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# Synthesis of tryptophan-containing 2,5-diketopiperazines *via* sequential C–H activation: total syntheses of tryprostatin A, maremycins A and B†

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Indole 2,5-diketopiperazines (DKPs) are an important type of metabolic cyclic dipeptides containing a tryptophan (Trp) unit possessing a range of interesting biological activities. The intriguing structural features and divergent activities have stimulated tremendous efforts towards their efficient synthesis. Herein, we report the development of a unified strategy for the synthesis of three Trp-containing DKPs, namely tryprostatin A, and maremycins A and B, *via* a sequential C–H activation strategy. The key Trp skeletons were synthesized from the inexpensive, readily available alanine *via* a Pd(II)-catalyzed  $\beta$ -methyl C(sp<sup>3</sup>)–H monoarylation. A subsequent C2-selective prenylation of the resulting 6-OMe-Trp by Pd/norbornene-promoted C–H activation led to the total synthesis of tryprostatin A in 12 linear steps from alanine with 25% overall yield. Meanwhile, total syntheses of maremycins A and B were successfully accomplished using a sequential Pd-catalyzed methylene C(sp<sup>3</sup>)–H methylation as the key step in 15 linear steps from alanine.

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## Introduction

2,5-Diketopiperazines (DKPs), an important type of cyclic dipeptides derived from the condensation of two amino acids, represent a large family of biologically active molecules with a rigid conformation and high stability *in vivo*.<sup>1</sup> Among them, indole DKP alkaloids that contain a tryptophan (Trp) unit are particularly interesting (Fig. 1). The intriguing structural features of these compounds enhance their binding affinity and specificity with biological targets, thus making them suitable for drug discovery with various biological activities such as antitumor, antibacterial, antiviral, antioxidant, and immunomodulatory.<sup>2</sup> Representative members include tryprostatin A (1), maremycin A and B (2), cyclotryprostatin A and B (3), thaxtomina A (4) and norgeamide A and B (5) (Fig. 1).

Tryprostatin A (1) was isolated as a secondary metabolite from a marine fungal strain *Aspergillus fumigatus* BM939.<sup>3a–c</sup> It was reported that tryprostatin A completely inhibited cell cycle progression of tsFT210 cells in the G2/M phase.<sup>3</sup> The total synthesis of tryprostatin A has been achieved by several groups with the emphasis on the asymmetric preparation of the chiral

6-OMe-Trp moiety and a C2-selective prenylation of the indole ring.<sup>4,5</sup> The Cook group first employed the Fischer indole/Schöllkopf protocol to furnish the chiral 6-OMe-Trp unit followed by bromination/lithiation/prenylation to install the C2 prenyl group, leading to the total synthesis in 13 steps with 14.7% overall yield.<sup>4a–d</sup> In 2010, the Fukuyama group elegantly applied radical-mediated indole synthesis/Pd-catalyzed prenylation to accomplish the total synthesis of tryprostatin in 11 linear steps with 30% yield.<sup>4e</sup>

Maremycin A and B (2a and 2b) are indole DKP alkaloids isolated from the culture broth of marine *Streptomyces* strain B 9173, featuring a 3-hydroxyindolin-2-one structure derived from (2*S*,3*S*)- $\beta$ -methyltryptophan ( $\beta$ -MeTrp).<sup>6</sup> The intriguing structure has stimulated tremendous efforts on the total synthesis.<sup>7</sup>

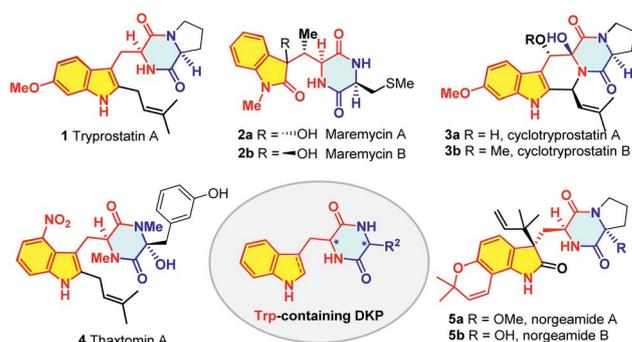


Fig. 1 Representative Trp-containing DKP natural products.

