



Cite this: *Chem. Commun.*, 2022, 58, 10861

Received 27th July 2022,
Accepted 27th August 2022

DOI: 10.1039/d2cc04199e

rsc.li/chemcomm

Selective NMR detection of *N*-methylated amines using cavitand-decorated silica nanoparticles as receptors†‡

Andrea Cesari,^{§a} Daniele Rosa-Gastaldo,^{§a} Alessandro Pedrini,^{§b} Federico Rastrelli,^{§a} Enrico Dalcanale,^{§b} Roberta Pinalli^{§b} and Fabrizio Mancin^{§a}*

We report a strategy for the realization of NMR chemosensors based on the spontaneous self-assembly of lower rim pyridinium-functionalized tetraphosphonate cavitands on commercial silica nanoparticles. These nanohybrids enable the selective detection of physiologically relevant *N*-methylated amines, with a limit of detection of 31 μ M, via STD-based NMR experiments, achieving for the first time fine structural selectivity in nanoparticle-assisted NMR chemosensing.

Nuclear Magnetic Resonance (NMR) spectroscopy shares with a few other techniques the advantage of identifying analytes according to their spectral fingerprint. This virtually eliminates the possibility of false positives in analytic applications and allows the identification of unknown species.^{1,2a,b} However, in most of the cases, the number of signals obtained by analysing a mixture is overwhelming, and overlapping prevents the individuation of the single components. To overcome this issue, we perfected over the last years the “nanoparticle-assisted NMR chemosensing” technique.³ This protocol combines the recognition capabilities of nanoparticle receptors with NOE-based NMR experiments, which enable to transfer magnetization between the nanoparticles and the bound analytes. In this way, it is possible to tag and extract only the target signals from the spectrum of the mixture analysed.

Over the years, this technique has been gradually improved reaching limits of detection (LOD) in the low μ M range.³ This required the design of high-performance NMR experiments^{4,5} and

the optimization of the nanoreceptors.^{6,7} Beside recognizing the analyte, the fundamental role of the nanoreceptor is to enable an efficient magnetization transfer thanks to its long correlation time (τ_c).⁸ In this perspective, the 2 nm ligand-coated gold nanoparticles (AuNPs, $\tau_c \approx 10$ ns) used as first-generation self-assembled nanoreceptors were underperforming. In fact, although theory predicts that optimal NOE enhancements should be reached at τ_c longer than 1 ns,⁹ the finite association lifetime between the analyte and the receptor itself moves this threshold towards longer correlation times (about 1 μ s),⁹ which are attained by nanoscopic objects with diameters larger than 12 nm.¹⁰ A relevant improvement was hence achieved when a second-generation nanoreceptors was conceived, based on the self-assembly of charged AuNPs on larger (15–20 nm) silica nanoparticles bearing the opposite charge.⁹

In the evolution of these chemosensing systems, a still lacking feature is selectivity. Indeed, AuNPs used so far as receptors could only distinguish broad classes of analytes (cations, anions).⁶ To address this issue we reasoned that the self-assembly of charged supramolecular hosts on the surface of silica nanoparticles would result in nanohybrid receptors large enough to allow the effective transfer of magnetization (Fig. 1a), providing a general strategy to transfer the selectivity of supramolecular hosts to NMR chemosensing.

To test this hypothesis, we selected a tetraphosphonate cavitand bearing four propyl pyridinium moieties at the lower rim (Tiiii-Py, Fig. 1b).¹¹ Tetraphosphonate cavitands are well known for their ability to selectively recognize biologically relevant *N*-methylated ammonium derivatives.¹¹ Such features allowed the successful coupling of these hosts with different techniques for the detection of tumour markers^{12–14} and psychoactive substances.^{15,16} To form the nanohybrids, we selected commercial LUDOX HS-30 silica nanoparticles, which are readily available, cheap, and have an average size of 13–15 nm (Fig. S1 and S2, ESI†). The four positive charges of the Tiiii-Py should grant its adsorption on the negatively charged surface of silica nanoparticles. The sensing performances were tested on three different phenethylamine analogues (Fig. 1b), since this

