



Cite this: *J. Mater. Chem. B*, 2021, 9, 7030

Polysaccharide nanoparticles: from fabrication to applications

Alexander Plucinski,[†] Zan Lyu [†] and Bernhard V. K. J. Schmidt ^{*}

Polysaccharides have attracted considerable attention in a broad range of applications in recent years, which is due to their remarkable features such as biocompatibility, biodegradability, renewable origin, and facile modification. Considerable research efforts have been focused on developing polysaccharide nanoparticles and to promote their applications in various areas and biomedicine in particular. The present review highlights the properties of common polysaccharides used in nanoparticle formation as well as strategies to fabricate polysaccharide nanoparticles. Furthermore, the combination of polysaccharide nanoparticles and polymers is presented and brought into the context of applications. Finally, applications of polysaccharide nanoparticles as nano-delivery system, Pickering emulsion stabilisers, and material reinforcing agent in the fields of nanomedicine, cosmetics, and food system are highlighted. Moreover, this review describes and critically discusses present limitations and drawbacks in the preparation and use of polysaccharide nanoparticles, revealing directions to develop polysaccharide nanoparticles for further utilisation in various applications in the future.

Received 22nd March 2021,
Accepted 23rd April 2021

DOI: 10.1039/d1tb00628b

rsc.li/materials-b

1. Introduction

Hydrophilic polymers and water-based polymer systems have gained significant attention over the past decades due to their applications in various interdisciplinary areas *e.g.* drug-delivery,¹ tissue-engineering,² catalysis,³ membrane technology,⁴ aggregate formation^{5,6} or phase separations like aqueous two-phase systems (ATPS).⁷ Especially, nanoparticle (NP) based technology⁸ is a significant area in biomedical,⁹ pharmaceutical,⁹ cosmetic/cosmeceutical^{10,11} and food industry.¹² As such, NPs have

been prepared and utilised in the areas such as targeted substance delivery, emulsion stabilisation, imaging on the molecular scale for diagnostic applications, and material reinforcement.^{13–18} Moreover, emulsion stabilisers based on NPs have been extensively studied for application as cosmetic formulations,¹⁹ in oil-in-water (O/W) emulsions^{20,21} and in water-in-water (W/W) emulsions.^{22,23} The transformation of traditional food and agriculture sectors is also significantly promoted by nanotechnology. In addition to transporting bioactive substances like nutrients, numerous novel applications such as smart and active packaging, nanosensors, nanopesticides and nanofertilisers have been invented for improving food quality and agricultural output.¹²

Polysaccharides are a significant class of hydrophilic polymers with natural origin and biocompatibility that find frequent use

School of Chemistry, University of Glasgow, G12 8QQ Glasgow, UK.

E-mail: bernhard.schmidt@glasgow.ac.uk

[†] These authors contributed equally.



Alexander Plucinski

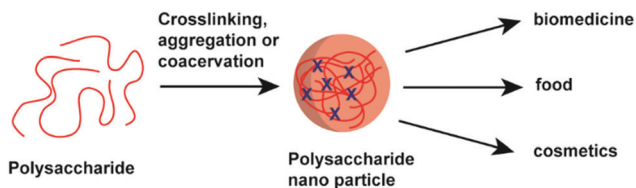
Alexander Plucinski has obtained his chemistry bachelor degree in 2016 and master degree in 2018 at the University of Potsdam. Since 2019, he is a PhD student under the supervision of Dr Bernhard V. K. J. Schmidt at the University of Glasgow. His research focusses on the self-assembly and phase separation of hydrophilic polymers in aqueous environment.



Zan Lyu

Zan Lyu received his dual BSc degree from Changzhou University, China and St. Francis Xavier University, Canada in 2019. In 2020, he received his master's degree in Chemistry at the School of Chemistry, University of Glasgow.





Scheme 1 Overview of polysaccharide NP formation and their applications.

in water-based polymer systems and in nanotechnology in particular, which is mainly due to their favourable properties in biological systems, *e.g.* biodegradability, biocompatibility, and low-toxicity. These properties constitute considerable requirements for the utilisation of NPs and thus polysaccharides represent an ideal class of building blocks for NP fabrication.²⁴ For example, in the field of medical therapy, polysaccharide-based NPs have the advantages of high loading efficiencies, fast drug release rates and good targeting ability as well as high stability and low toxicity in physiological environment.^{9,25,26} In addition to biodegradability and biocompatibility, polysaccharides have gained considerable attention due to their abundance, facile processing and sustainable feedstocks (refer to Section 2).¹⁹ The chemical functionalisation of polysaccharides is mainly achieved by using the free carboxyl and hydroxyl groups distributed along the backbone of the polysaccharides. The utilisation of these reactive groups allows the fabrication of suitable polysaccharide derivatives with determined properties (*e.g.* hydrophobicity, solubility), promoting further use of polysaccharides in specific application areas.^{25,27,28}

Based on common mechanisms such as ionic crosslinking, covalent crosslinking, self-assembly of hydrophobically modified polysaccharides, polyelectrolyte complexation, and forming polysaccharide–drug conjugates (refer to Section 3),²⁵ many researchers have designed different approaches to synthesise polysaccharide-based NPs with controlled size, morphology, and structure (Scheme 1).²⁹ The key point of the choice of the synthetic route is to optimise the final properties of NPs designed for a specific application. In addition, a series of

factors such as physicochemical parameters of the polysaccharides used, polysaccharide chemical composition, NPs size, and surface morphology, can be used as a guide during the synthesis process.³⁰ Sometimes, misunderstandings occur because of the misuse of the term NP referring to both NP and nanocrystal, which are indeed different types of materials. NPs are amorphous particles while nanocrystals are crystalline.³¹ Therefore, polysaccharide nanocrystals will not be discussed detailedly in the following sections, but some useful applications will be mentioned.

As polysaccharide-based NPs have a broad range of possible applications, understanding the mechanism of action is highly important. In this review, the properties of several common polysaccharides (refer to Section 2) and practical methods for preparing their NPs will be discussed (refer to Section 3). Also, the development of polysaccharide/polymer NP systems (refer to Section 4) is discussed, highlighting the advantages of the NP system obtained from a polysaccharide and synthetic polymer combination. Finally, on the basis of the preceding sections, the emphasis of this review is to survey some useful applications of polysaccharide-based NPs in nanomedicine, cosmetics, and food (refer to Section 5). At last, we will summarise the developments in the field of polysaccharide NPs and give an outlook for future directions.

2. Polysaccharides

Polysaccharides are natural polymers consisting of monosaccharide units linked by glycosidic bonds with chitosan (CS), cellulose (CL), starch, hyaluronic acid (HA), and dextran (DEX) being typical examples.^{25,32} Similar to proteins and glycosaminoglycans, which are also common natural polymers, polysaccharides can be extracted from various sources, for instance, from plant origin, microbial origin and animal origin.^{25,33} The biological properties and activities of different polysaccharides are diverse due to their different chemical structures (Fig. 1) and ionic nature. Thus, their functions and applications are directly related to their chemical and biochemical specifications.^{25,28} The properties of common polysaccharides which are widely used in nanomedicine, cosmetics, food, and other areas are summarised in Table 1, and more information is detailed in the following sections.

2.1 Chitin and chitosan

Chitin (CH) is the primary component of the crustaceans' exoskeleton, and it is also the second most abundant natural polysaccharide.^{25,45} A commonly employed derivative of CH is chitosan (CS), which is a linear polysaccharide obtained from the partial *N*-deacetylation of CH with a deacetylation degree of over 40% under alkaline conditions and elevated temperature (Fig. 1a).⁴⁶ CH has a hydrophobic linear structure composed of β -(1,4)-linked *N*-acetyl-D-glucosamine units, while the structure of CS is mainly based on β -(1,4)-linked glucosamine and *N*-acetyl glucosamine units.^{28,47}

CH is insoluble in water or in other common solvents. However, CS produced from the *N*-deacetylation process of CH has increased aqueous solubility. Acidic conditions promote the



Bernhard V. K. J. Schmidt

block copolymer self-assembly, metal–organic framework/polymer hybrids and carbon nitride/polymer hybrid materials.

Bernhard V. K. J. Schmidt completed his PhD in 2013 with Prof. Barner-Kowollik at the Karlsruhe Institute of Technology and a PostDoc with Prof. Hawker at the University of California, Santa Barbara. Afterwards he joined the department of Prof. Antonietti at the Max Planck Institute of Colloids and Interfaces as a Group Leader and finished his Habilitation in 2020. Since 2019, he is Lecturer in Synthetic Polymer Chemistry at the University of Glasgow. His research focusses on



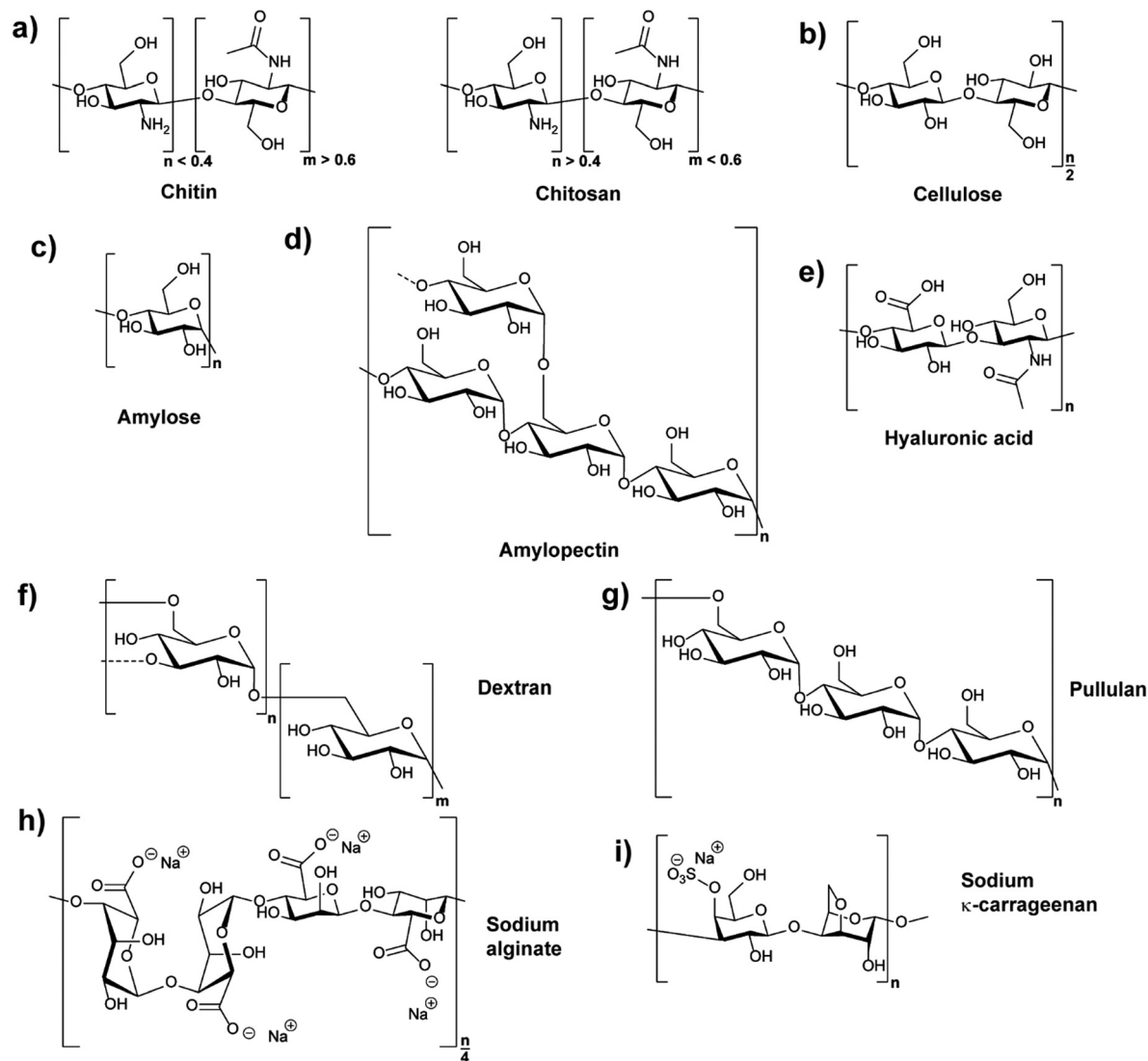


Fig. 1 Chemical structures of polysaccharides: (a) chitin (CH) and its derivative from *N*-deacetylation chitosan (CS), (b) cellulose (CL), (c) and (d) structures of two different polymers coexisting in starch, amylose and amylopectin (dashed lines indicate branches), (e) hyaluronic acid (HA), (f) dextran (DEX) (dashed lines indicate branches), (g) pullulan (PL), (h) sodium alginate (α -L-guluronic acid- α -L-guluronic acid- β -D-mannuronic acid- β -D-mannuronic acid sequence shown), and (i) κ -carrageenan (κ -CRG).

Table 1 Properties of common polysaccharides

Polysaccharide	Water solubility	Molar mass range/kDa	Source
Alginate	Soluble (sodium or potassium salt)	32–400 ³⁴	Algae, bacteria
Carrageenan (CRG)	Soluble in hot water	100–1000 ³⁵	Seaweed
Chitin (CH)	Insoluble ²⁸	1000–2500 ²⁸	Exoskeleton of non-mammals
Chitosan (CS)	Soluble under acidic conditions ²⁷	100–500	<i>N</i> -Deacetylation process of chitin
Cellulose (CL)	Insoluble ³⁶	50–2000 ^{37,38}	Higher plants
Dextran (DEX)	Soluble	3–2000 ^{39,40}	Bacteria
Starch	Insoluble in cold water ⁴¹	~100 for amylose, and 1000–10 000 for amylopectin	Higher plants
Hyaluronic acid (HA)	Soluble ⁴²	20–10 000 ^{28,43}	Mammals
Pullulan (PL)	Soluble	100–250 ⁴⁴	Bacteria

water solubility of CS due to protonation of the basic amino groups.²⁸ In addition, the conversion of CH to CS decreases the molecular weight (MW) from an average range of 1000–2500 kDa to 100–500 kDa. The lower MW of CS resulting from the deacetylation process can also facilitate the water solubility of

CS.^{25,28} The more frequent use of CS in NPs compared to CH can be deduced to the improved water solubility. Furthermore, the amine functions of CS can be employed for chemical reactions, *e.g.* functionalisation, and CS can be produced in many kinds of forms, such as powder, paste, and film. Frequently, CS is



utilised for the synthesis of widely used CS-based nanoparticles (CSNPs).^{28,33}

As the nature of CS favours chemical modifications compared to less reactive CH, the preparation of CS-based nanoparticles (CSNPs) can be conducted by various avenues. Effective chemical modifications enable further options to use the material compared to the natural compound. The chemical modifications are usually carried out at the sites of reactive amino groups of CS, and there is a wide range of examples of commonly used CS derivatives such as thiolated CS, sugar-bearing CS, and carboxyalkyl CS.^{25,48} These CS derivatives are designed to enhance specific properties of CS. For instance, CS modified by thiol groups can obtain improved mucoadhesive properties, while conjugating hydrophobic moieties (e.g. deoxycholic acid and cholesterol) to CS introduces an avenue to form CSNPs by self-assembly.^{45,48}

CS is widely investigated in many areas of application, for example, drug delivery, tissue engineering, and stabilising cosmetic ingredients.^{45,49,50} The positive charge density on CS is thought as significant in improving the cell uptake, but it is accompanied with higher toxicity.^{25,51} To solve this problem, the selection of suitable CS derivatives have attracted growing attention, in order to improve health benefits of CS.⁵¹

2.2 Cellulose

Cellulose (CL) is the primary constituent of plant cell walls, and it is the most abundant natural polysaccharide.²⁸ As the base fundamental unit of plants, CL is considered as human/animal/environmental-friendly, biocompatible, and biodegradable.^{33,52} CL is mainly extracted from plant origin, and its linear polysaccharide structure is formed by β -(1,4)-linked D-glucose units, giving a flat ribbon-like conformation (Fig. 1b).⁵²

The CL polymer and polymer fibrils are relatively stable due to their inter- and intra-chain hydrogen bonding network. Thus, the linkage of individual CL chains is stabilised by supramolecular interactions, giving the linear configuration and an axial stiffness in CL fibrils. On the basis of CL biosynthesis and extraction processes, various types of CL-based nanoparticles (CLNPs) and CL nanocomposite materials can be obtained. The main factors which cause the differentiation are the source materials of CL (e.g. wood, plant, and bacteria) and the deconstruction processes during extraction.^{52,53}

For the utilisation of CL, many applications in diverse research areas are described, such as food packaging, drug delivery, films, and reinforcing materials for polymer matrices.^{33,52} After appropriate surface functionalisation, the surface chemistry of CLNPs enables self-assembly, making CLNPs versatile materials with considerable mechanical properties.⁵²

2.3 Starch

In plant tubers and seed endosperm, starch is the primary storage carbohydrate which can be extracted from many resources such as corn and potato.⁵⁴ Chemically, two different polysaccharide structures are present in starch, which are amylose (Fig. 1c) and amylopectin (Fig. 1d). Amylose is a linear polymer formed by α -(1,4)-linked D-glucose units while amylopectin is a highly branched polymer composed of short chains of α -(1,4)-linked D-glucose

units with branches formed by α -(1,6) linkages at the branch positions.^{54,55}

The average MW of amylose is in the range of 10^5 Da while that of amylopectin is distributed from 10^6 to 10^7 Da. For example, rice starch has higher MW compared with those from corn, wheat, and potato, because rice starch has the lowest amylose content amongst them.⁵¹ Similar to CL, starch is insoluble in cold water due to the hydrogen bonding effect. Moreover, the high MW of starch decreases the solubility. In relation, amylopectin has higher solubility compared to amylose because of its highly branched structure, which is useful in separating amylose and amylopectin from starch granules.^{27,51}

Starch-based NPs (SNPs) can be prepared by various strategies such as precipitation, solvent evaporation, and emulsion cross-linking. Moreover, different preparation methods will result in variation of SNPs properties (e.g. shape and crystallinity).^{33,56} Compared with other polysaccharides, starch is unique because it has the property to convert to thermoplastic material in the presence of plasticisers. Also, starch has the advantages of stability, biocompatibility, and biodegradability, making it useful in drug delivery, tissue engineering, and food packaging applications.^{33,47}

2.4 Hyaluronic acid

As a typical extracellular matrix polymer, hyaluronic acid (HA) has broad applications in biomedical and nanotechnological areas. Regarding the structure of HA, it is a hydrophilic linear polysaccharide composed of alternately linked D-glucuronic and N-acetyl-D-glucosamine units *via* β -1,3 and β -1,4 glycosidic bonds (Fig. 1e).^{33,43} As HA is a negatively charged polysaccharide, hyaluronic acid-based NPs (HANPs) can be formed by using cationic molecules (e.g. CS) as ionic crosslinkers. Moreover, approaches such as making HA-cargo (e.g. drug) conjugates have also been developed to form HA-based nanocarriers.^{57,58} HA is mostly extracted from tissues, and it is significant in the areas of tissue repair, wound healing, food industry, and cosmetics. Moreover, HA is not only a good wound healing factor but also an adequate carrier for the delivery of antibiotics that aid in wound healing properties.^{15,28}

2.5 Dextran

Dextran (DEX) belongs to the first commercial exopolysaccharides produced by bacterial enzymes and is mostly obtained from *Leuconostoc mesenteroide*. It is branched, and the basic structure of the various types of dextran consists of main chains formed by (1,6)- α -D-glucose with various ratios of linkages and branches (Fig. 1f).⁵⁹ The ratios of α -(1,6) linkages can vary from 97% to 50%, and the branching can take place mainly in position α -(1,3), and occasionally in position α -(1,2) or α -(1,4).^{28,59} Because of the reactive hydroxyl chemistries of DEX, it is easy to be functionalised.²⁸ Furthermore, DEX is highly water-soluble and frequently used in ATPS.^{60,61} Depending on the strain of bacteria and conditions used, properties of DEX such as branching and molecular weight vary considerably. The MW of DEX ranges from 3000 Da to 2 000 000 Da, and the larger DEX



(> 60 000) are excreted poorly from the kidney and remain in the blood before being completely metabolised.^{39,62}

Non-ionic DEX-based NPs can be obtained by self-assembly, and due to the biocompatibility and biodegradability of DEX, NPs from DEX are discussed as suitable drug/gene nano-vehicles. In addition, the superior water solubility of Dextran-based NPs (DEXNPs) prevents cellular toxicity after completing the drug delivery process.⁶³ Similar to CS and HA, the advantages of DEXNPs can be promoted after suitable chemical modifications or forming DEX–drug conjugates. Typical DEX derivatives such as thiolated DEX, phosphorylated DEX, and dextran sulfate (DS) are all very useful in enhancing the targeting ability of DEXNPs.^{26,59,63} For example, the chemical modification of DEX to obtain DS enables interactions with lipoproteins and complexation with fibrinogen or DNA. Furthermore, modification with hydrophobic moieties introduces amphiphilicity, which is very important to form self-assembled and water dispersible DEXNPs in the field of drug delivery.²⁶

2.6 Pullulan

Pullulan (PL) is a non-ionic exopolysaccharide which is mainly obtained from the fermentation medium of the fungus-like yeast *Aureobasidium pullulans* under limiting conditions. PL has been widely used in various areas because it is non-toxic, non-mutagenic, non-carcinogenic, biocompatible, and bio-degradable.⁶⁴ The structure of PL is based on α -(1,6) linked maltotriose units (Fig. 1g), and the linkage between maltotriose units significantly gives structural flexibility of PL.^{65,66} PL is easily soluble in water and insoluble in most organic solvents. Compared with other polysaccharides, the PL aqueous solution is stable and has a relatively low viscosity. The MW of PL ranges from 100 kDa to 250 kDa, and its appearance is white or yellowish-white powder.⁴⁴ PL hydrogel NPs have been employed for gene delivery⁶⁷ or as composite with CS for vaccine delivery.^{68,69} Recently, PL was utilised as block in double hydrophilic block copolymers, which were utilised in turn for self-assembly, for example in the formation of completely hydrophilic capsules,⁷⁰ droplets^{71,72} or particles.⁷³

Similar to DEX-based NPs, PL-based NPs (PLNPs) can be prepared by self-assembly after the hydrophobic modification of PL. In the fields of drug and gene delivery, hydrophobically modified PL has been widely investigated to be a good nano-carrier. Moreover, by mixing hydrophobically modified PL with quantum dots, the formed PLNPs can also play a significant role in medical tracing areas such as tumour imaging.^{32,66} PL is also being used in the areas of food and cosmetics, for example to extend food shelf life and simplify food processing. For cosmetics, PL can be utilised in lotions, shampoos, and face masks.⁷⁴

2.7 Alginate

Alginate is a well-known polysaccharide produced by brown algae and bacteria consisting of α -L-guluronic acid and β -D-mannuronic acid building blocks that are linearly linked by 1,4-glycosidic linkages (Fig. 1h).⁷⁵ Alginate chains consist of blocks of either all α -L-guluronic acid, all β -D-mannuronic acid, or alternating α -L-guluronic acid– β -D-mannuronic acid subunits.⁷⁶ It is known to be biodegradable, non-toxic and abundant with

molar masses between 32 and 400 kDa.³⁴ The polymer is anionic due to the acid functions and frequently used in the form of sodium or potassium salts. A remarkable feature of alginate is its interaction of α -L-guluronic acid– α -L-guluronic acid subunits with divalent cations like calcium ions leading to hydrogel formation.⁷⁶ These hydrogels are utilised in various medical applications,⁷⁷ e.g. encapsulation of drugs and release⁷⁸ or as cell growth environment.⁷⁹ The complexation with Ca^{2+} can also be employed to form NPs for example *via* emulsion methods⁸⁰ or precipitation.⁸¹ Also, a combination with cationic CS enables NP formation *via* coacervation.⁸²

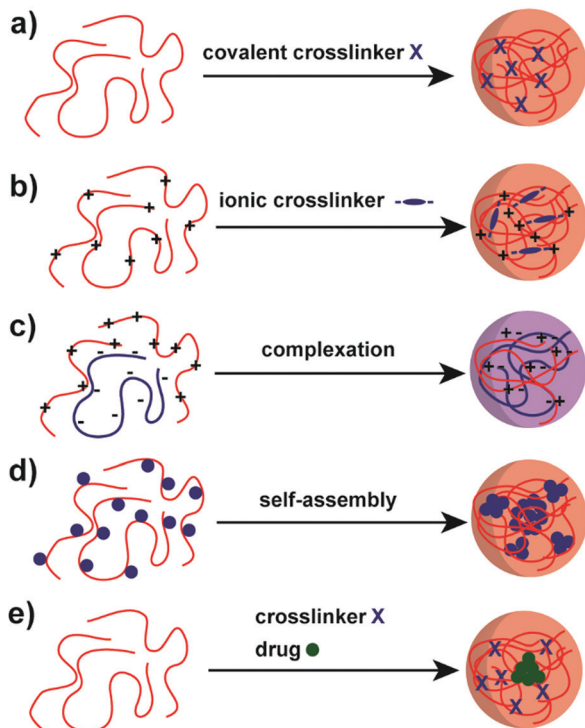
2.8 Carrageenan

Carrageenans (CRG) are linear sulphated polysaccharides and originate from seaweed.³⁵ The polymer structure consists mainly of 3-linked β -D-galactopyranose and 4-linked α -D-galactopyranose or 4-linked 3,6-anhydro- α -D-galactopyranose units (Fig. 1i with an example structure). CRGs are classified into six types based on the chemical structure (κ), (ι), (λ), (μ), (ν) and (θ)-CRG), *i.e.* the position of sulfate esters and the presence of anhydro units. Overall, the MW of commercial CRG ranges from 100 to 1000 kDa with solubility in hot water and partially cold water depending on substitution. CRGs form helices in aqueous environment leading to gelation that is exploited in food industry for thickening.⁸³ Due to its polyanionic character, CRG can be used in conjunction with CS to form NPs.^{84,85} Nevertheless, the biocompatibility and toxicity of CRG is currently under discussion and has to be followed closely for food and biomedical applications in particular.⁸⁶

3. Synthesis of polysaccharide nanoparticles

A variety of methods has been proposed to prepare NPs based on natural polysaccharides. The selection of a proper synthesis method depends on the targeted application and its requirements. For example, many factors such as thermal and chemical stability of the active agent, reproducibility of the release kinetic profiles, particle size, and stability of the final product and residual toxicity associated with the final product have to be taken into account before preparing drug delivery nanocarriers⁸⁷ and NPs for other applications. The avenue for the preparation of NPs is crucial for stability under the intended conditions and in particular the crosslinking chemistry has to be considered. For example, the native charges of polysaccharides can be exploited for the synthesis of polysaccharide NPs^{88–90} or covalent crosslinking can be implemented.^{91–93} Mizrahy and Peer discussed the main mechanisms during the synthesis of polysaccharide NPs in their review in detail, which are categorised as chemical (covalent) crosslinking, physical (ionic) crosslinking, polyelectrolyte-complexation and self-assembly (Scheme 2).²⁵ Due to the repeated number of monomer units on the polysaccharide a full network can be achieved. Hence, a complete gelation of the reaction medium has to be suppressed *via* the preparation conditions, e.g. in dispersed systems.