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In 2008, Tamura and co-workers reported the first total synthesis of maremycin A using [3 + 2] cycloaddition as the key step in 9 linear steps with 10.6% yield.<sup>7a,e</sup> In 2011, the Melchiorre group reported the synthesis of the enantioenriched 3-substituted 3-hydroxyoxindoles by organocatalyzed addition of *N*-Me dioxindole to crotonaldehyde and accomplished the total synthesis of maremycin A in 7 linear steps with 15% yield.<sup>7b</sup> Shortly after, Scheidt and co-workers achieved a concise synthesis of maremycin B *via* a key *N*-heterocyclic carbene/Lewis acid-catalyzed formal [3 + 2] addition in 6 linear steps with 17% yield.<sup>7c</sup> Another strategic synthesis of maremycin A and B was accomplished by Jia and co-workers in 2012, featuring a chiral pool strategy starting from *L*-isoleucine followed by a Pd-catalyzed indole synthesis (14 steps, 4.8% yield for maremycin A; 13 steps, 8.2% yield for maremycin B).<sup>7d</sup> The biosynthesis of the key  $\beta$ -MeTrp unit has also been achieved by Lin,<sup>7f,g</sup> Buller and Arnold.<sup>7h</sup>

Although Trp is the least abundant of the 20 proteinogenic amino acids, its unique structure renders it to be easily derivatized both enzymatically and chemically. Thus, Trp has been found to be an important biosynthetic precursor to a variety of microbial natural products.<sup>8</sup> Structurally, Trp-containing DKPs are characterized by a Trp or its derivatives, as exemplified in Fig. 1. For example, tryprostatin A is composed of a proline and an unnatural 2-prenyl-6-OMe-Trp unit; maremycins A and B are composed of an *S*-methylcysteine (*S*-Me-Cys) and an unnatural AA derived from  $\beta$ -MeTrp. Obviously, the concise synthesis of these Trp-containing DKP alkaloids highly relies on the development of a general and straightforward strategy to access various substituted Trps.

In recent years, C–H activation has revolutionized the retrosynthetic logic of natural products.<sup>9</sup> Our group has long been interested in promoting the practical synthesis of natural products *via* C–H activation.<sup>10</sup> Herein, we report the total synthesis of three Trp-containing DKP alkaloids, namely tryprostatin A, and maremycins A and B. The key Trp skeletons were synthesized from the inexpensive, readily available alanine *via* a Pd(*II*)-catalyzed  $\beta$ -methyl C(sp<sup>3</sup>)-H monoarylation.<sup>11,12</sup> A subsequent C2-selective prenylation of the resulting 6-OMe-Trp by Pd/norbornene-promoted C–H activation led to the total

synthesis of tryprostatin A. Meanwhile, total syntheses of maremycins A and B were successfully accomplished using a sequential Pd-catalyzed methylene C(sp<sup>3</sup>)-H methylation as the key step.

## Results and discussion

### Retrosynthetic analysis based on a sequential C–H activation strategy

Our retrosynthetic analysis is depicted in Fig. 2. The DKP moiety could be formed by the condensation of two amino acids [*e.g.*  $\beta$ -MeTrp (**I**) + *S*-Me-Cys (**II**)  $\rightarrow$  maremycins; 2-prenyl-6-OMe-Trp (**III**) + Pro (**IV**)  $\rightarrow$  tryprostatin A]. We reasoned that the syntheses of the two key Trp units,  $\beta$ -MeTrp (**I**) and 2-prenyl-6-OMe-Trp (**III**), were a good platform to verify the brevity and practicality of the sequential C–H activation strategy. Specifically, our recently developed Pd(*II*)-catalyzed  $\beta$ -methyl C(sp<sup>3</sup>)-H monoarylation of an alanine derivative (**6**)<sup>11</sup> followed by methylene C(sp<sup>3</sup>)-H methylation<sup>13</sup> directed by 8-aminoquinoline would enable the facile construction of  $\beta$ -MeTrp (**I**).<sup>14</sup> 2-Prenyl-6-OMe-Trp (**III**) could be assembled by the same  $\beta$ -methyl C(sp<sup>3</sup>)-H monoarylation strategy while the requisite prenyl moiety could be installed by a C2-selective C–H prenylation. The inspiration for this came from previous reports by Bach and co-workers<sup>15</sup> on Pd(*II*)/norbornene-promoted C2-selective C–H alkylation of indole.<sup>16</sup> Different from previous synthetic strategies, this sequential C–H activation logic enables the asymmetric synthesis of indole DKPs from the inexpensive and commercially available *L*-alanine as the chiral pool, avoiding the employment of expensive chiral catalysts/auxiliaries.

