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# MoS<sub>2</sub> nanosheets for the detoxification of Hg<sup>2+</sup> in living cells†

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A simple and green one-step method for the simultaneous exfoliation and functionalization of few-layered MoS<sub>2</sub> nanosheets was developed via the ultrasonic treatment of bulk MoS<sub>2</sub> powder in sodium phytate-containing aqueous solution. The as-prepared MoS<sub>2</sub> nanosheets show high stability, low cytotoxicity and high adsorption capacity for Hg<sup>2+</sup> in aqueous solution. The maximum adsorption capacity for Hg<sup>2+</sup> is about 7.5 times and 3.7 times that of bulk MoS<sub>2</sub> powder and unfunctionalized MoS<sub>2</sub> nanosheets, respectively. The adsorption kinetics and thermodynamics indicate that the adsorption of Hg<sup>2+</sup> is a monolayer chemical adsorption. Finally, the as-prepared MoS<sub>2</sub> nanosheets were applied for the removal of Hg<sup>2+</sup> in HepG2 cells and show a similar detoxification effect as that of the common antidote *meso*-2,3-dimercaptosuccinic acid, indicating the potential application of MoS<sub>2</sub> nanosheets for biological detoxification.

## 1. Introduction

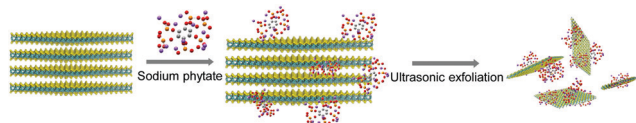
The mercury ion (Hg<sup>2+</sup>) is a highly toxic ion that can cause gastrointestinal mucosal and kidney failure, central nervous system damage and even death.<sup>1,2</sup> Hg<sup>2+</sup> contamination has become increasingly serious in our living environment and ecosystem. The development of efficient technologies for the removal of Hg<sup>2+</sup> has received tremendous attention. Traditional Hg<sup>2+</sup> removal technologies include chemical precipitation,<sup>3</sup> electrochemical processes,<sup>4</sup> adsorption,<sup>5</sup> membrane separation,<sup>6</sup> ion exchange<sup>7</sup> and solvent extraction.<sup>8</sup> Among these technologies, adsorption has the advantages of simple operation and low cost. Owing to the large number of sulfur (S) atoms exposed on the surface and the strong soft-soft interaction between Hg<sup>2+</sup> and S, molybdenum disulfide (MoS<sub>2</sub>), a typical layered transition-metal dichalcogenide (TMDC)<sup>9</sup> composed of a hexagonal plane with Mo atoms sandwiched by two sulfur atoms (S–Mo–S), can be used as a promising adsorbent for the removal of Hg<sup>2+</sup>.<sup>10</sup> Therefore, MoS<sub>2</sub> nanosheets with widened interlayer spacing,<sup>10</sup> two-dimensional (2D) MoS<sub>2</sub> nanosheets,<sup>11</sup>

oxygen-incorporated MoS<sub>2</sub> nanosheets,<sup>12</sup> Au/Fe<sub>3</sub>O<sub>4</sub>/MoS<sub>2</sub> aerogels,<sup>13</sup> d-MoS<sub>2</sub>/Fe<sub>3</sub>O<sub>4</sub> nanohybrids,<sup>14</sup> and cellulose/MoS<sub>2</sub>/Fe<sub>3</sub>O<sub>4</sub> composites<sup>15</sup> have been shown to be excellent adsorbents for the removal of Hg<sup>2+</sup> from aqueous solutions. Although these MoS<sub>2</sub>-based adsorbents show high adsorption capacity for Hg<sup>2+</sup>, they are still in the proof-of-concept phase and none of the MoS<sub>2</sub>-based adsorbents have been applied to remove Hg<sup>2+</sup> in real complex environmental and biological samples.

Bulk MoS<sub>2</sub> has strong chemical bonding within its layers and weak van der Waals forces between its layers. To expose more surfaces with S atoms to the outside for adsorption, the exfoliation of bulk MoS<sub>2</sub> into single- or few-layered nanosheets is necessary. To date, chemical exfoliation with alkali metal compounds and liquid-phase exfoliation are recognized as effective methods for the scale-up production of MoS<sub>2</sub> nanosheets.<sup>16</sup> However, chemical exfoliation requires a harsh and strong intercalating reagent (such as *n*-butyl lithium) followed by ultrasonic exfoliation under inert atmosphere.<sup>17,18</sup> Moreover, the preparation procedure is expensive, dangerous, long and tedious. Liquid-phase exfoliation of bulk MoS<sub>2</sub> via facile ultrasonic treatment in organic solvents, such as ethanol,<sup>19,20</sup> 2-propanol,<sup>21</sup> *N*-methyl-2-pyrrolidone,<sup>22</sup> acetone and alcohols (ethanol, methanol and isopropanol),<sup>23</sup> or in aqueous solution with auxiliary reagents such as thiolated surfactants,<sup>24</sup> polystyrene-polyacrylamide,<sup>25</sup> bovine serum albumin,<sup>26</sup> poly(3,4-ethylenedioxythiophene):poly-(styrenesulfonate),<sup>27</sup> Nafion,<sup>28</sup> and polymeric ionic liquids,<sup>29</sup> has been successfully developed and is widely applied in the fields of catalysis, organic solar cells, antibacterial materials, conductive films, *etc.* However, these organic solvents, surfactants, and polymers are mostly toxic, and will thus cause adverse effects in biological applications. Therefore, a facile method for the

