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Palladium-catalyzed C–H glycosylation and retro Diels–Alder tandem reaction *via* structurally modified norbornadienes (smNBDs)†

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This report describes palladium-catalyzed C–H glycosylation and retro Diels–Alder tandem reaction *via* structurally modified norbornadienes (smNBDs). smNBDs were proposed to regulate the reactivity of the aryl-norbornadiene-palladacycle (ANP), including its high chemoselectivity and regioselectivity, which were the key to constructing C2 and C3 unsubstituted C4-glycosidic indoles. The scope of this substrate is extensive; the halogenated six-membered and five-membered glycosides were applied to the reaction smoothly, and *N*-alkyl (primary, secondary and tertiary) C4-glycosidic indoles can also be obtained by this method. In terms of mechanism, the key ANP intermediates characterized by X-ray single-crystal diffraction and further controlled experiments proved that the migration-insertion of smNBDs with phenylpalladium intermediate endows them with high chemo- and regioselectivity. Finally, density functional theory (DFT) calculation further verified the rationality of the mechanism.

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Introduction

A variety of glycoconjugates exhibit a range of essential biological functions, such as immune response, intercellular communication, and the bioactive modulation of natural products.^{1–3} Their unstable biochemical properties, such as glycosidic linkages undergoing hydrolysis in the living body, limit their practical development. Notably, the *C*-aryl glucoside skeleton, which is widely found in nature, has high biological metabolism stability, and *C*-glycosyl indole derivatives, which are widely used in the field of medicinal chemistry, have unique biological activities.^{4–14} In 1994, the Hofsteenge group first discovered α -*C*-mannosyltryptophan (α -*C*-Man-Trp) **I** from the Trp7 of ribonuclease.^{15–17} Due to their unique structure and biological activities such as cellular signaling and communication, these compounds have attracted considerable attention. Compounds **II** and **III** with 3-indolyl-*C*-glycoside skeleton are sodium-dependent glucose cotransporter 2 (SGLT2) inhibitors and are considered potential drugs for the therapy of type 2

diabetes.^{18,19} Compounds **IV** and **V** with 2-indolyl-*C*-glycoside skeleton were isolated from *Isatis indigotica* and they show significant cytotoxic activities against human liver cancer HepG2 cells, human myeloid leukemia Mata cells and human myeloid leukemia HL-60 (Scheme 1a).²⁰

Due to indolyl-*C*-glycosides' DNA modulatory properties and great potential in disease treatment, a variety of synthetic methods have been developed to construct these valuable scaffolds. These compounds are mainly synthesized using the following two approaches. The first method is to construct a 2-indolyl-*C*-glycoside skeleton by Larock indole synthesis with 2-iodoanilines and glycosyl alkyne as the substrate.^{21–27} The second approach is the transition-metal-catalyzed direct C–H functionalization of indole,^{28–35} and the most efficient is the Pd-catalyzed *ortho*-directed C–H glycosylation reactions to construct 2-indolyl-*C*-glycoside skeleton using glycosyl chloride

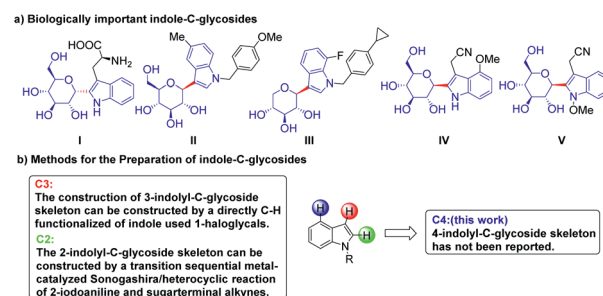
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Scheme 1 The study of glycosidic indoles was limited to C2 and C3 position substitution.



donors (Scheme 1b).³⁶ However, the construction of 4-indole-C-glycosides has not been reported. Therefore, the development of an efficient C–H glycosylation reaction to construct a 4-glycosylindole skeleton has important application value.

The Catellani reaction was discovered in 1997,³⁷ then Lautens expanded the chemical compatibility of Pd/NBE chemistry by utilizing phosphine ligands in 2000.³⁸ Later, it was named the Catellani–Lautens reaction system. The mechanism of the Catellani reaction is rather lengthy,^{39–49} so there are a lot of competitive reactions to produce a variety of by-products.⁵⁰ For example, the reductive elimination of aryl-norbornene-palladacycle (ANP) intermediate may take place to form the byproduct benzocyclobutene.^{51,52} In recent years, structurally modified norbornene (smNBE) strategy provides a good solution to this problem.