### Total synthesis of tryprostatin A *via* a sequential C–H activation strategy

We have previously reported the practical synthesis of  $\beta$ -aryl- $\alpha$ -amino acids through a Pd(*II*)-catalyzed  $\beta$ -methyl C(sp<sup>3</sup>)-H monoarylation of 8-aminoquinoline derived alanine **6** with a broad range of aryl iodides,<sup>11</sup> which has been successfully applied to the total synthesis of tetrahydroisoquinoline alkaloids.<sup>10c,d</sup> The robustness of this protocol encouraged us to examine if 3-iodoindole derivatives were compatible with this

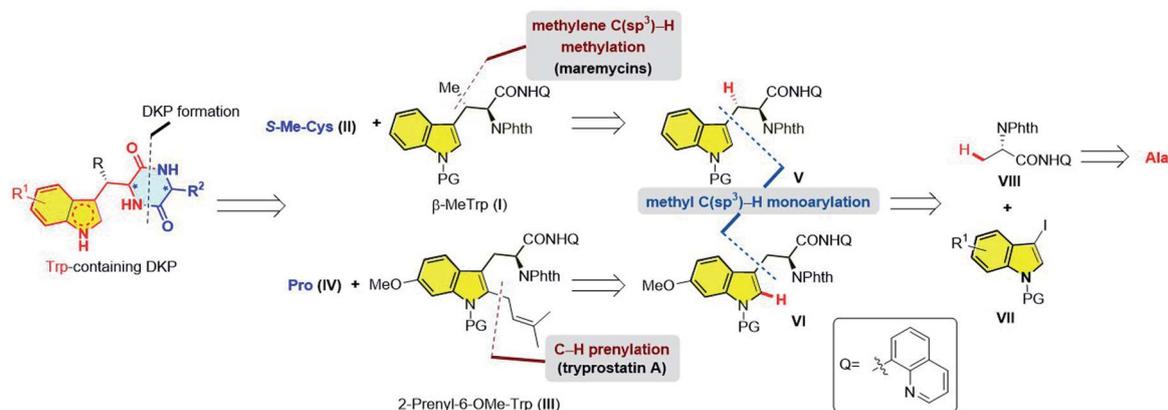
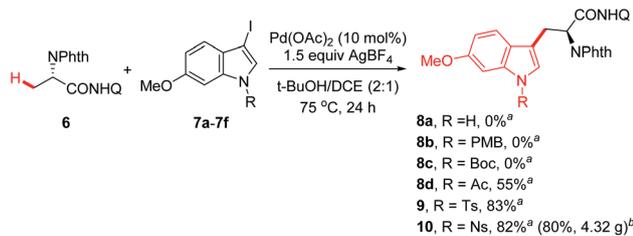


Fig. 2 Retrosynthetic analysis.





Scheme 1 Synthesis of tryptophan derivatives by Pd(II)-catalyzed C(sp<sup>3</sup>)-H monoarylation. <sup>a</sup>0.2 mmol **6**. <sup>b</sup>8 mmol **6**.

