



Cite this: *Mater. Adv.*, 2022, 3, 4765

Eco-friendly synthesis of carbon nanotubes and their cancer theranostic applications

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Carbon nanotubes (CNTs) with attractive physicochemical characteristics such as high surface area, mechanical strength, functionality, and electrical/thermal conductivity have been widely studied in different fields of science. However, the preparation of these nanostructures on a large scale is either expensive or sometimes ecologically unfriendly. In this context, plenty of studies have been conducted to discover innovative methods to fabricate CNTs in an eco-friendly and inexpensive manner. CNTs have been synthesized using various natural hydrocarbon precursors, including plant extracts (e.g., tea-tree extract), essential oils (e.g., eucalyptus and sunflower oil), biodiesel, milk, honey, and eggs, among others. Additionally, agricultural bio-wastes have been widely studied for synthesizing CNTs. Researchers should embrace the usage of natural and renewable precursors as well as greener methods to produce various types of CNTs in large quantities with the advantages of cost-effectiveness and environmentally benign features. In addition, multifunctionalized CNTs with improved biocompatibility and targeting features are promising candidates for cancer theranostic applications owing to their attractive optical, chemical, thermal, and electrical properties. This perspective discusses the recent developments in eco-friendly synthesis of CNTs using green chemistry-based techniques, natural renewable resources, and sustainable catalysts, with emphasis on important challenges and future perspectives and highlighting techniques for the functionalization or modification of CNTs. Significant and promising cancer theranostic applications as well as their biocompatibility and cytotoxicity issues are also discussed.

Received 24th March 2022,
Accepted 14th May 2022

DOI: 10.1039/d2ma00341d

rsc.li/materials-advances

1. Introduction

Single-walled (SW) and multi-walled (MW) carbon nanotubes (CNTs) are two main categories of CNTs, based on the number of graphene sheets comprising the cylindrical tubes. These cylindrical nanostructures can be employed for various pharmaceutical and biomedical applications owing to their distinct physicochemical properties and multiple functionalization capabilities (Fig. 1). CNTs exhibit attractive interfacial area with cellular membrane, multifunctional surfaces, and needle-like shapes, making them promising candidates for delivery of

therapeutic and diagnostic agents.^{1,2} In this context, various natural resources and (bio)renewable materials such as plant biomass or animal bioproducts can be employed for the fabrication of nanostructures like CNTs.³ However, more explorations are still needed on green chemistry-based solutions to reduce or discontinue the use of toxic and harmful agents with environmental and human hazards. The greener synthesis and functionalization of CNT-based nanostructures are in their infancy stages, and some challenges such as optimization of conditions, biocompatibility improvements, (bio)renewability, biodegradation, pharmacokinetics, isolation/purification, toxicity/biosafety, and *in vivo/in vitro* analyses need to be systematically studied to obtain CNTs with higher purity standards and up-scalable potentials;^{4–7} these may be equally applicable to other carbon-based nanostructures such as graphene, graphene oxide, fullerenes, and nanodiamonds, among others.^{8,9} The standardization procedures are vital in transforming lab-scale into industrial and up-scalable production, taking into account environmentally friendly aspects and green chemistry principles.

CNTs have been synthesized using assorted techniques like laser-ablation, spray pyrolysis, chemical vapor deposition, carbon arc discharge, and flame synthesis, among others.¹⁰ These synthesis methods have some limitations and drawbacks,

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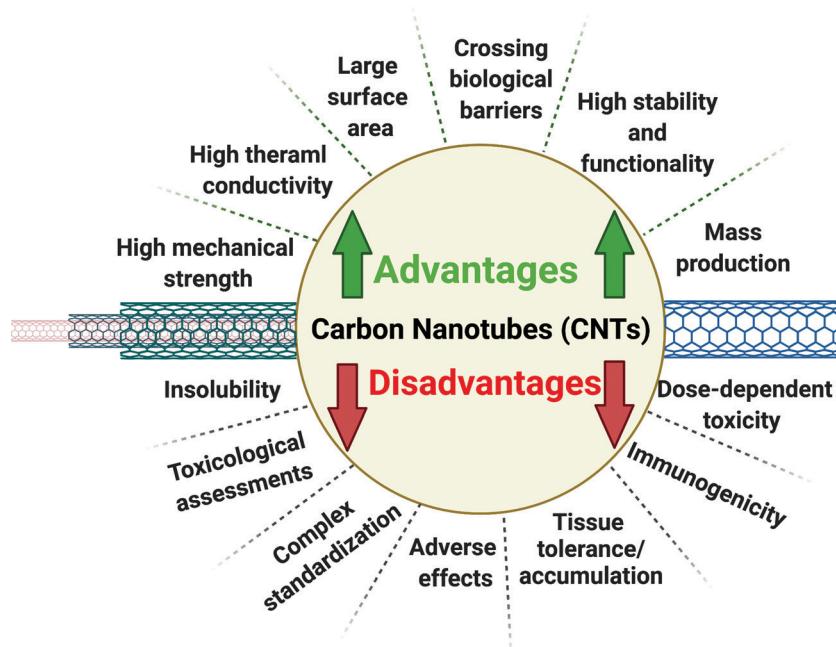


Fig. 1 Some salient advantages and limitations of CNTs.

such as dependence on expensive equipment, chemical catalysts, or high energy consumption, and inclusion of toxic chemical impurities in the ensuing CNTs.^{11,12} On the other hand, the application of greener techniques for the fabrication of CNTs with suitable properties has been explored to avoid the deployment of toxic/hazardous chemical materials and complicated instruments. Researchers have studied the utilization of low-cost and sustainable raw agricultural materials and biowastes to synthesize carbon-based nanostructures *via* simple and greener methods by preventing further pollutions/contaminants. For environmentally benign synthesis of CNTs with different sizes and structures, sustainable and natural carbon feedstocks/resources, sustainable catalysts, eco-friendly synthesis techniques should be applied, and purification procedures and gas emissions need to be within the tenets of the green chemistry principles.¹³

CNTs with good penetrability, nano-needle shapes, hollow monolithic structures, optico-electrical properties, and high drug loading capacity have shown promising applicability for targeted cancer therapy, photothermal ablation, and drug/gene delivery.^{14,15} These nanostructures with their unique sizes and morphologies as well as their high stability can be considered as attractive non-polymeric candidates for cancer nanotheranostics and anticancer nanocarriers applications.^{16,17} Also, their ability to easily penetrate cell membranes has provided an opportunity to consider them as efficient nanocarriers for drug delivery systems and theranostics.¹⁸ CNTs can effectively cross the biological barriers, allowing their applications in the delivery of therapeutically active molecules. The interaction between CNTs and cells is one of the important factors for evaluating their future biomedical potentials. Functionalized CNTs have exhibited a great potential to be taken up by a wide range of cells and can intracellularly traffic *via* different cellular

barriers.¹⁹ Besides, unique mechanical, thermal, optical, and electric features such as good heat conduction, tensile strength, and flexibility make CNTs different from the other carbon-based nanomaterials.²⁰ CNTs have also been explored for designing a variety of magnetic resonance imaging (MRI) contrast agents *via* the incorporation of gadolinium chelates.²¹ These carbon nanomaterials can also serve in cell imaging due to fluorescence in the near-infrared region; they do not affect the viability of the diagnosed cells. The strong absorption of CNTs in near-infrared regions can enable their additional deployments in photothermal therapy.²² These nanomaterials can transform the laser energy to acoustic signals and show excellent resonant Raman scattering and photoluminescence in near-infrared region, offering great opportunities in cancer diagnostics.²³ One of the main differences between SWCNTs and MWCNTs is the suitable electrical features of the former; the properties of MWCNTs mainly depend on the number of layers.²¹ However, some disadvantages/limitations namely the poor solubility and dispersibility of CNTs need to be surmounted for their biological and biomedical applications.^{24,25}

Surface multifunctionalization or modification tactics have been studied for improving the solubility/dispersibility and biocompatibility of CNTs.²⁶⁻²⁸ Thus, the biocompatibility of CNTs can be improved and their toxicity can be reduced by applying various functional groups *via* covalent or non-covalent bindings;²⁹ the covalent functionalization create strong binding of groups with suitable biocompatibility on the surface of CNTs.^{30,31} Herein, recent advances on the synthesis of various types of CNTs have been covered with a focus on eco-friendly and sustainable synthesis techniques, natural renewable feedstocks, and green catalysts. The functionalization or modification of CNTs based on eco-friendly protocols are discussed as



well as recent advances related to their cancer theranostic applications, focusing on important challenges and future perspectives encompassing biocompatibility and toxicity issues. The importance of sustainable methods with the explicit reduction in the usage of hazardous chemical agents and biocompatibility enhancement of ensued products is felt more than before. In the general domain of CNTs, however, there is a long way to go before realizing the benefits of greener synthesis and functionalization of these nanostructures.

2. Eco-friendly synthesis and functionalization

Greener synthesis and functionalization techniques can minimize the deployment of hazardous/toxic agents and expensive and rare materials/equipment to avoid adverse environmental impacts.³² Catalytic procedure has been broadly studied for increasing the yield of production and reducing the reaction temperature and time.³³ The chemical vapor deposition technique with large-scale production potentials has been typically initiated by decomposition of hydrocarbon gases or organic solvents, when the nano-sized catalysts are generated in a heated furnace. However, this technique may suffer from high energy consumption and toxic/hazardous chemicals utilization; several techniques such as electrostatic spray-assisted or combustion chemical vapor deposition have been studied wherein safer reagents can be utilized. *Via* the optimization of the reaction conditions and applying renewable precursors/catalysts, different types of CNTs with unique properties can be achieved.³⁴

2.1. Microwave (MW)-assisted synthesis

Several techniques such as conventional pyrolysis, hydrothermal carbonization pretreatment, cyclic oxidation, combustion method, and chemical vapor deposition have been reported for the synthesis of carbon materials from biomass, but are beleaguered by some noticeable disadvantages of high temperature usage, complex processes/intricacy of scale-up, low yield/efficiency, problems in process control, and likelihood of contaminations and emission of air pollutants during the synthesis procedure.^{35–38} On the other hand, the safer synthesis of CNTs, and the exact growth mechanisms of these ensued nanostructures from natural precursors/catalysts have been not entirely and analytically investigated.^{39,40} Microwave (MW)-assisted production approaches with mass production capabilities can be deployed for the assembly of CNTs with added benefits of lower cost, fast reaction time, simple/time-saving purification, and eco-friendliness;⁴¹ MW plasma chemical vapor deposition technique with advantages of non/low pollutions, significant reactivity, rapid heating, and controllability has garnered attention in the fabrication of CNTs.⁴² Despite these advantages, it still faces some important challenges regarding the quality of obtained CNTs under atmospheric pressure and low temperature that should be tuned. The effects of crucial factors such as the reacting gases (*e.g.*, H₂ pretreatment), nitrogen-containing additive (nitrogen-comprising gas), and appropriate MW power should

be analytically evaluated.^{38,43} For instance, it was indicated that the higher ionization degree could be attained with elevated MW power; additional carbon atoms could be localized for thickening of CNTs.³⁸ With an enhancement in the MW power and the subsequent increase in substrate temperature, the agglomeration of catalyst particles can transpire, thus culminating in the formation of CNTs with larger diameters. Nonetheless, in some instances, longer and thinner CNTs could be obtained by increasing the MW power due to the formation of significant amount of etching species (*e.g.*, atomic hydrogen in the plasma) in the presence of too high MW power; outer walls of CNTs were etched away, thus reducing the diameter of ensued CNTs.⁴⁴

MWCNTs (~10–40 nm) were synthesized *via* a MW-assisted technique with advantages of improved specific surface area in CNTs and enhanced yield of their production, providing optimal and inexpensive synthesis possibilities;⁴¹ MW irradiation has been deployed for the synthesis of CNTs with fewer imperfections compared to the conventional heating techniques.⁴⁵ The effortless in-core heating of carbon-based sources can be performed under a MW field to allow their distortion *via* MW heating for acquiring newer carbon-based structures (*e.g.*, CNTs).⁴⁶ MWCNTs were rapidly synthesized on a graphite surface with ferrocene (as a catalyst),⁴⁷ where deployment of MW technique highlighted the benefits of low temperature, and shorter reaction time, for decomposition of the ferrocene powder on abundant iron as catalyst and hydrocarbons (carbon source) to obtain CNTs.⁴⁷

MWCNTs on biochar substrates were synthesized *via* MW-assisted chemical vapor deposition technique at 600 °C by applying nickel as a catalyst; temperature was much lower than the routine chemical vapor deposition techniques. The carbon-containing species found in the volatiles probably served as carbon sources originating the growth of CNTs on the biochar surfaces, wherein inorganic species in biomass structures functioned as catalysts.⁴⁸ Further, gumwood biomass was employed to synthesize MWCNTs (~50 nm) *via* a MW-induced technique, in which char particles and minerals (in char particles) represented as substrates and catalysts, respectively.⁴⁹ The volatiles underwent the thermal and/or catalytic cracking procedure on the surface of char, generating the self-assembled amorphous carbon nanospheres onto MWCNTs structures under the MW irradiation conditions.⁴⁹ Besides, the dissociation of biomass and formation of carbon nanomaterials significantly depended on the temperature and pressure. For instance, the rice husk powders produced the biogas including CH₄ when they were irradiated by MW (~200 °C) and plasma irradiation. This temperature could sufficiently crack the pyrolysis of CH₄ to generate amorphous carbon layers as nucleation sites on the surface of nickel catalyst.⁵⁰ Notably, physicochemical characteristics of CNTs can be affected by production temperature of biomass. As an example, CNTs were synthesized utilizing biochar (the precursor) under the MW irradiation wherein higher concentration of CNTs with smaller hydrodynamic diameter could be obtained from biochars prepared at 600 °C. Also, the prepared CNTs from biochar of wheat straw and



hazelnut hulls exhibited a higher degree of wall graphitization, thus signifying high quality of CNTs.⁵¹

Typically, the heat transfer in conventional heating includes the process of conduction, convection, and radiation, but MW-assisted techniques stimulate the volumetric heating of the feedstock through bulk energy transfer. Additionally, the crucial advantages of MW-induced heating over conventional heating comprise volumetric/selective heating, low material pre-processing costs, uniform distribution of temperature, fast startup/shut down processes, shorter processing time, rapid heating and non-contact heating, low heat losses, and high efficiency of energy.^{20,22,23} In one study, MWCNTs were synthesized using MW-assisted pyrolysis technique in a bench scale pyrolysis reactor at 600 W power level which reached up to a temperature of 500 °C. MW pyrolysis of bagasse was performed with and without iron (Fe) and cobalt (Co) as susceptors/catalysts. As a result, by inserting Fe, high yield of H₂ and CH₄ gases could be obtained in addition to CO₂ and CO.⁵² Compared to the conventional high temperature techniques (800–1000 °C) utilizing graphite/carbon fibers or hydrocarbon gases as precursors for manufacturing CNTs, MW-assisted technique featured relatively low temperature (400–500 °C) at medium MW power (600 W).⁵² Further, in this study, the authors stated that high temperature furnace treatment and chemical vapor deposition methods yielded better quality and finer MWCNTs (~10–20 nm) compared to the MW-assisted pyrolysis technique. But, MW-assisted pyrolysis exhibited some important advantages such as short processing time (<15 min) and necessity of moderately low process temperatures. It appears that more elaborate studies are required for producing CNTs in high yields from lignocellulosic biomass *via* MW pyrolysis.⁵² Some important examples of CNTs prepared *via* MW-assisted chemical vapor deposition and MW pyrolysis techniques with advantages of uniform heating, higher yields, and improved mass transfer of volatiles are summarized in Table 1.

