

Cite this: *Anal. Methods*, 2022, 14, 843

A novel electrochemical sandwich-like immunosensor based on carboxyl $\text{Ti}_3\text{C}_2\text{T}_x$ MXene and rhodamine b/gold/reduced graphene oxide for *Listeria monocytogenes*

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Listeria monocytogenes (LM) is one of the most common food-borne pathogens and can induce a series of diseases with a high mortality rate to humans; hence, it is very necessary to develop a highly sensitive method for LM detection. Based on this need, a new sandwich-like electrochemical immunosensing platform was developed herein by preparing carboxyl $\text{Ti}_3\text{C}_2\text{T}_x$ MXene (C- $\text{Ti}_3\text{C}_2\text{T}_x$ MXene) as the sensing platform and rhodamine b/gold/reduced graphene oxide (RhB/Au/RGO) as the signal amplifier. The high conductivity and large surface area of C- $\text{Ti}_3\text{C}_2\text{T}_x$ MXene make it a desirable nanomaterial to fix the primary antibody of LM (PAb), while the prepared Au/RGO/RhB nanohybrid is dedicated to assembling the secondary antibody (SAb) of LM, offering an amplified response signal. Through the use of RhB molecule as the signal probe, the experiments showed that the peak currents of RhB increase along with an increase in the concentration of LM from 10 to 10^5 CFU mL^{-1} , and an extremely low limit of detection (2 CFU mL^{-1}) was obtained on the basis of the proposed immunosensing platform after optimizing various conditions. Hence, it is confirmed that the developed sandwich-like immunosensor based on C- $\text{Ti}_3\text{C}_2\text{T}_x$ MXene and RhB/Au/Gr has great application in the detection of LM and other analytes.

Received 30th November 2021
Accepted 20th January 2022

DOI: 10.1039/d1ay02029c

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1. Introduction

As one of the most common food-borne pathogens, *Listeria monocytogenes* (LM) can induce various serious diseases (e.g. meningitis, febrile gastroenteritis, abortion and septicemia) to human health.^{1–3} LM can survive easily in a series of foods including water, cheese, milk, meats and raw vegetables.^{4,5} In the USA, a zero-tolerance policy was formed for the presence of LM in food; in Canada, only 100 CFU g^{-1} was allowed.^{6,7} Consequently, developing effective methods to achieve sensitive detection of LM is considerably important. Up to now, several conventional methods (e.g. dynamic light scattering immunoassay, lateral-flow enzyme immunoconcentration, enzyme-linked immunosorbent assay and quartz crystal microbalance immunosensors) have been developed for LM detection.^{4,8–11} There is no doubt that these methods are very important in detecting LM, but they suffer from drawbacks which include time-consuming process, expensive equipment or complex operation. Compared to the general methods, electrochemical technology has many

incomparable merits since it is very simple, low-cost, easy to operate, and time-saving.¹² In the last few years, there have been some interesting electrochemical sensing methods proposed to detect LM,^{13–15} while developing a sensing platform with higher performance is still a major challenge.

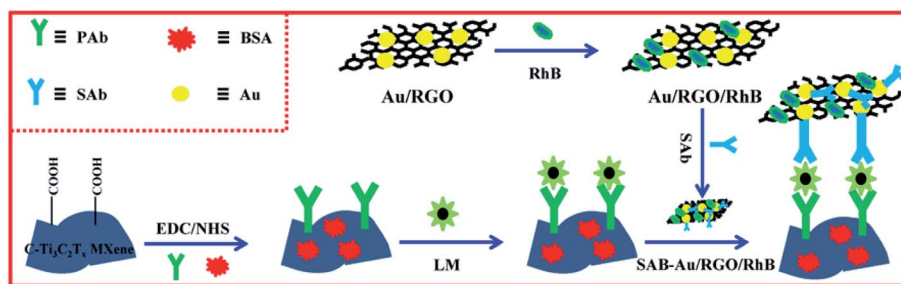
Resulting from the superior signal amplification feature, which causes improvement in sensitivity, the sandwich-like electrochemical immunosensing strategy has aroused considerable attention in recent years.^{16–18} As one of the crucial factors, designing suitable materials as sensing platform, which is dedicated to immobilize antibodies coupled with improved electron transfer, is of great significance.^{19,20} MXenes, known as transition metal carbides/carbonitrides, are an emerging representative 2D nanomaterials in recent years due to their excellent properties.^{21–23} MXenes were discovered *via* exfoliating the “A” laminar component from the MAX phase, which represents the corresponding components in compounds, where “M” reflects the transition metal, “A” refers to the elements in group IIIA/IVA, and “X” is assigned to C/N.^{24–29} The high electrical conductivity and large surface area of MXenes make MXenes-based nanomaterials have important applications in many fields.^{30–32} In fact, presently MXenes have been applied widely in energy storage and conversion, sensors and biomedicine,^{33–36} but there are only a few reports focused on their application in electrochemical immunosensors.

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Scheme 1 Illustrations for constructing the sandwich-like immunosensor of LM based on C-Ti₃C₂T_x MXene and RhB/Au/RGO.

