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## Ascorbate-assisted nitric oxide release from photocontrollable nitrosonium ion releasers for potent *ex vivo* photodilation†

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**We found that *N*-nitrosoaminoanisole derivatives tethered to dyes work as photocontrollable nitrosonium cation releasers and are converted to potent nitric oxide releasers in the presence of sodium ascorbate. The *N*-nitrosoaminoanisole derivative **2** worked as a more potent photodilating reagent *ex vivo* than previously reported nitric oxide releasers.**

In mammals, reduced blood flow may lead to circulatory system disease and organ failure,<sup>1</sup> and drugs that improve blood flow by relaxing blood vessels are therefore promising candidates for the treatment of various cardiovascular diseases and stroke. Nitric oxide (NO) is an endogenous signaling molecule that causes a potent but transient relaxation of vessels,<sup>2</sup> so chemicals that enable spatiotemporal control of NO concentration are of great interest. In particular, many studies have focused on photocontrollable NO releasers,<sup>3</sup> but so far, there have been few reports of successful optical control of blood vessel dilation.<sup>4</sup> Our group has developed a series of visible-light-controlled NO releasers, composed of a light-harvesting antenna moiety and an *N*-nitrosoaminophenol moiety for NO release. As shown in Fig. 1a, photoinduced electron transfer (PeT) to the antenna moiety from the *N*-nitrosoaminophenol moiety generates a phenoxyl radical intermediate that releases NO to form a stable quinone. Our design has achieved high cross section ( $\epsilon_{\text{NO}}$ : 365) as a red light controllable NO releaser ( $\lambda$ : 660 nm) and enabled us to regulate vasodilation of *ex vivo* rat aorta, as well as the intracavernous pressure of rats *in vivo* (Table S1, ESI†).<sup>5</sup> Interestingly, during structural optimization of a green-light-controllable NO releaser, we found that the hydroxyl group of NO-Rosa5 (**1**) is necessary for NO release, and furthermore, that

the *O*-methylated compound, NO-Rosa6 (**2**), released nitrosonium ion (NO<sup>+</sup>) instead of NO (Fig. 1c).<sup>5c</sup> A plausible mechanism of NO<sup>+</sup> release from **2** is shown in Fig. 1b. After PeT, the radical cation releases NO<sup>+</sup> via a mesolytic cleavage mechanism.<sup>6</sup> In the present work, we compared the NO- and NO<sup>+</sup>-releasing abilities of **1** and **2**, as well as a red-light-controllable NO releaser, NORD-1 (**3**) and its *O*-methylated derivative, NORD-2 (**4**). Unexpectedly, we found that **2** and **4** afforded NO, but not NO<sup>+</sup>, more efficiently and quickly than **1** and **3** in the presence of 1 equivalent of sodium ascorbate (SA), a biologically ubiquitous reductant. In *ex vivo* assay using rat aorta strips, **2** induced vasodilation more quickly than **1**, and was effective even in the absence of exogenously added ascorbic acid.

We previously found that light irradiation induced little release of NO from **2**, but nevertheless, decomposition of **2** was faster than that of **1**.<sup>5c</sup> Decomposition product analysis revealed that **2** released NO<sup>+</sup> instead of NO *in vitro*. Since NO<sup>+</sup> can directly cause *S*- or *N*-nitrosylation, controlled NO<sup>+</sup> release is expected to be a good approach for biological investigation of nitrosylated proteins and other biological nitrosylated molecules.<sup>7</sup> Thus, we considered that **4**, which is an *O*-methylated derivative of the red-light-controllable NO releaser **3**, might also be a photocontrollable NO<sup>+</sup> releaser based on the structural analogy with **2**. So, we prepared these compounds according to the previous method (see Supporting Information) and confirmed that these compounds efficiently harvest green or red light, respectively (Fig. S1, ESI†).