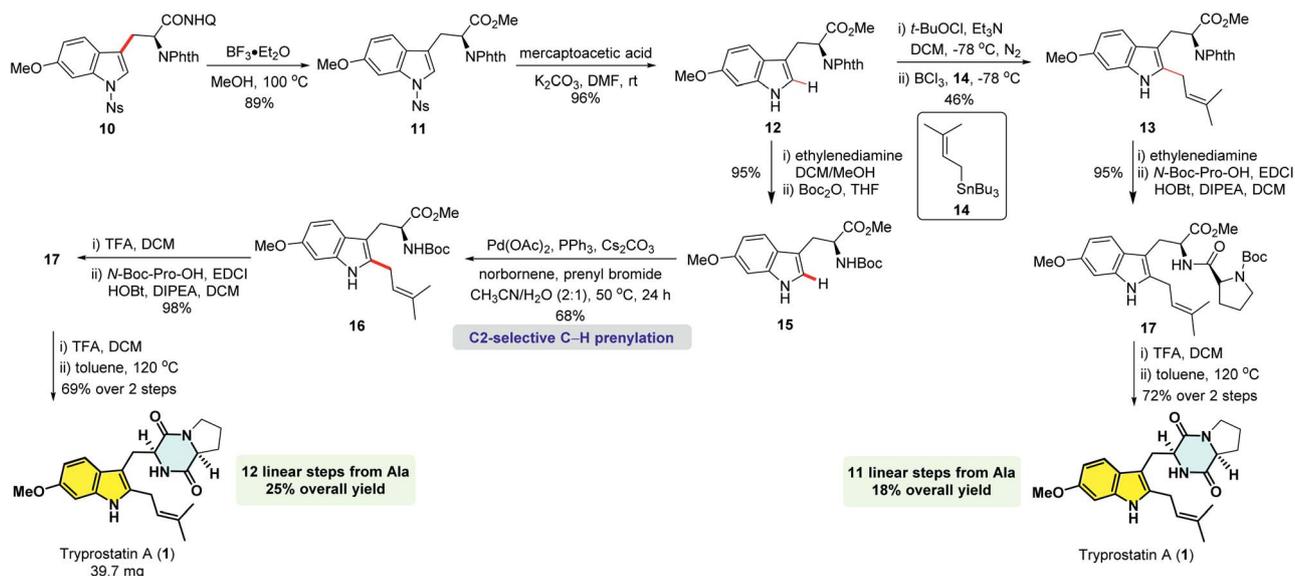
protocol. We commenced the synthesis with the examination of different N-protected 3-iodo-6-methoxyindoles with the monoarylation of alanine derivative **6** (Scheme 1).<sup>11</sup> We were delighted to find that the use of Ac, Ts and Ns as the protecting groups gave the desired 6-OMeTrps in moderate to high yields (**8d**, R = Ac, 55%; **9**, R = Ts, 83%; **10**, R = Ns, 82%), whereas the use of free indole or *N*-Boc and *N*-PMB-protected ones resulted in no detectable products (**8a–8c**). Considering the scalable synthesis (Scheme 1, **10**, 80%, 4.32 g) and ease of removal, *N*-Ns-protected iodoindole **7f** was chosen as the iodination reagent. Of note, the *N*-Phth protecting group is crucial for the high efficiency of C–H arylation as reported in previous studies.<sup>11,12</sup> Removal of the 8-aminoquinoline group of Trp **10** was completed by treating with BF<sub>3</sub>·Et<sub>2</sub>O in methanol (Scheme 2).<sup>12b,17</sup> Trp derivative **11** was obtained in 89% yield along with the recovery of 8-aminoquinoline in 80% yield. Subsequent removal of the *p*-Ns group with mercaptoacetic acid/K<sub>2</sub>CO<sub>3</sub> gave *N*-phthaloyl-6-OMeTrp **12** in 96% yield.<sup>18</sup>

With *N*-phthaloyl-6-OMeTrp **12** in hand, we set out to install the C2-prenyl moiety. The selective introduction of prenylation at the C2 position of the indole ring is a daunting challenge. In the synthesis of tryprostatin B, Danishefsky group elegantly developed a C2-selective nucleophilic prenylation of *in situ*

generated 3-chloroindolenine with prenylstannane and BCl<sub>3</sub> in 83% yield.<sup>19</sup> However, when we tried to apply this method to the C2-prenylation of *N*-phthaloyl-6-OMeTrp **12**, the desired C2-prenylation product **13** was only obtained in 46% yield, largely due to the formation of chlorinated byproducts caused by the electron-donating methoxy group at the C6-position.

