent stereocontrol (Scheme 11).⁴⁶

The use of acetylenic epoxides as substrates for the synthesis of oxygen heterocycles was further extended by Hanaoka and co-workers (Table 3).⁴⁷ Both *cis* and *trans*-acetylenic epoxides with electron donating substituents on the alkyne were cleanly converted to the corresponding THPs through activation with $\text{BF}_3 \cdot \text{Et}_2\text{O}$, in the absence of the cobalt complex (entries C and D, Table 3). This is analogous to the results of Nicolaou (Scheme 2) whereby inversion of stereochemistry at the propargylic position is observed.²³ In contrast, substrates containing electron withdrawing or electron neutral groups on the alkyne gave predominantly the 5-*exo* products.

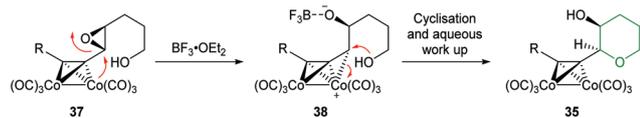
**Scheme 6** Synthesis of the J ring of ciguatoxin (top) and B ring of gambiertoxin 4B (bottom).^{39,40}



Scheme 8 Synthesis of B ring of ciguatoxin C (top) and BC rings of (-)-brevisin (bottom). MP = 4-methoxyphenyl.^{42,43}



Scheme 9 Cyclisation of racemic *trans*-acetylenic epoxides.⁴⁴



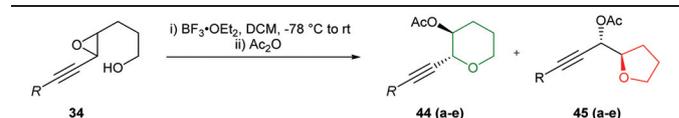
Scheme 10 Mechanism for regioselective cyclisation of 4,5-epoxy alcohols 37.⁴⁵

Forsyth showcased the value of this procedure in 2000 during the total synthesis of thysiferol (Scheme 12).⁴⁸ Cyclisation of trisubstituted epoxide 46 with $\text{BF}_3 \cdot \text{Et}_2\text{O}$ gave THP 47 in 91% isolated yield and with complete regiocontrol.

In further studies, Hanaoka *et al.* illustrated that trisubstituted epoxides with an additional methyl group at either end of the epoxide (*i.e.* propargylic or homopropargylic positions) also led to 6-*endo* cyclisation using either the cobalt or Lewis acid mediated protocols.⁴⁹ However, poor stereoselectivity was often observed for these classes of substrates using either procedure.

In 2004, the synthesis of THPs using an intramolecular Nicholas reaction was reported by Martin *et al.* (Scheme 13).⁵⁰ Activation of a dicobalt complexed propargylic alcohol 49 with

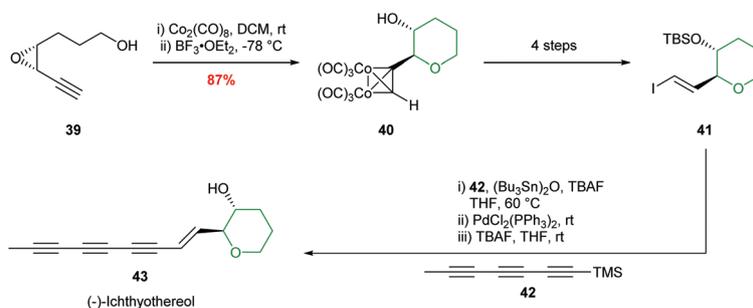
Table 3 Alternative procedure for cyclisation of acetylenic epoxides 34⁴⁷



Entry	R	Yield	44 : 45
A	H	92%	10 : 90
B	TMS	91%	62 : 38
C	<i>n</i> Bu	96%	95 : 5
D	Ph	94%	100 : 0
E	Bz	96%	1 : 99

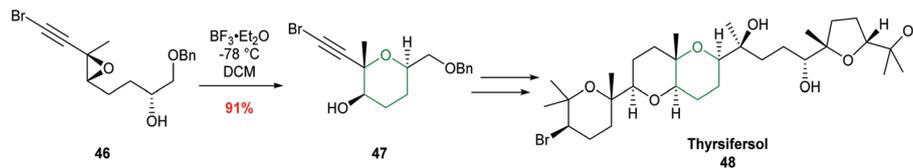
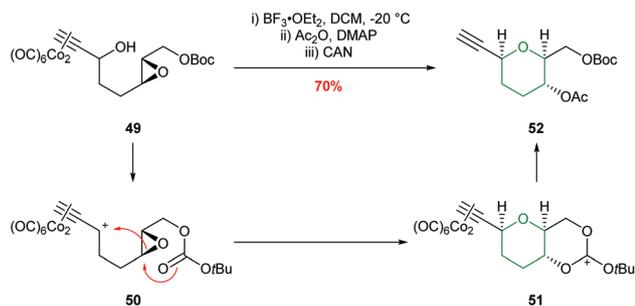
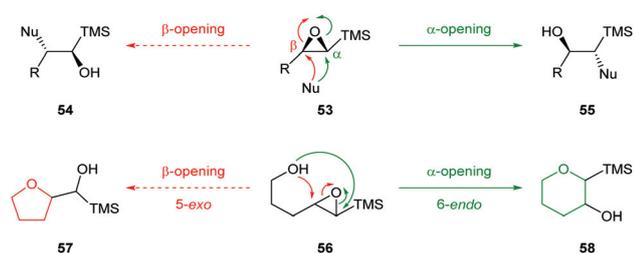
$\text{BF}_3 \cdot \text{Et}_2\text{O}$ generates carbocation 50 which is trapped by the oxirane oxygen. Although this process does not involve intramolecular epoxide ring opening of an acetylenic epoxide, but instead reversed reactivity of an epoxide attacking the propargylic center, the formal 6-*endo* product is still formed.

2.1.3 Epoxysilanes. Epoxysilanes 53 are readily synthesised by epoxidation of vinyl silanes and undergo nucleophilic ring opening α to silicon (Scheme 14).⁵¹ This is in contrast to classical reactivity where silicon directs nucleophilic attack to the β -position due to stabilisation of the resultant positive charge through hyperconjugation. Computational studies by Paquette and Tomoda indicated the origin of α -opening is weakening of the $\sigma_{\text{C-O}}$ bond α to silicon.^{52,53} The groups of Schaumann and Jamison have investigated the use silicon to direct intramolecular epoxide ring opening of 4,5-epoxy alcohols (Scheme 14).

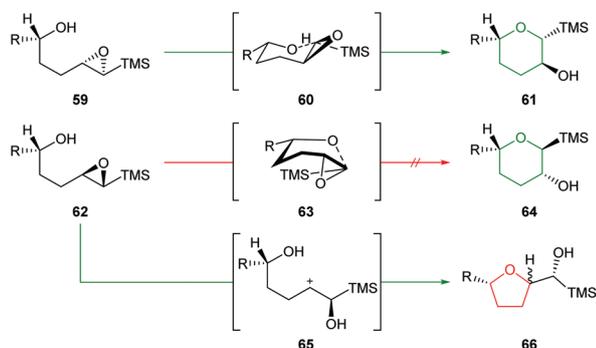


Scheme 11 Total synthesis of (-)-ichthyothereol 43.⁴⁶



Scheme 12 Total synthesis of thyriferol by Forsyth.⁴⁸Scheme 13 Intramolecular Nicholas reaction developed by Martín.⁵⁰Scheme 14 Inter and intramolecular nucleophilic ring opening of epoxy silanes **53** and **56**.⁵¹

Initially, Schaumann *et al.* conducted acid catalysed cyclisations of *trans*-4,5-epoxy alcohols which showed that 1,4-*anti* diastereoisomers **59** afforded the 6-*endo* products **61** whereas 1,4-*syn* diastereoisomers **62** yielded two epimeric 5-*exo* products **66** (Scheme 15).⁵⁴ These results were rationalised by the ability of the 1,4-*anti* diastereoisomers to adopt a chair-like

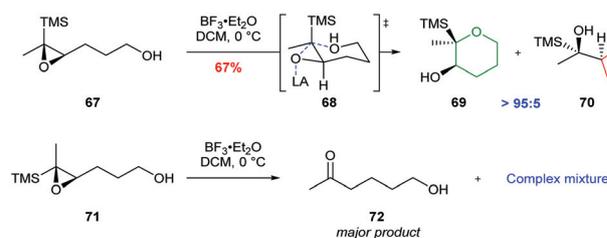
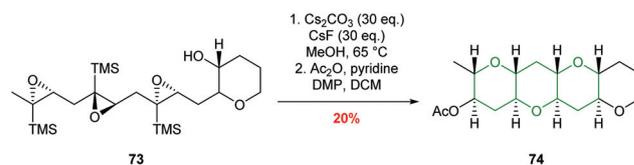
Scheme 15 Intramolecular ring opening of 1,4-*anti* (top) and 1,4-*syn* (bottom) epoxy alcohols.⁵⁴

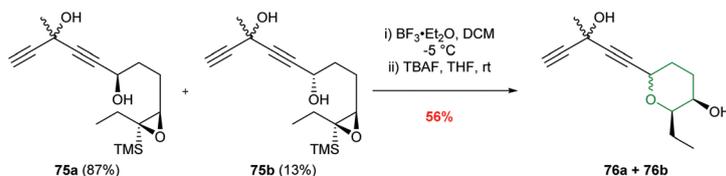
conformation **60** in the transition state whilst cyclisation of 1,4-*syn* diastereoisomers would require a boat-like transition state **63**. To avoid this high energy pathway, the reaction instead could proceed *via* an S_N1 mechanism, facilitated by the β-cation stabilisation effect of silicon.

Jamison further investigated cyclisations of epoxysilanes as a strategy for THP synthesis in ladder polyethers.⁵⁵ In this case, trisubstituted epoxysilane **67**, which places silicon in the axial position during cyclisation, gave THP **69** as the major product, whereas the diastereomer **71** produced a complex mixture of products (Scheme 16). The use of epoxysilanes as substrates is of particular value for the selective synthesis of THPs as the silyl directing group may be readily removed *via* a TBAF promoted Brook rearrangement.

Jamison used this chemistry to good effect in iterative cascades, creating fused THP rings in excellent stereoselectivities and yields (Scheme 17).^{56,57} It was found that while BF₃·Et₂O was effective in the synthesis of isolated tetrahydropyrans, poor selectivity was obtained in cascade reactions. In contrast, the use of mildly basic conditions (Cs₂CO₃ and CsF in MeOH) facilitated the formation of fused THPs with concomitant deprotection of the silyl directing groups *via* a homo-Brook rearrangement (Scheme 17).

Jauch and co-workers used BF₃·Et₂O in the conversion of epoxysilane **75** to create the 6-*endo* product in 81% yield (Scheme 18).⁵⁸ The silyl directing group was subsequently

Scheme 16 Cyclisation of *cis*-epoxysilane **67** and *trans*-epoxysilane **71** by Jamison.⁵⁵Scheme 17 Epoxide-opening cascade directed by silicon.^{56,57}

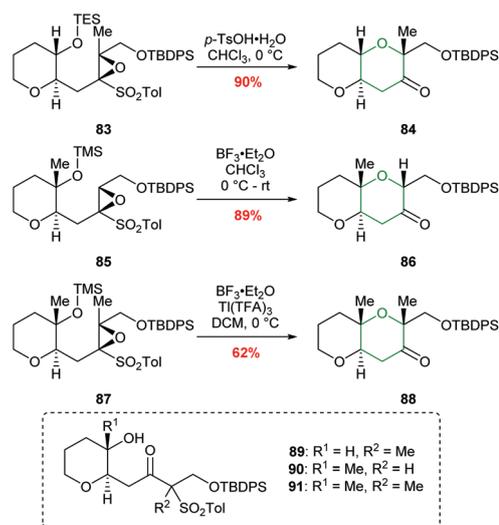


Scheme 18 Synthesis of eastern THP ring of the jerangolids by Jauch.⁵⁸

removed with TBAF to furnish THP **76** required for the construction of the jerangolids.

2.1.4 Epoxysulfones. The use of alkenic and acetylenic epoxides as well as epoxysilanes as substrates for IERO all deliver the THPs as the major products by stabilising the 6-*endo* transition state. An alternative approach was developed by Mori, where judicious incorporation of a sulfonyl functional group on epoxide **77** led to destabilisation of the 5-*exo* transition state **81** and therefore formation of the 6-*endo* product **80** was favoured (Scheme 19).^{59,60} Upon treatment of epoxysulfone **77** with Brønsted acid, a sequence of silyl deprotection, 6-*endo* cyclisation and finally loss of phenylsulfonate afforded THP **80** in 80% yield. This process cleanly removes the sulfonyl directing group *in situ* and generates a ketone which can serve as a valuable functional group for further synthetic manipulations. The strong electron withdrawing nature of the sulfonyl group disfavours nucleophilic attack α to sulfur, which would proceed *via* the high energy transition state **81**, and 5-*exo* product **82** was not detected.