The NO-releasing ability of these compounds was investigated using an NO electrode. A solution of each compound (10  $\mu\text{M}$ ) was irradiated with green (568 nm) or red (660 nm) LED light and the NO concentration was recorded by using an electrochemical detector equipped with an NO electrode. As shown in Fig. 2a and b, **1** and **3** released NO efficiently, whereas **2** and **4** did not. It was also confirmed that the light irradiation did not induce any amperometric response in the absence of these compounds (Fig. S2, ESI†). The proposed radical intermediates formed in the PeT process were detected by means of electron spin resonance (ESR) spectroscopy. The signal of **2** (Fig. 2d) is assigned to the charge-separated (CS) state generated by intramolecular PeT. The signal of **1** (Fig. 2c) was stronger than that of **2**. This indicates

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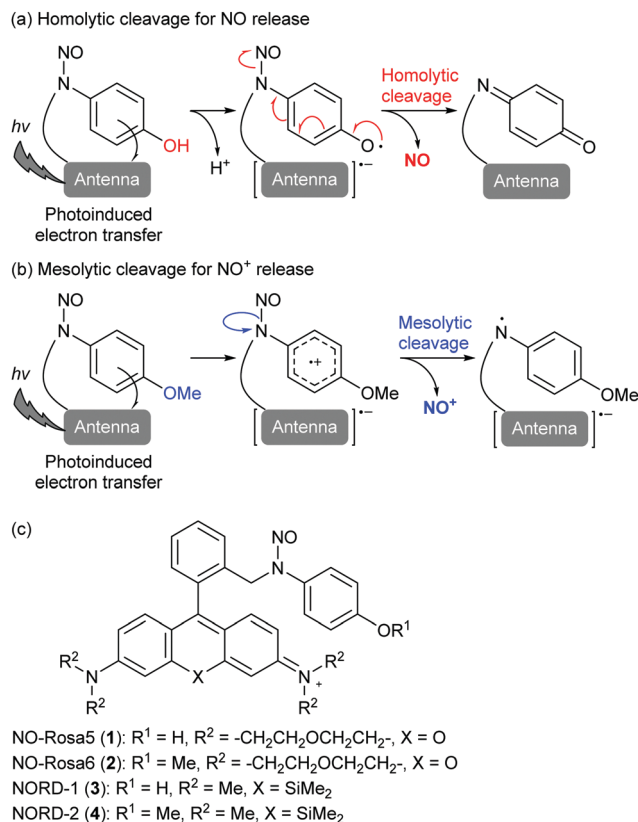


Fig. 1 Plausible mechanisms for photoinduced release of NO (a), and NO<sup>+</sup> (b); (c) structures of NO-Rosa5 (1), NO-Rosa6 (2), NORD-1 (3), and NORD-2 (4).

that the signal of the phenoxyl radical [1(-H<sup>•</sup>)] generated by deprotonation of the donor radical cation moiety overlaps with the signal of the short-lived CS state. The SOMO of 1(-H<sup>•</sup>) consists of the N-N antibonding orbital (Fig. S3m and n, ESI<sup>†</sup>). The NO release occurred by cleavage of the N-N bond of 1(-H<sup>•</sup>) ( $d(\text{N-N}) = 1.357 \text{ \AA}$ ), which is elongated compared to that of 2 (1.333  $\text{\AA}$ ) (Fig. S3a and b, ESI<sup>†</sup>).<sup>8</sup> Similar behaviors were observed in the cases of 3 and 4 (Fig. 2e, f and Fig. S3c, d, ESI<sup>†</sup>). On the other hand, as shown in Fig. S4 (ESI<sup>†</sup>), the absorbance of 2 or 4 during photoirradiation decreased more quickly than that of 1 or 3, respectively. This result suggests that 2 and 4 released a different species, that is NO<sup>+</sup>, and the resulting amino group of the aniline moiety attacked the 9-position of the xanthene moiety to form a colorless closed (spiro) form (Scheme S1a, ESI<sup>†</sup>). In contrast, 1 and 3 released NO, and the resulting semiquinone radical would disproportionate to form a colorless closed compound and a still-colored aldehyde (Scheme S1b, ESI<sup>†</sup>). Next, to investigate the NO<sup>+</sup>-releasing ability of these PeT-driven compounds, 2,3-diaminonaphthalene (DAN) was employed as an NO<sup>+</sup> fluorescence probe.<sup>9</sup> DAN has also been utilized as an alternative fluorescence NO probe to Griess reagent; it reacts with NO<sup>+</sup> derived from NO<sub>2</sub><sup>-</sup> under acidic conditions to form 2,3-naphthotriazole (NAT), which shows strong fluorescence. DAN can be employed for the direct detection of NO<sup>+</sup> without acidic treatment. A solution of each PeT-driven compound (10  $\mu\text{M}$ ) and DAN (10  $\mu\text{M}$ ) was photoirradiated and then the fluorescent products were analyzed by HPLC. As shown in Fig. 2g, h and

