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Preparation of iminosugars from aminopolyols via selective oxidation using galactose oxidase†

Kathryn Yeow,^a Marianne B. Haarr,^a Jimmy Muldoon^b and Elaine O'Reilly^{a*}

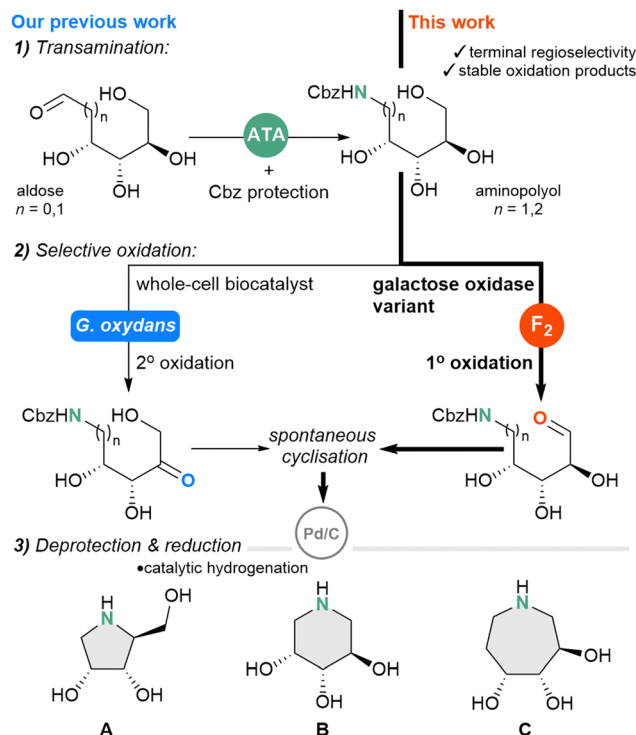
Minimally protected aminopolyols are novel substrates for the galactose oxidase variant F₂. Site-selective oxidation proceeds at the terminal primary alcohol, followed by spontaneous cyclisation to afford stable hemiaminal/hemiacetal anomers of the piperidine and azepane scaffolds, with isolated yields of up to 94%. Simultaneous deprotection and reduction occurred readily to afford valuable and biologically relevant iminosugars.

Iminosugars are polyhydroxylated alkaloids and sugar mimics, containing a nitrogen atom in place of the endocyclic oxygen (Scheme 1). These naturally occurring compounds are of pharmaceutical importance because they interact with and inhibit carbohydrate processing enzymes. Known mechanisms of action include mimicking the oxocarbenium-ion transition state of glycoside hydrolysis,¹ chaperoning misfolded proteins,² and allosteric chaperoning.³ Therapeutic applications are wide-ranging and include the treatment of type II diabetes (miglitol),⁴ Gaucher's disease (miglustat)⁵ and viral infections.^{6,7} Whilst the drug-like properties of iminosugars are advantageous, such as chemical and metabolic stability, and good oral bioavailability, first-generation candidates suffer from lack of selectivity, potency and give rise to undesirable side effects.^{8,9} The absence of efficient syntheses for preparing structurally diverse derivatives has been highlighted as a limitation to the advancement of second-generation iminosugars as therapeutics.⁸ A robust biocatalytic route for the preparation of iminosugar compounds has not yet been developed and has the potential to greatly simplify and streamline their synthesis.

Through a biocatalytic retrosynthetic analysis, we propose a sequential, three-step chemo-enzymatic cascade, whereby monosaccharides undergo transamination, selective oxidation,

and reduction, *via* transaminase (TA), oxidoreductase and catalytic hydrogenation steps, respectively (Scheme 1).

In support of this biomimetic strategy, a biosynthetic gene cluster that initiates 1-deoxynorijirimycin synthesis has been reported and encodes for a TA, alcohol dehydrogenase, and phosphorylase; transforming fructose-6-phosphate into an iminosugar scaffold.^{10,11} The prospect of accessing aminopolyols from readily available monosaccharides is intriguing, as naturally occurring D-sugars are a major component of plant



Scheme 1 Biocatalytic routes to pyrrolidine-(A), piperidine-(B) and azepane-(C) type iminosugars from monosaccharides. Previous work includes transamination¹² and *G. oxydans* mediated secondary oxidation²⁰ ATA = amine transaminase, F₂ = galactose oxidase variant F₂.