MW-assisted techniques can be deployed for converting renewable biomass resources to value-added CNTs with industrial potential. However, the mechanism of formation and transformation of biomass molecular structures into CNTs, *e.g.*, the conversion of cellulose into CNTs and its interaction with MW, the evaluation of elemental composition of biomass, in-depth study on optimization and scale-up process (yield

optimization and establishment of the MW heating efficiency), and the selection of appropriate catalysts need optimization in future to improve the present techniques for synthesizing CNTs without defects in structure, one of the main lingering challenges.^{36,37,53} MW-assisted synthesis techniques exhibit faster heating rates as the material couple strongly with the electromagnetic energy under MW irradiation.^{37,54,55} The fast heating rate under MW irradiation stimulates and enhances the thermal decomposition reactions.⁵⁶ Dissimilar to the conventional heating, this can offer the selective heating due to the transformation of energy at a molecular scale. Additionally, MW heating generates hot spots that emerge from either the inhomogeneity of the MW field or the dielectric property and shape of feedstock particles. Consequently, the particle core is hotter than the particle surface temperature, as only the bulk temperature is measured; the size, shape, and chemical structure of materials can affect the extent of MW absorption, the heating intensity, and hot spots. The production of hot spots has considerable influences on the composition and distribution of final products.^{37,54,55}

After forming CNTs structures, they need to be purified and functionalized for further applications; MW-assisted protocols are favorable in view of rapid coupling of carbonaceous materials with MWs. The conventional techniques applied for the purification of CNTs include thermal, chemical, and sonication treatments.^{39,40} For instance, it has been revealed that physical properties such as thermal conductivity and diffusivity of the MWCNTs were highly enhanced by applying MW irradiation-based purification technique.⁵⁷ The purification of MWCNTs *via* MW oxidation techniques can reduce the related structural defects and improve the dispersity of these nanostructures; higher purification efficiency could be realized compared to the typical oxidation technique.⁵⁸ Furthermore, a two-step eco-friendly technique deployed MW irradiation for the purification of MWCNTs with the benefit of residual metal catalysts elimination,⁵⁹ the functionalization of CNTs using MW irradiation can decrease the reaction time,⁶⁰ and help improve their solubility.⁶¹

2.2. Vapor-assisted ozone treatment

A process for functionalizing of CNTs has been reported *via* a vapor-assisted ozone treatment; this technique is capable of

Table 1 Some selected examples of CNTs synthesized from biomass using MW irradiation

CNTs	Catalysts	Synthesis method	Biomass	Ref.
MWCNTs (50–200 nm)	Nickel	MW-chemical vapor deposition	Rice husk	50
MWCNTs (17–100 nm)	Ferrocene		Oat hulls, hazelnut hulls, wheat straw, and rapeseed cake	51
MWCNTs (50 nm)	Nickel (Ni)		Pine nutshell	48
MWCNTs (50–100 nm)	Mineral matter in char particles from biomass		Gumwood	49
MWCNTs (50–100 nm)	—	MW pyrolysis at low temperature (600 °C); self-extrusion of volatiles	Palm kernel shell cellulose	53
MWCNTs (~20 ± 10 nm, 20 wt% Fe and 50 ± 20 nm, 33.3 wt% Fe)	Fe and Co	MW pyrolysis at low temperature (400–500 °C)	Sugarcane bagasse	52



introducing extensive level of oxygen groups with a dominant population of carboxyl groups at near ambient temperature.⁶² The oxidative modified CNTs were obtained through a simple ozone treatment process under the solvent vapors comprising H_2O , H_2O_2 , and $\text{C}_2\text{H}_5\text{OH}$; the oxygenated groups on CNTs were considerably enhanced by applying the suitable functionalization temperature (~ 40 °C) and incremental partial vapor pressure. The ozone treatment under the vapor has been performed by applying compressed air as the carrier gas, providing environmentally benign functionalization procedure.⁶² Additionally, the suspension of CNTs was prepared deploying an ozone treatment to produce ultra-high performance concrete/CNT composites.⁶³ This ozone treatment could improve the CNTs dispersion in aqueous media *via* the formation of oxygen-bearing carboxylic groups on the surfaces of CNTs. Therefore, the interfacial interaction between CNTs and ultra-high-performance concrete could be improved, instigating significant nucleation influence at initial ages. Ozone treatment offered multiple nucleation sites and double steric repulsion *via* the accelerated degree of CNTs dispersion, resulting in the enhanced hydration at initial ages and increased compressive strength at later ages.⁶³

2.3. Ultrasonic irradiation

A process was reported for directly functionalization of SWCNTs using poly(ethylene glycol) grafted polymers in aqueous media *via* Diels–Alder click reaction at room temperature under ultrasonic irradiation.⁶⁴ The prepared CNT-based nanosystems with improved drug loading capacity could successfully deliver doxorubicin to the targeted tumor cells; these nanosystems with pH-dependent behavior exhibited suitable drug release rate at acidic condition (pH = 5.5) compared to the physiological condition (pH = 7.4). The functionalized SWCNTs delivery system had no noticeable cytotoxicity against the normal cell lines, but demonstrated significant antitumor effects against HeLa cancerous cells, providing promising cancer/tumor-targeted chemotherapy options.⁶⁴ Ultrasonic irradiation technique has been introduced for the chemical treatment of carboxylated MWCNTs–COOH with thiamine to generate poly(vinyl chloride)/thiamine-MWCNTs nanocomposites with significant thermal stability and improved mechanical properties.⁶⁵ Besides, polymers could be grafted on the surface of MWCNTs using Diels–Alder reaction in water *via* heating-stirring and ultrasound-assisted method. As a result, significant reaction rate enhancement could be attained, which was ~ 12 times higher than the one under the conventional heating-stirring condition without losing the efficiency of grafting process, thus offering a simple and environmentally benign direct functionalization technique applicable on larger scales.⁶⁶

MWCNTs have been synthesized *via* the sonication of CHCl_3 , CH_2Cl_2 , and CH_3I after the addition of silicon nanowires under ambient conditions. It was revealed that ultrasonic irradiation could facilitate the decomposition of solvent molecules and their subsequent reaction with the hydrogen-terminated silicon surfaces of the nanowires.⁶⁷ Similarly, a sonochemical method was deployed for the synthesis of high-purity SWCNTs.

The mixture solution was pulsed with high-intensity ultrasound for 20 min. Silica particles acted as nucleation sites and the ultrasound could facilitate the decomposition of ferrocene to produce Fe nanomaterials for catalyzing the growth of nanotubes.⁶⁸ Although acceptable results can be found in limited studies, but still the quality of CNTs produced *via* ultrasound-assisted techniques did not match the superiority of CNTs obtained by traditional tactics. Therefore, it is unlikely that ultrasound-assisted production of CNTs will find industrial applications unless significant improvements and optimization can be attained; however, ultrasound-assisted techniques have been widely deployed for the surface modification and dispersion/solubilization of CNTs.⁶⁹

3. Sustainable feedstocks and catalysts

Typically, carbon resources utilized in the synthesis of carbon nanomaterials are not renewable, and their production *via* conventional techniques often leads to the formation of hazardous or toxic waste/chemicals, generating possible adverse environmental impacts; innovative strategies and technologies are thus highly necessitated for the simpler, safer, low-cost, and sustainable fabrication of these nanostructures from renewable carbon resources with environmentally benign features, and lower sulfur/heavy metal levels, and greenhouse gas emissions.^{37,53} Carbon sources and synthesis techniques can affect the morphology and characteristics of CNTs, as well as their growth rate.^{39,70} Natural resources (e.g., plant-based hydrocarbons) and agricultural waste hydrocarbons have been explored for the eco-friendly synthesis of CNTs as low cost, renewable, and sustainable materials.^{71,72} Finding suitable and eco-friendly catalysts with low cost is crucial in producing high-quality CNTs.⁷³ On the other hand, natural mineral-derived catalysts with advantages of sustainability should be further considered for fabricating carbon-based nanomaterials such as CNTs, because of their low cost, eco-friendliness, and abundant accessibility.⁷⁴ For instance, MWCNTs were synthesized using a benign method comprising stone garnet sand (as low cost catalyst and support) and common household gas (as carbon source).⁷⁵ Vertically aligned CNTs (~ 4 –30 nm) could be gotten in high yield utilizing Fe/vermiculite catalyst on a pilot plant fluidized bed reactor chemical vapor deposition.⁷⁶ Natural clays have also been deployed as catalysts with the advantages of their inherently strong acidity, ion-exchange capability, adsorption, and large surface area.⁷⁷ Siliceous breccia was utilized as a natural catalyst for manufacturing CNTs without any pretreatment process through the chemical vapor deposition technique (750 °C, 30 min);⁷⁸ siliceous breccia with iron oxide-hydroxides functioned as catalysts for dissociating the precursors of hydrocarbon and producing the CNTs.⁷⁸ Pumice (a volcanic rock) and laterite (a clay material) as natural catalysts have been evaluated for fabricating graphitized CNTs; these CNTs were employed as reinforcement constituents in nanocomposite materials.⁷⁹ CNTs were also fabricated using biochar (a renewable and low-cost substrate) *via* MW-assisted chemical vapor deposition technique. Biochar produced *via* pyrolysis of biomass in the absence of oxygen demonstrated



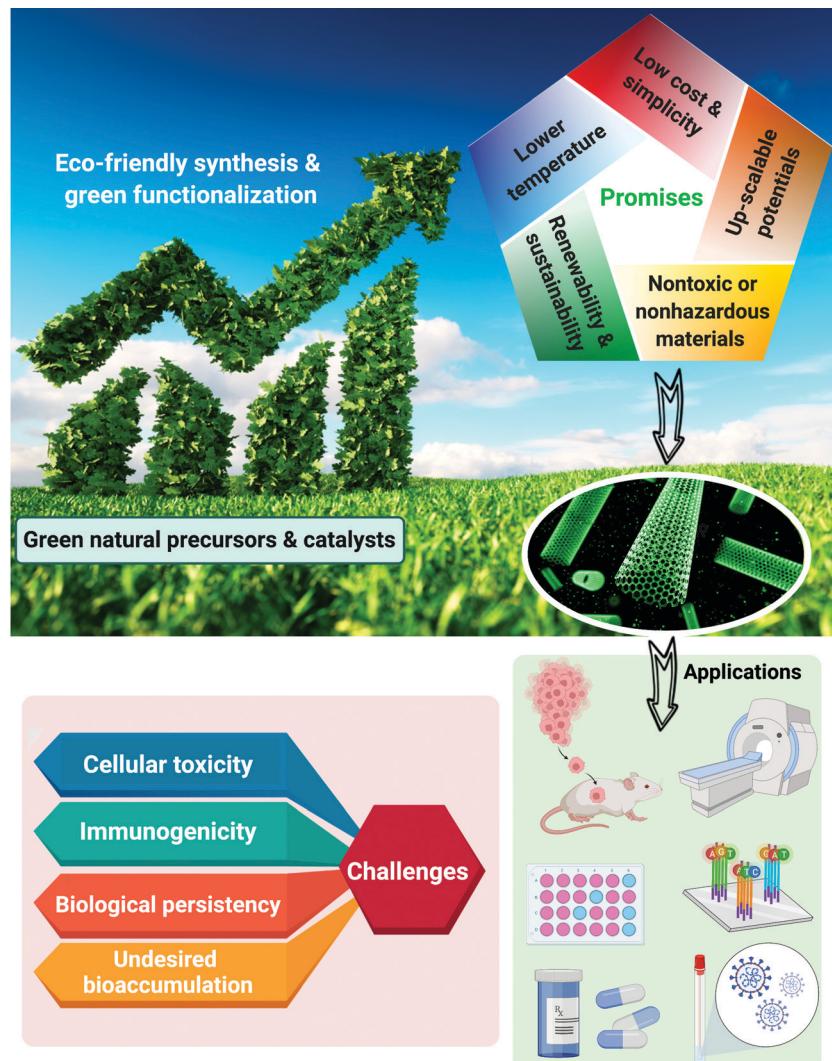


Fig. 2 Natural precursors or catalysts can be utilized for production and functionalization of CNTs, which can lead to significant advantages and promising applications, while some challenges still remain.