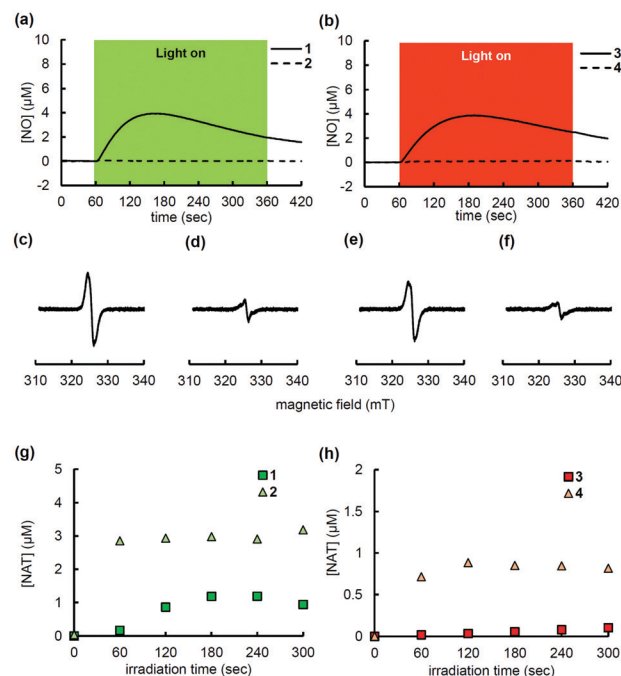
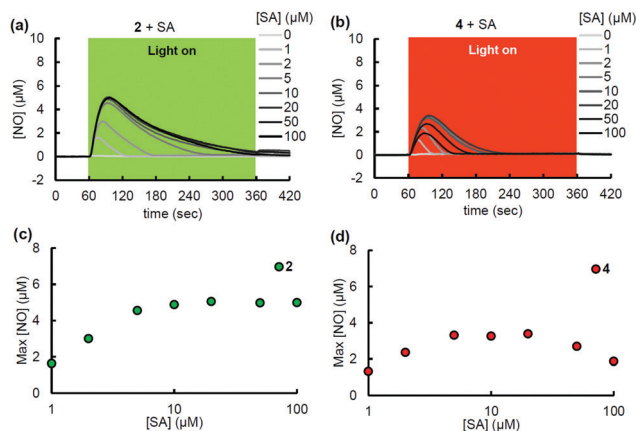


Fig. 2 NO release detected by an NO electrode from a 10  $\mu\text{M}$  solution of 1 or 2 (a), and 3 or 4 (b); ESR spectrum of the charge-separated state in an irradiated 10  $\mu\text{M}$  solution 1 (c), 2 (d), 3 (e) or 4 (f); NO<sup>+</sup> detection using 10  $\mu\text{M}$  2,3-diaminonaphthalene (DAN) in an irradiated solution of 10  $\mu\text{M}$  1 or 2 (g), or 3 or 4 (h).