^a Dipartimento di Scienze Chimiche, Università di Padova, via Marzolo 1, 35131 Padova, Italy. E-mail: fabrizio.mancin@unipd.it

^b Dipartimento di Scienze Chimiche, della Vita e della Sostenibilità Ambientale and INSTM Udr Parma - Università di Parma, Parco Area delle Scienze, 17/A, 43124, Parma, Italy. E-mail: roberta.pinalli@unipr.it

† This paper is dedicated to Prof. Paolo Scrimin, on the occasion of its 70th birthday.

‡ Electronic supplementary information (ESI) available: Experimental details, additional NMR experiments, binding isotherms. See DOI: <https://doi.org/10.1039/d2cc04199e>

§ These authors equally contributed to the study.



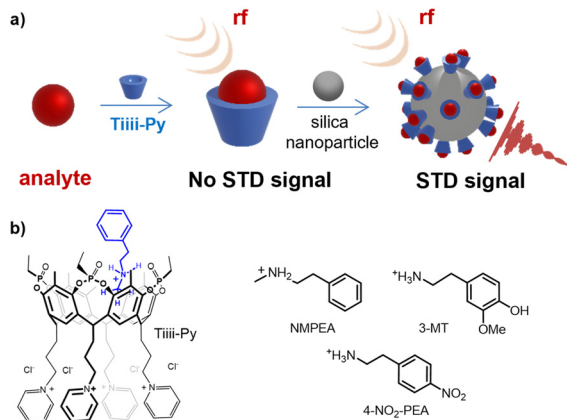


Fig. 1 (a) Sensing scheme studied in this work; (b) left: chemical structure of TiIII-Py cavitand (with included NMPEA), right: analytes used in this work.

family of compounds is of great relevance as neurotransmitters, drugs and even biomarkers.¹⁷

At first, we investigated the association of TiIII-Py to LUDOX nanoparticles. Preliminary ¹H-NMR experiments showed that the addition of LUDOX to a 0.5 mM TiIII-Py solution in buffered D₂O (phosphate buffer 5 mM, pD = 7.0) resulted in the progressive disappearance of the TiIII-Py signals, reaching saturation at 400 cavitand molecules per particle (Fig. S3 and S4, ESI†). To obtain more information on the interaction of TiIII-Py with silica nanoparticle we used 3-methoxytyramine (3-MT, Fig. 1b) as probe molecule. Being a primary amine, 3-MT is not recognized by TiIII-Py and indeed no changes were observed in the NMR spectrum when the two were mixed (Fig. 2a and b). On the contrary, broadening of the 3-MT signals was observed when it was added to a suspension of LUDOX (Fig. 2c). The subsequent addition of TiIII-Py caused the re-sharpening of the 3-MT signals (Fig. 2d) to linewidths similar to those recorded in samples containing 3-MT alone (Fig. 2a).

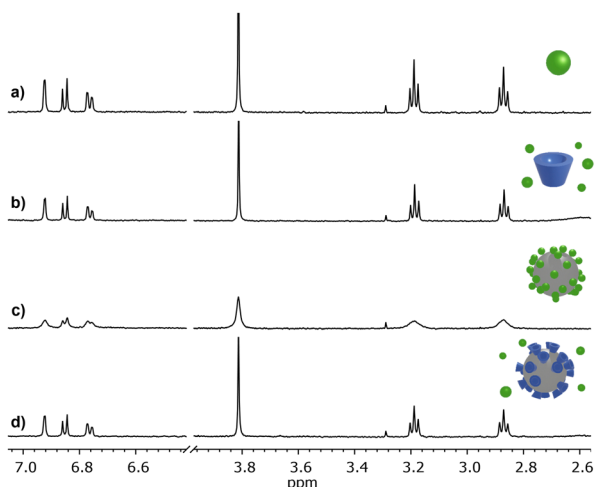


Fig. 2 ¹H-NMR subspectra of (a) 3-MT (green sphere) 0.25 mM; (b) 3-MT 0.25 mM and TiIII-Py (blue basket) 0.1 mM; (c) 3-MT 0.25 mM and LUDOX (grey sphere) 0.15 μM; (d) 3-MT 0.25 mM, TiIII-Py 0.1 mM and LUDOX 0.15 μM (Conditions: 500 MHz, 25 °C, phosphate buffer 5 mM, pD = 7.0, D₂O).