high porosity, allowing suitable dispersion of nanocatalysts on its surface thus stimulating the generation of CNTs.⁴⁸

Plant extracts as natural precursors can be deployed for sustainable synthesis of CNTs (Fig. 2).⁸⁰ They serve as greener catalysts for fabricating CNTs with the advantages of cost-effectiveness, renewability, and abundance,⁸¹ thereby avoiding the application of potentially hazardous metal catalysts (Table 2). Besides, plant extracts as green catalysts can be converted into porous and activated carbons with considerable nucleation sites for growing CNT-based nanostructures. The activated carbons with porous surfaces illustrated larger surface area than transition metals. Thus, nucleation sites for growing CNTs became larger, and the activated carbon could decompose hydrocarbons at a lower temperature; the growth of CNTs using plant extracts has been reported below ~ 575 °C, whereas the typical reactions applying transition metal catalysts were performed at $\sim 700\text{--}1200$ °C.⁴⁰ By applying a wall-nut extract catalyst, high yield of CNTs production could be obtained at chemical vapor

deposition temperature of 575 °C.⁸⁰ Qu *et al.*⁸² reported the greener formation of CNTs-Cu/ZnO nanocomposites utilizing *Brassica juncea* as a plant source of Cu, Zn, and C. The prepared CNTs (~ 80 nm) had middle-hollow structures and were not at all crystalline, with a few defects in the walls. They provided an innovative greener plant-based strategy for the development of CNTs; however, future investigations should be focused on their surface functionalization and optimization process.⁸² Besides, the nanostructures of MWCNTs ($\sim 80\text{--}90$ nm) were fabricated *via* chemical vapor deposition using *Cocos nucifera* Linn (coconut oil) where nitrogen gas was utilized as an inert atmosphere and a suitable carrier for the evaporated precursor.⁸³

Olive (*Olea europaea*) and coconut oils served as natural carbon precursors and NiCl_2 functioned as catalyst for the eco-friendly synthesis of CNTs ($\sim 27\text{--}31$ nm) through a simple and cost-effective pyrolysis technique at low temperatures. Consequently, uniformed SWCNTs from olive oil and the carbon rods from coconut oil were successfully obtained. The uniformity in



Table 2 Some plant-based renewable precursors/catalysts for the synthesis of CNTs

Plant-based resources	CNT-Based nanostructures	Techniques	Sizes (nm)	Ref.
Olive oil (<i>Olea europaea</i>) <i>Brassica juncea</i> L.	SWCNTs CNTs-Cu/ZnO nanocomposites	Pyrolysis method —	~27 Outer diameter of ~80	84 82
Coconut oil (<i>Cocos nucifera</i> Linn.)	MWCNTs	Chemical vapor deposition	Diameter of ~80–90	83
Eucalyptus oil	SWCNTs	Spray pyrolysis method	Diameter of ~0.79–1.71	86
Natural palm oil	Vertically aligned CNTs; SWCNTs	Thermal catalytic chemical vapor method	SWCNTs: ~0.6–1.2	32
Fresh bamboo culms	MWCNTs	Chemical vapor deposition	Diameter of less than 20	87
Rice straw (raw rice straw & neutral pulp)	MWCNTs	Pyrolysis method	Raw rice straw: ~15–40 Neutral pulp: 14.6–47.9	88
Turpentine oil (pine tree)	MWCNTs	Spray pyrolysis method	~15–40	89
Sesame oil (<i>Sesamum indicum</i>)	Branched nitrogen (N)-doped CNTs	Spray pyrolysis-assisted chemical vapor deposition	~30–60	90
<i>Cinnamomum camphora</i> (camphor)	MWCNTs	Chemical vapor deposition	~10	72
<i>Azadirachta indica</i> (neem oil)	MWCNTs	Spray pyrolysis method	~15–30	91
<i>Cynodon dactylon</i> , <i>Rosa</i> , <i>Azadirachta indica</i> , <i>Juglans regia</i>	MWCNTs	Chemical vapor deposition	~8–15	80

SWCNTs was not reported by using coconut oil, because of high proportion of saturated fat content (~82.5%) and less reactivity in comparison with the unsaturated hydrocarbons.⁸⁴ These CNTs should be further evaluated for agricultural, biotechnological and biomedical applications.⁸⁵

By applying a simple spray pyrolysis technique, SWCNTs (~0.79–1.71 nm) were fabricated through catalytic decomposition of the eucalyptus oil on a high silica-zeolite support impregnated with Fe/Co catalyst at 850 °C.⁸⁶ Suriani *et al.*³² synthesized vertically aligned CNTs with a diameter of ~0.6–1.2 nm on silicon substrates *via* thermal catalytic-chemical vapor reactor technique utilizing natural palm oil (as carbon source); these CNTs had high purity of ~90%.³² Additionally, the deployment of chemical vapor deposition technique was reported for synthesizing MWCNTs (<20 nm) using bamboo charcoals (as natural catalyst or substrate) derived from the heat-treated fresh bamboo culms at 1000–1500 °C. It was revealed that Mg₂SiO₄ and mostly calcium silicate were responsible for growing the MWCNTs.⁸⁷ The minerals in the bamboo charcoal served as catalysts similar to the transition metals for nucleating CNTs. Basta *et al.*⁸⁸ illustrated the eco-friendly fabrication of MWCNTs (~14.6–47.9 nm) from hydrochars of rice straw *via* a simple pyrolysis technique. The pyrolysis of rice straw-alkaline pulp and sulfite pulp hydrochars resulted in almost needles-like CNTs produced in between graphene nanosheets, with diameters ranging from 2.5–6.8 nm and 4–8 nm, respectively.⁸⁸ Besides, aligned CNTs (~70–130 μm in length) were fabricated using turpentine oil (as a low-cost plant-based precursor) and ferrocene mixture *via* a simple spray pyrolysis technique. These CNTs with outer diameters between ~15 and 40 nm should be further evaluated for biomedical applications.⁸⁹

Sesame oil as a natural and low-cost hydrocarbon precursor was utilized for manufacturing aligned-stack of branched nitrogen-doped CNTs *via* a simple spray pyrolysis-assisted chemical vapor deposition technique; ferrocene (C₁₀H₁₀Fe) and acetonitrile (CH₃CN) were utilized as the catalyst and nitrogen-doping agent, respectively for growing the N-doped aligned CNTs.⁹⁰ Neem oil

obtained from the seeds of *Azadirachta indica* was utilized as a carbon source to produce aligned CNTs nanostructures *via* a simple spray pyrolysis method. The main constituents of neem oil are hydrocarbons with less oxygen, offering the precursors in spray pyrolysis production of CNTs; the aligned CNT bundles were grown directly inside the quartz tubes. These CNTs were found to be multi-walled structures with an inner diameter of ~15–30 nm.⁹¹

4. Carbon-based nanomaterials with biomedical potentials

Overall, carbon-based nanomaterials with their attractive mechanical, optical, thermal, and electronic features can be simply functionalized;⁹² these materials have shown excellent potentials in bio- and nanomedicine because of their suitable electrical conductivity, biocompatibility, and large surface area in addition to their unique thermal and mechanical features.^{93,94} For instance, the electronic properties of CNTs mainly include high surface-to-volume ratio, the possibility of surface molecular absorption, fouling resistance, and stimulation ability to fast electron transfer kinetics.⁹⁵ These nanomaterials can be loaded with targeted drugs or molecularly therapeutic agents *via* hydrophobic interactions or π–π stacking due to their inherent hydrophobic nature, providing promising drug delivery nanosystems with high sensitivity/selectivity, targeting properties, sustained drug release behaviors, stability, and bio-clearance. Carbon-based nanomaterials can be further conjugated with a variety of imaging agents or targeted moieties with high densities, offering great opportunities for solving the limitations of detection and sensitivity towards the cancerous cells, especially for multimodal cancer theranostics.⁹⁶ These nanomaterials exhibited intrinsic properties that make them promising candidates for biomedical applications (Fig. 3A). As an example, CNTs exhibited unique optical absorption in the near-infrared region as well as easy functionalization, which can enhance their *in vivo* applications; they also showed bioimaging and tracing



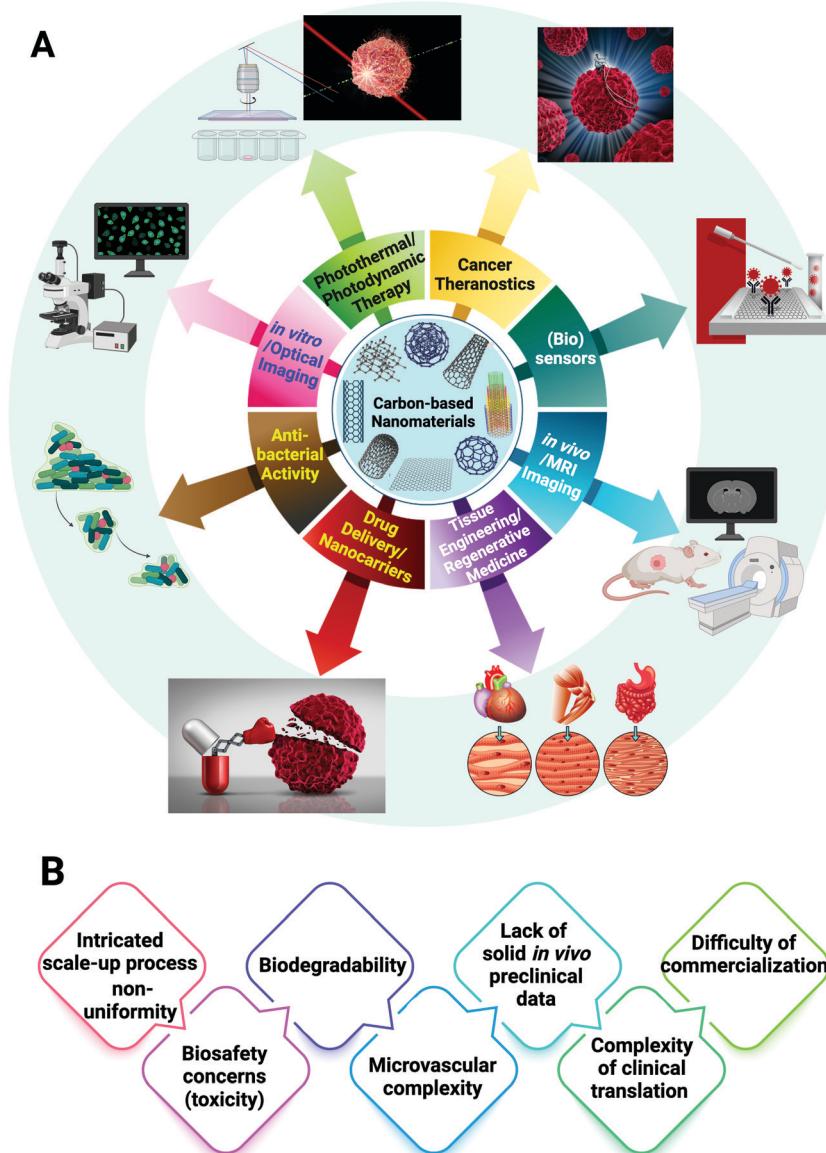


Fig. 3 (A) Biomedical promises and (B) important challenges of carbon-based nanomaterials.

performances combined with drug delivery (Fig. 3A).⁹⁷ However, since CNT-delivery systems with their needle (fiber)-like shapes could be bio-persistent and showed asbestos-like pathogenicity,⁹⁸ the biosafety issues and potential toxicity of these carbon-based nanomaterials should be the priority of research, especially for clinical translations (Fig. 3B).^{97,99} Currently, various studies have focused on the improvement of their biocompatibility through surface functionalization as well as targeted bioecological interactions to provide biocompatible nano-systems with enough susceptibility for enzymatic degradation and non/low immune system reactions.^{100–102} In this section, we focus on cancer theranostic applications of CNTs.