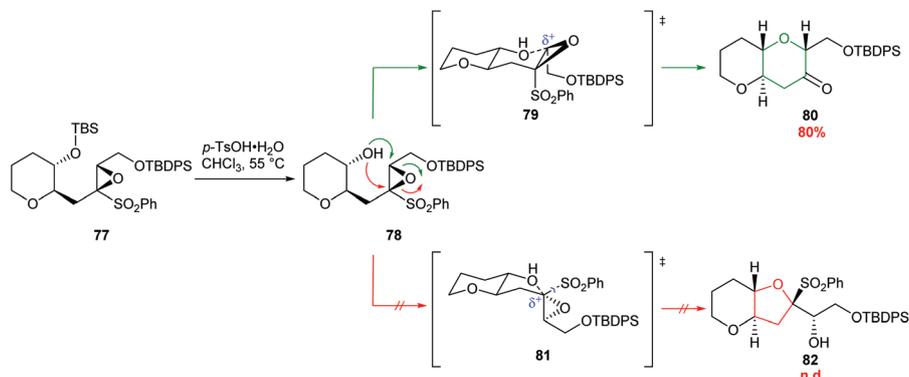
A common structural feature of polyether marine toxins is the presence of methyl groups on *trans*-fused polytetrahydropyran ring systems. Mori constructed these scaffolds in excellent yields from epoxysulfones (Scheme 20).⁶¹ For example, treatment of epoxysulfone **77** with *p*-TsOH at 55 °C gave THP **80** in 80% yield (Scheme 19). Alternative reaction conditions were required when the substrate possessed either a tetrasubstituted epoxide (**83**) or a silyl protect tertiary alcohol (**85** and **87**) rather than a secondary alcohol (Scheme 20). Treatment of epoxide **83** with TsOH at 0 °C gave **84** in 90% yield whereas at higher temperatures 1,2-sulfonyl migration occurred leading to ketone **89** as the major product. For tertiary silyl ether **85**,



Scheme 20 Conversion of methyl substituted epoxy sulfones **83**, **85** and **87** to the corresponding THP products.⁶¹

Lewis acidic conditions were required to facilitate clean conversion to the THP **86**. Finally, for the formation of dimethyl-substituted THP **88** from **87**, $\text{BF}_3 \cdot \text{Et}_2\text{O}$ was used to promote cyclisation, and $\text{Ti}(\text{TFA})_3$ was added to suppress the problematic 1,2-sulfonyl rearrangement.

The relative stereochemistry of the epoxysulfone substrate proved to be important in the outcome of reactions with Brønsted acids with cyclisation only occurring with substrates such as **78**, whilst **92**, **93** and **94** were unreactive (Fig. 4). These differences in reactivity were proposed to be due to steric inter-



Scheme 19 6-*endo* cyclisation of epoxysulfone **77**.⁵⁹



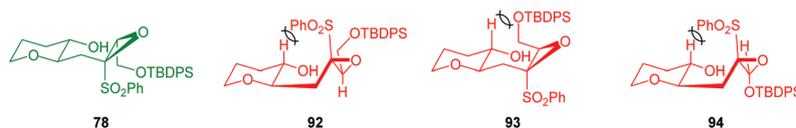
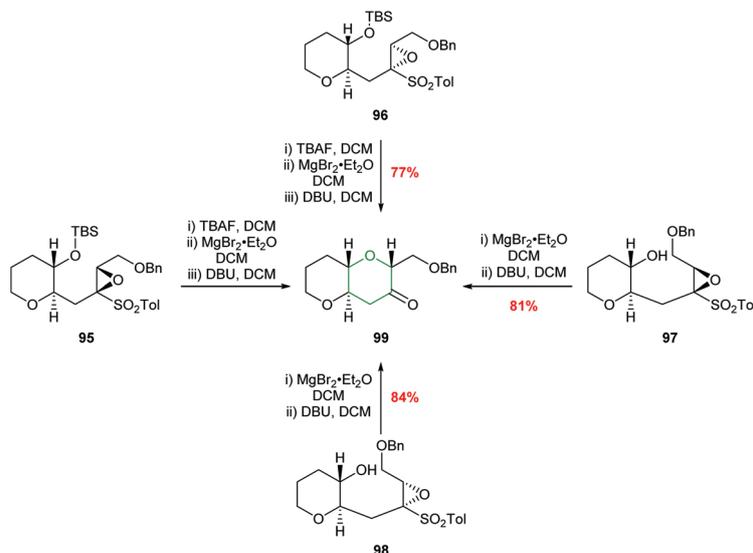


Fig. 4 Proposed transition states and corresponding steric clashes for the intramolecular epoxide ring opening of epoxysulfones **78**, **92**, **93** and **94**.⁶²



Scheme 21 Alternative procedure for the synthesis of THP **99**.⁶²

actions preventing the unreactive isomers adopting the 6-*endo* transition states.⁶² To expand the substrate scope, Mori used alternative reaction conditions to effect cyclisation and deliver THPs (Scheme 21). Thus, the silyl ether of the epoxysulfone (**95** or **96**) was first deprotected with TBAF and the resultant alcohols treated with $\text{MgBr}_2 \cdot \text{Et}_2\text{O}$ and finally addition of DBU gave bicyclic ketone **99** in good yield and excellent stereocontrol.⁶² Although this is not an intramolecular epoxide ring opening process, the formal 6-*endo* product is obtained.

Sulfonyl directed intramolecular epoxide ring opening has been widely used by Mori and others to prepare THP containing natural products and some examples are shown in Scheme 22.^{63–68}

2.1.5 Trialkyl substituted epoxides. The ability of methyl groups to stabilise positive charged intermediates of epoxide ring-opening reactions in the synthesis of polycyclic polyethers has been reviewed previously.²¹ McDonald and co-workers reported that the nature of the terminating nucleophile plays an important role in the outcome of such cascade processes.^{69–71} Gagné *et al.* investigated the gold catalysed cyclisation of allenic epoxides **108** and **110** with methyl groups located at different ends of the trisubstituted epoxide (Scheme 23).⁷² By switching the position of the methyl group, 5-*exo* and 6-*endo* cyclisations occurred selectively.

A further example of this regiocontrol was demonstrated by Holton in the total synthesis of hemibrevetoxin B reported in

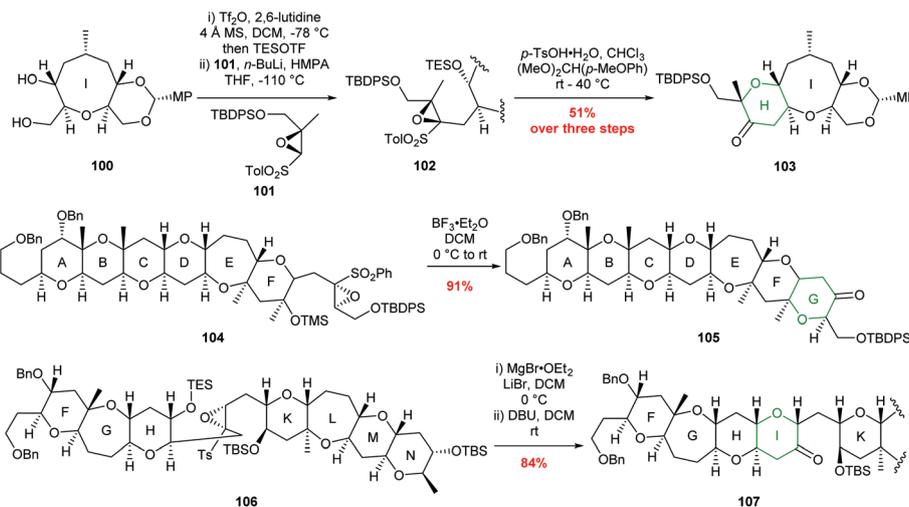
2002 (Scheme 24).⁷³ Reaction of **112** with *N*-(phenylseleno)phthalimide promotes a cascade process leading to formation of 6- and 7-membered oxygen heterocycles in 83% yield. The use of hexafluoroisopropanol (HFIP) as a solvent was proposed to be important to increase the charge separation in the reaction and therefore improve selectivity.

More recently, Qu and co-workers demonstrated how trialkyl substituted epoxides are converted to the corresponding THP products through reaction with 1-ethyl-3-methylimidazolium tetrafluoroborate ($[\text{EMIM}]\text{BF}_4$) in perfluoro-*tert*-butanol (PFTB).⁷⁴ In contrast, disubstituted epoxides formed solely the THF products. These contrasting results indicate the important role that an extra substituent on the epoxide can play in controlling 6-*endo* cyclisation. Many selective cyclisation cascades have been used to good effect to generate the core structures of marine polycyclic polyether natural products.

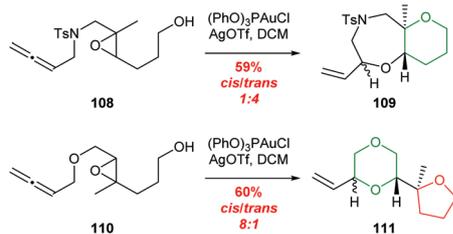
2.2 Reagent controlled formation of THPs from 4,5-epoxy alcohols

2.2.1 TIPSOTf mediated cyclisations. Although controlling cyclisation using directing groups is effective, it imposes constraints on the substrates which can be used. It would be ideal to switch between the formation of the 5-*exo* and 6-*endo* products by simply altering the reaction conditions. This was elegantly demonstrated by Morimoto and co-workers during





Scheme 22 Examples of sulfonyl directed intramolecular epoxide ring opening in total synthesis. H ring of ciguatoxin (top), G ring of gambierol (middle) and I ring of gymnocin-A (bottom). MP = 4-methoxyphenyl.^{65,66,68}



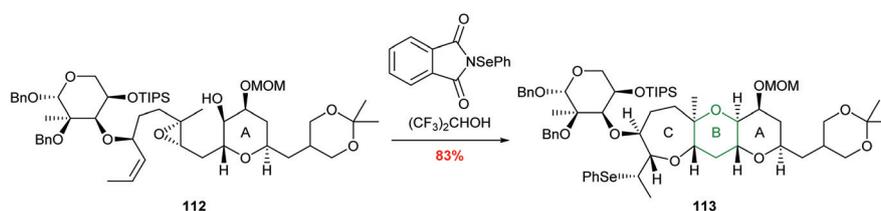
Scheme 23 Gold-catalysed cyclisation of allenic epoxides **108** and **110**.⁷²

investigations into the synthesis of various oxasqualenoids (Scheme 25).^{75,76} Interestingly, despite the presence of a trisubstituted epoxide, Brønsted acid catalysed cyclisation of epoxy-alcohol **114** in DCM gave 5-*exo* product **115** in which attack occurred at the less substituted end of the epoxide. In contrast treatment of the same substrate **114**, with TIPSOTf gave tetrahydropyran **116** in good yield. Morimoto proposed that the 6-

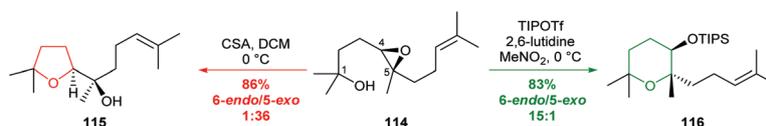
endo product was formed preferentially due to steric repulsion between the bulky silyl group and the substituents on C-5 in the 5-*exo* transition state. Further investigations revealed that steric bulk at C-5 was required to maintain the preference of the 6-*endo* product. However, this process was limited to tertiary alcohols due to formation of triflates with other epoxy alcohols.

In 2007, Morimoto demonstrated the value of these reagent-controlled cyclisations to create both 5 and 6 membered rings with excellent selectivity in the total synthesis of (+)-enshuol **121** (Scheme 26).⁷⁷ Whilst this methodology still imposes constraints on the substrate structure, it demonstrates the power of using different reagents to control selectivity.

2.2.2 Use of chiral phosphoric acid. Pseudomonic acid A is an antibiotic produced by the bacterium *Pseudomonas fluorescens*. It is unstable under acidic conditions due to attack of the 7-hydroxyl group onto the 10,11-epoxide giving a mixture of bicyclic products (Scheme 27). In 2020, Nagorny and co-workers reported the reagent controlled cyclisation of methyl

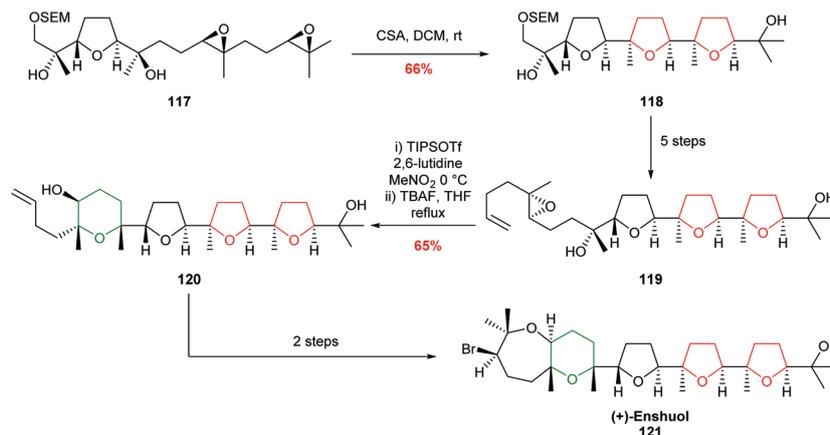


Scheme 24 Synthesis of the B and C rings of hemibrevetoxin B.⁷³

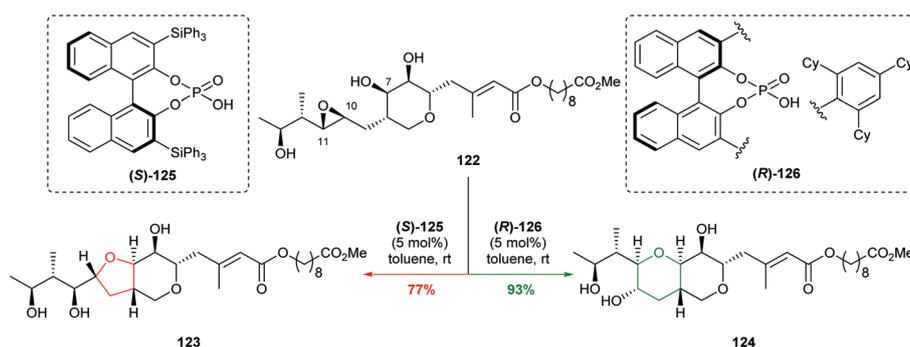


Scheme 25 Reagent-controlled selectivity in cyclisations of 4,5-epoxy alcohol **114**.⁷⁵





Scheme 26 Total synthesis of (+)-enhuol by TIPSOTf promoted 6-endo cyclisation.⁷⁷



Scheme 27 Cyclisations of epoxide **122** using chiral phosphoric acid catalysts.⁷⁸

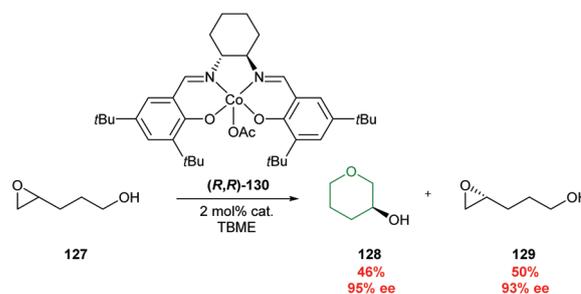
pseudomonte A **122** using chiral phosphoric acid catalysts to give either **123** or **124** in 77% and 93% yield respectively (Scheme 27).⁷⁸

Through a combination of mechanistic and computational studies, Nagorny proposed the selectivity originated from steric clashes between the catalyst and substrate set up by a hydrogen bond network. In particular, steric interactions between (*R*)-**126** and **122** increase the energy of the 5-*exo* transition state, whilst no such interactions occur between (*S*)-**125** and **122**. Although this is a substrate specific case, it elegantly illustrates the use of non-covalent interactions with a chiral catalyst in intramolecular attack on epoxides. With the aid of computational methods and experimental design, this work may pave the way forward for developing new catalysts for controlling cyclisation of a variety of 4,5-epoxy alcohols.