Taken together, the above NMR experiments confirmed the absorption of TiIII-Py on the surface of the nanoparticles to form TiIII-Py@LUDOX nanohybrids. The grafting of small molecules to nanoparticles causes the broadening of their NMR signals because of the reduction of the tumbling rate.¹⁸ Obviously, if surface binding and dissociation are fast with respect to the NMR timescale, the extent of the observed broadening depends on the strength of the interaction, since the apparent lineshape is (approximately) averaged over the populations of the bound and unbound species. 3-MT has only one positive charge and consequently it is expected to adsorb weakly to the nanoparticles.^{18–20} For this reason, the broadening of its signals was moderate. On the other hand, TiIII-Py has four positive charges that ensure a strong adsorption onto the silica surface.^{19,20} The consequent full adsorption of TiIII-Py to the nanoparticles resulted in broadening of its signals beyond detection. Because of its high affinity for silica, TiIII-Py completely displaced the adsorbed 3-MT molecules from the particles when it was added to the mixture of 3-MT and LUDOX, restoring the sharpness of 3-MT signals (Fig. 2).

Interaction of polycharged species with charged nanoparticles can lead to the formation of a corona but also to nanoparticles crosslinking and aggregation.¹⁹ To investigate this point, we performed Dynamic Light Scattering (DLS) experiments. These showed that subsequent additions of TiIII-Py (up to 100 μM) to a 1.5 μM suspension of LUDOX in 5 mM phosphate buffer (pH = 7.0) resulted in a moderate nanoparticles hydrodynamic diameter increase (from 12.8 to 17.4 nm), as expected in the case of the formation of a layer of TiIII-Py on the surface of the nanoparticles. This diameter increase was accompanied by the formation of a small number of larger aggregates (Table S1, ESI†). However, no relevant precipitation was observed for several hours in the conditions of the NMR sensing experiments.

Having established the formation of the TiIII-Py@LUDOX nanohybrids, we investigated their ability to bind *N*-methylammonium ions. In the first experiment, ¹H-NMR spectra of *N*-methyl phenethylamine (NMPEA, 0.5 mM in D₂O buffered at pD = 7.0 with 5 mM phosphate buffer) were recorded in the presence of LUDOX, TiIII-Py and the TiIII-Py@LUDOX hybrids (Fig. 3a–d). As in the case of 3-MT, addition of LUDOX to a solution of NMPEA caused the broadening of all the signals of the analyte (Fig. 3d). On the other hand, the addition of the sole TiIII-Py (Fig. 3b) caused the broadening only of the signals arising from the aliphatic residues, as well as small upfield shifts. These effects are known in the case of the formation of the TiIII-Py/*N*-methylated ammonium host-guest complexes, and are ascribed to the shielding effect produced on the analyte by the inclusion in the aromatic cavity of TiIII-Py.¹¹ When the spectrum is recorded in the presence of the TiIII-Py@LUDOX hybrids (Fig. 3c), the trends of broadening and chemical shift perturbations were similar to those observed in the presence of only TiIII-Py, but less pronounced. This confirmed that silica-grafted TiIII-Py was still capable to bind NMPEA.