Scheme 2 Avenues for the preparation of polysaccharide NPs: (a) covalent crosslinking, (b) ionic crosslinking, (c) complexation of oppositely charged polysaccharides, (d) self-assembly of hydrophobically (blue sphere) modified polysaccharides and (e) polysaccharide–drug (green sphere) conjugate.²⁵

In addition to the crosslinking process itself, the formation of NPs relies on the formation of materials on the nano scale *via* various preparative techniques. In this section, important and widely used methods such as nanoprecipitation, complex coacervation, and emulsification will be discussed. Also, the microfluidic technique has proven to be a useful direction for the preparation of polysaccharide NPs, both in emulsion and nanoprecipitation approaches.⁹⁴ Other related methods developed by researchers will be reviewed as well. The relationship between the properties (*e.g.* diameter) of the polysaccharide NPs synthesised and the methods utilised will be discussed to showcase how the methods affect the final properties.

3.1 Crosslinking and aggregation strategies

As for common polymer NPs, one of the main concerns in the fabrication of polysaccharide NPs is the crosslinking of the polysaccharide to form stable particles. Crosslinking has a considerable influence on the polysaccharide NP properties and hence applications, *e.g.* mechanical properties, swellability or drug-encapsulation/release.

Covalent crosslinking introduces irreversible crosslinking points leading to highly stable structures (Scheme 2a) that can withstand chemical stress, *e.g.* solvent changes or a broad pH range.²⁵ The crosslinker molecules have to be multifunctional in order to react twice with a polysaccharide molecule or with other crosslinkers to form a connection between polysaccharide molecules. Common crosslinkers for polysaccharides are dialdehydes

that link *via* acetal formation. Other options include multifunctional carboxylic acids forming ester or amide bonds. An avenue to introduce degradability or even stimulus response is the utilisation of cleavable crosslinkers.⁹⁵ One of the issues with covalent crosslinking is the potential toxicity of remaining non-linked small molecule crosslinkers that might hinder biomedical applications.

Another way to introduce crosslinks is *via* a supramolecular pathway for charged polysaccharides, *e.g.* in ionic crosslinking (Scheme 2b). For that, oppositely charged small ionic molecules with multiple functionalities are added, *e.g.* tripolyphosphate (TPP), or ions with multiple vacancies for interaction, for example Ca^{2+} .⁹⁶ The crosslinking proceeds *via* electrostatic interactions that connect the crosslinker with the polysaccharide. In contrast to covalent crosslinking, ionic crosslinking features a lower chemical stability, *e.g.* against changes in pH or ionic strength, and also physical stability, *e.g.* against temperature changes. However, this defined instability can be exploited to enable degradation on purpose, for example in drug-delivery applications.⁹⁵

A crosslinking method of high relevance is coacervation or polyelectrolyte complexation of charged polysaccharides. In order to form coacervate structures, oppositely charged polysaccharides and/or polyelectrolytes are employed. In a similar way to ionic crosslinking, electrostatic interactions lead to crosslinking (Scheme 2c). As no toxic small molecule crosslinkers are used, the coacervate approach is considered highly biocompatible at least if the employed polyelectrolytes/polysaccharides are biocompatible. The obtained stability of the NPs relies on charge density and distribution of charges along the chain, as the interaction between both polyelectrolytes/polysaccharides is the main factor for stability.⁹⁷ Similar to ionic crosslinking, pH, ionic strength and temperature play a crucial role for the stability and preparation of coacervate particles. Polysaccharides like HA, CS and alginate are utilised frequently for coacervate formation. Coacervation/polyelectrolyte complexation can be also utilised in the formation of layer-by-layer assemblies.^{98,99}

Self-assembly of polysaccharide chains can be employed for NPs formation as well, which is another supramolecular avenue.¹⁰⁰ Commonly, hydrophilic polysaccharides are functionalised with hydrophobic moieties (bile acids, fatty acids or cholesterol) that will aggregate in aqueous environment leading to micellar structures (Scheme 2d).^{101,102} As such, NPs with hydrophobic core and hydrophilic polysaccharide shell are obtained. The core can perform as a reservoir for hydrophobic drugs or other molecules. The properties of the micellar NPs can be tailored *via* the utilised polysaccharide conjugate, *e.g.* MW or hydrophobic to hydrophilic ratio, but also *via* the preparation process, *e.g.* the kinetics of micelle formation or the utilised solvent system. The approach was brought to the next level by Wich and coworkers,¹⁰³ who synthesised an amphiphilic block copolymer of DEX and hydrophobic acetalated DEX and formed micellar NPs that way. As such, a completely polysaccharide-based amphiphilic block copolymer was presented.

At last, a related structure to micelles should be discussed, namely polysaccharide–drug conjugates (Scheme 2e).⁴⁸ Instead of adding non-functional hydrophobic molecules to drive supramolecular aggregation, hydrophobic drugs can be conjugated as



well to the polysaccharide. As such, polysaccharide NPs can be obtained that serve as drug-delivery vehicles directly. An important characteristic is the connection of drug and polysaccharide that should be cleavable in the body at best at the right point of action.¹⁰⁴ In principle, also other methods of aggregation or crosslinking can be combined with polysaccharide-drug conjugates.

3.2 Nanoprecipitation

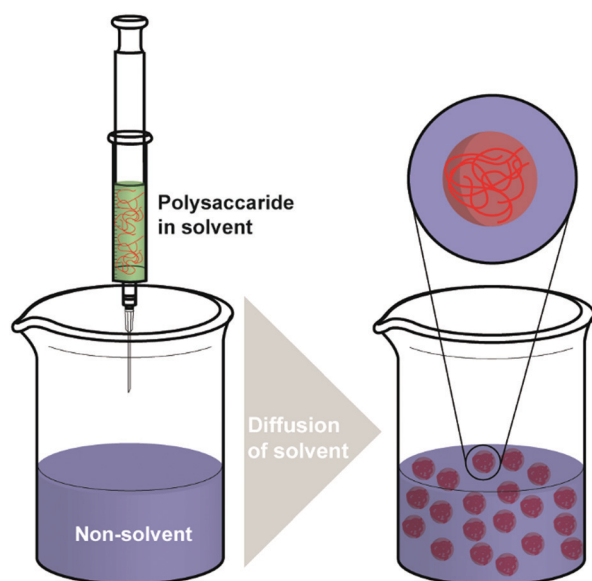
Nanoprecipitation is one of the first developed techniques for encapsulating drug molecules, and it is also known as solvent displacement or interfacial deposition. Nanoprecipitation has been widely used in the formation of many kinds of NPs such as polymeric NPs, drug NPs, drug-loaded protein NPs and inorganic NPs.¹⁰⁵ Though nanoprecipitation has advantages of being simple, reproducible, fast, and economic, it is still challenging to encapsulate water soluble compounds with this method.¹⁰⁶

In the nanoprecipitation process, two miscible phases are required, which are an aqueous phase and an organic/oil phase.¹⁰⁷ A typical procedure (Scheme 3) of nanoprecipitation consists of dissolving hydrophobic solutes in a water-miscible solvent wherein they have high solubility at first. Subsequently, this solution is added to a significant excess amount of an anti-solvent such as water or buffer solutions. Because the solubility of hydrophobic solutes decreases in the aqueous solution, particles of hydrophobic compounds will precipitate. Finally, particles of hydrophobic molecules are obtained after removing the excess solvent by evaporation, dialysis, or lyophilisation.¹⁰⁵ The work of Barresi and coworkers showed that mixing of phases has a significant effect on controlling the particle size.¹⁰⁸ Briefly, a larger population of smaller NPs can be obtained from good mixing conditions, while poor mixing produced larger NPs. The NP formation also relies considerably on the surface tension difference between aqueous phase (with high surface tension)

and organic phase (with low surface tension). The difference in surface tension leads to continuous solvent vortices formation at the interface of both liquids, and the organic solvent diffuses from low surface tension, causing gradual precipitation of the polymer (e.g. polysaccharides) on the organic surface and formation of nanocapsules.¹⁰⁹

Zhao and coworkers discussed three widely used nanoprecipitation methods in their work, which are traditional nanoprecipitation, flash nanoprecipitation, and micro-fluidic nanoprecipitation.¹⁰⁵ To control the size and morphological structure of the NPs prepared, key parameters such as mixing temperature, solvent/antisolvent ratios, and properties of the chosen polymers are very important. Operation of traditional nanoprecipitation is easy, quick, and affordable. However, it has a lack of control over mixing and has wider size distributions of large particle sizes.¹⁰⁵ Flash nanoprecipitation is a simple, rapid and scalable method. Turbulence-based intensive mixing can be achieved within few milliseconds by using flash nanoprecipitation mixers. Therefore, flash nanoprecipitation produces better NP quality with smaller NP size and narrow size distribution.^{105,110} The limitation of flash nanoprecipitation is that the particles produced do not have adequate stability for some applications.¹⁰⁵ Micro-fluidic nanoprecipitation is usually laminar flow-based and operated in continuous flow, its high-speed diffusion-based mixing (at milliseconds or even microseconds) results from a reduced mixing path (down to several tens of nanometres). By changing the configuration of the micromixer or controlling the ratio of flow rates, physicochemical properties of NPs (e.g. size, charge, morphology) can be controlled. However, an issue is system blocking caused by solid particle accumulation. Moreover, small volume and low flow rates in microfluidics result in relatively low productivity.^{105,111}

As a simple and rapid method, nanoprecipitation has been used to prepare various types of polysaccharide NPs. Tay and coworkers synthesised SNPs with sizes in the range between 300 nm and 400 nm by adding dissolved starch solution into excess ethanol under controlled conditions (Fig. 2a).¹¹² The presence of appropriate surfactant was also an important parameter in modulating the shape and size of the produced SNPs (Fig. 2b and c). NPs of hydrophobically modified CL were also synthesised by using nanoprecipitation method. Kulterer *et al.* prepared cellulose acetate (CA) NPs by developing an efficient nanoprecipitation technique based on using good CA solvents as a modifier for the nonsolvent.¹¹³ The size of NPs was reduced by sonication, efficient agitation, and optimisation of process parameters such as tetrahydrofuran (THF) content, temperature and pH value. Spherical NPs of 60 nm with high yield were obtained by adding good polymer solvents (THF) to an anti-solvent (water). A nanoprecipitation process for the formation of DEX NPs was described by Weiss and coworkers.¹¹⁴ Initially, DEX, PL and starch were methacrylated *via* methacrylic anhydride to enable crosslinking and the hydrophile-lipophile balance (HLB) value measured. In the next step, the modified polysaccharides were dissolved in acetone or THF, precipitated in water and crosslinked under UV light. The obtained particles were subjected to atomic force microscopy (AFM) measurements in order to



Scheme 3 Schematic representation of the nanoprecipitation method.



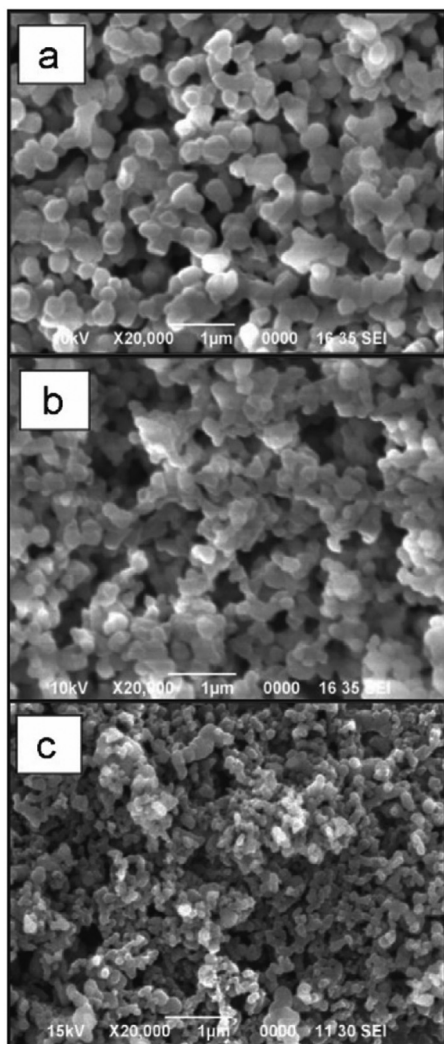


Fig. 2 Examples of polysaccharide NPs obtained by the nanoprecipitation method observed *via* Scanning Electron Microscopy (SEM) (scale bar 1 μm): (a) micrograph of precipitated SNPs prepared by addition of 1 mL of 1% starch into 20 mL of ethanol, (b) micrographs of precipitated SNPs prepared by addition of 1 mL of 1% starch into 20 mL of ethanol with 4% of cetrimeronium bromide surfactant and (c) micrographs of precipitated SNPs prepared by addition of 1 mL of 1% starch into 20 mL of ethanol with 4% of Tween 80 surfactant. (Reproduced with permission from ref. 112. Copyright Elsevier, 2011.)

obtain mechanical properties. Interestingly, the Young's moduli of particles correlated with the HLB value of the utilised polysaccharide, which might be due to the difference in water swelling for particles with different HLB value. Nicolas and coworkers utilised nanoprecipitation in the formation of polysaccharide capsules employing a template.¹¹⁵ Various polysaccharides were employed, *e.g.* DEX, PL, DS or HA, and crosslinked *via* isophorone isocyanate during the nanoprecipitation process. Mygliol as templating agent was dissolved together with the crosslinker in acetone and the polysaccharides added in aqueous solution at once. In addition to forming capsules from one polysaccharide component, blends were introduced as well as multilayers.

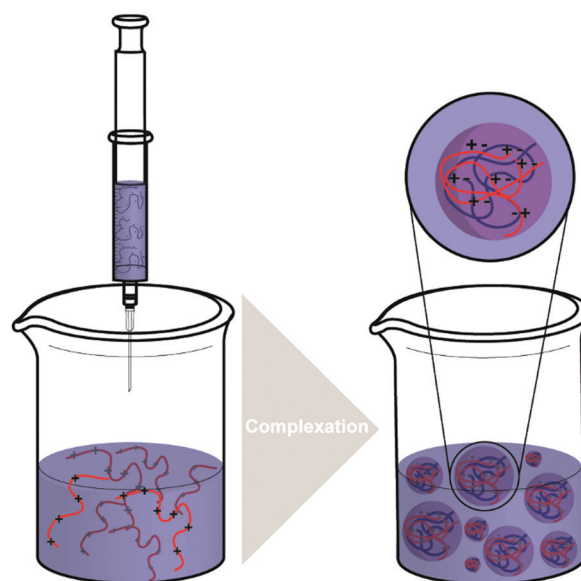
Overall, nanoprecipitation is easy to conduct with fast operation processing and simple equipment requirement. However, issues such as low productivity and system blocking during the NPs formation need more attention in future development.¹⁰⁵ Thus, improved control for fast processing is required and more complex structures have to be investigated in the future.

3.3 Complex coacervation

The complex coacervation method is carried out by mixing aqueous solutions of two polymers with opposite charges. It is a suitable method for the preparation of NPs due to its simplicity and mild preparation conditions without the need of organic solvents.¹⁹ To synthesise and enhance natural polysaccharide NPs, complex coacervation with oppositely charged polyelectrolytes has been widely used.¹¹⁶

The mixing of solutions containing oppositely charged polyelectrolytes (polycation and polyanion) (Scheme 4) establishes an equilibrium between the dense liquid phase and the dilute solution phase. The dense liquid phase is also called the coacervate, which is polyelectrolyte-rich and frequently produced from proteins and polysaccharides.^{117,118} According to the theoretical model of Voorn and Overbeek, the association of polyelectrolytes is significantly determined by the electrostatic interaction and the entropy gain during the complex coacervation process.^{118,119} Related to this, many important modes of polymer complex coacervation including the Voorn–Overbeek theory were reviewed by Sing.¹²⁰

An interesting feature of the complex coacervation process of polyelectrolytes, is that the net charge of the NPs formed can be modulated by adding one polyelectrolyte in excess to the other, giving a core–shell structure with the excess component segregated at the outer shell. The physical properties of the polyelectrolyte-complex formed by coacervation can be affected by many factors such as pH of the solution, temperature, ionic strength,



Scheme 4 The typical processes of forming polysaccharide NPs such as CS/HA NPs by complex coacervation process.



charge density, and the molar mass of polyelectrolytes.^{32,106} For instance, Delair and coworkers found improved stability of the polyelectrolyte-complex formed by DS (polyanion) and CS (poly-cation) by remaining the charge ratio R (n^+/n^-) under 0.6.¹²¹

For polyelectrolyte-complex formation, CS is the most commonly used cationic polysaccharide. Besides, various negatively charged polysaccharides such as HA, DS, and carboxymethyl cellulose can be combined with CS.³² Chen and coworkers prepared NPs made of oleoyl-carboxymethyl-chitosan (OCMCS) and HA for gene delivery using the coacervation process.¹²² The results obtained from their work showed that a charge ratio of 5 (n^-/n^+) and a weight ratio of 4 (OCMCS/HA) were the optimal conditions giving the smallest (165 nm), positively charged (+14.2 mV) and monodisperse NPs (Fig. 3a). Ferreira and coworkers developed a pH sensitive CS/DS NP system for the delivery of insulin using complex coacervation.¹²³ The optimum synthesis conditions had a DS:CS mass ratio of 1.5:1 at pH 4.8, and the stable spherical NPs produced had a mean diameter of 500 nm. It was also found that the insulin released by CS/DS NP system maintained its immunogenic bioactivity, indicating a bright future for the CS/DS NPs as a potential oral delivery system for insulin. Dou and coworkers¹²⁴ described the formation of complex coacervates as well. At first, CS/DEX nanoparticles were formed *via in situ* crosslinking and self-assembly. CS and DEX were dissolved in water and radicals formed on polysaccharide backbones *via* addition of cerium(IV) ions that initiated the formation of

poly(methyl acrylate) (PMA) and crosslinking *via* diallyldisulfide. Due to the formation of hydrophobic grafts an amphiphilic structure was generated that assembled into NPs. In a subsequent step, these particles were combined with HA to obtain a larger super structure *via* coacervation (Fig. 3b). The formation of multi-particle aggregates or HA-surface modified particles could be tailored *via* the ratio of HA and particle precursors. Finally, the particles were employed to target cancer cells *via* the lipid raft-dependent endocytosis pathway.

The influence of CS acetylation degree on the synthesis, stability, and performance of CS/HA NPs prepared by electrostatic complex coacervation was studied by Furlani *et al.*¹²⁵ A crosslinking mechanism mediated by TPP was involved because the inter-polyelectrolyte interaction between CS and HA alone was not sufficient to form spherical particles. The synthesis of NPs was carried out as follows. First, CS, sodium hyaluronate, and TPP were dissolved individually in deionised water. After solubilisation, HA/TPP solution was prepared by adding TPP solution dropwise to the hyaluronate solution. Then, the formation of NPs was conducted by adding the HA/TPP solution dropwise to CS solution under stirring. The results of their work proved that CS acetylation degree determines the NPs stability, and CS with specific fractions of acetylated units such as 0.16 and 0.63 were found to promote the formation of homogenous NPs. Recently, Tirelli and coworkers studied the formation of CS/HA-based nano particles with respect to the effects of template addition.¹²⁶ A two-step templating process was compared with a direct polyelectrolyte complexation approach. The two-step templating was performed *via* ionotropic gelation of CS with TPP and subsequent incubation with HA. It could be shown that HA can quantitatively exchange with TPP in the second step, which was accompanied with aggregation of particles. In contrast to the direct process of CS/HA complexation, larger particles were generated. Furthermore, the utilisation of low MW CS induced the formation of a HA corona.

Another key feature of complex coacervation is the preservation of the structure and property of biochemical drugs including genes and proteins, which is due to the mild conditions required for coacervate formation and their native environment.¹²⁷ In addition, the complex coacervation process enables the formation of polysaccharide NPs with high loading of biomolecular cargo.¹²⁸ Although there has been considerable progress in the theory of coacervation,^{120,129} the prediction of outcomes from the preparation procedure is challenging,¹³⁰ *e.g.* the control over NPs size and shape. This is mainly due to the various parameters in coacervate preparation like pH, ionic strength, polymer concentration, temperature and polymer type/MW. To improve the performance of NPs in the drug delivery system, the mixing of polysaccharide with other polymers can also be achieved using complex coacervation method.¹⁹ As such, access to additional features is possible, *e.g.* structures with poly(ethylene glycol) (PEG) corona.¹³¹

3.4 Emulsion-based methods

As a regular method for preparing polymer NPs,^{132,133} emulsion-based procedures are also a versatile method used for the synthesis

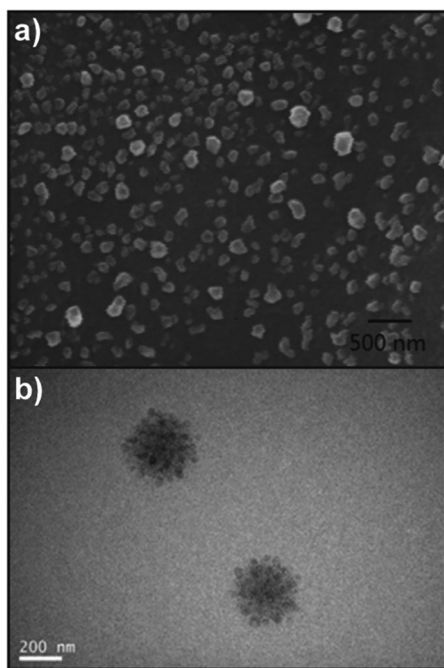
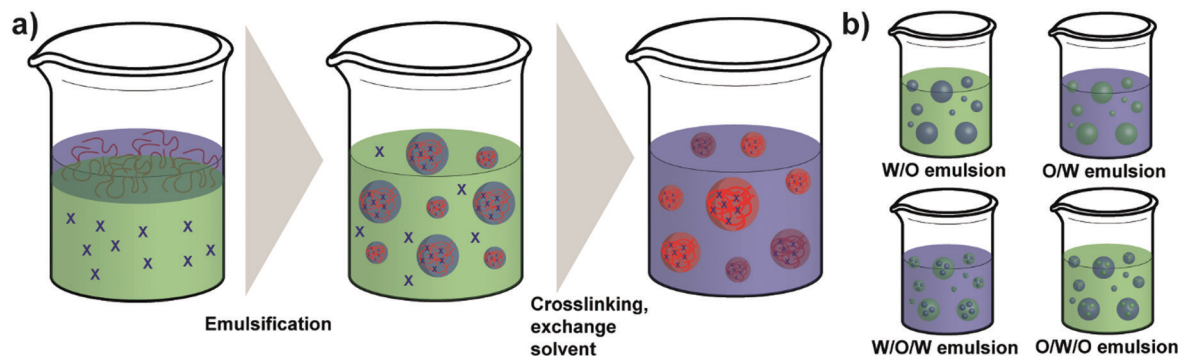


Fig. 3 Examples of polysaccharide NPs obtained by using the complex coacervation method: (a) oleoyl-carboxymethyl-chitosan (OCMCS)-hyaluronic acid (HA)/DNA NPs with a charge ratio of 5 (n^-/n^+) and a weight ratio of 4 (OCMCS/HA) (scale bar 500 nm) and (reproduced with permission from ref. 122. Copyright Elsevier, 2013) and (b) TEM image of HA/CS/DEX superstructure NPs formed *via* coacervation of CS/DEX NPs with HA. (Reproduced with permission from ref. 124. Copyright American Chemical Society, 2020.)