2.2.3 Cobalt and vanadium catalysed cyclisations. While the transition metal catalysed methods described earlier using palladium and rhodium required an α,β -unsaturated ester adjacent to the epoxide to facilitate 6-*endo* cyclisation (section 2.1.1), other more general metal catalysed cyclisations of 4,5-epoxy alcohols have been developed. In 1999, Jacobsen reported the use of Co^{III}(salen) catalyst (*R,R*)-**130** in the IERO of racemic epoxide **127**, with complete 6-*endo* selectivity (Scheme 28).⁷⁹ The chiral catalyst was selective for one enantio-

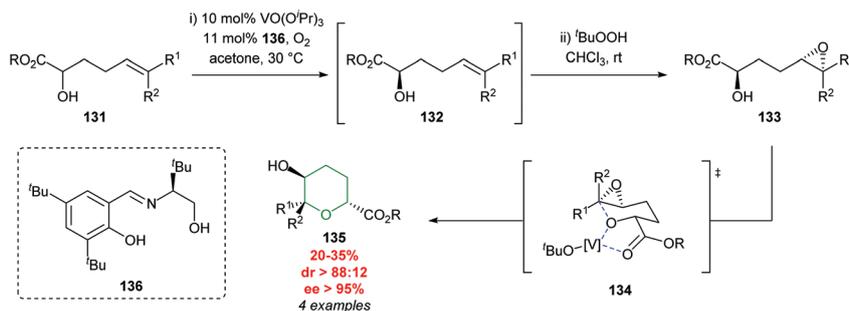
mer of epoxide **127**, facilitating a kinetic resolution giving a separable mixture of THP **128** in 46% yield and 95% ee along with (*R*)-epoxide **129** (50% yield, 93% ee). The origin of this 6-*endo* selectivity was not discussed, although a mechanism for an analogous process has been proposed.⁸⁰

Later in 2006, Toste developed a vanadium catalysed kinetic resolution starting from unsaturated alcohols (Scheme 29).⁸¹ The approach proceeded *via* an initial resolution of α -hydroxy ester **131**, followed by subsequent diastereoselective epoxidation of **132** and finally regioselective intramolecular epoxide ring opening to afford THP products **135**. Cyclisation is pro-



Scheme 28 Kinetic resolution of 4,5-epoxy alcohols through cobalt catalysis.⁷⁹



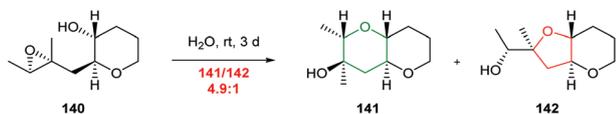
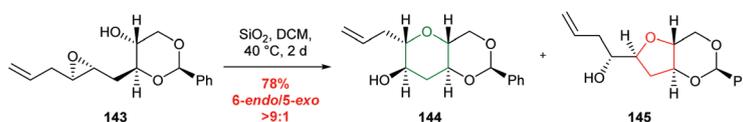
Scheme 29 Vanadium catalysed synthesis of 2,5-*trans*-tetrahydropyrans.⁸¹

posed to proceed *via* coordination of the vanadium complex to the hydroxy ester **133** *via* a chair like transition state **134**.⁴ To the best of our knowledge, neither the cobalt or vanadium catalysed processes have been used in the total synthesis of natural products.

2.2.4 Templated cyclisations in water. When investigating *endo*-selective epoxide ring opening cascades for the synthesis of marine polycyclic polyethers, Jamison proposed that regio-control may be achieved using a THP template within the starting material.⁸² Whilst poor selectivity was observed using Brønsted acids and bases as well as Lewis acidic conditions in organic solvents (entries 1–3, Table 4), when cyclisation was conducted in water, excellent 6-*endo* selectivity was achieved (entry 4).⁸³ Qu and co-workers revealed that a combination of the aqueous environment and the template was required, as

Table 4 Cyclisation of THP templated epoxy alcohol **137**⁸³

Entry	Conditions	138 : 139
1	Cs ₂ CO ₃ , MeOH	1 : 2.7
2	BF ₃ ·Et ₂ O, DCM	1.4 : 1
3	AcOH, toluene	1.6 : 1
4	H ₂ O	>10 : 1

Scheme 30 Water promoted 6-*endo* cyclisation of 4,5-epoxy alcohol **140**.⁷⁰Scheme 31 Silica gel promoted 6-*endo* cyclisation of benzylidene acetal templated epoxide **143**.⁸⁷

linear substrates afforded primarily THF products.⁸⁴ Jamison *et al.* conducted detailed mechanistic studies whereby the role of the template and water was interrogated, confirming the role of the THP template.^{85,86}

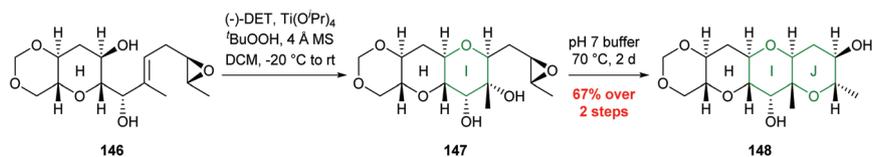
Whilst a methyl substituent on a trisubstituted epoxide can direct nucleophilic attack as described in section 2.1.5, interestingly Jamison demonstrated that cyclisation of THP templated substrates proceeded to give *endo* products such as **141** under aqueous conditions (Scheme 30).⁷⁰

Building on the use of the THP ring template, Jamison developed a benzylidene acetal template to achieve selective 6-*endo* cyclisation which has the advantage that following reaction, the template was readily removed (Scheme 31).⁸⁷ Both water and silica gel were used to promote the desired cyclisation.

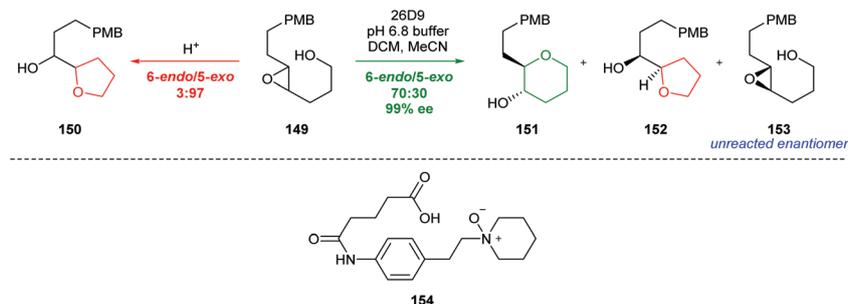
This methodology was used to good effect in the synthesis of the HIJK ring system of gymnocin A through a polycyclisation cascade in water, and the J ring of yessotoxin (Scheme 32).^{87,88}

2.2.5 Antibody catalysis. In 1993, Lerner *et al.* showed that antibodies could be used to catalyse the 6-*endo* cyclisation of 4,5-epoxy alcohol **149** (Scheme 33).⁸⁹ A transition state mimic **154** was designed such that the anionic charge of the *N*-oxide would recruit cationic amino acid residues to promote epoxide ring opening, and cationic charge would induce the formation of anionic residues to stabilise a buildup of positive charge in the 6-*endo* transition state. Antibodies were produced in mice against the transition state mimic **154** and then purified. Of the 26 antibodies obtained, antibody 26D9 exhibited excellent catalytic activity, converting epoxide **149** to THP **151** as the major product. Importantly, antibody 26D9 was selective for one enantiomeric epoxide, resulting in a combined selective cyclisation and resolution. Computational studies indicated the antibody catalyst must lower the energy of the 6-*endo* transition state by more than 3.6 kcal mol⁻¹ relative to the 5-*exo*





Scheme 32 Synthesis of J ring of yessotoxin by Jamison.⁸⁸



Scheme 33 Cyclisation of epoxy alcohol **149** under acidic conditions and antibody catalysis (top) and transition state mimic **154** used to produce antibody catalyst 26D9.^{89,91}

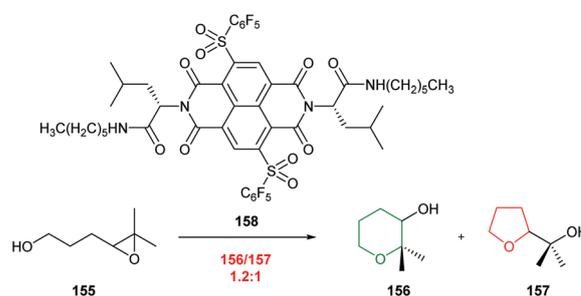
transition state.⁹⁰ Further studies by Lerner yielded the novel antibody 5C8, which also showed excellent catalytic activity. Through the use of X-ray crystallography and docking studies, an acid–base catalysed mechanism was proposed.⁹¹

A similar approach was investigated by Chiosis, but poor selectivity was achieved.⁹² Nevertheless, both these studies illustrate the power of antibody catalysis to overcome the inherent preference for 5-*exo* cyclisation of 4,5-epoxy alcohols.

2.2.6 π -Anion catalysis. The recent development of strategies to stabilise anions through aromatic π -surfaces provides an exciting opportunity for controlling regioselectivity of reactions such as cyclisations of 4,5-epoxy alcohols.^{93,94} Matile and co-workers reported that π -anion surfaces can promote 5-*exo* cyclisation of 4,5-epoxy alcohols. Nevertheless, a slight preference for 6-*endo* cyclisation was observed when epoxide **155** was cyclised in the presence of naphthalene diimide (NDI) catalyst **158** (Scheme 34). Although the origin of this selectivity is unclear, this preliminary result illustrates the potential to develop alternative π -anion catalysts to overcome inherent 5-*exo* reactivity to form THF rings.

2.3 Further methods

2.3.1 Expansion of THF rings. During the total synthesis of lasalocid A, Kishi revealed a valuable rearrangement which converted the 5-*exo* product, generated by an intramolecular epoxide ring opening, to the corresponding THF ring (Scheme 35).⁹⁵ For example, following synthesis of THF **161**, the secondary alcohol was converted to mesylate **162** and subsequent silver carbonate promoted ring expansion afforded THF **163** in 65% yield, which was used as an intermediate in the synthesis of lasalocid A. Whilst this strategy requires extra synthetic transformations, no directing group was required.



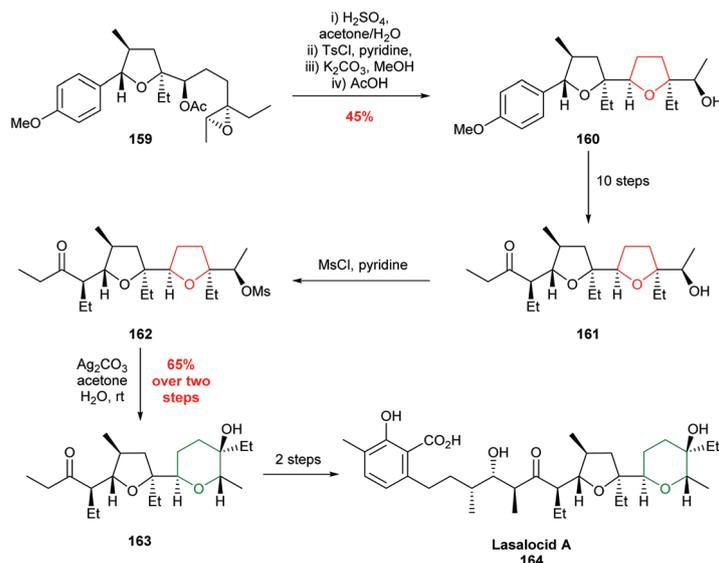
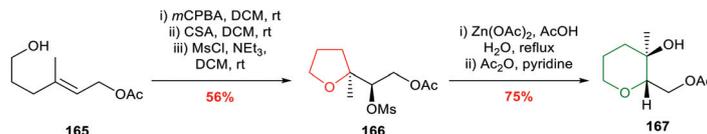
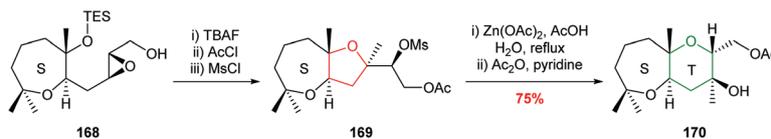
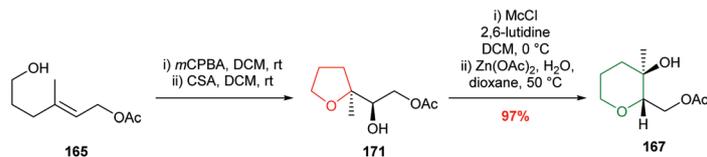
Scheme 34 Cyclisation of 4,5-epoxy alcohol **155** catalysed by NDI **158**.⁹³

Nakata and co-workers reported further investigations into this ring expansion and first converted unsaturated alcohol **165** to an epoxide which cyclised to the expected THF on treatment with camphorsulfonic acid in DCM (Scheme 36).⁹⁶ The key ring expansion step was performed by heating mesylate **166** in AcOH and water in the presence of Zn(OAc)₂ giving, after acetylation, THF **167** in 75% yield.

