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Hydrogenation of CO₂ to formic acid in biphasic systems using aqueous solutions of amino acids as the product phase†

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Carbon capture and utilization is considered a promising approach for introducing CO₂ into the chemical value chain, especially in combination with bioenergy applications (BECCU). We report here on the catalytic hydrogenation of CO₂ to formic acid in a biphasic reaction system using aqueous solutions of amino acids as the product phase and possible capture solutions for biogenic CO₂. The molecular structure of the ruthenium catalyst and the catalyst phase were matched through a combined design process identifying *n*-dodecanol (lauryl alcohol) as the preferred "green" solvent. A total turnover number (TON) of over 100 000 mole HCOOH per mole of catalyst (46 582 g HCOOH per g of Ru) with minimal contamination of the aqueous phase with metal or organic solvent was obtained. The resulting aqueous solutions attained almost quantitative conversions with up to 0.94 mol formic acid per mol amino acid (ca. 108 g HCOOH per kg). Such solutions may find use directly, or after upgrading, in agricultural applications without the need for energy intensive and costly isolation of pure formic acid.

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Introduction

Formic acid (FA) is one of the most widely discussed target products for hydrogenation of CO₂ with a significant potential to reduce the carbon footprint as compared to its fossil-based production.^{1–8} However, major challenges are the energy demand for CO₂ capture and release^{9,10} as well as isolation of the formic acid product from the reaction mixture.¹¹ In the present paper, we demonstrate a concept to integrate carbon capture technologies based on proteinogenic amino acids with catalytic CO₂ conversion to yield HCOOH solutions with a potential for agricultural applications, minimizing both upstream and downstream unit operations. This opens the possibility for the valorization of waste CO₂ streams from biogas production directly with local and decentralized applications in the same industrial sector.

The most advanced technologies for CO₂ capture from flue gases rely on basic organic amines, such as monoethanolamine (MEA) or methyldiethanolamine (MDEA).^{12,13} In the search for bio-based alternatives, amino acids and their salts have been investigated.^{14–17} Their applicability in reversible

carbon capture has been demonstrated with higher reaction rates and CO₂-loading for the respective potassium salts as compared to the neutral amino acids.^{17–19} Scrubbing solutions based on amino acid salts have been applied already at the pilot plant scale using PostCapTM technology.²⁰

Formic acid has a number of applications in relation to biological processes, *e.g.* as a biomass preservative in silage, or as an additive in the food or pharma industry.²¹ In academic research, formic acid has been shown to enhance microbial growth and/or product formation rates during fermentation processes.^{22–25}

Most notably in the present context, mixtures of formic acid with proteinogenic amino acids have potential applications as livestock feed. Methionine^{26,27} and arginine²⁸ are limiting amino acids in poultry feed, and supplements of these amino acids are commercially available.²⁹ Similarly, arginine^{30–32} as well as methionine and histidine are essential amino acids for pig breeding.^{33,34} At the same time, formic acid and formic acid salts are used as preservatives in animal feeds.^{35–37} Aqueous solutions of mixtures of amino acids and formic acid may therefore constitute attractive target products for use in animal farming.

We therefore envisage the use of catalytic CO₂ hydrogenation as the molecular relay integrating upstream CO₂ capture in aqueous amino acid solutions directly with the potential for downstream use of the resulting product phases (Fig. 1).

The essential prerequisite to establishing potential value chains from biogenic CO₂ sources to agricultural applications

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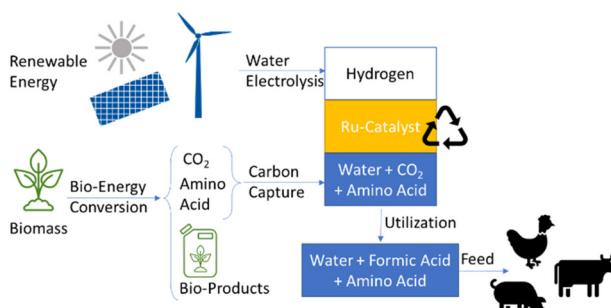


Fig. 1 Envisaged BECCU concept from biomass and renewable energy to amino acid-formic acid solutions as livestock feed.

is the effective separation of the organometallic catalyst from the aqueous product solution. The catalyst phase must be chosen to avoid the leaching of metals or ligands as well as cross-contamination with organic solvents into the aqueous phase. A tailored biphasic reaction system enabling such a fully integrated process scheme is presented in this paper.

Results and discussion

The catalytic hydrogenation of CO_2 in the presence of aqueous amino acid solutions has been demonstrated recently using homogeneous catalysts in THF/water mixtures in the context of reversible hydrogen storage.^{38,39} For the use of aqueous product solutions most directly in the applications envisaged here, essential parameters are the choices of amino acid and solvent used as the catalyst phase (Scheme 1). However, several concepts for liquid–liquid biphasic systems to separate organometallic catalysts from very polar formic acid product phases have been reported.^{40–49} Based on the experience in our laboratories,^{50–52} we initiated a systematic study to design a

catalytic system combining high catalyst productivity with effective retention and separation from the amino-acid-based product phase.