4.1. CNTs with cancer theranostic potentials

Nano-based structures and formulations with multifunctionality are being employed to produce innovative nano-delivery

systems as the conventional drugs or drug delivery systems have suffered from low specificity, selectivity, efficiency, and high adverse effects.^{103,104} For cancer therapy, engineered nanostructures have been designed to specifically target cancerous cells and reduce the possible toxic effects of drugs.^{105–107} In this context, various innovative nanocarriers and nanosystems with targeted and controlled anticancer properties have been designed using CNTs (Tables 3 and 4).^{13,104,108–111} But, more elaborative studies are still required for ascertaining the biocompatibility, cytotoxicity, carcinogenicity, and pharmacokinetics of these CNT-based nano-systems.¹¹² Wong *et al.*¹¹³ have reviewed the drug delivery potentials of CNTs, focusing on their drug loading and cellular uptake capacity as well as the other important features; the main challenges for the CNT-based delivery systems are hydrophobicity and potential toxicity (in biological systems). The surface functionalization of CNTs

Table 3 CNTs for cancer theranostic applications: important properties, approaches, and related anticancer pathways

CNTs-based nanosystems	Properties	<ul style="list-style-type: none"> - Strong optical absorption - The potential to convert the absorbed light into thermal heat - Phototherapy agents - High potential for generating reactive oxygen species (ROS) - High potential for delivering therapeutic/diagnostic agents - CNTs-Based drug delivery in cancer therapy - CNTs mediated photothermal therapy - CNTs mediated photodynamic therapy - CNTs-Based combined photothermal therapy (photodynamic-photothermal, chemo-photothermal, and immune-photothermal therapy) - CNTs for sonodynamic therapy - CNTs with anti-metastatic properties and functionality - Stimulation of immune system - Inhibition of cancer/tumor cells - Regulation of angiogenesis - Photodynamic/photothermal effects - Targeted/controlled anticancer therapeutics delivery
	Approaches	
	Anticancer pathways	

can enhance the biocompatibility and targeting features, which are crucial for cancer theranostics.^{114,115} For instance, a cancer therapy nanosystem was constructed by applying platinum nanoparticles supported on polybenzimidazole functionalized polymers and MWCNTs.¹¹⁶ Consequently, the prepared nanosystem demonstrated remarkable inhibitory activity on the epithelial–mesenchymal transition and cell cycle markers of cancerous cells; they had specific cytotoxicity on breast cancerous cells, but not on the adult stem cells.¹¹⁶ Further, hyaluronic acid as CD44 receptor targeting ligand and α -tocopheryl succinate for synergistic effects were employed for the functionalization of MWCNTs and the ensued structures were assessed for the targeted delivery of doxorubicin chemotherapeutic agent. Good biosafety and improved cellular placement as well as targeted anticancer effects could be obtained against breast cancerous cells.¹¹⁷ Significant cellular uptake, synergistic effects, growth inhibitory results, and total apoptotic ratio were reported by this nanoformulation.¹¹⁷

The salient advantageous features for CNT-based drug delivery systems encompass pH-dependent behavior, specificity/selectivity, prolonged/controlled release features, reduced side effects, high drug loading capacity, and reduced administration dose/frequency.¹³⁷ These materials with their unique architectures have been explored for targeted delivery of anticancer and chemotherapeutic agents (Table 5);¹³⁸ the improved release behavior and multifunctionality could be attained especially by MWCNTs for cancer therapy and diagnosis owing to sustained drug release in related cancer tissues and high surface areas for functioning groups. In addition, pH sensitive polymers (e.g., chitosan) demonstrated high targeting delivery properties with the advantages of biocompatibility, high tumor selectivity, pH-responsive behaviour, and chemical versatility.¹²⁵ Cirillo *et al.*¹²⁵ fabricated methotrexate-loaded chitosan-MWCNTs nano-hybrid platforms with pH-dependent, high sensitivity/selectivity, and sustained delivery features and evaluated their applicability against H1299 human lung cancerous cells compared to MRC-5 human lung fibroblast cells. In another study, an innovative nanoplatform comprised epirubicin-loaded magnetic-MWCNTs with controlled/sustained release and improved cytotoxicity effects.¹³⁹ Besides, MWCNTs were derivatized with naringenin

against lung cancerous cells.¹⁴⁰ These functionalized CNTs demonstrated pH- dependency and prolonged release behaviour as well as low toxicity effects on non-malignant cells compared to free naringenin; efficient anticancer effects on malignant lung cells could be achieved, *in vitro*.¹⁴⁰ In addition, the formulated chitosan–folate conjugated MWCNTs illustrated some important advantages of sustained release, biocompatibility, and biosafety features for targeted docetaxel delivery to the lung cancerous cells, *in vitro* (Fig. 4). This platform internalized into the cells *via* a folate receptor-mediated endocytic pathway with 89 fold more efficacy than the commercial docetaxel in A549 cells; the drug encapsulation efficacy was more than 79%. Histopathological analyses demonstrated no inflammatory or pathologic effects of this nanosystem.¹⁴¹

Functionalized CNTs have shown appropriate target localization, which is vital for efficient and targeted cancer diagnosis and therapy. For instance, bisphosphonate–CNT conjugates have been constructed for targeting regions of active bone metabolism and were prepared by applying covalent functionalization or latently generating reactive polymer–nanotube complexes (Fig. 5A and B),¹⁵³ which provided higher quality colloidal dispersions. Low cytotoxicity as well as proper biocompatibility profile could be achieved using both prepared bisphosphonate–CNT conjugates. These conjugates were radio-labelled with ^{99m}Tc in significant radiochemical yield ($\sim 80\text{--}92\%$). The biodistribution evaluations illustrated that these complexes had speedy blood clearance after 1 h; interestingly, superb bone localization of latently reactive polymer–nanotube complexes could be attained in comparison to covalent functionalization.¹⁵³ The functionalised MWCNTs were explored for gemcitabine drug delivery *via* a targeted lymph node system with magnetic properties, *in vitro* and *in vivo*. This nanosystem with high antitumor effects and drug loading capacity are suitable for the inhibition/regression of the metastasis in lymph node under the magnetic field with high efficacy and low side effects (Fig. 5C).¹⁴⁶

The treated C6 glioma cells with DNA wrapped SWCNTs ($5\text{ }\mu\text{g mL}^{-1}$) were exposed to bi-phasic electric pulses (6.6 V m^{-1} , 200 Hz, pulse duration 1 ms).¹⁵⁴ The low-frequency and low-strength electric field stimulation of glioma cells exposed to



Table 4 Some notable examples of nanoformulations or nanosystems constructed from CNTs for cancer theranostic applications

Type of CNTs	Anticancer drugs/agents	Advantages and properties	Ref.
SWCNTs	Doxorubicin	<ul style="list-style-type: none"> - Improved targeted delivery - High intracellular accumulation of drug - Suitable cytotoxicity effects with prolonged release behavior - High loading potentials 	118
	Paclitaxel	<ul style="list-style-type: none"> - High efficacy against cell proliferation with enhanced apoptosis rate of anticancer drug in hypoxic environment - Improved chemotherapeutic effects of paclitaxel through hypoxia-inducible factor (HIF)-1α downregulation and apoptosis-related/autophagy-associated proteins upregulation 	119
	Camptothecin	<ul style="list-style-type: none"> - Improved anticancer therapeutic efficiency with selective inhibitory effects of $\alpha_V\beta_3$-expressed cancerous cells, while inducing low cytotoxicity to $\alpha_V\beta_3$-negative cancerous cell up to 3.78- and 3.02-fold in two and three dimensional culture 	120
	Curcumin	<ul style="list-style-type: none"> - Improved blood concentration of curcumin (~18-fold) with remarkable inhibitory effects against cancerous cells and related tumor growth - No noticeable toxicity - Enhanced delivery features 	121
	HIF-1 α small interfering RNA (siRNA)	<ul style="list-style-type: none"> - Effective suppression of tumor growth - High specificity/selectivity and low toxicity - Selective inhibition of cellular HIF-1α performance 	122
	Cyclin A ₂ siRNA	<ul style="list-style-type: none"> - Improved reduction of cell proliferation with induction of apoptosis in chronic myelogenous leukaemia K562 cells - Depletion of cyclin A₂ can inhibit the proliferation of targeted cells and stimulate apoptosis of them - Induction of apoptosis - Over-expression and uptake of the P53 within the MCF-7 cells 	123
MWCNTs	p53 plasmids	<ul style="list-style-type: none"> - High selectivity with low toxicity - pH-Responsive and prolonged release behaviour - High efficacy and targeting properties - Prolonged and pH dependent release behaviour - Low haemolytic toxicity - High cytotoxicity against cancerous cells - Improved biocompatibility and pharmacokinetics - High cellular uptake with specificity - High cytotoxicity against cancerous cells - Combinatorial therapeutic effects against pancreatic cancer cells - High efficiency for drug delivery - High induced cancerous cell death effects - High selectivity and targeting properties - Improved photothermal effects with biocompatibility - High induced cancerous cell death - Targeted antitumor effects - High accumulation of CREKA peptide in tumors (~6 fold) - Efficient crossing from the cell membrane - Targeted antitumor effects with synergistic activity - Low toxicity - Significant inhibitory influences - High apoptosis rate and drug-loading capacity - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties - Efficient drug delivery - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Antitumor proliferation effects - Targeted cytotoxicity against cancerous cell lines 	124
	Methotrexate	<ul style="list-style-type: none"> - High selectivity with low toxicity - pH-Responsive and prolonged release behaviour - High efficacy and targeting properties - Prolonged and pH dependent release behaviour - Low haemolytic toxicity - High cytotoxicity against cancerous cells - Improved biocompatibility and pharmacokinetics - High cellular uptake with specificity - High cytotoxicity against cancerous cells - Combinatorial therapeutic effects against pancreatic cancer cells - High efficiency for drug delivery - High induced cancerous cell death effects - High selectivity and targeting properties - Improved photothermal effects with biocompatibility - High induced cancerous cell death - Targeted antitumor effects - High accumulation of CREKA peptide in tumors (~6 fold) - Efficient crossing from the cell membrane - Targeted antitumor effects with synergistic activity - Low toxicity - Significant inhibitory influences - High apoptosis rate and drug-loading capacity - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties - Efficient drug delivery - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Antitumor proliferation effects - Targeted cytotoxicity against cancerous cell lines 	125
	Gemcitabine	<ul style="list-style-type: none"> - High selectivity with low toxicity - pH-Responsive and prolonged release behaviour - High efficacy and targeting properties - Prolonged and pH dependent release behaviour - Low haemolytic toxicity - High cytotoxicity against cancerous cells - Improved biocompatibility and pharmacokinetics - High cellular uptake with specificity - High cytotoxicity against cancerous cells - Combinatorial therapeutic effects against pancreatic cancer cells - High efficiency for drug delivery - High induced cancerous cell death effects - High selectivity and targeting properties - Improved photothermal effects with biocompatibility - High induced cancerous cell death - Targeted antitumor effects - High accumulation of CREKA peptide in tumors (~6 fold) - Efficient crossing from the cell membrane - Targeted antitumor effects with synergistic activity - Low toxicity - Significant inhibitory influences - High apoptosis rate and drug-loading capacity - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties - Efficient drug delivery - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Antitumor proliferation effects - Targeted cytotoxicity against cancerous cell lines 	126
	Carboplatin	<ul style="list-style-type: none"> - High selectivity with low toxicity - pH-Responsive and prolonged release behaviour - High efficacy and targeting properties - Prolonged and pH dependent release behaviour - Low haemolytic toxicity - High cytotoxicity against cancerous cells - Improved biocompatibility and pharmacokinetics - High cellular uptake with specificity - High cytotoxicity against cancerous cells - Combinatorial therapeutic effects against pancreatic cancer cells - High efficiency for drug delivery - High induced cancerous cell death effects - High selectivity and targeting properties - Improved photothermal effects with biocompatibility - High induced cancerous cell death - Targeted antitumor effects - High accumulation of CREKA peptide in tumors (~6 fold) - Efficient crossing from the cell membrane - Targeted antitumor effects with synergistic activity - Low toxicity - Significant inhibitory influences - High apoptosis rate and drug-loading capacity - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties - Efficient drug delivery - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Antitumor proliferation effects - Targeted cytotoxicity against cancerous cell lines 	127
	Pemetrexed and quercetin	<ul style="list-style-type: none"> - High selectivity with low toxicity - pH-Responsive and prolonged release behaviour - High efficacy and targeting properties - Prolonged and pH dependent release behaviour - Low haemolytic toxicity - High cytotoxicity against cancerous cells - Improved biocompatibility and pharmacokinetics - High cellular uptake with specificity - High cytotoxicity against cancerous cells - Combinatorial therapeutic effects against pancreatic cancer cells - High efficiency for drug delivery - High induced cancerous cell death effects - High selectivity and targeting properties - Improved photothermal effects with biocompatibility - High induced cancerous cell death - Targeted antitumor effects - High accumulation of CREKA peptide in tumors (~6 fold) - Efficient crossing from the cell membrane - Targeted antitumor effects with synergistic activity - Low toxicity - Significant inhibitory influences - High apoptosis rate and drug-loading capacity - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties - Efficient drug delivery - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Antitumor proliferation effects - Targeted cytotoxicity against cancerous cell lines 	110
	Recombined ricin A chain	<ul style="list-style-type: none"> - High selectivity with low toxicity - pH-Responsive and prolonged release behaviour - High efficacy and targeting properties - Prolonged and pH dependent release behaviour - Low haemolytic toxicity - High cytotoxicity against cancerous cells - Improved biocompatibility and pharmacokinetics - High cellular uptake with specificity - High cytotoxicity against cancerous cells - Combinatorial therapeutic effects against pancreatic cancer cells - High efficiency for drug delivery - High induced cancerous cell death effects - High selectivity and targeting properties - Improved photothermal effects with biocompatibility - High induced cancerous cell death - Targeted antitumor effects - High accumulation of CREKA peptide in tumors (~6 fold) - Efficient crossing from the cell membrane - Targeted antitumor effects with synergistic activity - Low toxicity - Significant inhibitory influences - High apoptosis rate and drug-loading capacity - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties - Efficient drug delivery - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Antitumor proliferation effects - Targeted cytotoxicity against cancerous cell lines 	128
	Gold nanostars	<ul style="list-style-type: none"> - High selectivity with low toxicity - pH-Responsive and prolonged release behaviour - High efficacy and targeting properties - Prolonged and pH dependent release behaviour - Low haemolytic toxicity - High cytotoxicity against cancerous cells - Improved biocompatibility and pharmacokinetics - High cellular uptake with specificity - High cytotoxicity against cancerous cells - Combinatorial therapeutic effects against pancreatic cancer cells - High efficiency for drug delivery - High induced cancerous cell death effects - High selectivity and targeting properties - Improved photothermal effects with biocompatibility - High induced cancerous cell death - Targeted antitumor effects - High accumulation of CREKA peptide in tumors (~6 fold) - Efficient crossing from the cell membrane - Targeted antitumor effects with synergistic activity - Low toxicity - Significant inhibitory influences - High apoptosis rate and drug-loading capacity - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties - Efficient drug delivery - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Antitumor proliferation effects - Targeted cytotoxicity against cancerous cell lines 	129
	CREKA peptide (a tumor homing peptide)	<ul style="list-style-type: none"> - Targeted antitumor effects - High accumulation of CREKA peptide in tumors (~6 fold) 	130
	Gemcitabine and lentinan	<ul style="list-style-type: none"> - Efficient crossing from the cell membrane - Targeted antitumor effects with synergistic activity 	131
	Tamoxifen and lentinan	<ul style="list-style-type: none"> - Low toxicity - Significant inhibitory influences - High induced cancerous cell death effects - High selectivity and targeting properties - Improved photothermal effects with biocompatibility - High induced cancerous cell death - Targeted antitumor effects - High accumulation of CREKA peptide in tumors (~6 fold) - Efficient crossing from the cell membrane - Targeted antitumor effects with synergistic activity - Low toxicity - Significant inhibitory influences - High apoptosis rate and drug-loading capacity - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties - Efficient drug delivery - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Antitumor proliferation effects - Targeted cytotoxicity against cancerous cell lines 	132
Hybrid (SWCNTs and MWCNTs)	Trastuzumab and pertuzumab	<ul style="list-style-type: none"> - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties - Low toxicity - Significant inhibitory influences - High apoptosis rate and drug-loading capacity - High inhibitory activity against SK-BR-3 (a human breast cancer cell line) with targeting and selectivity properties 	133
	Doxorubicin	<ul style="list-style-type: none"> - High internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) 	134
	Serine/threonine-protein kinase (PLK1) siRNA	<ul style="list-style-type: none"> - High silencing of polo-like kinase 1 (PLK-1) in HeLa cells - Low toxicity - Good biocompatibility - Internalization of hybrid-CNTs in MCF-7 (a breast cancer cell line) and MDA-MB-231 (a breast cancer cell line) 	135
	Non-coding negative siRNA	<ul style="list-style-type: none"> - Antitumor proliferation effects - Targeted cytotoxicity against cancerous cell lines 	136