^a School of Chemistry, University College Dublin, Belfield, Dublin 4, Ireland.
E-mail: elaine.oreilly@ucd.ie, marianne.haarr@ucd.ie

^b Mass Spectrometry Facility, School of Chemistry, University College Dublin, Belfield, Dublin 4, Ireland

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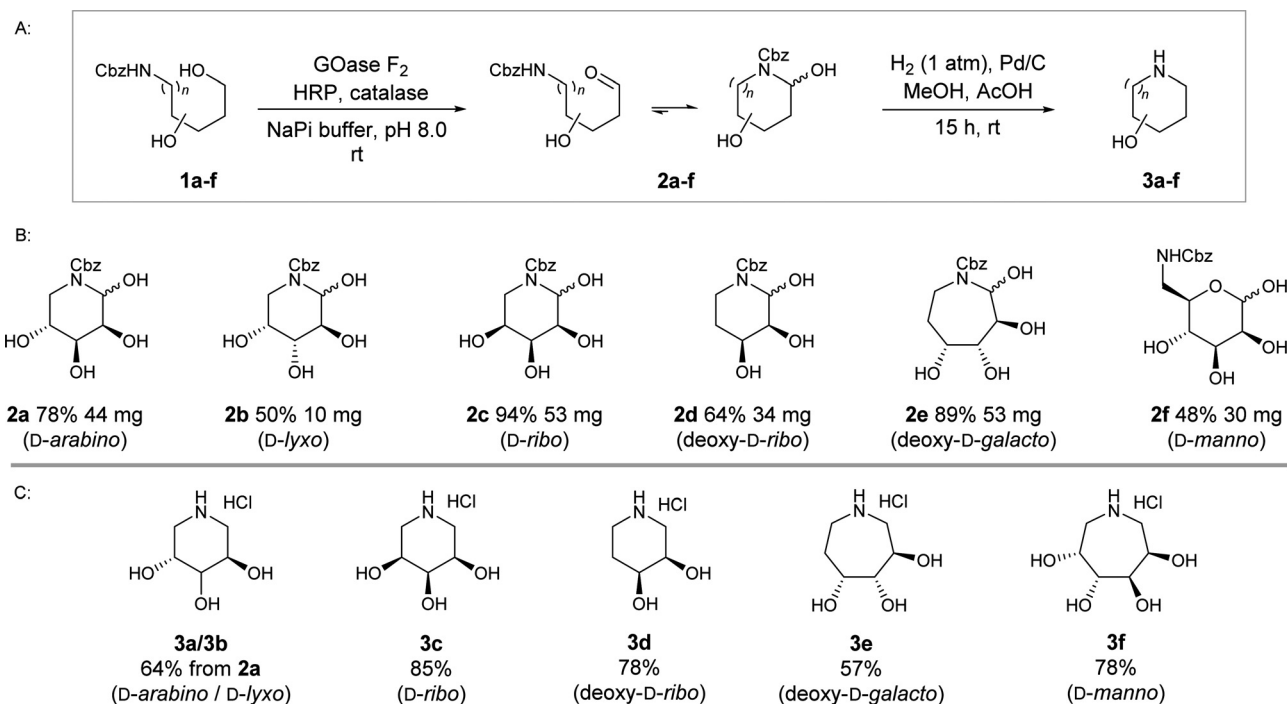


biomass and considered waste products from industry and agriculture, therefore utilising this feedstock represents a useful valorisation approach. Recently, we reported that TAs can catalyse the direct amination of aldoses to aminopolyols (Scheme 1);¹² a study revealing activity towards ketoses was reported shortly after.¹³

With the first key transaminase step established to afford the aminopolyols from aldoses, we then sought a biocatalytic strategy for the selective oxidation, which must proceed regioselectively at the C5- or C6-OH positions to enable cyclisation. Synthetic approaches for regioselective oxidation of polyol species include the use of transition-state metal (Ru, Pd)^{14,15} and photoredox catalysis,¹⁶ and has even been applied to unprotected carbohydrates, yet scope remains limited to cyclic substrates. The challenge of achieving regioselective oxidation persists for linear polyol species, for which there are fewer variations in the electronic and steric environments between hydroxyl groups. Building on early reports,^{17–19} we recently employed *Gluconobacter oxydans* as a whole-cell biocatalyst, which displayed regioselectivity towards the secondary alcohol positions (Scheme 1).²⁰ Since the enzyme(s) responsible for the oxidation was uncharacterised, we next wished to identify specific oxidoreductases that could add to our toolkit of selective aminopolyol oxidation biocatalysts. We reasoned that it would also be valuable to find an enzyme to carry out the primary aminopolyol oxidation to complement the secondary oxidation mediated by *G. oxydans*, allowing more piperidine-type iminosugars to be derived from the pentose monosaccharides.