In 2011, Bach and co-workers elegantly reported a Pd(II)/norbornene-promoted C2-selective C–H alkylation of indoles<sup>15a,b</sup> and extended it to the regioselective alkylation at the C2 indole position of tryptophan.<sup>15c</sup> Inspired by these pioneering studies, we rationalized that a Pd-catalyzed, norbornene-mediated C2-selective prenylation might be possible by finely tuning the conditions. Firstly, *N*-phthaloyl-6-OMeTrp **12** was converted to *N*-Boc-6-OMeTrp **15** in 95% yield over two steps. Screening experiments were conducted with **15** and prenyl bromide (Table S1, see the ESI† for details). Although the previous conditions only gave trace C2-prenylation product **16**, we found that the prenylation reaction was dramatically affected by bases and solvents, with a stronger base, Cs<sub>2</sub>CO<sub>3</sub>, and a mixture of acetonitrile and water as the solvent, being the optimal conditions. Finally, the prenylation product **16** was obtained in 68% yield when the reaction was conducted under the following optimized conditions: Pd(OAc)<sub>2</sub> (10 mol%), PPh<sub>3</sub> (20 mol%), 3 equiv. prenyl bromide, 5 equiv. norbornene, 3 equiv. Cs<sub>2</sub>CO<sub>3</sub>, CH<sub>3</sub>CN/H<sub>2</sub>O (2 : 1) at 50 °C for 24 hours under N<sub>2</sub>.

Cleavage of *N*-Boc followed by coupling with *N*-Boc-Pro-OH, gave dipeptide **17** in 98% yield. Subsequently, deprotection of *N*-Boc followed by an intramolecular cyclization accomplished the total synthesis of tryprostatin A (**1**). The <sup>1</sup>H and <sup>13</sup>C NMR spectra of our synthetic tryprostatin A (**1**) were identical to that reported by Osada.<sup>3b</sup> The optical rotation values also matched (our synthetic product: [α]<sub>D</sub><sup>20</sup> = –67.5 (c 0.825, CHCl<sub>3</sub>); lit.<sup>3b</sup> [α]<sub>D</sub><sup>27</sup> = –69.7 (c 0.70, CHCl<sub>3</sub>)). The concise total synthesis of tryprostatin A was achieved in 12 linear steps from alanine with



Scheme 2 Synthesis of tryprostatin A.



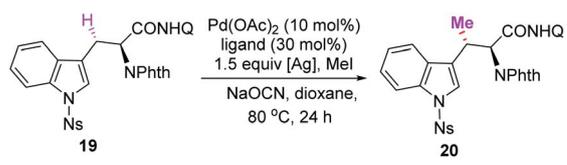
25% overall yield using Pd(II)-catalyzed monoarylation of methyl C(sp<sup>3</sup>)-H and Pd/norbornene-promoted C-H prenylation as key steps.

Removal of the *N*-phthaloyl group from **13** with ethylenediamine, followed by coupling with *N*-Boc-Pro-OH also furnished dipeptide **17** in 95% yield. Thus, an alternative total synthesis of tryprostatin A could also be achieved in 11 linear steps from alanine with 18% overall yield using Danishefsky's C2-selective nucleophilic prenylation as a key step.