SWCNTs could lead to enhanced CNTs accumulation inside the cells.¹⁵⁴ In another study, the coupled SWCNTs with hyaluronic

acid and chlorin e6 were explored against colon cancer using photodynamic therapy.¹⁵⁵ The nanosystem improved the efficacy



Table 5 Some selected examples of CNT-based nanosystems for the delivery of anticancer agents

Nanosystems	Anticancer agents	Targeting agents	Cancer/tumor	Ref.
Chitosan-modified SWCNTs	Doxorubicin	Folic acid	Liver cancer	142
MWCNTs functionalized with poly(acrylic acid) and decorated with iron oxide magnetic nanoparticles	Doxorubicin	Folic acid	Human glioblastoma cells	143
Polyoxyl 35 castor oil noncovalent modified SWCNTs	Doxorubicin	—	Sarcoma tumor	144
Polycitric acid-polyethylene glycol-polycitric acid functionalized MWCNTs	Cisplatin	—	Colon adenocarcinoma tumor	145
Functionalized MWCNTs	Gemcitabine	Magnetic particles	Cancer lymph node metastasis	146
Functionalized SWCNTs with piperazine-polyethylenimine derivative	siRNA	—	Human breast cancer cells	147
Poly(ethylene glycol) grafted polymers-SWCNTs	Doxorubicin	—	HeLa cancer cells	64
Radiolabelled SWCNTs	Radionuclide	Thiolated antibodies	Burkitt lymphoma	148
Transactivator of transcription (TAT) peptide-chitosan-MWCNTs	Doxorubicin	—	Bel-7402 cells	149
Functionalized SWCNTs -CONH-(CH ₂) ₆ -NH ³⁺ Cl ⁻ functionalized SWCNTs	Taxoid	Biotin	Leukemia cells	150
MWCNTs-COOH	Telomerase reverse transcriptase siRNA-Pemetrexed and quercetin	—	Lewis lung tumors; human HeLa cells	151
MWCNTs coated with silica & chitosan Glycopolymer decorated MWCNTs	Doxorubicin	—	Human tumor cells (breast and pancreatic cells)	110
	Doxorubicin	Folic acid	Breast cancer	111
			Breast cancer	152

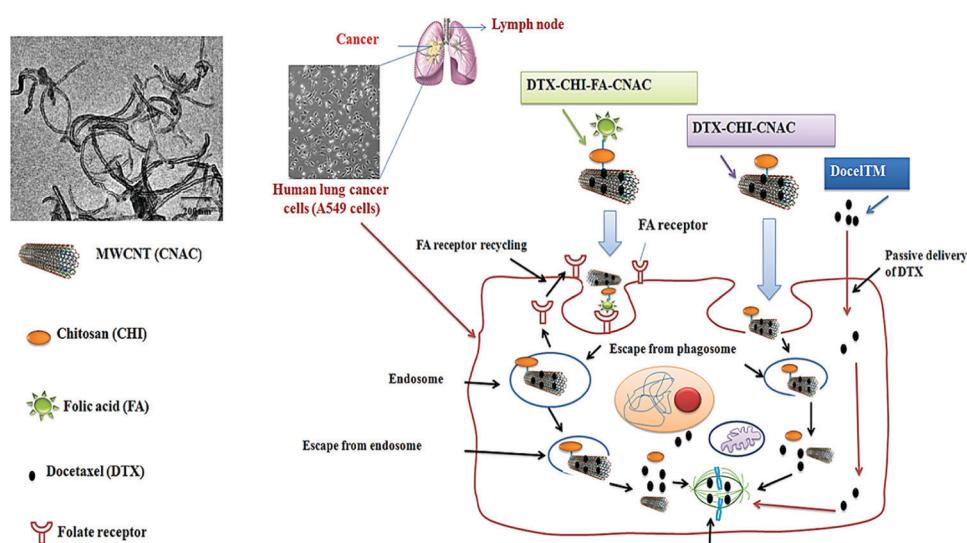


Fig. 4 Chitosan-folate conjugated MWCNTs for targeted and sustained delivery of docetaxel anticancer drug to the lung. Reproduced with permission from ref. 141 Copyright 2017 Elsevier.

of photodynamic therapy and triggered the death in colon cancerous cells.¹⁵⁵ For evaluating the tumor detectability and fluorescence image-guided surgery effects on post-operative survivals, an innovative custom-built reflectance/fluorescence imaging nanosystem was introduced.¹⁵⁶ The composed contrast agent from SWCNTs was an intraperitoneal injectable nano-molecular probe coupled to an M13 bacteriophage delivering a modified peptide attached to the extracellular protein over-expressed in ovarian cancer, SPARC protein. This imaging nanosystem could detect second near-infrared window fluorescence

with real-time video imagery potentials for better controlling the intra-operative tumor surgical removal; the microscopic detection of tumors with high quality and resolution has been reported.¹⁵⁶

5. Cytotoxicity and biocompatibility

Although CNTs can be desirable candidates in medicine and particularly for biomedical applications, there is not a unified consensus among the scientific community in terms of their



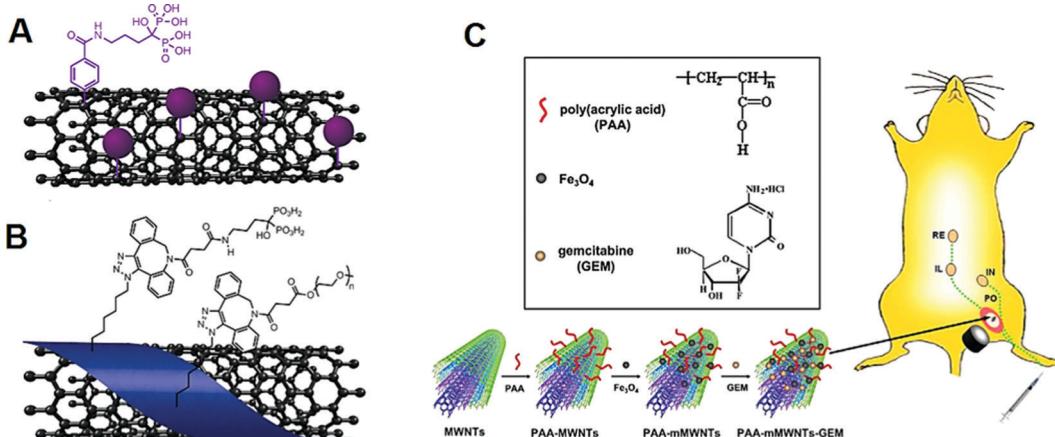


Fig. 5 Chemical structures of the prepared (A) covalently functionalized and (B) non-covalently functionalized SWCNTs. Reproduced with permission from ref. 153 copyright 2020 American Chemical Society. (C) MWCNTs for magnetic lymphatic gemcitabine drug delivery. PO: popliteal lymph node; IN: inguinal lymph node; IL: para-iliac lymph node; RE: renal hilar lymph nodes. Reproduced with permission from ref. 146 Copyright 2011 Elsevier.

toxic effects on cells.⁹ An undeniable fact is that there are no standard guidelines in scientific publications to calibrate and compare the result of one study with another. The most important and challenging issue is the ability to prepare CNTs of uniform length and diameter, because only after this accomplishment that the biological functional and cytotoxicity of CNTs can be improved. This hurdle gets even more critical while deploying SWCNTs, as chirality plays a key role in displaying the proper bio-function of the CNTs. Only if all these criteria are met, can the applications of CNTs in general be taken into green chemistry arena. Thus, it is imperative that all the CNTs should be the same length/diameter/chirality so that the toxicity profile of each one of the CNTs can be considered the same, thus enabling the full comprehension of the toxicological effects of all CNTs. However, it appears that this message is not well-understood and conveyed even in scientific studies nowadays.

If the physical properties (including aspect ratio known as the ratio of length to diameter, and area), dose, time of exposure, purity, and chemical agents for functionalization of CNTs are not well-engineered, then they can have fatal consequences because the CNTs can accumulate in different tissues/organs (such as brain, spleen, heart, kidney, lung, liver, *etc.*), produce reactive oxygen species (ROS) and eventually damage the healthy cells.^{12,157,158} This sequence of events happens through a process called phagocytosis (Fig. 6). Indeed, by exposing or entering inappropriate CNTs into the living body, the immune system recognizes them as foreign agents and attempts to phagocytose them by immune cells (macrophages) (Fig. 6). The effect of two different structural type of CNTs, short/tangled MWCNTs and long/rigid MWCNTs, have been studied on phagocytosis by macrophages and their following cleaning from the living tissues, to establish the safety profile of CNTs, *in vivo*.⁹⁸ The results revealed that low aspect ratio MWCNTs can be engulfed by macrophages before their clearance by draining lymph vessels, while high aspect ratio MWCNTs cannot be cleared and therefore gets accumulated in tissues,

which in turn promote carcinogenesis (Fig. 6a).¹⁵⁹ The efficiency of phagocytosis for varied geometries of CNTs was also mapped (Fig. 6b). As depicted in Fig. 6b, an incomplete phagocytosis occurs for large aspect ratio MWCNTs (upper left corner, long and thick CNTs, $Mph < 1$), while an effective phagocytosis was discerned for low aspect ratio of MWCNTs (lower left corner, short and small CNTs, $Mph > 1$).¹⁶⁰ Other factors that can influence the safety of CNTs *in vivo* includes the increased solubility of CNTs which in turn can prevent their aggregation and facilitate their clearance from the body and consequently less accumulation in the tissues/organs (Fig. 6c).¹⁵⁹ To overcome the aforementioned challenges, significant advances have been made in slicing both SWCNTs and MWCNTs to specific lengths with adherence to green chemistry tenets, for example, deploying continuous flow thin film microfluidics to ensure that the processing is scalable (Fig. 6d).¹⁶¹

The surface area, retention time, fibre dimensions, (bio)-persistency, and reactivity/inherent toxicity can affect the pathogenicity, biocompatibility, and toxicity of CNTs.¹⁶² In the context of medical and biomedical use, precise cellular analyses and specific histopathological assessments need to be performed to evaluate the biocompatibility and pathogenicity of these nano-structures;¹¹² though the chemically-functionalized CNTs displayed low toxicity, the possible scepticism due to non-biodegradability should be considered. It was reported that MWCNT-altered polyvinylchloride surfaces induced direct platelet activation/aggregation *in vitro*, and could also produce massive thrombosis in an *in vivo* rabbit model.¹⁶³ Pulmonary biocompatibility evaluations of administered CNTs in mice demonstrated severe decrease in antioxidants such as catalase, superoxide dismutase, and glutathione, and could induce the oxidants such as oxidative stress, lipid peroxidation, and myeloperoxidase (MPO).¹⁶⁴

The lack of stimulating acute adverse reactions (*e.g.*, cell detachment, tissue necrosis, and apoptosis), non-acute inflammatory reactions, non-toxicity of chemical components of metabolism, and non-toxic or without longstanding tissue bioaccumulation are some of the important features, which