Another key factor in the sandwich-like immunosensor is designing a desirable nanostructure as signal amplifier, which is also very important for improving sensing sensitivity. As another star material, reduced graphene oxide (RGO) possesses many excellent advantages such as large surface area, high electrical conductivity and chemical stability; moreover, RGO can easily be functionalized.^{37–40} Presently, various RGO-based nanostructures have been designed as signal amplifiers to construct electrochemical immunosensors.^{41,42} For instance, Wang *et al.*⁴³ prepared horseradish peroxidase (HRP)/gold nanoparticles (Au) on TiO₂/graphene as signal amplifier for constructing an immunosensor of carcinoembryonic antigen based on the electrocatalytic current of HRP/H₂O₂ toward hydroquinone reduction. In addition, similar ways for signal amplification based on RGO and electrocatalysis of HRP were also developed for squamous cell carcinoma antigen,⁴⁴ prostate specific antigen⁴⁵ and cardiac troponin I.⁴⁶ Obviously, the preparation methods and sensing mechanisms of these signal amplifiers are relatively complex.

Herein, carboxyl Ti₃C₂T_x MXene (C-Ti₃C₂T_x MXene) and gold/RGO/rhodamine b (Au/RGO/RhB) nanostructures were prepared respectively as sensing platform and signal amplifier. By functionalizing Ti₃C₂T_x MXene with carboxyl, the formed C-Ti₃C₂T_x MXene can immobilize LM primary antibody (PAb) *via* a simple amidation reaction to capture LM targets. As for Au/RGO/RhB, RhB was non-covalently functionalized on the Au/RGO surface *via* π - π stacking to be used as a desirable probe since it exhibits sensitive electrochemical oxidation current, and the secondary antibody (SAB) was assembled on the Au surface. On the basis of the above principle, a new sandwich-like electrochemical immunosensing platform was constructed for detecting LM (Scheme 1) coupled with an excellent sensing performance under the optimized conditions.

2. Experimental section

2.1 Preparation of C-Ti₃C₂T_x MXene

Ti₃C₂T_x MXene was first prepared by etching the aluminum layer from Ti₃AlC₂ according to a previous method.⁴⁷ Briefly, 1.0 g Ti₃AlC₂ powder was added slowly to 20.0 mL HF solution under stirring at ~35 °C and maintained for ~24 h. After

centrifuging, washing and drying of the precipitate, Ti₃C₂T_x MXene was obtained successfully.

Next, C-Ti₃C₂T_x MXene was prepared according to a previous work with some modifications.⁴⁸ In brief, 0.16 g 4-aminobenzoic acid and 0.14 g sodium hydroxide were first added to 50.0 mL water (~2 °C), and then 0.26 g sodium nitrite (7.6 mM) was added slowly. Subsequently, 6.0 mL HCl solution was quickly added to this solution under stirring and kept for ~40 min. Then, the desired diazonium salt solution was added to 20.0 mL of Ti₃C₂T_x solution (5.0 mg mL⁻¹) and allowed to stir for ~5 hours. Finally, the solution was centrifuged, washed and dried to obtain the C-Ti₃C₂T_x MXene product.

2.2 Preparation of Au/RGO/RhB

Before the preparation of Au/RGO/RhB, Au/RGO was prepared *via* a common method.^{49,50} For preparing RhB/Au/RGO, 20.0 mL of Au/RGO dispersion (0.5 mg mL⁻¹) was mixed with 20.0 mL of RhB solution (0.25 mg mL⁻¹) and stirred continuously for ~30 min. Owing to the π - π stacking between RhB and RGO, the RhB molecules were functionalized non-covalently on the RGO surface. After centrifugation, rinsing and vacuum-drying, the Au/RGO/RhB product was obtained successfully.

2.3 Construction of the sandwich-like immunosensor

SAb (30.0 μ L) was added to 4.0 mL Au/RGO/RhB dispersion (0.5 mg mL⁻¹) and incubated at room temperature for ~2 hour to assemble SAb on Au surface, forming SAB-Au/RGO/RhB. Next, 10.0 μ L C-Ti₃C₂T_x MXene suspension of (0.5 mg mL⁻¹) was dropped onto a cleaned glassy carbon electrode (GCE) and a 1 : 1 mixture of 1-ethyl-3-[3-dimethylaminopropyl] carbodiimide hydrochloride (0.1 M) with *N*-hydroxysuccinimide mixture (0.2 M) was used to activate the carboxyl group for 30 min. Sequentially, PAb solution (8.0 μ L) was dropped and assembled on the activated C-Ti₃C₂T_x MXene/GCE surface, denoting the formed electrode as PAb/Ti₃C₂T_x MXene/GCE. Phosphate buffer solution (PBS) was then used to rinse PAb/Ti₃C₂T_x MXene/GCE to remove the unbound PAb, and 4.0 μ L BSA (0.1 g mL⁻¹) was used to prevent nonspecific adsorption by incubating PAb/Ti₃C₂T_x MXene/GCE with BSA to form BSA/PAb/Ti₃C₂T_x MXene/GCE. For LA sensing, the constructed BSA/PAb/Ti₃C₂T_x MXene/GCE reacted with LM at different concentrations and dipped in PBS to eliminate nonspecific LM cell.



Finally, LM/BSA/PAb/Ti₃C₂T_x MXene/GCE was combined with SAb-Au/RGO/RhB to produce a sandwich-like structure, which was then measured by an electrochemical method.

3. Results and discussion

3.1 Characterization of C-Ti₃C₂T_x MXene and Au/RGO/RhB

Fig. 1 shows the scanning electron microscopy (SEM) images of Ti₃AlC₂ MAX phase precursor and Ti₃C₂T_x MXene. It is noted that the pristine Ti₃AlC₂ particles exhibit densely aligned layered structures with smooth surface (Fig. 1A). After etching with HF, the produced Ti₃C₂T_x layered nanostructures become very evident and exhibit loosely packed accordion-like morphology compared to the initial Ti₃AlC₂ sample (Fig. 1B).