### Total syntheses of maremycins A and B via a sequential C(sp<sup>3</sup>)-H activation strategy

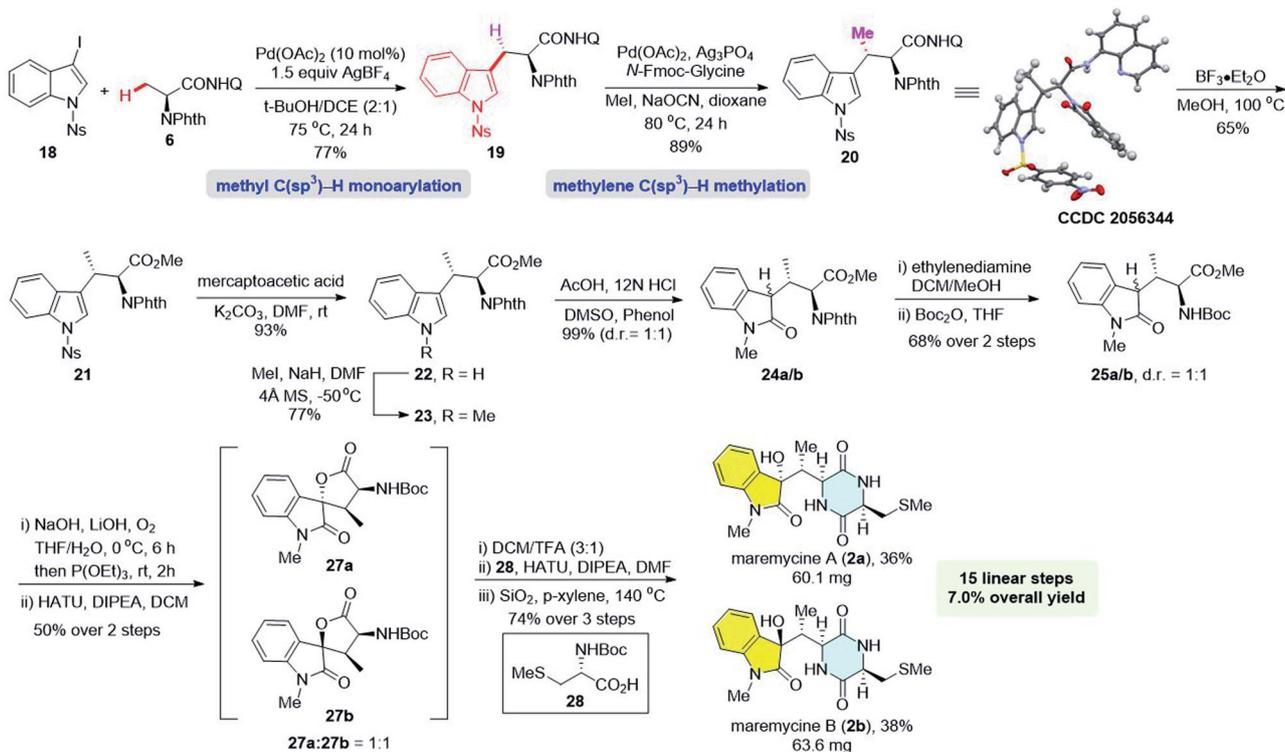
Encouraged by these results, the total syntheses of maremycins A and B via sequential C(sp<sup>3</sup>)-H activation was conducted. The major challenge was the efficient synthesis of the β-MeTrp unit (Fig. 2, I). We envisioned that the key unit could be obtained from 8-aminoquinoline-derived alanine **6** via a sequential Pd(II)-catalyzed methyl C(sp<sup>3</sup>)-H monoarylation followed by methylene C(sp<sup>3</sup>)-H methylation. Following our discovery in the synthesis of tryprostatin A, *N*-Ns-3-iodoindole **18** was used as an efficient arylation reagent to react with alanine derivative **6**, giving Trp **19** in 82% yield (Scheme 3, 10.55 g), which was subjected to a subsequent Pd(II)-catalyzed methylene C(sp<sup>3</sup>)-H methylation. However, unsatisfactory yield and stereoselectivity were obtained under previously established conditions for aliphatic amides (Table 1, entry 1, 29% yield, d.r. = 3 : 1).<sup>12a</sup> Screening of ligands was carried out since we have observed that ligands are essential for both the reactivity and diastereoselectivity. Several sulfonamides were tested and moderate

Table 1 Pd(II)-catalyzed methylene C(sp<sup>3</sup>)-H methylation<sup>a</sup>



Entry <sup>a</sup>	Ligand	[Ag]	d.r.	Yield (%)
1	<i>p</i> -ClPhSO <sub>2</sub> NH <sub>2</sub>	Ag <sub>2</sub> CO <sub>3</sub>	3 : 1	29
2	PhSO <sub>2</sub> NH <sub>2</sub>	Ag <sub>2</sub> CO <sub>3</sub>	3 : 1	64
3	<i>p</i> -MePhSO <sub>2</sub> NH <sub>2</sub>	Ag <sub>2</sub> CO <sub>3</sub>	5 : 2	56
4	<i>p</i> -CF <sub>3</sub> PhSO <sub>2</sub> NH <sub>2</sub>	Ag <sub>2</sub> CO <sub>3</sub>	2 : 1	44
5	PivOH	Ag <sub>2</sub> CO <sub>3</sub>	1 : 1	80
6	AdCO <sub>2</sub> H	Ag <sub>2</sub> CO <sub>3</sub>	9 : 7	66
7	<i>N</i> -Boc-glycine	Ag <sub>2</sub> CO <sub>3</sub>	3 : 1	44
8	<i>N</i> -Cbz-glycine	Ag <sub>2</sub> CO <sub>3</sub>	3 : 1	90
9	<i>N</i> -Fmoc-glycine	Ag <sub>2</sub> CO <sub>3</sub>	12 : 1	40
10	<i>N</i> -Fmoc-glycine	AgOAc	12 : 1	52
11	<i>N</i> -Fmoc-glycine	Ag <sub>3</sub> PO <sub>4</sub>	12 : 1	89(84) <sup>b,c</sup>
12	<i>N</i> -Fmoc-glycine	AgOCN	12 : 1	77
13	<i>N</i> -Fmoc-glycine	AgOTf	12 : 1	93(52) <sup>b</sup>
14	<i>N</i> -Fmoc-glycine	AgTFA	12 : 1	62
15	<i>N</i> -Fmoc-glycine	AgOMs	12 : 1	68
16	<i>N</i> -Fmoc-glycine	AgBF <sub>4</sub>	12 : 1	0
17	None	Ag <sub>2</sub> CO <sub>3</sub>	—	Trace