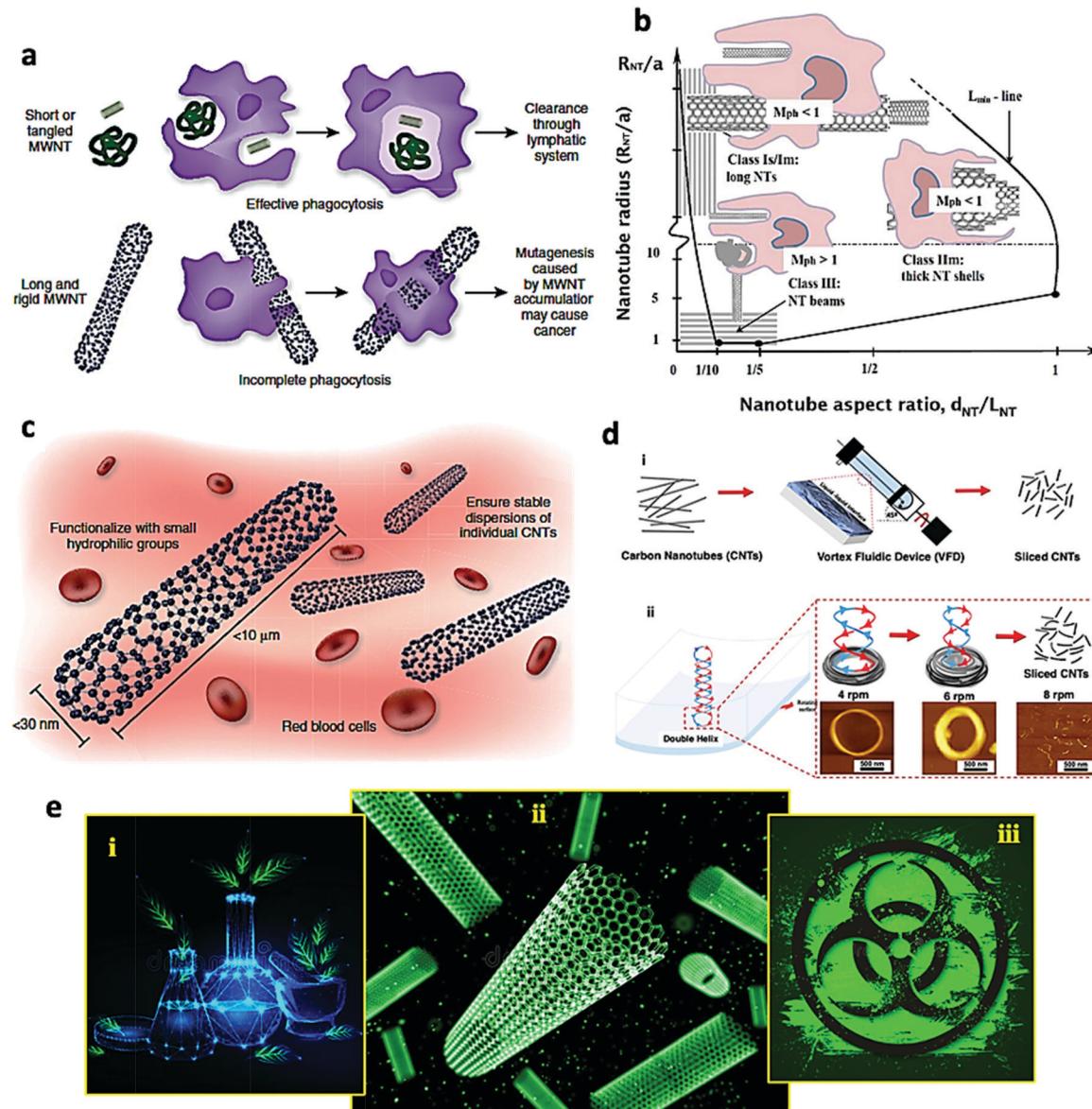


Fig. 6 Some important factors affecting the safety of CNTs *in vivo*: (a and b) the effect of CNTs structure on the efficiency of phagocytosis by macrophages and clearing from tissues, *in vivo*. Reproduced with permission from ref. 159 Copyright 1969, Nature Publishing Group and ref. 160 Copyright 2017 Elsevier. (c) Other factors such as the solubility and aggregation of CNTs influence the safety profile of CNTs *in vivo*. Reproduced with permission from ref. 159 Copyright 1969, Nature Publishing Group (d) (i) schematic illustration of the process for slicing both SWCNTs and MWCNTs via a vortex fluidic device (VFD) technique, and (ii) double helical topological fluid flow coiling at 4k and 6k rpm, with CNTs slicing at 8k rpm. Reproduced with permission from ref. 161 Copyright 2021 American Chemical Society (e) CNTs produced via greener processes could be a promising, cost-effective, eco-friendly, and leading to improve the biocompatibility and biodistribution of the CNTs, while exploiting the use of natural, renewable and sustainable resources instead of toxic chemicals.

should be considered for further applications of CNTs.¹⁶⁵ In one study, results obtained from cellular tests indicated the high level of cellular viability in contact with CNTs and the minor collagen generation enhancement, as well as the absence of pro-inflammatory interleukin 6 cytokine and the lack of free radicals induction from the prepared CNTs.¹⁶⁶ The evaluation of a single-chirality DNA-encapsulated SWCNTs complex upon intravenous administration exhibited suitable short- and long-term biocompatibility. After histopathological analyses, no alterations upon administration of CNTs were detected; also,

the route of administration, purity, types, and functionalization affected the biocompatibility and biodistribution of these nanostructures.¹⁶⁷

Biocompatible polymeric materials for the functionalization of CNTs can help to improve their biocompatibility, preventing their possible toxicity towards living organisms. For instance, biocompatible glycopolymers decorated onto MWCNTs could successfully improve their biocompatibility and anticancer delivery potentials.¹⁵² For preventing low dispersity and possible cytotoxicity, a new technique without utilizing any artificial

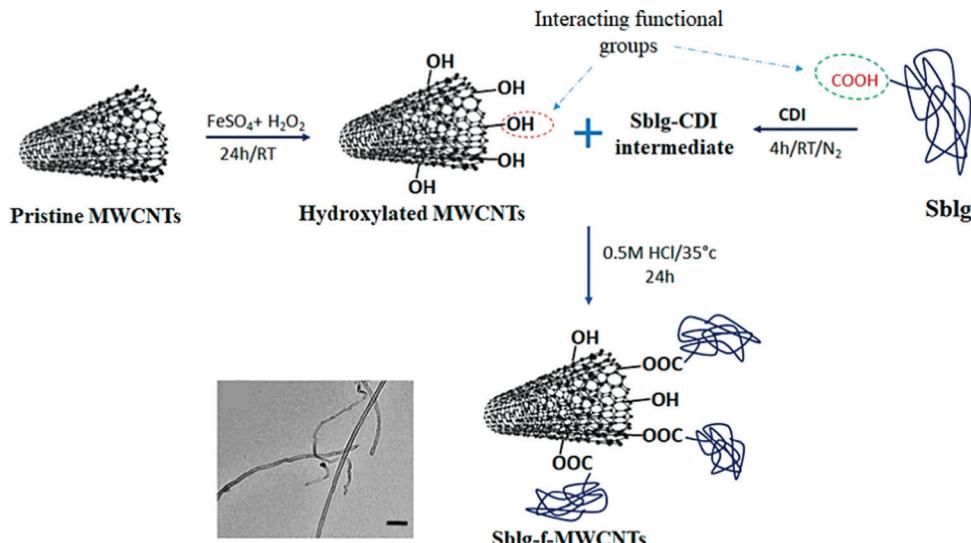


Fig. 7 The preparation process of succinylated β -lactoglobulin-functionalized MWCNTs. CDI: N,N' -carbonyldiimidazole, Sblg: succinylated β -lactoglobulin. Reproduced with permission from ref. 170 Copyright 2019 American Chemical Society.

chemical agents was reported for the production of biocompatible CNT ink with high stability up to months; these CNTs were stabilized by sustainable silk sericin protein, and the hybrid sericin–CNTs structures were formed *via* non-covalent interactions.¹⁶⁸ Some organic materials like tannic acid could effectively improve the dispersibility of CNTs and decrease their cytotoxicity, thus improving their biocompatibility.¹⁶⁹ The attachment of biocompatible polymers such as polyethylene-glycol (PEG) on CNTs was another strategy for improving the biocompatibility and dispersity, which are vital for their future clinical and biomedical applications; but this can cause the degradation, the formation of anti-PEG antibodies, and low cellular uptake.¹⁷⁰ Accordingly, Jain *et al.*¹⁷⁰ reported the surface functionalization of MWCNTs by applying the bovine-milk-derived protein succinylated β -lactoglobulin instead of PEG, providing superior biocompatibility, dispersion stability, and *in vitro* cell uptake. The succinylated β -lactoglobulin-functionalized MWCNTs demonstrated enhanced aqueous colloidal stability and half-maximal inhibitory concentration (IC_{50}) values (Fig. 7).¹⁷⁰

6. Concluding remarks and future perspectives

CNTs have been explored for various biological and biomedical applications owing to their attractive physicochemical features such as high thermo-electrical conductivity, large surface areas, multifunctionality, and significant mechanical strength. However, preparing them on a large scale is either expensive or sometimes ecologically unfriendly. In this context, plenty of studies have been conducted to discover innovative methods to fabricate CNTs in an eco-friendly and inexpensive manner. A benign and sustainable alternative to address the above-mentioned concerns would be to consider plant extracts and agriculture wastes as renewable, natural, and biodegradable

raw materials, which can reduce or eliminate the greenhouse gases, and exploitation of unsafe/toxic agents to help grow sustainable and greener technologies.

Various types of functionalized CNTs can be prepared *via* environmentally benign and sustainable techniques utilizing inexpensive, renewable, and natural resources as catalysts and abundant hydrocarbon precursors; these natural precursors and waste carbon have efficiently produced CNTs with desired stability and size/morphology. However, after the production of CNTs, suitable functionalization and tailoring of their surface chemistry are crucial for improving their toxicity, biocompatibility, and physical features; comprehensive *in vivo* and *in vitro* analyses are also vital for their future clinical and biomedical applications. These CNTs may suffer from some intrinsic problems, including poor solubility in aqueous solutions or organic solvents. The covalently functionalized CNTs exhibited approved pharmacokinetics and biodistribution properties, which are vital for the clinical and biomedical applications. The toxicity of CNTs depends on the type of nanotubes, administration methods, dose, and targeted tissue types, and it can be minimized by using greener surface functionalization or synthesis techniques; biodistribution, toxicological, and biosafety assessments are essential for future clinical and biomedical applications of these CNT-based nanosystems.

Conflicts of interest

The author(s) declare no competing interest.

Acknowledgements

E. M. would like to acknowledge the support from the National Institute of Biomedical Imaging and Bioengineering (5T32EB009035).