Fig. S5 (ESI<sup>†</sup>), a larger amount of NAT was found in the irradiated solution of 2 or 4, as compared with 1 or 3, respectively. These results indicated that *O*-methylation converted the PeT-driven NO releasers into photocontrollable NO<sup>+</sup> releasers. In terms of the reaction mechanism, it is assumed that NO<sup>+</sup> release is not due to destabilization of the N-N bond *via* formation of a radical intermediate such as 1 and 3, but rather is due to the strong polarization of the N-N bond caused by radical cation formation after PeT.

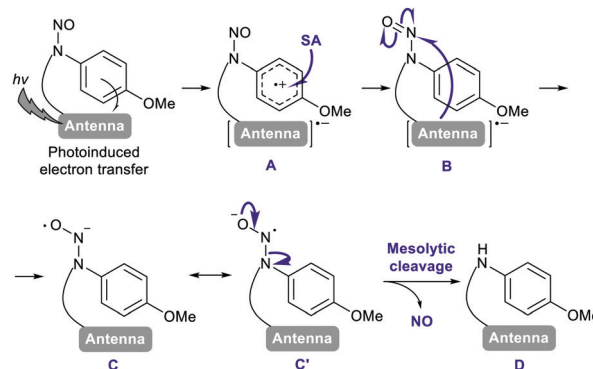
Various redox-active substances exist in the biological milieu, and we next examined the effects of some of them on the photoreactivity of the PeT-driven compounds working *via* radical mechanisms. First, the effect of SA, a major one-electron reductant that exists both inside and outside cells was examined.<sup>10</sup> While SA hardly affected the NO release from 1 and 3, significant NO release was unexpectedly observed from 2 and 4 in the presence of SA (Fig. 3a, b and Fig. S6, ESI<sup>†</sup>). Other biological reductants examined, glutathione (GSH), nicotine adenine dinucleotide (NADH), and tocopherol, did not affect the NO release from PeT-driven compounds (Fig. S7, ESI<sup>†</sup>). The ascorbate-assisted NO release was SA-concentration-dependent, and reached a plateau at 10  $\mu\text{M}$  SA (Fig. 3c and d). At SA concentrations over 10  $\mu\text{M}$ , the maximum NO concentration derived from 2 was unaffected, while that derived from 4 decreased with increasing concentration of ascorbate. The ascorbate-assisted NO release from 2 and 4 seemed quick and efficient, so we determined the quantum yield  $\phi_{\text{NO}}$  using an NO electrode and a chemical actinometer based on ammonium tetrathiocyanatodiamminechromate(III), known as Reinecke's





**Fig. 3** (a and b) Detection of ascorbate-assisted NO release from a solution of each compound (10  $\mu\text{M}$ ) in the presence of the indicated concentration of sodium ascorbate (SA) using an NO electrode; (c and d) plots of maximum [NO] against SA concentration.