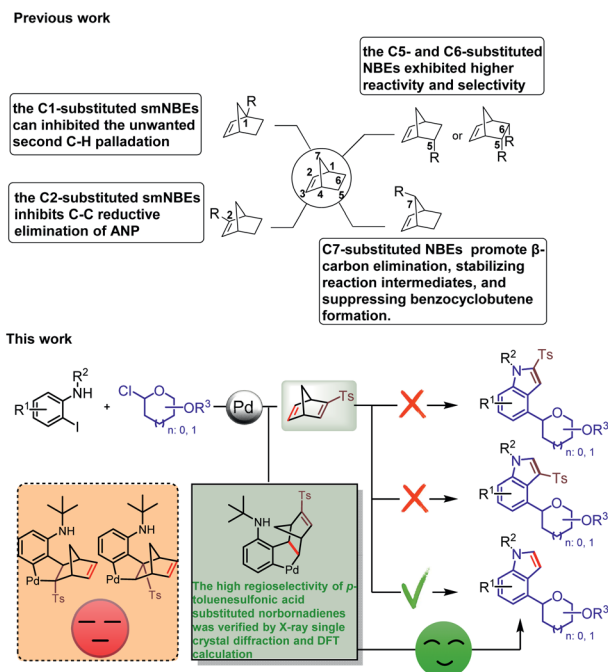
In 2015, Dong *et al.* first reported the smNBE (C5-amide-substituted),^{53,54} and since then, Yu,⁵⁵ Zhou,^{56–60} Gu,⁶¹ Ding,⁶² Cheng,⁶³ Liang⁶⁴ and others have developed numerous C2-, C5- and C6-substituted NBEs that exhibited unique reactivity and higher selectivity compared to the simple unsubstituted NBE. The amide-substituted NBE was reported by Dong *et al.*^{65–68} and the ester-substituted NBE reported by Yu *et al.* was the most effective type of C2-substituted smNBEs that greatly inhibit the C–C reductive elimination of ANP to form the by-product benzocyclobutanes and effectively reduce the overall activation barrier. Notably, Yu *et al.* made a breakthrough in using the C2-substituted smNBEs and quinoline-type ligand to realize *meta*-C–H activation with guiding groups. In 2018, Dong *et al.* developed a series of C1-substituted smNBEs that significantly inhibited the β -carbon elimination of NBE in *ortho*-unsubstituted and inhibited the unwanted second C–H

palladation.^{69,70} Recently, Dong *et al.* discovered that C7-substituted NBE can promote β -carbon elimination, stabilizing reaction intermediates, and tetrahydrobenzo[*b*]azepine derivatives were efficiently synthesized from *ortho*-unsubstituted iodobenzene (Scheme 2).⁷¹

Although high achievements have been made in the study of palladium/structurally modified norbornene (Pd/smNBEs) system, the palladium/structurally modified norbornadiene (Pd/NBD) system has not been reported yet. The main reason is that the migration-insertion reaction between smNBDs and phenylpalladium intermediates has four potential reaction sites, which is more challenging than our previous work to synthesize C4-aminated indoles using structurally unsubstituted NBD.⁷² In this reaction, we first used structurally modified norbornadiene (smNBDs) as the coupling partner of the Catellani reaction to realize the *ortho*-C–H glycosylation reaction and used the reverse Diels–Alder reaction of norbornadiene to realize the synthesis of C4-glycoside indole.

Results and discussion

Initially, we used *N*-(*tert*-butyl)-2-iodoaniline (**1a**) as a model substrate and glycosyl chloride donors (**2a**) as an *ortho*-glycosylation reagent, using palladium/norbornadiene (NBD) cooperative catalysis to investigate this reaction (Table 1). Unfortunately, the desired product (**4a**) was isolated in a 23% yield (entries 1) with NBD (**N1**, 3.5 equiv.), Pd(OAc)₂ (10 mol%), tri(2-furyl)phosphane (TFP, 20 mol%), and Cs₂CO₃ (4.0 equiv.) in 1,4-dioxane under Ar at 90 °C (12 h)–150 °C (24 h), and most of the reaction proceeded toward the intramolecular Buchwald coupling without *ortho*-glycosylation. We hypothesized that a wide spectrum of NBDs may have significant effects on the



Scheme 2 Synthesis of C4-glycosidic indoles *via* palladium/structurally modified norbornadiene (smNBDs).

Table 1 Optimization of reaction conditions^a

N1 (NBD), 23 % byproduct GC yield 19 % ^b	N2 , 19 %	N3 , 7 %	N4 , 5 %
N5 , 24 %	N6 , 11 %	N7 , 66 %	N8 , 79 %
N9 , 62 %	N10 , 49 %	N11 , trace	N12 , trace
			N13 , trace