Our attention turned towards oxidase enzymes of which, many carbohydrate oxidases have been characterised in literature.²¹

In comparison to the nicotinamide coenzyme requirements of dehydrogenases, oxidases use molecular O₂ as the mild oxidant for the regeneration of the reduced enzyme. Galactose oxidase (GOase) from *Fusarium graminearum* is a well characterised copper-dependent oxidase, and the wild-type enzyme regioselectively oxidises the primary C6-OH of D-galactopyranose. In comparison, other carbohydrate oxidases tend to favour secondary C2-OH oxidation.²² GOase has been the subject of several engineering projects, delivering variants with outcomes such as increased expression and stability in *Escherichia coli* (*E. coli*) (M₁ variant),²³ activity towards non-natural substrates, such as glucose (M₃ variant)²⁴ and fructose (R330K variant)²⁵ as well as activity towards aromatic secondary alcohols (M_{3–5} variant)²⁶ and glycans on glycoproteins (F₂ variant) (ESI,† Fig. S2).²⁷ Following these advances, optimising standard operating conditions for GOase has been pursued in a step towards realising its industrial relevance.²⁸ Recently, GOase F₂ was evolved further for the desymmetrizing oxidation step in the enzymatic cascade for the synthesis of islatravir.²⁹ For this project, the F₂ variant was particularly attractive for its ability to catalyse the terminal selective oxidation of glucosides, showing that activity towards a polyhydroxylated scaffold is retained.³⁰ If the site-selective oxidation of aminopolyols can be performed with the same regioselectivity, the resulting aminoaldehyde products will cyclise to afford the valuable piperidine and azepane ring systems (see Scheme 2) and therefore,



Scheme 2 (A) Chemoenzymatic synthesis of piperidine- and azepane-type iminosugars. (B) Substrate scope of the oxidation products and isolated yields, products **2a–e** exist predominantly in hemiaminal form, whereas **2f** is in hemiacetal form. (C) Iminosugars **3a–f** are isolated as the HCl salt.



the F₂ variant was the biocatalyst candidate selected for this study.

In view of developing a biocatalytic cascade, it was important that selected sugar starting materials are substrates for the initial transamination. The panel of substrates **1a–f** (Scheme 2, ESI† Fig. S1) were derived from D-aldoses arabinose, lyxose, ribose, 2-deoxy-ribose, mannose, and 2-deoxy-galactose and reflect the rich stereochemical landscape that can be accessed through sugar precursors. Aminopolyols were N-Cbz protected to enable isolation and characterisation of the oxidation products.

The enzyme GOase F₂ was recombinantly expressed in *E. coli* BL21(DE3) according to modified protocols³¹ and purified to homogeneity by streptavidin affinity chromatography, with modest yields of approximately 12 mg per litre of culture. In the initial set of experiments, we screened **1a–f** as substrates for the F₂-catalysed oxidation on an analytical scale (1 mL) (see ESI†, Section 3). Having established suitable conditions for reaction screening, we opted to monitor biotransformations directly using thin layer chromatography (TLC). TLC analysis indicated conversion of starting material for all the tested substrates after an overnight reaction (see ESI† Fig. S4). LC/MS analysis confirmed the formation of a mono-oxidation product and there was no evidence of side product formation, such as overoxidation to the carboxylic acid or the geminal diol species that has been previously reported.³⁰