References

- Z. Chen, A. Zhang, X. Wang, J. Zhu, Y. Fan, H. Yu and Z. Yang, *J. Nanomater.*, 2017, 2017, DOI: [10.1155/2017/3418932](https://doi.org/10.1155/2017/3418932).
- E. Mostafavi, P. Soltantabar and T. J. Webster, *Biomaterials in translational medicine*, Elsevier, 2019, pp. 191–212.
- S. Iravani, *Green Chem.*, 2011, **13**, 2638–2650.
- S. Vivekanandhan, M. Schreiber, S. Muthuramkumar, M. Misra and A. K. Mohanty, *J. Appl. Polym. Sci.*, 2017, **134**, 1–15.
- S. Iravani and R. S. Varma, *Green Chem.*, 2020, **22**, 2643–2661.
- S. Iravani and R. S. Varma, *Green Chem.*, 2020, **22**, 612–636.
- S. Iravani and R. S. Varma, *Environ. Chem. Lett.*, 2020, **18**, 703–727.
- M. Zarghami Dehaghani, F. Yousefi, S. M. Sajadi, M. Tajammal Munir, O. Abida, S. Habibzadeh, A. H. Mashhadzadeh, N. Rabiee, E. Mostafavi and M. R. Saeb, *Molecules*, 2021, **26**, 4920.
- H. Zare, S. Ahmadi, A. Ghasemi, M. Ghanbari, N. Rabiee, M. Bagherzadeh, M. Karimi, T. J. Webster, M. R. Hamblin and E. Mostafavi, *Int. J. Nanomed.*, 2021, **16**, 1681–1706.
- T. Saliev, *J. Carbon Res.*, 2019, **5**, 1–22.
- R. Shoukat and M. Imran Khan, *Microsyst. Technol.*, 2022, **28**, 885–901.
- S. Rathinavel, K. Priyadarshini and D. Panda, *Mater. Sci. Eng., B*, 2021, **268**, 115095.
- B. F. Dizaji, S. Khoshbakht, A. Farboudi, M. H. Azarbajian and M. Irani, *Life Sci.*, 2020, **257**, 118059.
- P. Utreja, S. Jain and A. K. Tiwary, *Curr. Drug Delivery*, 2010, **7**, 152–161.
- R. Singh and S. V. Torti, *Adv. Drug Delivery Rev.*, 2013, **65**, 2045–2060.
- S. Marchesan, K. Kostarelos, A. Bianco and M. Prato, *Mater. Today*, 2015, **18**, 12–19.
- M. Sheikhpour, M. Naghinejad, A. Kasaeian, A. Lohrasbi, S. S. Shahraeini and S. Zomorodbakhsh, *Int. J. Nanomed.*, 2020, **15**, 7063–7078.
- D. Chen, C. A. Dougherty, K. Zhu and H. Hong, *J. Controlled Release*, 2015, **210**, 230–245.
- K. Kostarelos, L. Lacerda, G. Pastorin, W. Wu, S. Wieckowski, J. Luangsivilay, S. Godefroy, D. Pantarotto, J.-P. Briand, S. Muller, M. Prato and A. Bianco, *Nat. Nanotechnol.*, 2007, **2**, 108–113.
- A. Sanginario, B. Miccoli and D. Demarchi, *Biosensors*, 2017, **7**, 9.
- I. Koćik, D. Jankowski and A. Jagusiak, *J. Carbon Res.*, 2022, **8**, 1–16.
- M. A. Deshmukh, J. Y. Jeon and T. J. Ha, *Biosens. Bioelectron.*, 2020, **150**, 111919.
- L. Tang, Q. Xiao, Y. Mei, S. He, Z. Zhang, R. Wang and W. Wang, *J. Nanobiotechnol.*, 2021, **19**, 423.
- S. Hossen, M. K. Hossain, M. K. Basher, M. N.-H. Mia, M. T. Rahman and M. J. Uddin, *J. Adv. Res.*, 2019, **15**, 1–18.
- Z. Liao, S. W. Wong, H. L. Yeo and Y. Zhao, *NanoImpact*, 2020, **20**, 100253.
- S. Niyogi, M. A. Hamon, H. Hu, B. Zhao, P. Bhowmik, R. Sen, M. E. Itkis and R. C. Haddon, *Acc. Chem. Res.*, 2002, **35**, 1105–1113.
- G. Jamalipour Soufi, P. Iravani, A. Hekmatnia, E. Mostafavi, M. Khatami and S. Iravani, *Comments Inorg. Chem.*, 2021, DOI: [10.1080/02603594.2021.1990890](https://doi.org/10.1080/02603594.2021.1990890).
- G. Jamalipour Soufi and S. Iravani, *Green Chem.*, 2020, **22**, 2662–2687.
- V. Negri, J. Pacheco-Torres, D. Calle and P. López-Larrubia, *Top. Curr. Chem.*, 2020, **378**, 15.
- Z. Chen, A. Zhang, X. Wang, J. Zhu, Y. Fan, H. Yu and Z. Yang, *J. Nanomater.*, 2017, 2017, 1–13.
- J. Simon, E. Flahaut and M. Golzio, *Materials*, 2019, **12**, 624.
- A. B. Suriani, A. A. Azira, S. F. Nik, R. Md Nor and M. Rusop, *Mater. Lett.*, 2009, **63**, 2704–2706.
- L. Deng, Q. Xu and H. Wu, *Proc. Environ. Sci.*, 2016, **31**, 662–667.
- B. M. Chufa, H. C. Ananda Murthy, B. A. Gonfa and T. Y. Anshebo, *Green Chem. Lett. Rev.*, 2021, **14**, 640–657.
- M. Ukai, H. Kameya, H. Nakamura and Y. Shimoyama, *Spectrochim. Acta, Part A*, 2008, **69**, 1417–1422.
- J. Zhang, A. Tahmasebi, J. E. Omoriyekomwan and J. Yu, *J. Anal. Appl. Pyrolysis*, 2018, **130**, 142–148.
- J. E. Omoriyekomwan, A. Tahmasebi, J. Dou, R. Wang and J. Yu, *Fuel Process. Technol.*, 2021, **214**, 106686.
- Y. Liu, J. He, N. Zhang, W. Zhang, Y. Zhou and K. Huang, *J. Mater. Sci.*, 2021, **56**, 12559–12583.
- R. Kumar, R. K. Singh and D. P. Singh, *Renewable Sustainable Energy Rev.*, 2016, **58**, 976–1006.
- B. Makgabutlane, L. N. Nthunya, M. S. Maubane-Nkadieng and S. D. Mhlanga, *J. Environ. Chem. Eng.*, 2021, **9**, 104736.
- E. A. Burakova, T. P. Dyachkova, A. V. Rukhov, E. N. Tugolukov, E. V. Galunin, A. G. Tkachev, A. A. Basheer and I. Ali, *J. Mol. Liq.*, 2018, **253**, 340–346.
- M. Chen, C. Chen, S. Shi and C. Chen, *Jpn. J. Appl. Phys.*, 2003, **42**, 614–619.
- J. Y. Lee and B. S. Lee, *Thin Solid Films*, 2002, **418**, 85–88.
- A. Hirata and N. Yoshioka, *Tribol. Int.*, 2004, **37**, 893–898.
- N. M. Mubarak, J. N. Sahu, E. C. Abdullah, N. S. Jayakumar and P. Ganesan, *Res. Chem. Intermed.*, 2016, **42**, 3257–3281.
- J. A. Menéndez, A. Arenillas, B. Fidalgo, Y. Fernández, L. Zubizarreta, E. G. Calvo and J. M. Bermúdez, *Fuel Process. Technol.*, 2010, **91**, 1–8.
- S. Guo, Q. Dai, Z. Wang and H. Yao, *Composites, Part B*, 2017, **124**, 134–143.
- J. Zhang, A. Tahmasebi, J. E. Omoriyekomwan and J. Yu, *Diamond Relat. Mater.*, 2019, **91**, 98–106.
- K. Shi, J. Yan, E. Lester and T. Wu, *Ind. Eng. Chem. Res.*, 2014, **53**, 15012–15019.
- Z. Wang, H. Ogata, S. Morimoto, J. Ortiz-Medina, M. Fujishige, K. Takeuchi, H. Muramatsu, T. Hayashi, M. Terrones, Y. Hashimoto and M. Endo, *Carbon*, 2015, **94**, 479–484.
- P. Hildago-Oporto, R. Navia, R. Hunter, G. Coronado and M. E. Gonzalez, *J. Environ. Manag.*, 2019, **244**, 83–91.



52 B. Debalina, R. B. Reddy and R. Vinu, *J. Anal. Appl. Pyrolysis*, 2017, **124**, 310–318.

53 J. E. Omoriyekomwan, A. Tahmasebi, J. Zhang and J. Yu, *Energy Convers. Manag.*, 2019, **192**, 88–99.

54 J. Li, J. Dai, G. Liu, H. Zhang, Z. Gao, J. Fu, Y. He and Y. Huang, *Biomass Bioenergy*, 2016, **94**, 228–244.

55 A. S. Rossi, M. S. Pereira, J. M. dos Santos, I. Petri Jr and C. H. Ataide, *Mater. Sci. Forum*, 2017, **899**, 528–533.

56 P. Ingole, A. Ranveer, S. Deshmukh and K. Deshmukh, *Int. J. Adv. Technol. Eng. Sci.*, 2016, **4**, 78–84.

57 A. K.-M. M. Haque, G. S. Oh, T. Kim, J. Kim, J. Noh, S. Huh, H. Chung and H. Jeong, *Mater. Res. Bull.*, 2016, **73**, 247–255.

58 J. Zheng, R. Bao, J. Yi and P. Yang, *Diamond Relat. Mater.*, 2016, **68**, 93–101.

59 V. Gomez, S. Irusta, O. B. Lawal, W. Adams, R. H. Hauge, C. W. Dunnill and A. R. Barron, *RSC Adv.*, 2016, **6**, 11895–11902.

60 Y. Ling and A. Deokar, in *Carbon Nanotubes Applications on Electron Devices*, ed. J. M. Marulanda, IntechOpen, UK (London), 2010, pp. 128–141.

61 S. P. Economopoulos, N. Karousis, G. Rotas, G. Pagona and N. Tagmatarchis, *Curr. Org. Chem.*, 2011, **15**, 1121–1132.

62 J. Luo, Y. Liu, H. Wei, B. Wang, K.-H. Wu, B. Zhang and D. S. Su, *Green Chem.*, 2017, **19**, 1052–1062.

63 M. Jung, S.-g Hong and J. Moon, *Mater. Des.*, 2020, **193**, 108813.

64 X. T. Cao, M. Prakash Patil, Q. T. Phan, C. M.-Q. Le, B.-H. Ahn, G.-D. Kim and K. T. Lim, *J. Ind. Eng. Chem.*, 2020, **83**, 173–180.

65 S. Mallakpour, A. Abdolmaleki and F. Azimi, *Ultrason. Sonochem.*, 2017, **39**, 589–596.

66 C. M.-Q. Le, X. T. Cao and K. T. Lim, *Ultrason. Sonochem.*, 2017, **39**, 321–329.

67 C. P. Li, B. K. Teo, X. H. Sun, N. B. Wong and S. T. Lee, *Chem. Mater.*, 2005, **17**.

68 S.-H. Jeong, J.-H. Ko, J.-B. Park and W. Park, *J. Am. Chem. Soc.*, 2004, **126**, 15982–15983.

69 S. E. Skrabalak, *Phys. Chem. Chem. Phys.*, 2009, **11**, 4930–4942.

70 A. D. Goswami, D. H. Trivedi, N. L. Jadhav and D. V. Pinjari, *J. Environ. Chem. Eng.*, 2021, **9**, 106118.

71 V. Romanovicz, B. A. Berns, S. D. Carpenter and D. Carpenter, *Int. J. Energy Power Eng.*, 2013, **7**, 665–668.

72 M. Kumar and Y. Ando, *J. Phys.: Conf. Ser.*, 2007, **61**, 129.

73 T. U. Wani, R. Mohi-Ud-Din, T. A. Wani, R. H. Mir, A. M. Itoo, F. A. Sheikh, N. A. Khan and F. H. Pottoo, *Curr. Pharm. Biotechnol.*, 2021, **22**, 793–807.

74 S. K. Verma, A. K. Das, S. Gantait, V. Kumar and E. Gurel, *Sci. Total Environ.*, 2019, **667**, 485–499.

75 M. Endo, K. Takeuchi, Y. A. Kim, K. C. Park, T. Ichiki, T. Hayashi, T. Fukuyo, S. Iinou, D. S. Su, M. Terrones and M. S. Dresselhaus, *ChemSusChem*, 2008, **1**, 820–822.

76 Q. Zhang, M.-Q. Zhao, J.-Q. Huang, Y. Liu, Y. Wang, W.-Z. Qian and F. Wei, *Carbon*, 2009, **47**, 2600–2610.

77 W.-D. Zhang, I. Y. Phang and T. X. Liu, *Adv. Mater.*, 2006, **18**, 73–77.

78 A. Kumar, Y. Kostikov, M. Zanatta, G. D. Sorarù, B. Orberger, G. D. Nessim and G. Mariotto, *Diamond Relat. Mater.*, 2019, **97**, 107433.

79 S. Malik, *Polyhedron*, 2018, **152**, 90–93.

80 N. Tripathi, V. Pavelyev and S. S. Islam, *Appl. Nanosci.*, 2017, **7**, 557–566.

81 D. Janas, *Sustainability*, 2020, **12**, 4115.

82 J. Qu, C. Luo, Q. Cong and X. Yuan, *J. Mater. Cycles Waste Manag.*, 2014, **16**, 162–166.

83 S. Paul and S. K. Samdarshi, *New Carbon Mater.*, 2011, **26**, 85–88.

84 Z. Abdel Hamid, A. Abdul Azim, F. Abdel Mouez and S. S. Abdel Rehim, *J. Anal. Appl. Pyrolysis*, 2017, **126**, 218–229.

85 D. K. Patel, H.-B. Kim, S. D. Dutta, K. Ganguly and K.-T. Lim, *Materials*, 2020, **13**, 1679.

86 P. Ghosh, R. A. Afre, T. Soga and T. Jimbo, *Mater. Lett.*, 2007, **61**, 3768–3770.

87 J. Zhu, J. Jia, F. L. Kwong, D. H. Leung Ng and S. C. Tjong, *Biomass Bioenergy*, 2012, **36**, 12–19.

88 V. F. Lotfy, N. A. Fathy and A. H. Basta, *J. Environ. Chem. Eng.*, 2018, **6**, 6263–6274.

89 K. Awasthi, R. Kumar, R. S. Tiwari and O. N. Srivastava, *J. Exp. Nanosci.*, 2010, **5**, 498–508.

90 R. Kumar, R. K. Singh and R. S. Tiwari, *Mater. Des.*, 2016, **94**, 166–175.

91 R. Kumar, R. S. Tiwari and O. N. Srivastava, *Nanoscale Res. Lett.*, 2011, **6**, 92.

92 S. Augustine, J. Singh, M. Srivastava, M. Sharma, A. Das and B. D. Malhotra, *Biomater. Sci.*, 2017, **5**, 901–952.

93 M. Derakhshi, S. Daemi, P. Shahini, A. Habibzadeh, E. Mostafavi and A. A. Ashkarraan, *J. Funct. Biomater.*, 2022, **13**, 27.

94 M. Ashrafizadeh, H. Saebfar, M. H. Gholami, K. Hushmandi, A. Zabolian, P. Bikarannejad, M. Hashemi, S. Daneshi, S. Mirzaei, E. Sharifi, A. P. Kumar, H. Khan, H. H. Sheikh Hossein, M. Vosough, N. Rabiee, V. K. Thakur, P. Makvandi, Y. K. Mishra, F. R. Tay, Y. Wang, A. Zarrabi, G. Orive and E. Mostafavi, *Expert Opin. Drug Delivery*, 2022, **19**, 355–382.

95 A. Vasilescu, A. Hayat, S. Gáspár and J.-L. Marty, *Electroanalysis*, 2018, **30**, 2–19.

96 J. Saleem, L. Wang and C. Chen, *Adv. Healthcare Mater.*, 2018, **7**, 1800525.

97 K. Park, *ACS Nano*, 2013, **7**, 7442–7447.

98 C. A. Poland, R. Duffin, I. Kinloch, A. Maynard, W. A. Wallace, A. Seaton, V. Stone, S. Brown, W. MacNee and K. Donaldson, *Nat. Nanotechnol.*, 2008, **3**, 423–428.

99 Y. Grosse, D. Loomis, K. Z. Guyton, B. Lauby-Secretan, F. E. Ghissassi, V. Bouvard, L. Benbrahim-Tallaa, N. Guha, C. Scoccianti, H. Mattock and K. Straif, *Lancet Oncol.*, 2014, **15**, 1427–1428.

100 D. Chen, C. A. Dougherty, K. Zhu and H. Hong, *J. Controlled Release*, 2015, **210**, 230–245.

101 F. M. Tonelli, V. A. Goulart, K. N. Gomes, M. S. Ladeira, A. K. Santos, E. Lorençon, L. O. Ladeira and R. R. Resende, *Nanomedicine*, 2015, **10**, 2423–2450.

102 K. Bhattacharya, S. P. Mukherjee, A. Gallud, S. C. Burkert, S. Bistarelli, S. Bellucci, M. Bottini, A. Star and B. Fadeel, *Nanomedicine*, 2016, **12**, 333–351.



