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Decarboxylative oxygenation of carboxylic acids with O₂ via a non-heme manganese catalyst†

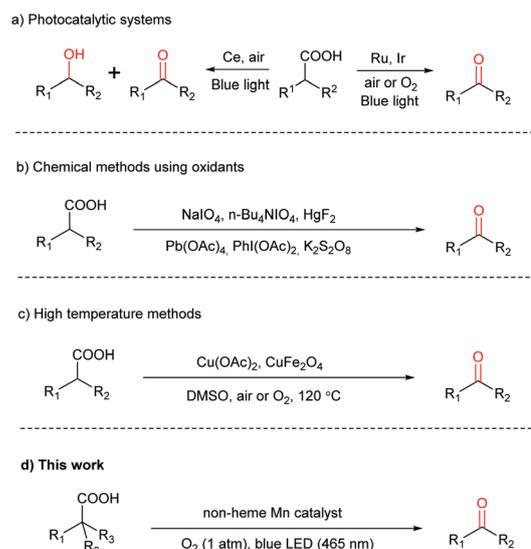
Renpeng Guan, Elliot L. Bennett, Zhiliang Huang * and Jianliang Xiao *

Reported here is a new protocol for the decarboxylative oxygenation of carboxylic acids using a non-heme manganese catalyst under blue light irradiation with O₂ as the sole oxidant. Featuring mild reaction conditions, the protocol allows readily available carboxylic acids to be converted into a wide variety of valuable aldehydes, ketones and amides. Mechanistic studies indicate that the decarboxylation and oxygenation involves the formation of active Mn-oxygen species.

Introduction

Carboxylic acids are one of the most important feedstocks in organic, polymer and material synthesis.^{1,2} They are widely available in nature and industry, inexpensive, stable, and non-toxic in general.^{3–5} Moreover, many carboxylic acids, such as amino acids, fatty acids and sugar acids, can be derived directly from natural resources. Therefore, a great number of transformations of carboxylic acids have been reported in the past few decades.^{6–9} In particular, the construction of useful carbon–carbon and carbon–heteroatom bonds through decarboxylation has received significant attention.^{10–24} However, direct decarboxylative oxygenation of carboxylic acids to the corresponding carbonyl compounds, *i.e.* aldehydes and ketones, with high efficiency and wide substrate scope under mild conditions remains limited. Aldehydes and ketones are the fundamental building blocks of organic chemistry,^{25–30} and they can be synthesized *via* a variety of methods, such as carbonylation of alkenes and oxidation of hydrocarbons.^{31,32} However, these methods often rely on harsh conditions such as high temperature, high pressure, and toxic reagents. For instance, whilst hydroformylation is widely used in industry to prepare aldehydes, the cobalt catalyst necessitates >200 atm syngas and >100 °C temperature and although much milder conditions are employed for Rh-based catalysts, the metal is expensive and toxic.³³ Therefore, exploring rapid synthesis of aldehydes and ketones under mild conditions with cheap substrates and catalysts is a worthy endeavor. Given the easy availability of various carboxylic acids, they are well suited for this endeavor.

Decarboxylative oxygenation of carboxylic acids is a challenging reaction, requiring a catalyst not only capable of promoting decarboxylation but also oxidation. So far, there have been only three reports, demonstrating the reaction *via* photoredox catalysis using molecular oxygen (O₂) under mild conditions. Two of these require precious Ru or Ir photocatalysts,^{34,35} while the third produces a mixture of alcohols and ketones/aldehydes as the products (Scheme 1a).³⁶ Other methods have been reported; however, they make use of stoichiometric strong oxidants, *e.g.* NaIO₄,³⁷ *n*-Bu₄NIO₄,³⁸ HgF₂,³⁹ Pb(OAc)₄,⁴⁰ PhI(OAc)₂⁴¹ and K₂S₂O₈⁴² (Scheme 1b), or rely on a high reaction temperature (Scheme 1c).^{25,43} Herein, we report a highly selective decarboxylative oxygenation of carboxylic acids to aldehydes or ketones by O₂ (1 atm), promoted by a non-heme



Scheme 1 Decarboxylative oxygenation of carboxylic acids.

Department of Chemistry, University of Liverpool, Liverpool L69 7ZD, UK.

E-mail: Z.Huang49@liverpool.ac.uk, J.Xiao@liverpool.ac.uk

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Mn(II) catalyst and visible light under mild conditions (Scheme 1d).

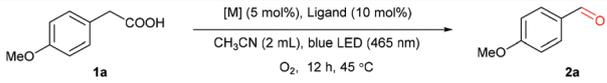
The use of benign and inexpensive O₂ for selective oxidation with first-row, biologically relevant metal complexes as catalysts remains a significant challenge for the synthetic chemist.^{44–46} Over the past a few decades, some biomimetic Fe and Mn complexes bearing heme and non-heme polydentate ligands have been developed to catalyze the oxidation of hydrocarbons, alcohols as well as carboxylic acids with O₂.^{37,44,47–51} However, significant drawbacks remain in most cases, *e.g.* the need for stoichiometric co-reductants and unsatisfactory selectivity due to poorly controlled radical-type pathways. To date, only a few biomimetic metal complexes are known that can catalyze the aerobic oxidation in a controlled manner without using a co-reductant.^{52–57} Following on from our recent study of aerobic cleavage of alkenes with a non-heme Mn(II) complex,⁵⁸ we investigated the decarboxylative oxygenation of carboxylic acids to aldehydes/ketones by O₂ with this and related complexes. Our findings are detailed below.