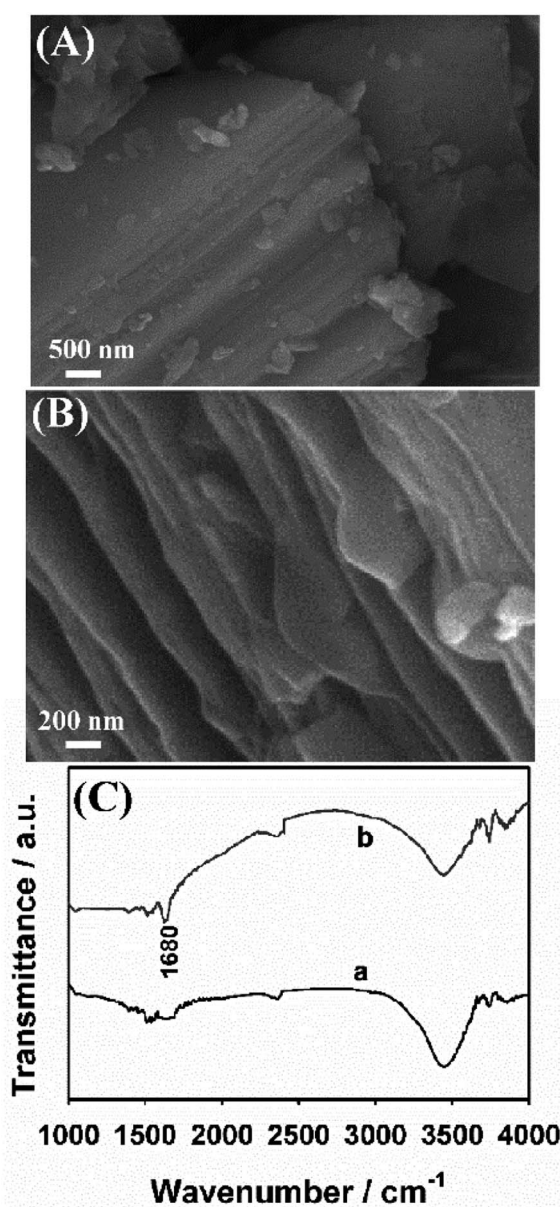


Fig. 1 The SEM images of (A) Ti₃AlC₂ and (B) Ti₃C₂T_x; (C) FT-IR spectra of Ti₃C₂T_x MXene (a) and C-Ti₃C₂T_x MXene (b).

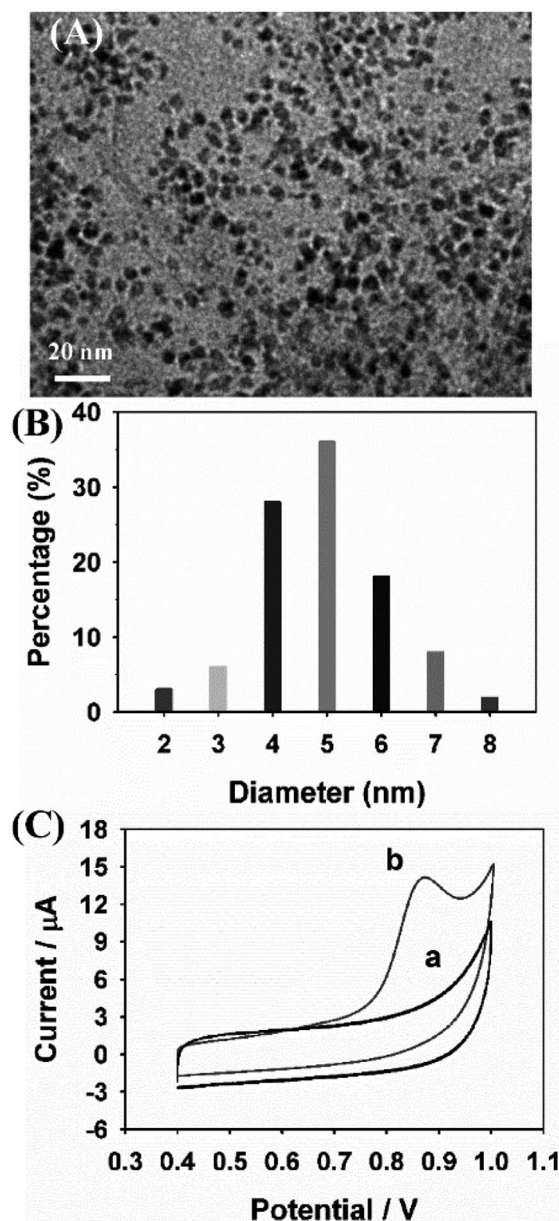


Fig. 2 (A) The TEM image of Au/RGO hybrid, (B) the particle size distribution of AuNPs, and (C) the CV responses of Au/RGO (a) and Au/RGO/RhB (b).