Scheme 5 Schematic overview over the emulsion process for polysaccharide NP formation: (a) formation of polysaccharide NPs via water-in-oil emulsion and crosslinking (external gelation) and (b) types of emulsions used to prepare polysaccharide NPs (green: organic solvent; blue: water).

of polysaccharide NPs.¹⁹ The term emulsification is defined as a process of forming a metastable dispersion (*e.g.* oil and water) also called emulsion that would separate into two phases in equilibrium. In biological systems different uses of emulsions are found, for example, lipoprotein forms an emulsion for the delivery and metabolism of fat in the body.^{30,134}

Generally, depending on the type of dispersed phase and dispersion medium, emulsions are categorised as O/W direct emulsion and water-in-oil (W/O) inverse emulsion, in which water or oil is the continuous phase, respectively. Besides, more complex multiple emulsions such as oil-in-water-in-oil emulsion and water-in-oil-in-water emulsion can also be obtained by dispersing the O/W or W/O emulsions in oil or water using specific techniques (Scheme 5).¹³⁴ In addition, W/W emulsions can be formed as well (refer to Section 4.3).^{135,136} Considering the size of droplets, emulsions formed can be classified as microemulsion, miniemulsion and macroemulsion. Microemulsions are thermodynamically stable with droplet sizes ranging from 10 to 100 nm. Miniemulsions and macroemulsions are both thermodynamically unstable with droplet sizes between 100 nm and 1 μm and larger than 1 μm , respectively.³⁰

Different from the nanoprecipitation method mentioned above, which is a simple one-step method, preparation of organic NPs such as polysaccharide NPs using emulsification method is based on a two-step procedure. At first the emulsion is prepared as mentioned above and subsequently, NPs can be generated from the emulsion via various methods, such as solvent evaporation, solvent diffusion, and reverse salting-out.¹³⁷ In particular, the formation of polysaccharide NPs by emulsification method is conducted as follows: a solution of polysaccharide is firstly formed by dissolving polysaccharides in deionised water or other solvents. Secondly, the solution is dispersed in an oil phase to fabricate O/W or W/O emulsions by stirring or employing ultrasound. Polysaccharide NPs can finally be prepared by internal or external gelation.¹⁹ In external gelation, the crosslinkers diffuse from an external source into the polysaccharide emulsion, *i.e.* from the continuous phase. While for internal gelation, the crosslinkers are already present inside the droplets before gelation occurs.¹³⁸

To obtain the emulsification system, low-energy emulsification techniques and high-energy emulsification techniques are

commonly used. The former one is based on the expansion of low-energy stirring routes in constant progress. It can be classified as spontaneous emulsification, emulsion inversion point method, and phase inversion temperature method. For high-energy emulsification, high-energy mechanical stirring processes such as sonication and microfluidic methods are employed, the applied stress predominately determines the size of the nanodroplets produced.³⁰ Ravi *et al.* successfully prepared Lopinavir loaded pullulan acetate NPs (PANPs) for oral delivery using O/W emulsion-solvent-evaporation method in their study.¹³⁹ Firstly, Lopinavir and pullulan acetate were dissolved in methylene chloride giving the organic phase. Subsequently, to form the emulsification system, the organic phase was added dropwise to an aqueous poly(vinyl alcohol) (PVA) solution under gentle magnetic stirring, where PVA acts as steric stabiliser for the PANPs. After a 15 min high-speed homogenisation, the primary emulsion was centrifuged to obtain the PANPs. Further purification was finally conducted to remove the excess free drug and PVA before storage. According to the results of their work, the size of PANPs prepared was about 197 nm, high entrapment efficiency ($\sim 75\%$) and monodisperse particles were also obtained. Moreover, a tissue distribution study proved that Lopinavir loaded PANPs could be a remarkable approach in treating HIV infection.

In the study of Park's group, doxorubicin (DOX) loaded HA-based nanocarriers for the intracellular DOX delivery were synthesised by using the O/W emulsion method (Fig. 4a).¹⁴⁰ The HA-SS-DDT-DOX NPs formed were based on an amphiphilic HA derivative, the HA-SS-DDT, which was prepared by modifying HA with dodecanethiol (DDT) via disulfide linkage. The primary O/W emulsion was prepared by adding a solution of DOX HCl and triethylamine in chloroform, into the aqueous HA-SS-DDT solution. After evaporation of chloroform, the solution was filtered, dialysed, and lyophilised to obtain HA-SS-DDT-DOX NPs with a diameter around 334 nm. It was found that the synthesised NPs were stable under physiological conditions and DOX was effectively encapsulated with loading efficiency $> 70\%$. Moreover, the HA-SS-DDT NPs were effectively taken up by squamous cell carcinoma (SCC7) cells followed by fast release of DOX, indicating the promising future of HA-SS-DDT NPs as a potential DOX carrier (Fig. 4b). Sagis and coworkers utilised sodium alginate to form NPs via an emulsion process.¹⁴¹



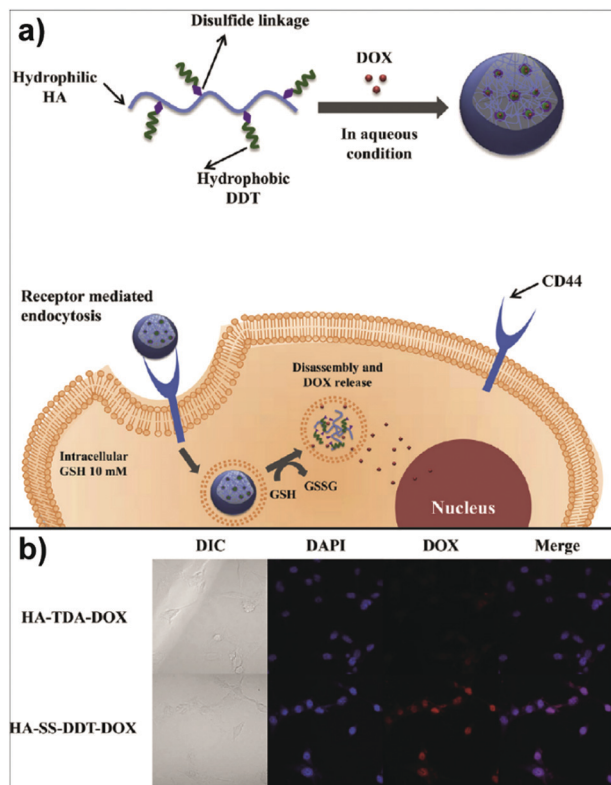


Fig. 4 Polysaccharide NPs obtained by using the emulsification method: (a) schematic approach of HA-SS-DDT-DOX utilisation in cancer treatment and (b) fluorescence microscopic images of SCC7 cells treated with HA-TDA-DOX and HA-SS-DDT-DOX showing the release of DOX in cells in the case of HA-SS-DDT-DOX. (Reproduced with permission from ref. 140. Copyright Elsevier, 2014.)

Alginate was dissolved in water and the aqueous solution emulsified in an oil with polyglycerol polyricinoleate as surfactant. Then, CaCl_2 nanoparticles were added to induce an external gelation process of the alginate *via* Ca^{2+} /alginate complexation.

Another kind of HA-based NPs were prepared by Min *et al.* with the O/W emulsion method.¹⁴² In aqueous conditions, HA-5 β -cholanolic acid conjugates were firstly used to form HANPs by self-assembly. Subsequently, a bioinert and hydrophobic perfluoropentane (PFP) solution was added to the prepared HANPs and emulsified to fabricate PFP-loaded HANPs. The analysis results demonstrated that the PFP-loaded HANPs were stable and robust and had a homogeneous as well as narrow size distribution with an average diameter of 350 nm. Sriamornsak and coworkers described the formation of dextrin NPs *via* an inverse emulsion approach.¹⁴³ Water-in-hexane emulsions were employed and dextrin crosslinked with glyoxal. Particle sizes were tailored *via* sonication time, surfactant type and concentration as well as HLB value. In a similar way, Gupta and coworkers formed nanoparticles from PL with sizes below 50 nm.¹⁴⁴

Emulsification methods have a considerable advantage in controlling the size and loading of the synthesised polysaccharide NPs due to the convenient adjustment of emulsion droplet sizes and the option to include cargoes in specific phases of the emulsions. Nevertheless, emulsion methods are more complicated

than nanoprecipitation and complex coacervation caused by the additional step of emulsification. Furthermore, the requirements of large amounts of organic solvents limit the utilisation of emulsification method.¹⁹

3.5 Other methods

In addition to the three important methods introduced above, there have been other practical methods such as desolvation and dialysis, developed by researchers for preparing polysaccharide NPs, which are also appropriate in producing stable and mono-disperse polysaccharide NPs with controllable size.¹⁹

When using the desolvation method, a desolvation factor such as salts or alcohols is slowly added into the solution of macromolecules to induce polymer precipitation.³⁰ For making drug nanocarriers, the selection of desolvating agents is based on the nature of the drug to be encapsulated inside the polysaccharide NPs.¹⁹ Amoabediny and coworkers synthesised curcumin loaded CSNPs and SNPs using the desolvation method in their work.¹⁴⁵ The desolvating agent was composed of absolute ethanol and predetermined concentrations of curcumin. The CSNPs and SNPs were formed by ethanol precipitation of an aqueous CS/starch solution. Mean sizes of the synthesised curcumin loaded CSNPs and SNPs were 66 and 61 nm with a maximum loading efficiency of 12% and 14%, respectively. The desolvation method can produce NPs directly in suspension in an aqueous medium, however, a further purification step is required because of the potential toxicity caused by desolvating agents.¹⁹

Related to the nanoprecipitation method, a solvent displacement mechanism is used in the dialysis method. Additional tools such as dialysis tubes or semi-permeable membranes are utilised to provide a physical barrier with appropriate molar mass cut off for polymers/polysaccharides.³⁰ Zhang and coworkers synthesised paclitaxel (PTX) loaded HA NPs by using dialysis method.¹⁴⁶ HA was modified first by coupling with aminated retinoic acid. Next, PTX solution in ethanol was added into the solution of retinoic acid modified HA (HRA). The obtained solution was then dialysed against distilled water, and the PTX-loaded HRA NPs were finally collected after filtration and lyophilisation. The particle size of the prepared PTX-loaded HRA NPs was around 149 nm, and the inhibition effect showed by PTX-loaded HRA NPs on tumour growth proved its future application in cancer chemotherapy. Compared to nanoprecipitation, the dialysis method relies on a slow exchange of solvent and thus more likely generates thermodynamic products.

Synthesis of NPs using ionic gelation is regularly performed with charged polysaccharides in aqueous medium in very dilute solution. For instance, CSNPs can be produced by ionic gelation because it is positively charged at neutral pH and can form NPs by crosslinking with small molecules (*e.g.* the monomers to form polymers) with opposite charge *via* electrostatic interaction.^{19,30} Alonso and coworkers successfully entrapped DOX into CSNPs through the ionic gelation of CS with TPP.¹⁴⁷ CS-DOX complex was firstly prepared by adding DOX into the CS aqueous solution at pH 5.5. Subsequently, a suitable amount of TPP was mixed with the CS-DOX complex to form the DOX loaded CSNPs. According to the results obtained from their study, though the complexation efficiency was low, no complex dissociation was



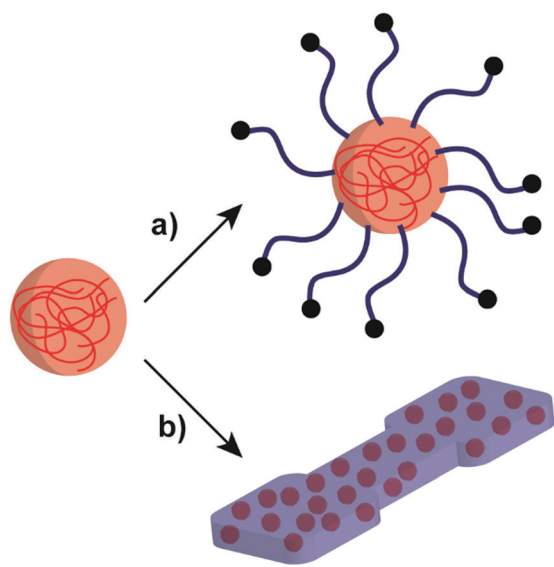
found after the formation of NPs. Moreover, the usefulness of CSNPs as the DOX carrier and to transport it into the cells in its active form was demonstrated.

Spray-drying is a technique notably used in the production of powders, granules or agglomerates from the mixture of drug and excipient solutions as well as suspensions.^{19,87} A typical spray-drying process is based on drying of aerosol droplets in contact with a hot drying gas to evaporate the moisture and to form the solid product.³⁰ Li *et al.* described the preparation of NPs formed by modified starch in their study.¹⁴⁸ The results showed that state-of-the-art spray drying equipment could lower the size of the produced dried particles by an order of magnitude attaining submicron sizes. Also, there are many factors possible to affect the size of the NPs produced. In the review of Agnihotri *et al.*,⁸⁷ various parameters such as the size of the nozzle, spray flow rate, and atomisation pressure are discussed that can be controlled to obtain desired the particle size.

Including what has been introduced above, there are many other practical techniques for polysaccharide NP synthesis that complement the most common methods and cannot be covered in the present review. Overall, further studies are still required to develop more efficient methods/techniques in the future.

4. Polysaccharide nanoparticles and polymers

Developing NP systems formed by polysaccharides and polymers have potential applications in many areas such as drug delivery, Pickering emulsifiers, and improving the properties of polymer-based materials.^{70,149–151} As such, polysaccharide/polymer NP systems combine the advantages of both polysaccharides and polymers, bringing benefits to overcome difficult challenges that belong to the individual materials (Scheme 6).



Scheme 6 Schematic representation of the polysaccharide NP incorporation with polymer systems: (a) polymer-grafted polysaccharide NP and (b) polysaccharide NPs in a polymer matrix.

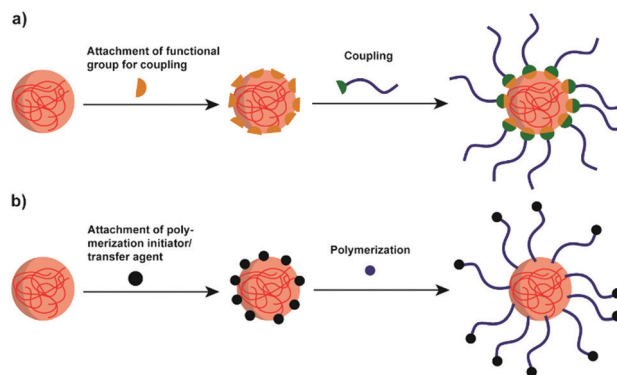
For example, poly(ethyl acrylate) (PEA) or PMA polymers can introduce flexibility to rigid starch-based materials for further utilisation.^{152,153}

This section will mainly focus on the methods of attaching polymers to polysaccharide NPs and inclusion of polysaccharide NPs into polymer structures. The system formed by the inclusion of polysaccharide NPs into the bulk of polymers is also known as a typical core-shell-structured hybrid or composite material. The core-shell structure is based on a sufficient modification of the surface of polysaccharide NPs by synthetic polymers, giving a polysaccharide NP core and a polymer shell.¹⁵⁰ Moreover, in some cases, hydrophobic polymer NPs can also be used to form a core modified by outer hydrophilic polysaccharides. In the field of drug/gene delivery, the hydrophobic-hydrophilic core-shell structure plays a very significant role in enhancing the stability, water solubility and biocompatibility of NP structures.^{154–156} The polymer core-polysaccharide shell structure will not be discussed in detail in this section as it is not directly related to polysaccharide NPs, more related information of how the core-shell structure with a polysaccharide shell is utilised can be found in the literature.^{157,158}

4.1 Polymers attached to polysaccharide NPs

Though polysaccharide NPs have been widely investigated in many research areas, for several applications, their hydrophilicity precludes further use of them. For example, to enhance the properties of hydrophobic polylactide (PLA), starch/PLA composites have been considered as an efficient choice. However, the hydrophobic PLA is immiscible with native hydrophilic starch, causing incompatibility and weak mechanical properties.¹⁵³ Thus, improved compatibilisation *via* polymer attachment is a considerable direction.

To effectively combine the advantages of polysaccharide NPs and polymers, graft modification/copolymerisation methods are necessary. Two types of grafting reactions can be conducted to form the polymer grafted polysaccharide system, *i.e.* the “grafting to” and “grafting from” reactions.¹⁵⁹ In “grafting to” reactions (Scheme 7a), functionalised polymers are prepared in



Scheme 7 Schematic representation of the two grafting approaches. (a) The “grafting to” approach involving the coupling between the polysaccharide NPs and the pre-formed polymer chains (blue line). (b) The “grafting from” approach relying on polymerisation initiated at the NP surface (initiator/transfer agent: black sphere; monomer: blue sphere).



advance to be reacted with functional groups present on the polysaccharide interface/polysaccharide backbone. Moreover, the polymer chains to be grafted can be fully characterised before attachment. In “grafting from” reactions (Scheme 7b) the polymerisation of monomers takes place directly on the polysaccharide interface started from the polysaccharide backbone, which minimises the steric hindrance. In this case, the full characterisation of the grafted chain becomes challenging if not impossible.¹⁶⁰

According to Cunningham and coworkers, SNPs could be modified with synthetic polymers by using the two grafting reactions mentioned above *via* nitroxide-mediated polymerisation.^{31,159} The “grafting from” approach was carried out in three main steps under inert conditions, which are SNP-macroinitiator preparation, “grafting from” reaction, and acid hydrolysis (purification). Graft modified SNP with poly(methyl methacrylate-*co*-styrene) (P(MMA-*co*-S)), PMA, and poly(acrylic acid) were successfully obtained. In addition, the feasibility of varying the amount of grafted polymer on the SNP surface was demonstrated.³¹

As well as SNPs, CSNPs were also studied for forming core-shell-structured hybrids. In the work of Tang *et al.*, two types of polymers were investigated to modify the surface of CS nanospheres (spherical CSNPs) *via* an ATRP “grafting from” process.¹⁶¹ The used polymers were the homopolymer poly(methyl methacrylate) (PMMA) and amphiphilic block copolymer PMMA-*b*-poly(poly(ethylene glycol) methyl ether methacrylate) (PMMA-*b*-P(PEGMA)). The obtained structure had a CS nanosphere core and a densely grafted outer PMMA or PMMA-*b*-P(PEGMA) layer. The CSNP-*g*-(PMMA-*b*-P(PEGMA)) core-shell structure described the inclusion of a polysaccharide core in multipolymer layers. According to the results of the stability studies, the block copolymer grafted nanospheres CSNP-*g*-(PMMA-*b*-P(PEGMA)) were more stable in both THF and deionised water compared to the Br functionalised CSNP and CSNP-*g*-PMMA, indicating the potential use for the synthesis of CSNPs based core-shell structure for *in vivo* biomedical applications and industrial-scale polymerisation.

Yuan *et al.* grafted poly(hydroxy ethyl methacrylate) (PHEMA) from crosslinked CS microspheres *via* an ATRP approach.¹⁶² There, CS spheres were conjugated with an ATRP initiator and HEMA polymerised. In the following step, the hydroxyl groups from the HMA units were exploited to attach melamine in amide formation *via* a carbonyl diimidazole activation. As such, microspheres were obtained that featured considerable and rapid Cu(II) adsorption from aqueous solution. Moreover, the microspheres could be recycled *via* exchange with H⁺ ions. Another route for the formation of polymer decorated CS microspheres was presented by Peixão *et al.* cerium(IV) initiation was employed to graft (*N*-isopropylacrylamide) (NIPAM) directly from the crosslinked microspheres.¹⁶³ Additionally, the microspheres were embedded with magnetite nanoparticles to introduce magnetic features. The final microspheres featured thermo-responsivity due to PNIPAM and could be easily separated from liquid media *via* magnetic field. In particular, the system was utilised to remove oil from water, *i.e.* the microsphere material was added to oily water to capture the oil and separate with a

magnet. At ambient temperature a removal efficiency of 85% was observed, while an efficiency of 97% was observed above 40 °C. This difference is due to the fact that the PNIPAM is more hydrophobic above its lower critical solution temperature and thus more susceptible to absorb oil.

Similarly, “grafting to” approaches were also investigated to obtain grafted polysaccharide NPs, *e.g.* SNP under inert conditions.¹⁵⁹ SNP was firstly functionalised with 4-vinylbenzyl chloride (VBC), and in a separate step, the *N*-*tert*-butyl-*N*-[1-diethylphosphono-(2,2-dimethylpropyl)nitroxide] (SG1)-capped P(MMA-*co*-S) was synthesised. Finally, the prepared SNP-VBC and SG1-capped P(MMA-*co*-S) were reacted to form graft modified SNP. The “grafting to” approach allows accurate and precise characterisation of the precursors and the grafted SNP. Wang and coworkers synthesised core-shell NPs based on an SNP-based core and a PNIPAM brush shell using surface-initiated single-electron transfer living radical polymerisation (SI-SET-LRP) in their work.¹⁵⁰ SNPs were firstly initiator-modified by Br groups using an esterification reaction (Fig. 5a). Subsequently, PNIPAM chains were grafted to initiator-modified SNPs by using SI-SET-LRP. Compared with the initial SNPs, the SNP-*g*-PNIPAMs revealed an evolution in particle morphology (Fig. 5b-i), proving the successful synthesis.

A “grafting-to” approach was employed by Lin and coworkers to form polysaccharide NPs with heparin core and PEG shell.¹⁶⁴

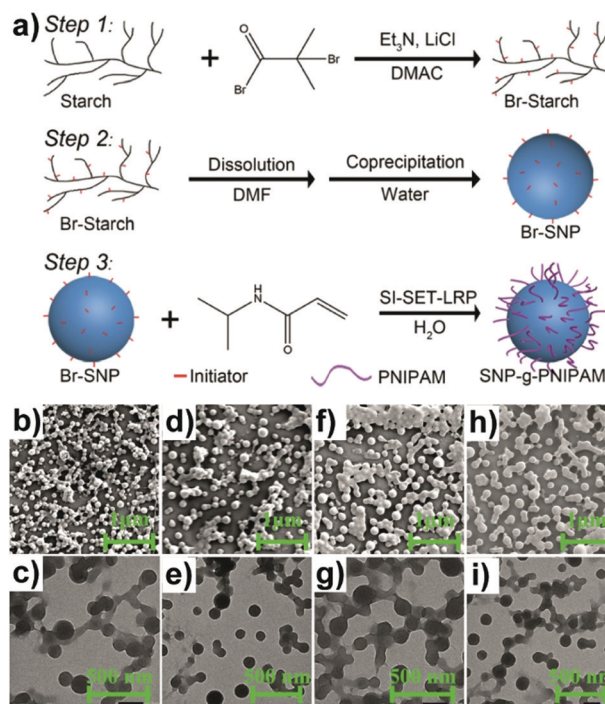


Fig. 5 (a) Schematic overview of PNIPAM grafting from SNPs with the degree of substitution of the Br group around 1.1 per saccharide unit, (b) and (c) SNP-*g*-PNIPAM after 15 min (SEM top: scale bar 1 μ m, TEM bottom: scale bar 500 nm), (d) and (e) SNP-*g*-PNIPAM after 30 min (SEM top, TEM bottom), (f) and (g) SNP-*g*-PNIPAM after 60 min (SEM top, TEM bottom), (h) and (i) SNP-*g*-PNIPAM after 120 min (SEM top, TEM bottom). (Reproduced with permission from ref. 150. Copyright American Chemical Society, 2019.)



The anionic polysaccharide heparin was combined with poly-(lysine) to form NPs *via* coacervate formation under ultrasound treatment. To enable conjugation with PEG, the heparin chains were modified with tetrazine units. Finally, norbornene end functionalised PEG was added to the NPs to enable grafting. Subsequently, cell interactions with functionalised NPs were investigated, where the grafting could be used to tailor macrophage uptake *in vitro*.