Next, we sought to isolate enough material for full characterisation, as up until this point, neither the site of oxidation nor the form of the oxidised products had been determined. Furthermore, the compounds are not readily accessible commercially or *via* chemical synthesis, and calibration standards for quantifying reaction conversion were required. For these reasons, biotransformations were scaled up to 10 mL, with substrate loading at 20 mM (0.2 mmol),[‡] relatively high compared to what has typically been reported for GOase-catalysed reactions. From the analytical-scale reactions it was evident that the GOase F₂ variant precipitates readily from the reaction mixture, and consequently, over time activity likely would diminish. To circumvent a drop off in the reaction progression and to drive the reactions towards completion, we opted to cumulatively add enzyme over the course of the reaction. The progress of the reaction and timeline of sequential additions was informed by TLC analysis. Through these efforts, oxidation products **2a–f** were isolated in 48–94% yield, corresponding to up to 53 mg of the D-ribo **2c** and 2-deoxy-D-galacto **2e** compounds. It was not possible to achieve complete conversion with all biotransformations within a reasonable time frame, meaning D-lyxo **1b** and D-manno **1f** compounds were isolated in yields of approximately 50%. Despite long reaction times, we observed no side product formation, and that the oxidation products were remarkably stable.

Characterisation of the compounds by NMR spectroscopy was initially challenging due to the presence of anomeric mixtures, but analysis of 2D NMR spectra enabled resolution between configurational anomers. The spontaneous cyclisation was supported by the appearance of the characteristic cyclic

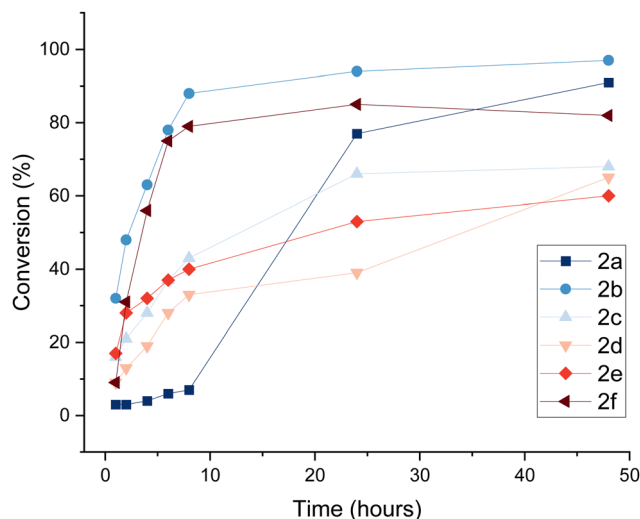


Fig. 1 Time course analysis of the GOase F₂-catalysed oxidation of N-Cbz-protected aminopolyols. Conversion to products **2a–f** was determined by LC/MS analysis, monitoring product formation at timepoints up to 48 hours, and plotted as the average of triplicate reactions (1 mL) (for individual plots with error bars, see ESI†, Section 4).

hemiaminal or hemiacetal peak in the range δ 5.1–5.8 ppm. From this, the ratio of hemiaminal anomers was determined, with product ratios differing across the substrate panel. The absolute stereochemistry of each anomer was not ascertained in this study, with the view that upon deprotection and imine reduction, the anomers converge to a single product.

To gain a better understanding of the reaction progress, we determined the reaction conversion after a single addition of enzyme and monitored the reaction progress over time on analytical scale (Fig. 1, Table 1). After 8 h, conversion of substrates **1b–f** to oxidised species **2b–f** ranged from 33–88%. The initially slow reaction progress of entry 1 can be attributed to the poor solubility of the aminopolyol **1a** in buffer at ambient temperature. Despite protein precipitation in the reaction mixture, enzymatic activity was still evident after 24 hours. The results show that aminopolyols **1a–f** are good substrates for F₂, reaching up to 97% conversion on analytical scale.

Table 1 Conversion data for GOase F₂-catalysed oxidation of N-Cbz-protected aminopolyols

	Substrate	Absolute configuration	Product ^a	Conversion [%]		
				8 h	24 h	48 h
1	1a	D-arabino	2a	7	77	91
2	1b	D-lyxo	2b	88	94	97
3	1c	D-ribo	2c	43	66	68
4	1d	2-deoxy-D-ribo	2d	33	39	65
5	1e	2-deoxy-D-galacto	2e	40	53	60
6	1f	D-manno	2f	79	85	82

Reaction conditions: substrate (0.2 mmol, 20 mM), catalase (0.1 mg mL⁻¹), HRP Type I (0.1 mg mL⁻¹), CuSO₄ (50 μ M), NaPi (100 mM, pH 8), purified GOase F₂ (2 mg mL⁻¹) at 20 °C. ^a A mixture of cyclic hemiaminal and hemiacetal anomers are obtained.