Binding of NMPEA to TiIII-Py@LUDOX was confirmed as well by a ¹H-NMR titration performed on a sample containing TiIII-Py@LUDOX (0.25 mM TiIII-Py) in D₂O buffered with phosphate at pD = 7.0, providing a binding constant of (3.1 ± 0.5)·



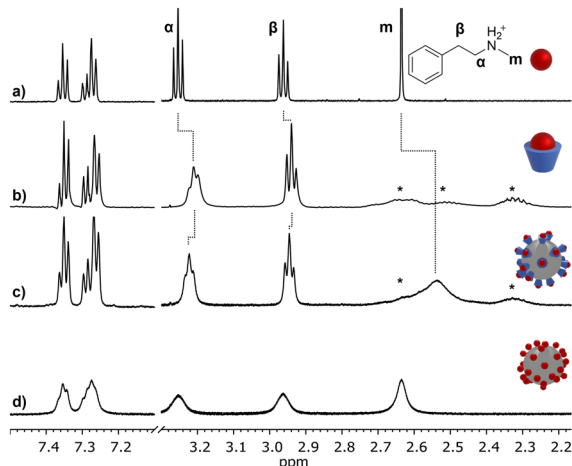


Fig. 3 ^1H -NMR subspectra of: (a) NMPEA (red sphere) 0.5 mM; (b) NMPEA 0.5 mM and Tiii-Py (blue basket) 0.25 mM; (c) NMPEA 0.5 mM, Tiii-Py 0.25 mM and LUDOX (grey sphere) 0.15 μM ; (d) NMPEA 0.5 mM and LUDOX 0.15 μM . Asterisks denote the Tiii-Py signals. Dashed lines indicate the shifts of the signals (Conditions: 500 MHz, 25 $^\circ\text{C}$, phosphate buffer 5 mM, pD = 7.0, D_2O).

10^3 M^{-1} (Fig. S5 and S6, ESI ‡), consistent with the ones previously reported for similar systems in water.¹¹

The ability of the Tiii-Py@LUDOX nanoconjugates to selectively bind *N*-methylammonium moieties, as well as the possibility to employ them in NMR chemosensing, was finally confirmed with saturation transfer difference (STD) NMR experiments on a sample containing 1 mM NMPEA and 0.25 mM Tiii-Py or Tiii-Py@LUDOX in buffered D_2O (phosphate buffer 5 mM, pD = 7.0). This is a steady-state NOE experiment where the spin populations of the analytes interacting with the nanoparticles are altered by continuous irradiation of a selected region of the spectrum (which, in this case, includes only Tiii-Py signals).⁶ As a result, in the spin diffusion limit the intensity of the analytes' signals decreases, and subtraction from an off-resonance reference spectrum reveals only the signals of the interacting analytes. As reported in Fig. 4, no STD signal was observed when the sole Tiii-Py was added to the NMPEA solution (Fig. 4b). This result confirmed that the small size of the Tiii-Py-NMPEA host-guest complexes prevented an effective transfer of saturation. By the contrary, clear STD analyte signals were present (average S/N = 17 for the aromatic region) in the spectra of same sample containing the Tiii-Py@LUDOX nanohybrids (Fig. 4d). The relevant implications of this result are many: (i) they provide another proof of the analyte binding to Tiii-Py@LUDOX since, in the absence of interaction, no STD signal could be observed; (ii) they provide further evidence for the formation of the Tiii-Py@LUDOX hybrids, since the cavitand alone does not produce any STD signal; (iii) they confirm that the formation of Tiii-Py@LUDOX increases the efficiency of the saturation transfer from the cavitand to the analyte. Control experiments performed on samples containing glucose and phenylalanine as potential interferents in the presence of Tiii-Py@LUDOX nanohybrids produced approximately zero STD response (Fig. S7, ESI ‡),