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† Electronic supplementary information (ESI) available: Adsorption spectra of MoS<sub>2</sub> nanosheets at different conditions, XRD pattern, Raman spectra, adsorption spectra of MoS<sub>2</sub> nanosheets with different reagents, effect of pH on the adsorption amount, Langmuir adsorption isotherms, XPS spectra of MoS<sub>2</sub> nanosheets, adsorption selectivity, viability of HepG2 cells, comparison of yields of MoS<sub>2</sub> nanosheets, adsorption dynamics model parameters, the maximal adsorption capacity and correlation coefficient of Langmuir isotherms, comparison of the maximum adsorption capacity of MoS<sub>2</sub> for Hg<sup>2+</sup>. See DOI: 10.1039/d2ma00058j



**Scheme 1** Schematic illustration of the exfoliation of bulk MoS<sub>2</sub> and the functionalization of MoS<sub>2</sub> nanosheets by phytate.

exfoliation and functionalization of MoS<sub>2</sub> nanosheets with a green and cost-effective auxiliary reagent to reinforce their biocompatibility and functionality for biological application is highly desired.<sup>30,31</sup>

Herein, we report a facile green method to simultaneously exfoliate and functionalize MoS<sub>2</sub> nanosheets *via* ultrasonic treatment of bulk MoS<sub>2</sub> in sodium phytate-containing aqueous solution (Scheme 1). Sodium phytate is the sodium salt of phytic acid (myo-inositol-1,2,3,4,5,6-hexakisphosphate) extracted from rice bran and is a natural chelating agent. The as-prepared MoS<sub>2</sub> nanosheets show high stability in phosphate buffer solution in the pH range of 2.0–12.0 and low cytotoxicity. Taking advantage of the high binding affinity between Hg<sup>2+</sup> and S sites on the surface of MoS<sub>2</sub> sheets<sup>10,11</sup> and the biocompatible, biodegradable, and nontoxic phytate,<sup>32</sup> the as-prepared MoS<sub>2</sub> nanosheets were used as an adsorbent for the removal of Hg<sup>2+</sup> from water and as a detoxifier for the removal of Hg<sup>2+</sup> in HepG2 cells. Our results indicate that the as-prepared MoS<sub>2</sub> nanosheets display a detoxification effect that is comparable to that of *meso*-2,3-dimercaptosuccinic acid (DMSA), a heavy metal detoxifier approved by the Food and Drug Administration of USA for the treatment of lead and mercury toxicity both in children and adults. To the best of our knowledge, this work is the first example to use TMDC for heavy metal detoxification in living cells.

## 2. Experimental

### 2.1. Chemical and materials

Bulk MoS<sub>2</sub> powder and DMSA were purchased from Shanghai Aladdin Biochemical Technology. Sodium phytate was bought from Sangon Biotech (Shanghai) Co., Ltd. Hg<sup>2+</sup> standard solution (1000 µg mL<sup>-1</sup>) in 1.0 mol L<sup>-1</sup> HNO<sub>3</sub> was obtained from the National Nonferrous Metals and Electronic Materials Analysis and Testing Center of China, and was diluted to the desired concentration with water. Dulbecco's modified Eagle's medium (DMEM), phosphate buffered solution (PBS, 10 mmol L<sup>-1</sup>, pH 7.4) and Cell Counting Kit-8 (CCK-8) were purchased from GE Healthcare Life Sciences HyClone laboratories, Inc. Without specification, the metal ions were nitrate salts and anions were sodium salts. All water used was ultrapure water (18.2 MΩ cm, Millipore).

### 2.2. Exfoliation and functionalization of MoS<sub>2</sub> nanosheets by phytate

200 mg Bulk MoS<sub>2</sub> powder was added to 40 mL aqueous solution containing 40 mg sodium phytate. The suspension was ultrasonically treated for 35 h. The dispersion was centrifuged at 5000 rpm for 20 min to remove large particles, and the supernatant was further centrifuged at 12 000 rpm for 10 min to

remove excessive sodium phytate. The precipitate was collected and redispersed in water. The removal of sodium phytate in the supernatant was repeated three times, and the collected precipitate was redispersed in water for further use. In addition, bulk MoS<sub>2</sub> powder was separately exfoliated in Na<sub>2</sub>HPO<sub>4</sub> solution and water under the same conditions.