Finally, iminosugars **3a–f** were readily obtained from the oxidation products **2a–f** by hydrogenation using Pd/C, in which the deprotection and imine reduction steps occur concomitantly (Scheme 2). After 16 h reaction time we observed complete conversion to the desired products **3a–f**, regardless of the anomeric hemiaminal/hemiacetal distribution of the precursors **2a–f**.

We have demonstrated that minimally protected aminopolyols, key intermediates for iminosugar synthesis, are novel substrates for the galactose oxidase variant F₂. This report represents further progress from our previously reported chemoenzymatic route towards monosaccharide-derived iminosugars.²⁰ Having identified the F₂ variant as a suitable oxidase, protein engineering and optimisation studies can now follow to extend the substrate scope, efficiency, stability, and expression of this catalyst. The necessity for sequential addition of enzyme, due to poor stability, enabled enough material to be isolated for characterisation and further analysis, but could potentially be avoided by using immobilised enzyme. Biocatalytic and chemoenzymatic cascades offer enormous advantages over more traditional stepwise synthetic approaches to complex molecules and are being considered as viable industrial approaches.³² Future efforts will focus on eliminating the need for the *N*-Cbz protecting group on the aminopolyol and identifying and introducing an appropriate imine reductase (IRED) in the last step of the cascade to enable a one-pot fully enzymatic cascade from monosaccharides to iminosugars.

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Conflicts of interest

There are no conflicts of interest to declare.

Notes and references

† The batch prep reaction of *D*-lyxo substrate is carried out on 5 mL scale.