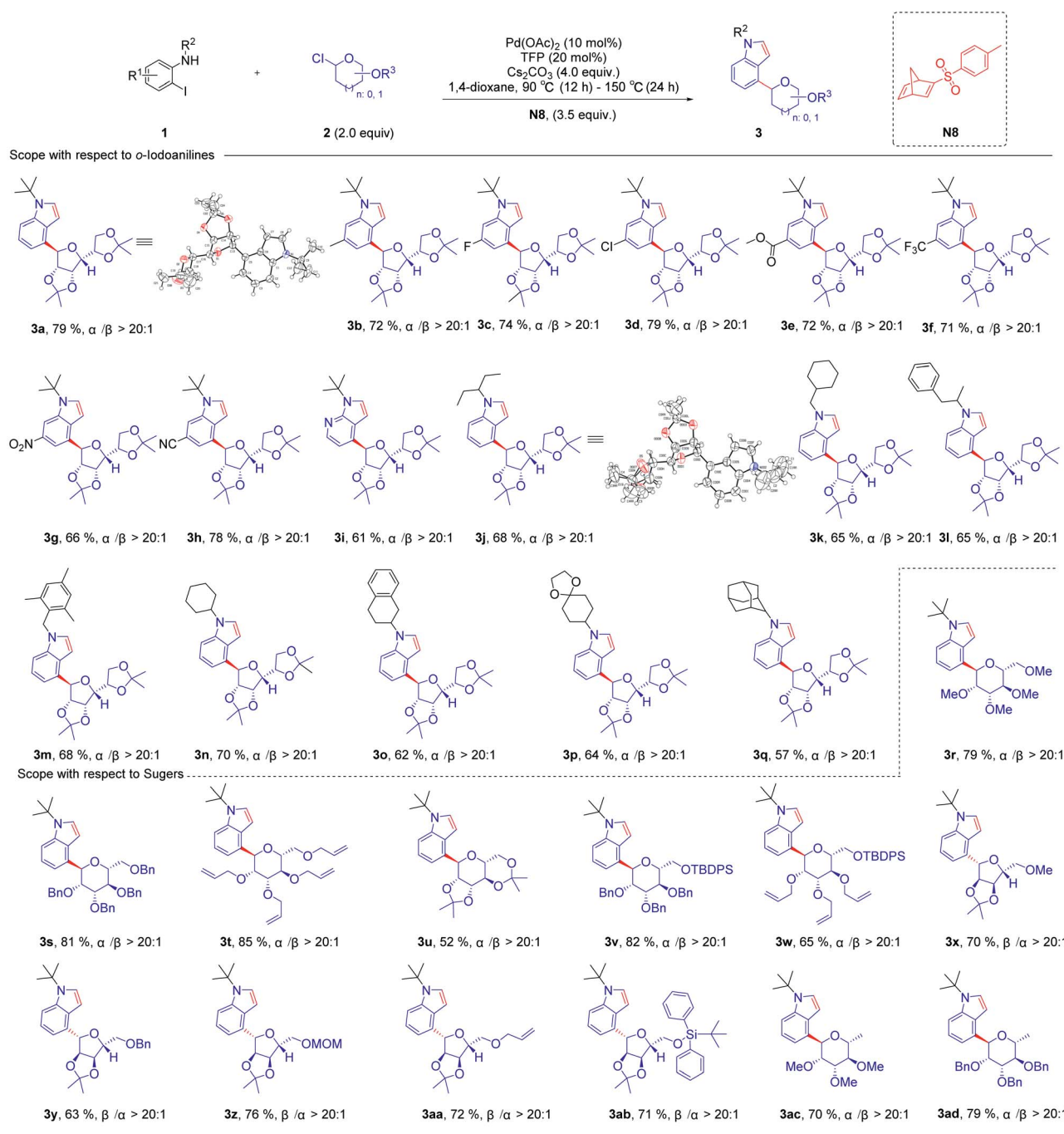
^a Reaction conditions unless otherwise noted: **1a** (0.2 mmol), **2a** (0.2 mmol), **N** (3.5 equiv.), Pd(OAc)₂ (10 mmol%), TFP (20 mmol%), and Cs₂CO₃ (4.0 equiv.) in 1,4-dioxane, 90 °C (12 h)–150 °C (24 h) isolated yields.^b The yields and ratios of the byproduct were observed by GC-MS.



reactivity of this transformation. Then, we tried a series of modified NBDs and found that the intramolecular Buchwald coupling without *ortho*-glycosylation by-product was inhibited. After intensive investigation, the NBD with substitution *para*-toluenesulfonyl groups (**N8**) was the optimal *ortho*-C–H glycosylation coupling partner for the Catellani reaction, and it gave an isolated yield of 79%.

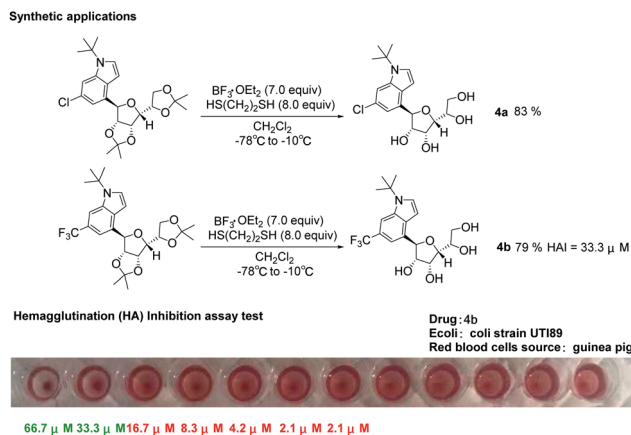
With optimized reaction conditions in hand, we first examined the substrate scope of *o*-iodoanilines. The reactions of *o*-iodoanilines, electron-donating group (–Me), halogen (–F, Cl), and strongly electron-withdrawing group (–NO₂, CN, CO₂Et, CF₃) substituents performed smoothly, affording the desired 4-glycosylindole products (**3b–3h**) in 66–79% yield. It is noteworthy to mention that 3-iodopyridine-2-amine as a heteroaromatic substrate reacted favourably and 4-glycosyl-7-

Table 2 Substrate scope of reaction^a



^a Reaction conditions unless otherwise noted: **1** (0.2 mmol), **2** (0.4 mmol), **N8** (3.5 equiv.), Pd(OAc)₂ (10 mol%), TFP (20 mol%), and Cs₂CO₃ (4.0 equiv.) in 1,4-dioxane, 90 °C (12 h)–150 °C (24 h) isolated yields.