<sup>a</sup> Reaction conditions: **19** (0.15 mmol), Pd(OAc)<sub>2</sub> (10 mol%), ligand (30 mol%), [Ag] (1.5 equiv.), MeI (10.0 equiv.), NaOCN (2.0 equiv.), 1,4-dioxane, 80 °C, 24 h. Isolated yield. <sup>b</sup> 4.0 mmol scale. <sup>c</sup> 1.0 equiv. Ag<sub>3</sub>PO<sub>4</sub>.



Scheme 3 Total syntheses of maremycins A and B.



yields with poor diastereoselectivities were observed (entries 2–4). The use of pivalic acid and 1-adamantanecarboxylic acid could significantly improve the yields, albeit with very low diastereoselectivities (entries 5 and 6).

It has been reported that *N*-monoprotected amino acids could act as efficient ligands to promote C–H activation reactions.<sup>20</sup> Further investigations revealed that the diastereoisomeric ratio could be significantly improved to 12 : 1 when using *N*-Fmoc-glycine as the ligand (entry 9, 40% yield). Various silver salts were then tested (entries 10–16). Although AgOTs was found to be the most efficient additive when the reaction was conducted in 0.15 mmol scale (entry 13, 93% yield), it was unsuitable for the scale-up reaction (4 mmol, 52% yield). Fortunately, Ag<sub>3</sub>PO<sub>4</sub> was found to be the optimal oxidant for the scale-up reaction, giving the same high yields (entry 11, 0.15 mmol scale, 89% yield; 4 mmol, 84% yield, 12 : 1 d.r.). Trace product was observed when the reaction was conducted in the absence of the ligand, indicating the remarkable facilitation of ligands (entry 17). The absolute configuration of methylated product **20** was confirmed unambiguously by X-ray crystallographic analysis (Scheme 3, CCDC 2056344†).<sup>21</sup>

Removal of the 8-aminoquinoline directing group under BF<sub>3</sub>·Et<sub>2</sub>O/MeOH gave methyl ester **21** in 65% yield.<sup>12b,17</sup> After removal of the *N*-Ns group followed by *N*-methylation of indole, Trp **23** was obtained in 72% yield over two steps. The direct oxidation of *N*-methyl-indole **23** to 3-hydroxy-2-indolone under IBX/CeCl<sub>3</sub> turned out to be unsuccessful due to the steric bulkiness of the β-methyl group.<sup>22</sup> Inspired by Jia's work,<sup>7d</sup> a two-step oxidation sequence was adopted. *N*-Methyl-indole **23** was first oxidized by DMSO in an acidic environment to furnish indolone **24a/b** in 99% yield as a 1 : 1 mixture of diastereomers. The *N*-Phth group was then transformed to *N*-Boc to give **25a/b**, which was oxidized with O<sub>2</sub> and subsequently underwent lactonization to give the spiro-lactone **27**.<sup>23</sup>