### 2.3. Adsorption of Hg<sup>2+</sup> by MoS<sub>2</sub> nanosheets.

MoS<sub>2</sub> nanosheets exfoliated by sodium phytate (6.4 mg) were added to 30 mL Hg<sup>2+</sup> solution with different concentrations (100 ng mL<sup>-1</sup>, 1 µg mL<sup>-1</sup>, 10 µg mL<sup>-1</sup> and 20 µg mL<sup>-1</sup>). The mixture solution was shaken at a rate of 300 rpm at room temperature (25 °C). An aliquot of 2 mL mixture solution was taken out at a certain time interval. After centrifugation at 12 000 rpm for 20 min, the supernatant was collected and filtered by a 0.22 µm filter to remove large particles. The concentration of residual Hg<sup>2+</sup> in the filtrate was determined by ICP-MS.

### 2.4. Cytotoxicity assay and detoxification experiment

For the cytotoxicity assay, human hepatoma cells (HepG2) were treated with 0.25% trypsin for 1 min. After the trypsin solution was discarded, HepG2 cells were dispersed in DMEM medium by blowing. Then, HepG2 cells (1 × 10<sup>5</sup> cell in each well) were plated in a 96-microwell microplate and cultured in DMEM at 37 °C in a humidified 5% CO<sub>2</sub> incubator for 24 h. After the cells were washed with PBS (10 mmol L<sup>-1</sup>, pH 7.4) three times to remove the secretions and culture medium, 100 µL MoS<sub>2</sub> nanosheets exfoliated by sodium phytate (0, 50, 100, 250, 500 and 1000 µg mL<sup>-1</sup>), Hg<sup>2+</sup> (0, 0.5, 1, 2, 5 and 10 µg mL<sup>-1</sup>), or DMSA (0, 50, 100, 150 and 200 µg mL<sup>-1</sup>) in DMEM were each added and cultured at 37 °C in a humidified 5% CO<sub>2</sub> incubator for 12 h. After being washed with PBS (10 mmol L<sup>-1</sup>, pH 7.4) three times, 10 µL CCK-8 and 90 µL DMEM were added to each well and the cells were incubated in the dark for 2 h. Finally, the absorbance of each well at 450 nm was recorded. Each experiment was repeated five times. The relative survival rate of the cells was calculated by taking the cell survival rate of the blank as 100%.

To investigate the detoxification effect of the MoS<sub>2</sub> nanosheets exfoliated by sodium phytate, 100 µL Hg<sup>2+</sup> (2 µg mL<sup>-1</sup>) in DMEM was added to HepG2 cells (1 × 10<sup>5</sup> cell) and incubated in the dark for 2 h. After the cells were washed with PBS (10 mmol L<sup>-1</sup>, pH 7.4) three times, 11 µL MoS<sub>2</sub> nanosheets or DMSA in DMEM (0, 50, 100, 150 and 200 µg mL<sup>-1</sup>) was further added. After further incubation for 12 h, the cells were washed with PBS (10 mmol L<sup>-1</sup>, pH 7.4) three times. The CCK-8 assay was used to detect the relative survival rate of the cells. Each experiment was repeated five times.

## 3. Results and discussion

### 3.1. Optimization of the exfoliation conditions

Bulk MoS<sub>2</sub> powder was ultrasonically exfoliated in sodium phytate aqueous solution. The exfoliation conditions (such as ultrasonication time, concentrations of sodium phytate and bulk MoS<sub>2</sub> powder) were optimized. The absorbance of the



MoS<sub>2</sub> nanosheets gradually increased with increasing time, and reached the platform until the ultrasonic time was up to 35 h (Fig. S1A, ESI†). The exfoliation yield, namely the mass ratio of the collected MoS<sub>2</sub> nanosheets to bulk MoS<sub>2</sub> powder, was increased with the concentration of sodium phytate until its concentration reached 1 mg mL<sup>-1</sup> (Fig. S1B, ESI†). However, when the concentration of sodium phytate exceeded 1 mg mL<sup>-1</sup>, the exfoliation yield was decreased. Similarly, the exfoliation yield was increased with the concentration of bulk MoS<sub>2</sub> powder until its concentration reached 8 mg mL<sup>-1</sup> (Fig. S1C, ESI†). The exfoliation yield was reduced instead with the further increase of bulk MoS<sub>2</sub> powder. Therefore, the optimal conditions for the exfoliation of MoS<sub>2</sub> nanosheets are 35 h ultrasonic time, 1 mg mL<sup>-1</sup> sodium phytate and 8 mg mL<sup>-1</sup> bulk MoS<sub>2</sub> powder. Under the optimal conditions, the exfoliation yield is 18.1%, which is 8 times that of the MoS<sub>2</sub> nanosheets exfoliated in pure water. As shown in Table S1 (ESI†), the ultrasonic exfoliation yield is also higher than those by ultrasonic exfoliation in most solvents. The effect of pH on the stability of MoS<sub>2</sub> nanosheets was also investigated (Fig. S1D, ESI†). MoS<sub>2</sub> nanosheets can be dispersed stably in phosphate buffer (10 mmol L<sup>-1</sup>) over a pH range of 2.0–12.0, indicating that the functionalization of phytate on the surface of the MoS<sub>2</sub> nanosheets endows their high stability in aqueous solution.