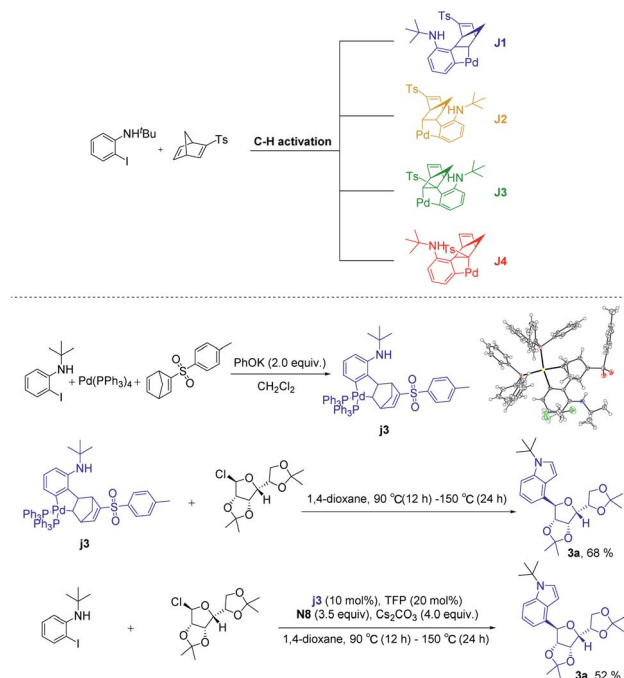




Scheme 3 Synthetic applications and biochemical experiment.

azaindole (**3i**) was obtained in 61% yield. Subsequently, a range of the groups on the nitrogen atom of *o*-iodoaniline was also examined under the optimal reaction conditions. Chain alkane (**3j**), 3-phenylbutane-2-yl (**3l**) containing aromatic hydrocarbons, benzyl (**3m**), cycloalkane (**3n**), tetrahydronaphthalene (**3o**), and adamantane (**3q**) with a large steric hindrance were outstandingly tolerated and afforded the desired products in excellent yields. It can be seen that this method synthesizes a variety of *N*-alkyl substituted indoles, which are difficult to generate by the direct coupling reaction of indole with tertiary or secondary carbon. Inspired by these encouraging results, we then studied a range of glycosyl chlorides. Tetramethyl-, tetrabenzyl-, and tetraallyl-protected α -mannosyl chlorides performed excellently and delivered the corresponding product (**3r–3t**) in 79–85% yield. The corresponding α -*C*-aryl mannoside (**3u**) can be obtained using 2,3,4,6-di-*O*-isopropylidene- α -mannosyl chloride as a substrate in 52% yield. Other mannosyl chlorides with TBDPS are methyl- and benzyl-, such as **3v** and **3w**, affording the desired products in 65–82% yields with exclusive α -selectivity. Delightfully, α -ribofuranosyl chlorides with different functional groups, such as methyl-, benzyl-, methoxymethyl-, allyl- and TBDPS can also deliver the corresponding products in 63–76% yield (**3x–3ab**). In addition, α -rhamnosyl chloride can also be used as a suitable substrate for this reaction, and the corresponding targeted product can be obtained with an excellent yield of 70–79% (Table 2).

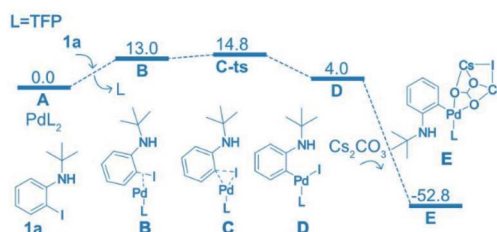
It can be known from the introduction that pharmaceutical compounds and natural products usually have unprotected carbohydrate skeleton structures, so we conducted deprotection experiments on 4-glycosylindole. We tried using ethylene dithioglycol and boron trifluoride ether for deprotection. Finally, we found that by adding boron trifluoride ether as the Lewis acid at -78 °C, ethylene dithioglycol and DCM as the solvent to react at -10 °C, the yield of the deprotected product (**4a–4b**) was 79–83%. Urinary tract infections (UTIs) are mostly caused by Gram-negative uropathogenic *Escherichia coli* (UPEC) bacteria. *C*-Mannosides have good inhibitory activity against the type 1 pilus adhesion, FimH.^{73,74} After obtaining the compounds with a more stable and unprotected structure of 4-



Scheme 4 Structurally modified norbornadiene (smNBDs) has four potential reaction sites.

indole-*C*-glycosides, we tested the bioactivity of compounds **4a** and **4b**. The hemagglutination (HA) inhibition assay test was performed on guinea pig red blood cells using clinical *E. coli* strain UTI89. We found that the **4b** compound has moderate activity (HAI = 33.3 μ M) in the hemagglutination (HA) inhibition test. Here, we disclosed that α -D-mannofuranose provided a possible new structure for the inhibitory activity against FimH compared with traditional α -D-mannopyranose (Scheme 3).