Results and discussion

We commenced our study by examining the decarboxylative oxygenation of 4-methoxybenzeneacetic acid (**1a**) with O₂ (1 atm) (Table 1). Based on our previous work,⁵⁸ the photocatalytic system comprised of Mn(OTf)₂ and **L4** was initially tested, as it was proven to be effective for O₂ activation in the presence of blue light. To our delight, the desired oxygenation product **2a** could be obtained in 90% yield (Table 1, entry 6). [Mn(dtbbpy)₂(OTf)₂] formed *in situ* is likely to be the catalytic species, as the isolated [Mn(dtbbpy)₂(OTf)₂] complex displayed a similar activity (Table 1, entry 1). Unsurprisingly, the ligand-free Mn(OTf)₂, which is less likely to produce activated oxygen species from O₂ ($E_{\text{Mn(III)/Mn(II)}}^0 = 1.56 \text{ V/SHE}$, $E_{\text{O}_2/\text{H}_2\text{O}}^0 = 0.82 \text{ V/SHE}$, neutral conditions), showed no catalytic activity for the target reaction (Table 1, entry 2). The effect of ligand was further explored by testing a range of substituted bipyridines (**L1–L8**), revealing **L4** to give the best yield (Table 1, entries 3–10). Meanwhile, evaluation of the solvents showed that the aerobic oxygenation reaction presented the highest reactivity in acetonitrile (ESI, Table S1†). The combinations of **L4** with other metal salts, such as Cu(OTf)₂, Fe(OTf)₂, and CoCl₂, were ineffective, affording much lower yields of the target product (Table 1, entries 11–13). In addition, MnCl₂ is less effective than Mn(OTf)₂, as can be seen in Table 1, entry 14. As is clear, both O₂ and blue light play important roles in this transformation; in their absence, no target product was observed (Table 1, entry 15, and Fig. S3 in ESI†). The screening established the following optimized conditions: Mn(OTf)₂ (5 mol%), **L4** (10 mol%) as ligand, O₂ as oxidant in CH₃CN at 45 °C with blue light irradiation.

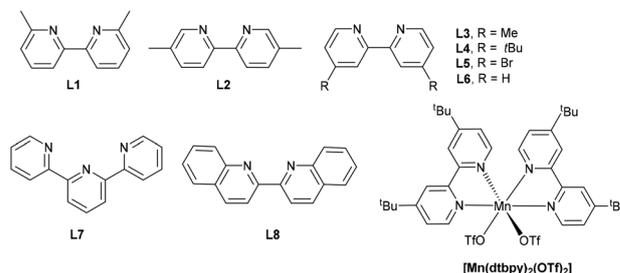
Under the optimized reaction conditions, a series of benzylic carboxylic acids were first examined to test the generality of this decarboxylative oxygenation protocol. As shown in Scheme 2, phenyl acetic acids bearing electron-withdrawing or

Table 1 Optimization of reaction conditions^{a,b}



Entry	[M]	Ligand	Yield (%)
1	[Mn(dtbbpy) ₂ (OTf) ₂]	—	87
2	Mn(OTf) ₂	—	0
3	Mn(OTf) ₂	L1	33
4	Mn(OTf) ₂	L2	46
5	Mn(OTf) ₂	L3	61
6	Mn(OTf) ₂	L4	90
7	Mn(OTf) ₂	L5	41
8	Mn(OTf) ₂	L6	64
9	Mn(OTf) ₂	L7	14
10	Mn(OTf) ₂	L8	41
11	Cu(OTf) ₂	L4	4
12	Fe(OTf) ₂	L4	16
13	CoCl ₂	L4	42
14	MnCl ₂	L4	53
15 ^c	[Mn(dtbbpy) ₂ (OTf) ₂]	—	0
16 ^d	Mn(OTf) ₂	L4	62

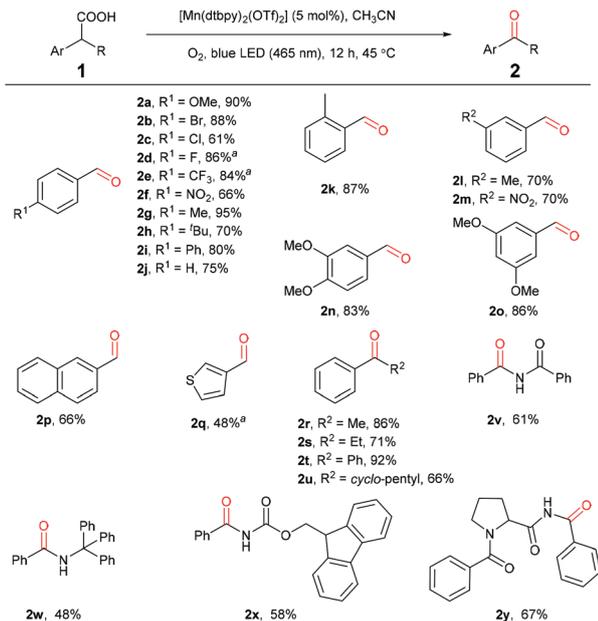
^a Reaction conditions: **1a** (0.5 mmol), [M] (5 mol%), ligand (10 mol%), CH₃CN (2 mL), blue light (465 nm), 45 °C, O₂ (1 atm), 12 h. ^b NMR yields, determined using mesitylene (20 μL) as internal standard. ^c N₂ (1 atm). ^d Air instead of O₂.