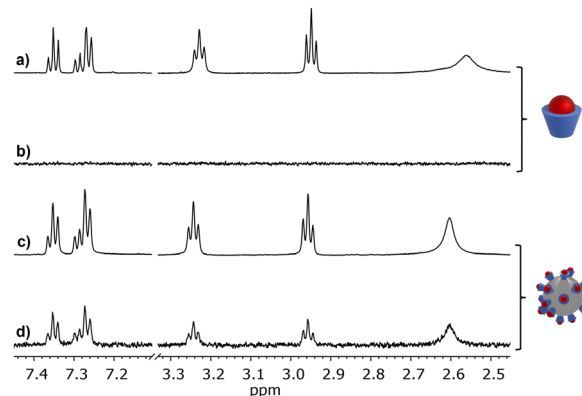


Fig. 4 (a) ^1H -NMR subspectrum of NMPEA (red sphere) 1 mM and Tiii-Py (blue basket) 0.25 mM; (b) STD subspectrum of the same sample; (c) ^1H -NMR subspectrum of NMPEA 0.5 mM, Tiii-Py 0.25 mM and LUDOX (grey sphere) 0.15 μM ; (d) STD subspectrum of the same sample. On resonance saturation was performed at 8.75 ppm (Conditions: 500 MHz, 25 $^\circ\text{C}$, phosphate buffer 5 mM, pD = 7.0, D_2O , 32 scans, acquisition time 5 min).

confirming the role of the cavitand in the selective detection of *N*-methylated amines. The main limit of STD-based detection is that saturation is hardly diffused across interrupted dipole-dipole coupling networks of spins and requires close analyte-receptor contacts. This can substantially affect the effectiveness of chemosensing experiments based on STD if the analyte binding sites include a limited number of hydrogen atoms and/or these belong to different spin system than the one saturated. This limitation is overcome by our recently developed HPwSTD experiment,^{3,4} which uses high power radiofrequency pulses to saturate both the nanohybrid spins and the slow tumbling water molecules belonging to the nanohybrid solvation shell. This ensures a larger source of saturation to be transferred to the analytes and, at the end, greater sensitivity.

To test the performances of the Tiii-Py@LUDOX nanohybrids with the HPwSTD experiment, a mixture of three different amines, namely NMPEA, 3-MT, and 4-nitrophenethylamine (4- NO_2 -PEA), each at 0.25 mM concentration, was added to a suspension of Tiii-Py@LUDOX in buffered D_2O (5 mM phosphate buffer, pD = 7.0). In this case, the selectivity of the sensing system was challenged by the fact that all the analytes are phenethylamines, with the sole NMPEA being *N*-methylated. As expected, no HPwSTD signals from the three analytes were detected in samples devoid of the silica nanoparticles (Fig. 5(a.2)). On the other hand, broad and strong signals from all the analytes were detected in samples containing the sole silica nanoparticles (Fig. 5(a.3)). Indeed, the use of solvation water as saturation source enables also these inorganic particles as NMR nanoreceptors, but they obviously feature small selectivity. It is worth noting that the broadening of NMPEA signals appears more relevant than that of the other analytes. This likely due to the greater affinity of *N*-methylated ammoniums for silica with respect to primary ammonium ions.¹⁸ Still, the affinity difference is not enough to produce relevant differences in the intensities of the STD signals.