According to the above experimental results, the structure-modified norbornadienes (smNBDs) can effectively regulate the reaction activity of aryl-norbornadiene-palladacycle (ANP), thus successfully realizing the *ortho* C–H glycosylation. Due to the unique structural characteristics of norbornadiene, the retro-Diels–Alder reaction could further be realized to construct the indole skeleton. However, the modified norbornadiene has two olefins and four migration insertion sites. In theory, four isomers may be formed, among which **J1** and **J2** have migratory-insertion on the non-*p*-toluenesulfonyl side olefins, while **J3** and **J4** are migratory-insertions on the side of the *p*-toluene sulfonic acid. Therefore, we first carried out experimental verification.

Fig. 1 Computed free energy surface of the oxidation addition reaction. The relative free energies are presented in kcal mol⁻¹.

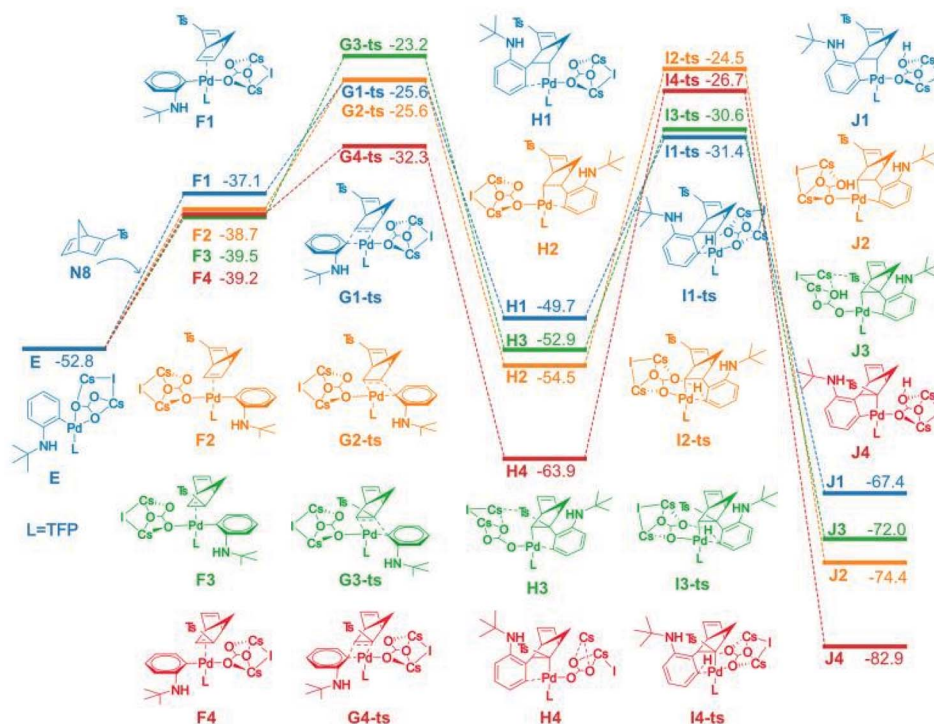


Fig. 2 Computed free energy surface for alkene insertion and C–H activation process. The relative free energies are presented in kcal mol⁻¹.

Under the system of potassium phenolate as the base, we successfully obtained the single crystal of the ANP intermediate (**3j**) and did not find other types of isomers through NMR spectroscopy. Then, we carried out the reaction with an equivalent or catalytic amount of intermediate (**3j**). Finally, the target products were obtained in good yield (Scheme 4). The experimental results directly proved that the migration-insertion of palladium compounds with *p*-toluenesulfonyl substituted norbornadiene was highly chemo- and regioselective, and the mechanism of promoting the activity of the ANP intermediate was inductive effect. In addition, the C2, C3 position unsubstituted C4-glycosidic indoles indirectly proved that the intramolecular Buchwald coupling step occurred on the non *p*-toluenesulfonyl side olefins.

Density functional theory (DFT) was used to study the reaction selectivity. At the initial stage, **1a** attacks the Pd(0) catalyst **A** and one ligand leaves forming the complex **B**, as shown in Fig. 1. This process is roughly endothermic around 13.0 kcal mol⁻¹. Then, oxidative addition takes place through the transition state of **C-ts** to form intermediate **D** with a barrier of 14.8 kcal mol⁻¹ from **A** to **C-ts**. Cs₂CO₃ attacks the Pd center generating complex **E**, as suggested in Liu's work,⁷⁵ releasing around 56.8 kcal mol⁻¹. The C=C of **N8** coordinates with Pd forming four isomers **F1**, **F2**, **F3** and **F4**, as shown in Fig. 2.