This methodology was used in the synthesis of a model compound **170** with the ST rings of maitotoxin (Scheme 37).^{34,97}

Zinc acetate in refluxing acetic acid was required for the ring expansion of mesylates **166** and **169**. However, Nakata showed that by using chloromethylsulfonate (Mc) as the leaving group, rather than mesylate, ring expansion was achieved without the need for acetic acid (Scheme 38).^{98,99} These modified conditions were used in the efficient conversion of THF **171** to THF **167**. Furthermore, it was shown that ring expansion may be performed in the absence of a Lewis acid, albeit giving 6-membered rings in lower yield.



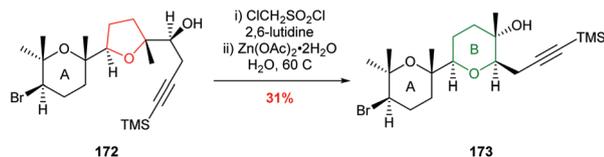
Scheme 35 Total synthesis of lasalocid A.⁹⁵Scheme 36 Preparation of THP **167** by ring expansion of 5-*exo* cyclisation product.⁹⁶Scheme 37 Synthesis of model ST rings of maitotoxin.^{34,97}Scheme 38 Alternative conditions for the rearrangement-ring expansion of 5-*exo* product **171**. Mc = chloromethylsulfonate.⁹⁹

McDonald used this ring expansion strategy to create the B ring of 15,28-dideoxy-15,28-didehydrothyrseol in 31% yield over 2 steps from alcohol **172** (Scheme 39).¹⁰⁰

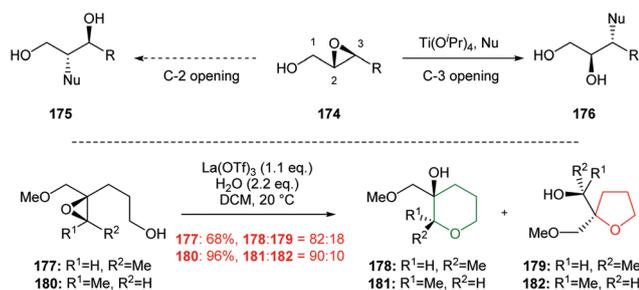
2.3.2 Cyclisation of 4,5-epoxy-4-methoxymethyl-1-hexanols. The intermolecular reaction of nucleophiles with 2,3-epoxy alcohols in the presence of $\text{Ti}(\text{O}^i\text{Pr})_4$ is an established method for achieving regioselective epoxide ring opening at the C-3 position (Scheme 40).¹⁰¹ Murai and co-workers investigated intramolecular variants, whereby chelation of 4,5-epoxy-4-

methoxymethyl-1-hexanols **177** and **180** with Lewis acids would promote 6-*endo* over 5-*exo* cyclisation (Scheme 40).¹⁰² Optimisation studies revealed that $\text{La}(\text{OTf})_3$ with 2.2 equivalents (eq.) of water in DCM converted both *cis* and *trans* epoxides **177** and **180** to the corresponding THP products **178** and **181** respectively. These conditions proved to be highly specific, and changing solvent, water content or Lewis acid reversed the regioselectivity of epoxide ring opening or resulted in low yields. Murai proposed that the observed selectivity was due to

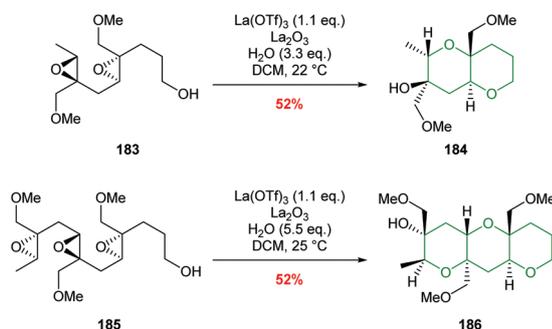




Scheme 39 Synthesis of B ring in 15,28-dideoxy-15,28-didehydrothyrseanol.¹⁰⁰



Scheme 40 Inter- and intramolecular Lewis acid catalysed regioselective epoxide ring opening.^{101,102}



Scheme 41 La(OTf)₃ catalysed polyepoxide ring opening cascades.¹⁰³

water coordinating to La inducing an optimal chelation bite angle between the epoxide and methoxymethyl oxygen, which in turn leads to selective attack. However, no mechanistic or computational studies were reported to support this hypothesis.

Two examples of the use of La(OTf)₃ mediated cascade reactions have been reported leading to bicyclic and tricyclic products **184** and **186** (Scheme 41).¹⁰³ There remains opportunities for exploitation of this methodology in natural product synthesis.

2.4 Summary of synthetic methods

In summary, many methods have been developed for the preparation of oxygen heterocycles from 4,5-epoxy alcohols. Structural features may be incorporated into the substrate, such as double and triple bonds, epoxysilanes and epoxysulfones, which favour formation of tetrahydropyrans *via* either stabilisation of a 6-*endo* transition state or destabilization of a 5-*exo* transition state, to afford tetrahydropyrans in high yields.

These strategies have been used in natural product synthesis. There are exciting prospects for the use of reagents and catalysts in IERO to control 5-*exo* and 6-*endo* cyclisation of 4,5-epoxy alcohols. Both Lewis acids and chiral phosphoric acids have been used in regioselective cyclisations but to date these methods have not been widely used in the total synthesis of natural products. However, taking inspiration from nature might facilitate the optimisation of catalysts and development of novel methods for reagent controlled 6-*endo* cyclisation for THP formation. In the next section the biosynthesis of THPs *via* cyclisations of 4,5-epoxy alcohols are reviewed.

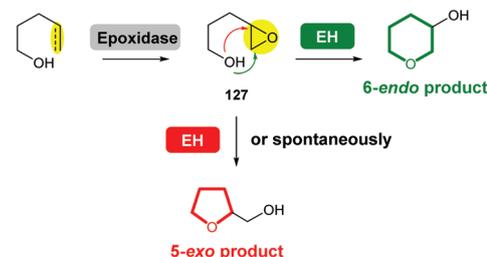
3. Biosynthetic methods

Compared with the diversity of synthetic methods for the preparation of saturated oxygen heterocycles, a more limited set of reactions are used in natural product biosynthesis. However, epoxide formation/intramolecular epoxide-opening cascades are common processes used in the biosynthesis of many natural products. In some cases, epoxide hydrolases (EHs) have been identified which control the stereochemical outcome of cyclisation. When EHs are absent, labile epoxide biosynthetic intermediates **127** often spontaneously cyclise to generate 5-*exo* THF products (Scheme 42).

Interestingly, the biosynthesis of some natural products with structures containing THF rings has also been reported to be mediated by EHs, or in some cases, the 5-*exo* cyclisation is significantly accelerated by the presence of an EH. While this review focuses on THP formation, the biosynthesis of THF rings *via* a similar epoxide formation/epoxide-opening cascade reaction is topical and therefore is also discussed.

3.1 Formation of THPs from 4,5-epoxy alcohols catalysed by epoxide hydrolases (EHs)

3.1.1 Lasalocid A. Lasalocid A isolated from *Streptomyces lasaliensis* is one of the important ionophore antibiotics among commercially available anticoccidial agents.¹⁰⁴ Formation of the terminal THP ring in lasalocid A has attracted significant attention from researchers in the field and was amongst the earliest examples reported of intramolecular epoxide ring opening to generate a 6-membered ring in natural product biosynthesis.



Scheme 42 Biosynthesis of THPs and THFs *via* epoxide formation/epoxide-opening cascade reactions. EH: epoxide hydrolase.



Using *in vivo* and *in vitro* studies of the flavin-dependent monooxygenase (FMO) Lsd18, Minami *et al.* demonstrated that Lsd18 is responsible for oxidising the diene precursor **187** to afford bisepoxyprelasalocid A **188** (Scheme 43).¹⁰⁵ In search for the enzyme responsible for selective epoxide-opening, Oikawa and co-workers identified Lsd19 as the putative epoxide hydrolase from the lasalocid biosynthetic gene cluster. *In vitro* turnover assays of bisepoxyprelasalocid A **188** with purified Lsd19 showed conversion of the synthetic substrate into lasalocid A (Scheme 43).¹⁰⁴ In the absence of Lsd19, the synthetic bisepoxyprelasalocid A **188**, which was obtained in only 3% yield, spontaneously cyclises to generate the THF ring of isolasalocid A **189**.¹⁰⁴ As discussed in section 2.3.1, Kishi synthesised the THP ring of lasalocid A by initial 5-*exo* cyclisation of an analogous epoxide **159** followed by ring expansion (Scheme 35). Whilst an elegant approach, this contrast between the synthesis and biosynthesis of the lasalocid THP ring highlights how nature can elegantly employ EH enzymes to readily overcome inherent selectivity for 5-*exo* cyclisation of 4,5-epoxy alcohols.

Concurrent *in vivo* studies by the Leadlay group on the targeted deletion of the *lsd19* gene (previously identified as *lasB*) in the lasalocid producing strain further supported the role of Lsd19 in directing biosynthesis towards lasalocid A rather than towards isolasalocid A.¹⁰⁶ In the Δ *lasB* mutant of *Streptomyces lasaliensis*, production of lasalocid A was no longer detected, and isolasalocid A was the exclusive product.¹⁰⁶

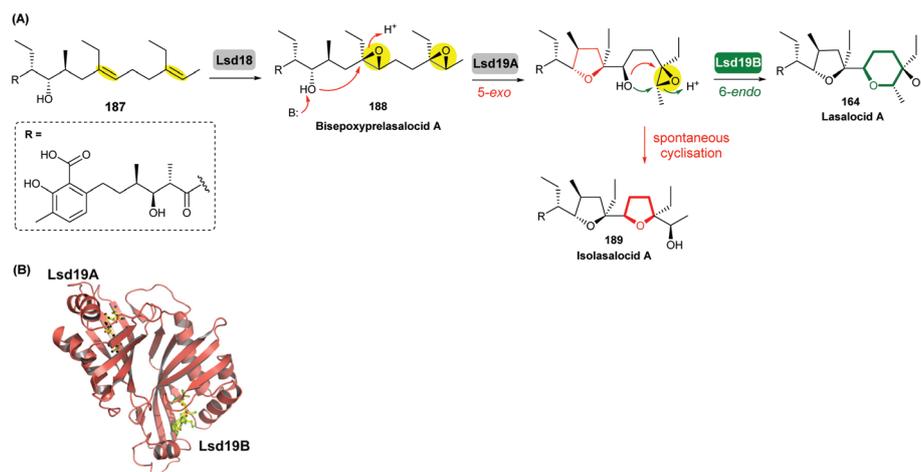
To gain a better understanding of how Lsd19 catalyses THP formation, Hotta *et al.* determined the X-ray crystal structure of Lsd19 in complex with its substrate and a product analogue to 1.59 Å resolution (Scheme 43).¹⁰⁷ The epoxide hydrolase consists of two catalytic domains including the N-terminal region Lsd19A and the C-terminal region Lsd19B. The 5-*exo* and 6-*endo* epoxide-opening steps are catalysed by Lsd19A and Lsd19B, respectively. On the basis of structural analysis and quantum mechanical calculations, a mechanism of general

acid–base catalysis has been proposed in which an aspartate residue acts as a base that activates the hydroxyl group on the substrate for a nucleophilic attack on the epoxide carbon, while a tyrosine residue acts as an acid to stabilise the negative charge that develops on the epoxide oxygen during the transition state of the reaction.¹⁰⁷

3.1.2 Aurovertin. Aurovertins are polyketides isolated from fungal species such as *Calcarisporium arbuscula* that exhibit significant inhibition of oxidative phosphorylation.¹⁰⁸ Structurally, aurovertins contain an unusual dioxabicyclo-octane ring that is formed *via* iterative epoxidations and epoxide-openings of a polyene precursor.