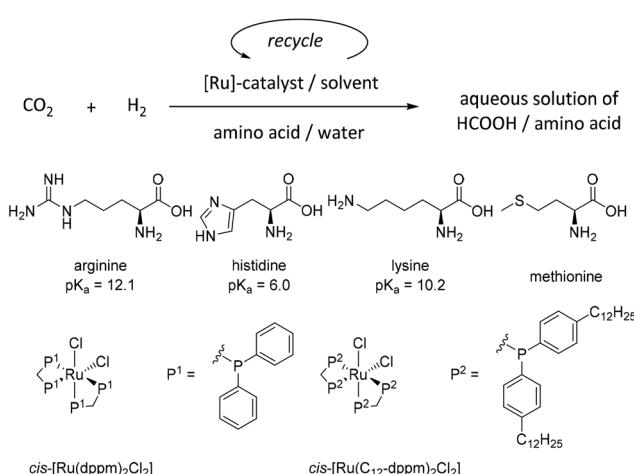
The organometallic complex *cis*-[Ru(dppm)₂Cl₂]⁵⁰ (dppm = bis-diphenylphosphinomethane) and its analogous congener tagged with hydrophobic side chains *cis*-[Ru(C₁₂-dppm)₂Cl₂]⁵³ were selected as catalyst precursors to cover a broad range of potential solvents for catalyst separation and recycling. Previous studies showed that they form the corresponding monohydride complexes under hydrogen pressure, which can be assumed as catalytically active species in line with what is generally known about Ru-phosphine catalysts in this field.⁵⁴

The proteinogenic amino acids arginine (Arg), histidine (His), lysine (Lys), and methionine (Met) were used in the aqueous phase (Scheme 1). In a first screening, the amino acids Arg, Lys, and His carrying basic side groups were studied because amines are known to capture CO_2 as well as stabilize HCOOH, effectively shifting the reaction equilibrium to the formic acid side. As the catalyst phase, the solvent methyl isobutyl carbinol (MIBC) in combination with *cis*-[Ru(dppm)₂Cl₂] as catalyst precursor was evaluated due to the high activity shown in previous studies on biphasic systems using aqueous MEA solutions as the capture and product phase.⁵⁰ The volume ratio between MIBC and water was adjusted to 2 : 3 with amino acid concentrations of 0.86 M for Arg and Lys and 0.25 M for His, based on their respective solubilities. The catalyst concentration in the MIBC phase was 0.5 mM corresponding to loadings of 1 : 2580 and 1 : 750, respectively.

The reactions were carried out under initial partial pressures (at room temperature) of 30 bar CO_2 and 60 bar H_2 and a reaction temperature of 70 °C. The reaction times were determined by monitoring the pressure drop until no further decrease of pressure was observed (ESI Fig. S3†). The final concentration of formic acid and the corresponding ratio of HCOOH to amino acid were determined by ¹H NMR spectroscopy. The initial rates, as measured from the pressure drop, are given as mol HCOOH per mol ruthenium and hour (*i.e.*, turnover frequency, TOF [h⁻¹]). The results are shown together with the results for MEA as the benchmark in Fig. 2.

A formic acid/amino acid ratio of approximately 1 : 1 was reached with Arg and His, corresponding to the thermodynamic limit for amine bases in an aqueous solution.^{1,55–57} For Lys, the reaction did not proceed beyond a ratio of 0.5 : 1. Given the long reaction time in this case, this may reflect, at least in part, catalyst deactivation. Most notably, the productivity with Arg matched that of MEA very well and the remarkable initial rate of 23 000 h⁻¹ even surpassed the benchmark case significantly.

While the organic solvent MIBC allowed for a very fast reaction, it also resulted in significant cross-solubility into the aqueous phase of 0.32 mmol or 2.2%. In order to minimize any mutual cross-solubility, the nonpolar solvent *n*-tetradecane was investigated, which has also been used for CO_2 hydrogenation in biphasic systems previously.⁵⁸ The lipophilic-tagged ligand bis(bis(4-dodecylphenyl)phosphanyl)methane (C₁₂-dppm) was used to ensure solubility and exclusive partitioning



Scheme 1 Reaction system for the catalytic conversion of CO_2 and H_2 to formic acid, featuring pre-catalysts *cis*-[Ru(dppm)₂Cl₂] and *cis*-[Ru(C₁₂-dppm)₂Cl₂] in methyl isobutyl carbinol (MIBC) and the amino acids used in the present study.