The alkene insertion occurred with a barrier of 27.2, 27.1, 29.6, and 20.4 kcal mol⁻¹ through **G1-ts**, **G2-ts**, **G3-ts** and **G4-ts**, respectively, while releasing 12.6, 14.2, 15.0, and 24.7 kcal mol⁻¹ forming **H1**, **H2**, **H3** and **H4**, respectively. The benzene ring coordinates with the Pd for **H1**, **H2**, **H3** and **H4**. The barrier of the reverse process of the alkene insertion was 24.1, 27.3, 31.3, and 31.6 kcal mol⁻¹ from the complex **H** series

through the **G-ts** series. We found that the alkene insertion is a reversible process. C–H activation occurred with barriers of 18.3, 28.4, 23.9, and 37.2 kcal mol⁻¹ for **H1**, **H2**, **H3**, and **H4**, respectively, generating five-membered ring complex **J1**, **J2**, **J3**, and **J4**. The C–H activation barrier for **H1** was 18.3 kcal mol⁻¹, which is the smallest barrier in the C–H activation barriers. The C–H activation occurred from **H1** to **I2-ts** most favorably generating **K1**, agreeing with the experiment. The reverse C–H activation barrier is -36.0, -49.9, -41.4, and 56.2 kcal mol⁻¹ from complex **J** series to complex **H** series.

In conclusion, we directly proved that the migratory insertion of *p*-toluenesulfonyl substituted norbornadiene with the phenylpalladium intermediate was highly chemo- and regioselective manner by X-ray single-crystal diffraction and density functional theory (DFT) calculation. Achieving high chemo- and regioselectivity was crucial for the subsequent C–H glycosylation, and it can be inferred that the regulation of the norbornadiene *p*-toluenesulfonyl group on the reaction was the induction effect. In particular, the chemo- and regioselectivity of *p*-toluenesulfonyl substituted norbornadiene was also very important for the formation of target products, avoiding the formation of *p*-toluenesulfonyl substituted indoles. Of course, the key step of C–H glycosylation, which is the transition state between ANP intermediates and halogenated glycosides, was complex and still under further exploration.

Conclusion

This report describes the palladium-catalyzed C–H glycosylation and retro Diels–Alder tandem reaction *via* structurally modified norbornadiene (smNBDs). A highly functionalized 4-



glycosylindole was synthesized *via* the three-component cross-coupling of *o*-iodoaniline, glycosyl chloride and structurally modified norbornadiene. And the skeleton structure of unprotected glycosyl indole was obtained by the deprotection experiment. In terms of mechanism, the key ANP intermediates characterized by X-ray single-crystal diffraction and further control experiment proved that the migration-insertion reaction of smNBDs with phenyl palladium intermediate has high chemo- and regioselectivity. Finally, density functional theory (DFT) calculations were used to study the main five-membered aryl-norbornadiene-palladacycle (ANP) intermediate formation process.

Data availability

All Data associated with this article are available in the ESI.†

Author contributions

Y. A. and B.-S. Z. performed the methodology, synthesis, characterization, analysis, and wrote the manuscript, and these authors contributed equally. Y.-N. D., Z. Z., X.-Y. G. and X.-S. L. analyzed the data, discussed the results. X. W. performed the biological activity experiment. Y. L. provided carried out DFT calculations. Y.-M. L. designed the project and supervised the whole experiment. All authors read and approved the final manuscript.

Conflicts of interest

The authors declare no conflicts of interest.