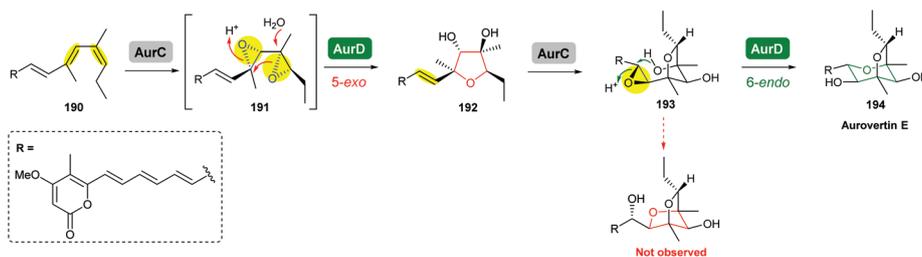
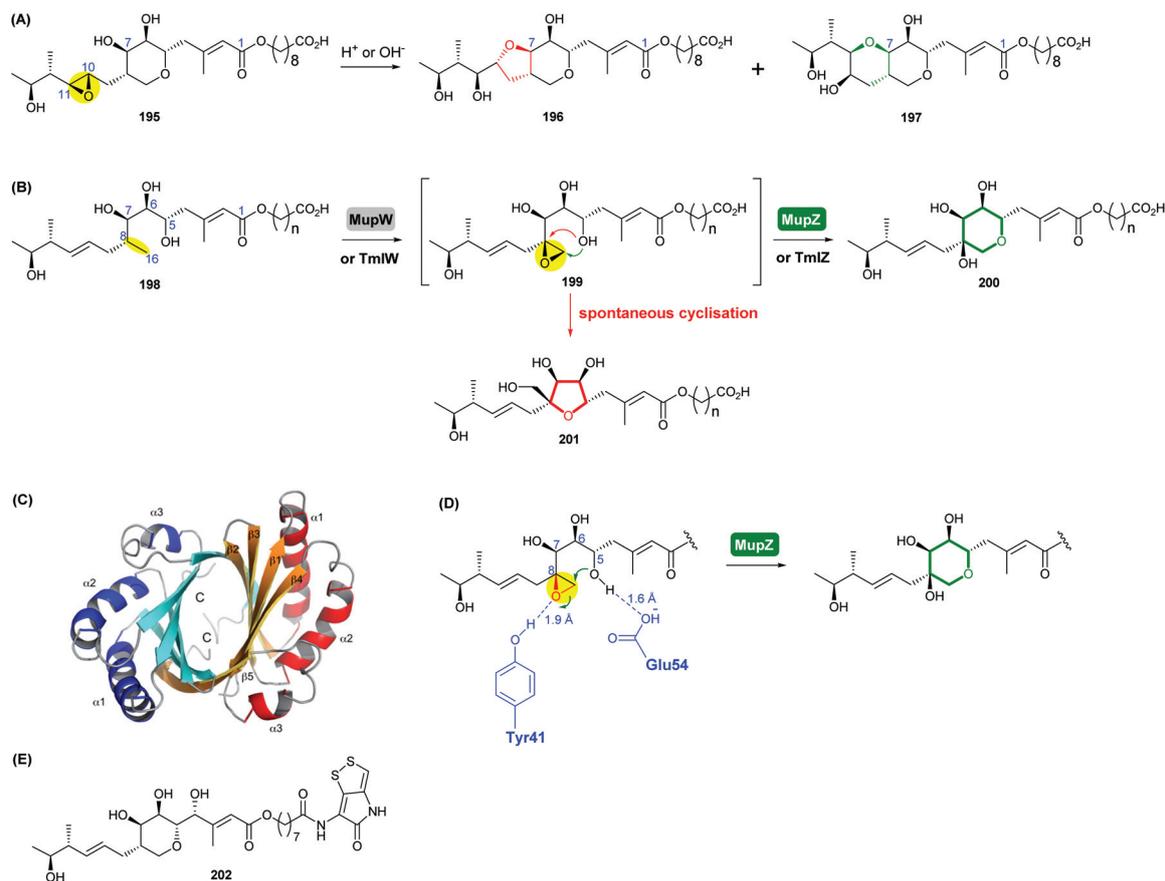
The terminal triene portion **190** of the polyene precursor firstly undergoes bisepoxidation catalysed by the flavin-dependent monooxygenase AurC to form a proposed bisepoxide **191**, which is then transformed into an isolable THF intermediate **192** through a 5-*exo* epoxide-opening step catalysed by the epoxide hydrolase AurD (Scheme 44). This stable THF intermediate then undergoes another round of epoxidation (by AurC) to give **193** and in this case cyclisation occurs *via* 6-*endo* ring opening (by AurD) to yield the dioxabicyclo-octane ring product **194**.¹⁰⁸

3.1.3 Mupirocin/thiomarinol. Mupirocin, produced by *Pseudomonas fluorescens* NCIMB 10586, is a clinically important antibiotic against Gram-positive bacteria, including methicillin-resistant *Staphylococcus aureus* (MRSA).¹⁰⁹ The major component of mupirocin (pseudomonic acid A, **195**) bears a 10,11-epoxide which outside the pH range of 4–9, undergoes intramolecular attack by the 7-hydroxyl group to yield THF and THP rearrangement products **196** and **197** (Scheme 45).¹⁰⁹ As discussed earlier in the synthetic methods section, Nagorny and co-workers recently reported the reagent controlled opening of this 10,11-epoxide to yield either the THF or THP as the major product (Scheme 27, section 2.2.2).⁷⁸ However, of particular interest is the pharmacophoric THP ring of mupirocin, which is biosynthesised *via* a unique



Scheme 43 (A) THP ring formation in lasalocid A biosynthesis; (B) Crystal structure of Lsd19 with bound substrate (left half) and product (right half) analogues shown by stick models.^{104–107}



Scheme 44 Proposed late-stage biosynthesis of aurovertins.¹⁰⁸Scheme 45 (A) Rearrangement products of pseudomonic acid A (**195**) formed under either acidic or basic conditions; (B) THP ring formation in mupirocin/thiomarolin biosynthesis; (C) crystal structure of MupZ; (D) proposed mechanism of MupZ-catalysed THP ring formation; (E) structure of thiomarolin A (**202**).^{109,110}

epoxide formation/epoxide opening cascade starting from a non-activated alkane precursor.¹¹⁰ Notably, all the other epoxidases reviewed herein use alkene precursors as substrates.

Using whole-cell biotransformations, Crump, Willis and co-workers showed that the C8–C16 single bond in the acyclic precursor **198** is oxidised by the Rieske non-haem oxygenase MupW to give the corresponding 8,16-epoxide **199** (presumably through an 8,16-alkene intermediate), which spontaneously cyclises giving a five-membered tetrahydrofuran ring product **201** (Scheme 45).¹¹⁰ When both MupW and the epoxide hydrolase MupZ are included in the biotransformation

system, the hydroxylated THP ring product **200** is then generated. Deletion of the *mupZ* gene in the mupirocin producing strain abolished production of the THP ring metabolites and the THF ring compounds were produced instead. Based on X-ray crystallographic studies, molecular modelling and mutagenesis experiments of MupZ, the 6-endo epoxide-opening has been proposed to proceed through an acid–base mechanism, in which the Glu54 residue initially deprotonates the 5-hydroxyl group of the substrate (Scheme 45D), then the Tyr41 residue protonates the epoxide oxygen and subsequently stabilises the developing transition state, leading to the 6-endo



THP ring product **200**.¹¹⁰ This mechanism of general acid-base catalysis was found to be similar to that of the THP ring formation in lasalocid A biosynthesis.¹⁰⁷

The thiomarinols, produced by marine bacteria such as *Pseudoalteromonas* sp. SANK 73390, are a group of natural products that closely resemble mupirocin and also show significant antibiotic activities against MRSA.¹¹¹ Their structures are assembled on a highly equivalent polyketide backbone to mupirocin but are esterified by an 8-hydroxynonanoic acid chain and bear an additional non-ribosomal peptide synthase (NRPS)-derived pyrroline moiety (Scheme 45E). TmlW/TmlZ are the MupW/MupZ equivalents in thiomarinol biosynthesis and each share over 55% sequence identity to their counterparts in the mupirocin pathway. Using the same whole-cell bio-transformation approach, Wang *et al.* recently demonstrated that thiomarinol biosynthesis employs the same THP ring forming mechanism that requires the dual action of the Rieske non-haem oxygenase TmlW and the epoxide hydrolase TmlZ.¹¹¹ It was also shown that these two pairs of enzymes are cross compatible between the mupirocin and thiomarinol systems.¹¹¹

3.1.4 Xiamenmycin. Xiamenmycins are benzopyran natural products isolated from *Streptomyces xiamenensis* 318 with anti-fibrotic and anti-inflammatory activities.¹¹² Biosynthesis of the tetrahydropyran ring moiety in xiamenmycins requires the cooperation of a flavin-dependent monooxygenase (XimD) and a SnoaL-like cyclase (XimE).¹¹²

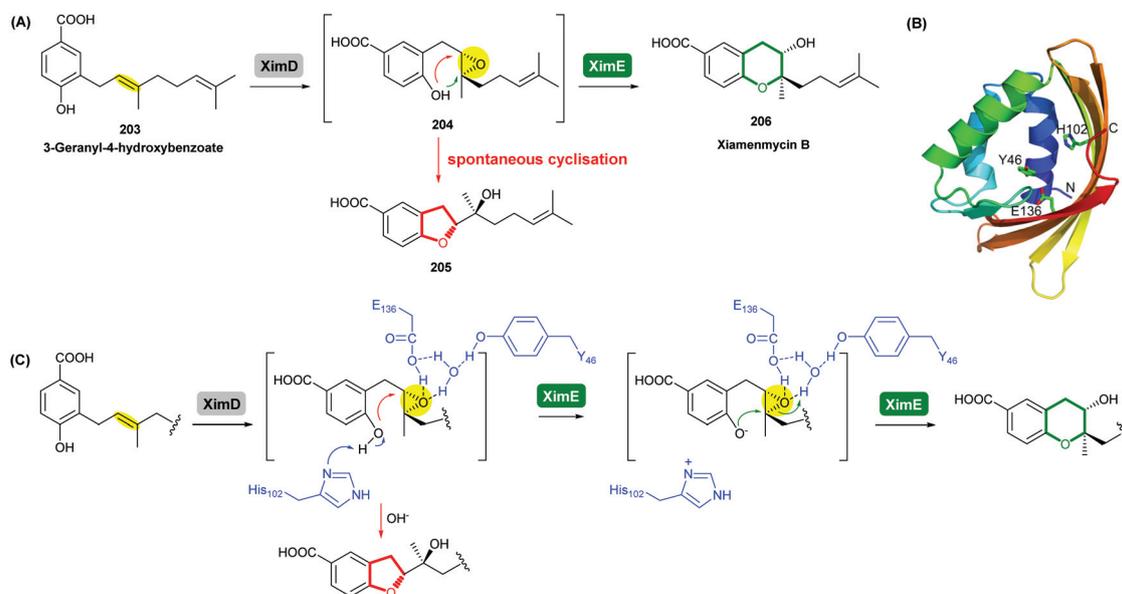
Using *in vitro* enzymatic assays, He *et al.* demonstrated that the 3-geranyl-4-hydroxybenzoate precursor **203** is converted by XimD to an unstable epoxide intermediate **204**, which spontaneously transforms to a benzofuran product **205**. Although the extra methyl substituent on the epoxide intermediate may have a directing effect that has been applied in synthetic

efforts to favour the THP ring formation (see section 2.1.5 for examples), no THP ring product was reported in this case. However, when XimE was added in the assay, the 6-membered ring benzopyran xiamenmycin B **206** was formed as the major product (Scheme 46).¹¹² Crystallographic structures of XimE, with and without product, suggested a synergistic mechanism in which residues E136, Y46, and H102 are functionally important to catalysis, substrate binding, and structural stabilisation (Scheme 46). Notably, in studies on substrate scope, these two enzymes exhibited high promiscuity capable of producing 14 products with 6 different benzoheterocyclic scaffolds.¹¹² In a follow-up study, this pair of substrate-promiscuous enzymes were further employed for combinatorial biosynthesis of a series of benzoheterocyclic derivatives with greater structural diversity.¹¹³

3.2 Formation of THPs from 4,5-epoxy alcohols in which no EH has been identified

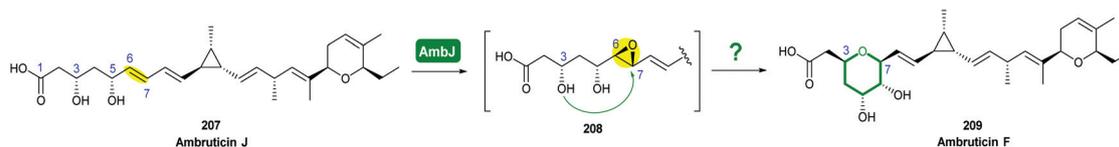
3.2.1 Ambruticin. The ambruticins are a group of myxobacterial polyketides produced by various *Polyangium cellulorum* and *Sorangium cellulorum* strains with potent antifungal activities against a broad range of pathogens.¹¹⁴ Their structures feature a poly-olefinic skeleton, with an embedded trisubstituted cyclopropane and two oxygen heterocycles – a tetrahydropyran and dihydropyran.