electron-donating groups on the phenyl ring are all suitable substrates (**2a–2o**), affording the desirable aldehyde products in good to excellent yields. During our investigation, we noticed that phenyl acetic acids bearing electronic-withdraw groups were usually less reactive. For example, *para*-F-substituted acid **1d** and *para*-CF₃-substituted acid **1e** showed low reactivity for the oxidation under above standard conditions, giving the corresponding aldehydes in 35% and 17% yields, respectively. However, the oxidation was improved by introducing a catalytic amount of sodium acetate as an additive (Scheme 2. See ESI for the detailed reaction optimization, Table S2†). 2-Naphthyl acetic acid and S-heterocycle acetic acid can be oxidized selectively to afford **2p** and **2q** in moderate yields, respectively. Secondary benzylic carboxylic acids are also efficiently converted to the corresponding ketone products in good to excellent yields under the standard conditions (**2r–2u**). It is worth noting that a range of amino acids including a dipeptide can be oxidized to the corresponding amide products in a highly selective manner, showing potential applications in bio-conjugate chemistry (**2v–2y**).

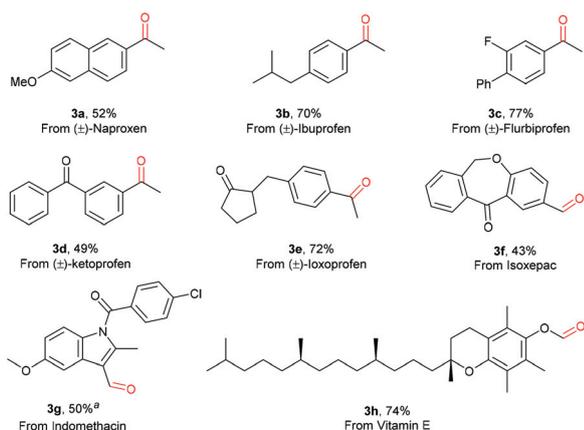
Notably, a variety of drug molecules with functionalized aryl acetic acid scaffolds were also selectively oxidized by O₂





Scheme 2 Oxidation of mono- and di-substituted benzylic carboxylic acids. Reaction conditions: **1** (0.5 mmol), $[\text{Mn}(\text{dtbbpy})_2(\text{OTf})_2]$ (5 mol%) (*in situ* prepared), CH_3CN (2 mL), blue light (465 nm), 45 °C, O_2 (1 atm), 12 h. Isolated yields are given. ^a CH_3COONa (30 mol%) was added.

under the conditions developed. As can be seen in Scheme 3, the oxidation of naproxen, ibuprofen, flurbiprofen, ketoprofen, ioxoprofen, isoxepac and indomethacin all proceeded successfully, affording the corresponding aldehydes or ketones in good yields (**3a–3g**). Moreover, this photo-Mn protocol showed its ability in late-stage decarboxylative oxygenation of vitamin E, affording the aldehyde product **3h** in 74% yield. The relatively active benzylic C–H bonds, tertiary C–H bonds, and indole ring, which are prone to oxidation, remained intact, showing the high chemoselectivity of this oxidation protocol.

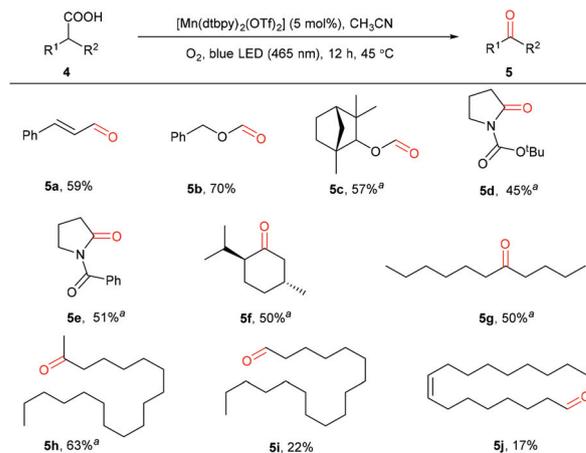


Scheme 3 Oxidation of pharmaceutical molecules. Reaction conditions: Carboxylic acid (0.5 mmol), $[\text{Mn}(\text{dtbbpy})_2(\text{OTf})_2]$ (5 mol%), CH_3CN (2 mL), blue light (465 nm), 45 °C, O_2 (1 atm), 12 h. Isolated yields are given. ^a CH_3COONa (30 mol%) was added.

The success in late-stage decarboxylative oxygenation of drugs or natural products demonstrates the practical applicability of this oxidation protocol in accessing new bioactive molecules.