Another avenue to obtain polysaccharide NPs with polymer-covered surfaces is *via* self-assembly approaches.¹⁶⁵ For example, Park and coworkers grafted PEG and 5 β -cholanic acid onto HA.¹⁶⁶ Due to the hydrophobic 5 β -cholanic acid and the hydrophilic PEG, an amphiphilic structure was obtained that was capable of forming micellar particles in aqueous environment with PEG on the surface. The formed particles featured reduced uptake in the liver, prolonged blood circulation, and improved tumour targetability due to the PEG functionalisation, which was shown *via in vivo* cell internalisation studies. A similar strategy was implemented by various researchers in the past years to obtain PEGylated particles for applications in the biomedical field, *e.g.* DOX delivery,^{167–171} theranostic NPs to target colon cancer,¹⁷²

as a stabiliser for deacetyl mycoepoxydiene nanocrystals,¹⁷³ gene delivery^{166,174,175} or sorafenib delivery.¹⁷⁶ To enhance the stability of the formed particles, Park and coworkers introduced covalent crosslinking.¹⁷⁷ In addition to PEG and 5 β -cholanic acid, methacrylamide units were grafted onto HA. The methacrylamide units were crosslinked *via* UV light after encapsulation of PTX. Sustained release of the drug was observed as well as improved cell internalisation leading to an overall enhanced therapeutic efficacy. Later on, a bioresponsive system was introduced (Fig. 6),¹⁷⁸ where HA was grafted with PEG that was attached *via* a matrix metalloproteinase 9 (MMP9) cleavable connection (pep) as well as conjugated to ovalbumin (OVA) as foreign antigen. In such a way, the PEG corona facilitated efficient transport but was cleaved at the tumour site *via* MMP9 enabling cell uptake leading to foreign antigen presentation on tumour cells. As such, tumour growth was inhibited considerably.

PEGylated DEX NPs were synthesised by Xiao and coworkers, who grafted PEG onto DEX in the first step and crosslinked *via* 3,3'-dithiodipropionic acid in the second step.¹⁷⁹ Next, DOX was encapsulated and drug-release studied *in vitro*, where the disulfide-based crosslinker can be cleaved *via* a redox stimulus.

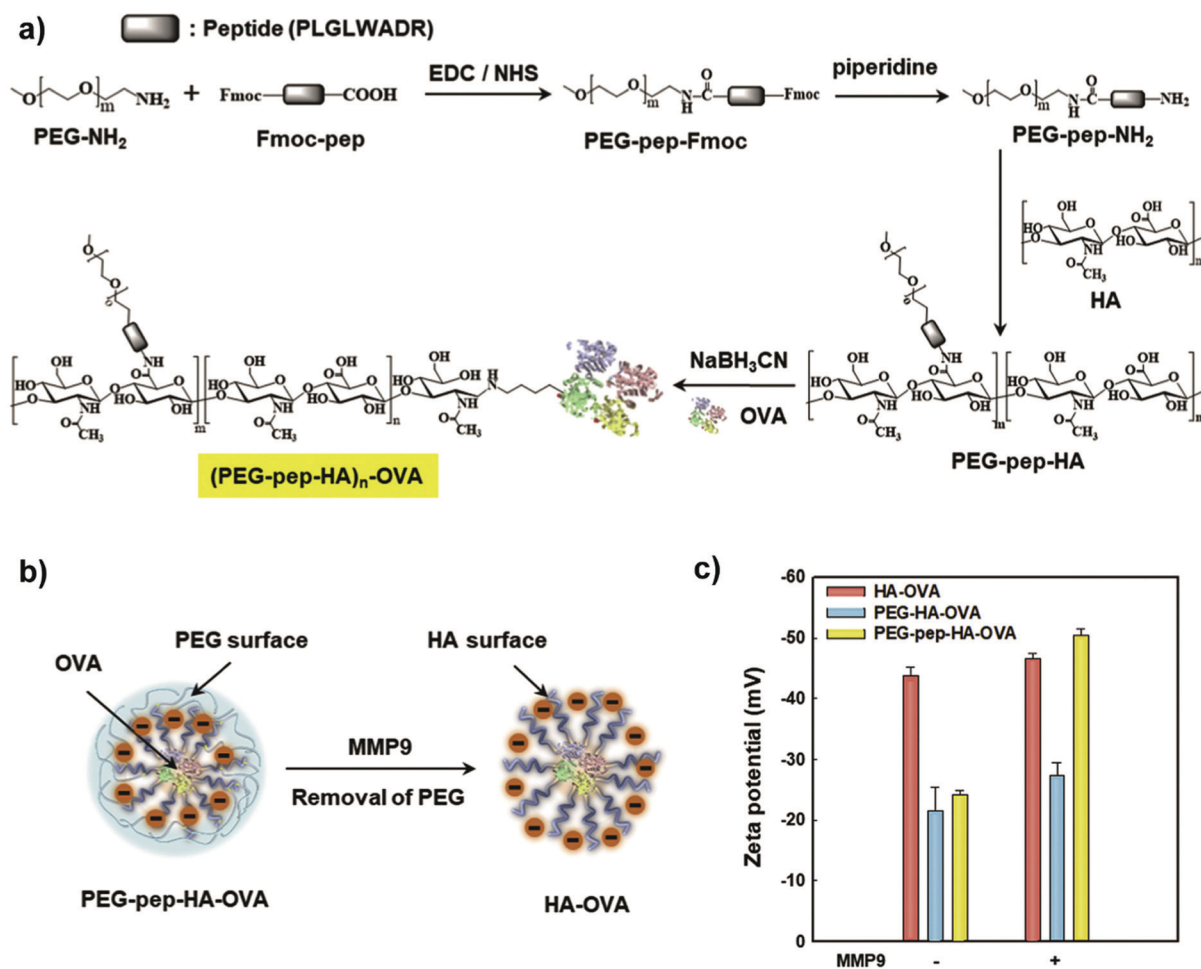


Fig. 6 Preparation and characterisation of the PEG-pep-HA-OVA conjugate: (a) synthetic route, (b) schematic illustration of MMP9-responsive removal of PEG from the PEG-pep-HA-OVA and (c) zeta potential values of the PEG-pep-HA-OVA with or without MMP-9. (Reproduced with permission from ref. 178. Copyright Elsevier, 2017.)



Pérez-Álvarez and coworkers described the formation of HA-based nanogels crosslinked *via* PEG spacers in a microemulsion approach.¹⁸⁰ The formed nanogels featured pH response due to the HA content, which was revealed by the swelling ratio at different pH. Wich and coworkers fabricated NPs based on spermine functionalised acetalated DEX.¹⁸¹ In a subsequent step, PEG or DEX were grafted onto the NP surface and their stability studied over time. In contrast to PEG grafting, DEX functionalised NPs aggregated over time. With respect to biological activity, PEG grafting led to attenuated particle binding and uptake. DEX grafting showed an opposite effect with enhanced passive targeting of dendritic cells and their stimulation. This combination is desired for vaccination avenues based on dendritic cells aiming to deliver an antigen and an adjuvant to dendritic cells at the same time in order to induce immune response.

Also, CS-based NPs were PEGylated in order to improve circulation in the body and stability. CS-coated nanocapsules were PEGylated by de la Fuente and coworkers.¹⁸² Initially, a nanoemulsion method was utilised to form CS capsules with lipophilic core (oleic acid). In the next step, the formed capsules were grafted with PEG *via* amide bonds. The authors showed that the PEG grafting hindered unspecific protein adsorption, *in vitro* studies showed low cytotoxicity and considerable internalisation in Vero cells. Bikiaris and coworkers synthesised PEG-grafted CS for the use as protein delivery system.¹⁸³ For the particle formation, coacervation with poly(glutamic acid) or ionic crosslinking with TPP was utilised. As protein to deliver, bovine serum albumin was encapsulated in the NPs. The observed release characteristics could be tailored *via* PEG MW, crosslinker content and type. Increased amounts of PEG on the particle surface and increased MW led to a faster release, while crosslinking with the small molecule TPP slowed the release down.

PEGylated CS NPs for the delivery of PTX were studied by Jain and coworkers.¹⁸⁴ At first PTX loaded CS NPs were formed *via* solvent evaporation method together with PVA. In the next step PEG was grafted onto the particles *via* amide formation and the remaining hydroxyl group at the other end of the PEG chain was used to conjugate transferrin. As such, the particles featured a drug, a stabilising corona and a cancer cell targeting moiety (transferrin). A considerable therapeutic effect was observed *in vivo* due to sustained release profile, greater intracellular uptake, promoted cell cytotoxicity and prolonged systemic circulation time. Hartmann and coworkers utilised CS NPs for gene delivery.¹⁸⁵ At first, CS was grafted with PEG and then combined with unfunctionalised CS as well as small interfering RNA (siRNA) in slightly acidic environment. Due to the negative charge of siRNA and the positive charges of CS, polyplexes were formed, similar to the coacervation technique. The formed NPs featured the cargo, *i.e.* siRNA in the core and PEG in the shell. The authors studied the effect of PEGylation degree on NP properties. Interestingly, physicochemical properties revealed only minor changes, while biological activity differed significantly pointing towards an optimum between PEG stabilisation of NP transport in the body and activity of the carrier in gene transfection. Also, other cases of PEGylated CS-based NPs were utilised for gene delivery, *e.g.* for the delivery of siRNA.^{186–188}

A nanoprecipitation avenue was utilised by Ganachaud and coworkers to form core-shell particles.¹⁸⁹ A polysaccharide (PL or DEX) was combined with a glycopolymer, *i.e.* poly(*N*-[7-(α -D-mannopyranosyloxy)heptyl]methacrylamide-*co*-glycidyl methacrylate), and mygliol. The polysaccharide and glycopolymer were dissolved in water and precipitated into acetone containing mygliol as a template for the core and crosslinked *via* isophorone diisocyanate. The technique enabled formation of mono- or multi-layered glyco nanocapsules through simultaneous or sequential nanoprecipitation, respectively. The prepared nanocapsules could be loaded with a drug (Camptothecin), covered with biotin or magnetic nanoparticles attached. Due to the incorporation of DEX, enzymatic degradation of the capsules could be achieved *via* dextranase.

An integrated self-assembly of polysaccharide-based double hydrophilic block copolymers and crosslinking was described by Schmidt and coworkers.¹⁹⁰ A completely hydrophilic block copolymer of PL and P(PEGMA) was synthesised and the self-assembly studied in solution showing considerable effect from the length of the P(PEGMA) block. In a subsequent step, PL was crosslinked *via* sodium trimetaphosphate to obtain stable NPs that could withstand dilution and organic solvent. In a similar way, the block copolymer of PL and poly(*N,N*-dimethylacrylamide) was studied,⁹¹ which showed particle formation. In order to stabilise the structures, crosslinking was performed as well. In this case, the PL block was oxidised with periodate to form aldehyde functions along the backbone that were cross-linked *via* imine formation with cystamine. As such, NPs were synthesised that could be broken *via* pH or redox stimulus. Akiyoshi and coworkers described the formation of vesicles based on polymer grafted polysaccharides.¹⁹¹ PL was grafted with poly(propylene oxide) and the thermoresponsivity of the PPO blocks used to turn vesicle formation on or off. The vesicle size could be controlled by polymer concentration.

The examples shown above from previous developments of polysaccharide NP/polymer systems describe how polymer chains can be attached to the surface of polysaccharide NPs and how a combination of polymers and polysaccharides can improve the applicability of polysaccharide NPs. Future directions certainly will consider other polymer shells and incorporation of polymers into polysaccharide NPs as a way to tailor NPs properties. However, issues such as toxicity and poor biodegradability caused by polymers with high molar mass limit the applications of these conjugated systems, which has to be addressed in future research.⁴³

4.2 Inclusion of polysaccharide NPs into polymer structures

The inclusion of a polysaccharide NP core into a polymer shell or matrix (Scheme 6b) has received much attention due to the wide applications in diverse areas.¹⁵⁰ The synergistic effect resulting from combining the properties of the core and the shell imparts new functionality to the final obtained material.¹⁵³

Wang *et al.* prepared core-shell NPs which had a SNP hard core and a PEA outer shell to strengthen the toughness of PLA-based nanocomposites.¹⁵³ Native starch was firstly esterified with acryloyl chloride and stearyl chloride to introduce hydrophobicity



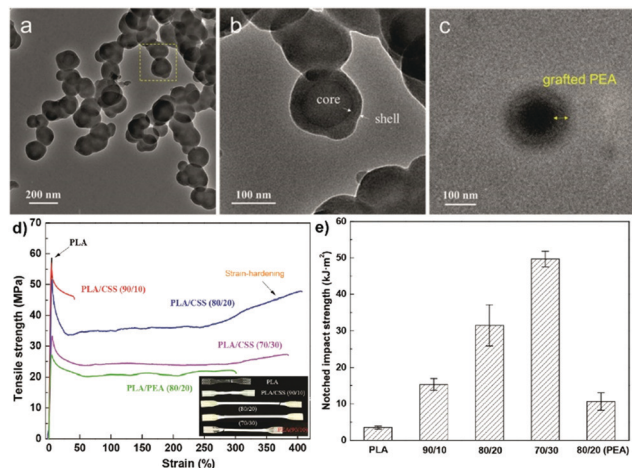


Fig. 7 (a–c) TEM images of SNP-*g*-PEA core-shell NPs, (d) stress–strain curves of PLA and PLA/SNP-*g*-PEA core-shell NP composite (inset a shows optical photo of the samples), (e) notched impact strengths of PLA and PLA/SNP-*g*-PEA core-shell NP composite. (Reproduced with permission from ref. 153. Copyright American Chemical Society, 2018.)

and polymerisable units. After that, a graft emulsion polymerisation was followed using potassium persulfate as the initiator. By using the “grafting from” method, ethyl acrylate monomers were polymerised from the surface of SNPs, forming the SNP-*g*-PEA core-shell NPs (Fig. 7a–c). Next, the particles were combined with PLA *via* melt-blending. Compared with pure PLA, elongation at break and notched impact strength were significantly improved after adding 20 wt% SNP-*g*-PEA core-shell NPs (Fig. 7d and e). The same authors also substituted PEA with poly(glycidyl methacrylate), where the epoxy group can act as a crosslinking point for PLA end groups *via* ring-opening.¹⁹² In a recent follow-up work, the same group showed even enhanced mechanical properties of blended PLA by implementation of two polymer layers around the SNPs, namely PEA and PMMA, formed *via* emulsion polymerisation.¹⁹³

By using a similar emulsion polymerisation synthesis method, Liu *et al.* synthesised SNP-based core-shell NPs with a PMA outer shell to improve the mechanical properties of poly(propylene carbonate) (PPC).¹⁵² The benefits resulting from the addition of 20 wt% SNP-*g*-PMA core-shell NPs to neat PPC were shown *via* enhanced tensile strength, increased toughness, and increased thermal degradation temperature for the PPC/SNP-*g*-PMA composite. Moreover, the increased glass transition temperature (T_g) demonstrated that PPC/SNP-*g*-PMA composite had higher rigidity compared to that of the neat PPC, providing a direction for a more comprehensive modification and a wider application range of PPC. Recently, SNP-*g*-PMA was utilised as compatibiliser for blends of PPC and PLA.¹⁹⁴ The blends were formed *via* melt-blending and the compatibilisation was investigated *via* AFM, dynamic scanning microcalorimetry and rheology. Improved mechanical and thermal properties were obtained, *e.g.* compared to a control blend PPC/PLA (60/40), the elongation at break of PPC/PLA/SNP-*g*-PMA (60/40/20) was enhanced from 15% to 272% without demoting tensile strength. Additionally, the heat distortion temperature of PPC/PLA/SNP-*g*-PMA (60/40/20) was increased by 21 °C to 47 °C compared to PPC. Jiang and coworkers presented an

avenue to enhance the mechanical properties of poly(propylene) (PP) *via* incorporation of SNPs.¹⁹⁵ Thermoplastic starch acetate and PP were melt-blended, leading to SNP formation *in situ* due to the addition of poly(ethylene octane) grafted with maleic anhydride. As such, higher notched impact strength and elongation at break of the blend material compared to the plain PP were observed.

For the further use of the inclusion of polysaccharide NPs in polymer structure, Somavarapu and coworkers described a particle-within-particle system formed by CS-based NPs such as CS/TPP NPs incorporated in polymer microparticles.¹⁹⁶ By using a modified W/O/W double emulsion method, Ranibizumab was loaded into CSNPs with a size of 17–350 nm that were entrapped in poly(D,L-lactide-co-glycolide) (PLGA) microparticles. The CSNPs loaded PLGA microparticles significantly showed improved Ranibizumab loading and release profile, demonstrating the promising future for this novel system as an adequate nano-carrier. Teng and coworkers presented the fabrication of PLGA and poly(cyclohexane-1,4-diyl acetone dimethylene ketal) microspheres with included HA/CS NPs. A double emulsion process was performed to obtain the composite microspheres. HA/CS NPs were loaded with siRNA as a treatment for rheumatoid arthritis. The deposition of siRNA loaded particles in the polymer matrix prevented siRNA degradation and improved circulation time *in vivo*. Nevertheless, the observed release was not maintained over 8 days, which is needed as rheumatoid arthritis is a chronic disease requiring long-term treatment. The characteristics of the carrier showed interesting and effective delivery properties and might be useful for the treatment of other diseases but further optimisation is needed.

Berkland and coworkers utilised NPs based on polysaccharides and PLGA for hydrogel formation.¹⁹⁷ Alginate and CS containing PLGA NPs were generated *via* coprecipitation first. In the next step, the oppositely charged NPs were combined to form hydrogels *via* mixing and charge-based attraction of particles. The gels proved to be sensitive to external force and could be injected. Furthermore, the interactions with human umbilical cord mesenchymal stem cells were studied *in vitro* and high biocompatibility was indicated.

Overall, polysaccharide NPs can lead to a considerable improvement in properties when combined with a polymer matrix, *e.g.* mechanical properties but also cargo encapsulation and release. In addition, polysaccharide NPs act as filler material from renewable resources, which is another advantage. More research on polysaccharide/polymer systems is essential to promote them for broader application areas and overcome the challenges when using them.

4.3 Aqueous two-phase system templated polysaccharide microparticles

ATPS are formed from specific mixtures of two hydrophilic polymers or salts at elevated concentration.^{60,61} A commonly used system is the combination of PEG and DEX. These systems feature a completely water-based environment and very low surface tensions between the two phases.^{7,198} In recent years, ATPS have been utilised frequently for the separation of biomacromolecules as well as nanoparticles^{199–201} or as environment for



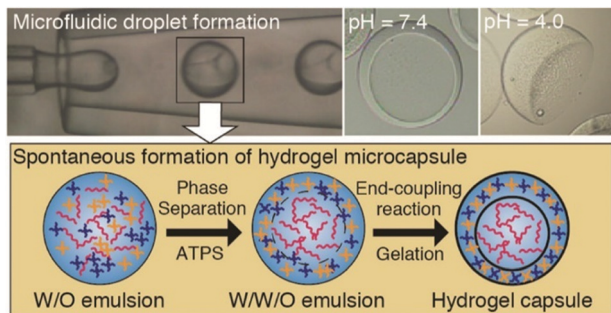


Fig. 8 Formation of PEG/DEX capsules *via* ATPS templating in a microfluidic system. (Reproduced with permission from ref. 204. Copyright American Chemical Society, 2019.)

enzymatic catalysis.²⁰² In addition to macroscopic ATPS, dispersed systems, *i.e.* water-in-water emulsions, have been investigated as well^{135,136} that can be employed as template for the formation of microparticles in aqueous environment.²⁰³ Huck and coworkers synthesised microparticles *via* microfluidics with droplets containing the ATPS from DEX and poly(ethylene glycol) diacrylate (PEGDA) that were formed in oil.²⁰³ The diacrylate phase was photopolymerised to obtain stable particles. In such a way, DEX core/PEGDA shell particles were produced and the particle shape could be tailored *via* the DEX/PEGDA ratio, *e.g.* core/shell or particles with socket. Another crosslinking technique was explored by Ono and coworkers, namely amide formation *via* active esters and amines (Fig. 8).²⁰⁴ In a microfluidic avenue, droplets of PEG/DEX were formed in oil. After phase separation, the PEG phase settled as the shell of the aqueous droplets. Notably, the PEG solution was fed into the system *via* two streams as two PEG species were employed, *i.e.* tetra-arm PEG with either NH₂ or *N*-hydroxy succinimide ester functionality that crosslinked immediately after phase separation. In such a way, crosslinking was performed directly without light or other stimuli, revealing PEG/DEX capsules.

The fabrication of Janus particles was described by Shum and coworkers.²⁰⁵ Here, also microfluidics were utilised to form droplets containing PEG and DEX in an oil. In order to form half-spherical particles, the partitioning behaviour of agarose in the ATPS was exploited that preferably enriches in the PEG phase. As agarose solidifies at low temperatures, the PEG phase was stabilised against dissolution in water, while the DEX phase dissolved revealing PEG half spheres at temperatures below 17 °C. PNIPAM/alginate based microparticles were presented by Arkedani and coworkers, who used Ba²⁺ for ionic cross-linking of alginate and microfluidics for droplet formation and thermoresponsive PNIPAM for templating.²⁰⁶ A completely water-based approach was introduced by van Esch and coworkers.²⁰⁷ Three streams were employed in microfluidics, *i.e.* a PEG-containing stream for the core, a DEX-containing stream for the shell and a PEG-containing stream as continuous phase. PEG-in-DEX droplets were formed in a PEG-solution and the DEX phase crosslinked *via* photo-induced thiol-yne reaction. As such, permeable capsules were synthesised in a completely aqueous system.

ATPS provide a useful avenue to template the formation of polysaccharide based microparticles in a completely water-based environment. As such, organic solvents can be excluded, which is highly interesting for bio-related applications.

5. Applications of polysaccharide nanoparticles

In various areas of research and application, nano-sized delivery systems based on polysaccharide NPs have been widely used in transporting biomedical “cargoes”, for example, drugs, genes, polypeptides, proteins, vaccines or nucleic acids. Moreover, polysaccharide NPs can be used in other areas of applications such as Pickering emulsion stabilisers/emulsifiers and as reinforcing agents in various materials due to their environmental-friendly properties. For instance, when polysaccharide NPs are used to enhance other biodegradable and renewable polymers, efficient and eco-friendly materials with adequate thermo-mechanical properties can be obtained, which is crucial for the development of materials without any environmental issues and overcoming the disadvantages of the original material, *e.g.* fragility.^{19,56}

For example, Iijima and coworkers utilised polysaccharide NPs as precursors for polysaccharide films that were studied with respect to interactions with biological cells.²⁰⁸ CS was combined with anionic polysaccharides to form NPs and subsequently the dispersions were cast into molds and dried at elevated temperature to obtain particle-based films. Depending on the anionic polysaccharide type cell adhesion could be tailored.

Practical applications of polysaccharide NPs in the fields of nanomedicine, cosmetics, and food systems are summarised in the following sections and discussed according to their suitability and required properties for the respective application.

5.1 Polysaccharide nanoparticles in nanomedicine

In biomedicine and biomaterials engineering, polysaccharide NPs have significant advantages and potential applications due to their biocompatibility and biodegradability.^{19,209} Especially in the field of nanomedicine, polysaccharide NPs can be exploited due to their favourable properties in biological systems, *e.g.* due to their mucoadhesive properties and cell uptake.^{210,211} As such, applications like drug-delivery and tissue engineering are discussed frequently but also other therapeutic developments have been presented in recent years. For example, Park and coworkers described the utilisation of CS-based nanoparticles for photodynamic therapy.²¹² There, the photosensitiser Pheophorbide A was conjugated to CS *via* a reactive oxygen species sensitive thioketal linker. The amphiphilic CS-conjugate self-assembled into NPs in aqueous environment leading to a self-quenching of the photosensitiser. The presence of reactive oxygen species in tumour environment resulted in thioketal bond cleavage and release of the photosensitiser. The photosensitiser was activated by near infrared irradiation in turn with a toxic effect. Another approach to photodynamic therapy was described by Wich and coworkers.²¹³ A fully polysaccharide-based



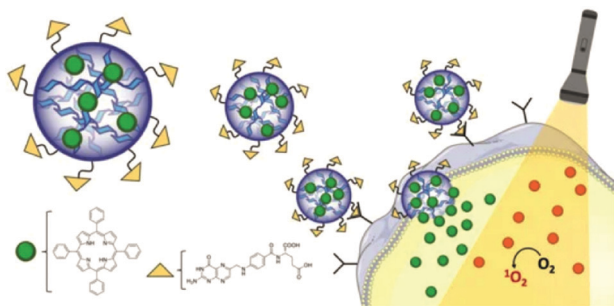


Fig. 9 Mechanism of folic acid targeted photodynamic therapy: DEX NPs loaded with tetraphenylporphyrin are taken up via endocytosis and release the photosensitiser for phototoxic effect.²¹⁴ (Licensed under CC-BY 2019.)

block copolymer consisting of DEX and acetalated DEX blocks was synthesised *via* a copper free click reaction based on thiol exchange leading to disulfide connected blocks. Next, micelles were formed and phthalocyanine zinc as photosensitiser encapsulated, where the block copolymer acted as surfactant for the zinc complex. Delivery of the payload was investigated *in vitro*, showing the block copolymer stabilised the photosensitiser with increased bioavailability. Upon irradiation with NIR light, a strong concentration dependent toxic effect was observed. It should be mentioned that the disulfide block connection introduced redox responsive degradation of the micellar NPs. In a subsequent contribution, DEX NPs were conjugated with folic acid to improve specific uptake of the photosensitiser into cancer cells (Fig. 9).²¹⁴

Alonso and coworkers studied CS/DS or CS/HA coacervate NPs loaded with an HIV peptide antigen candidate, *i.e.* HIV protease cleavage site 5 peptide (PCS5).²¹⁵ The authors investigated various factors in their NPs and the effect on immune response. For example, the attachment of the antigen *via* ionic interactions, cleavable or non-cleavable conjugation was investigated as well as the presence of immunomodulatory molecules such as poly(inosinic acid):poly(cytidylic acid) (poly(I:C)). All NP formulations showed a generation of antibody responses. Overall, NPs with PCS5 conjugation and poly(I:C) incorporation featured the strongest activation of antigen-presenting cells. Sacco and coworkers investigated the effect of CS acetylation on the immune cell response.¹²⁵ CS was combined with short HA in a coacervate formation for the synthesis of NPs. The particles were studied regarding stability at different pH and osmolarity, where only one type of acetylated CS showed stability over a broad range of conditions. In the subsequent immune response studies, the stable formulation induced minor immunogenic response while less stable compositions induced tumour necrosis factor- α production by macrophages. The study revealed that the hydrophilic/hydrophobic balance of polysaccharides is major factor in interactions with biological systems.