The biosynthetic pathway was proposed by Reeves and co-workers through analysis of the biosynthetic gene cluster and characterisation of compounds produced by gene knockout strains.¹¹⁴ Disruption of the *ambJ* gene that encodes for a flavin-dependent epoxidase yielded ambruticin J lacking the THP ring. Thus, the THP ring formation was proposed to occur through epoxidation of ambruticin J **207** by AmbJ, fol-



Scheme 46 (A) THP ring formation in xiamenmycin biosynthesis; (B) crystal structure of XimE; (C) proposed mechanism of XimE-catalysed pyran ring formation.¹¹²





Scheme 47 Proposed formation of the hydroxylated THP in ambruticin biosynthesis.¹¹⁴

lowed by selective epoxide-opening of **208** at C-7 to give ambruticin F (Scheme 47). It is not known whether this cyclisation is under enzymatic or substrate control, but to-date no epoxide hydrolase has been identified in the ambruticin gene cluster.¹¹⁴ Nevertheless, as demonstrated by the work of Nicolaou (section 2.1), alkenic epoxides such as epoxy-ambruticin J **208** readily under IERO to form THP products due to stabilisation of the 6-endo transition state by the adjacent π -system.²² Our more recent studies involving cyclisation of model 4,5-epoxy alcohols are in accord with the proposal that both the 8,9-alkene and 5-alcohol of epoxy-ambruticin J **208** may play a role in controlling 6-endo cyclisation.¹¹⁵ In 2021 the total synthesis of ambruticin J was reported enabling further biosynthetic studies to be carried out to elucidate the role of AmbJ.^{115,116}

3.2.2 Abyssomicin. The abyssomicins are spirotreronate marine natural products isolated from various *Verrucosipora* and *Streptomyces* species.¹¹⁷ To date, ~30 abyssomicins have been isolated and characterised, and they display a wide range of promising bioactivities including antibacterial and antiviral activities.¹¹⁸

Combining *in vivo* gene inactivation with *in vitro* biochemical studies, Li *et al.* showed that the ether ring formation and hydroxylation in abyssomicin 2 biosynthesis is catalysed by the cytochrome P450 AbmV.¹¹⁸ This was proposed to result from a domino reaction sequence involving, (i) epoxidation of the 11,12-alkene in abyssomicin 6 to generate an epoxide intermediate **210** followed by, (ii) epoxide ring-opening *via* nucleophilic attack of the tetronate OH upon C12 to afford abyssomicin 2 (Scheme 48).



Scheme 48 Proposed late stage pathway for abyssomicin biosynthesis.¹¹⁸

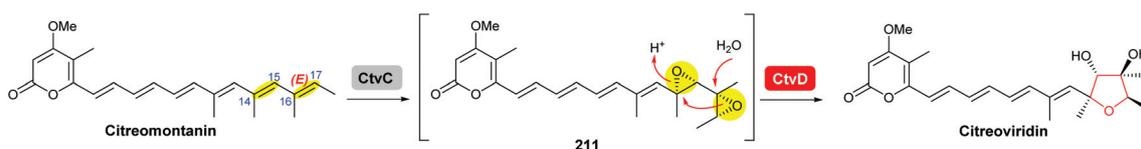
3.3 Formation of THFs catalysed by EHS

3.3.1 Citreoviridin. Citreoviridin is a highly reduced polyketide product isolated from several *Penicillium* species. It has been shown to inhibit the mitochondrial ATP synthetase system.¹¹⁹ Citreoviridin has a close structural similarity to the aurovertins (see section 3.1.2) and THF ring formation in citreoviridin involves a bisepoxidation step to generate **211** followed by cyclisation similar to aurovertin biosynthesis.¹⁰⁸

Using *Aspergillus nidulans* as a heterologous expression host, Wang and co-workers reconstituted the pathway and demonstrated that four genes, *ctvA*, *ctvB*, *ctvC*, and *ctvD*, are sufficient to produce citreoviridin.¹¹⁹ Overexpression of the two genes *ctvA* and *ctvB* produced citreomontanin. In order to form the THF ring with the correct stereochemistry, the terminal alkene of citreomontanin with an *E*-16,17-alkene needs to undergo isomerisation to yield the *Z*-16,17 isomer, a step that could be catalysed by the flavin-dependent monooxygenase CtvC (Scheme 49). Bisepoxidation by CtvC then forms (17*R*,16*R*,15*S*,14*R*)-bisepoxide **211**. Addition of *ctvC* to the overexpression system generated a mixture of new products, with citreoviridin being the major one among other unidentified products (possibly due to spontaneous hydrolysis and degradation of the unstable bisepoxide intermediate). Finally, when *ctvD* was added to generate the *ctvABCD* overexpression strain, the only observed product was citreoviridin (Scheme 49).¹¹⁹

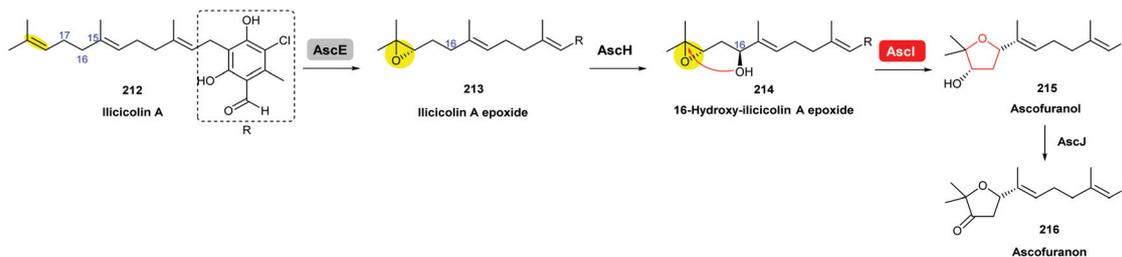
3.3.2 Ascofuranone. Ascofuranone is a meroterpenoid produced by various filamentous fungi including *Acremonium egyptiacum* and has been shown in recent studies to be a promising drug candidate against African trypanosomiasis and a potential anticancer lead compound.¹²⁰

Using gene knockout experiments and *in vitro* enzymatic turnover assays, Araki *et al.* demonstrated that AscE is a P450 monooxygenase that catalyses stereoselective epoxidation of the terminal olefin of ilicicolin A to produce ilicicolin A epoxide **213**, which is then hydroxylated by another P450 monooxygenase AscH to yield the 16-hydroxy-ilicicolin A epoxide intermediate **214** (Scheme 50). This epoxy alcohol **214**



Scheme 49 Proposed late stages biosynthesis of citreoviridin.¹¹⁹





Scheme 50 Proposed late-stage biosynthesis of ascofuranone.¹²⁰

is then cyclised by AscI to form the THF ring in ascofuranol **215**, and finally oxidation of the secondary alcohol to a ketone by AscJ delivers the product ascofuranone **216**. It is interesting to note that the 16-hydroxy-illicicolin A epoxide intermediate **214** can also be converted into ascofuranol nonenzymatically under acidic conditions. AscI shares 29% and 27% identities, respectively, to the epoxide hydrolases AurD and CtvD in aurovertin and citreoviridin biosynthesis (see previous sections).¹²⁰

3.3.3 Aurachin. Aurachins are a group of myxobacterial quinoline alkaloids produced by *Stigmatella aurantiaca* Sg a15 that exhibit antibacterial and antifungal among many other biological activities.¹²¹ The proposed late-stage biosynthesis of aurachin A involves epoxidation of the farnesyl side chain followed by an EH-catalysed epoxide-opening to form the THF ring product.¹²²

Based on isotopic feeding studies, gene inactivation experiments and bioinformatic analysis, a biosynthetic sequence has been proposed for the late stage transformation of aurachin B to aurachin A (Scheme 51).^{122,123} The FAD-dependent monooxygenase AuaJ is believed to catalyse epoxidation of the 2',3'-alkene on the farnesyl chain to give **217**. Intramolecular attack of the epoxide intermediate **217** by the 3-hydroxyl would then result in the epoxide-opening and cyclisation to afford the THF ring. The proposed epoxide intermediate **217** has not been detected from the cell extracts, which may be due to rapid turnover. Although this process was proposed to be catalysed by an epoxide hydrolase, the function of AuaI remains to be confirmed by further studies.

3.3.4 Penigequinolone/aspokinolone. Penigequinolones are insecticidal quinolone alkaloids produced by various *Penicillium* and *Aspergillus* species. Aspokinolones are closely related analogues of penigequinolones and the two pathways share most of their biosynthetic machinery (Scheme 52).¹²⁴

Yaequinolone C has been proven to be an off-pathway shunt product found during studies on penigequinolone bio-

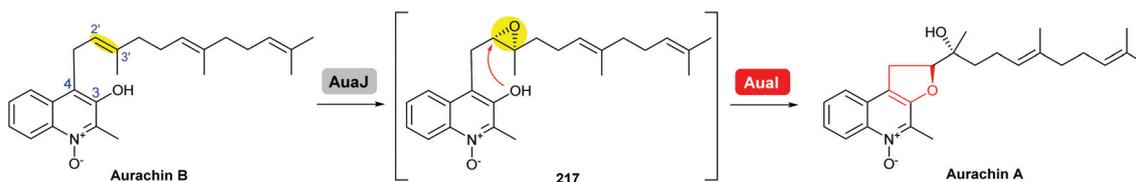
synthesis.¹²⁵ Formation of the THF ring in yaequinolone C has been shown by Zou *et al.* to result from epoxidation of the terminal olefin of the precursor **218** catalysed by the flavin-dependent monooxygenase PenE, followed by the PenJ-catalysed epoxide-opening of **219** to afford yaequinolone C (Scheme 52).¹²⁶ It was found that yaequinolone C could also be formed when the alkene precursor **218** was incubated with only PenE, suggesting the epoxide-opening step could happen spontaneously. When the biotransformation was performed with both PenE and PenJ, the conversion of the precursor **218** to yaequinolone C was significantly elevated (~10-fold), indicating the role of PenJ as an epoxide hydrolase in enhancing the rate of the 5-*exo* cyclisation step.¹²⁶

The corresponding flavin-dependent monooxygenase/epoxide hydrolase pair AsqG/AsqB in the aspoquinolone pathway have been shown to be functionally equivalent in their ability to catalyse the same transformations.¹²⁵ These two pairs of enzymes were also found to be cross compatible in mix-and-match experiments,¹²⁵ which is similar to the mupirocin/thiomarinol case.¹¹¹

In the presence of further enzymes, epoxide intermediate **219** undergoes further modifications to yield the final metabolites penigequinolones and aspoquinolones.¹²⁵

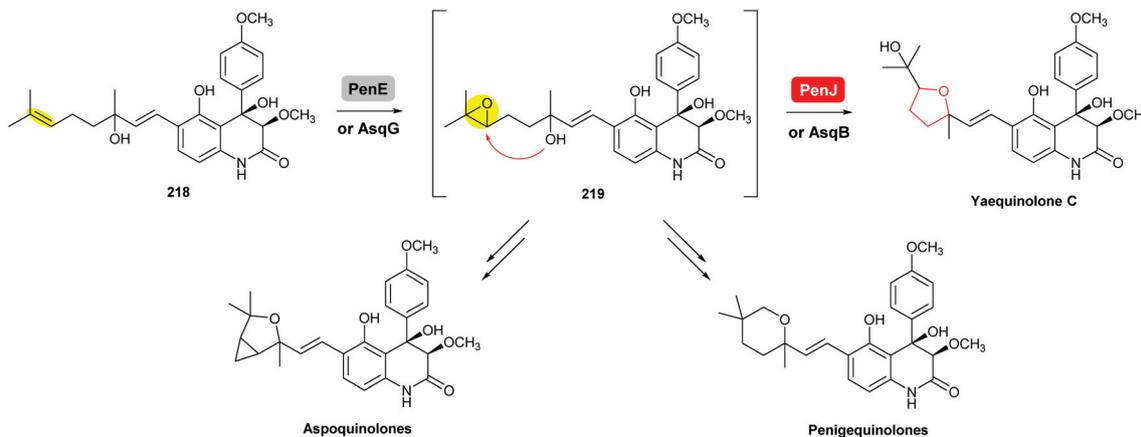
3.3.5 Monensin. Monensin A is a polyether ionophore isolated from *Streptomyces cinnamonensis* and has been widely used in veterinary medicine and in animal husbandry.¹²⁷ Formation of the THF rings in monensin have been shown to proceed through epoxidation of an *E,E,E*-triene precursor **220** followed by an epoxide-opening cascade reaction (Scheme 53).

Deletion of the *monCI* gene, which encodes a putative flavin-dependent epoxidase from the monensin biosynthetic gene cluster, led to accumulation of an *E,E,E*-triene precursor **220** in mutant strains of *Streptomyces cinnamonensis*, suggesting MonCI is responsible for formation of the triepoxide intermediate **221**.¹²⁷ Subsequent epoxide-opening of the

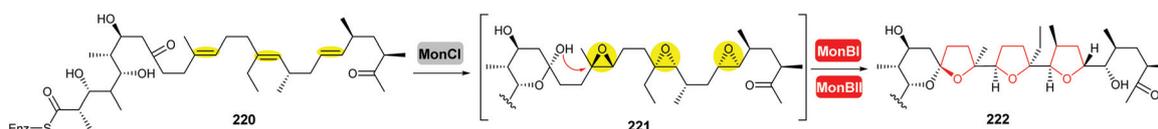


Scheme 51 Proposed late-stage biosynthesis of aurachin A.^{122,123}





Scheme 52 THF ring formation in yaequinolone C biosynthesis.^{125,126}



Scheme 53 THF ring formation in monensin biosynthesis.^{127,128}

trioxide intermediate was originally thought to be catalysed by MonCII but was later found to be controlled by the MonBI and MonBII epoxide hydrolases.¹²⁸ Using structurally simple substrate analogues, Oikawa and co-workers demonstrated remarkable synergistic effect between MonBI and MonBII in catalysing epoxide-opening cascade reactions.¹²⁹ When used alone in turnover reactions, MonBI was inactive and MonBII was only weakly active. However, the epoxide opening activity was dramatically enhanced with the addition of MonBI to the MonBII reaction mixture.¹²⁹

Other epoxide hydrolases that have been proposed to be involved in THF ring biosynthesis include SalBI/SalBII and from salinomycin pathway,¹³⁰ NigBI/NigBII from the nigericin pathway,¹³¹ NanI from the nanchangmycin,¹³² MadI from the maduramicin pathway,¹³³ and Pak24/Pak25 from the K-41A pathway.¹³⁴

3.4 Summary of biosynthetic methods

THP and THF rings are often essential structural moieties for bioactivities of many natural products and the formation of these oxygen heterocycles *via* epoxide formation/epoxide-opening cascade reactions has been demonstrated in a variety of bacterial and fungal natural product biosynthetic pathways. The epoxidation step is commonly catalysed by a flavin-dependent monooxygenase on an alkene substrate. However, interestingly in the mupirocin/thiomarinol systems, a Rieske non-haem oxygenase catalyses oxidation of a non-activated alkane precursor to an epoxide *via* an alkene. For regioselective intramolecular opening of epoxide biosynthetic intermediates, an epoxide hydrolase (EH) is often employed to yield the 6-*endo*

cyclisation THP product. Only three protein structures (Lsd19, MupZ and XimE) have been reported for these EHs so far and X-ray crystallographic studies have suggested a similar mechanism of general acid-base catalysis. In the cases of ambruticins and abyssomicins where EHs have not been identified, further studies would be required to determine whether their THP ring formation steps are under enzymatic or substrate control. Cyclisation of 4,5-epoxy alcohols to generate THFs has been shown to either occur spontaneously or to be mediated and significantly accelerated by EHs, although no protein structure has been reported for this type of EHs yet. These epoxidation/epoxide-opening cascade reactions are summarised in Table 5.