### 3.2. Characterization of MoS<sub>2</sub> nanosheets

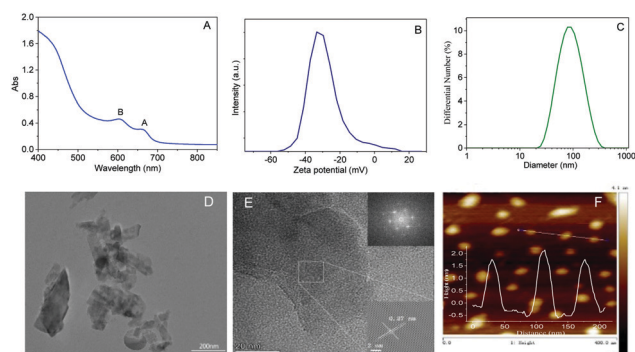
There are two distinctive absorption peaks at 665 nm and 603 nm (A and B excitons, respectively) in the absorption spectrum of the MoS<sub>2</sub> nanosheets solution (Fig. 1A), which are typical characteristics of the hexagonal symmetry space group of 2H-MoS<sub>2</sub>, corresponding to the direct excitonic transition of MoS<sub>2</sub> at the *K*-points of the Brillouin zone.<sup>23,28</sup> According to the relationship between the mean number of layers and the wavelength of the A-exciton ( $\lambda_A$ ) proposed by Coleman and co-workers,<sup>31</sup> the mean number of layers is estimated to be less than 3. The potential of MoS<sub>2</sub> nanosheets was measured to be -33.4 mV (Fig. 1B), indicating that the functionalization by

phytate endows the MoS<sub>2</sub> nanosheets with abundant negative charges and high hydrophilicity and dispersity in aqueous solution. Moreover, the MoS<sub>2</sub> nanosheets show monodispersity with the most probable size of about 90 nm (Fig. 1C).

As observed from the low-resolution TEM image (Fig. 1D), MoS<sub>2</sub> nanosheets are composed of sheet-like structures from monolayers to few layers with lateral sizes from several dozens to a few hundreds of nanometers. The ordered lattice and parallel fringe can be clearly observed in the high-resolution TEM (HRTEM) image (Fig. 1E). The lattice spacing of 0.27 nm is indexed to the (100) plane of MoS<sub>2</sub>.<sup>10,27,30</sup> The selected area electron diffraction (SAED) pattern (inset of Fig. 1E) further confirms the single crystalline nature of the hexagonal symmetry structure of MoS<sub>2</sub> nanosheets. The thickness of the MoS<sub>2</sub> nanosheets is less than 2 nm, which is observed from the AFM image (Fig. 1F). According to the Bragg equation, the interlayer distance of the MoS<sub>2</sub> nanosheets was calculated to be 0.62 nm.<sup>23,33</sup> Therefore, the layer number is about 2 to 3, which is in agreement with the result from the absorption spectrum.

As shown in the XRD patterns (Fig. S2A, ESI†), MoS<sub>2</sub> nanosheets maintain high crystallinity as bulk MoS<sub>2</sub> powder, indicating that there is no phase conversion during ultrasonic exfoliation. The diffraction peaks at 14.4°, 32.7°, 39.6°, 44.3°, 49.8°, 58.3° and 60.4° correspond to the (002), (100), (103), (006), (105), (110) and (008) crystal faces of 2H-MoS<sub>2</sub> (JCPDS No. 24-0513), respectively. The relative intensity of the (002) peaks in the MoS<sub>2</sub> nanosheets exfoliated by sodium phytate is weaker than bulk MoS<sub>2</sub> powder, indicating the successful exfoliation of the MoS<sub>2</sub> nanosheets.<sup>34</sup> Raman spectra (Fig. S2B, ESI†) show the characteristic in-plane (*E*<sub>2g</sub><sup>1</sup>) and out-of-plane (*A*<sub>1g</sub>) vibrational modes of MoS<sub>2</sub> at about 380 cm<sup>-1</sup> and 405 cm<sup>-1</sup>.<sup>11,23</sup> Compared with bulk MoS<sub>2</sub> powder, the vibration modes of MoS<sub>2</sub> nanosheets are red-shifted due to the weakening of the van der Waals force between the layers,<sup>35</sup> which also suggests the successful exfoliation of bulk MoS<sub>2</sub> into few-layered nanosheets.<sup>36</sup> There is also a rather weak second-order scattering process near 450 cm<sup>-1</sup>, attributed to the longitudinal acoustic vibrational mode of the 2H phase monolayer MoS<sub>2</sub> nanosheets.<sup>37</sup>