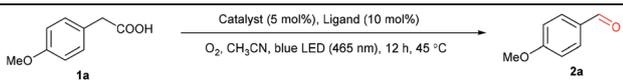
The more challenging enoic acids and aliphatic carboxylic acids were also tested.^{59,60} As can be seen in Scheme 4, the C=C double bonds in enoic acids, usually sensitive to oxidation conditions, remained intact during the oxidation (**5a**). Notably, long-chain and cyclic aliphatic acids with different functionalities, including ether, carbamate, and amide, are suitable to give the corresponding carbonyl products with blue or UV light irradiation (**5b–5e**), highlighting the high chemoselectivity and hence great application potential of this method. Secondary aliphatic acids without other functional groups were also oxidatively decarboxylated to furnish the target carbonyls in 50%–63% yields (**5f–5h**) under the irradiation of UV-light. Notably, the primary stearic acid and oleic acid could be oxidized to the corresponding aldehydes; however, low yields, as well as low conversions were observed (**5i** and **5j**). In comparison with Macmillan's work in 2019,³⁵ this protocol shows a wider substrate scope. Not only can the cyclic aliphatic acids be oxidized selectively, but also long-chain fatty acids, saturated and unsaturated, could be converted, which are usually challenging substrates.⁶¹

Although the detailed mechanism of the decarboxylative oxygenation reaction is unclear, a series of experiments were performed to shed light on possible reaction pathways. As reported by Boger's group,⁶² singlet oxygen (¹O₂) can efficiently transfer pyrrole-2-carboxylic acids to the corresponding ketones in the presence of rose bengal. Therefore, it is important to determine whether ¹O₂ plays a role in the photo-Mn enabled oxidative decarboxylation. As shown in Table 2, a range of well-known photosensitizers (Table 2, entries 1–4), such as Eosin Y disodium salt, $\text{Ru}(\text{bpy})_3$, $\text{Ir}(\text{dFppy})_3$ and rose bengal,^{58,63} which can generate ¹O₂ under blue light, were used individually as replacement catalysts for the decarboxyla-



Scheme 4 Oxidation of aliphatic acids. Reaction conditions: **4** (0.5 mmol), $[\text{Mn}(\text{dtbbpy})_2(\text{OTf})_2]$ (5 mol%), CH_3CN (2 mL), blue LED light (465 nm), 45 °C, O_2 atmosphere (1 atm), 12 h. Isolated yields are given. ^a DCE/ CH_3CN (1 : 1, 2 mL), ultraviolet light (365 nm).



Table 2 Decarboxylative oxidation of **1a** in the presence of well-known photosensitizers or singlet oxygen trap^{a,b}


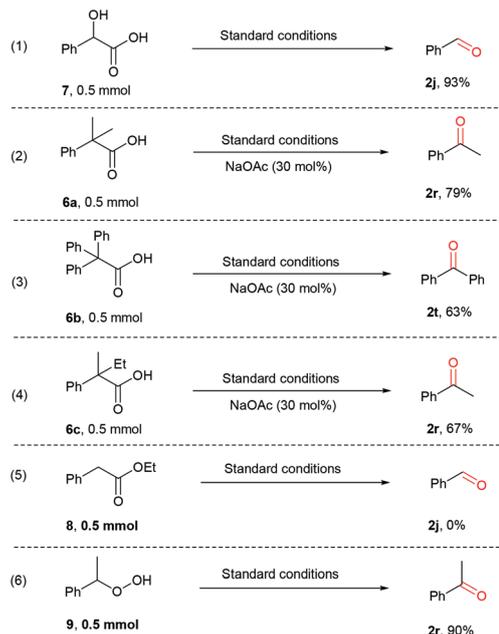
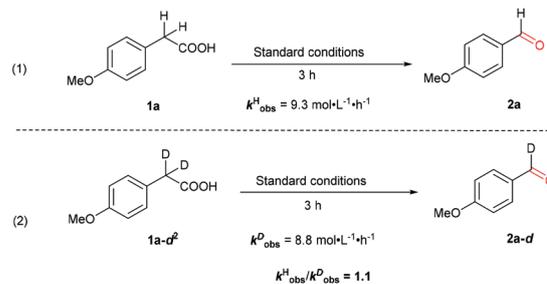
Entry	Catalyst	Yield (%)
1	Eosin Y disodium salt	37
2	[Ru(bpy) ₃ ·6H ₂ O]	19
3	[Ir(dFppy) ₃]	0 ^c
4	Rose bengal	4
5 ^d	[Mn(dtbpy) ₂ (OTf) ₂]	70

^a Reaction conditions: Acid (0.5 mmol), catalyst (5 mol%), CH₃CN (2 mL), blue light (465 nm), 45 °C, O₂ (1 atm), 12 h. ^b NMR yields, determined using mesitylene as internal standard. ^c Conversion was observed under the conditions reported in ref. 35 (see Scheme S2, ESI†). ^d DPA (0.1 mmol) was added.

tive oxygenation of **1a** under standard reaction conditions. However, these photosensitizers showed a significantly lower catalytic effect. Furthermore, when an ¹O₂ trap, 9,10-diphenylanthracene (DPA) which is known to react rapidly with ¹O₂ to give an endoperoxide product ($k \approx 1.3 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$),⁶⁴ was added to the oxidative decarboxylation of **1a** under the standard conditions (Table 2, entry 5), **2a** was obtained in 70% yield, and no endoperoxide was detected. These observations appear to rule out the involvement of ¹O₂ in the reaction pathway.