Two most relevant applications of polysaccharide nanoparticles in biomedicine, *i.e.* drug/gene delivery and tissue engineering, are discussed in detail below.

5.1.1 Polysaccharide NPs for drug/gene delivery. In recent years, the mechanism of the circulation of polymeric drug/gene-loaded NPs in human bodies has been broadly investigated. After the injection into the body, drug/gene-loaded NPs cross the

epithelial barriers and circulate in the blood vessels before reaching the target site. The drug accumulation in targeting sites (*e.g.* tumours) usually results from the enhanced permeation and retention (EPR) effect, which is based on eased penetration of NPs into pathological tissues due to more accessible fenestrations in the tissue.³² The EPR effect is a unique phenomenon of pathological tissues such as solid tumours related to their anatomical and pathophysiological differences from normal tissues. Moreover, potential severe adverse effects to healthy tissue caused by systemically-delivered active drugs can be prevented by using drug-loaded NPs *via* the EPR effect.^{58,216}

Furthermore, polysaccharide NPs were utilised to improve the solubility of poorly water-soluble drugs, transport the active drug molecules to a targeted pathological site, and release drugs controllably and sustainably for drug/gene delivery.¹⁹ Also, various useful theranostic systems based on polysaccharide NPs were developed, as summarised by Lee and coworkers, *e.g.* fluorescence imaging and drug delivery, magnetic resonance imaging and drug delivery or near-infrared fluorescence imaging and drug delivery.⁵⁸

The polysaccharides discussed above (Section 2) can effectively be used to develop suitable delivery systems. Aminabhavi and coworkers described diverse pharmaceutical applications of CS particulate systems, including cancer therapy, gene delivery, mucosal delivery, topical delivery, ocular delivery and colon targeted drug delivery.⁸⁷ Yameen's group developed novel enzyme-responsive DEX based oligoester crosslinked NPs for the controlled release of an anticancer drug, 5-fluorouracil (5-FU).²¹⁷ The results obtained from their study showed that 5-FU was effectively encapsulated in the DEXNPs (Fig. 10), and the synthesised DEXNPs maintained their stability at pH conditions representative of specific *in vivo* environments (*e.g.* stomach, small intestine, and colon). Moreover, as reported by the studies of release kinetics and colon cancer cell viability, it

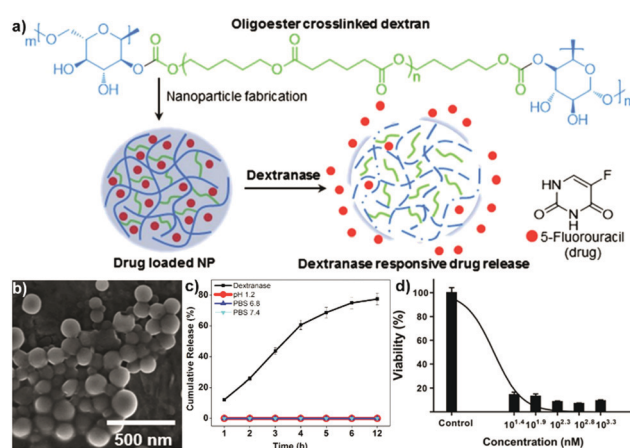


Fig. 10 (a) Schematic overview for nanoprecipitation assisted fabrication of DEXNPs in the presence of 5-FU, as well as depiction of dextranase responsive drug release, (b) SEM image of 5-FU loaded DEXNPs, (c) release kinetics of DEXNPs at simulated pH 1.2, PBS 6.8, PBS 7.4 & dextranase (15 U mL⁻¹) and (d) cytotoxicity of DEXNPs against HCT 116 colon cancer cell line after 1 h of exposure of DEXNPs to dextranase. (Reproduced with permission from ref. 217. Copyright Elsevier, 2020.)



was found that 75% (± 4) of the encapsulated 5-FU was released in 12 h in the presence of dextranase and the used HCT116 cell displayed a substantial cell viability decrease. These results demonstrated the potential applications of the studied DEXNPs as a specific 5-FU delivery system in response to the dextranase enzyme. In addition to the DOX-loaded HANPs discussed in Sections 3 and 4, other polysaccharides and their derivatives were also investigated by different research groups for their utilisation in DOX delivery. Based on their studies, CSNPs,^{48,87,147} CLNPs,^{218,219} SNPs,^{220,221} and DEXNPs^{63,222,223} proved to be potential nanocarriers for antitumour drugs such as DOX in order to enhance their therapeutic effects.

Some polymer NPs with polysaccharide shell have also attracted attention to be used as adequate nanocarriers. For encapsulating and protecting Astaxanthin, Xu and coworkers utilised CS oligosaccharides to form a cationic hydrophilic shell around the anionic hydrophobic PLGA NPs.¹⁴⁹ The results of their work showed that the core-shell NPs formed had improved water solubility and stability compared to the PLGA NPs without a CS oligosaccharide shell, making the developed NPs suitable for their future applications in transporting hydrophobic nutraceuticals and drugs. A polysaccharide NP based on DS/CS for the delivery of insulin was described by Neufeld and coworkers.²²⁴ The NPs were designed for oral dosage and provide improved intestinal absorption. An improved performance compared to oral insulin solution revealing 2–3-fold improvement of oral relative pharmacological availability was observed. Heinze and coworkers described the attachment of a histone deacetylase inhibitor into CL or DEX-based NPs, *i.e.* valproic acid.²²⁵ The acid was attached *via* esterification and NPs formed *via* nanoprecipitation or emulsion method. The formed particles could be used for valproate release *via* enzymatic cleavage of the ester bond, which was studied *in vitro* and in shell-less Hen's egg tests. Overall, the nanocarriers revealed good biocompatibility. An alginate-based system for the delivery of NO and Ag NPs was described by Seabra and coworkers.²²⁶ Alginate NPs were synthesised *via* gelation with Ca^{2+} and a precipitation. Furthermore, Ag NPs synthesised from AgNO_3

and tea extract and mercaptosuccinic acid (a NO acceptor) were encapsulated into the alginate NPs. The particles could be loaded with NO *via* nitrosation of mercaptosuccinic acid and utilised subsequently for antibacterial purpose. The particles showed antibacterial activity against three bacteria strains indicating a synergistic effect of NO release and Ag NPs, while not being toxic to Vero cells.

Gene delivery is a frequently investigated application of polysaccharide NPs.^{227–229} Especially, CS complexation with siRNA or DNA for gene delivery has been studied frequently.^{230–232} For example, Leong and coworkers synthesised CS-based nanoparticles with DNA loading.²³³ Thus, CS/DNA NPs were formed *via* a coacervation process. The particles were PEGylated and conjugated with transferrin to enhance stability and cell uptake. Minimal cytotoxicity was observed and gene transfection in various cell types *e.g.* HEK293, HeLa or IB-3-1, which could be tailored *via* the surface functionalisation. Fréchet and coworkers described gene delivery based on DEX NPs.^{234,235} DEX was partially oxidised *via* periodate, acetalated and conjugated with spermine by reductive amination. The carrier particles were generated *via* a double emulsion technique and directly loaded with siRNA. The cationic spermine functionalisation acted as a handle to complex with siRNA for high loading efficiency. Gene transfection was studied *via* knockdown of luciferase expression in HeLa cells *in vitro*. As the spermine/siRNA interaction is sensitive to pH, selective release was observed after internalisation in the cell only.

A NP based on PL was described by Letourneur and coworkers (Fig. 11).²³⁶ Cationic modified PL was combined with Fucoidan in a coacervation process to form NPs and microRNA adsorbed at the surface. The system promoted microRNA delivery inside cells and selectively targeted towards cells with P-selectin overexpression at the surface of activated platelets and endothelial cells in atherosclerotic-related diseases. Amiji and coworkers studied HA-based gene delivery.²³⁷ HA was grafted with hexyl chains to introduce hydrophobicity, PEG to enhance particle stability and poly(ethylene imine) (PEI) to facilitate interaction with siRNA. Tumour necrosis factor- α -specific siRNA was encapsulated and the uptake as well as down regulation of tumour

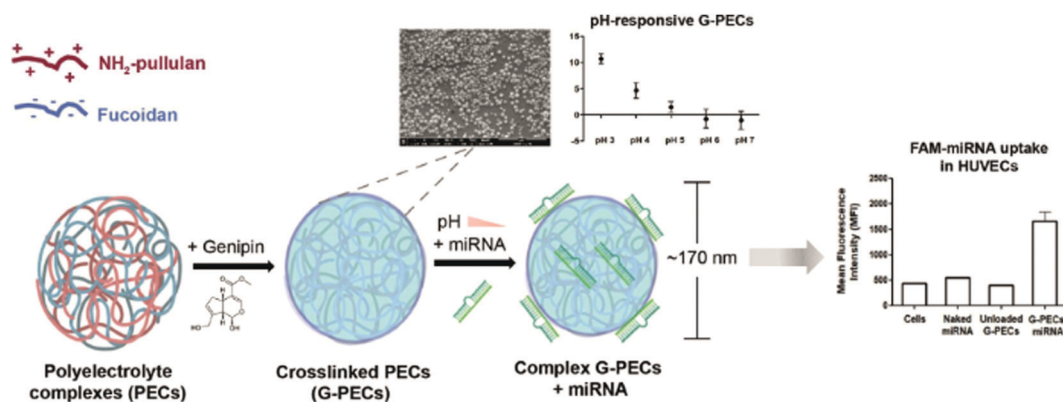


Fig. 11 Formation of NH_2 -PL/fucoidan coacervate NPs and loading with microRNA (miRNA) (left), particle characterisation *via* SEM and pH response (top) and mean fluorescence intensity (MFI) of fluorescein amidite (FAM) measured by flow cytometry analysis revealing the uptake in human umbilical vein endothelial cells (HUVECs) (right). (Reproduced with permission from ref. 236. Copyright Elsevier, 2021.)



necrosis factor- α in macrophages observed. As such, an improved survival rate was observed in the mouse model. Gene carriers based on alginate/ Ca^{2+} complexation have been described as well. Peng and coworkers formed alginate NPs *via* W/O microemulsion and crosslinking *via* Ca^{2+} .⁸⁰ In order to test gene delivery, GFP-encoding plasmids were encapsulated and the degree of endocytosis by NIH 3T3 cells was investigated. Ca-alginate NPs with an average size around 80 nm in diameter were very efficient gene carriers compared to plasmid DNA (pDNA) condensed by PEI.

NPs from CRG for drug-delivery purposes have been described as well, where frequently a complexation with CS was used for NP stabilisation. Grenha *et al.* described the formation of NPs from κ -CRG and CS *via* a complex coacervation process.²³⁸ Particles in a size range from 400 to 700 nm were described with adjustable properties according to the ration of the two polysaccharides. In the next step, loading with OVA as well as release was probed. A sustained and controlled release of OVA over three weeks was observed. A similar structure was described by Chung and coworkers, who used ι -CRG and CS-based NPs for the loading and delivery of insulin *via* oral administration.²³⁹ Abdelghany *et al.* produced κ -CRG and CS-based NPs for tuberculosis treatment *via* ethionamide encapsulation.²⁴⁰ The CRG content was varied to study the effect on particle size, zeta potential and drug release. In particular, it was found that CRG stabilised the particles and enhanced ethionamide entrapment. A sustained release was observed and considerable antimycobacterial activity. A CRG-based capsule was described by Tánori and coworkers, who utilised an emulsion process to form capsules from κ -CRG and CS.²⁴¹ At first olive oil nano emulsion droplets were formed and κ -CRG/CS deposited at the interface *via* layer-by-layer assembly. The oil droplets were loaded with the lipophilic anti-inflammatory drug Diflunisal and the release studied. A zero-order release profile was observed for three- and four-layer systems, while first order release kinetics were observed for one and two layers as well as a Fickian diffusion for emulsion droplets without polysaccharide coating.

To enhance the cellular delivery of genetic materials, NPs with polysaccharide coating were described as well, for example HA-PEI, HA-PEG, and HA-PEI/HA-PEG were synthesised by Amiji and coworkers in order to encapsulate pDNA.¹⁵⁵ The negatively charged HA formed the hydrophilic shell of the NPs, and the combination of HA, PEI and PEG has a considerable effect on gene expression results because of the ability of HA-PEI/HA-PEG NPs to protect pDNA from the environment. In addition, the appropriate pDNA accessibility of HA-PEI/HA-PEG NPs could result in controlled release of pDNA and improved transfection in cells, which might be useful for the delivery of genetic material for cancer therapy. Liu *et al.* designed a novel core-shell nanoparticle system based on PL and poly(β -amino)ester (PBAE) for the codelivery of pDNA expressing green fluorescent protein (pEGFP) and the model chemotherapy agent methotrexate (MTX).¹⁵⁶ The MTX-PL/PBAE/pEGFP NPs prepared had a PL shell and a PBAE/pEGFP core. Also, they were found to exhibit a strong hepatoma-targeting property at both cellular and animal levels, indicating its potential for combination of chemotherapy and gene therapy. Although coating drug/gene-loaded

hydrophobic polymer NPs with a hydrophilic polysaccharide shell can improve their stability and water solubility for improved delivery, information of the safety and efficacy of the polysaccharide/polymer systems still requires more research.¹⁴⁹

Drug/gene delivery systems based on polysaccharide NPs have great potential in the treatment of cancer and other diseases such as diabetes. However, further study is crucial to overcome several challenges in their use. For example, the drawbacks resulting from the reliance on the EPR effect for passive targeting during the drug/gene delivery: heterogeneity of the EPR effect caused by factors such as variations in NP size can result in uncertainty in delivery efficiency. Moreover, burst release and compromised mechanical properties caused by incomplete degradation of polysaccharide NPs is an issue that should not be ignored.²⁴² Thus, ongoing development of targeted delivery systems is required to improve drug/gene delivery efficiency.

5.1.2 Polysaccharide NPs for tissue engineering. Tissue engineering aims at developing biological substitutes that restore, maintain, or improve tissue function by using the principles of engineering in life sciences. In tissue engineering, cells and cell carriers are two primary components, which play significant roles in developing the biological substitutes. The cell carriers are usually referred to as scaffolds, and their functions involve delivering cells to a desirable area and inducing local tissue regeneration, as barriers to protect transplanted cells or tissues from host immune attack, or as reactors to stimulate host cell recruitment, homing, and differentiation.^{37,243} Tissue engineering covers the generation of scaffolds, hydrogels, and particulate systems from biocompatible polymers to transport growth factors, which are soluble secreted proteins important for tissue regeneration.^{49,244} The efficiency of tissue engineering is improved by controlled release of therapeutic factors to specific sites, which is thought as a special case of drug delivery.³² During the delivery and release process of protein-based growth factors, polysaccharide-based micro-/nanoparticles are a suitable carrier material because they can protect the growth factors, give a versatile release profile, and reduce the possibility of the side effects.²⁴²

Mandal and coworkers prepared dual growth factor loaded CSNPs by using epidermal growth factor (EGF), fibroblast growth factor (FGF) and CS.²⁴⁵ The increased proliferation of fibroblasts *in vitro* demonstrated the successful delivery of growth factors from CSNP. Moreover, non-toxicity of growth factors loaded CSNPs and controlled release were observed, and the attachment and growth of fibroblast in EGF/FGF-CSNP system were significantly improved. These results indicated the promising utilisation of this dual growth factors loaded nanoparticle system in tissue engineering area.

In the work of Alonso and coworkers, CS/HA based NPs were synthesised by using TPP ionic gelation for transporting vascular endothelial growth factor (VEGF) and platelet derived growth factor (PDGF-BB).⁵⁷ The results demonstrated that the CS/HA based nanocarriers were stable in biological media without any cytotoxicity. According to their *in vitro* release studies, PDGF-BB could be released from CS/HA NPs sustainably in one week, while the release of VEGF was uncontrollable. This report already



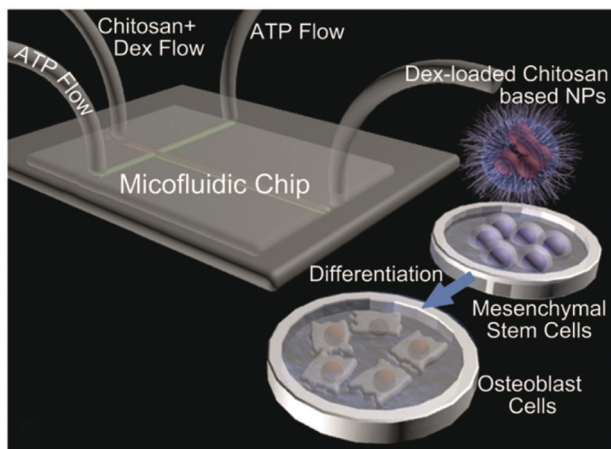


Fig. 12 Schematic representation of a cross-junction microfluidic device used to hydrodynamically focus the chitosan–dexamethasone flow using a sheath flow of adenosine triphosphate-containing water for the preparation of CS-based NPs and their use in cell growth. (Reproduced with permission from ref. 248. Copyright Elsevier, 2015.)

showed the future of injectable PDGF-BB loaded CS/HA nanoparticle system in the fields of tissue engineering and treating ischemia-related diseases but further research is needed in the control of growth factor release. Wu and coworkers described a novel DEX- and gelatin-based nanoparticle system (DGNPs) for encapsulating and transporting the growth factor bone morphogenetic protein (BMP).²⁴⁶ The mono-dispersed DGNPs formed had a mean diameter of about 54 nm and good swelling and degradation properties in the target media. According to their *in vitro* release studies, more than 90% of the loaded BMP was maintained for more than 12 days and released in a sustained manner. These positive results proved that the formed DGNPs have the potential to be biodegradable and biocompatible carriers for protein-based growth factors. Goncalves and coworkers presented NPs based on CS and poly(γ -glutamic acid) with diclofenac as non-steroidal anti-inflammatory drug for the treatment of degenerated intervertebral disks.²⁴⁷

Tayebi and coworkers described a microfluidic avenue for the preparation of CS/dexamethasone NPs that were utilised as adjuvants in cell growth (Fig. 12).²⁴⁸ The effect of the adenosine triphosphate-crosslinked particles on the osteogenic differentiation of mesenchymal stem cells was studied. Due to the tailored size of the NPs improved differentiation was observed *via* enhanced dexamethasone release.

It should be noted that polysaccharide NPs with hydrophilic properties can be utilised as adequate growth factor nanocarriers, but several issues (*e.g.* safety, cost, efficacy) need to be addressed for development of successful growth factor therapies. The pre-clinical results from *in vitro* cellular study and *in vivo* animal model may not perfectly simulate the physiology and pathophysiology in humans, which has to be studied thoroughly to enable future applications.^{242,244} Thus, further studies are required to enhance the control over polysaccharide NP-governed tissue growth processes.

5.2 Polysaccharide nanoparticles in cosmetics

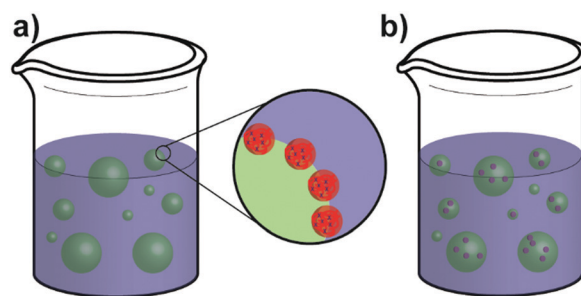
Cosmetics are consumer products to be placed in contact with the external parts of the human body, with teeth or the mucous membranes of the oral cavity. The primary utilisations of cosmetics include cleaning, change of appearance, and body odour correction.²⁴⁹ The use of polysaccharide-based NPs is a specific branch area of applying nanotechnology in cosmetics. Many researchers have investigated the applications of polysaccharide-based NPs in improving the properties of cosmetic products, for instance, using them as cosmetic ingredients to stabilise Pickering emulsions or deliver useful substances.¹⁰

5.2.1 Polysaccharide NPs for Pickering emulsion stabilisation.

Emulsions are widely used in various fields such as pharmaceuticals, drug delivery or food industry.²⁵⁰ To form various cosmetic and personal care products such as antiperspirants, deodorants, and anti-aging products, emulsions are also frequently used.²⁵¹ As introduced in Section 3, an emulsion is a dispersion of two immiscible liquids. In the absence of an adequate interfacial stabiliser, the formed emulsions are unstable and tend to undergo phase separation, causing creaming in the emulsion. To solve this issue, surfactants and solid NPs are used because they can reduce the interfacial tension between the two phases and improve emulsion stability.^{252,253}

Emulsions stabilised with solid NPs are also referred to as Pickering emulsions (Scheme 8). Though surfactants can enhance both the kinetic and thermodynamic stability of the emulsions, their toxicity, high cost, and impractical recovery lead to issues.^{251,252} However, solid NPs such as polysaccharide NPs used in Pickering emulsions can be environment-friendly and biologically compatible. Besides, Pickering emulsions can impart improved stability against coalescence. These advantages cause that Pickering emulsions can replace the surfactant stabilised emulsion to produce long-term stabilised emulsions to be applied in cosmetics.^{251,252,254} In the review of Yuan and coworkers, various types of solid particles utilised in forming Pickering emulsions were listed and discussed in detail.²⁵⁰ For example, nanoscale particles derived from natural polysaccharides such as CH nanocrystals,¹⁹ CL nanocrystals,^{255,256} and starch nanocrystals^{256–258} were found to be potential Pickering emulsion stabilisers due to their superior performance in various areas.

Biodegradable and antibacterial CS is widely used in cosmetics,²⁵⁹ and CSNPs have been investigated by many researchers



Scheme 8 (a) Structure of a polysaccharide NP stabilised Pickering emulsion and (b) Pickering emulsion with cargo loaded droplets.



for their utilisation in stabilising Pickering emulsions. Tong's group described a simple, straightforward reversible, and long-term stable (> 2 months) Pickering emulsion system based on CS.²⁶⁰ At high pH conditions (pH > 6), CS is water-insoluble and the CSNPs were formed *in situ* by aggregation. These CS aggregates adsorbed at the interface of oil and water to stabilise the O/W emulsions, forming an excellent stabilised Pickering emulsion system. While at low pH conditions (pH < 6), CS is water-soluble and the demulsification phenomenon occurred. The results obtained from this study demonstrated that CS is a practical pH-sensitive emulsifier, and the Pickering emulsions formed by CS could be useful to fabricate a broad range of reversible emulsion systems.

The abundant, low-cost, and biodegradable starch also has potential applications in stabilising cosmetic Pickering emulsions.^{261,262} However, the broad size distribution of starch particles negatively affects its performance as an emulsifier.²⁵⁰ Therefore, various modifications (*e.g.* hydrophobic modification) of starch have been investigated to increase the possibility of SNPs to adsorb at the oil–water interface during emulsification.²⁶¹ Wang and coworkers described a novel particulate emulsifier based on starch–acetic anhydride–phthalic anhydride nanospheres for undecanol- and glyceryl trioctanoate-water emulsions.^{263,264} According to the results from confocal laser scanning microscopy, the nanospheres were adsorbed at the liquid interface, forming a dense film around the dispersed droplets. Besides, it was found that salt addition could cause emulsion coalescence, and the O/W emulsions could be inverted to W/O emulsions by varying the water/oil volume ratio and pH of aqueous dispersion, respectively. These results demonstrated the promising future of using biocompatible SNPs as practical Pickering emulsions stabilisers.