- D. L. Zechel, A. B. Boraston, T. Gloster, C. M. Boraston, J. M. Macdonald, D. M. G. Tilbrook, R. V. Stick and G. J. Davies, *J. Am. Chem. Soc.*, 2003, **125**, 14313–14323.
- C. H. Hill, A. H. Viuff, S. J. Spratley, S. Salamone, S. H. Christensen, R. J. Read, N. W. Moriarty, H. H. Jensen and J. E. Deane, *Chem. Sci.*, 2015, **6**, 3075–3086.
- D. D'Alonzo, M. De Fenza, C. Porto, R. Iacono, M. Huebecker, B. Cobucci-Ponzano, D. A. Priestman, F. Platt, G. Parenti, M. Moracci, G. Palumbo and A. Guaragna, *J. Med. Chem.*, 2017, **60**, 9462–9469.
- L. J. Scott and C. M. Spencer, *Drugs*, 2000, **59**, 521–549.
- C. E. M. Hollak, D. Hughes, I. N. Van Schaik, B. Schwierin and B. Bembi, *Pharmacoevidenciol. Drug Saf.*, 2009, **18**, 770–777.
- D. S. Alonzi, K. A. Scott, R. A. Dwek and N. Zitzmann, *Biochem. Soc. Trans.*, 2017, **45**, 571–582.
- K. Prichard, D. Campkin, N. O'Brien, A. Kato, G. W. Fleet and M. I. Simone, *Chem. Biol. Drug Des.*, 2018, **92**, 1171–1197.
- G. Horne, F. X. Wilson, J. Tinsley, D. H. Williams and R. Storer, *Drug Discovery Today*, 2011, **16**, 107–118.
- R. J. Nash, A. Kato, C. Y. Yu and G. W. Fleet, *Future Med. Chem.*, 2011, **3**, 1513–1521.
- L. F. Clark, J. V. Johnson and N. A. Horenstein, *ChemBioChem*, 2011, **12**, 2147–2150.
- K. D. Kang, Y. S. Cho, J. H. Song, Y. S. Park, J. Y. Lee, K. Y. Hwang, S. K. Rhee, J. H. Chung, O. Kwon and S. Il Seong, *J. Microbiol.*, 2011, **49**, 431–440.
- R. Cairns, A. Gomm, J. Ryan, T. Clarke, E. Kulcinskaja, K. Butler and E. O'Reilly, *ACS Catal.*, 2019, **9**, 1220–1223.
- F. Subrizi, L. Benhamou, J. M. Ward, T. D. Sheppard and H. C. Hailes, *Angew. Chem., Int. Ed.*, 2019, **58**, 3854–3858.
- M. Jäger, M. Hartmann, J. G. De Vries and A. J. Minnaard, *Angew. Chem., Int. Ed.*, 2013, **52**, 7809–7812.
- C. K. Hill and J. F. Hartwig, *Nat. Chem.*, 2017, **9**, 1213–1221.
- D. J. Gorelik, V. Dimakos, T. Adrianov and M. S. Taylor, *Chem. Commun.*, 2021, **57**, 12135–12138.
- M. Schedel, in *Biotechnology: Biotransformations II*, ed. D. R. Kelly, WILEY-VCH Verlag GmbH; Weinheim, 2nd edn, 2000, vol. 8b, ch. 7, pp 295–311.
- M. K. Sethi, A. Kumar, N. Maddur, R. Shukla and L. N. Vemula, *J. Mol. Catal. B: Enzym.*, 2015, **112**, 54–58.
- N. G. Oikonomakos, C. Tiraidis, D. D. Leonidas, S. E. Zographos, M. Kristiansen, C. U. Jessen, L. Nørskov-Lauritsen and L. Agius, *J. Med. Chem.*, 2006, **49**, 5687–5701.
- J. Kuska, F. Taday, K. Yeow, J. Ryan and E. O'Reilly, *Catal. Sci. Technol.*, 2021, **11**, 4327–4331.
- S. Savino and M. W. Fraaije, *Biotechnol. Adv.*, 2021, **51**, 107634.
- P. Ragaini, P. Bollella, G. Pankratova, C. K. Peterbauer, P. Conghaile, D. Leech, B. Haghighi and L. Gorton, *ChemElectroChem*, 2019, **6**, 801–809.
- L. Sun, I. P. Petrounia, M. Yagasaki, G. Bandara and F. H. Arnold, *Protein Eng., Des. Sel.*, 2001, **14**, 699–704.
- L. Sun, T. Bulter, M. Alcalde, I. P. Petrounia, F. H. Arnold, F. H. Arnold, L. Sun, T. Bulter and M. Alcalde, *ChemBioChem*, 2002, **8**, 781–783.
- S. E. Deacon, K. Mahmoud, R. K. Spooner, S. J. Firbank, P. F. Knowles, S. E. V. Phillips and M. J. McPherson, *ChemBioChem*, 2004, **5**, 972–979.
- F. Escalettes and N. J. Turner, *ChemBioChem*, 2008, **9**, 857–860.
- J. B. Rannes, A. Ioannou, S. C. Willies, G. Grogan, C. Behrens, S. L. Flitsch and N. J. Turner, *J. Am. Chem. Soc.*, 2011, **133**, 8436–8439.
- A. Toftgaard Pedersen, W. R. Birmingham, G. Rehn, S. J. Charnock, N. J. Turner and J. M. Woodley, *Org. Process Res. Dev.*, 2015, **19**, 1580–1589.
- M. A. Huffman, A. Fryszkowska, O. Alvizo, M. Borra-Garske, K. R. Campos, K. A. Canada, P. N. Devine, D. Duan, J. H. Forstater, S. T. Grosser, H. M. Halsey, G. J. Hughes, J. Jo, L. A. Joyce, J. N. Kolev, J. Liang, K. M. Maloney, B. F. Mann, N. M. Marshall, M. McLaughlin, J. C. Moore, G. S. Murphy, C. C. Nawrat, J. Nazor, S. Novick, N. R. Patel, A. Rodriguez-Granillo, S. A. Robaire, E. C. Sherer, M. D. Truppo, A. M. Whittaker, D. Verma, L. Xiao, Y. Xu and H. Yang, *Science*, 2019, **366**, 1255–1259.
- S. C. Cosgrove, A. P. Matthey, M. Riese, M. R. Chapman, W. R. Birmingham, A. J. Blacker, N. Kapur, N. J. Turner and S. L. Flitsch, *ACS Catal.*, 2019, **9**, 11658–11662.
- J. Vilím, T. Knaus and F. G. Mutti, *Angew. Chem., Int. Ed.*, 2018, **57**, 14240–14244.
- A. R. Alcántara, P. Domínguez de María, J. A. Littlechild, M. Schürmann, R. A. Sheldon and R. Wohlgenuth, *ChemSusChem*, 2022, **15**, e202102709.