Previous studies on decarboxylative oxygenation of phenylacetic acids by Mn-based enzymes indicate that the hydroxylation of C–H bond in the benzylic position is the initiation step of this oxidation reaction, which affords 2-hydroxy-2-phenylacetic acid as the key intermediate for the further oxidative decarboxylation process.^{65–67} Knowing this, several control experiments were performed to determine whether a similar hydroxylation process occurs as the key step. Firstly, the potential intermediate 2-hydroxy-2-phenylacetic acid (**7**) was subjected to the standard reaction conditions, affording **2j** in 93% yield (Scheme 5, eqn (1)). This result indicates that **7** could be involved in the oxidative decarboxylation reaction under the current conditions. However, when the corresponding benzylic C–H bonds were substituted by two alkyl or phenyl groups, the desired decarboxylative oxygenation proceeded smoothly as well (Scheme 5, eqn (2)–(4)). In all cases, the ketone products were obtained, apparently *via* a C–C bond cleavage process due to the formation of the strong C=O bond. This indicates the involvement of a radical fragmentation pathway. Additionally, no reactions happened when ethyl 2-phenylacetate (**8**) was employed (Scheme 5, eqn (5)). Meanwhile, **2r** was afforded when (1-hydroperoxyethyl)benzene (**9**) was employed (Scheme 5, eqn (6)). These observations indicate that the acid moiety is involved and peroxide intermediates may be formed in the decarboxylative oxygenation reaction.

Further insight was obtained by following the kinetic isotope effect (KIE) (Scheme 6). Parallel reactions using acids **1a** and **1a-d²** afforded the initial rates $k_{\text{obs}}^{\text{H}} = 9.3$ and $k_{\text{obs}}^{\text{D}} =$

**Scheme 5** Control experiments to shed light on the mechanism. Conditions: Substrate (0.5 mmol), [Mn(dtbpy)₂(OTf)₂] (5 mol%), CH₃CN (2 mL), blue light (465 nm), 45 °C, O₂ (1 atm), 12 h, CH₃COONa (30 mol%) where applicable; NMR yields, determined using mesitylene as internal standard.**Scheme 6** Kinetic isotope effect studied under the optimized conditions.

8.8 mol L⁻¹ h⁻¹, respectively. Clearly, the cleavage of the C–H bond on the benzylic position is less likely to be involved in the turnover limiting step, considering the KIE value of *ca.* 1.1. This result lends further support to the notion that the oxidative decarboxylation is not initiated by the benzylic C–H bond hydroxylation.

Subsequently, the reaction of carboxylic acids with the [Mn(dtbpy)₂(OTf)₂] complex was studied with UV-Vis spectroscopy. As shown in Fig. 1, there is an obvious decrease of the absorbance of **1a** at 230 nm when 1 equivalent of [Mn(dtbpy)₂(OTf)₂] was added. In the meantime, compared with that of [Mn(dtbpy)₂(OTf)₂], the spectrum of the mixture of [Mn(dtbpy)₂(OTf)₂] and **1a** showed a significant absorption decrease at around 295 nm and 306 nm. These observations indicate that [Mn(dtbpy)₂(OTf)₂] reacts with **1a** to form a new



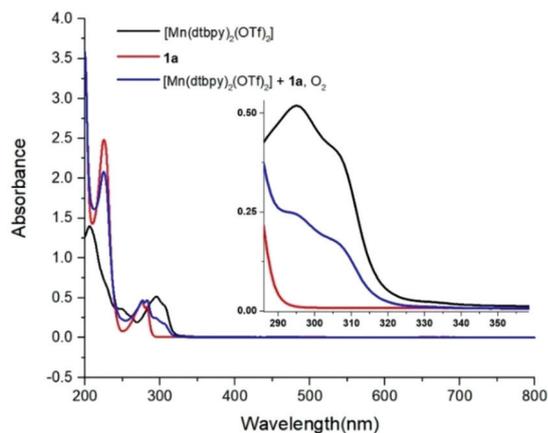


Fig. 1 UV-Vis spectra of $[\text{Mn}(\text{dtbp})_2(\text{OTf})_2]$, **1a** and the mixture of $[\text{Mn}(\text{dtbp})_2(\text{OTf})_2]$ and **1a** without blue light irradiation (concentration of $[\text{Mn}(\text{dtbp})_2(\text{OTf})_2]$ and **1a**: 25 μM in CH_3CN).

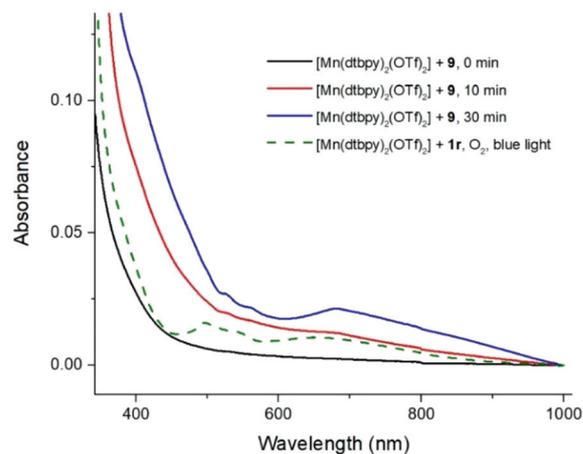


Fig. 3 *In situ* UV-Vis monitoring of the mixture of $[\text{Mn}(\text{dtbp})_2(\text{OTf})_2]$ and **9** and the UV-vis spectra of the mixture of $[\text{Mn}(\text{dtbp})_2(\text{OTf})_2]$ and **1r** with blue light irradiation for 2 h (concentration of each compound: 25 μM in CH_3CN).

$\text{Mn}(\text{II})$ species. Further structural information of the new $\text{Mn}(\text{II})$ species was obtained by HRMS, which suggests that the carboxylate-coordinated $[\text{Mn}(\text{dtbp})_2(\mathbf{1a}\text{-H})(\text{OTf})]$ is likely to be formed by losing a HOTf from the parent complex. Fig. 2 shows the experimental and calculated mass spectra of the molecular ion resulting from the loss of a ^-OTf fragment from the $[\text{Mn}(\text{dtbp})_2(\mathbf{1a}\text{-H})(\text{OTf})]$ complex.