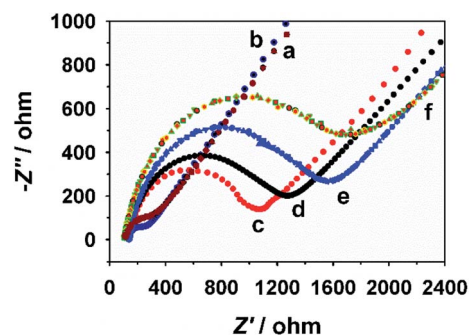


Fig. 3 The EIS plots of GCE (a), C-Ti₃C₂T_x MXene/GCE (b), PAb/Ti₃C₂T_x MXene/GCE (c), BSA/PAb/Ti₃C₂T_x MXene/GCE (d), LM/BSA/PAb/Ti₃C₂T_x MXene/GCE (e), and SAb-Au/RGO/RhB/LM/BSA/PAb/Ti₃C₂T_x MXene/GCE (f).



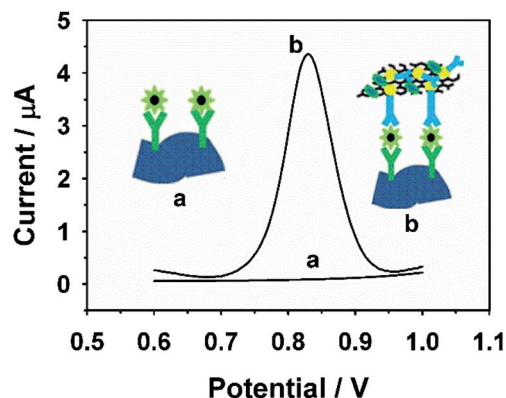


Fig. 4 The DPV response of LM/BSA/PAb/Ti₃C₂T_x MXene/GCE (a) and SAb-Au/RGO/RhB/LM/BSA/PAb/Ti₃C₂T_x MXene/GCE (b) in 0.1 M PBS.

Fourier transform infrared (FT-IR) spectroscopy was used to characterize the formation of C-Ti₃C₂T_x MXene (Fig. 1C). Compared with Ti₃C₂T_x MXene, C-Ti₃C₂T_x MXene possesses a typical and strong characteristic peak of carboxyl presented at ~1680 cm⁻¹.

Fig. 2A shows the transmission electron microscopy (TEM) images of Au/RGO hybrids; it was found that Au nanoparticles were distributed uniformly on the RGO surface with an average diameter of ~5.0 nm (Fig. 2B), and ICP-AES demonstrated that the content of Au is ~5.7%. The abundant and uniform Au present on the Au/RGO surface is very important to assemble SAb and improve the sensing capability. The functionalization

of RhB towards Au/RGO can be easily demonstrated by cyclic voltammetry (CV). As shown in Fig. 2C, there is no peak current present between 0.4 and 1.0 V, while Au/RGO/RhB shows a remarkable oxidation peak current at ~0.87 V since RhB is an electroactive molecule.

3.2 Electrochemical evaluation of the sandwich-like immunosensor

Firstly, electrochemical impedance spectroscopy (EIS) was introduced to evaluate the procedures in constructing the immunosensing platform, which was performed in a 5.0 mM [Fe(CN)₆]^{3-/4-} solution (Fig. 3). Generally, biomolecules including antibody, LM cells and BSA are non-conductive and could restrain electron channel to increase the R_{CT} values. As shown in Fig. 3, the charge-transfer resistance (R_{CT}) at C-Ti₃C₂T_x MXene/GCE is smaller than that of bare GCE due to the high conductivity of Ti₃C₂T_x MXene. When C-Ti₃C₂T_x MXene/GCE was assembled successively with PAB, LM and SAb-Au/RGO/RhB/LM, the R_{CT} values at PAb/Ti₃C₂T_x MXene/GCE, BSA/PAb/Ti₃C₂T_x MXene/GCE, LM/BSA/PAb/Ti₃C₂T_x MXene/GCE and SAb-Au/RGO/RhB/LM/BSA/PAb/Ti₃C₂T_x MXene/GCE in turn show an increase. These results indicate that the developed immunosensing layers were fabricated successfully.

The feasibility of the proposed immunosensor was studied by differential pulse voltammetry (DPV) technique. As shown in Fig. 4 (LM concentration, 10⁶ CFU mL⁻¹), there is no peak current observed at LM/BSA/PAb/Ti₃C₂T_x MXene/GCE in 0.1 M

