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CORRECTION

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Correction: A testis-derived macroporous 3D scaffold as a platform for the generation of mouse testicular organoids

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Correction for 'A testis-derived macroporous 3D scaffold as a platform for the generation of mouse testicular organoids' by Tohid Rezaei Topraggaleh *et al.*, Biomater. Sci., 2019, **7**, 1422–1436, DOI: 10.1039/C8BM01001C.

We regret to inform the reader that an error related to data handling/labelling during the preparation of the representative images in Fig. 2C and E (control) and negative control of vimentin and Ki67 in Fig. 7 resulted in incorrect figure panel placements in the original article. We therefore replace the Fig. 2E (control) and Fig. 7 (negative control for Ki67) images with the correct images. The corrected Fig. 2 and 7 are shown below.

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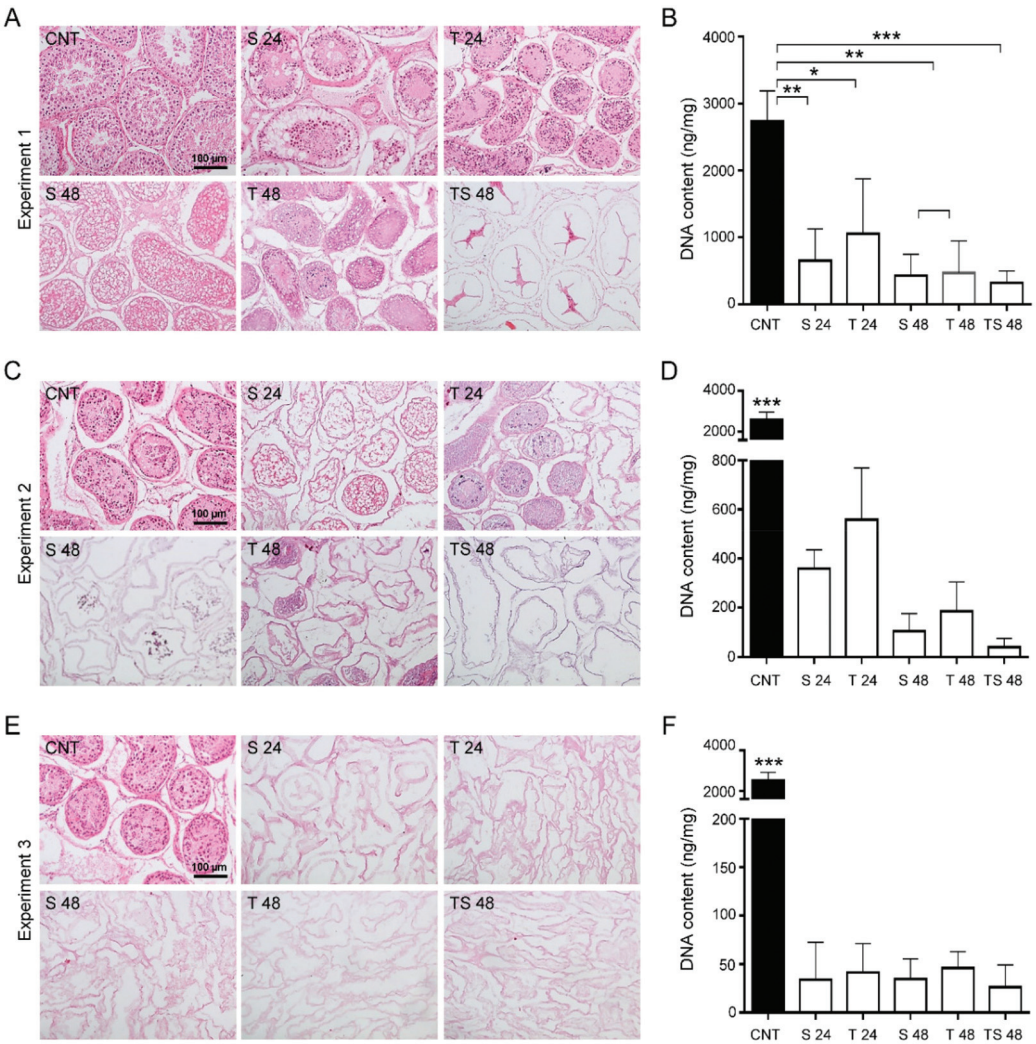

Fig. 2 Histological analysis and quantification of DNA content in decellularized testicular fragments. Experiment 1: Fresh testicular fragments were decellularized by a solution containing 1% (w/v) SDS and 1% (v/v) Triton X-100 in PBS with five experimental groups. (I) S 24; SDS for 24 h, (II) T 24; Triton X-100 for 24 h, (III) S 48; SDS for 48 h, (IV) T 48; Triton X-100 for 48 h, (V) TS 48; Triton X-100 for 24 h and SDS for another 24 h. In all groups, the process was followed by washing the samples with PBS for 24 h. Experiment 2: Testicular fragments were frozen before the decellularization procedure. All of the five decellularization protocols were similar to experiment 1. Experiment 3: In this experiment the detergents were dissolved in distilled water (instead of PBS). All of the protocols were similar to those in experiment 2. H&E staining of the native testicular tissue (CNT) and DTFs of experiment 1 (A), experiment 2 (C), and experiment 3 (E). The DNA quantification of samples in experiment 1, 2, and 3 (B, D, and E respectively) showed that the DNA contents of TS 48 (experiment 2) and all of the protocols in experiment 3 were less than 50 ng per mg of dry weight. Data are expressed as the mean ± SD, and include 3 biological replicates, $*P < 0.05$, $**P < 0.01$, and $***P < 0.001$.