We envisioned that a Mn-peroxide intermediate may be formed by the reaction of $[\text{Mn}(\text{dtbp})_2(\mathbf{1a}\text{-H})(\text{OTf})]$ with O_2 in the decarboxylative oxidation under blue light.^{34,67} Thus, a mixture of $[\text{Mn}(\text{dtbp})_2(\text{OTf})_2]$ and **1r**, which was expected to afford $[\text{Mn}(\text{dtbp})_2(\mathbf{1r}\text{-H})(\text{OTf})]$, was followed by UV-Vis spectroscopy with or without blue light irradiation. As shown in Fig. 3, an obvious absorption at *ca.* 685 nm was observed, when the mixture was irradiated by blue light under O_2 (Fig. 3, green dash line). Notably, a similar absorption band was also found in the mixture of $[\text{Mn}(\text{dtbp})_2(\text{OTf})_2]$ and **9** without blue light irradiation under N_2 , which increased over time and then reached saturation before beginning to decrease (Fig. S4 in ESI[†]). Finally, the product **2r** was formed in 60% yield after 60 min (Scheme S1[†]). These observations indicate that a peroxide intermediate may be involved and complexed with the catalyst in the process of decarboxylative oxidation. Comparing

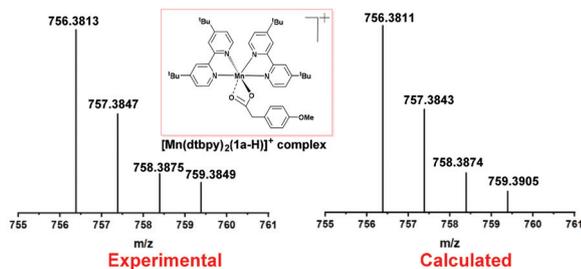
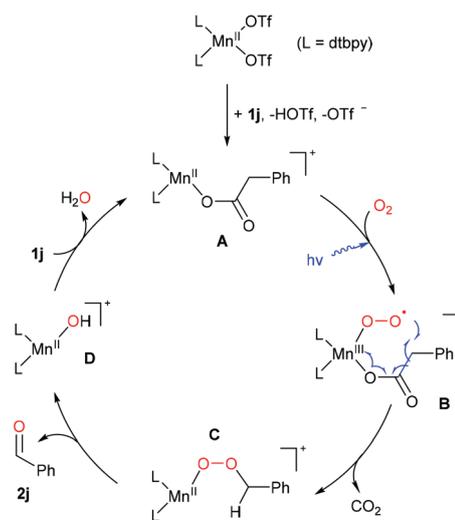


Fig. 2 HRMS spectrum of $[\text{Mn}(\text{dtbp})_2(\mathbf{1a}\text{-H})(\text{OTf})]$ formed in the *in situ* reaction of $[\text{Mn}(\text{dtbp})_2(\text{OTf})_2]$ with **1a**.

with the previous literature, the absorption at 685 nm is likely to arise from a Mn-peroxide species.^{68–70}

Based on the observations above and previous literature,^{25,36} a plausible mechanism is proposed by taking the oxidation of phenylacetic acid (**1j**) as an example. As shown in Scheme 7, **1j** firstly reacts with the $\text{Mn}(\text{II})$ precatalyst to form the intermediate **A**, which is subsequently oxidized by O_2 to afford the key species **B** under blue light irradiation.^{44,71–73} The unstable superoxide radical attacks the benzylic carbon of the coordinated acid driven by the release of CO_2 , resulting in the formation of the $\text{Mn}(\text{II})$ -peroxide **C**. Further decomposition of the unstable species **C** produces the carbonyl product and a $\text{Mn}(\text{II})\text{-OH}$ species **D**, which reacts with phenylacetic acid, regenerating **A**.



Scheme 7 A plausible mechanism of decarboxylative oxygenation of carboxylic acids (possible coordination of solvent or other molecules is omitted).



Conclusions

In conclusion, a selective, practical decarboxylative oxygenation protocol has been developed, which makes use of a non-heme Mn(II) complex as the catalyst and molecular oxygen as the oxidant under blue light irradiation. With this protocol, readily available carboxylic acids, including benzylic acids, amino acids, and pharmaceutical molecules, can be easily converted to a wide range of valuable aldehydes, ketones, and amides in good yields under mild conditions. Further mechanistic studies and applications of this catalytic system are underway in our laboratory and will be reported in due course.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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Notes and references