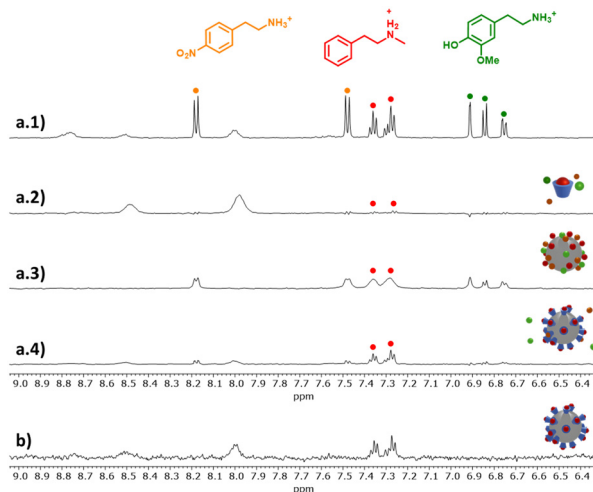


Fig. 5 (a.1) ^1H -NMR subspectrum of a solution of NMPEA (red sphere), 4- NO_2 -PEA (orange sphere), 3-MT (green sphere) 0.25 mM each and Tiiii-Py (blue basket) 0.1 mM; (a.2) HPwSTD subspectrum of a solution of NMPEA, 4- NO_2 -PEA, 3-MT 0.25 mM each and LUDOX (grey sphere) 0.15 μM ; (a.3) HPwSTD subspectrum of a solution of NMPEA, 4- NO_2 -PEA, 3-MT 0.25 mM each and Tiiii-Py 0.1 mM and LUDOX 0.15 μM ; (a.4) HPwSTD subspectrum of a solution of NMPEA, 4- NO_2 -PEA, 3-MT 0.25 mM each, Tiiii-Py 0.1 mM and LUDOX 0.15 μM , 512 scans. (Conditions: 500 MHz, 25 $^\circ\text{C}$, phosphate buffer 5 mM, pH = 7.0, $\text{H}_2\text{O}/\text{D}_2\text{O}$ 90 : 10). Full spectra are reported in Fig. S8, ESI ‡ .

Eventually, the HPwSTD spectra collected from the samples containing both Tiiii-Py and LUDOX (Fig. 5(a.4)) showed the presence of the sole signals of NMPEA, with the other analytes barely detectable.

Hence, the cavitand gives to the Tiiii-Py@LUDOX hybrids the ability to differentiate protonated *N*-methylated amines from protonated primary amines with very similar structure, introducing a remarkable discriminating capability. In addition, the restoration of the hyperfine structure of the signals, with respect to the sample containing only LUDOX, confirms that, notwithstanding the higher affinity for silica of *N*-methylated amines,¹⁸ their direct binding to the nanoparticles is prevented by the surface saturation with the cavitand.

In a final experiment, we verified the possibility to detect *N*-methylated amines at physiologically relevant concentrations. A sample containing 50 μM NMPEA in buffered water (5 mM phosphate buffer, pD = 7.0) was analysed with the HPwSTD protocol. Also in this case, analyte's signals were detected in the spectrum with a S/N value of 9.1 (Fig. 5(b)). Concentration dependent experiments allowed to determine a LOD value of 31 μM (Fig. S9, ESI ‡). Such performance is similar or even better than that of other sensing systems based on the Tiiii-Py cavitands (Table S2, ESI ‡),^{12–16} with the additional bonus of the virtual cancellation of any possibility of false positives due to the recording of the whole analytes' NMR spectra.

In conclusion, we have here demonstrated as the non-covalent adsorption of the Tiiii-Py cavitand to large silica nanoparticles results in an efficient and selective NMR chemosensing systems. This strategy provides a new tool in the field of

NMR spectral editing and chemosensing with nanoparticles. Application of this protocol to the detection of relevant metabolites in biological samples is under investigation.

This work was funded by the Italian Association for Cancer Research (AIRC) under the Investigators Grants scheme (IG 25003). It also benefited from the equipment and framework of the COMP-HUB Initiative, funded by the 'Departments of Excellence' program of the Italian Ministry for Education, University and Research (MIUR, 2018–2022).

Conflicts of interest

There are no conflicts to declare.