As the properties of starch depend on its origin, the possibilities of native starches extracted from different sources such as taro, wheat, waxy maize, rice, and potato as Pickering emulsions stabilisers were also studied by other groups.^{265,266} For instance, Sun's group prepared four different types of SNPs to stabilise the soybean O/W emulsion system.²⁶¹ Their results showed that the SNPs with near-neutral wettability were synthesised from sweet potato, corn, and tapioca. The near-neutral wettability promoted the adsorption of SNPs at the oil–water interface and the Pickering emulsion stabilisation. In addition, it was also found that the SNPs size could affect the stability of the emulsion, the corn-derived SNPs were less stable than the other two because of the smaller particle size and the caused deviated contact angle from 90°.

A combination of NPs derived from CS and starch for the formation of Pickering emulsions was described by Matos and coworkers.²⁶⁷ Due to their considerable difference in size (nm *vs.* micron), the order of stabiliser addition had a profound effect on the obtained droplet sizes of oil in water. When CS particles are added first, both particle types perform stabilisation, while in the case of simultaneous addition, stabilisation is predominantly due to the larger starch NP addition that displace CS particles. Wang and coworkers employed starch NPs as Pickering stabiliser for the fabrication of microgels.²⁶⁸ At first starch was hydrophobised *via* butyl glycidyl ether and

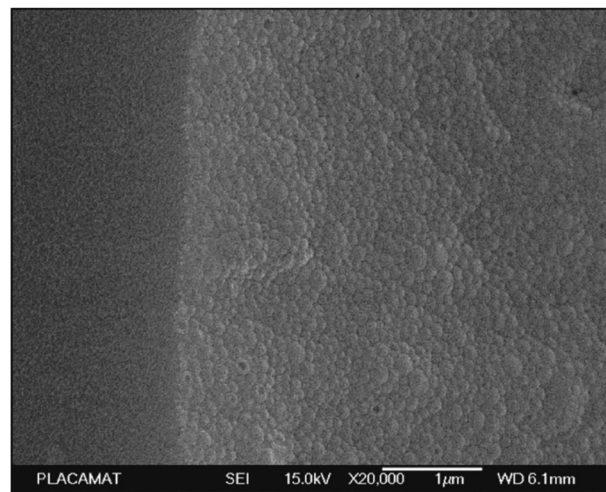


Fig. 13 Cryo-SEM image of the surface of a dodecane droplet in water stabilised by acetalated DEX NPs. (Reproduced with permission from ref. 269. Copyright American Chemical Society, 2020.)

palmitate, then NPs were formed *via* nanoprecipitation. In the next step, an inverse Pickering emulsion was formed with *n*-hexane as the continuous phase and an aqueous solution of acrylamide, acrylic acid and initiator as dispersed phase. After homogenisation the polymerisation was started at 30 °C and well-defined microgels decorated with starch NPs were obtained. Héroquez and coworkers fabricated Pickering emulsions of medium chain triglyceride or dodecane in water *via* acetalated DEX NPs (Fig. 13).²⁶⁹ The acetalated DEX NPs were formed *via* nanoprecipitation and had a diameter around 200 nm. As acetals can be hydrolysed under acidic conditions the particles featured pH responsivity, leading to destabilisation of the emulsion after hydrolysis.

In summary, applying polysaccharide NPs in Pickering emulsions can reduce the tendency of emulsion coalescence and introduce high stability, biocompatibility, and non-toxicity, which will provide support to developing stabilised Pickering emulsions used in different areas such as cosmetics.

5.2.2 Polysaccharide NPs for delivery of cosmetic ingredients.

Polysaccharide NPs such as CSNPs, HANPs, and SNPs can also be introduced as nanocarriers in cosmetical care products (for hair or body care) due to their biodegradability. In many cases, bioactive substances in cosmetic formulations such as vitamins are not stable, *e.g.* they are sensitive to temperature, pH, light and oxidation. Therefore, encapsulation and protection are required to avoid unwanted degradation and to ensure target-specific release.²⁷⁰

Kim *et al.* successfully encapsulated a cosmetical vitamin A alcohol, *e.g.* retinol, into CSNPs for cosmetic and pharmaceutical applications.^{271,272} As such, the solubility issues of retinol in water was solved by incorporating it into CSNPs, and the study showed that the retinol water solubility was increased more than 1600-fold. Inspired by this, Harris *et al.* prepared CS hydrochloride (HCS)–TPP based NPs for the encapsulation of yerba mate (*Ilex paraguariensis* (ILE)) extract.²⁷³ The results demonstrated that the CSNPs involved encapsulation method is a promising technique



Their work was on the basis of the preparation and characterisation of polysaccharide based NPs which had a PEG/methylcellulose (MC) core including anthocyanidin and sodium acetate, and a CS/gallotannin-based shell. The PEG/MC core and anthocyanidin/sodium acetate acted as the smart part of the NPs. As such, a system with T_g corresponding in the range of the unacceptable temperature for product storage was designed *via* the combination of PEG and MC combining a very high and a very low T_g . Anthocyanidin and sodium acetate were mixed to indicate colour change, the neutral colourless anthocyanidin under action of sodium acetate turned purple with increasing temperature of PEG/MC mixture. The active part of the particles was the CS/gallotannin-based shell, which takes the advantage of antibacterial CS and photoprotective and antioxidative gallotannin. The analysis results showed that the prepared SANPs turned from colourless to purple at about 20 °C and 0 °C respectively. Moreover, the SANPs displayed effective antibacterial activity, UV light absorption, antioxidative activity, and non-toxicity, demonstrating the promising application of the prepared polysaccharide-based SANPs in the food packaging area.

The influence of polysaccharide NPs such as CSNPs on the improvement of the physical/chemical properties of food packaging materials such as mechanical strength and thermal stability were investigated by various groups.^{281,282} For example, the addition of polysaccharide NPs enables reduction of the water vapour permeability (WVP) of food packaging materials, which has attracted frequent attention as it is a vital issue for the development of sustainable packaging materials. NPs are considered to be suitable for reducing WVP because they distort the diffusion pathways for the water vapour through the packaging film.^{283,284} In the area of mechanical improvements, Mattoso and coworkers described the fabrication of nanocomposite films of CS NPs in pectin matrix (Fig. 15).²⁸² CS NPs were combined with high methyl (HDM) and low methyl (LDM) pectin matrices. Next, films were casted that showed homogeneous, compact, and continuous properties. The addition of CSNPs improved the mechanical properties considerably, *e.g.* tensile strength increased from 30.81 to 46.95 MPa and from 26.07 to 58.51 MPa for HDM pectin/CSNP and LDM pectin/CSNP, respectively.

Sun and coworkers investigated the effects of taro-derived SNPs content on the properties of corn starch-based packaging films.²⁸⁵ Their work showed that the addition of taro SNPs to the corn starch films increased the tensile strength of films from 1.11 MPa to 2.87 MPa. It was also found that strong interactions between taro SNPs and films improved the thermal

stability of starch films. Moreover, the WVP of the films was decreased after addition of taro SNPs, and the degree of the WVP reduction correlated with the concentration of the taro SNPs. These results demonstrated the potential of incorporating taro SNPs into corn starch films for developing biodegradable starch-based films with improved properties. A similar study was conducted by the same group.²⁸⁶ Potato-derived SNPs were incorporated into pea starch-based nanocomposite films. The mechanical properties of the films were found to be enhanced by the addition of potato SNPs, and particularly for the incorporation of 6% potato SNPs, the film tensile strength increased the most (from 8.8 MPa to 15.0 MPa). Compared to control samples, the melting temperature of the nanocomposite films increased with the increased potato SNPs content. For the WVP and water-solubility of the nanocomposite films, the incorporation of 6% potato SNPs led to the most significant reduction of the WVP value but it increased again with higher content values. The phenomenon was assigned to the abundant hydroxyl groups in excessive potato SNPs showing more affinity for water than starch. The lowest water-solubility of the nanocomposite films was also obtained at the potato SNPs content of 6%. The improved reduction of water-solubility was considered to be related to the high tensile strength values and the entanglement between SNPs and starch chains, which decreased the nanocomposites' solubility. The results discussed above proved that adding potato SNPs into the pea starch films could be applied in developing degradable food packaging, while WVP and water-solubility can be minimised *via* the optimisation of material composition.

Polysaccharide NPs play a significant role in developing improved food packaging materials. They can combine the advantages of both intelligent packaging and active packaging. Moreover, issues such as high WVP and limited mechanical properties caused by using polysaccharide films can also be overcome by the addition of polysaccharide NPs.^{282,285,286}

5.3.2 Polysaccharide NPs for the delivery of ingredients in food. Many of the common food constituents containing beneficial biological activities, such as nutraceuticals, vitamins, and nutrients, are highly lipophilic molecules. As such, there are various factors limiting the incorporation and delivery of pure lipophilic bioactive substances, for instance, chemical instability, poor bioavailability, poor water solubility, and the interactions between the ingredients. To overcome these challenges in the applications of food and nutrition, nanotechnological approaches provide a direction for the encapsulation and controlled-release of active food ingredients (Scheme 9).^{277,287–289}

Different examples of suitable nanoscale delivery systems for transporting food constituents were listed and discussed in the review of McClements,²⁸⁷ such as micelles, micro/nanoemulsions, liposomes, microgels, and biopolymer NPs (*e.g.* protein NPs and polysaccharide NPs). For the delivery systems based on polysaccharides, their properties such as thermal stability, digestibility, and antioxidant capacity, depend significantly on the components used to construct a nanoscale delivery system.

Shimoni's group entrapped a hydrophobic nutraceutical, the water-insoluble isoflavone Genistein, into enzymatically synthesised DEXNPs.²⁹¹ The dextransucrase generated spherical DEXNPs were

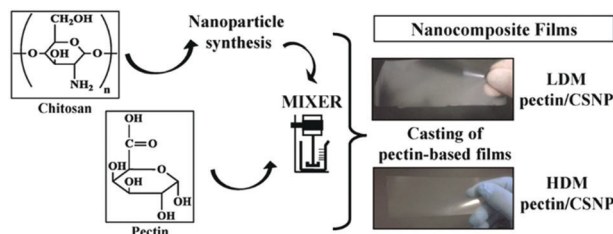
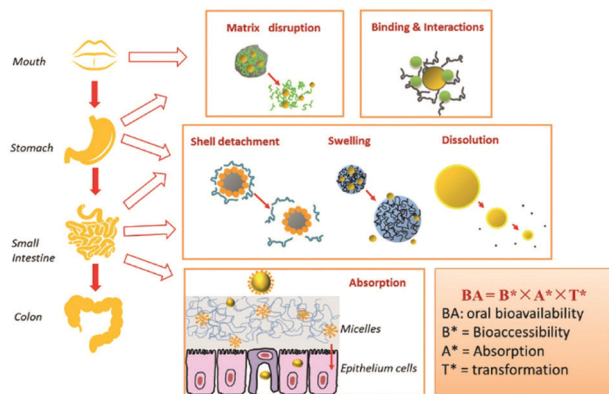


Fig. 15 Overview of nanocomposite formation from pectin and CS NPs and casted films. (Reproduced with permission from ref. 282. Copyright Elsevier, 2016.)





Scheme 9 Overview of the design of food ingredient delivery systems to control bioavailability *via* body-particle interactions. (Reproduced with permission from ref. 290. Copyright John Wiley and Sons, 2017.)

prepared under optimised conditions (pH 5.2–6 and sucrose concentration > 0.5 M). Although the bioavailability of the DEXNPs was increased due to the smaller particle size, larger surface area, and increased solubility caused by the Genistein inclusion, it was found that the loading efficiency of Genistein was very low. The limitation in Genistein loading was considered to result from the fact that the Genistein was only included in dextran particles without forming complexes with the dextran chains. The study provided a good guide to develop nanosized vehicles to incorporate and stabilise the hydrophobic bioactive compounds in foods, which was useful to establish more platforms for delivering water insoluble bioactive chemicals in food and/or pharmaceuticals.

Developing delivery systems based on polysaccharide NPs can also be combined with the food packaging applications mentioned in Section 5.3.1. Nieto-Suaza *et al.* prepared curcumin-loaded SNPs as the reinforcing agent to be incorporated into the aloe vera gel-banana starch composite films.²⁹² The results obtained from their work showed that the addition of curcumin-loaded SNPs could enhance several properties of the composite films. For instance, with the increased concentration of the curcumin-loaded SNPs, the important WVP of the composite films decreased. One reason for that phenomenon has been discussed above, which is the distortion of the water diffusion path caused by the addition of SNPs. Besides, the existence of the hydrophobic curcumin also played an important role in reducing WVP because it can limit the interaction between water and the polymeric matrix. Moreover, it was observed that the antimicrobial and antioxidative curcumin could be released from the composite films to different food simulants, such as simulate hydrophobic, simulate hydrophilic, and simulate lipophilic foods. The curcumin release was favoured in simulated lipophilic foods and could be controlled by modifying the polarity of the SNPs by acetylation. The group's work combined the effective utilisations of polysaccharide NPs in food delivery systems and food packaging, indicating the promising future of developing biodegradable composite films that can incorporate bioactive substances in food industries.

Furthermore, there are other practical polysaccharide NPs related food delivery systems widely used in transporting bioactive substances. For example, abundant and antimicrobial

CSNPs have been widely used as nanocarriers for transporting tea polyphenols, which are nutraceuticals bringing multi-health benefits including cancer prevention, antioxidant activities, antibacterial activities, and immune enhancement. However, some associated challenges can be caused by factors such as the presence of ions, pH, and enzymes in the gastrointestinal tract. The uncontrollable size of CSNPs resulted from fluctuations of these factors may affect the stability of the tea polyphenols loaded CSNPs.^{293,294} Gao and coworkers fabricated zein-propylene glycol alginate-tea saponin (TS) ternary complex nanoparticles for the delivery of resveratrol.²⁹⁵ The presence of TS on the particles significantly improved resveratrol loading as well as the environmental stability of resveratrol. The best release characteristics in the small intestine were obtained for a ratio of zein to TS of 4:3 as observed *via* an *in vitro* digestion model. As such, the novel protein-polysaccharide-surfactant ternary complex enabled efficient delivery of resveratrol.

McClements and coworkers summarised several examples of forming protein-polysaccharide NPs based delivery systems by conjugating polysaccharide (*e.g.* CS, DEX) with another common biopolymer protein (*e.g.* Whey Protein Isolate, OVA).²⁹⁰ The functional properties of proteins can be improved by conjugation to polysaccharides, giving tailored gastrointestinal protection and physicochemical stability. However, regulatory issues may be caused by using conjugates compared to all-natural ingredients, and the production costs may increase due to the additional ingredients and processing operations required to fabricate conjugates.

NPs based on natural polysaccharides can fulfil most of the requirements to be applied in food-grade delivery systems due to their biodegradability and biocompatibility. Despite several advantages like effective encapsulation of bioactive substances and improved stability against enzymes in the gastrointestinal tract, drawbacks such as the aggregation of polysaccharide NPs and burst release caused by small particle size and large surface area need more attention. Therefore, future studies are necessary to overcome these challenges and fabricate polysaccharide nanocarriers with improved properties for delivery systems in food.^{293,294}

6. Conclusion and outlook

Polysaccharide NPs have found considerable attention in the past decade mainly for biomedical applications but also use in cosmetics and food has been implemented. The utilised polysaccharides are obtained from various natural sources making them prime candidates for sustainable nanotechnology. Furthermore, polysaccharides have favourable biological properties and activities depending on their chemical structures. Key features are the remarkable biodegradability, biocompatibility, and low-toxicity of polysaccharides that support their broad applications in biological environment – a key driver for future developments.

The basis for polysaccharide NPs are the preparation methods. A variety of synthesis methods for preparing NPs with polysaccharides and their derivatives were presented in here, highlighting the broad range of options in polysaccharide NP synthesis. The choice of a suitable synthesis method depends on the specific



application and its requirements. For instance, to fabricate excellent polysaccharide NPs based nanocarriers for *in vivo* delivery systems, controllable size, charge, surface morphology, loading capacity, and substances release are important. These requirements can mostly be satisfied by choosing one of the common fabrication methods, *e.g.* nanoprecipitation, complex coacervation, emulsification, or other practical methods. However, there are some limitations of these methods that have to be taken into account, such as the requirement of large amounts of organic solvents for the emulsification method and the potential toxicity issue when using the desolvation method, for example. Although, the fabrication of polysaccharide NPs has been investigated thoroughly in recent years, novel approaches are still sought after. Especially, the replacement of organic solvents with harmless and/or sustainable alternatives could be a considerable direction as well as improved control over particle size and shape with new template methods.

A direction in polysaccharide NPs with considerable growth potential is the combination with polymeric materials. For example, to attach polymers to polysaccharide NPs to form polysaccharide/polymer NP systems, “grafting to” and “grafting from” reactions are applied. In “grafting to” reactions, functionalised and fully characterised polymer is prepared in advance before attachment. While in “grafting from” reactions, monomers polymerise directly on the interface of polysaccharide NPs. After graft modification, polysaccharide–polymer based core–shell NPs are obtained that feature new properties to be tailored *via* the chemistry of the grafted polymer. The inclusion of a polysaccharide NP core into the polymer shell can introduce improved stability against solvents and stimuli response indicating the potential use of polysaccharide–polymer based core–shell NPs in biomedical applications. Furthermore, inclusion of polysaccharide NPs into polymer materials leads to enhancement of the mechanical properties with possible implications for polymer materials production on an industrial-scale. The field of polysaccharide/polymer hybrid and composite materials is still underrepresented in the literature. Nevertheless, various directions can be foreseen, for example tailoring of polymer surface wettability *via* polysaccharide NP introduction or control of drug release from polysaccharide NPs *via* responsive polymer shells.

As effective nano-sized carriers, polysaccharide NPs have broad applications in transporting substances. When utilised in nanomedicine, cosmetics, and food areas, polysaccharide NPs introduce various benefits for the transport of drugs, genes and other hydrophobic and lipophilic substances *via* efficient protective encapsulation, improving the bioavailability and chemical stability of these substances. In nanomedicine, the EPR effect allows the targeted delivery of drug/gene-loaded polysaccharide NPs to pathological tissues, and various *in vitro* release studies demonstrated that the loaded drugs and genes could be released in a controlled and sustained manner. These useful results also play a significant role in tissue engineering applications which are based on growth factors delivery. Moreover, it was proven that polysaccharide NPs could be practical for the delivery and controlled-release of cosmetic and food ingredients, showing the promising future of polysaccharide NPs based nanocarriers. Certainly, in the

future advanced nanocarriers will be developed that allow more targeted delivery as well as tailored release profiles. Here functionalisation of polysaccharide NPs with targeting moieties and polymers could be an avenue.

On the other hand, polysaccharide NPs can be designed as Pickering emulsion stabilisers and material reinforcing agent in cosmetics and food packaging areas due to their environmental-friendly properties. According to former studies, emulsions stabilised by polysaccharide NPs showed their potential to produce long-term stabilised formulations in cosmetics. After adding polysaccharide NPs to the packaging films, the mechanical properties were enhanced accompanied by lower WVP values, indicating the potential of incorporating polysaccharide NPs into other materials for developing food packaging materials with improved properties. Also, in the stabilisation of emulsions and introduction in food packaging further tailored functionalisation could have a positive effect on overall materials properties, *e.g.* when interaction of polysaccharide NPs and matrix materials are improved.

Overall, it can be concluded that polysaccharide NPs have proven to be very important future biological nanocarriers, emulsifiers, and material reinforcing additives. However, more research is still required to work out issues when fabricating and using them. For instance, it is important to promote the preclinical results from *in vitro* cellular studies and *in vivo* animal studies to the human system. Also, aggregation of polysaccharide NPs and burst release of substances are worth mentioning. Therefore, further deep studies are necessary to promote the applications of polysaccharide NPs and ensure their safety and efficacy in the future.

Abbreviations

AFM	Atomic force microscopy
ATPS	Aqueous two-phase system
BMP	Bone morphogenetic protein
CA	Cellulose acetate
CH	Chitin
CL	Cellulose
CLNPs	Cellulose-based nanoparticles
CRG	Carrageenan
CS	Chitosan
CSNPs	Chitosan-based nanoparticles
DEX	Dextran
DEXNPs	Dextran-based nanoparticles
DDT	Dodecanethiol
DGNPs	DEX- and gelatin-based nanoparticle system
DOX	Doxorubicin
DS	Dextran sulfate
EPR	Enhanced permeation and retention
FAM	Fluorescein amidite
HA	Hyaluronic acid
HANPs	Hyaluronic acid-based nanoparticles
HDM	High methyl pectin matrix
HRA NPs	NPs formed by coupling HA with aminated retinoid acid (ATRA)



HUVEC	Human umbilical vein endothelial cell
LDM	Low methyl pectin matrix
MC	Methylcellulose
MFI	Mean fluorescence intensity
MTX	Methotrexate
MMP9	Metalloproteinase 9
MW	Molecular weight
MXS	Minoxidil sulphate
NP	Nanoparticle
OCMCS	Oleoyl-carboxymethyl-chitosan
OVA	Ovalbumin
O/W	Oil-in-water
PBAE	Poly(β -amino)ester
PCS-5	HIV protease cleavage site 5 peptide
PDGF-BB	Platelet derived growth factor
pDNA	Plasmid DNA
PEA	Poly(ethyl acrylate)
PEI	Poly(ethylene imine)
PEG	Poly(ethylene glycol)
PEGDA	Poly(ethylene glycol)diacrylate
PEGFP	Plasmid DNA expressing green fluorescent protein
pep	MMP9 cleavable connection
PFP	Perfluoropentane
Poly(I:C)	Poly(inosinic acid):poly(cytidylic acid)
PLA	Poly(lactide)
PLGA	Poly(D,L-lactide-co-glycolide)
PMA	Poly(methyl acrylate)
PMMA	Poly(methyl methacrylate)
PMMA- <i>b</i> -P(PEGMA)	Poly(methyl methacrylate)- <i>b</i> -poly(poly(ethylene glycol) methyl ether methacrylate)
P(MMA-co-S)	Poly(methyl methacrylate-co-styrene)
PNIPAM	Poly(<i>N</i> -isopropylacrylamide)
PPC	Poly(propylene carbonate)
P(PEGMA)	Poly(poly(ethylene glycol) methyl ether methacrylate)
PTX	Paclitaxel
SANPs	Smart and active nanoparticles
SEM	Scanning electron microscopy
SG1	<i>N</i> -tert-Butyl- <i>N</i> -[1-diethylphosphono-(2,2-dimethylpropyl)nitroxide]
siRNA	Small interfering RNA
SI-SET-LRP	Surface-initiated single-electron transfer living radical polymerisation
SNPs	Starch-based nanoparticles
TEM	Transmission electron microscopy
T_g	Glass transition temperature
THF	Tetrahydrofuran
TPP	Triphosphosphate
TS	Tea saponin
VBC	4-Vinylbenzyl chloride
VEGF	Vascular endothelial growth factor
W/O	Water-in-oil
W/W	Water-in-water
WVP	Water vapour permeability
5-FU	5-Fluorouracil

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

B. S. and A. P. acknowledge funding from the University of Glasgow and the German Research Foundation (grant no. SCHM 3282/3-1).