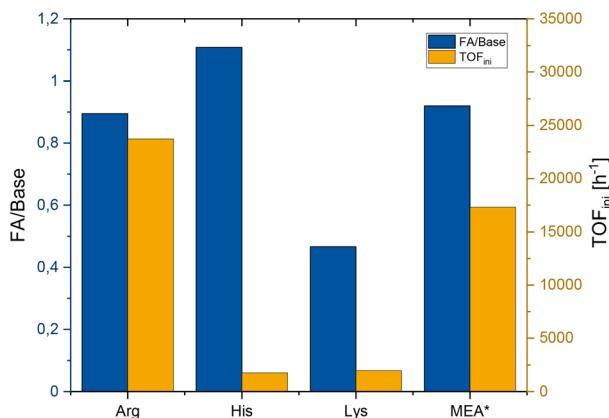


Fig. 2 Formic acid/amino acid ratio and initial TOF for Arg, His and Lys. Conditions: 30 bar CO₂, 60 bar H₂, 2.58 mmol Arg/Lys or 0.75 mmol His in water (3 mL), 1 μ mol *cis*-[Ru(dppm)₂Cl₂] in MIBC (2 mL), 70 °C, reaction time until the pressure was constant. *Values for comparison from Scott *et al.* with MEA (7.9 mmol MEA in 2 mL H₂O, 4.1 μ mol *cis*-[Ru(dppm)₂Cl₂] in 1.5 mL MIBC, 30 bar CO₂, 60 bar H₂, 70 °C).⁵⁰

of the ruthenium catalyst in the organic phase (Fig. 3).¹¹ Quantitative conversion for Arg and His was reached again with the more basic amino acid. For Lys, only small amounts of formic acid were obtained even with prolonged reaction times.

In general, however, the rates were nearly two orders of magnitude lower with tetradecane/*cis*-[Ru(C₁₂-dppm)₂Cl₂] than with MIBC/*cis*-[Ru(dppm)₂Cl₂]. In order to separate the electronic effect of the ligand modification from the solvent effects, the C₁₂-tagged complex was also used with MIBC as the catalyst phase, resulting in a TOF_{ini} of 2827 h⁻¹. While this confirmed the expected reduced activity of the more electron rich C₁₂-tagged complex,^{59,60} it also revealed a major contribution of the chosen solvent. Therefore, a detailed screening of solvents possessing a miscibility gap with water using the same pre-catalyst *cis*-[Ru(C₁₂-dppm)₂Cl₂] under identical conditions with Arg as the amino acid was undertaken.

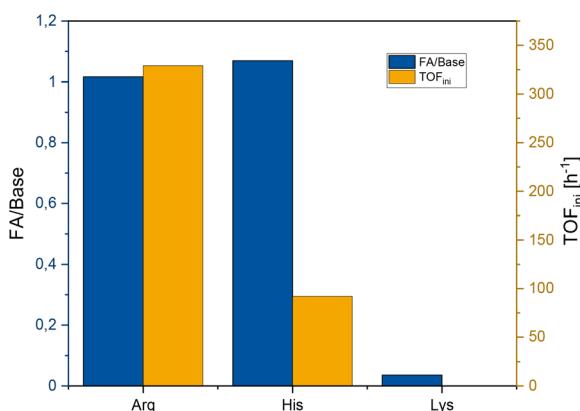


Fig. 3 Formic acid/base ratio and initial TOF for Arg, His and Lys. Conditions: 30 bar CO₂, 60 bar H₂, 2.58 mmol Arg/Lys or 0.75 mmol His, respectively, in water (3 mL), 1 μ mol *cis*-[Ru(C₁₂-dppm)₂Cl₂] in tetradecane (2 mL), 70 °C, reaction time until the pressure was constant.

The resulting initial activities are shown in Fig. 4. The results demonstrate the drastic influence of the chosen organic phase spanning two orders of magnitude in the rate between the lowest initial TOF observed with hexanol and the highest activity with ethyl acetate. No obvious correlations between specific functional groups or structural units in the solvents are apparent. Also, solvatochromic and other solvent parameters (ESI Fig. S6–S10†), gas solubilities (ESI Fig. S11 and S12†) or cross-solubility into the aqueous phase (ESI, Fig. S13†) did not show any clear trend. Mass transfer limitations⁶¹ were excluded by variation of the catalyst concentration. As the activity did not increase significantly at a lower catalyst loading, the capacity of all catalytically active centers was sufficiently utilized for selected solvents from either side of the reactivity range (ESI, Fig. S14†).

An interesting volcano-type trend results from the correlation of TOF_{ini} values with dielectric constants of the solvents, as shown in Fig. 5. Of the three top-performing solvents, *n*-dodecanol – also known as lauryl alcohol – best meets the design criteria for the target application: it shows practically no cross-solubility in water, it is a fatty alcohol derived from biomass, and it is compatible with biological applications, for example in consumer care products. Therefore, this “green” solvent was used for further optimization.

The solubility limit of arginine in the neutral, aqueous solution is about 1 M, due to the betaine form of the amino acid. We anticipated that the solubility would be significantly higher at low pH resulting from the presence of CO₂ at the beginning and the formation of HCOOH at the end of the reaction. We therefore explored the possibility of using over-saturated CO₂ solutions to achieve higher product concentrations.