- 1 M. Simonetta, S. Carra, L. Ebersson, J. Koskikallio, R. Sneeden, V. Kucherov, L. Yanovskaya, L. Bergelson, M. Shemyakin and H. Kwart, *Carboxylic acids and esters*, Wiley, 1969.
- 2 M. Večeřa and J. Gasparič, *Detection and Identification of Organic Compounds*, Springer, 1971, pp. 247–293.
- 3 L. J. Goossen, N. Rodriguez and K. Goossen, *Angew. Chem., Int. Ed.*, 2008, **47**, 3100–3120.
- 4 J. Xuan, Z. G. Zhang and W. J. Xiao, *Angew. Chem., Int. Ed.*, 2015, **54**, 15632–15641.
- 5 F. Julia-Hernandez, M. Gaydou, E. Serrano, M. van Gemmeren and R. Martin, *Top. Curr. Chem.*, 2016, **374**, 45.
- 6 C. Shen, P. Zhang, Q. Sun, S. Bai, T. S. Hor and X. Liu, *Chem. Soc. Rev.*, 2015, **44**, 291–314.
- 7 Y. Wei, P. Hu, M. Zhang and W. Su, *Chem. Rev.*, 2017, **117**, 8864–8907.
- 8 J. Pritchard, G. A. Filonenko, R. van Putten, E. J. Hensen and E. A. Pidko, *Chem. Soc. Rev.*, 2015, **44**, 3808–3833.
- 9 A. Tortajada, M. Borjesson and R. Martin, *Acc. Chem. Res.*, 2021, **54**, 3941–3952.
- 10 T. P. Yoon, M. A. Ischay and J. Du, *Nat. Chem.*, 2010, **2**, 527–532.
- 11 J. A. Kautzky, T. Wang, R. W. Evans and D. W. C. MacMillan, *J. Am. Chem. Soc.*, 2018, **140**, 6522–6526.
- 12 N. Rodriguez and L. J. Goossen, *Chem. Soc. Rev.*, 2011, **40**, 5030–5048.
- 13 M. Pichette Drapeau and L. J. Goossen, *Chem. – Eur. J.*, 2016, **22**, 18654–18677.
- 14 S. Bhadra, W. I. Dzik and L. J. Goossen, *J. Am. Chem. Soc.*, 2012, **134**, 9938–9941.
- 15 C. P. Johnston, R. T. Smith, S. Allmendinger and D. W. MacMillan, *Nature*, 2016, **536**, 322–325.
- 16 S. Ventre, F. R. Petronijevic and D. W. MacMillan, *J. Am. Chem. Soc.*, 2015, **137**, 5654–5657.
- 17 A. Noble, S. J. McCarver and D. W. MacMillan, *J. Am. Chem. Soc.*, 2015, **137**, 624–627.
- 18 L. Chu, C. Ohta, Z. Zuo and D. W. MacMillan, *J. Am. Chem. Soc.*, 2014, **136**, 10886–10889.
- 19 Z. Zuo and D. W. MacMillan, *J. Am. Chem. Soc.*, 2014, **136**, 5257–5260.
- 20 F. Bu, L. Lu, X. Hu, S. Wang, H. Zhang and A. Lei, *Chem. Sci.*, 2020, **11**, 10000–10004.
- 21 Y. Yuan, Z. Zhou, L. Zhang, L. S. Li and A. Lei, *Org. Lett.*, 2021, **23**, 5932–5936.
- 22 Y. Liang, F. Strieth-Kalthoff, P. Bellotti and F. Glorius, *Chem. Catal.*, 2021, **1**, 1427–1436.
- 23 C. Zheng, Y. Wang, Y. Xu, Z. Chen, G. Chen and S. H. Liang, *Org. Lett.*, 2018, **20**, 4824–4827.
- 24 L. Wang, T. Wang, G.-J. Cheng, X. Li, J.-J. Wei, B. Guo, C. Zheng, G. Chen, C. Ran and C. Zheng, *ACS Catal.*, 2020, **10**, 7543–7551.
- 25 Q. Feng and Q. Song, *J. Org. Chem.*, 2014, **79**, 1867–1871.
- 26 M. Liu, H. Wang, H. Zeng and C. J. Li, *Sci. Adv.*, 2015, **1**, e1500020.
- 27 J. Li, H. Wang, Z. Qiu, C. Y. Huang and C. J. Li, *J. Am. Chem. Soc.*, 2020, **142**, 13011–13020.
- 28 L. Lv, D. Zhu and C. J. Li, *Nat. Commun.*, 2019, **10**, 715.
- 29 L. Lv, D. Zhu, J. Tang, Z. Qiu, C.-C. Li, J. Gao and C.-J. Li, *ACS Catal.*, 2018, **8**, 4622–4627.
- 30 H. Wang, X. J. Dai and C. J. Li, *Nat. Chem.*, 2017, **9**, 374–378.
- 31 B. Cornils, W. A. Herrmann and M. Rasch, *Angew. Chem., Int. Ed. Engl.*, 1994, **33**, 2144–2163.
- 32 R. Bruckner, *Advanced organic chemistry: reaction mechanisms*, Elsevier, 2001.
- 33 K. Weissermel and H.-J. Arpe, *Industrial organic chemistry*, John Wiley & Sons, 2008.
- 34 Y. Sakakibara, P. Cooper, K. Murakami and K. Itami, *Chem. – Asian J.*, 2018, **13**, 2410–2413.
- 35 T. M. Faraggi, W. Li and D. W. C. MacMillan, *Isr. J. Chem.*, 2019, **60**, 410–415.
- 36 S. Shirase, S. Tamaki, K. Shinohara, K. Hirose, H. Tsurugi, T. Satoh and K. Mashima, *J. Am. Chem. Soc.*, 2020, **142**, 5668–5675.
- 37 M. Araghi and F. Bokaei, *Polyhedron*, 2013, **53**, 15–19.
- 38 S. Tangestaninejad and V. Mirkhani, *J. Chem. Res.*, 1998, **32**, 820–821.
- 39 S. Farhadi, P. Zaringhadam and R. Z. Sahamieh, *Tetrahedron Lett.*, 2006, **47**, 1965–1968.
- 40 Y. Pocker and B. C. Davis, *J. Am. Chem. Soc.*, 1973, **95**, 6216–6223.
- 41 K. Gholamreza and A. Roxana, *Chem. Res. Chin. Univ.*, 2008, **24**, 464–468.