The survey XPS spectrum (Fig. 2A) indicates there are Mo, S, P, C and O elements in the MoS<sub>2</sub> nanosheets exfoliated by phytate, indicating that phytate is functionalized on the surface of the MoS<sub>2</sub> nanosheets. In the Mo 3d core-level XPS spectrum of MoS<sub>2</sub> nanosheets exfoliated by sodium phytate (Fig. 2B), the peaks at 229.3 eV and 232.5 eV can be attributed to Mo<sup>4+</sup> 3d<sub>5/2</sub> and Mo<sup>4+</sup> 3d<sub>3/2</sub> of 2H-MoS<sub>2</sub>. The weak peaks at 235.7 eV and 233.0 eV can be ascribed to Mo<sup>6+</sup> 3d<sub>3/2</sub> and Mo<sup>6+</sup> 3d<sub>5/2</sub>, respectively. This also can be observed in the Mo 3d core-level XPS spectra of the MoS<sub>2</sub> nanosheets exfoliated by H<sub>2</sub>O and bulk MoS<sub>2</sub> powder, and may come from MoO<sub>3</sub> or MoO<sub>4</sub><sup>2-</sup>,<sup>23,30</sup> indicating that the surface of the MoS<sub>2</sub> nanosheets is partially oxidized on exposure to air.<sup>23,38</sup> The weak feature peak at 226.6 eV originated from the S 2s.<sup>39</sup> In the S 2p core-level XPS spectrum (Fig. 2C), the peaks at 162.2 eV and 163.4 eV can be attributed to S 2p<sub>3/2</sub> and S 2p<sub>1/2</sub>, respectively. In addition, there exists a weak peak at 133.8 eV (Fig. 2D), corresponding to the



**Fig. 1** (A) Absorption spectrum, (B) zeta potential distribution, (C) size distribution, (D) TEM image, (E) HRTEM and (F) AFM images of MoS<sub>2</sub> nanosheets exfoliated by sodium phytate. Insets of (E) and (F) are the SAED pattern and the height profile of MoS<sub>2</sub> nanosheets exfoliated by sodium phytate, respectively. The concentration of MoS<sub>2</sub> nanosheets in (A) is 1.12 mg mL<sup>-1</sup>.





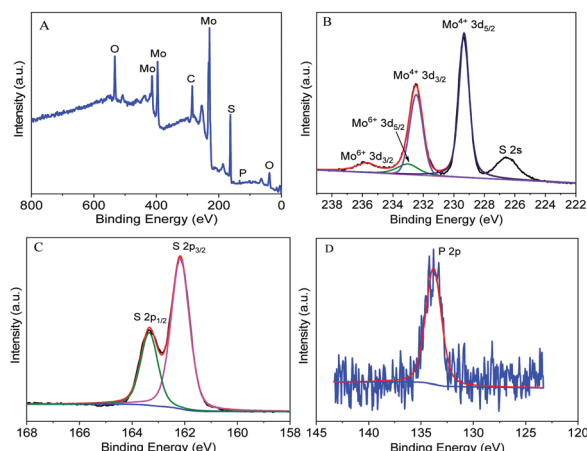


Fig. 2 The survey (A), Mo 3d (B), S 2p (C) and P 2p (D) core-level XPS spectra of MoS<sub>2</sub> nanosheets exfoliated by sodium phytate.

binding energy of P 2p. From the above discussion, there exists a weak peak at 133.8 eV (Fig. 2D), corresponding to the binding energy of P 2p. From the above discussion, the MoS<sub>2</sub> nanosheets are successfully exfoliated from bulk MoS<sub>2</sub> powder in sodium phytate solution, and the phytate molecules are adsorbed onto the surface of the MoS<sub>2</sub> nanosheets.

### 3.3. Exfoliation mechanism

To investigate the effect of phytate on the exfoliation of MoS<sub>2</sub> nanosheets, bulk MoS<sub>2</sub> powder was also ultrasonically exfoliated in Na<sub>2</sub>HPO<sub>4</sub> solution and ultra-pure water with the same ultrasonic time. The exfoliation yield is positively correlated with the absorption intensity of the MoS<sub>2</sub> nanosheets solution. As shown in Fig. S3 (ESI<sup>†</sup>), the exfoliation yield is on the order of ultra-pure water < Na<sub>2</sub>HPO<sub>4</sub> solution < sodium phytate. The exfoliation yield in the Na<sub>2</sub>HPO<sub>4</sub> solution is higher than that in ultra-pure water, indicating that the phosphate ion can promote the exfoliation of the MoS<sub>2</sub> nanosheets. The exfoliation yield in sodium phytate is higher than that in Na<sub>2</sub>HPO<sub>4</sub> solution, even though the molar concentration of Na<sub>2</sub>HPO<sub>4</sub> is nearly 20-folds that of sodium phytate. There are six phosphate groups in the phytate molecule, which is more efficient than Na<sub>2</sub>HPO<sub>4</sub> in adsorbing on the surface of the MoS<sub>2</sub> nanosheets and inserting into the interlayers of bulk MoS<sub>2</sub>, resulting in high exfoliation yield. The exfoliation process of bulk MoS<sub>2</sub> by sodium phytate is proposed in Scheme 1. The van der Waals force between each layer of bulk MoS<sub>2</sub> is destroyed by ultrasonication, and phytate molecules intercalate into the edges of the crystal interlayers to facilitate the production of MoS<sub>2</sub> nanosheets. After exfoliation, the adsorption of phytate on the MoS<sub>2</sub> nanosheets can be stably dispersed in water.