In Scheme 2, pictures of the reaction mixture of 12 mmol Arg with 2 mL water (corresponding to 6 M upon dissolution) and 2 mL catalyst solution are shown before the reaction, after pressurization with CO₂, and after the reaction. Before pressur-

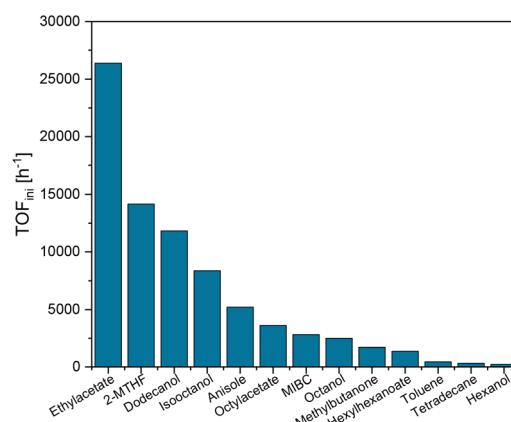


Fig. 4 Initial TOFs for the biphasic CO₂ hydrogenation using aqueous Arg and *cis*-[Ru(C₁₂-dppm)₂Cl₂] as catalyst in various solvents. Conditions: 30 bar CO₂, 60 bar H₂, 2.58 mmol Arg in water (3 mL), 1 μ mol *cis*-[Ru(C₁₂-dppm)₂Cl₂] in the indicated solvent (2 mL), 70 °C. TOF_{ini} calculated from initial linear pressure drop; see the ESI, Fig. S5.†



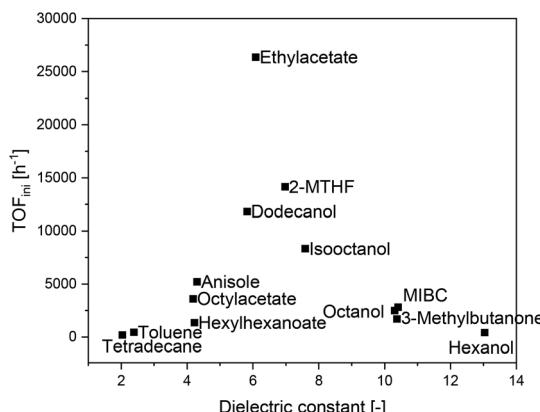


Fig. 5 Correlation between the dielectric constant and TOF_{ini} from the experiments described in Fig. 4.



Scheme 2 Pictures of the Arg, water and catalyst solution in the autoclave before the reaction (left), after saturation with CO_2 (middle) and after the reaction with H_2 (right).

ization, large amounts of undissolved Arg were dispersed in water below the yellow catalyst phase. After saturation with CO_2 , mixture became turbid but most of the solid particles were dissolved. At the end of the hydrogenation when the pressure reached a constant value after 1.5 h, both phases had become clear and arginine was completely dissolved. Also, a phase expansion of the aqueous solution was visible. Analysis of the aqueous phase revealed a FA/base ratio of 0.77. The TOF_{ini} was estimated to be $15\ 345\ \text{h}^{-1}$ under these conditions. Since the volume and density do not remain constant, the wt% was more reliable in terms of concentration than the molar amounts. The obtained product solution consisted of 9.4 wt% FA and 46.4 wt% Arg.

In order to mimic the CO_2 capture step, a CO_2 saturated solution with Arg, corresponding to a 6 M concentration, was prepared on a liter scale. This concentration was found to provide an optimum balance between the final concentration and the formation rate TOF_{ini} (Table S5 and Fig. S15†). The capture capacity of Arg was determined as 0.94 mol CO_2 per mol Arg under 2 bar.³⁸ The catalytic hydrogenation was performed in a largely automated semi-batch process with a 100 mL autoclave⁵¹ using 20 mL of catalyst phase containing only 10 μmol of the Ru-complex and 50 g of the separately prepared CO_2 saturated aqueous solution. The autoclave was pressurized with H_2 at 90 bar and heated to 70 °C while stirring using a magnetic stir bar. The set-up allowed continuous feeding of H_2 gas at isobaric conditions while monitoring the

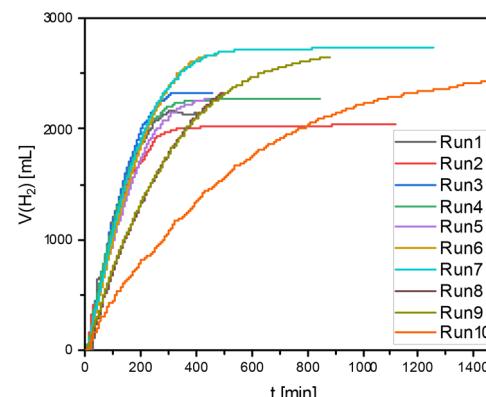


Fig. 6 Delivered H_2 during the recycling experiments in the semi-continuous setup. Conditions: 90 bar H_2 , 70 °C, 10 μmol *cis*-[Ru(C_{12} -dppm)₂Cl₂] in *n*-dodecanol (20 mL); ca. 50 g of CO_2 -saturated 6 M Arg-solution added and removed in each run.