- 42 T. B. Mete, T. M. Khopade and R. G. Bhat, *Tetrahedron Lett.*, 2017, **58**, 2822–2825.
- 43 T. Rahman, G. Borah and P. K. Gogoi, *Catal. Lett.*, 2020, **150**, 2267–2272.
- 44 S. Sahu and D. P. Goldberg, *J. Am. Chem. Soc.*, 2016, **138**, 11410–11428.
- 45 B. Zhu, T. Shen, X. Huang, Y. Zhu, S. Song and N. Jiao, *Angew. Chem., Int. Ed.*, 2019, **58**, 11028–11032.
- 46 Y. F. Liang and N. Jiao, *Acc. Chem. Res.*, 2017, **50**, 1640–1653.
- 47 S. Paria, L. Que Jr. and T. K. Paine, *Angew. Chem., Int. Ed.*, 2011, **50**, 11129–11132.
- 48 R. A. Baglia, J. P. T. Zaragoza and D. P. Goldberg, *Chem. Rev.*, 2017, **117**, 13320–13352.
- 49 K. A. Prokop and D. P. Goldberg, *J. Am. Chem. Soc.*, 2012, **134**, 8014–8017.
- 50 K. A. Prokop, H. M. Neu, S. P. de Visser and D. P. Goldberg, *J. Am. Chem. Soc.*, 2011, **133**, 15874–15877.
- 51 S. Fukuzumi, H. Kotani, K. A. Prokop and D. P. Goldberg, *J. Am. Chem. Soc.*, 2011, **133**, 1859–1869.
- 52 S. Riaño, D. Fernández and L. Fadini, *Catal. Commun.*, 2008, **9**, 1282–1285.
- 53 S. Barroso, G. Blay, I. Fernández, J. R. Pedro, R. Ruiz-García, E. Pardo, F. Lloret and M. C. Muñoz, *J. Mol. Catal. A: Chem.*, 2006, **243**, 214–220.
- 54 M. Guo, J. Zhang, L. Zhang, Y. M. Lee, S. Fukuzumi and W. Nam, *J. Am. Chem. Soc.*, 2021, **143**, 18559–18570.
- 55 M. Guo, Y. M. Lee, S. Fukuzumi and W. Nam, *Coord. Chem. Rev.*, 2021, **435**, 213807.
- 56 G. Li, P. A. Kates, A. K. Dilger, P. T. Cheng, W. R. Ewing and J. T. Groves, *ACS Catal.*, 2019, **9**, 9513–9517.
- 57 W. Liu, X. Huang, M. J. Cheng, R. J. Nielsen, W. A. Goddard 3rd and J. T. Groves, *Science*, 2012, **337**, 1322–1325.
- 58 Z. Huang, R. Guan, M. Shanmugam, E. L. Bennett, C. M. Robertson, A. Brookfield, E. J. L. McInnes and J. Xiao, *J. Am. Chem. Soc.*, 2021, **143**, 10005–10013.
- 59 A. Uttry and M. van Gemmeren, *Synthesis*, 2019, 479–488.
- 60 K. Kiyokawa, S. Yahata, T. Kojima and S. Minakata, *Org. Lett.*, 2014, **16**, 4646–4649.
- 61 N. A. Herman and W. Zhang, *Curr. Opin. Chem. Biol.*, 2016, **35**, 22–28.
- 62 D. L. Boger and C. M. Baldino, *J. Org. Chem.*, 2002, **56**, 6942–6944.
- 63 M. Scholz, R. Dedic, T. Breitenbach and J. Hala, *Photochem. Photobiol. Sci.*, 2013, **12**, 1873–1884.
- 64 Y. You, *Org. Biomol. Chem.*, 2018, **16**, 4044–4060.
- 65 D. Sheet and T. K. Paine, *Chem. Sci.*, 2016, **7**, 5322–5331.
- 66 M. L. Neidig, A. Decker, O. W. Choroba, F. Huang, M. Kavana, G. R. Moran, J. B. Spencer and E. I. Solomon, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**, 12966–12973.
- 67 O. W. Choroba, D. H. Williams and J. B. Spencer, *J. Am. Chem. Soc.*, 2000, **122**, 5389–5390.
- 68 N. Kitajima, H. Komatsuzaki, S. Hikichi, M. Osawa and Y. Moro-oka, *J. Am. Chem. Soc.*, 2002, **116**, 11596–11597.
- 69 T. A. Jackson, A. Karapetian, A. F. Miller and T. C. Brunold, *Biochemistry*, 2005, **44**, 1504–1520.
- 70 S. Groni, G. Blain, R. Guillot, C. Policar and E. Anxolabehere-Mallart, *Inorg. Chem.*, 2007, **46**, 1951–1953.
- 71 J. A. Kovacs, *Acc. Chem. Res.*, 2015, **48**, 2744–2753.
- 72 M. K. Coggins, X. Sun, Y. Kwak, E. I. Solomon, E. Rybak-Akimova and J. A. Kovacs, *J. Am. Chem. Soc.*, 2013, **135**, 5631–5640.
- 73 M. Sankaralingam, Y. M. Lee, S. H. Jeon, M. S. Seo, K. B. Cho and W. Nam, *Chem. Commun.*, 2018, **54**, 1209–1212.