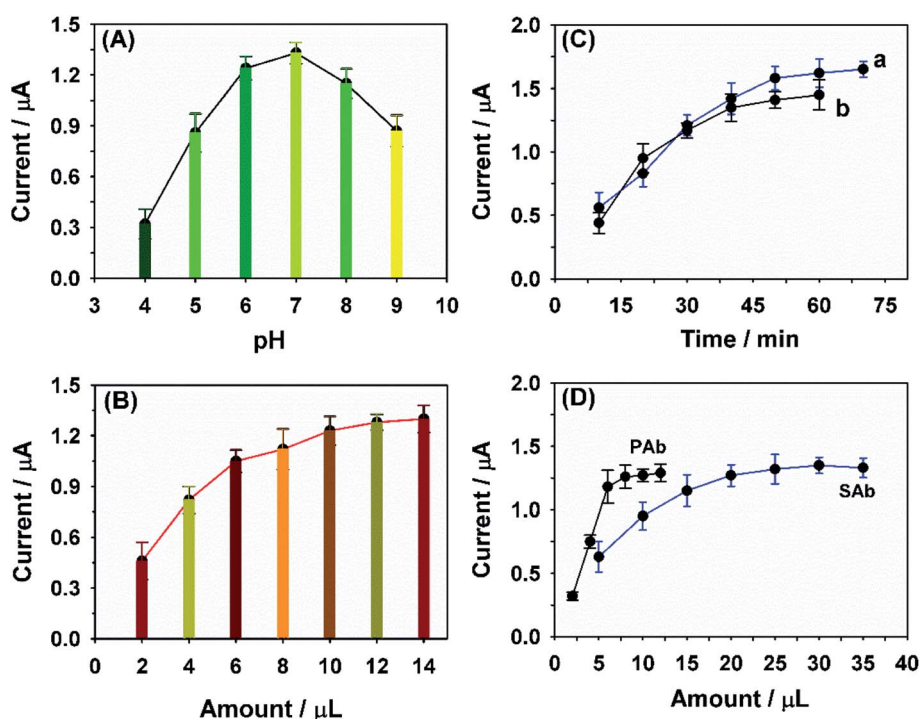


Fig. 5 Influence of the (A) pH value of PBS; (B) amount of Ti₃C₂T_x MXene; (C) reaction time of LM with BSA/PAb/Ti₃C₂T_x MXene/GCE (a) and LM/BSA/PAb/Ti₃C₂T_x MXene with SAb-Au/RGO/RhB (b); (D) amount of PAb and SAb. The specific values of the error bar for (A), (B), (C) and (D) are 0.1 ± 0.03, 0.17 ± 0.04, 0.09 ± 0.03 and 0.08 ± 0.03, respectively.



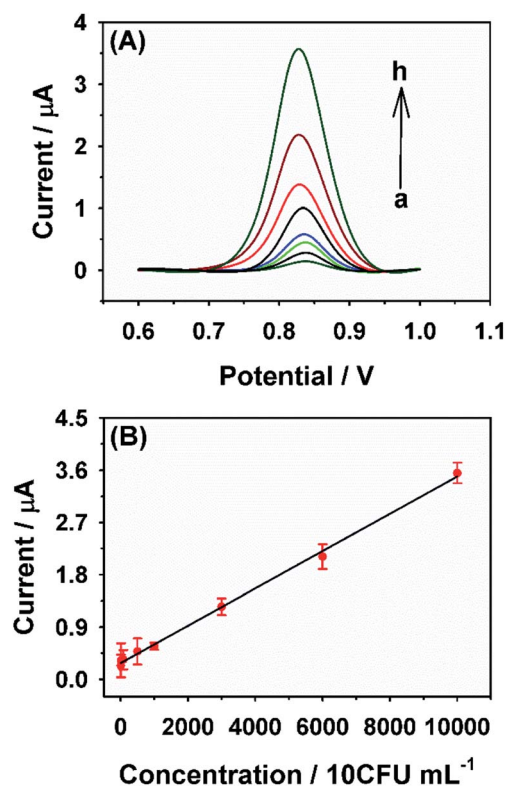


Fig. 6 (A) The DPV responses of LM with various concentrations in 0.1 M PBS (from a to h: 10, 100, 800, 5×10^3 , 10^4 , 3×10^4 , 6×10^4 and 10^5 CFU mL^{-1}) based on the developed immunosensor; (B) the calibration plot of LM. The specific value of the error bar for (B) is 0.18 ± 0.06 .

PBS (pH 7.0). As for SAb-Au/RGO/RhB/LM/BSA/PAb/ $\text{Ti}_3\text{C}_2\text{T}_x$ MXene/GCE, it exhibits an obvious peak current of RhB due to the capture of SAb-Au/RGO/RhB resulting from the antigen-antibody reaction, indicating that the developed immunosensing method is feasible for LM detection. In order to achieve high sensitivity in sensing LM, a series of controlled experiments were performed.

In general, excessive acidity or alkalinity could enable various proteins to become unstable, thus the effect from PBS needs to be evaluated. As shown in Fig. 5A, the DPV peak currents of SAb-Au/RGO/RhB/LM/BSA/PAb/ $\text{Ti}_3\text{C}_2\text{T}_x$ MXene/GCE increase along with the increase in pH value from 4.0, and the maximum peak current presents at pH value of 7.0, thus PBS with the value of 7.0 was selected for all the experiments. Meanwhile, the $\text{C-Ti}_3\text{C}_2\text{T}_x$ MXene amount coated on the GCE

surface is important for the sensing capability, and Fig. 5B shows that the peak currents of RhB increase along with the increase in the amount from 2.0 to 10.0 μL . When the used amount exceeds 10.0 μL , there is little change in the RhB peak currents, indicating that 10.0 μL $\text{C-Ti}_3\text{C}_2\text{T}_x$ MXene dispersion was enough. Furthermore, the incubation time between LM with different antibodies was studied. As shown in Fig. 5C, the studies demonstrated that the optimum length for reacting LM with PAb was 50 min, while the time for reacting LM with SAb was 40 min. The concentration of LM used for the controlled experiments was 3×10^4 CFU mL^{-1} . In addition, the amounts of PAb and SAb used in the construction of the immunosensor were also studied, and it can be observed from Fig. 5D that the best amounts of PAb and SAb were 8.0 and 30.0 μL , respectively.