H_2 flow (see Fig. 6). After about 5 h, consumption of H_2 stopped, the stirring was discontinued to allow phase separation, and the aqueous phase was removed for analysis under pressure at 70 °C. The autoclave was charged with a new batch of CO_2 -saturated arginine solution and the reaction repeated without any catalyst make-up in between. Over ten runs, the final formic acid-to-amino acid ratio reached the 1:1 limit ($0.91 \pm 0.03 : 1$ on average, 9.0–10.8 wt% FA) indicating practically quantitative conversion of the captured CO_2 to FA by catalytic hydrogenation. The data correspond to a total TON of 102 282 corresponding to 46.6 kg HCOOH per g of ruthenium.

The rate of formation corresponded to a TOF_{ini} of $3357\ \text{h}^{-1}$ in the first run, which was significantly lower than on the 10 mL scale reflecting most likely mass transfer limitations due to less effective mixing. The system proved very robust, however, and the rates only declined after eight repetitive batches. In run ten, almost 50% of the initially observed rate was still maintained and the thermodynamic concentration limit was reached upon prolonged reaction time. The contamination of the aqueous product phase with ruthenium was determined by ICP-MS measurements (Table S7†). While a notable leaching of Ru into the product phase was noted after the first reaction (0.50 ppm), only trace amounts were detected in all further runs (0.01–0.08 ppm). Over the entire process, 3.3% of the originally charged Ru was leached into the product phase with a total loss of only 1% in runs 2–10 demonstrating proficient catalyst retention. The low leaching values cannot explain the reduced catalytic activity indicating that the chemical deactivation of the catalyst also plays a role under small-scale laboratory conditions. After the last run, 15.5 g of catalyst phase was recovered corresponding to 95% of the initially charged *n*-dodecanol proving also the excellent phase separation and negligible cross-solubility.

While basic amino acids such as Arg have the potential to act as scrubbing agents for CO_2 capture, other amino acids are also very important as nutrients in animal feed. In particular,



methionine is an important amino acid for livestock feed for poultry in which formic acid is a prominent preservative.^{26,62} Therefore, we also examined the application of the biphasic system for CO₂ hydrogenation with Met in the aqueous phase. The water phase was saturated with Met at its maximum solubility (0.33 M) and *n*-dodecanol containing 0.5 mM of *cis*-[Ru(C₁₂-dppm)₂Cl₂] was used as the catalyst phase. The reaction was conducted under standard conditions (30 bar CO₂, 60 bar H₂, 70 °C) for 16 h led to a formic acid-to-amino acid ratio of 0.6:1, corresponding to a TON of 595. This result demonstrates that the catalytic system is also compatible with neutral amino acids, opening a broad spectrum of possible compositions for applications of these solutions.

Amino acids are often applied in combinations for feed supplements. The formation of formic acid was therefore investigated using an aqueous solution containing a mixture of the three essential amino acids, namely arginine, histidine and methionine at their saturation limits (Arg 0.86 M, His 0.25 M, Met 0.33 M).⁶³ Under standard conditions (3 mL H₂O, 30 bar CO₂, 60 bar H₂, 70 °C, 0.5 mM *cis*-[Ru(C₁₂-dppm)₂Cl₂] in 2 mL dodecanol), a concentration of 0.86 M FA was achieved after 2.5 h.

Conclusion

In summary, we have demonstrated that the hydrogenation of CO₂ to formic acid can be performed effectively in biphasic systems comprising aqueous solutions of amino acids as the product phase together with an organic phase containing an organometallic catalyst. Besides the choice of catalyst, a major tuning factor of these biphasic systems lies in the respective organic phase. The bio-based solvent *n*-dodecanol (lauryl alcohol) was identified as a preferred solvent due to a combination of high catalyst activity, negligible water miscibility, and its benign character. The use of CO₂-saturated Arg solutions as would be obtained from CO₂ capture was successfully demonstrated to integrate the upstream carbon dioxide source with the downstream product phase directly. Excellent retention of the catalyst was achieved leading to metal contamination in the lower ppm range in the aqueous phase. In repetitive batch experiments, a total turnover number of more than 100 000 was achieved with initial turnover frequencies in the 10³ h⁻¹ range. Solutions of up to 0.94 mol FA per mol amino acid were obtained using the basic amino acid Arg corresponding to 108 g of newly formed HCOOH per kg solution.