4. Future outlook

4.1 Combining chemical synthesis and biosynthesis

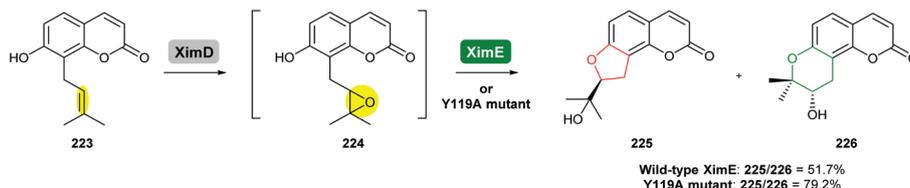
Despite advances in chemical methodology aided by computational studies, the total synthesis of complex natural products remains a challenging endeavour. Modular approaches to the total synthesis of target compounds often provide flexibility for the assembly of libraries of natural product analogues but may rely on the use of protecting groups, precious metals, toxic reagents and solvents whilst generating substantial amounts of unwanted by-products. On the other hand, enzyme-catalysed transformations offer clean and selective methods to generate target compounds but there can be issues to overcome in terms of narrow substrate specificities and scale-up. Hybrid strategies combining chemical



Table 5 THP and THF ring formation via epoxidation/epoxide-opening cascade reactions in natural product biosynthesis

Natural product	Strain	Substrate	Epoxidase	Epoxidase (PDB ID)	Product	Ref.
Lasaloid A	<i>Streptomyces lasaliensis</i>		LsdH8 (FMO)	Lsd19 (PDB ID: 3RGA)		104–107
Aurovertin	<i>Calcarisporium arbuscula</i>		AurC (FMO)	AurD		108
Mupirocin	<i>Pseudomonas fluorescens</i>		MupW (Rieske non-haem oxygenase)	MupZ (PDB ID: 6FXD)		110
Thiomarinol	<i>Pseudoalteromonas</i> sp.		TmlW (Rieske non-haem oxygenase)	TmlZ		111
Xiamenmycin	<i>Streptomyces xiamenensis</i>		XimD (FMO)	XimE (PDB ID: 6ISK)		112
Ambruticin	<i>Sorangium cellulosum</i>		AmbJ (FMO)	Not reported		114
Abyssomicin	<i>Streptomyces koyangensis</i>		AbmV (P450)	Not reported		118
Citreoviridin	<i>Penicillium</i> sp.		CtvC (FMO)	CtvD		119
Ascofuranone	<i>Acremonium egyptiacum</i>		AscE (P450)	AscI		120
Aurachin	<i>Stigmatella aurantiaca</i>		AuaJ (FMO)	AuaI		122
Penigequinolone	<i>Penicillium</i> & <i>Aspergillus</i> sp.		PenE (FMO)	PenJ		125 and 126
Aspoquinolone	<i>Aspergillus nidulans</i>		AsqG (FMO)	AsqB		126
Monensin	<i>Streptomyces cinnamonensis</i>		MonCI (FMO)	MonBI, MonBII		127–129





Scheme 54 Formation of pyranocoumarins by engineered XimE.¹¹³

synthesis and biotransformations continue to provide exciting new opportunities to gain efficient access to supplies of complex natural products and their analogues that are not accessible by individual synthetic or biotechnological approaches.¹³⁵

4.2 Enzyme engineering

In a world with rising demands for clean, efficient, and selective catalysts, biosynthetic enzymes will continue to provide a repertoire of powerful tools to generate complex molecules and their derivatives. However, the catalytic potential of these enzymes is still far from fully explored.¹³⁶

In a recent study, Xu and co-workers engineered XimE from xiamenmycin biosynthesis to improve its catalytic activity for the preparation of angular pyranocoumarins by site-directed mutagenesis (Scheme 54).¹¹³ Guided by the crystallographic structure of XimE and molecular docking, the Y119A mutant of XimE was generated. The proportion of THP product 226 to THF product 225 increased from 51.7% in wild-type XimE to 79.2% in the Y119A mutant.¹¹³

As further epoxide hydrolases are discovered, rational engineering of these enzymes to expand their substrate scope, enhance their catalytic activity and to possibly switch their regioselectivity in epoxide-opening process will be a fascinating area of research. For enzymes to be utilised in combination with organic synthesis, it will be necessary to optimise their stability, substrate scope and performance under a variety of conditions such as in the presence of organic solvents. We anticipate such efforts will tune and diversify the functions of these enzymes to provide biocatalysts that bring the benefits of nature's biosynthetic machinery to chemical synthesis.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We thank the EPSRC Bristol Chemical Synthesis Centre for Doctoral Training (EP/L015366/1) for a studentship to JIB and the BBSRC/EPSRC for funding through the Bristol Centre for Synthetic Biology (BB/L01386X/1) to LW.

References

- R. D. Taylor, M. MacCoss and A. D. G. Lawson, *J. Med. Chem.*, 2014, **57**, 5845.
- M. A. Perry, S. D. Rychnovsky and N. Sizemore, Synthesis of Saturated Oxygenated Heterocycles I, in *Topics in Heterocyclic Chemistry*, ed. J. Cossy, Springer, Berlin, Heidelberg, 2014, vol. 35, pp. 43–95.
- O. V. Salomatina, O. I. Yarovaya and V. A. Barkhash, *Russ. J. Org. Chem.*, 2005, **41**, 155.
- I. Larrosa, P. Romea and F. Urpí, *Tetrahedron*, 2008, **64**, 2683.
- I. Vilotijevic and T. Jamison, *Mar. Drugs*, 2010, **8**, 763.
- F. Vetica, P. Chauhan, S. Dochain and D. Enders, *Chem. Soc. Rev.*, 2017, **46**, 1661.
- C. Olier, M. Kaafarani, S. Gastaldi and M. P. Bertrand, *Tetrahedron*, 2010, **66**, 413.
- X. Han, G. Peh and P. E. Floreancig, *Eur. J. Org. Chem.*, 2013, **2013**, 1193.
- A. Deiters and S. F. Martin, *Chem. Rev.*, 2004, **104**, 2199.
- P. A. Clarke and S. Santos, *Eur. J. Org. Chem.*, 2006, **2006**, 2045.
- N. M. Nasir, K. Ermanis and P. A. Clarke, *Org. Biomol. Chem.*, 2014, **12**, 3323.
- K. Fujiwara and A. Murai, *Bull. Chem. Soc. Jpn.*, 2004, **77**, 2129.
- H. Fuwa, *Mar. Drugs*, 2016, **14**, 65.
- F. Hemmerling and F. Hahn, *Beilstein J. Org. Chem.*, 2016, **12**, 1512.
- S. Meng, G.-L. Tang and H.-X. Pan, *ChemBioChem*, 2018, **19**, 2002.
- M.-C. Tang, Y. Zou, K. Watanabe, C. T. Walsh and Y. Tang, *Chem. Rev.*, 2017, **117**, 5226.
- J. E. Baldwin, *J. Chem. Soc., Chem. Commun.*, 1976, 734.
- I. V. Alabugin and K. Gilmore, *Chem. Commun.*, 2013, **49**, 11246.
- K. Gilmore, R. K. Mohamed and I. V. Alabugin, *Wiley Interdiscip. Rev.: Comput. Mol. Sci.*, 2016, **6**, 487.
- J. Na, K. N. Houk, C. G. Shevlin, K. D. Janda and R. A. Lerner, *J. Am. Chem. Soc.*, 1993, **115**, 8453.
- I. Vilotijevic and T. F. Jamison, *Angew. Chem., Int. Ed.*, 2009, **48**, 5250.
- K. C. Nicolaou, M. E. Duggan, C.-K. Hwang and P. K. Somers, *J. Chem. Soc., Chem. Commun.*, 1985, **111**, 1359.
- K. C. Nicolaou, C. V. C. Prasad, P. K. Somers and C. K. Hwang, *J. Am. Chem. Soc.*, 1989, **111**, 5330.



- 24 J. Coxon, M. Hartshorn and W. Swallow, *Aust. J. Chem.*, 1973, **26**, 2521.
- 25 I. Kadota, H. Ueno and Y. Yamamoto, *Tetrahedron Lett.*, 2003, **44**, 8935.
- 26 K. Sato and M. Sasaki, *Angew. Chem., Int. Ed.*, 2007, **46**, 2518.
- 27 P. A. Evans and V. S. Murthy, *Tetrahedron Lett.*, 1999, **40**, 1253.
- 28 P. Neogi, T. Doundoulakis, A. Yazbak, S. C. Sinha, S. C. Sinha and E. Keinan, *J. Am. Chem. Soc.*, 1998, **120**, 11279.
- 29 M. Kanemoto, M. Murata and T. Oishi, *J. Org. Chem.*, 2009, **74**, 8810.
- 30 Y. Sakamoto, G. Matsuo, H. Matsukura and T. Nakata, *Org. Lett.*, 2001, **3**, 2749.
- 31 H. Fukui and I. Shiina, *Org. Lett.*, 2008, **10**, 3153.
- 32 R. K. Bressin, S. Osman, I. Pohorilets, U. Basu and K. Koide, *J. Org. Chem.*, 2020, **85**, 4637.
- 33 H. Matsukura, M. Morimoto, H. Koshino and T. Nakata, *Tetrahedron Lett.*, 1997, **38**, 5545.
- 34 T. Nakata, *J. Synth. Org. Chem., Jpn.*, 1998, **56**, 940.
- 35 H. Matsukura, N. Hori, G. Matsuo and T. Nakata, *Tetrahedron Lett.*, 2000, **41**, 7681.
- 36 M. Morimoto, H. Matsukura and T. Nakata, *Tetrahedron Lett.*, 1996, **37**, 6365.
- 37 B. M. Trost and A. Tenaglia, *Tetrahedron Lett.*, 1988, **29**, 2927.
- 38 T. Suzuki, O. Sato and M. Hiram, *Tetrahedron Lett.*, 1990, **31**, 4747.
- 39 T. Oishi, K. Maeda and M. Hiram, *Chem. Commun.*, 1997, 1289.
- 40 T. Suzuki, O. Sato, M. Hiram, Y. Yamamoto, M. Murata, T. Yasumoto and N. Harada, *Tetrahedron Lett.*, 1991, **32**, 4505.
- 41 J. D. Ha, E. Y. Shin, S. K. Kang, J. H. Ahn and J.-K. Choi, *Tetrahedron Lett.*, 2004, **45**, 4193.
- 42 M. Inoue, F. Saito, M. Iwatsu, Y. Ishihara and M. Hiram, *Tetrahedron Lett.*, 2007, **48**, 2171.
- 43 K. W. Armbrust, M. G. Beaver and T. F. Jamison, *J. Am. Chem. Soc.*, 2015, **137**, 6941.
- 44 C. Mukai, Y. Ikeda, Y. Sugimoto and M. Hanaoka, *Tetrahedron Lett.*, 1994, **35**, 2179.
- 45 B. J. Teobald, *Tetrahedron*, 2002, **58**, 4133.
- 46 C. Mukai, N. Miyakoshi and M. Hanaoka, *J. Org. Chem.*, 2001, **66**, 5875.
- 47 C. Mukai, Y. Sugimoto, Y. Ikeda and M. Hanaoka, *Tetrahedron Lett.*, 1994, **35**, 2183.
- 48 I. C. González and C. J. Forsyth, *J. Am. Chem. Soc.*, 2000, **122**, 9099.
- 49 C. Mukai, S. Yamaguchi, I. J. Kim and M. Hanaoka, *Chem. Pharm. Bull.*, 2001, **49**, 61.
- 50 F. R. P. Crisóstomo, T. Martín and V. S. Martín, *Org. Lett.*, 2004, **6**, 565.
- 51 P. F. Hudrlik, J. P. Arcoleo, R. H. Schwartz, R. N. Misra and R. J. Rona, *Tetrahedron Lett.*, 1977, **18**, 591.
- 52 W. E. Fristad, T. R. Bailey, L. A. Paquette, R. Gleiter and M. C. Boehm, *J. Am. Chem. Soc.*, 1979, **101**, 4420.
- 53 T. Ikeno, J. Harada and S. Tomoda, *Chem. Lett.*, 1999, **28**, 409.
- 54 G. Adiwidjaja, H. Flörke, A. Kirschning and E. Schaumann, *Tetrahedron Lett.*, 1995, **36**, 8771.
- 55 T. P. Heffron and T. F. Jamison, *Org. Lett.*, 2003, **5**, 2339.
- 56 G. L. Simpson, T. P. Heffron, E. Merino and T. F. Jamison, *J. Am. Chem. Soc.*, 2006, **128**, 1056.
- 57 T. P. Heffron, G. L. Simpson, E. Merino and T. F. Jamison, *J. Org. Chem.*, 2010, **75**, 2681.
- 58 J. Lenhof, M. Hutter, V. Huch and J. Jauch, *Eur. J. Org. Chem.*, 2020, **2020**, 5833.
- 59 Y. Mori, K. Yaegashi and H. Furukawa, *J. Am. Chem. Soc.*, 1996, **118**, 8158.
- 60 Y. Mori, *Chem. Pharm. Bull.*, 2019, **67**, 1.
- 61 Y. Mori, H. Furuta, T. Takase, S. Mitsuoka and H. Furukawa, *Tetrahedron Lett.*, 1999, **40**, 8019.
- 62 Y. Mori, K. Yaegashi and H. Furukawa, *Tetrahedron Lett.*, 1999, **40**, 7239.
- 63 Y. Mori, K. Yaegashi and H. Furukawa, *J. Am. Chem. Soc.*, 1997, **119**, 4557.
- 64 Y. Mori, K. Nogami, H. Hayashi and R. Noyori, *J. Org. Chem.*, 2003, **68**, 9050.
- 65 M. Inoue, S. Yamashita, A. Tatami, K. Miyazaki and M. Hiram, *J. Org. Chem.*, 2004, **69**, 2797.
- 66 H. Furuta, Y. Hasegawa and Y. Mori, *Org. Lett.*, 2009, **11**, 4382.
- 67 T. Sakai, A. Ishihara and Y. Mori, *J. Org. Chem.*, 2017, **82**, 3976.
- 68 T. Sakai, S. Matsushita, S. Arakawa, K. Mori, M. Tanimoto, A. Tokumasu, T. Yoshida and Y. Mori, *J. Am. Chem. Soc.*, 2015, **137**, 14513.
- 69 F. E. McDonald, X. Wang, B. Do and K. I. Hardcastle, *Org. Lett.*, 2000, **2**, 2917.
- 70 C. J. Morten and T. F. Jamison, *J. Am. Chem. Soc.*, 2009, **131**, 6678.
- 71 F. Bravo, F. E. McDonald, W. A. Neiwert, B. Do and K. I. Hardcastle, *Org. Lett.*, 2003, **5**, 2123.
- 72 M. A. Tarselli, J. L. Zuccarello, S. J. Lee and M. R. Gagné, *Org. Lett.*, 2009, **11**, 3490.
- 73 A. Zakarian, A. Batch and R. A. Holton, *J. Am. Chem. Soc.*, 2003, **125**, 7822.
- 74 F. Li, S. Ren, P. Li, P. Yang and J. Qu, *Angew. Chem., Int. Ed.*, 2020, **59**, 18473.
- 75 Y. Morimoto, Y. Nishikawa, C. Ueba and T. Tanaka, *Angew. Chem., Int. Ed.*, 2006, **45**, 810.
- 76 Y. Morimoto, Y. Nishikawa and M. Takaishi, *J. Am. Chem. Soc.*, 2005, **127**, 5806.
- 77 Y. Morimoto, H. Yata and Y. Nishikawa, *Angew. Chem., Int. Ed.*, 2007, **46**, 6481.
- 78 S. Wang, A. J. Arguelles, J. Tay, M. Hotta, P. M. Zimmerman and P. Nagorny, *Chem. – Eur. J.*, 2020, **26**, 4583.
- 79 M. H. Wu, K. B. Hansen and E. N. Jacobsen, *Angew. Chem., Int. Ed.*, 1999, **38**, 2012.