Under the above optimized controlled conditions, the quantitative sensing of LM was performed using DPV based on the proposed SAb-Au/RGO/RhB/LM/BSA/PAb/ $\text{Ti}_3\text{C}_2\text{T}_x$ MXene sandwich-like immunosensor. From Fig. 6, it is observed that the peak currents of RhB probe linearly increase along with the increase in concentration of LM from 10 and 10^5 CFU mL^{-1} . The linear equation expression is $I_p (\mu\text{A}) = 0.281 + 0.003C$ (CFU mL^{-1}) ($R = 0.998$) and limit of detection (LOD) is calculated to be 2 ± 0.3 CFU mL^{-1} (S/N, 3). Compared with most previous works, the present work exhibits better sensing performance including linearity range (LR) and LOD (Table 1); this can attributed mainly to the desirable properties of $\text{Ti}_3\text{C}_2\text{T}_x$ MXene (large surface area and conductivity) and the signal amplification capacity of Au/RGO/RhB.

3.3 Specificity, reproducibility and stability of the developed immunosensing platform

For further evaluating the specificity of the developed sandwich-like immunosensor, the response signals of RhB were studied through selecting other pathogens other than LM. As exhibited in Fig. 7A, the obtained results revealed a significant peak current of RhB observed only from LM (10^4 CFU mL^{-1}); as for the other pathogens including *Escherichia coli* (Es), *Vibrio parahaemolyticus* (Vp), *Shigella* (Sh), *Salmonella enteritidis* (Se) and *Staphylococcus aureus* (Sac) coupled with a high concentration (5×10^4 CFU mL^{-1}), the presented peak currents of RhB are nearly negligible, demonstrating that the developed sandwich-like immunosensor offers excellent specificity for detecting LM.

Next, for evaluating the reproducibility of the immunosensor, the electrochemical responses of 8 fabricated sensors were measured parallelly (Fig. 7B), and the results display that

Table 1 Comparison of various immunosensing platforms with the present method for LM

Electrodes	Assay label	Measuring method	Linear range/CFU mL^{-1}	LOD/CFU mL^{-1}	Reference
Gold electrode	HRP	Amperometry	10^2 to 10^6	10^2	6
Interdigitated array microelectrode	Urease	EIS	1.9×10^3 to 1.9×10^6	1.6×10^3	51
TiO_2 nanowire bundle gold microelectrode	Label-free	EIS	—	10^2	52
Au NPs/screen-printed carbon electrode	HRP	Amperometry	2.25×10 to 2.25×10^5	2.25×10^2	53
CNT fibers electrode	HRP	Cyclic voltammetry	10^2 to 10^5	1.07×10^2	54
$\text{C-Ti}_3\text{C}_2\text{T}_x$ MXene/GCE	RhB	DPV	10 to 10^5	2 ± 0.3	This work



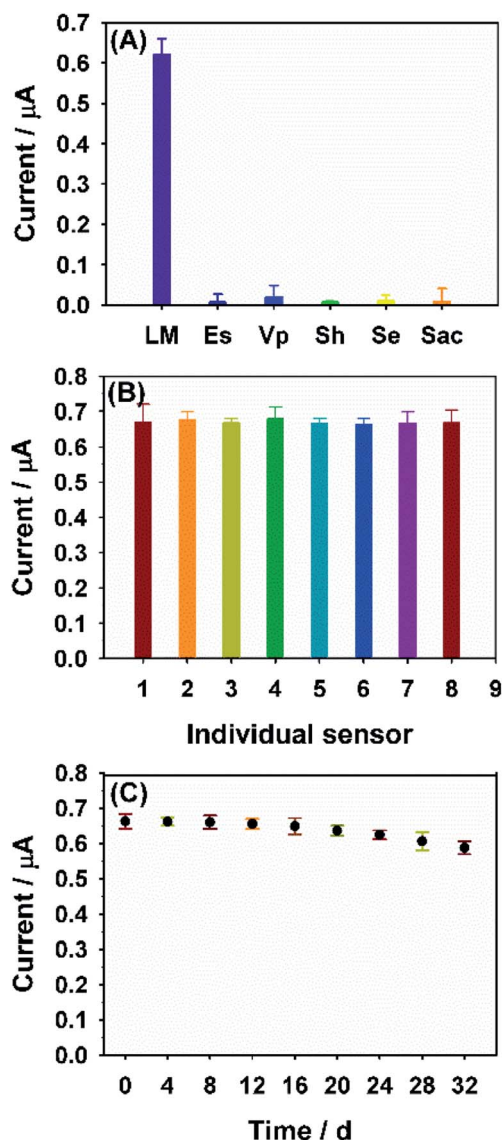


Fig. 7 (A) Specificity, (B) reproducibility and (C) stability for LM detection based on the constructed sandwich-like immunosensor. The specific values of the error bar for (A), (B) and (C) are 0.02 ± 0.006 , 0.03 ± 0.005 and 0.02 ± 0.004 , respectively.

the relative standard deviation value from their current response was 4.36%. Furthermore, in order to study the immunosensor stability, the modified electrodes and SAB-Au/RGO/RhB were stored at 4 °C for further use; the research results showed almost no decrease in the current response ($\sim 5.32\%$) after storing for 32 days (Fig. 7C). These results reveal that the proposed immunosensor possesses satisfactory reproducibility and good stability.