### 3.4. Adsorption of Hg<sup>2+</sup> by MoS<sub>2</sub> nanosheets

The strong soft-soft interaction between Hg<sup>2+</sup> and S atoms and the powerful chelation effect of phytate on the surface of the MoS<sub>2</sub> nanosheets inspired us to utilize MoS<sub>2</sub> nanosheets as adsorbents for the removal of Hg<sup>2+</sup>. The removal efficiency (RE)

and adsorption capacity ( $q_e$ ) were calculated from eqn (1) and (2), respectively.

$$RE = \frac{(C_0 - C_e)}{C_0} \quad (1)$$

$$q_e = \frac{(C_0 - C_e) \times V}{m} \quad (2)$$

where  $C_0$  ( $\mu\text{g mL}^{-1}$ ) is the initial concentration of Hg<sup>2+</sup>,  $C_e$  ( $\mu\text{g mL}^{-1}$ ) is the equilibrium concentration of Hg<sup>2+</sup> after adsorption,  $q_e$  ( $\text{mg g}^{-1}$ ) is the adsorption capacity of the MoS<sub>2</sub> nanosheets at equilibrium time,  $V$  (mL) is the solution volume, and  $m$  (g) is the mass of the adsorbent.

The effect of pH on the adsorption of Hg<sup>2+</sup> by MoS<sub>2</sub> nanosheets exfoliated by sodium phytate was first investigated. As shown in Fig. S4 (ESI<sup>†</sup>), the adsorption capacity is basically unchanged over the pH range of 4.0–6.0. When the pH is higher than 6.0, the adsorption capacity is slightly decreased due to the hydrolysis of Hg<sup>2+</sup>. Therefore, the following adsorption experiments were conducted under acidic condition. The effect of the initial concentration of Hg<sup>2+</sup> on the adsorption by MoS<sub>2</sub> nanosheets was investigated. As shown in Fig. 3A, the adsorption amount is gradually increased with the increasing initial concentration of Hg<sup>2+</sup>, and the adsorption reaches equilibrium after 1 h. However, the RE (Fig. S5, ESI<sup>†</sup>) is almost unchanged due to the low initial Hg<sup>2+</sup> concentration ( $C_0$ ). The adsorption kinetics was fitted by the pseudo-second-order kinetic model (3).<sup>11,13</sup>

$$\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{t}{q_e} \quad (3)$$

$k_2$  ( $\text{g mg}^{-1} \text{h}^{-1}$ ) is the pseudo-second-order adsorption rate constant,  $q_t$  ( $\text{mg g}^{-1}$ ) is the adsorption amount of Hg<sup>2+</sup> at time  $t$  (h), and  $q_e$  ( $\text{mg g}^{-1}$ ) is the adsorption capacity at equilibrium. As shown in Fig. 3B and Table S2 (ESI<sup>†</sup>), the calculated values ( $q_{e,\text{cal}}$ ) are close to the experimental values ( $q_{e,\text{exp}}$ ), which indicates that the adsorption of Hg<sup>2+</sup> by MoS<sub>2</sub> nanosheets is mainly chemical adsorption.

The adsorption thermodynamics was fitted by the Langmuir isotherm model (4).<sup>11,13</sup>

$$\frac{c_e}{q_e} = c_e \frac{1}{q_{\text{max}}} + \frac{1}{K_L q_{\text{max}}} \quad (4)$$

$c_e$  is the equilibrium concentration ( $\mu\text{g mL}^{-1}$ ) of Hg<sup>2+</sup>.  $q_e$  and  $q_{\text{max}}$  are the equilibrium adsorption capacity and the maximum

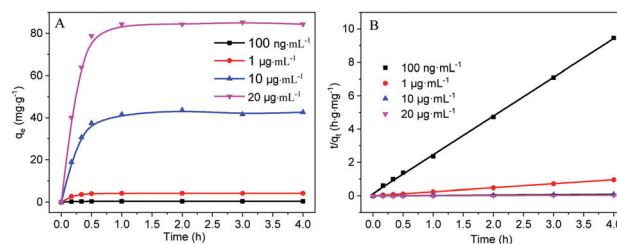


Fig. 3 (A) Adsorption of Hg<sup>2+</sup> by MoS<sub>2</sub> nanosheets exfoliated by sodium phytate with different initial concentrations of Hg<sup>2+</sup>. (B) Pseudo-second-order kinetic curves for Hg<sup>2+</sup> adsorption.