Acknowledgements

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References

- 1 F. Berti and R. Adamo, *Chem. Soc. Rev.*, 2018, **47**, 9015–9025.
- 2 R. A. Dwek, *Chem. Rev.*, 1996, **96**, 683–720.
- 3 A. Varki, *Glycobiology*, 2017, **27**, 3–49.
- 4 D. E. Levy and C. Tang, *The chemistry of C-glycosides*, Elsevier, 1995.
- 5 K. Krohn, A. Agoos and C. Bäuerlein, *J. Carbohydr. Chem.*, 2003, **22**, 579–592.
- 6 T. Bililign, B. R. Griffith and J. S. Thorson, *Nat. Prod. Rep.*, 2005, **22**, 742–760.
- 7 J. Štambaský, M. Hocek and P. Kočovský, *Chem. Rev.*, 2009, **109**, 6729–6764.
- 8 L. Nicolas, E. Izquierdo, P. Angibaud, I. Stansfield, L. Meerpoel, S. Reymond and J. Cossy, *J. Org. Chem.*, 2013, **78**, 11807–11814.
- 9 X. Cai, K. Ng, H. Panesar, S.-J. Moon, M. Paredes, K. Ishida, C. Hertweck and T. G. Minehan, *Org. Lett.*, 2014, **16**, 2962–2965.
- 10 N. Taniguchi, T. Endo, W. Hart, G. P. H. Seeberger and C. H. Wong, *Glycoscience: Biology and Medicine Glycoscience: Biology and Medicine*, Springer, 2015.
- 11 E. De Clercq, *J. Med. Chem.*, 2016, **59**, 2301–2311.
- 12 Y. Yang and B. Yu, *Chem. Rev.*, 2017, **117**, 12281–12356.
- 13 K. Kitamura, Y. Ando, T. Matsumoto and K. Suzuki, *Chem. Rev.*, 2018, **118**, 1495–1598.
- 14 Q. Sun, H. Zhang, Q. Wang, T. Qiao, G. He and G. Chen, *Angew. Chem., Int. Ed.*, 2021, **60**, 19620–19625.
- 15 J. Hofsteenge, D. R. Mueller, T. de Beer, A. Loeffler, W. J. Richter and J. F. G. Vliegthart, *Biochemistry*, 1994, **33**, 13524–13530.
- 16 S. Manabe and Y. Ito, *J. Am. Chem. Soc.*, 1999, **121**, 9754–9755.
- 17 T. Nishikawa, M. Ishikawa, K. Wada and M. Isobe, *Synlett*, 2001, 0945–0947.
- 18 C.-H. Yao, J.-S. Song, C.-T. Chen, T.-K. Yeh, T.-C. Hsieh, S.-H. Wu, C.-Y. Huang, Y.-L. Huang, M.-H. Wang, Y.-W. Liu, C.-H. Tsai, C. R. Kumar and J.-C. Lee, *Eur. J. Med. Chem.*, 2012, **55**, 32–38.
- 19 N. Kerru, A. Singh-Pillay, P. Awolade and P. Singh, *Eur. J. Med. Chem.*, 2018, **152**, 436–488.
- 20 S. Yonekubo and N. Fushimi, E. P. Appl. EP1813611A1, August 1, 2007.
- 21 T. Nishikawa, *Synlett*, 1999, 123–125.
- 22 T. Nishikawa, Y. Koide, A. Kanakubo, H. Yoshimura and M. Isobe, *Org. Biomol. Chem.*, 2006, **4**, 1268–1277.
- 23 C. Wiebe, C. Schlemmer, S. Weck and T. Opatz, *Chem. Commun.*, 2011, **47**, 9212–9214.
- 24 C. Wiebe, S. Fusté de la Sotilla and T. Opatz, *Synthesis*, 2012, **44**, 1385–1397.
- 25 A. Yepremyan and T. G. Minehan, *Org. Biomol. Chem.*, 2012, **10**, 5194–5196.
- 26 P. Subramanian and K. P. Kaliappan, *Eur. J. Org. Chem.*, 2013, 595.
- 27 F. Zhang, D. Mu, L. Wang, P. Du, F. Han and Y. Zhao, *J. Org. Chem.*, 2014, **79**, 9490–9499.
- 28 E. Y. Aguilera and M. S. Sanford, *Organometallics*, 2019, **38**, 138–142.
- 29 P. J. Cabrera and M. S. Sanford, *J. Am. Chem. Soc.*, 2018, **140**, 5599–5606.
- 30 M. R. Luzung, C. A. Lewis and P. S. Baran, *Angew. Chem., Int. Ed.*, 2009, **48**, 7025–7029.
- 31 Q. Michaudel, D. Thevenet and P. S. Baran, *J. Am. Chem. Soc.*, 2012, **134**, 2547–2550.
- 32 Q. Wang, Y. Fu, W. Zhu, S. An, Q. Zhou, S.-F. Zhu, G. He, P. Liu and G. Chen, *CCS Chem.*, 2020, **2**, 1729–1736.
- 33 S. Zhang, Y.-H. Niu and X.-S. Ye, *Org. Lett.*, 2017, **19**, 3608–3611.
- 34 Q. Wang, S. An, Z. Deng, W. Zhu, Z. Huang, G. He and G. Chen, *Nat. Catal.*, 2019, **2**, 793–800.
- 35 J. Liu, X. Xiao, P. Han, H. Zhou, Q.-S. Yin and J.-S. Sun, *Org. Biomol. Chem.*, 2020, **18**, 8834.