References

- P.-F. Cui, W.-R. Zhuang, X. Hu, L. Xing, R.-Y. Yu, J.-B. Qiao, Y.-J. He, F. Li, D. Ling and H.-L. Jiang, *Chem. Commun.*, 2018, **54**, 8218–8221.
- J. Zhu and R. E. Marchant, *Expert Rev. Med. Devices*, 2011, **8**, 607–626.
- Z. Ge, D. Xie, D. Chen, X. Jiang, Y. Zhang, H. Liu and S. Liu, *Macromolecules*, 2007, **40**, 3538–3546.
- N. Kocherginsky, C. L. Tan and W. F. Lu, *J. Membr. Sci.*, 2003, **220**, 117–128.
- S. Matoori and J.-C. Leroux, *Mater. Horiz.*, 2020, **7**, 1297–1309.
- M. Antonietti and S. Förster, *Adv. Mater.*, 2003, **15**, 1323–1333.
- B. V. K. J. Schmidt, *Macromol. Chem. Phys.*, 2018, **219**, 1700494.
- R. K. Ibrahim, M. Hayyan, M. A. AlSaadi, A. Hayyan and S. Ibrahim, *Environ. Sci. Pollut. Res.*, 2016, **23**, 13754–13788.
- R. Seyedebrahimi, S. Razavi and J. Varshosaz, *J. Cluster Sci.*, 2020, **31**, 99–108.
- A. Mihrianyan, N. Ferraz and M. Strømme, *Prog. Mater. Sci.*, 2012, **57**, 875–910.
- I. P. Kaur, M. Kapila and R. Agrawal, *Ageing Res. Rev.*, 2007, **6**, 271–288.
- X. He, H. Deng and H.-M. Hwang, *J. Food Drug Anal.*, 2019, **27**, 1–21.
- N. Kamaly, B. Yameen, J. Wu and O. C. Farokhzad, *Chem. Rev.*, 2016, **116**, 2602–2663.
- V. Wagner, A. Dullaart, A.-K. Bock and A. Zweck, *Nat. Biotechnol.*, 2006, **24**, 1211–1217.
- G. Chen, I. Roy, C. Yang and P. N. Prasad, *Chem. Rev.*, 2016, **116**, 2826–2885.
- L. Zhang, F. Gu, J. Chan, A. Wang, R. Langer and O. Farokhzad, *Clin. Pharmacol. Ther.*, 2008, **83**, 761–769.
- B. Yameen, W. I. Choi, C. Vilos, A. Swami, J. Shi and O. C. Farokhzad, *J. Controlled Release*, 2014, **190**, 485–499.
- C. Ornelas-Megiatto, P. N. Shah, P. R. Wich, J. L. Cohen, J. A. Tagaev, J. A. Smolen, B. D. Wright, M. J. Panzner, W. J. Youngs and J. M. Fréchet, *Mol. Pharmaceutics*, 2012, **9**, 3012–3022.
- J. Yang, S. Han, H. Zheng, H. Dong and J. Liu, *Carbohydr. Polym.*, 2015, **123**, 53–66.
- M. V. Tzoumaki, T. Moschakis, V. Kiosseoglou and C. G. Biliaderis, *Food Hydrocolloids*, 2011, **25**, 1521–1529.
- B. P. Binks and C. P. Whitby, *Colloids Surf., A*, 2005, **253**, 105–115.
- B. T. Nguyen, T. Nicolai and L. Benyahia, *Langmuir*, 2013, **29**, 10658–10664.
- M. Pavlovic, A. Plucinski, L. Zeininger and B. V. K. J. Schmidt, *Chem. Commun.*, 2020, **56**, 6814–6817.



- 24 Z. Liu, Y. Jiao, Y. Wang, C. Zhou and Z. Zhang, *Adv. Drug Delivery Rev.*, 2008, **60**, 1650–1662.
- 25 S. Mizrahy and D. Peer, *Chem. Soc. Rev.*, 2012, **41**, 2623–2640.
- 26 R. Heo, D. G. You, W. Um, K. Y. Choi, S. Jeon, J.-S. Park, Y. Choi, S. Kwon, K. Kim and I. C. Kwon, *Biomaterials*, 2017, **131**, 15–26.
- 27 M. Q. Guo, X. Hu, C. Wang and L. Ai, in *Polysaccharides: Structure and Solubility*, ed. Z. Xu, IntechOpen, 2017, pp. 7–21.
- 28 A. D. Baldwin and K. L. Kiick, *Pept. Sci.*, 2010, **94**, 128–140.
- 29 M. Gericke, P. Schulze and T. Heinze, *Macromol. Biosci.*, 2020, **20**, 1900415.
- 30 R. Brayner, F. Fiévet and T. Coradin, in *Nanomaterials: A Danger or a Promise? A chemical and biological perspective*, ed. J. Allouche, Springer-Verlag, 2013, pp. 27–74.
- 31 J. C. Cazotti, A. T. Fritz, O. Garcia-Valdez, N. M. Smeets, M. A. Dubé and M. F. Cunningham, *Carbohydr. Polym.*, 2020, **228**, 115384.
- 32 S. K. Nitta and K. Numata, *Int. J. Mol. Sci.*, 2013, **14**, 1629–1654.
- 33 S. Raveendran, A. K. Rochani, T. Maekawa and D. S. Kumar, *Materials*, 2017, **10**, 929.
- 34 K. Y. Lee and D. J. Mooney, *Prog. Polym. Sci.*, 2012, **37**, 106–126.
- 35 L. Li, R. Ni, Y. Shao and S. Mao, *Carbohydr. Polym.*, 2014, **103**, 1–11.
- 36 B. Medronho, A. Romano, M. G. Miguel, L. Stigsson and B. Lindman, *Cellulose*, 2012, **19**, 581–587.
- 37 L. Bacakova, K. Novotná and M. Parizek, *Physiol. Res.*, 2014, **63**, S29.
- 38 D. Klemm, B. Heublein, H. P. Fink and A. Bohn, *Angew. Chem., Int. Ed.*, 2005, **44**, 3358–3393.
- 39 L. Liu, Y. Ding, S. Liu, S. Wang, Y. Fang and M. Lyu, *Qual. Assur. Saf. Crops Foods*, 2019, **11**, 53–59.
- 40 E. Khalikova, P. Susi and T. Korpela, *Microbiol. Mol. Biol. Rev.*, 2005, **69**, 306–325.
- 41 D. Domene-López, J. C. García-Quesada, I. Martin-Gullon and M. G. Montalbán, *Polymers*, 2019, **11**, 1084.
- 42 I. Šimkovic, M. Hricovíni, L. Šoltés, R. Mendichi and C. Cosentino, *Carbohydr. Polym.*, 2000, **41**, 9–14.
- 43 G. Tripodo, A. Trapani, M. L. Torre, G. Giammona, G. Trapani and D. Mandracchia, *Eur. J. Pharm. Biopharm.*, 2015, **97**, 400–416.
- 44 R. S. Singh, G. K. Saini and J. F. Kennedy, *Carbohydr. Polym.*, 2008, **73**, 515–531.
- 45 K. Janes, P. Calvo and M. Alonso, *Adv. Drug Delivery Rev.*, 2001, **47**, 83–97.
- 46 A. Singla and M. Chawla, *J. Pharm. Pharmacol.*, 2001, **53**, 1047–1067.
- 47 A. Nešić, G. Cabrera-Barjas, S. Dimitrijević-Branković, S. Davidović, N. Radovanović and C. Delattre, *Molecules*, 2020, **25**, 135.
- 48 J. H. Park, G. Saravanakumar, K. Kim and I. C. Kwon, *Adv. Drug Delivery Rev.*, 2010, **62**, 28–41.
- 49 S. M. Ahsan, M. Thomas, K. K. Reddy, S. G. Sooraparaju, A. Asthana and I. Bhatnagar, *Int. J. Biol. Macromol.*, 2018, **110**, 97–109.
- 50 B. R. Shah, Y. Li, W. Jin, Y. An, L. He, Z. Li, W. Xu and B. Li, *Food Hydrocolloids*, 2016, **52**, 369–377.
- 51 T. Kean and M. Thanou, *Adv. Drug Delivery Rev.*, 2010, **62**, 3–11.
- 52 R. J. Moon, A. Martini, J. Nairn, J. Simonsen and J. Youngblood, *Chem. Soc. Rev.*, 2011, **40**, 3941–3994.
- 53 M. A. Hubbe, O. J. Rojas, L. A. Lucia and M. Sain, *BioResources*, 2008, **3**, 929–980.
- 54 K. Chi, H. Wang and J. M. Catchmark, *Food Hydrocolloids*, 2020, **103**, 105696.
- 55 R. Zafar, K. M. Zia, S. Tabasum, F. Jabeen, A. Noreen and M. Zuber, *Int. J. Biol. Macromol.*, 2016, **92**, 1012–1024.
- 56 A. Dufresne and J. Castaño, *Starch/Staerke*, 2017, **69**, 1500307.
- 57 Y. Parajó, I. d'Angelo, A. Welle, M. Garcia-Fuentes and M. J. Alonso, *Drug Delivery*, 2010, **17**, 596–604.
- 58 M. Swierczewska, H. S. Han, K. Kim, J. Park and S. Lee, *Adv. Drug Delivery Rev.*, 2016, **99**, 70–84.
- 59 D. M. Suflet, G. C. Chitanu and J. Desbrières, *Carbohydr. Polym.*, 2010, **82**, 1271–1277.
- 60 Y. Chao and H. C. Shum, *Chem. Soc. Rev.*, 2020, **49**, 114–142.
- 61 C. R. Mace, O. Akbulut, A. A. Kumar, N. D. Shapiro, R. Derda, M. R. Patton and G. M. Whitesides, *J. Am. Chem. Soc.*, 2012, **134**, 9094–9097.
- 62 M. Naessens, A. Cerdobbel, W. Soetaert and E. J. Vandamme, *J. Chem. Technol. Biotechnol.*, 2005, **80**, 845–860.
- 63 A. Banerjee and R. Bandopadhyay, *Int. J. Biol. Macromol.*, 2016, **87**, 295–301.
- 64 N. H. Silva, C. Vilela, A. Almeida, I. M. Marrucho and C. S. Freire, *Food Hydrocolloids*, 2018, **77**, 921–930.
- 65 C. A. Cozzolino, G. Castelli, S. Trabattoni and S. Farris, *Food Packag. Shelf Life*, 2016, **8**, 50–55.
- 66 V. D. Prajapati, G. K. Jani and S. M. Khanda, *Carbohydr. Polym.*, 2013, **95**, 540–549.
- 67 M. Gupta and A. K. Gupta, *J. Controlled Release*, 2004, **99**, 157–166.
- 68 E. Cevher, S. K. Salomon, A. Makrakis, X. W. Li, S. Brocchini and H. O. Alpar, *J. Microencapsulation*, 2015, **32**, 755–768.
- 69 E. Cevher, S. K. Salomon, S. Somavarapu, S. Brocchini and H. O. Alpar, *J. Microencapsulation*, 2015, **32**, 769–783.
- 70 J. Willersinn, A. Bogomolova, M. B. Cabré and B. V. K. J. Schmidt, *Polym. Chem.*, 2017, **8**, 1244–1254.
- 71 S. M. Brosnan, H. Schlaad and M. Antonietti, *Angew. Chem., Int. Ed.*, 2015, **54**, 9715–9718.
- 72 R. B. Lira, J. Willersinn, B. V. K. J. Schmidt and R. Dimova, *Macromolecules*, 2020, **53**, 10179–10188.
- 73 J. Willersinn and B. V. K. J. Schmidt, *J. Polym. Sci., Part A: Polym. Chem.*, 2017, **55**, 3757–3766.
- 74 S. Farris, I. U. Unalan, L. Introzzi, J. M. Fuentes-Alventosa and C. A. Cozzolino, *J. Appl. Polym. Sci.*, 2014, **131**, 40539.
- 75 J. P. Paques, E. van der Linden, C. J. van Rijn and L. M. Sagis, *Adv. Colloid Interface Sci.*, 2014, **209**, 163–171.
- 76 A. D. Augst, H. J. Kong and D. J. Mooney, *Macromol. Biosci.*, 2006, **6**, 623–633.
- 77 M. S. Hasnain, A. K. Nayak, M. Kurakula and M. N. Hoda, *Alginates in Drug Delivery*, Elsevier, 2020, pp. 129–152.



- 78 L. Wells and H. Sheardown, *Eur. J. Pharm. Biopharm.*, 2007, **65**, 329–335.
- 79 M. B. Evangelista, S. X. Hsiang, R. Fernandes, P. Sampaio, H.-J. Kong, C. C. Barrias, R. Salema, M. A. Barbosa, D. J. Mooney and P. L. Granja, *Biomaterials*, 2007, **28**, 3644–3655.
- 80 J. O. You and C. A. Peng, *Macromol. Symp.*, 2005, **219**, 147–153.
- 81 A. Zahoor, S. Sharma and G. Khuller, *Int. J. Antimicrob. Agents*, 2005, **26**, 298–303.
- 82 S. K. Motwani, S. Chopra, S. Talegaonkar, K. Kohli, F. J. Ahmad and R. K. Khar, *Eur. J. Pharm. Biopharm.*, 2008, **68**, 513–525.
- 83 R. Yegappan, V. Selvaprithiviraj, S. Amirthalingam and R. Jayakumar, *Carbohydr. Polym.*, 2018, **198**, 385–400.
- 84 A. L. Daniel-da-Silva, L. Ferreira, A. M. Gil and T. Trindade, *J. Colloid Interface Sci.*, 2011, **355**, 512–517.
- 85 S. Rodrigues, A. M. R. da Costa and A. Grenha, *Carbohydr. Polym.*, 2012, **89**, 282–289.
- 86 S. David, C. S. Levi, L. Fahoum, Y. Ungar, E. G. Meyron-Holtz, A. Shpigelman and U. Lesmes, *Food Funct.*, 2018, **9**, 1344–1352.
- 87 S. A. Agnihotri, N. N. Mallikarjuna and T. M. Aminabhavi, *J. Controlled Release*, 2004, **100**, 5–28.
- 88 C. Alvarez-Lorenzo, B. Blanco-Fernandez, A. M. Puga and A. Concheiro, *Adv. Drug Delivery Rev.*, 2013, **65**, 1148–1171.
- 89 S. P. Strand, S. Lelu, N. K. Reitan, C. de Lange Davies, P. Artursson and K. M. Vårum, *Biomaterials*, 2010, **31**, 975–987.
- 90 J. Valente, P. Pereira, A. Sousa, J. Queiroz and F. Sousa, *Polymers*, 2021, **13**, 793.
- 91 J. Willersinn and B. V. K. J. Schmidt, *Polym. Chem.*, 2018, **9**, 1626–1637.
- 92 T. Harifi and M. Montazer, *Carbohydr. Polym.*, 2012, **88**, 1125–1140.
- 93 E. Campos, P. Coimbra and M. Gil, *Polym. Bull.*, 2013, **70**, 549–561.
- 94 E. Chiesa, R. Dorati, S. Pisani, B. Conti, G. Bergamini, T. Modena and I. Genta, *Pharmaceutics*, 2018, **10**, 267.
- 95 J. Berger, M. Reist, J. M. Mayer, O. Felt, N. Peppas and R. Gurny, *Eur. J. Pharm. Biopharm.*, 2004, **57**, 19–34.
- 96 K. Janes, P. Calvo and M. Alonso, *Adv. Drug Delivery Rev.*, 2001, **47**, 83–97.
- 97 J. Berger, M. Reist, J. M. Mayer, O. Felt and R. Gurny, *Eur. J. Pharm. Biopharm.*, 2004, **57**, 35–52.
- 98 S. Ye, C. Wang, X. Liu, Z. Tong, B. Ren and F. Zeng, *J. Controlled Release*, 2006, **112**, 79–87.
- 99 L. Richert, P. Lavalle, E. Payan, X. Z. Shu, G. D. Prestwich, J.-F. Stoltz, P. Schaaf, J.-C. Voegel and C. Picart, *Langmuir*, 2004, **20**, 448–458.
- 100 S. Wang, F. Fontana, M.-A. Shahbazi and H. A. Santos, *Chem. Commun.*, 2021, DOI: 10.1039/D1CC00811K.
- 101 C. Allen, D. Maysinger and A. Eisenberg, *Colloids Surf., B*, 1999, **16**, 3–27.
- 102 E. Akiyama, N. Morimoto, P. Kujawa, Y. Ozawa, F. M. Winnik and K. Akiyoshi, *Biomacromolecules*, 2007, **8**, 2366–2373.
- 103 B. B. Breitenbach, I. Schmid and P. R. Wich, *Biomacromolecules*, 2017, **18**, 2839–2848.
- 104 D. A. Ossipov, *Expert Opin. Drug Delivery*, 2010, **7**, 681–703.
- 105 Y. Liu, G. Yang, D. Zou, Y. Hui, K. Nigam, A. P. Middelberg and C.-X. Zhao, *Ind. Eng. Chem. Res.*, 2019, **59**, 4134–4149.
- 106 A. Maaz, W. Abdelwahed, I. Tekko and S. Trefi, *Int. J. Acad. Sci. Res.*, 2015, **3**, 1–12.
- 107 C. J. M. Rivas, M. Tarhini, W. Badri, K. Miladi, H. Greige-Gerges, Q. A. Nazari, S. A. G. Rodríguez, R. Á. Román, H. Fessi and A. Elaissari, *Int. J. Pharm.*, 2017, **532**, 66–81.
- 108 F. Lince, D. L. Marchisio and A. A. Barresi, *J. Colloid Interface Sci.*, 2008, **322**, 505–515.
- 109 K. Miladi, S. Sfar, H. Fessi and A. Elaissari, *Polymer nanoparticles for nanomedicines*, Springer, 2016, pp. 17–53.
- 110 C. G. Barreras-Urbina, B. Ramírez-Wong, G. A. López-Ahumada, S. E. Burrueal-Ibarra, O. Martínez-Cruz, J. A. Tapia-Hernández and F. Rodríguez Felix, *Int. J. Food Prop.*, 2016, **19**, 1912–1923.
- 111 S. Ding, N. Anton, T. F. Vandamme and C. A. Serra, *Expert Opin. Drug Delivery*, 2016, **13**, 1447–1460.
- 112 S. F. Chin, S. C. Pang and S. H. Tay, *Carbohydr. Polym.*, 2011, **86**, 1817–1819.
- 113 M. R. Kulterer, M. Reischl, V. E. Reichel, S. Hribernik, M. Wu, S. Köstler, R. Kargl and V. Ribitsch, *Colloids Surf., A*, 2011, **375**, 23–29.
- 114 E. Aschenbrenner, K. Bley, K. Koynov, M. Makowski, M. Kappl, K. Landfester and C. K. Weiss, *Langmuir*, 2013, **29**, 8845–8855.
- 115 X. Yan, R. A. N. S. Ramos, P. Alcouffe, L. E. Munoz, R. O. Bilyy, F. Ganachaud and J. Bernard, *Biomacromolecules*, 2019, **20**, 3915–3923.
- 116 M. Leonard, M. R. De Boisseson, P. Hubert, F. Dalencon and E. Dellacherie, *J. Controlled Release*, 2004, **98**, 395–405.
- 117 E. Kizilay, A. B. Kayitmazer and P. L. Dubin, *Adv. Colloid Interface Sci.*, 2011, **167**, 24–37.
- 118 F. Vecchies, P. Sacco, E. Decleva, R. Menegazzi, D. Porrelli, I. Donati, G. Turco, S. Paoletti and E. Marsich, *Biomacromolecules*, 2018, **19**, 3936–3944.
- 119 J. T. G. Overbeek and M. Voorn, *J. Cell. Comp. Physiol.*, 1957, **49**, 7–26.
- 120 C. E. Sing, *Adv. Colloid Interface Sci.*, 2017, **239**, 2–16.
- 121 A. Drogoz, L. David, C. Rochas, A. Domard and T. Delair, *Langmuir*, 2007, **23**, 10950–10958.
- 122 Y. Liu, M. Kong, X. J. Cheng, Q. Q. Wang, L. M. Jiang and X. G. Chen, *Carbohydr. Polym.*, 2013, **94**, 309–316.
- 123 B. Sarmiento, A. Ribeiro, F. Veiga and D. Ferreira, *Colloids Surf., B*, 2006, **53**, 193–202.
- 124 L. Liu, Y. Xu, P. Zhang, J. You, W. Li, Y. Chen, R. Li, B. Rui and H. Dou, *Langmuir*, 2020, **36**, 8580–8588.
- 125 F. Furlani, P. Sacco, E. Decleva, R. Menegazzi, I. Donati, S. Paoletti and E. Marsich, *ACS Appl. Mater. Interfaces*, 2019, **11**, 9794–9803.
- 126 A. Gennari, J. M. R. de la Rosa, E. Hohn, M. Pelliccia, E. Lallana, R. Donno, A. Tirella and N. Tirelli, *Beilstein J. Nanotechnol.*, 2019, **10**, 2594–2608.
- 127 N. Eghbal and R. Choudhary, *LWT*, 2018, **90**, 254–264.
- 128 Y. P. Timilsena, T. O. Akanbi, N. Khalid, B. Adhikari and C. J. Barrow, *Int. J. Biol. Macromol.*, 2019, **121**, 1276–1286.



- 129 C. E. Sing and S. L. Perry, *Soft Matter*, 2020, **16**, 2885–2914.
- 130 L. Zhou, H. Shi, Z. Li and C. He, *Macromol. Rapid Commun.*, 2020, **41**, 2000149.
- 131 J. R. Magana, C. Sproncken and I. K. Voets, *Polymers*, 2020, **12**, 1953.
- 132 J. P. Rao and K. E. Geckeler, *Prog. Polym. Sci.*, 2011, **36**, 887–913.
- 133 Q. Cao, T. Heil, B. Kumru, M. Antonietti and B. V. K. J. Schmidt, *Polym. Chem.*, 2019, **10**, 5315–5323.
- 134 K. Sakamoto, R. Lochhead, H. Maibach and Y. Yamashita, *Cosmetic science and technology: theoretical principles and applications*, Elsevier, 2017.
- 135 J. Esquena, *Curr. Opin. Colloid Interface Sci.*, 2016, **25**, 109–119.
- 136 J. Zhang, J. Hwang, M. Antonietti and B. V. K. J. Schmidt, *Biomacromolecules*, 2018, **20**, 204–211.
- 137 C. Vauthier and K. Bouchemal, *Pharm. Res.*, 2009, **26**, 1025–1058.
- 138 B. Lupo, A. Maestro, J. M. Gutiérrez and C. González, *Food Hydrocolloids*, 2015, **49**, 25–34.
- 139 P. R. Ravi, R. Vats, J. Balija, S. P. N. Adapa and N. Aditya, *Carbohydr. Polym.*, 2014, **110**, 320–328.
- 140 J. M. Shin, S. R. Hwang, R. Heo, G. Saravanakumar and J. H. Park, *Polym. Degrad. Stab.*, 2014, **109**, 398–404.
- 141 J. P. Paques, E. van der Linden, C. J. van Rijn and L. M. Sagis, *Food Hydrocolloids*, 2013, **31**, 428–434.
- 142 H. S. Min, S. Son, T. W. Lee, H. Koo, H. Y. Yoon, J. H. Na, Y. Choi, J. H. Park, J. Lee and M. H. Han, *Adv. Funct. Mater.*, 2013, **23**, 5518–5529.
- 143 S. Manchun, C. R. Dass and P. Sriamornsak, *Carbohydr. Polym.*, 2014, **101**, 650–655.
- 144 M. Gupta and A. K. Gupta, *J. Pharm. Pharm. Sci.*, 2004, **7**, 38–46.
- 145 A. Maghsoudi, F. Yazdian, S. Shahmoradi, L. Ghaderi, M. Hemati and G. Amoabediny, *Mater. Sci. Eng., C*, 2017, **75**, 1259–1267.
- 146 J. Yao, L. Zhang, J. Zhou, H. Liu and Q. Zhang, *Mol. Pharmaceutics*, 2013, **10**, 1080–1091.
- 147 K. A. Janes, M. P. Fresneau, A. Marazuela, A. Fabra and M. A. J. Alonso, *J. Controlled Release*, 2001, **73**, 255–267.
- 148 X. Li, N. Anton, C. Arpagaus, F. Belleteix and T. F. Vandamme, *J. Controlled Release*, 2010, **147**, 304–310.
- 149 C. Liu, S. Zhang, D. J. McClements, D. Wang and Y. Xu, *J. Agric. Food Chem.*, 2019, **67**, 5113–5121.
- 150 X. Pei, K. Zhai, C. Wang, Y. Deng, Y. Tan, B. Zhang, Y. Bai, K. Xu and P. Wang, *Langmuir*, 2019, **35**, 7222–7230.
- 151 A. Plucinski, J. Willersinn, R. B. Lira, R. Dimova and B. V. K. J. Schmidt, *Macromol. Chem. Phys.*, 2020, **221**, 2000053.
- 152 L. Liu, Y. Wang, Q. Hu, T. Li, P. Ma, H. Zhang, M. Du, M. Chen and W. Dong, *ACS Sustainable Chem. Eng.*, 2019, **7**, 13081–13088.
- 153 Y. Wang, Q. Hu, T. Li, P. Ma, S. Zhang, M. Du, M. Chen, H. Zhang and W. Dong, *Ind. Eng. Chem. Res.*, 2018, **57**, 13048–13054.
- 154 H. Kim, M. Shin, S. Han, W. Kwon and S. K. Hahn, *Biomacromolecules*, 2019, **20**, 2889–2903.
- 155 H. M. Aldawsari, H. K. Dhaliwal, B. M. Aljaeid, N. A. Alhakamy, Z. M. Banjar and M. M. Amiji, *Mol. Pharmaceutics*, 2018, **16**, 128–140.
- 156 Y. Liu, Y. Wang, C. Zhang, P. Zhou, Y. Liu, T. An, D. Sun, N. Zhang and Y. Wang, *ACS Appl. Mater. Interfaces*, 2014, **6**, 18712–18720.
- 157 C. Lemarchand, R. Gref and P. Couvreur, *Eur. J. Pharm. Biopharm.*, 2004, **58**, 327–341.
- 158 C. Lemarchand, R. Gref, C. Passirani, E. Garcion, B. Petri, R. Müller, D. Costantini and P. Couvreur, *Biomaterials*, 2006, **27**, 108–118.
- 159 J. C. Cazotti, A. T. Fritz, O. Garcia-Valdez, N. M. Smeets, M. A. Dubé and M. F. Cunningham, *Biomacromolecules*, 2020, **21**, 4492–4501.
- 160 A. Dufresne, *Molecules*, 2010, **15**, 4111–4128.
- 161 F. Tang, L. Zhang, J. Zhu, Z. Cheng and X. Zhu, *Ind. Eng. Chem. Res.*, 2009, **48**, 6216–6223.
- 162 S. Yuan, P. Zhang, Z. Yang, L. Lv, S. Tang and B. Liang, *Int. J. Biol. Macromol.*, 2018, **109**, 287–302.
- 163 M. V. G. Paixão, R. C. S. da Luz and R. de Carvalho Balaban, *J. Mol. Liq.*, 2019, **286**, 110792.
- 164 K. Peuler, N. Dimmitt and C.-C. Lin, *Carbohydr. Polym.*, 2020, **234**, 115901.
- 165 N. V. Rao, J. G. Rho, W. Um, P. K. Ek, V. Q. Nguyen, B. H. Oh, W. Kim and J. H. Park, *Pharmaceutics*, 2020, **12**, 931.
- 166 K. Y. Choi, K. H. Min, H. Y. Yoon, K. Kim, J. H. Park, I. C. Kwon, K. Choi and S. Y. Jeong, *Biomaterials*, 2011, **32**, 1880–1889.
- 167 M. Xu, J. Qian, A. Suo, H. Wang, X. Yong, X. Liu and R. Liu, *Carbohydr. Polym.*, 2013, **98**, 181–188.
- 168 H.-J. Cho, I.-S. Yoon, H. Y. Yoon, H. Koo, Y.-J. Jin, S.-H. Ko, J.-S. Shim, K. Kim, I. C. Kwon and D.-D. Kim, *Biomaterials*, 2012, **33**, 1190–1200.
- 169 K. Y. Choi, H. Y. Yoon, J.-H. Kim, S. M. Bae, R.-W. Park, Y. M. Kang, I.-S. Kim, I. C. Kwon, K. Choi and S. Y. Jeong, *ACS Nano*, 2011, **5**, 8591–8599.
- 170 C. Teng, Z. Chai, Z. Yuan, L. Ren, C. Lin, Z. Yan, W. He, C. Qin, L. Yang and X. Han, *Nanomedicine*, 2020, **24**, 102105.
- 171 H. S. Han, J. Lee, H. R. Kim, S. Y. Chae, M. Kim, G. Saravanakumar, H. Y. Yoon, D. G. You, H. Ko and K. Kim, *J. Controlled Release*, 2013, **168**, 105–114.
- 172 K. Y. Choi, E. J. Jeon, H. Y. Yoon, B. S. Lee, J. H. Na, K. H. Min, S. Y. Kim, S.-J. Myung, S. Lee and X. Chen, *Biomaterials*, 2012, **33**, 6186–6193.
- 173 J. Zhao, Y. Wang, Y. Ma, Y. Liu, B. Yan and L. Wang, *Carbohydr. Polym.*, 2019, **203**, 356–368.
- 174 S. Ganesh, A. K. Iyer, D. V. Morrissey and M. M. Amiji, *Biomaterials*, 2013, **34**, 3489–3502.
- 175 S. Ganesh, A. K. Iyer, F. Gattaceca, D. V. Morrissey and M. M. Amiji, *J. Controlled Release*, 2013, **172**, 699–706.
- 176 L. Mo, J. G. Song, H. Lee, M. Zhao, H. Y. Kim, Y. J. Lee, H. W. Ko and H.-K. Han, *Nanomedicine*, 2018, **14**, 557–567.
- 177 H. Y. Yoon, H. Koo, K. Y. Choi, I. C. Kwon, K. Choi, J. H. Park and K. Kim, *Biomaterials*, 2013, **34**, 5273–5280.
- 178 J. M. Shin, S. J. Oh, S. Kwon, V. Deepagan, M. Lee, S. H. Song, H.-J. Lee, S. Kim, K.-H. Song and T. W. Kim, *J. Controlled Release*, 2017, **267**, 181–190.