Based on these results, aqueous solutions of amino acids and formic acid resulting from the hydrogenation of CO₂ can be envisaged for use directly or by further upgrading for high value applications without the need for energy intensive isolation of the HCOOH product from the reaction mixture. This would allow the replacement of the present fossil-based formic acid with a product based on biogenic CO₂ for certain applications. The possible connection of the energetic use of biomass in biogas units and the upgrading of the inevitable

CO₂ byproduct to produce solutions that are used in farming directly may open novel opportunities for the concept of bio-energy conversion and carbon capture and utilization (BECCU) for local value chains contributing to a reduced CO₂ footprint in the agricultural sector.

Experimental

Repetitive hydrogenation in the semi-continuous reaction setup

The flow-scheme of the semi-continuous reaction setup⁵¹ controlled by LabView™ software (National Instruments) is shown in Fig. 7 and described in more detail in the ESI.† The catalyst *cis*-[Ru(C₁₂-dppm)₂Cl₂] was weighed into a Schlenk-tube (23.0 mg) and dissolved in *n*-dodecanol (10 mL). This solution was then added into the 100 mL autoclave, the Schlenk-tube was rinsed twice with *n*-dodecanol (5 mL) and the rinsing solutions were also added to the autoclave. The autoclave was sealed and connected to the plant. The substrate solution was prepared by weighing arginine (157.0 g) for multiple runs into the 900 mL high pressure reservoir, which was then evacuated three times and flushed with argon. Afterwards, water (150 mL) was added in argon counterflow. The high-pressure reservoir was positioned on a precision scale balance, connected to the plant, pressurized with CO₂ to 30 bar and stirred at rt until the pressure remained constant.

Capillaries and a HPLC-pump were flushed with the aqueous arginine solution up to the substrate inlet of the autoclave. Then, the autoclave was filled with 50 g of the substrate solution and pressurized with hydrogen (90 bar) over a bypass. The autoclave was then heated to 70 °C and the attained pressure (105 bar) was set as the fixed pressure in LabView™. The reaction was started by turning on the stirrer while the pressure was kept constant by dosing H₂ via a mass-flow con-

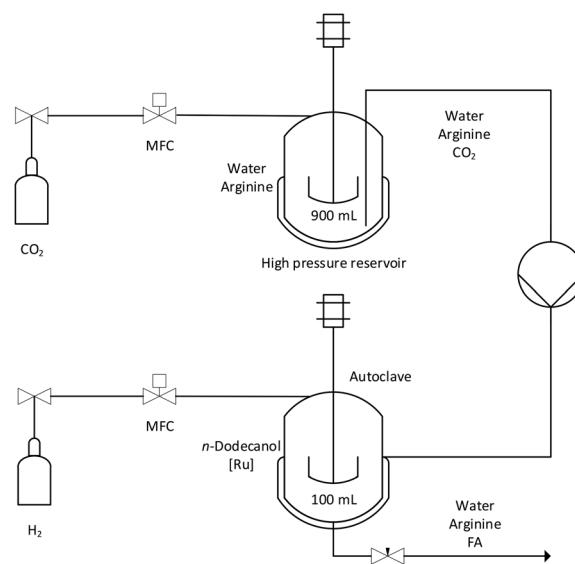


Fig. 7 Flow-scheme of the semi-continuous plant.



troller (MFC). Data recording was started simultaneously. The reaction was stopped as soon as no further flow of H₂ was detected. The stirrer was stopped and the phases were allowed to settle and separate for 2 min. The product solution was removed through the bottom fine dosing valve at the reaction temperature. Through the integrated window, the phase boundary was observed to avoid losses of the catalyst phase. The removed product phase was weighed and about 0.1 g of this solution was mixed with maleic acid (10 mg) as the internal standard and analyzed by ¹H NMR spectroscopy. For the subsequent run, a fresh substrate solution was added into the autoclave with the HPLC pump so that the previous reaction pressure was obtained again. Afterwards the stirrer, the H₂ delivery *via* the MFC, and the data recording were started again. This procedure was repeated nine times. After the last run, the catalyst phase was also removed and analyzed. The ruthenium contents of the aqueous phases were quantified by ICP-MS.

Author contributions

G. Franciò and W. Leitner conceived the idea and designed the experiments; N. Guntermann conducted the experiments under the supervision of G. Franciò and W. Leitner; N. Guntermann prepared the first draft; W. Leitner and G. Franciò revised the manuscript; all the authors endorsed the final version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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References