Notes and references

- Z. Xu, C. Liu, S. Zhao, S. Chen and Y. Zhao, *Chem. Rev.*, 2019, **119**, 195–230 and references therein.
- (a) Z. Xu, S. Gu, Y. Li, J. Wu and Y. Zhao, *Anal. Chem.*, 2022, **94**, 8285–8292; (b) C. Dong, Z. Xu, L. Wen, S. He, J. Wu, Q.-H. Deng and Y. Zhao, *Anal. Chem.*, 2021, **93**, 2968–2973.
- F. De Biasi, F. Mancin and F. Rastrelli, *Prog. Nucl. Magn. Res. Spectrosc.*, 2020, **117**, 70–88 and references therein.
- F. De Biasi, D. Rosa-Gastaldo, X. Sun, F. Mancin and F. Rastrelli, *J. Am. Chem. Soc.*, 2019, **141**, 4870–4877.
- F. De Biasi, B. B. Mascitti, Ě. Kupĉe and F. Rastrelli, *J. Magn. Res.*, 2022, **338**, 107190.
- (a) M.-V. Salvia, F. Ramadori, S. Springhetti, M. Diez-Castellnou, B. Perrone, F. Rastrelli and F. Mancin, *J. Am. Chem. Soc.*, 2015, **137**, 886–892; (b) L. Gabrielli, D. Rosa-Gastaldo, M. V. Salvia, S. Springhetti, F. Rastrelli and F. Mancin, *Chem. Sci.*, 2018, **9**, 4777–4784.
- X. Sun, L. Riccardi, F. De Biasi, F. Rastrelli, M. De Vivo and F. Mancin, *Angew. Chem., Int. Ed.*, 2019, **58**, 7702–7707.
- D. Neuhaus and M. P. Williamson, *Nuclear Overhauser Effect in Structural and Conformational Analysis*, Wiley-VCH, 2000.
- F. De Biasi, D. Rosa-Gastaldo, F. Mancin and F. Rastrelli, *Chem. Commun.*, 2021, **57**, 3002–3005.
- S.-H. Bae, H. J. Dyson and P. E. Wright, *J. Am. Chem. Soc.*, 2009, **131**, 6814–6821.
- R. Pinalli, G. Brancatelli, A. Pedrini, D. Menozzi, D. Hernández, P. Ballester, S. Geremia and E. Dalcaneale, *J. Am. Chem. Soc.*, 2016, **138**, 8569–8580.
- M. Dionisio, J. M. Schnorr, V. K. Michaelis, R. G. Griffin, T. M. Swager and E. Dalcaneale, *J. Am. Chem. Soc.*, 2012, **134**, 6540–6543.
- G. Valenti, E. Rampazzo, E. Biavardi, E. Villani, G. Fracasso, M. Marcaccio, F. Bertani, D. Ramarli, E. Dalcaneale, F. Paolucci and L. Prodi, *Faraday Discuss.*, 2015, **185**, 299–309.
- E. Biavardi, C. Tudisco, F. Maffei, A. Motta, C. Massera, G. G. Condorelli and E. Dalcaneale, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 2263–2268.
- D. Masseroni, E. Biavardi, D. Genovese, E. Rampazzo, L. Prodi and E. Dalcaneale, *Chem. Commun.*, 2015, **51**, 12799–12802.
- E. Biavardi, S. Federici, C. Tudisco, D. Menozzi, C. Massera, A. Sottini, G. G. Condorelli, P. Bergese and E. Dalcaneale, *Angew. Chem., Int. Ed.*, 2014, **53**, 9183–9188.
- I. R. N. Verly, R. Leen, J. R. Meinsma, G. K. J. Hooijer, C. D. Savci-Heijink, J. van Nes, M. Broekmans, R. J. A. Wanders, A. B. P. van Kuilenburg and G. A. M. Tytgat, *Eur. J. Cancer*, 2019, **111**, 21–29.
- (a) B. Zhang, M. Xie, L. Bruschweiler-Li, K. Bingol and R. Bruschweiler, *Anal. Chem.*, 2015, **87**, 7211–7217; (b) M. Xie and R. Bruschweiler, *J. Phys. Chem. Lett.*, 2020, **11**, 10401–10407.
- T. Bian, A. Gardin, J. Gemen, L. Houben, C. Perego, B. Lee, N. Elad, Z. Chu, G. M. Pavan and R. Klajn, *Nat. Chem.*, 2021, **13**, 940–949.
- G. Pieters, A. Cazzolaro, R. Bonomi and L. J. Prins, *Chem. Commun.*, 2012, **48**, 1916–1918.