- 36 Q. Wang, W. Zhu, Q. Sun, G. He and G. Chen, *Chin. J. Chem.*, 2021, **39**, 571–576.
- 37 M. Catellani, F. Frignani and A. Rangoni, *Angew. Chem., Int. Ed.*, 1997, **36**, 119–122.
- 38 M. Lautens and S. Piguel, *Angew. Chem., Int. Ed.*, 2000, **39**, 1045.
- 39 M. Catellani, E. Motti and S. Baratta, *Org. Lett.*, 2001, **3**, 3611–3614.
- 40 N. D. Ca, G. Sassi and M. Catellani, *Adv. Synth. Catal.*, 2008, **350**, 2179–2182.
- 41 M. Catellani, E. Motti and N. Della Ca', *Acc. Chem. Res.*, 2008, **41**, 1512–1522.
- 42 G. Maestri, N. Della Ca' and M. Catellani, *Chem. Commun.*, 2009, 4892–4894.
- 43 E. Motti, N. Della Ca', S. Deledda, E. Fava, F. Panciroli and M. Catellani, *Chem. Commun.*, 2010, **46**, 4291–4293.
- 44 M.-H. Larraufie, G. Maestri, A. Beaume, É. Derat, C. Ollivier, L. Fensterbank, C. Courillon, E. Lacôte and M. Catellani, *Angew. Chem., Int. Ed.*, 2011, **50**, 12253–12256.
- 45 G. Maestri, E. Motti, N. Della Ca', M. Malacria, E. Derat and M. Catellani, *J. Am. Chem. Soc.*, 2011, **133**, 8574–8585.
- 46 N. Della Ca', M. Fontana, E. Motti and M. Catellani, *Acc. Chem. Res.*, 2016, **49**, 1389–1400.
- 47 H.-G. Cheng, S. Chen, R. Chen and Q. Zhou, *Angew. Chem., Int. Ed.*, 2019, **58**, 5832–5844.
- 48 J. Wang and G. Dong, *Chem. Rev.*, 2019, **119**, 7478–7528.
- 49 K. Zhao, L. Ding and Z. Gu, *Synlett*, 2019, **30**, 129–140.
- 50 R. Li and G. Dong, *J. Am. Chem. Soc.*, 2020, **142**, 17859–17875.
- 51 G. Bocelli, M. Catellani and G. P. Chiusoli, *J. Organomet. Chem.*, 1985, **279**, 225–232.
- 52 M. Catellani and G. P. Chiusoli, *J. Organomet. Chem.*, 1985, **286**, 13–16.
- 53 Z. Dong, J. Wang, Z. Ren and G. Dong, *Angew. Chem., Int. Ed.*, 2015, **54**, 12664–12668.
- 54 Z. Wu, N. Fatuzzo and G. Dong, *J. Am. Chem. Soc.*, 2020, **142**, 2715–2720.
- 55 P.-X. Shen, X.-C. Wang, P. Wang, R.-Y. Zhu and J.-Q. Yu, *J. Am. Chem. Soc.*, 2015, **137**, 11574–11577.
- 56 S. Chen, Z.-S. Liu, T. Yang, Y. Hua, Z. Zhou, H.-G. Cheng and Q. Zhou, *Angew. Chem., Int. Ed.*, 2018, **57**, 7161–7165.
- 57 Z.-S. Liu, G. Qian, Q. Gao, P. Wang, H.-G. Cheng, Q. Wei, Q. Liu and Q. Zhou, *ACS Catal.*, 2018, **8**, 4783–4788.
- 58 Q. Gao, Y. Shang, F. Song, J. Ye, Z.-S. Liu, L. Li, H.-G. Cheng and Q. Zhou, *J. Am. Chem. Soc.*, 2019, **141**, 15986–15993.
- 59 Z.-S. Liu, G. Qian, Q. Gao, P. Wang, H.-G. Cheng, Y. Hua and Q. Zhou, *Tetrahedron*, 2019, **75**, 1774–1780.
- 60 P. Wang, S. Chen, Z. Zhou, H.-G. Cheng and Q. Zhou, *Org. Lett.*, 2019, **21**, 3323–3327.
- 61 W. Cai and Z. Gu, *Org. Lett.*, 2019, **21**, 3204–3209.
- 62 J. Liu, Q. Ding, W. Fang, W. Wu, Y. Zhang and Y. Peng, *J. Org. Chem.*, 2018, **83**, 13211–13216.
- 63 W. Lv, Y. Chen, S. Wen, D. Ba and G. Cheng, *J. Am. Chem. Soc.*, 2020, **142**, 14864–14870.
- 64 Y.-N. Ding, W.-Y. Shi, C. Liu, N. Zheng, M. Li, Y. An, Z. Zhang, C.-T. Wang, B.-S. Zhang and Y.-M. Liang, *J. Org. Chem.*, 2020, **85**, 11280–11296.
- 65 R. Li, F. Liu and G. Dong, *Org. Chem. Front.*, 2018, **5**, 3108–3112.
- 66 J. Wang, Z. Dong, C. Yang and G. Dong, *Nat. Chem.*, 2019, **11**, 1106–1112.
- 67 J. Wang, Y. Zhou, X. Xu, P. Liu and G. Dong, *J. Am. Chem. Soc.*, 2020, **142**, 3050–3059.
- 68 R. Li and G. Dong, *Angew. Chem., Int. Ed.*, 2018, **57**, 1697–1701.
- 69 J. Wang, R. Li, Z. Dong, P. Liu and G. Dong, *Nat. Chem.*, 2018, **10**, 866–872.
- 70 Z. Dong and G. Dong, *J. Am. Chem. Soc.*, 2013, **135**, 18350–18353.
- 71 X. Liu, J. Wang and G. Dong, *J. Am. Chem. Soc.*, 2021, **143**, 9991–10004.
- 72 B.-S. Zhang, Y. Li, Z. Zhang, Y. An, Y.-H. Wen, X.-Y. Gou, S.-Q. Quan, X.-G. Wang and Y.-M. Liang, *J. Am. Chem. Soc.*, 2019, **141**, 9731–9738.
- 73 M. Scharenberg, O. Schwardt, S. Rabbani and B. Ernst, *J. Med. Chem.*, 2012, **55**, 9810–9816.
- 74 L. Mydock-McGrane, Z. Cusumano, Z. Han, J. Binkley, M. Kostakioti, T. Hannan, J. S. Pinkner, R. Klein, V. Kalas, J. Crowley, N. P. Rath, S. J. Hultgren and J. W. Janetka, *J. Med. Chem.*, 2016, **59**, 9390–9408.
- 75 B.-W. Li, M.-Y. Wang, S. Fang and J.-Y. Liu, *Organometallics*, 2019, **38**, 2189–2198.