- 1 W. Leitner, *Angew. Chem., Int. Ed. Engl.*, 1995, **34**, 2207.
- 2 P. G. Jessop, T. Ikariya and R. Noyori, *Chem. Rev.*, 1995, **95**, 259.
- 3 C. Federsel, R. Jackstell and M. Beller, *Angew. Chem., Int. Ed.*, 2010, **49**, 6254.
- 4 P. G. Jessop, F. Joó and C.-C. Tai, *Coord. Chem. Rev.*, 2004, **248**, 2425.
- 5 W.-H. Wang, Y. Himeda, J. T. Muckerman, G. F. Manbeck and E. Fujita, *Chem. Rev.*, 2015, **115**, 12936.
- 6 J. Artz, T. E. Müller, K. Thenert, J. Kleinekorte, R. Meys, A. Sternberg, A. Bardow and W. Leitner, *Chem. Rev.*, 2018, **118**, 434.
- 7 J. Klankermayer, S. Wesselbaum, K. Beydoun and W. Leitner, *Angew. Chem., Int. Ed.*, 2016, **55**, 7296.
- 8 N. von der Assen, P. Voll, M. Peters and A. Bardow, *Chem. Soc. Rev.*, 2014, **43**, 7982.
- 9 A. Kätelhön, R. Meys, S. Deutz, S. Suh and A. Bardow, *Proc. Natl. Acad. Sci. U. S. A.*, 2019, **116**, 11187.
- 10 S. Deutz and A. Bardow, *Nat. Energy*, 2021, **6**, 203.
- 11 C. M. Jens, M. Scott, B. Liebergesell, C. G. Westhues, P. Schäfer, G. Franciò, K. Leonhard, W. Leitner and A. Bardow, *Adv. Synth. Catal.*, 2019, **361**, 307.
- 12 K. A. Mumford, Y. Wu, K. H. Smith and G. W. Stevens, *Front. Chem. Sci. Eng.*, 2015, **9**, 125.
- 13 R. S. Haszeldine, *Science*, 2009, **325**, 1647.
- 14 J. van Holst, S. R. A. Kersten and K. J. A. Hogendoorn, *J. Chem. Eng. Data*, 2008, **53**, 1286.
- 15 Y. Bian, S. Shen, Y. Zhao and Y. Yang, *J. Chem. Eng. Data*, 2016, **61**, 2391.
- 16 X. Wang, N. G. Akhmedov, D. Hopkinson, J. Hoffman, Y. Duan, A. Egbebi, K. Resnik and B. Li, *Appl. Energy*, 2016, **161**, 41.
- 17 A. Syalsabila, A. S. Maulud, H. Suleman and N. A. H. M. Nordin, *Int. J. Chem. Eng.*, 2019, **2019**, 1.
- 18 Z. Zhang, Y. Li, W. Zhang, J. Wang, M. R. Soltanian and A. G. Olabi, *Renewable Sustainable Energy Rev.*, 2018, **98**, 179.
- 19 V. S. Sefidi and P. Luis, *Ind. Eng. Chem. Res.*, 2019, **58**, 20181.
- 20 A. E. Reichl, R. Schneider, A. Ohligschläger, T. Rogalinski and S. Hauke, *Energy Procedia*, 2014, **63**, 6379.
- 21 J. Hietala, A. Vuori, P. Johnsson, I. Pollari, W. Reutemann and H. Kieczka, Formic Acid, in *Ullmann's Encyclopedia of Industrial Chemistry*, Wiley-VCH, Weinheim, 2016.
- 22 N. J. Claassens, I. Sánchez-Andrea, D. Z. Sousa and A. Bar-Even, *Curr. Opin. Biotechnol.*, 2018, **50**, 195.
- 23 J. La, G. de Cruz, F. Machens, K. Messerschmidt and A. Bar-Even, *ACS Synth. Biol.*, 2019, **8**, 911.
- 24 S. Zobel, J. Kuepper, B. Ebert, N. Wierckx and L. M. Blank, *Eng. Life Sci.*, 2017, **17**, 47.
- 25 L. M. Blank, T. Narancic, J. Mampel, T. Tiso and K. O'Connor, *Curr. Opin. Biotechnol.*, 2020, **62**, 212.
- 26 S. R. Fernandez, S. Aoyagi, Y. Han, C. M. Parsons and D. H. Baker, *Poul. Sci.*, 1994, **73**, 1887.
- 27 R. E. Warnick and J. O. Anderson, *Poul. Sci.*, 1968, **47**, 281.
- 28 Y. Han, H. Suzuki, C. M. Parsons and D. H. Baker, *Poul. Sci.*, 1992, **71**, 1168.
- 29 Supplementary feed for chickens, ducks, geese, quails and other poultry, "Röhnfried Hennengold", zu finden unter <https://stengel-fussring.com/produkte-nach-marken/roehnfried/roehnfried-hennengold-mineralfutter-mit-den-amino-saeuren-methionin-l-arginin-fluessig-1/>, 2022.
- 30 G. Wu, F. W. Bazer, T. A. Davis, L. A. Jaeger, G. A. Johnson, S. W. Kim, D. A. Knabe, C. J. Meininger, T. E. Spencer and Y.-L. Yin, *Livest. Sci.*, 2007, **112**, 8.



