Lab on a Chip



CORRECTION

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Correction: Quantifying nanotherapeutic penetration using a hydrogel-based microsystem as a new 3D *in vitro* platform

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Correction for 'Quantifying nanotherapeutic penetration using a hydrogel-based microsystem as a new 3D *in vitro* platform' by Saba Goodarzi *et al.*, *Lab Chip*, 2021, **21**, 2495–2510, DOI: 10.1039/D1LC00192B.

In Fig. 2E the legend of the graph should be inversed. The corrected legend is: white bars#2D, black bars#3D. The corrected figure is shown below.

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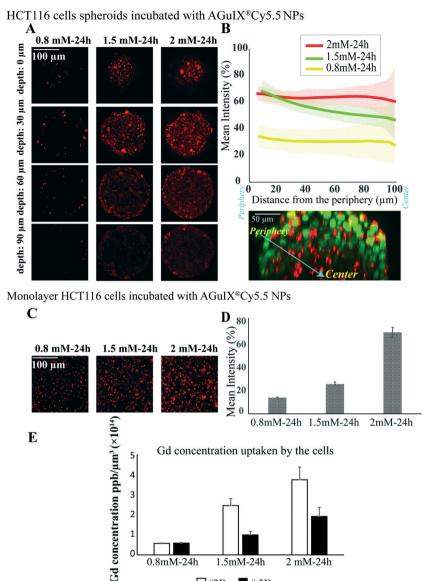


Fig. 2 Quantification of penetration and cellular uptake of AGulX®-Cy5.5 nanoparticles in HCT-116 tumour spheroids and monolayer cell culture. (A) Representative confocal fluorescence images of HCT-116 spheroids incubated with 0.8, 1.5 and 2 mM concentration of AGuIX®-Cy5.5 for 24 h for four different depths (0, 30, 60 and 90 µm). (B) Mean intensity along with standard deviation (light colors) of AGulX®-Cy5.5 as a function of the distance from the spheroid periphery (see the orthogonal view in the inset, green = nuclei, red = AGuIX®-Cy5.5) for 0.8 mM (yellow, N = 73), 1.5 mM (green, N = 68) and 2 mM (red, N = 121); three independent experiments. (C) Representative confocal fluorescence images of monolayer HCT-116 cells exposed to AGuIX®-Cy5.5 nanoparticles with 0.8, 1.5 and 2 mM concentration. (D) Quantification of the mean intensity of AGuIX®-Cy5.5 nanoparticles in maximal projection of confocal fluorescence images of monolayer cells after 24 h of incubation with different AGuIX®-Cy5.5 concentrations: 0.8 mM (yellow, N = 40), 1.5 mM (green, N = 40) and 2 mM (red, N = 40); three independent experiments. Error bars represent the standard deviations. (E) Mean and standard deviation of the concentration of Gd (ppb μm⁻³) uptaken by the cells after incubation with 0.8, 1.5 and 2 mM concentration of AGuIX® for 24 h in HCT-116 cell spheroids and monolayer cell culture measured with ICP-MS (N = 6, two independent experiments).

□#2D ■#3D

The Royal Society of Chemistry apologises for these errors and any consequent inconvenience to authors and readers.