adsorption capacity ( $\text{mg g}^{-1}$ ), respectively.  $K_L$  is the Langmuir constant. Three temperatures ( $25^\circ\text{C}$ ,  $30^\circ\text{C}$ ,  $40^\circ\text{C}$ ) were selected to investigate the adsorption thermodynamics. The fitting results are shown in Fig. S6 (ESI<sup>†</sup>) and listed in Table S3 (ESI<sup>†</sup>). The adsorption isotherm fitted the Langmuir model well, indicating that the adsorption of  $\text{Hg}^{2+}$  is a monolayer adsorption. The adsorption thermodynamics also indicates that the low temperature is in favor of  $\text{Hg}^{2+}$  adsorption and the highest  $q_{\text{max}}$  ( $313.48 \text{ mg g}^{-1}$ ) was obtained at room temperature ( $25^\circ\text{C}$ ). The Langmuir isotherm model was also used to compare the maximum adsorption capacity of  $\text{MoS}_2$  nanosheets exfoliated by sodium phytate, and  $\text{MoS}_2$  nanosheets exfoliated by  $\text{H}_2\text{O}$  and bulk  $\text{MoS}_2$  powder. The results are listed in Table S4 (ESI<sup>†</sup>). The  $q_{\text{max}}$  value obtained from the  $\text{MoS}_2$  nanosheets exfoliated by sodium phytate is about 7.5 times that of the  $\text{MoS}_2$  powder, and about 3.7 times that of the  $\text{MoS}_2$  nanosheets exfoliated by  $\text{H}_2\text{O}$ . This result indicates that besides the complexation of  $\text{Hg}^{2+}$  with the intrinsic S atoms of the  $\text{MoS}_2$  nanosheets,<sup>40</sup> the chelating effect of phytate on the surface of the  $\text{MoS}_2$  nanosheets plays the main role in  $\text{Hg}^{2+}$  adsorption. The interaction between the adsorbed  $\text{Hg}^{2+}$  ions and  $\text{MoS}_2$  nanosheets exfoliated by sodium phytate were further confirmed by XPS spectrum. As shown in Fig. S7 (ESI<sup>†</sup>), a new element (Hg) appears in the survey XPS spectrum of  $\text{MoS}_2$  nanosheets after  $\text{Hg}^{2+}$  adsorption. Furthermore, a new peak at  $161.9 \text{ eV}$  related to  $\text{S } 2\text{p}_{1/2}$  of  $\text{HgS}$  is observed in the  $\text{S } 2\text{p}$  core-level XPS spectrum, and the peaks at  $100.9 \text{ eV}$  and  $104.9 \text{ eV}$  are attributed to  $\text{Hg}^{2+} 4\text{f}_{7/2}$  and  $\text{Hg}^{2+} 4\text{f}_{5/2}$  of  $\text{HgS}$  in the  $\text{Hg } 4\text{f}$  core-level XPS spectrum,<sup>41</sup> which indicate the complex between S and  $\text{Hg}^{2+}$ . The chelating effect of phytate can be verified by the complex between O and  $\text{Hg}^{2+}$ . In the  $\text{Hg } 4\text{f}$  core-level XPS spectrum, the peaks at  $102.0 \text{ eV}$  and  $106.0 \text{ eV}$  are assigned to  $\text{HgO}$ .<sup>12</sup> In the O  $1\text{s}$  core-level XPS spectrum, the peaks at  $530.7 \text{ eV}$  and  $532.5 \text{ eV}$  are assigned to  $\text{MoO}_3$  and  $\text{HgO}$ , respectively, after  $\text{Hg}^{2+}$  adsorption.<sup>11</sup> However, without adsorption of  $\text{Hg}^{2+}$ , the peaks at  $530.4 \text{ eV}$  and  $531.6 \text{ eV}$  in the O  $1\text{s}$  core-level XPS spectrum might be deconvoluted into  $\text{MoO}_3$  and the adsorbed water, respectively.<sup>11</sup>

The adsorption of  $\text{MoS}_2$  nanosheets exfoliated by sodium phytate toward other heavy metal ions ( $\text{Pb}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Mn}^{2+}$  and  $\text{Zn}^{2+}$ ) and anions ( $\text{NO}_2^-$ ,  $\text{NO}_3^-$ ,  $\text{SO}_4^{2-}$  and  $\text{CO}_3^{2-}$ ) and their mixture with  $\text{Hg}^{2+}$  were investigated. As shown in Fig. S8 (ESI<sup>†</sup>), the  $\text{MoS}_2$  nanosheets show a certain adsorption on  $\text{Pb}^{2+}$  and a small amount of adsorption on  $\text{Cd}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Mn}^{2+}$  and  $\text{Zn}^{2+}$ . However, the  $\text{MoS}_2$  nanosheets show the best adsorption on  $\text{Hg}^{2+}$ . Moreover, the co-existing heavy metal ions and anions almost do not affect the adsorption of  $\text{Hg}^{2+}$ . In the co-existence of heavy ions and anions,  $\text{Hg}^{2+}$  ( $100 \text{ ng mL}^{-1}$ ) still could be reduced to less than  $1 \text{ ng mL}^{-1}$ , which is the permission limit of mercury in drinking water provided by the national standard of China. These results indicate the high selectivity of  $\text{MoS}_2$  nanosheets exfoliated by sodium phytate for  $\text{Hg}^{2+}$  adsorption due to the high affinity between  $\text{Hg}^{2+}$  and S on the surface of  $\text{MoS}_2$  nanosheets.<sup>10,11</sup>