- 31 D. Che, S. Adams, B. Zhao, G. Qin and H. Jiang, *Curr. Protein Pept. Sci.*, 2019, **20**, 736.
- 32 B. Tan, Y. Yin, Z. Liu, X. Li, H. Xu, X. Kong, R. Huang, W. Tang, I. Shizato, S. B. Smith, *et al.*, *Amino Acids*, 2009, **37**, 169.
- 33 National Research Council, Division on Earth and Life Studies, Board on Agriculture and Natural Resources, Committee on Nutrient Requirements of Swine, *Nutrient requirements of swine*, National Academies Press, Washington, D.C., 11th revised edn, 2012.
- 34 PIC North America, *Nutrient Specifications Manual 2016*, Hendersonville, TN 37075, 2016.
- 35 R. E. Muck, E. M. G. Nadeau, T. A. McAllister, F. E. Contreras-Govea, M. C. Santos and L. Kung, *J. Dairy Sci.*, 2018, **101**, 3980.
- 36 S. C. Ricke, D. K. Dittoe and K. E. Richardson, *Front. Vet. Sci.*, 2020, **7**, 563.
- 37 V. Bampidis, G. Azimonti, M. d. L. Bastos, H. Christensen, B. Dusemund, M. Kouba, M. Kos Durjava, M. López-Alonso, S. López Puente, F. Marcon, *et al.*, *EFSA J.*, 2019, **17**, e05645.
- 38 D. Wei, H. Junge and M. Beller, *Chem. Sci.*, 2021, **12**, 6020.
- 39 D. Wei, R. Sang, P. Sponholz, H. Junge and M. Beller, *Nat. Energy*, 2022, **7**, 438.
- 40 J. J. Anderson, D. J. Drury, J. E. Hamlin and A. G. Kent, WO8602066, BP Chemicals Limited, 1986.
- 41 R. G. Beevor, D. J. Gulliver, M. Kitson and R. M. Sorrel, EP0357243, BP Chemicals Limited, 1989.
- 42 M. J. Green, A. R. Lucy, M. Kitson and S. J. Smith, EP0329337, BP Chemicals Limited, 1989.
- 43 T. Schaub, O. Bey, A. Meier, D. M. Fries and R. Hugo, WO2013050367, BASF, 2011.
- 44 T. Schaub and R. A. Paciello, *Angew. Chem., Int. Ed.*, 2011, **50**, 7278.
- 45 A. Behr, P. Ebbinghaus and F. Naendrup, *Chem. Eng. Technol.*, 2004, **27**, 495.
- 46 C. A. Ohlin and G. Laurenczy, *High Pressure Res.*, 2003, **23**, 239.
- 47 Z. Zhang, Y. Xie, W. Li, S. Hu, J. Song, T. Jiang and B. Han, *Angew. Chem., Int. Ed.*, 2008, **47**, 1127.
- 48 J. Kothandaraman, A. Goeppert, M. Czaun, G. A. Olah and G. K. Surya Prakash, *Green Chem.*, 2016, **18**, 5831.
- 49 S. Kar, R. Sen, A. Goeppert and G. K. S. Prakash, *J. Am. Chem. Soc.*, 2018, **140**, 1580.
- 50 M. Scott, B. Blas-Molinos, C. Westhues, G. Franciò and W. Leitner, *ChemSusChem*, 2017, **10**, 1085.
- 51 W. Leitner, G. Franciò, M. Scott, C. Westhues, J. Langanke, M. Lansing, C. Hussong and E. Erdkamp, *Chem. Ing. Tech.*, 2018, **90**, 1504.
- 52 S. Wesselbaum, V. Moha, M. Meuresch, S. Brosinski, K. M. Thenert, J. Kothe, T. Vom Stein, U. Englert, M. Hölscher, J. Klankermayer, *et al.*, *Chem. Sci.*, 2015, **6**, 693.
- 53 M. Scott, C. G. Westhues, T. Kaiser, J. C. Baums, A. Jupke, G. Franciò and W. Leitner, *Green Chem.*, 2019, **21**, 6307.
- 54 M. Scott, Dissertation, RWTH Aachen, 2019.
- 55 W. L. F. Gassner, *J. Chem. Soc., Chem. Commun.*, 1993, 1465.
- 56 R. Tanaka, M. Yamashita and K. Nozaki, *J. Am. Chem. Soc.*, 2009, **131**, 14168.
- 57 Y. Himeda, N. Onozawa-Komatsuzaki, H. Sugihara and K. Kasuga, *J. Am. Chem. Soc.*, 2005, **127**, 13118.
- 58 N. Guntermann, H. G. Mengers, G. Franciò, L. M. Blank and W. Leitner, *Green Chem.*, 2021, **23**, 9860.
- 59 P. Zhang, S.-F. Ni and L. Dang, *Chem. – Asian J.*, 2016, **11**, 2528.
- 60 S.-F. Ni and L. Dang, *Phys. Chem. Chem. Phys.*, 2016, **18**, 4860.
- 61 P. J. Dyson and P. G. Jessop, *Catal. Sci. Technol.*, 2016, **6**, 3302.
- 62 P. T. H. van To, K. Masagounder and M. E. Loewen, *Comp. Biochem. Physiol., Part A: Mol. Integr. Physiol.*, 2021, **255**, 110908.
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