4. Conclusion

By preparing C-Ti₃C₂T_x MXene and Au/RGO/RhB respectively as the sensing platform and signal amplifier, a new sandwich-like electrochemical immunosensing platform was constructed herein for the sensitive determination of LM. Owing to the

superior properties of Ti₃C₂T_x MXene and Au/RGO/RhB, the developed SAB-Au/RGO/RhB/LM/BSA/PAb/Ti₃C₂T_x MXene sandwich-like immunosensor possesses wider LR and lower LOD than most previous works for detecting LM; thus it is believed that this sensor has great potential for application in LM analysis. Furthermore, through selecting other suitable antibodies other than LM antibodies, the sandwich-like immunosensor based on the prepared C-Ti₃C₂T_x MXene and Au/RGO/RhB shows broad application prospects for other targets.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was financially supported in part by Research Project of Military Logistics of PLA of China (No. CLB20J024), Foreign cooperation projects of Fujian Natural Science Foundation (No. 2019I0025), National Natural Science Foundation of China (No. 81772848), Joint Funds for the Innovation of Science and Technology from Fujian Province (No. 2017Y9127), and Fund from the 900th Hospital of Joint Logistics Support Force (No. 2020Q01, 2020Q06, 2020Z15, and 2020L17).

References

- 1 N. F. D. Silva, M. M. P. S. Neves, J. M. C. S. Magalhães, C. Freire and C. Delerue-Matos, *Trends Food Sci. Technol.*, 2020, **99**, 621–633.
- 2 J. Ding, J. Lei, X. Ma, J. Gong and W. Qin, *Anal. Chem.*, 2014, **86**, 9412–9416.
- 3 D. Patel, Y. Zhou and R. P. Ramasamy, *J. Electrochem. Soc.*, 2021, **168**, 057523.
- 4 X. Qi, Z. Wang, R. Lu, J. Liu, Y. Li and Y. Chen, *Food Chem.*, 2021, **338**, 127837.
- 5 S. Eissa and M. Zourob, *Microchim. Acta*, 2020, **187**, 486.
- 6 C. Cheng, Y. Peng, J. Bai, X. Zhang, Y. Liu, X. Fan, B. Ning and Z. Gao, *Sens. Actuators, B*, 2014, **190**, 900–906.
- 7 Q. Chen, J. Lin, C. Gan, Y. Wang, D. Wang, Y. Xiong, W. Lai, Y. Li and M. Wang, *Biosens. Bioelectron.*, 2015, **74**, 504–511.
- 8 B. Giménez, N. Graiver, L. Giannuzzi and N. Zaritzky, *Food Control*, 2021, **121**, 107602.
- 9 B. B. Tasbasi, B. C. Guner, M. Sudagidan, S. Ucak, M. Kavruk and V. C. Ozalp, *Anal. Biochem.*, 2019, **587**, 113449.
- 10 S. Wachiralurpan, K. Chansiri and P. A. Lieberzeit, *Sens. Actuators, B*, 2020, **308**, 127678.
- 11 Y. Zhao, S. Bu, C. Wang, C. Ma, Z. Li, W. Zhang and J. Wan, *Anal. Lett.*, 2021, **54**, 1603–1615.
- 12 J. Riu and B. Giussani, *Trac. Trends Anal. Chem.*, 2020, **126**, 115863.
- 13 X. Niu, W. Zheng, C. Yin, W. Weng, G. Li, W. Sun and Y. Men, *J. Electroanal. Chem.*, 2017, **806**, 116–122.
- 14 A. Liu, L. Shen, Z. Zeng, M. Sun, Y. Liu, S. Liu, C. Li and X. Wang, *Food Anal. Methods*, 2018, **11**, 215–223.