- 179 H. Lian, Y. Du, X. Chen, L. Duan, G. Gao, C. Xiao and X. Zhuang, *J. Colloid Interface Sci.*, 2017, **496**, 201–210.
- 180 S. Maiz-Fernández, L. Pérez-Álvarez, L. Ruiz-Rubio, R. Pérez González, V. Sáez-Martínez, J. Ruiz Pérez and J. L. Vilas-Vilela, *Polymers*, 2019, **11**, 742.
- 181 D. Bamberger, D. Hobernik, M. Konhäuser, M. Bros and P. R. Wich, *Mol. Pharmaceutics*, 2017, **14**, 4403–4416.
- 182 L. De Matteis, M. Alleva, I. Serrano-Sevilla, S. García-Embid, G. Stepien, M. Moros and J. M. De la Fuente, *Mar. Drugs*, 2016, **14**, 175.
- 183 S. A. Papadimitriou, D. S. Achilias and D. N. Bikiaris, *Int. J. Pharm.*, 2012, **430**, 318–327.
- 184 M. Nag, V. Gajbhiye, P. Kesharwani and N. K. Jain, *Colloids Surf., B*, 2016, **148**, 363–370.
- 185 A. Guţoaia, L. Schuster, S. Margutti, S. Laufer, B. Schlosshauer, R. Krastev, D. Stoll and H. Hartmann, *Carbohydr. Polym.*, 2016, **143**, 25–34.
- 186 W. E. Rudzinski, A. Palacios, A. Ahmed, M. A. Lane and T. M. Aminabhavi, *Carbohydr. Polym.*, 2016, **147**, 323–332.
- 187 H. Ragelle, S. Colombo, V. Pourcelle, K. Vanvarenberg, G. Vandermeulen, C. Bouzin, J. Marchand-Brynaert, O. Feron, C. Foged and V. Pr eat, *J. Controlled Release*, 2015, **211**, 1–9.
- 188 C. Corbet, H. Ragelle, V. Pourcelle, K. Vanvarenberg, J. Marchand-Brynaert, V. Pr eat and O. Feron, *J. Controlled Release*, 2016, **223**, 53–63.
- 189 X. Yan, P. Alcouffe, J. Bernard and F. Ganachaud, *Biomacromolecules*, 2020, **21**, 4591–4598.
- 190 N. Al Nakeeb, J. Willersinn and B. V. K. J. Schmidt, *Biomacromolecules*, 2017, **18**, 3695–3705.
- 191 T. Nishimura, S. Shishi, Y. Sasaki and K. Akiyoshi, *J. Am. Chem. Soc.*, 2020, **142**, 11784–11790.
- 192 X. Dong, Z. Wu, Y. Wang, T. Li, X. Zhang, H. Yuan, B. Xia, P. Ma, M. Chen and W. Dong, *Polym. Test.*, 2021, **93**, 106926.
- 193 X. Dong, Z. Wu, Y. Wang, T. Li, X. Zhang, H. Yuan, J. Jiang, B. Xia, P. Ma and M. Chen, *Mater. Lett.*, 2021, **289**, 129400.
- 194 X. Dong, L. Liu, Y. Wang, T. Li, Z. Wu, H. Yuan, P. Ma, D. Shi, M. Chen and W. Dong, *Carbohydr. Polym.*, 2021, **254**, 117321.
- 195 G. Wang, S. Li, Y. Feng, Y. Hu, G. Zhao and W. Jiang, *Carbohydr. Polym.*, 2020, **249**, 116795.
- 196 N. Elsaid, T. L. Jackson, Z. Elsaid, A. Alqathama and S. Somavarapu, *Mol. Pharmaceutics*, 2016, **13**, 2923–2940.
- 197 Q. Wang, S. Jamal, M. S. Detamore and C. Berkland, *J. Biomed. Mater. Res., Part A*, 2011, **96**, 520–527.
- 198 Y. Liu, R. Lipowsky and R. Dimova, *Front. Chem.*, 2019, **7**, 213.
- 199 M. Iqbal, Y. Tao, S. Xie, Y. Zhu, D. Chen, X. Wang, L. Huang, D. Peng, A. Sattar and M. A. B. Shabbir, *Biol. Proced. Online*, 2016, **18**, 1–18.
- 200 O. Akbulut, C. R. Mace, R. V. Martinez, A. A. Kumar, Z. Nie, M. R. Patton and G. M. Whitesides, *Nano Lett.*, 2012, **12**, 4060–4064.
- 201 P.-Å. Albertsson, *Nature*, 1958, **182**, 709–711.
- 202 M. Pavlovic, A. Plucinski, J. Zhang, M. Antonietti, L. Zeininger and B. V. K. J. Schmidt, *Langmuir*, 2020, **36**, 1401–1408.
- 203 S. Ma, J. Thiele, X. Liu, Y. Bai, C. Abell and W. T. Huck, *Small*, 2012, **8**, 2356–2360.
- 204 T. Watanabe, I. Motohiro and T. Ono, *Langmuir*, 2019, **35**, 2358–2367.
- 205 H. Yuan, Q. Ma, Y. Song, M. Y. Tang, Y. K. Chan and H. C. Shum, *Macromol. Chem. Phys.*, 2017, **218**, 1600422.
- 206 Y. Hu, S. Wang, A. Abbaspourrad and A. M. Ardekani, *Langmuir*, 2015, **31**, 1885–1891.
- 207 S. Mytnyk, I. Ziemecka, A. G. Olive, J. W. M. van der Meer, K. A. Totlani, S. Oldenhof, M. T. Kreutzer, V. van Steijn and J. H. van Esch, *RSC Adv.*, 2017, **7**, 11331–11337.
- 208 M. Yamazaki and K. Iijima, *Polymers*, 2020, **12**, 435.
- 209 A. Bianchera and R. Bettini, *Expert Opin. Drug Delivery*, 2020, **17**, 1345–1359.
- 210 S. Salatin and A. Yari Khosroushahi, *J. Cell. Mol. Med.*, 2017, **21**, 1668–1686.
- 211 S. Chayed and F. M. Winnik, *Eur. J. Pharm. Biopharm.*, 2007, **65**, 363–370.
- 212 S. Uthaman, Y. Kim, J. Y. Lee, S. Pillarisetti, K. M. Huh and I.-K. Park, *ACS Appl. Mater. Interfaces*, 2020, **12**, 28004–28013.
- 213 B. B. Breitenbach, E. Steiert, M. Konhäuser, L.-M. Vogt, Y. Wang, S. H. Parekh and P. R. Wich, *Soft Matter*, 2019, **15**, 1423–1434.
- 214 K. Butzbach, M. Konhäuser, M. Fach, D. N. Bamberger, B. Breitenbach, B. Epe and P. R. Wich, *Polymers*, 2019, **11**, 896.
- 215 T. G. Dacoba, R. W. Omange, H. Li, J. Crecente-Campo, M. Luo and M. J. Alonso, *ACS Nano*, 2019, **13**, 4947–4959.
- 216 J. Fang, H. Nakamura and H. Maeda, *Adv. Drug Delivery Rev.*, 2011, **63**, 136–151.
- 217 M. Abid, M. Naveed, I. Azeem, A. Faisal, M. F. Nazar and B. Yameen, *Int. J. Pharm.*, 2020, **586**, 119605.
- 218 S. Kumari, B. Ram, D. Kumar, S. Ranote and G. S. Chauhan, *Mater. Sci. Energy Technol.*, 2018, **1**, 22–28.
- 219 S. Javanbakht and H. Namazi, *Mater. Sci. Eng., C*, 2018, **87**, 50–59.
- 220 Y. Li, C. Liu, Y. Tan, K. Xu, C. Lu and P. Wang, *Carbohydr. Polym.*, 2014, **110**, 87–94.
- 221 H. Hu, Y. Li, Q. Zhou, Y. Ao, C. Yu, Y. Wan, H. Xu, Z. Li and X. Yang, *ACS Appl. Mater. Interfaces*, 2016, **8**, 30833–30844.
- 222 I. Wasiak, A. Kulikowska, M. Janczewska, M. Michalak, I. A. Cymerman, A. Nagalski, P. Kallinger, W. W. Szymanski and T. Ciach, *PLoS One*, 2016, **11**, e0146237.
- 223 B. Li, Q. Wang, X. Wang, C. Wang and X. Jiang, *Carbohydr. Polym.*, 2013, **93**, 430–437.
- 224 B. Sarmiento, A. Ribeiro, F. Veiga, D. Ferreira and R. Neufeld, *Biomacromolecules*, 2007, **8**, 3054–3060.
- 225 H. Lindemann, M. Kühne, C. Grune, P. Warncke, S. Hofmann, A. Koschella, M. Godmann, D. Fischer, T. Heinzl and T. Heinze, *Macromol. Biosci.*, 2020, **20**, 2000039.
- 226 A. L. Urzedo, M. C. Gonçalves, M. H. Nascimento, C. B. Lombello, G. Nakazato and A. B. Seabra, *Mater. Sci. Eng., C*, 2020, **112**, 110933.
- 227 M. S. Huh, E. J. Lee, H. Koo, J. Y. Yhee, K. S. Oh, S. Son, S. Lee, S. H. Kim, I. C. Kwon and K. Kim, *Top. Curr. Chem.*, 2017, **375**, 31.



- 228 S. Salatin and M. Jelvehgari, *Pharm. Sci.*, 2017, **23**, 84–94.
- 229 P. R. Wich and J. M. Fréchet, *Aust. J. Chem.*, 2012, **65**, 15–19.
- 230 H. Yasar, D.-K. Ho, C. De Rossi, J. Herrmann, S. Gordon, B. Loretz and C.-M. Lehr, *Polymers*, 2018, **10**, 252.
- 231 Y. Yang, X. Liu, D. Zhang, W. Yu, H. Xie, J. Zheng and X. Ma, *J. Controlled Release*, 2011, **152**, e160–e161.
- 232 O. S. Muddineti, A. Shah, S. V. K. Rompicharla, B. Ghosh and S. Biswas, *Int. J. Biol. Macromol.*, 2018, **118**, 857–863.
- 233 H.-Q. Mao, K. Roy, V. L. Troung-Le, K. A. Janes, K. Y. Lin, Y. Wang, J. T. August and K. W. Leong, *J. Controlled Release*, 2001, **70**, 399–421.
- 234 J. L. Cohen, S. Schubert, P. R. Wich, L. Cui, J. A. Cohen, J. L. Mynar and J. M. Fréchet, *Bioconjugate Chem.*, 2011, **22**, 1056–1065.
- 235 L. Cui, J. L. Cohen, C. K. Chu, P. R. Wich, P. H. Kierstead and J. M. Fréchet, *J. Am. Chem. Soc.*, 2012, **134**, 15840–15848.
- 236 F. C. Moraes, L. M. F. Ramirez, R. Aid, S. Benadda, M. Maire, C. Chauvierre, J. C. Antunes, F. Chaubet and D. Letourneur, *Int. J. Pharm.*, 2021, **597**, 120302.
- 237 V. Y. Kosovrasti, L. V. Nechev and M. M. Amiji, *Mol. Pharmaceutics*, 2016, **13**, 3404–3416.
- 238 A. Grenha, M. E. Gomes, M. Rodrigues, V. E. Santo, J. F. Mano, N. M. Neves and R. L. Reis, *J. Biomed. Mater. Res., Part A*, 2010, **92**, 1265–1272.
- 239 P. Sahoo, K. H. Leong, S. Nyamathulla, Y. Onuki, K. Takayama and L. Y. Chung, *React. Funct. Polym.*, 2017, **119**, 145–155.
- 240 S. Abdelghany, M. Alkhalwaldeh and H. S. Alkhatib, *J. Drug Delivery Sci. Technol.*, 2017, **39**, 442–449.
- 241 S. Rochín-Wong, A. Rosas-Durazo, P. Zavala-Rivera, A. Maldonado, M. E. Martínez-Barbosa, I. Vélaz and J. Tánori, *Polymers*, 2018, **10**, 760.
- 242 T. Miao, J. Wang, Y. Zeng, G. Liu and X. Chen, *Adv. Sci.*, 2018, **5**, 1700513.
- 243 C. Qi, X. Yan, C. Huang, A. Melerzanov and Y. Du, *Protein Cell*, 2015, **6**, 638–653.
- 244 P. Koria, *BioDrugs*, 2012, **26**, 163–175.
- 245 M. Rajam, S. Pulavendran, C. Rose and A. Mandal, *Int. J. Pharm.*, 2011, **410**, 145–152.
- 246 F.-m. Chen, Z.-w. Ma, G.-y. Dong and Z.-f. Wu, *Acta Pharmacol. Sin.*, 2009, **30**, 485–493.
- 247 G. Q. Teixeira, C. L. Pereira, F. Castro, J. R. Ferreira, M. Gomez-Lazaro, P. Aguiar, M. A. Barbosa, C. Neidlinger-Wilke and R. M. Goncalves, *Acta Biomater.*, 2016, **42**, 168–179.
- 248 M. M. Hasani-Sadrabadi, S. P. Hajrezaei, S. H. Emami, G. Bahlakeh, L. Daneshmandi, E. Dashtimoghadam, E. Seyedjafari, K. I. Jacob and L. Tayebi, *Nanomedicine*, 2015, **11**, 1809–1819.
- 249 I. Aranaz, N. Acosta, C. Civera, B. Elorza, J. Mingo, C. Castro, M. De los Llanos Gandía and A. Heras Caballero, *Polymers*, 2018, **10**, 213.
- 250 Y. Yang, Z. Fang, X. Chen, W. Zhang, Y. Xie, Y. Chen, Z. Liu and W. Yuan, *Front. Pharmacol.*, 2017, **8**, 287.
- 251 J. Tang, P. J. Quinlan and K. C. Tam, *Soft Matter*, 2015, **11**, 3512–3529.
- 252 U. Bains and R. Pal, *Appl. Sci.*, 2019, **9**, 4044.
- 253 K. W. Muhammad, N. Akhtar, M. S. K. Haji, R. Mustafa and G. Murtaza, *Lat. Am. J. Pharm.*, 2014, **33**, 731–738.
- 254 S. Simovic, N. Ghouchi-Eskandar and C. Prestidge, *J. Drug Delivery Sci. Technol.*, 2011, **21**, 123–133.
- 255 W. Wang, G. Du, C. Li, H. Zhang, Y. Long and Y. Ni, *Carbohydr. Polym.*, 2016, **151**, 1–8.
- 256 S. B. Haaj, W. Thielemans, A. Magnin and S. Boufi, *ACS Appl. Mater. Interfaces*, 2014, **6**, 8263–8273.
- 257 C. Li, P. Sun and C. Yang, *Starch/Staerke*, 2012, **64**, 497–502.
- 258 K. Wang, Y. Hong, Z. Gu, L. Cheng, Z. Li and C. Li, *Int. J. Biol. Macromol.*, 2020, **155**, 273–285.
- 259 J. Han, F. Chen, C. Gao, Y. Zhang and X. Tang, *Int. J. Biol. Macromol.*, 2020, **157**, 202–211.
- 260 H. Liu, C. Wang, S. Zou, Z. Wei and Z. Tong, *Langmuir*, 2012, **28**, 11017–11024.
- 261 S. Ge, L. Xiong, M. Li, J. Liu, J. Yang, R. Chang, C. Liang and Q. Sun, *Food Chem.*, 2017, **234**, 339–347.
- 262 X. Pei, K. Zhai, X. Liang, Y. Deng, Y. Tan, P. Wang and K. Xu, *Langmuir*, 2017, **33**, 3787–3793.
- 263 Y. Tan, K. Xu, C. Liu, Y. Li, C. Lu and P. Wang, *Carbohydr. Polym.*, 2012, **88**, 1358–1363.
- 264 Y. Tan, K. Xu, C. Niu, C. Liu, Y. Li, P. Wang and B. P. Binks, *Food Hydrocolloids*, 2014, **36**, 70–75.
- 265 P. Shao, H. Zhang, B. Niu and W. Jin, *Int. J. Biol. Macromol.*, 2018, **118**, 2032–2039.
- 266 C. Li, Y. Li, P. Sun and C. Yang, *Colloids Surf., A*, 2013, **431**, 142–149.
- 267 M. Matos, A. Marefati, R. Bordes, G. Gutiérrez and M. Rayner, *Carbohydr. Polym.*, 2017, **169**, 127–138.
- 268 K. Zhai, F. Zhang, C. Wang, X. Pei, Y. Tan, Y. Bai, B. Zhang, Y. Wang, K. Xu and P. Wang, *Polym. Adv. Technol.*, 2020, **31**, 1321–1329.
- 269 V. Maingret, C. Courrégelongue, V. Schmitt and V. Héroguez, *Biomacromolecules*, 2020, **21**, 5358–5368.
- 270 A. Ammala, *Int. J. Cosmet. Sci.*, 2013, **35**, 113–124.
- 271 M. P. Lupo, *Clin. Dermatol.*, 2001, **19**, 467–473.
- 272 D.-G. Kim, Y.-I. Jeong, C. Choi, S.-H. Roh, S.-K. Kang, M.-K. Jang and J.-W. Nah, *Int. J. Pharm.*, 2006, **319**, 130–138.
- 273 R. Harris, E. Lecumberri, I. Mateos-Aparicio, M. Mengibar and A. Heras, *Carbohydr. Polym.*, 2011, **84**, 803–806.
- 274 P. Ganesan and D.-K. Choi, *Int. J. Nanomed.*, 2016, **11**, 1987.
- 275 P. Morganti, M. Palombo, G. Tishchenko, V. E. Yudin, F. Guarneri, M. Cardillo, P. Del Ciotto, F. Carezzi, G. Morganti and G. Fabrizi, *Cosmetics*, 2014, **1**, 140–158.
- 276 B. N. Matos, T. A. Reis, T. Gratieri and G. M. Gelfuso, *Int. J. Biol. Macromol.*, 2015, **75**, 225–229.
- 277 Q. Huang, H. Yu and Q. Ru, *J. Food Sci.*, 2010, **75**, R50–R57.
- 278 C. Moraru, Q. Huang, P. Takhistov, H. Dogan and J. Kokini, *Global issues in food science and technology*, Elsevier, 2009, pp. 369–399.
- 279 A. Bahrami, R. Delshadi, E. Assadpour, S. M. Jafari and L. Williams, *Adv. Colloid Interface Sci.*, 2020, **278**, 102140.
- 280 A. S. Kritchenkov, A. R. Egorov, N. V. Dubashynskaya, O. V. Volkova, L. A. Zabodalova, E. P. Suchkova, A. V. Kurliuk,



- T. V. Shakola and A. P. Dysin, *Int. J. Biol. Macromol.*, 2019, **134**, 480–486.
- 281 L. Lin, Y. Gu and H. Cui, *Food Packag. Shelf Life*, 2019, **19**, 86–93.
- 282 M. V. Lorevice, C. G. Otoni, M. R. de Moura and L. H. C. Mattoso, *Food Hydrocolloids*, 2016, **52**, 732–740.
- 283 A. Arora and G. Padua, *J. Food Sci.*, 2010, **75**, R43–R49.
- 284 S. Shankar, L.-F. Wang and J.-W. Rhim, *Food Packag. Shelf Life*, 2019, **21**, 100363.
- 285 L. Dai, C. Qiu, L. Xiong and Q. Sun, *Food Chem.*, 2015, **174**, 82–88.
- 286 S. Jiang, C. Liu, X. Wang, L. Xiong and Q. Sun, *LWT-Food Sci. Technol.*, 2016, **69**, 251–257.
- 287 D. J. McClements, *J. Food Sci.*, 2015, **80**, N1602–N1611.
- 288 Y. D. Livney, *Curr. Opin. Food Sci.*, 2015, **3**, 125–135.
- 289 R. F. Gonçalves, J. T. Martins, C. M. Duarte, A. A. Vicente and A. C. Pinheiro, *Trends Food Sci. Technol.*, 2018, **78**, 270–291.
- 290 F. Liu, C. Ma, Y. Gao and D. J. McClements, *Compr. Rev. Food Sci. Food Saf.*, 2017, **16**, 76–95.
- 291 D. Semyonov, O. Ramon, Y. Shoham and E. Shimoni, *Food Funct.*, 2014, **5**, 2463–2474.
- 292 L. Nieto-Suaza, L. Acevedo-Guevara, L. T. Sánchez, M. I. Pinzón and C. C. Villa, *Food Struct.*, 2019, **22**, 100131.
- 293 P. Puligundla, C. Mok, S. Ko, J. Liang and N. Recharla, *J. Funct. Foods*, 2017, **34**, 139–151.
- 294 J. Liang, H. Yan, P. Puligundla, X. Gao, Y. Zhou and X. Wan, *Food Hydrocolloids*, 2017, **69**, 286–292.
- 295 Y. Wei, C. Li, L. Dai, L. Zhang, J. Liu, L. Mao, F. Yuan and Y. Gao, *Food Funct.*, 2020, **11**, 9973–9983.

