CORRECTION

[View Article Online](#)
[View Journal](#) | [View Issue](#)

Cite this: *Biomater. Sci.*, 2022, **10**, 1393

Correction: Antimicrobial peptide modification enhances the gene delivery and bactericidal efficiency of gold nanoparticles for accelerating diabetic wound healing

Song Wang,^a Chang Yan,^b Ximu Zhang,^c Dezhi Shi,^d Luxiang Chi,^{*b} Gaoxing Luo^{*a} and Jun Deng^{*a}

DOI: 10.1039/d2bm90013k
rsc.li/biomaterials-science

Correction for 'Antimicrobial peptide modification enhances the gene delivery and bactericidal efficiency of gold nanoparticles for accelerating diabetic wound healing' by Song Wang et al., *Biomater. Sci.*, 2018, **6**, 2757–2772, DOI: 10.1039/C8BM00807H.

The authors regret mistakes in Fig. 8 and in Fig. 9.

In Fig. 8a, panels a2 and a3 have partial overlap.

In Fig. 9e, the PBS treated panel is identical to the LL37 treated panel in Fig. 5c.

The correct figures are shown below:

^aInstitute of Burn Research, Southwest Hospital, State Key Lab of Trauma, Burn and Combined Injury, Third Military Medical University (Army Medical University), Chongqing 400038, China. E-mail: logxw@yahoo.com, djun.123@163.com

^bDepartment of Cardiology, Southwest Hospital, Third Military Medical University (Army Medical University), Chongqing 400038, China. E-mail: chi68754271@126.com

^cChongqing Key Laboratory of Oral Disease and Biomedical Sciences & Chongqing Municipal Key Laboratory of Oral Biomedical Engineering of Higher Education & Stomatological Hospital of Chongqing Medical University, Chongqing 401174, China

^dKey Laboratory of Three Gorges Reservoir Region's Eco-Environment, Ministry of Education, Faculty of Urban Construction and Environmental Engineering, Chongqing University, Chongqing 40005, PR China

Fig. 8 (A) Representative IHC-stained sections of granulation tissues for CD31. Scale bar: 50 μm . (B) Expression levels of VEGF in skin wounds treated with PBS, naked pDNAs, AuNPs/pDNAs, LL37, AuNPs@LL37 and AuNPs@LL37/pDNAs on the 10th day after injury ($n = 5$). (C) Quantitative analysis of the CD31⁺ microvessels. * and ** denote significant difference (compared with control groups) at $p < 0.05$ and $p < 0.01$ levels, respectively.

Fig. 9 Effects of AuNPs@LL37/pDNA on MRSA-infected diabetic wounds. (A) Treatment schedule for MRSA infected diabetic wounds. (B) Representative macroscopic images of wounds treated with PBS, vancomycin and AuNPs@LL37/pDNAs. The diameter of the green circle is 4 mm. Quantitative analysis of (C) wound closure rates ($n = 5$) and (D) the time to wound closure, defined as a wound area reduction of >50% ($n = 5$). (E) Images and (F) quantitative analysis of CFU from groups treated with PBS, vancomycin and AuNPs@LL37/pDNA. (G) H&E-stained sections of samples from wounds treated with PBS, vancomycin and AuNPs@LL37/pDNAs (14th day after injury). Scale bar: 200 μm (low magnification); 50 μm (high magnification). Quantitative analysis of (H) the infiltrated inflammatory cells ($n = 5$) and (I) the neocapillaries in wounds ($n = 5$). * and ** denote significant difference (compared with control groups) at $p < 0.05$ and $p < 0.01$ levels.

An independent expert has viewed the corrected images and has concluded that they are consistent with the discussions and conclusions presented.

The Royal Society of Chemistry apologises for these errors and any consequent inconvenience to authors and readers.