- 80 K. B. Hansen, J. L. Leighton and E. N. Jacobsen, *J. Am. Chem. Soc.*, 1996, **118**, 10924.
- 81 A. Blanc and F. D. Toste, *Angew. Chem., Int. Ed.*, 2006, **45**, 2096.
- 82 I. Vilotijevic and T. F. Jamison, *Science*, 2007, **317**, 1189.
- 83 C. J. Morten, J. A. Byers, A. R. Van Dyke, I. Vilotijevic and T. F. Jamison, *Chem. Soc. Rev.*, 2009, **38**, 3175.
- 84 Z. Wang, Y.-T. Cui, Z.-B. Xu and J. Qu, *J. Org. Chem.*, 2008, **73**, 2270.
- 85 J. A. Byers and T. F. Jamison, *J. Am. Chem. Soc.*, 2009, **131**, 6383.
- 86 J. A. Byers and T. F. Jamison, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 16724.
- 87 A. R. Van Dyke and T. F. Jamison, *Angew. Chem., Int. Ed.*, 2009, **48**, 4430.
- 88 L. C. Czabaniuk and T. F. Jamison, *Org. Lett.*, 2015, **17**, 774.
- 89 K. Janda, C. Shevlin and R. Lerner, *Science*, 1993, **259**, 490.
- 90 J. Na, K. N. Houk, C. G. Shevlin, K. D. Janda and R. A. Lerner, *J. Am. Chem. Soc.*, 1993, **115**, 8453.
- 91 K. Gruber, B. Zhou, K. N. Houk, R. A. Lerner, C. G. Shevlin and I. A. Wilson, *Biochemistry*, 1999, **38**, 7062.
- 92 G. Chiosis, *Tetrahedron Lett.*, 2000, **41**, 801.
- 93 X. Zhang, X. Hao, L. Liu, A.-T. Pham, J. López-Andarias, A. Frontera, N. Sakai and S. Matile, *J. Am. Chem. Soc.*, 2018, **140**, 17867.
- 94 M. Paraja and S. Matile, *Angew. Chem., Int. Ed.*, 2020, **59**, 6273.
- 95 T. Nakata, G. Schmid, B. Vranesic, M. Okigawa, T. Smith-Palmer and Y. Kishi, *J. Am. Chem. Soc.*, 1978, **100**, 2933.
- 96 T. Nakata, S. Nomura and H. Matsukura, *Tetrahedron Lett.*, 1996, **37**, 213.
- 97 K. Nagasawa, N. Hori, R. Shiba and T. Nakata, *Heterocycles*, 1997, **44**, 105.
- 98 T. Nakata, S. Nomura, H. Matsukura and M. Morimoto, *Tetrahedron Lett.*, 1996, **37**, 217.
- 99 N. Hori, K. Nagasawa, T. Shimizu and T. Nakata, *Tetrahedron Lett.*, 1999, **40**, 2145.
- 100 F. E. McDonald and M. A. Smart, *Heterocycles*, 2012, **86**, 535.
- 101 M. Caron and K. B. Sharpless, *J. Org. Chem.*, 1985, **50**, 1557.
- 102 K. Fujiwara, T. Tokiwano and A. Murai, *Tetrahedron Lett.*, 1995, **36**, 8063.
- 103 T. Tokiwano, K. Fujiwara and A. Murai, *Synlett*, 2000, 335.
- 104 Y. Shichijo, A. Migita, H. Oguri, M. Watanabe, T. Tokiwano, K. Watanabe and H. Oikawa, *J. Am. Chem. Soc.*, 2008, **130**, 12230.
- 105 A. Minami, M. Shimaya, G. Suzuki, A. Migita, S. S. Shinde, K. Sato, K. Watanabe, T. Tamura, H. Oguri and H. Oikawa, *J. Am. Chem. Soc.*, 2012, **134**, 7246.
- 106 L. Smith, H. Hong, J. B. Spencer and P. F. Leadlay, *ChemBioChem*, 2008, **9**, 2967.
- 107 K. Hotta, X. Chen, R. S. Paton, A. Minami, H. Li, K. Swaminathan, I. I. Mathews, K. Watanabe, H. Oikawa, K. N. Houk and C.-Y. Kim, *Nature*, 2012, **483**, 355.
- 108 X.-M. Mao, Z.-J. Zhan, M. N. Grayson, M.-C. Tang, W. Xu, Y.-Q. Li, W.-B. Yin, H.-C. Lin, Y.-H. Chooi, K. N. Houk and Y. Tang, *J. Am. Chem. Soc.*, 2015, **137**, 11904.
- 109 C. M. Thomas, J. Hothersall, C. L. Willis and T. J. Simpson, *Nat. Rev. Microbiol.*, 2010, **8**, 281.
- 110 L. Wang, A. Parnell, C. Williams, N. A. Bakar, M. R. Challand, M. W. van der Kamp, T. J. Simpson, P. R. Race, M. P. Crump and C. L. Willis, *Nat. Catal.*, 2018, **1**, 968.
- 111 L. Wang, Z. Song, P. R. Race, J. Spencer, T. J. Simpson, M. P. Crump and C. L. Willis, *Chem. Sci.*, 2020, **11**, 5221.
- 112 B. B. He, T. Zhou, X. L. Bu, J. Y. Weng, J. Xu, S. Lin, J. T. Zheng, Y. L. Zhao and M. J. Xu, *ACS Catal.*, 2019, 5391.
- 113 X. L. Bu, B. B. He, J. Y. Weng, C. C. Jiang, Y. L. Zhao, S. M. Li, J. Xu and M. J. Xu, *ACS Synth. Biol.*, 2020, **9**, 2282.
- 114 B. Julien, Z.-Q. Tian, R. Reid and C. D. Reeves, *Chem. Biol.*, 2006, **13**, 1277.
- 115 J. I. Bowen, L. Wang, M. P. Crump and C. L. Willis, *Org. Biomol. Chem.*, 2021, **19**, 6210.
- 116 K. Trentadue, C. Chang, A. Nalin and R. E. Taylor, *Chem. – Eur. J.*, 2021, **27**, 11126.
- 117 C. Sadaka, E. Ellsworth, P. Hansen, R. Ewin, P. Damborg and J. Watts, *Molecules*, 2018, **23**, 1371.
- 118 Q. Li, W. Ding, Z. Yao, J. Tu, L. Wang, H. Huang, S. Li and J. Ju, *Org. Lett.*, 2018, **20**, 4854.
- 119 T. S. Lin, Y. M. Chiang and C. C. C. Wang, *Org. Lett.*, 2016, **18**, 1366.
- 120 Y. Araki, T. Awakawa, M. Matsuzaki, R. Cho, Y. Matsuda, S. Hoshino, Y. Shinohara, M. Yamamoto, Y. Kido, D. K. Inaoka, K. Nagamune, K. Ito, I. Abe and K. Kita, *Proc. Natl. Acad. Sci. U. S. A.*, 2019, **116**, 8269.
- 121 Y. Katsuyama, K. Harmrolfs, D. Pistorius, Y. Li and R. Müller, *Angew. Chem., Int. Ed.*, 2012, **51**, 9437.
- 122 D. Pistorius, Y. Li, A. Sandmann and R. Müller, *Mol. Biosyst.*, 2011, **7**, 3308.
- 123 G. Höfle and B. Kunze, *J. Nat. Prod.*, 2008, **71**, 1843.
- 124 Y. Matsuda and I. Abe, *Nat. Prod. Rep.*, 2016, **33**, 26.
- 125 Y. Zou, M. Garcia-Borràs, M. C. Tang, Y. Hirayama, D. H. Li, L. Li, K. Watanabe, K. N. Houk and Y. Tang, *Nat. Chem. Biol.*, 2017, **13**, 325.
- 126 Y. Zou, Z. Zhan, D. Li, M. Tang, R. A. Cacho, K. Watanabe and Y. Tang, *J. Am. Chem. Soc.*, 2015, **137**, 4980.
- 127 A. Bhatt, C. B. W. Stark, B. M. Harvey, A. R. Gallimore, Y. A. Demydchuk, J. B. Spencer, J. Staunton and P. F. Leadlay, *Angew. Chem., Int. Ed.*, 2005, **44**, 7075.
- 128 A. R. Gallimore, C. B. W. Stark, A. Bhatt, B. M. Harvey, Y. Demydchuk, V. Bolanos-Garcia, D. J. Fowler, J. Staunton, P. F. Leadlay and J. B. Spencer, *Chem. Biol.*, 2006, **13**, 453.
- 129 K. Sato, A. Minami, T. Ose, H. Oguri and H. Oikawa, *Tetrahedron Lett.*, 2011, **52**, 5277.



- 130 M. E. Yurkovich, P. A. Tyrakis, H. Hong, Y. Sun, M. Samborsky, K. Kamiya and P. F. Leadlay, *ChemBioChem*, 2012, **13**, 66.
- 131 B. M. Harvey, T. Mironenko, Y. Sun, H. Hong, Z. Deng, P. F. Leadlay, K. J. Weissman and S. F. Haydock, *Chem. Biol.*, 2007, **14**, 703.
- 132 Y. Sun, X. Zhou, H. Dong, G. Tu, M. Wang, B. Wang and Z. Deng, *Chem. Biol.*, 2003, **10**, 431.
- 133 R. Liu, F. Fang, Z. An, R. Huang, Y. Wang, X. Sun, S. Fu, A. Fu, Z. Deng and T. Liu, *J. Ind. Microbiol. Biotechnol.*, 2020, **47**, 275.
- 134 J. Chen, J. Chen, C. Gui, C. Gui, Q. Wei, J. Liu, L. Ye, X. Tian, Y. C. Gu, Q. Li, J. Ju and J. Ju, *Org. Lett.*, 2020, **22**, 4627.
- 135 F. Hahn and F. M. Guth, *Nat. Prod. Rep.*, 2020, **37**, 1300.
- 136 K. Chen and F. H. Arnold, *Nat. Catal.*, 2020, **3**, 203.