103 A. V.-V. V. Ravi Kiran, G. Kusuma Kumari and P. T. Krishnamurthy, *J. Drug Delivery Sci. Technol.*, 2020, **59**, 101892.

104 S. Raj, S. Khurana, R. Choudhari, K. K. Kesari, M. A. Kamal, N. Garg, J. Ruokolainen, B. C. Das and D. Kumar, *Semin. Cancer Biol.*, 2021, **69**, 166–177.

105 C. C. Carrion, M. Nasrollahzadeh, M. Sajjadi, B. Jaleh, G. Jamalipour Soufi and S. Iravani, *Int. J. Biol. Macromol.*, 2021, **178**, 193–228.

106 M. Nasrollahzadeh, M. Sajjadi, S. Iravani and R. S. Varma, *Nanomaterials*, 2020, **10**, 1784, DOI: [10.3390/nano10091784](https://doi.org/10.3390/nano10091784).

107 M. Nasrollahzadeh, M. Sajjadi, S. Iravani and R. S. Varma, *Chemosphere*, 2021, **263**, 128005.

108 S. Iravani and R. S. Varma, *Mater. Adv.*, 2021, **2**, 2906–2917.

109 S. Iravani and R. S. Varma, *ACS Biomater. Sci. Eng.*, 2021, **7**, 1900–1913.

110 N. Badea, M. M. Craciun, A. S. Dragomir, M. Balas, A. Dinischiotu, C. Nistor, C. Gavan and D. Ionita, *Mater. Chem. Phys.*, 2020, **241**, 122435.

111 A. A.-G. El-Shahawy, N. Elnagar, M. Zohery, M. S. Abd Elhafeez and S. I. El-Dek, *Int. J. Polym. Mater. Polym. Biomater.*, 2021, DOI: [10.1080/00914037.2021.1925277](https://doi.org/10.1080/00914037.2021.1925277).

112 K. Aoki and N. Saito, *Nanomaterials*, 2020, **10**, 264.

113 M. A. Saleemi, Y. L. Kong, P. V.-C. Yong and E. H. Wong, *J. Drug Delivery Sci. Technol.*, 2020, **59**, 101855.

114 M. Eskandari, S. H. Hosseini, M. Adeli and A. Pourjavadi, *Iran. Polym. J.*, 2014, **23**, 387–403.

115 Y. Zhang, Y. Bai and B. Yan, *Drug Discovery Today*, 2010, **15**, 428–435.

116 M. R. Berber, H. Elkhenany, I. H. Hafez, A. El-Badawy, M. Essawy and N. El-Badri, *Nanomedicine*, 2020, **15**, 793–808, DOI: [10.2217/nnm-2019-0445](https://doi.org/10.2217/nnm-2019-0445).

117 N. J. Singhai, R. Maheshwari and S. Ramteke, *Colloids Interface Sci. Commun.*, 2020, **35**, 100235.

118 Y.-J. Gu, J. Cheng, J. Jin, S. H. Cheng and W.-T. Wong, *Int. J. Nanomed.*, 2011, **6**, 2889–2898.

119 Y. Wang, C. Wang, Y. Jia, X. Cheng, Q. Lin, M. Zhu, Y. Lu, L. Ding, Z. Weng and K. Wu, *PLoS One*, 2014, **9**, e104209.

120 B. Koh, S. B. Park, E. Yoon, H. M. Yoo, D. Lee, J. N. Heo and S. Ahn, *J. Pharm. Sci.*, 2019, **108**, 3704–3712.

121 H. Li, N. Zhang, Y. Hao, Y. Wang, S. Jia and H. Zhang, *Drug Delivery*, 2019, **26**, 1017–1026.

122 G. Bartholomeusz, P. Cherukuri, J. Kingston, L. Cognet, R. Lemos, T. K. Leeuw, L. Gumbiner-Russo, R. B. Weisman and G. Powis, *Nano Res.*, 2009, **2**, 279–291.

123 X. Wang, J. Ren and X. Qu, *ChemMedChem*, 2008, **3**, 940–945.

124 A. Karmakar, S. M. Bratton, E. Dervishi, A. Ghosh, M. Mahmood, Y. Xu, L. Mohammed Saeed, T. Mustafa, D. Casciano, A. Radominska-Pandya and A. S. Biris, *Int. J. Nanomed.*, 2011, **6**, 1045–1055.

125 G. Cirillo, O. Vittorio, D. Kunhardt, E. Valli, A. Farfalla, M. Curcio, U. G. Spizzirri and S. Hampel, *Materials*, 2019, **12**, 2889.

126 S. K. Prajapati, A. Jain, C. Shrivastava and A. K. Jain, *Int. J. Biol. Macromol.*, 2019, **123**, 691–703.

127 D. Salas-Treviño, O. Saucedo-Cárdenas, M. D. Loera-Arias, H. Rodríguez-Rocha, A. García-García, R. Montes-de-Oca-Luna, E. I. Piña-Mendoza, F. F. Contreras-Torres, G. García-Rivas and A. Soto-Domínguez, *Nanomaterials*, 2019, **9**, 1572.

128 X. Weng, M. Wang, J. Ge, S. Yu, B. Liu, J. Zhong and J. Kong, *Mol. BioSyst.*, 2009, **5**, 1224–1231.

129 Y. Zhu, Q. Sun, Y. Liu, T. Ma, L. Su, S. Liu, X. Shi, D. Han and F. Liang, *R. Soc. Open Sci.*, 2018, **5**, 180159.

130 B. Zhang, H. Wang, S. Shen, X. She, W. Shi, J. Chen, Q. Zhang, Y. Hu, Z. Pang and X. Jiang, *Biomaterials*, 2016, **79**, 46–55.

131 P. Zhang, W. Yi, J. Hou, S. Yoo, W. Jin and Q. Yang, *Int. J. Nanomed.*, 2018, **13**, 3069.

132 W. Yi, P. Zhang, J. Hou, W. Chen, L. Bai, S. Yoo, A. Khalid and X. Hou, *Int. J. Biol. Macromol.*, 2018, **120**, 1525–1532.

133 X. Zhang, J. Chen, Z. Weng, Q. Li, L. Zhao, N. Yu, L. Deng, W. Xu, Y. Yang, Z. Zhu and H. Huang, *Mol. Immunol.*, 2020, **119**, 48–58.

134 P. S.-O. Ozgen, S. Atasoy, B. Z. Kurt, Z. Durmus, G. Yigitte and A. Dag, *J. Mater. Chem. B*, 2020, **8**, 3123–3137.

135 A. Battigelli, J. T.-W. Wang, J. Russier, T. Da Ros, K. Kostarelos, K. T. Al-Jamal, M. Prato and A. J.-S. Bianco, *Small*, 2013, **9**, 3610–3619.

136 P. Singh, C. Samori, F. M. Toma, C. Bussy, A. Nunes, K. T. Al-Jamal, C. Ménard-Moyon, M. Prato, K. Kostarelos and A. Bianco, *J. Mater. Chem.*, 2011, **21**, 4850–4860.

137 M. S. Hasnain and A. K. Nayak, *Carbon Nanotubes for Targeted Drug Delivery*, Springer, 2019, pp. 1–9, DOI: [10.1007/978-981-15-0910-0_1](https://doi.org/10.1007/978-981-15-0910-0_1).

138 R. Maleki, H. H. Afrouzi, M. Hosseini, D. Toghraie, A. Piranfar and S. Rostami, *Comput. Methods Prog. Biomed.*, 2020, **186**, 105210.

139 N. Suo, M. Wang, Y. Jin, J. Ding, X. Gao, X. Sun, H. Zhang, M. Cui, J. Zheng, N. Li, X. Jin and S. Jiang, *Int. J. Nanomed.*, 2019, **14**, 1241–1254.

140 R. P. Morais, G. B. Novais, L. S. Sangenito, A. L.-S. Santos, R. Priefer, M. Morsink, M. C. Mendonça, E. B. Souto, P. Severino and J. C. Cardoso, *Int. J. Mol. Sci.*, 2020, **21**, 4557.

141 R. P. Singh, G. Sharma Sonali, S. Singh, S. Bharti, B. L. Pandey, B. Koch and M. S. Muthu, *Mater. Sci. Eng., C*, 2017, **77**, 446–458.

142 Z. Ji, G. Lin, Q. Lu, L. Meng, X. Shen, L. Dong, C. Fu and X. Zhang, *J. Colloid Interface Sci.*, 2012, **365**, 143–149.

143 Y. J. Lu, K. C. Wei, C. C.-M. Ma, S. Y. Yang and J. P. Chen, *Colloids Surf., B*, 2012, **89**, 1–9.

144 H. Liu, H. Xu, Y. Wang, Z. He and S. Li, *Drug Dev. Ind. Pharm.*, 2012, **38**, 1031–1038.

145 M. Adeli, F. Hakimpoor, M. Ashiri, R. Kabiri and M. Bavadi, *Soft Matter*, 2011, **7**, 4062–4070.

146 F. Yang, C. Jin, D. Yang, Y. Jiang, J. Li, Y. Di, J. Hu, C. Wang, Q. Ni and D. Fu, *Eur. J. Cancer*, 2011, **47**, 1873–1882.

147 M. Mohammadi, Z. Salmasi, M. Hashemi, F. Mosaffa, K. Abnous and M. Ramezani, *Int. J. Pharm.*, 2015, **485**, 50–60.



148 M. R. McDevitt, D. Chattopadhyay, B. J. Kappel, J. S. Jaggi, S. R. Schiffman, C. Antczak, J. T. Njardarson, R. Brentjens and D. A. Scheinberg, *J. Nucl. Med.*, 2007, **48**, 1180–1189.

149 X. Dong, Z. Sun, X. Wang and X. Leng, *Nanomedicine*, 2017, **13**, 2271–2280.

150 J. Chen, S. Chen, X. Zhao, L. V. Kuznetsova, S. S. Wong and I. Ojima, *J. Am. Chem. Soc.*, 2008, **130**, 16778–16785.

151 Z. Zhang, X. Yang, Y. Zhang, B. Zeng, S. Wang, T. Zhu, R. B.-S. Roden, Y. Chen and R. Yang, *Clin. Cancer Res.*, 2006, **12**, 4933–4939.

152 P. S.-O. Ozgen, S. Atasoy, B. Z. Kurt, Z. Durmus, G. Yigit and A. Dag, *J. Mater. Chem. B*, 2020, **8**, 3123–3137.

153 A. R. Genady, D. Fong, S. R. Slikboer, M. E. El-Zaria, R. Swann, N. Janzen, A. Faraday, S. A. McNelles, M. Rezvani, S. Sadeghi, A. Adronov and J. F. Valliant, *ACS Appl. Nano Mater.*, 2020, **3**, 11819–11824.

154 L. Golubewa, T. Kulahava, Y. Kunitskaya, P. Bulai, M. Shuba and R. Karpicz, *Biochem. Biophys. Res. Commun.*, 2020, **529**, 647–651.

155 P. Sundaram and H. Abrahamse, *Int. J. Mol. Sci.*, 2020, **21**, 4745.

156 L. Ceppi, N. M. Bardhan, Y. Na, A. Siegel, N. Rajan, R. Fruscio, M. G. D. Carmen, A. M. Belcher and M. J. Birrer, *ACS Nano*, 2019, **13**, 5356–5365.

157 A. M. Díez-Pascual, *Macromolecules*, 2021, **1**, 64–83.

158 R. Dubey, D. Dutta, A. Sarkar and P. Chattopadhyay, *Nanoscale Adv.*, 2021, **3**, 5722–5744.

159 K. Kostarelos, *Nat. Biotechnol.*, 2008, **26**, 774–776.

160 V. M. Harik, *Toxicol. Lett.*, 2017, **273**, 69–85.

161 T. M. Alharbi, Q. Li and C. L. Raston, *ACS Sustainable Chem. Eng.*, 2021, **9**, 16044–16051.

162 S. K. Smart, A. I. Cassady, G. Q. Lu and D. J. Martin, *Carbon*, 2006, **44**, 1034–1047.

163 A. M. Gaffney, M. J. Santos-Martinez, A. Satti, T. C. Major, K. J. Wynne, Y. K. Gun'ko, G. M. Annich, G. Elia and M. W. Radomski, *Nanomedicine*, 2015, **11**, 39–46.

164 P. Ravichandran, S. Baluchamy, R. Gopikrishnan, S. Biradar, V. Ramesh, V. Goornavar, R. Thomas, B. L. Wilson, R. Jeffers, J. C. Hall and G. T. Ramesh, *J. Biol. Chem.*, 2011, **286**, 29725–29733.

165 A. Bianco, K. Kostarelos and M. Prato, *Chem. Commun.*, 2011, **47**, 10182–10188.

166 J. Chłopek, B. Czajkowska, B. Szaraniec, E. Frackowiak, K. Szostak and F. Béguin, *Carbon*, 2006, **44**, 1106–1111.

167 T. V. Galassi, M. Antman-Passig, Z. Yaari, J. Jessurun, R. E. Schwartz and D. A. Heller, *PLoS One*, 2020, **15**, e0226791.

168 X. Liang, H. Li, J. Dou, Q. Wang, W. He, C. Wang, D. Li, J.-M. Lin and Y. Zhang, *Adv. Mater.*, 2020, **32**, 2000165.

169 X. Zhang, M. Liu, X. Zhang, F. Deng, C. Zhou, J. Hui, W. Liu and Y. Wei, *Toxicol. Res.*, 2015, **4**, 160–168.

170 S. Jain, S. M. Dongave, T. Date, V. Kushwah, R. R. Mahajan, N. Pujara, T. Kumeria and A. Popat, *ACS Biomater. Sci. Eng.*, 2019, **5**, 3361–3372.