salt (Table S2, ESI<sup>†</sup>).<sup>11</sup> As shown in Table S3 (ESI<sup>†</sup>), the  $\Phi_{\text{NO}}$  values of **2** ( $9.84 \times 10^{-3}$ ) and **4** ( $4.97 \times 10^{-3}$ ) in the ascorbate-assisted reaction were greater than those of **1** ( $0.919 \times 10^{-3}$ ) and **3** ( $3.30 \times 10^{-3}$ ), respectively, and SA hardly affected the  $\Phi_{\text{NO}}$  values of **1** and **3**. Notably, the ascorbate-assisted  $\Phi_{\text{NO}}$  of **2** was about ten times larger than that of **1**. To confirm whether  $\text{NO}^+$  reacts directly with SA, nitrosonium tetrafluoroborate ( $\text{NOBF}_4$ ) was added to a solution of SA, and the NO concentration was monitored. As shown in Fig. S8a (ESI<sup>†</sup>), no increase of NO was observed when a solution of  $\text{NOBF}_4$  in MeCN was added to an aqueous solution of SA. Further, no NO increment was observed when a solution of sodium nitrite ( $\text{NaNO}_2$ ), which would be formed after reaction of  $\text{NO}^+$  and  $\text{H}_2\text{O}$ , was added to a solution of SA (Fig. S8b, ESI<sup>†</sup>). These results indicate that SA did not react directly with  $\text{NO}^+$  or  $\text{NO}_2^-$ , but instead reacted with the photoreaction intermediate of **2** or **4**, resulting in the release of NO. The SA addition did not affect either the absorption spectrum or the fluorescence quantum yield  $\Phi_f$  of each compound, suggesting that SA did not interact with the ground or excited state of the antenna moiety but rather with the CS state (Fig. S9, ESI<sup>†</sup>). A plausible mechanism for NO release from **2** and **4** is shown in Fig. 4. After photoinduced electron transfer, the cation radical **A** would undergo one-electron reduction from SA. Then, the radical intermediate **B** of the antenna moiety would mediate one-electron reduction of *N*-nitrosoaniline to form oxyhydrazinyl radical **C** or **C'**. This intermediate **C'** would release NO in a mesolytic cleavage process to form **D**. In the case of the red-light-controllable compound **4**, Si-rhodamine could undergo two-electron reduction more easily than 10-oxa-rhodamine, so that excess SA might induce over-reduction, which would compete with the NO-releasing pathway (Scheme S2, ESI<sup>†</sup>).<sup>12</sup> Further, the nature of the photodecomposition products explains why **2** and **4** showed larger  $\Phi_{\text{NO}}$  values than **1** and **3**: the main photodecomposition products of **1** and **3** are both colorless closed-form products and still-colored aldehydes (Fig. S4b, ESI<sup>†</sup>), whereas those of **2** and **4** are only colorless closed-form products

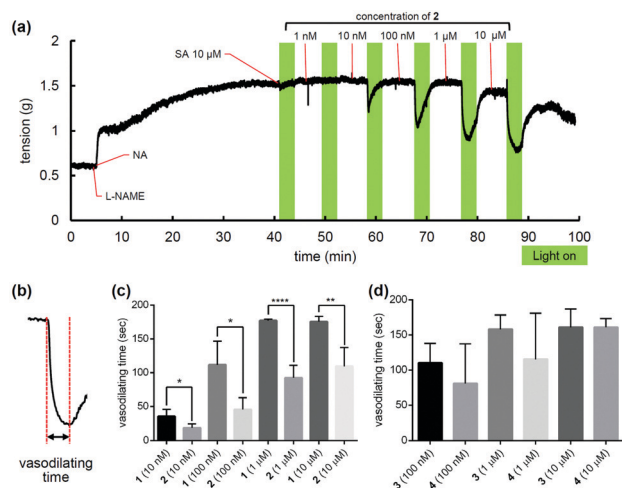


**Fig. 4** A plausible mechanism of ascorbate-assisted NO release from **2** or **4**.

(Scheme S4a, ESI<sup>†</sup>). In other words, inner filter effects could interfere with photoexcitation of remaining **1** and **3**.<sup>13</sup> Indeed, photodecomposition was faster at lower concentrations of **1** or **3**, probably because of a smaller inner filter effect (Fig. S10, ESI<sup>†</sup>).