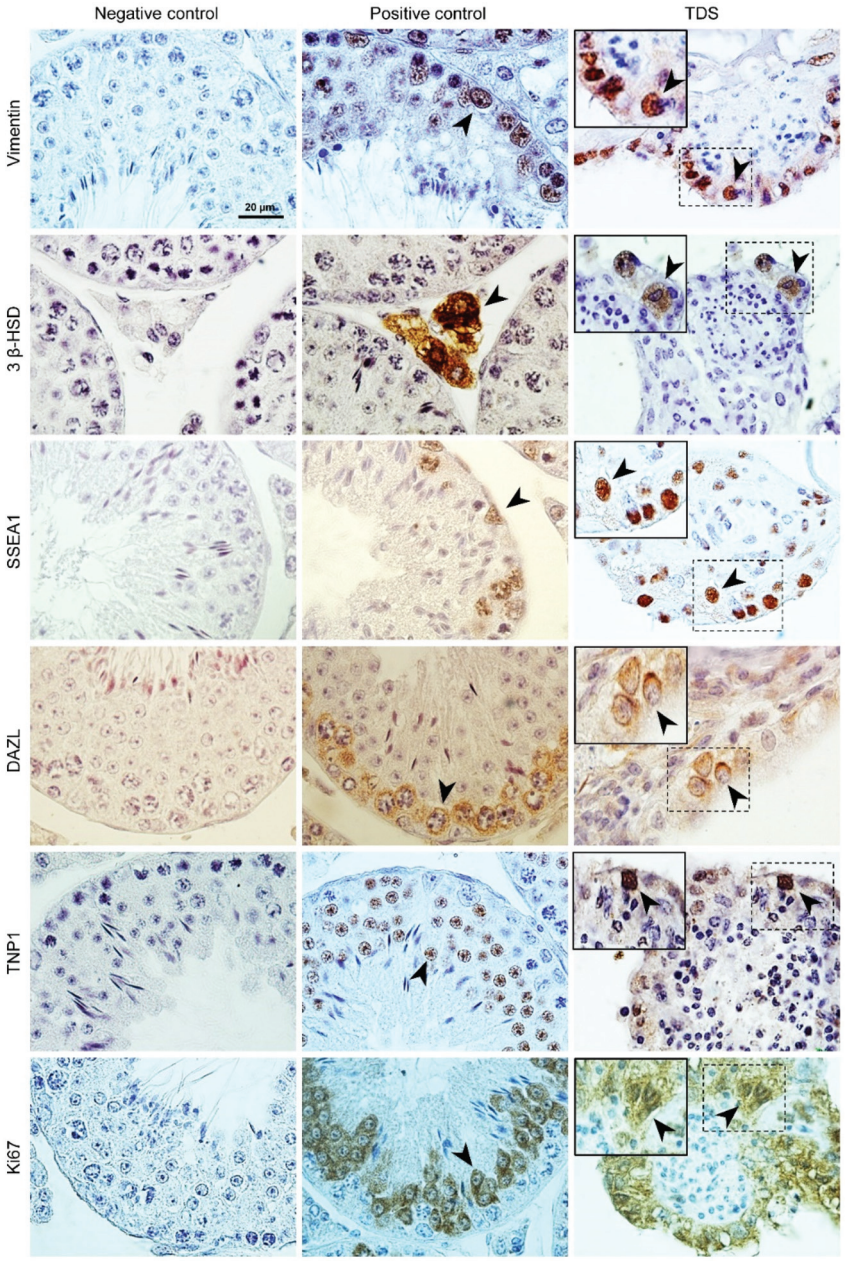

Fig. 7 Immunohistochemistry analysis of TOs. The immunohistochemical analysis of vimentin as a Sertoli cell marker showed the localization of vimentin-positive cells in the periphery of TOs. The expression of 3-beta-hydroxy dehydrogenase (3-β-HSD) as the Leydig cell marker was similar to that of Sertoli cells. The staining of stage-specific embryonic antigen 1 (SSEA1) was applied for the detection of spermatogonial cells in TOs. Numerous SSEA1 positive cells were randomly distributed in TOs. The expression of deleted in azoospermia-like (DAZL) protein as a marker of the early-stage development of spermatogonia was characterized in the periphery of TOs. Post-meiotic marker of transition protein 1 (TNP1) expression was identified in TOs after 30 days of the culture. The expression of Ki67 in the periphery of TOs indicated the proliferative activity of testicular cells in TOs following 30 days of the culture. For all markers, mature mouse testicular tissue was utilized as a positive control and the same staining procedure without primary antibodies was applied for negative control.

The Royal Society of Chemistry apologises for these errors and any consequent inconvenience to authors and readers