- 15 N. F. D. Silva, M. M. P. S. Neves, J. M. C. S. Magalhães, C. Freire and C. Delerue-Matos, *Talanta*, 2020, **216**, 120976.
- 16 Q. Guo, X. Zhang, F. Zhao, Q. Song, G. Su, Y. Tan, Q. Tao, T. Zhou, Y. Yu, Z. Zhou and C. Lu, *ACS Nano*, 2020, **14**, 2788–2797.
- 17 Y. Li, H. Shao, Z. Lin, J. Lu, L. Liu, B. Duployer, P. O. Å. Persson, P. Eklund, L. Hultman, M. Li, K. Chen, X.-H. Zha, S. Du, P. Rozier, Z. Chai, E. Raymundo-Piñero, P.-L. Taberna, P. Simon and Q. Huang, *Nat. Mater.*, 2020, **19**, 894–899.
- 18 M. Zhou, S. Ning, J. Liu, G. I. N. Waterhouse, L. Li, J. Dong and S. Ai, *Anal. Lett.*, 2021, **54**, 1769–1782.
- 19 J. Chen, P. Tong, L. Huang, Z. Yu and D. Tang, *Electrochim. Acta*, 2019, **319**, 375–381.
- 20 G. Cai, Z. Yu, P. Tong and D. Tang, *Nanoscale*, 2019, **11**, 15659–15667.
- 21 L. Wu, X. Lu, Dhanjai, Z.-S. Wu, Y. Dong, X. Wang, S. Zheng and J. Chen, *Biosens. Bioelectron.*, 2018, **107**, 69–75.
- 22 P. K. Kalambate, Dhanjai, A. Sinha, Y. Li, Y. Shen and Y. Huang, *Microchim. Acta*, 2020, **187**, 402.
- 23 Y. Xie, F. Gao, X. Tu, X. Ma, Q. Xu, R. Dai, X. Huang, Y. Yu and L. Lu, *J. Electrochem. Soc.*, 2019, **166**, B1673–B1680.
- 24 Y. Yi, D. Zhang, Y. Ma, X. Wu and G. Zhu, *Anal. Chem.*, 2019, **91**, 2908–2915.
- 25 X. Tu, F. Gao, X. Ma, J. Zou, Y. Yu, M. Li, F. Qu, X. Huang and L. Lu, *J. Hazard. Mater.*, 2020, **396**, 122776.
- 26 H. Zhang, Z. Wang, F. Wang, Y. Zhang, H. Wang and Y. Liu, *Anal. Chem.*, 2020, **92**, 5546–5553.
- 27 P. K. Kalambate, N. S. Gadhari, X. Li, Z. Rao, S. T. Navale, Y. Shen, V. R. Patil and Y. Huang, *Trac. Trends Anal. Chem.*, 2019, **120**, 115643.
- 28 Y. Yi, Y. Ma, F. Ai, Y. Xia, H. Lin and G. Zhu, *Chem. Commun.*, 2021, **57**, 7790–7793.
- 29 Y. Xia, Y. Ma, Y. Wu, Y. Yi, H. Lin and G. Zhu, *Microchim. Acta*, 2021, **188**, 1–9.
- 30 Y. Xia, Y. Zhao, F. Ai, Y. Yi, T. Liu, H. Lin and G. Zhu, *J. Hazard. Mater.*, 2021, 127974.
- 31 R. Zeng, W. Wang, M. Chen, Q. Wan, C. Wang, D. Knopp and D. Tang, *Nano Energy*, 2021, **82**, 105711.
- 32 R. Zeng, Z. Luo, L. Zhang and D. Tang, *Anal. Chem.*, 2018, **90**, 12299–12306.
- 33 Y. Yao, L. Lan, X. Liu, Y. Ying and J. Ping, *Biosens. Bioelectron.*, 2020, **148**, 111799.
- 34 M. Mohammadniaei, A. Koyappayil, Y. Sun, J. Min and M.-H. Lee, *Biosens. Bioelectron.*, 2020, **159**, 112208.
- 35 Y. Cai, J. Shen, G. Ge, Y. Zhang, W. Jin, W. Huang, J. Shao, J. Yang and X. Dong, *ACS Nano*, 2018, **12**, 56–62.
- 36 D. Huang, Y. Wu, F. Ai, X. Zhou and G. Zhu, *Sens. Actuators, B*, 2021, **342**, 130074.
- 37 L. Jothi, S. K. Jaganathan and G. Nageswaran, *Mater. Chem. Phys.*, 2020, **242**, 122514.
- 38 P. Zhao, M. Ni, Y. Xu, C. Wang, C. Chen, X. Zhang, C. Li, Y. Xie and J. Fei, *Sens. Actuators, B*, 2019, **299**, 126997.
- 39 K. Kunpatee, P. Chamsai, E. Mehmeti, D. M. Stankovic, A. Ortner, K. Kalcher and A. Samphao, *J. Electroanal. Chem.*, 2019, **855**, 113630.
- 40 R. Zhang, C. Zhang, F. Zheng, X. Li, C.-L. Sun and W. Chen, *Carbon*, 2018, **126**, 328–337.
- 41 S. Lv, K. Zhang, Y. Zeng and D. Tang, *Anal. Chem.*, 2018, **90**, 7086–7093.
- 42 Q. Zhou, Y. Lin, K. Zhang, M. Li and D. Tang, *Biosens. Bioelectron.*, 2018, **101**, 146–152.
- 43 Y. Chen, Y. Li, D. Deng, H. He, X. Yan, Z. Wang, C. Fan and L. Luo, *Biosens. Bioelectron.*, 2018, **102**, 301–306.
- 44 Y. Liu, H. Ma, J. Gao, D. Wu, X. Ren, T. Yan, X. Pang and Q. Wei, *Biosens. Bioelectron.*, 2016, **79**, 71–78.
- 45 M. Li, P. Wang, F. Li, Q. Chu, Y. Li and Y. Dong, *Biosens. Bioelectron.*, 2017, **87**, 752–759.
- 46 H. Lv, X. Zhang, Y. Li, Y. Ren, C. Zhang, P. Wang, Z. Xu, X. Li, Z. Chen and Y. Dong, *Microchim. Acta*, 2019, **186**, 1–10.
- 47 H. Wu, M. Almalki, X. Xu, Y. Lei, F. Ming, A. Mallick, V. Roddatis, S. Lopatin, O. Shekhah, M. Eddaoudi and H. N. Alshareef, *J. Am. Chem. Soc.*, 2019, **141**, 20037–20042.
- 48 P. Zhang, L. Wang, K. Du, S. Wang, Z. Huang, L. Yuan, Z. Li, H. Wang, L. Zheng and Z. Chai, *J. Hazard. Mater.*, 2020, **396**, 122731.
- 49 F. Cui and X. Zhang, *J. Electroanal. Chem.*, 2012, **669**, 35–41.
- 50 G. Goncalves, P. A. A. P. Marques, C. M. Granadeiro, H. I. S. Nogueira, M. K. Singh and J. Grácio, *Chem. Mater.*, 2009, **21**, 4796–4802.
- 51 D. Wang, Q. Chen, H. Huo, S. Bai, G. Cai, W. Lai and J. Lin, *Food Control*, 2017, **73**, 555–561.
- 52 R. Wang, C. Ruan, D. Kanayeva, K. Lassiter and Y. Li, *Nano Lett.*, 2008, **8**, 2625–2631.
- 53 D. Davis, X. Guo, L. Musavi, C. S. Lin, S. H. Chen and V. C. H. Wu, *Ind. Biotechnol.*, 2013, **9**, 31–36.
- 54 Y. Lu, Y. Liu, Y. Zhao, W. Li, L. Qiu and L. Li, *J. Nanomater.*, 2016, **2016**, 16.