NO can induce vasodilation of arteries by activating soluble guanylyl cyclase to promote production of cyclic GMP.<sup>2</sup> Therefore, we examined the vasorelaxant activity of these compounds by monitoring their effect on the tension of a rat aorta strip. Since the blood of healthy people contains at least 10  $\mu\text{M}$  SA, the photovasoilation experiments was conducted in the presence of 10  $\mu\text{M}$  SA.<sup>14</sup> The strip was pre-constricted with noradrenaline (NA, 10  $\mu\text{M}$ ), then SA (10  $\mu\text{M}$ ) was added, and 40  $\text{mW cm}^{-2}$  light irradiation was repeated with increasing concentrations of each PeT-driven compound (Fig. 5a). As shown in Fig. S11 (ESI<sup>†</sup>), the maximum vasodilation reached 60–80% of the NA-induced constriction for each compound. The  $\text{EC}_{50}$  values of vasodilation at 40  $\text{mW cm}^{-2}$  light irradiation were calculated to be approximately 10–100 nM. These values are consistent with previous reports that NO causes vasodilation in the 10–100 nM range.<sup>15</sup> The faster NO release from **2** than **1** was reflected in the vasodilating time, which is defined as the time to reach minimum tension in response to NO release (Fig. 5b). As shown in Fig. 5c and d, the vasodilating time of **2** was shorter than that of **1**, while there was no significant difference between those of **3** and **4**. This is probably because the  $\Phi_{\text{NO}}$  of **2** was nearly 10 times higher than that of **1**, while that of **4** was only about 1.3 times than that of **3**. Overall, the *in vitro* finding that SA made **2** and **4** better NO releasers is consistent with the outcomes of the *ex vivo* experiment. We also conducted a similar experiment using **2** without addition of SA, and **2** still showed potent vasodilation, as shown in Fig. S12 (ESI<sup>†</sup>). This can be explained by the presence of endogenous ascorbate in the cells of the aorta. These results indicate that the PeT-driven  $\text{NO}^+$  releasers could act as efficient photovaso dilators even without exogenous addition of SA. To confirm the mechanism of this vasodilation of **2**, we added a sGC inhibitor, ODQ, and confirmed that ODQ inhibited vasorelaxation induced by **2**. Importantly, the addition of a solution of  $\text{NaNO}_2$ , the final decomposition product of  $\text{NO}^+$ , induced no vasodilation at 10  $\mu\text{M}$ , and had little effect even at 100  $\mu\text{M}$





**Fig. 5** (a) Changes in the tension of rat aorta *ex vivo* induced by green-light-mediated NO release from **2** in the presence of a nitric oxide synthetase inhibitor, *N*<sup>G</sup>-nitro-L-arginine methyl ester (L-NAME, 10 μM). Rat aorta in a glass tube was treated with noradrenaline (NA, 10 μM), and sodium ascorbate (SA, 10 μM) followed by **2**. The tube was irradiated with a 568 nm LED (40 mW cm<sup>-2</sup>) for 3 min at each concentration; (b) definition of vasodilating time; (c and d) vasodilating time of each compound at each concentration. Data are expressed as mean ± SE (shown as error bars, *n* = 4). Statistical significance was examined by application of Student's *t*-test. \**p* < 0.05, \*\**p* < 0.01, and \*\*\*\**p* < 0.001.

(Fig. S13, ESI<sup>†</sup>), suggesting that NO, but not NO<sub>2</sub><sup>-</sup>, derived from **2** induced sGC-mediated vasodilation. **1** and **2** showed no significant cytotoxicity at 10 μM, though **3** and **4** suppressed cell proliferation, likely due to the lipophilicity of the dye (Fig. S14, ESI<sup>†</sup>).

We found that *O*-methylation of PeT-driven NO releasers converts them into NO<sup>+</sup> releasers, and also that these NO<sup>+</sup> releasers rapidly and efficiently release NO in the presence of SA at physiological concentrations. The NO release efficiency,  $\Phi_{\text{NO}}$ , of the *O*-methylated compounds was greater than those of previously reported NO releasers. Our results indicate that the higher  $\Phi_{\text{NO}}$  is due to a reduced inner filter effect. This finding could be helpful to improve the photoreaction efficiency in the development of other photoresponsive compounds. In an *ex vivo* photovasodilation test, these compounds could induce 60–80% of maximum vasodilation in response to light. The EC<sub>50</sub> values were in the 10–100 nM range. The photovasodilation of **2** was faster than that of **1**, and **2** was active even in the absence of additional (non-endogenous) SA. The structure-activity relationship data obtained in this study should be helpful in the development of compounds to control blood flow *in vivo*.

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## Conflicts of interest

There are no conflicts to declare.

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