### 3.5. Cell detoxification application

Before the cell detoxification experiment, MTT experiments were conducted to detect the cytotoxicity of  $\text{MoS}_2$  nanosheets

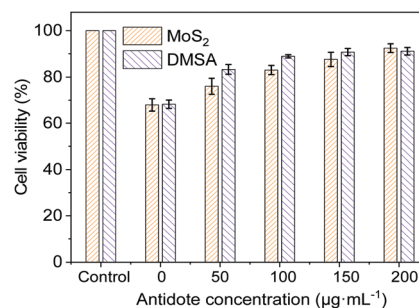


Fig. 4 Viability of HepG2 cells pre-treated with  $\text{Hg}^{2+}$  ( $2 \mu\text{g mL}^{-1}$ ) for 2 h followed by incubation with different concentrations of  $\text{MoS}_2$  nanosheets exfoliated by sodium phytate or DMSA for 12 h. The control was normal HepG2 cells.

exfoliated by sodium phytate,  $\text{Hg}^{2+}$  and DMSA. As shown in Fig. S9A (ESI<sup>†</sup>), the HepG2 cells maintain more than 95% cell proliferation capacity even though the concentration of the  $\text{MoS}_2$  nanosheets is up to  $250 \mu\text{g mL}^{-1}$ , indicating that the  $\text{MoS}_2$  nanosheets are highly biocompatible and have low toxicity.  $\text{Hg}^{2+}$  can induce a significant cytotoxicity in a dose-dependent manner (Fig. S9B, ESI<sup>†</sup>). Even low concentrations of  $\text{Hg}^{2+}$  can cause intoxication in HepG2 cells.  $2 \mu\text{g mL}^{-1}$  and  $5 \mu\text{g mL}^{-1}$   $\text{Hg}^{2+}$  resulted in the decrease of proliferation capacities of HepG2 cells to less than 70% and 30%, respectively. In addition, DMSA showed low cytotoxicity. The proliferation capacity was more than 90% in the DMSA concentration range of  $0\text{--}200 \mu\text{g mL}^{-1}$  (Fig. S9C, ESI<sup>†</sup>).

The high adsorption capacity for  $\text{Hg}^{2+}$  and low cytotoxicity of  $\text{MoS}_2$  nanosheets inspired us to investigate their detoxification effect on  $\text{Hg}^{2+}$  poisoning. HepG2 cells were incubated with  $\text{Hg}^{2+}$  ( $2 \mu\text{g mL}^{-1}$ ) to induce moderate cytotoxicity ( $\sim 70\%$  cell viability), and then incubated with different concentrations of  $\text{MoS}_2$  nanosheets exfoliated by sodium phytate. As shown in Fig. 4, with the increasing concentration of  $\text{MoS}_2$  nanosheets, the proliferation capacities of HepG2 cells gradually increased. When the concentration of  $\text{MoS}_2$  nanosheets reached  $200 \mu\text{g mL}^{-1}$ , the proliferation capacity of HepG2 cells was greater than 90%, which indicated that the cytotoxicity induced by  $\text{Hg}^{2+}$  was almost eliminated because the proliferation capacity of the HepG2 cells is comparable with that observed only in the presence of  $200 \mu\text{g mL}^{-1}$   $\text{MoS}_2$  nanosheets. Compared to the detoxifier DMSA, the  $\text{MoS}_2$  nanosheets show a similar detoxification effect, which indicates that the  $\text{MoS}_2$  nanosheets exfoliated by sodium phytate can be used as a potential detoxifier for  $\text{Hg}^{2+}$  poisoning.

## 4. Conclusions

In summary, the simultaneous exfoliation and functionalization of few-layered  $\text{MoS}_2$  nanosheets were achieved *via* ultrasonic treatment by using sodium phytate as an auxiliary reagent. The functionalization of phytate not only can facilitate the exfoliation of  $\text{MoS}_2$  nanosheets, but also endows  $\text{MoS}_2$  nanosheets with high stability in aqueous solution over a broad



pH range. This preparation procedure is simple, green, cost-effective, as well as high yield. The as-prepared MoS<sub>2</sub> nanosheets show the effective and selective adsorption for Hg<sup>2+</sup> in water, and can reduce Hg<sup>2+</sup> ion (100 ng mL<sup>-1</sup>) to a concentration below the Chinese national permission limit of mercury (1 ng mL<sup>-1</sup>) in drinking water. The as-prepared MoS<sub>2</sub> nanosheets also displayed low cytotoxicity for HepG2 cells, and showed a detoxification effect similar to that of the common detoxifier DMSA. The study may open a new application direction of TMDC in the development of novel therapeutics for the treatment of heavy metal poisoning.

## Author contributions

Shanshan Xing: conceptualization, validation, investigation, data curation, writing – original draft; Chunqiu Xia: methodology, investigation, writing – original draft; Xinyi Liu: methodology, data curation; Liangqia Guo: conceptualization, funding acquisition, writing – review & editing, supervision; Fengfu Fu: conceptualization, writing – review & editing.

## Conflicts of interest

There are no conflicts to declare.

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