Finally, deprotection of *N*-Boc in **27**, followed by condensation with **28** using HATU gave dipeptide product **29**, which upon treating with silica gel in boiling *p*-xylene, accomplished the total synthesis of maremycin A (**2a**) in 36% yield and maremycin B (**2b**) in 38% yield over three steps. All the spectroscopic data of our synthetic maremycin A were identical to those previously reported {see Table S2† for NMR data comparison; our synthetic product:  $[\alpha]_{\text{D}}^{20} = -111.3$  (*c* 0.24, MeOH); lit.<sup>6</sup>  $[\alpha]_{\text{D}}^{25} = -120.95$  (*c* 0.21, MeOH); lit.<sup>7d</sup>  $[\alpha]_{\text{D}}^{25} = -115.5$  (*c* 0.30, MeOH)}. The NMR spectroscopic data of maremycin B (**2b**) also matched those reported (see Table S3† for NMR comparison).<sup>6,7d</sup> However, the optical rotation of our synthetic **2b**  $\{[\alpha]_{\text{D}}^{20} = +69.8$  (*c* 0.78, MeOH) $\}$  was identical to that of Jia's synthetic sample  $\{[\alpha]_{\text{D}}^{25} = +78.3$  (*c* 0.28, MeOH) $\}$ <sup>7d</sup> but differed from that of Laatsch's report  $\{[\alpha]_{\text{D}}^{25} = +2.94$  (*c* 0.21, MeOH) $\}$ .<sup>6</sup>

## Conclusions

In summary, we have reported the development of a unified strategy for the synthesis of three Trp-containing DKPs, namely tryprostatin A, and maremycins A and B, via a sequential C–H activation strategy. The key Trp skeletons were synthesized from the inexpensive, readily available alanine via a Pd(II)-catalyzed β-

methyl C(sp<sup>3</sup>)–H monoarylation. A subsequent C2-selective prenylation of the resulting 6-OMe-Trp by Pd/norbornene-promoted C–H activation led to the total synthesis of tryprostatin A in 12 linear steps from alanine with 25% overall yield. Meanwhile, total syntheses of maremycins A and B were successfully accomplished using a sequential Pd-catalyzed methylene C(sp<sup>3</sup>)–H methylation as the key step in 15 linear steps from alanine. These achievements showcase the robustness of C–H activation in natural product total synthesis. Further application of the C–H activation strategy in the synthesis of other indole DKP alkaloids is ongoing in our laboratory.

## Data availability

Data for all compounds in this manuscript are available in the ESI,† which includes experimental details, characterisation and copies of <sup>1</sup>H and <sup>13</sup>C NMR spectra. Crystallographic data for compound **20** has been deposited under CCDC 2056344.

## Author contributions

X.-S. Yin and W.-Y. Qi contributed equally to this work.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

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## Notes and references

- (a) C. J. Dinsmore and D. C. Beshore, *Tetrahedron*, 2002, **58**, 3297–3312; (b) M. B. Martins and I. Carvalho, *Tetrahedron*, 2007, **63**, 9923–9932; (c) A. D. Borthwick, *Chem. Rev.*, 2012, **112**, 3641–3716; (d) Y. Wang, P. Wang, H. Ma and W. Zhu, *Expert Opin. Ther. Pat.*, 2013, **23**, 1415–1433; (e) P. Umberto and P. Silvia, *Targets Heterocycl. Syst.*, 2013, **17**, 215–239; (f) R.-M. Huang, X.-X. Yi, Y. Zhou, X. Su, Y. Peng and C.-H. Gao, *Mar. Drugs*, 2014, **12**, 6213–6235; (g) A. D. Borthwick and J. Liddle, *Med. Res. Rev.*, 2011, **31**, 576–604; (h) I. Bellezza, M. J. Peirce and A. Minelli, *Trends Mol. Med.*, 2014, **20**, 551–558.
- Y.-M. Ma, X.-A. Liang, Y. Kong and B. Jia, *J. Agric. Food Chem.*, 2016, **64**, 6659–6671.
- Isolation and biological activity of tryprostatin A: (a) C.-B. Cui, H. Kakeya, G. Okuda, R. Onose, M. Ubukata, I. Takahashi, K. Inoso and H. Osada, *J. Antibiot.*, 1995, **48**, 1382–1384; (b) C.-B. Cui, H. Kakeya and H. Osada, *J. Antibiot.*, 1996, **49**, 534–540; (c) C.-B. Cui, H. Kakeya, G. Okada, R. Onose and H. Osada, *J. Antibiot.*, 1996, **49**, 527–533; (d) H. Woehlecke, H. Osada, A. Herrmann and



- H. Lage, *Int. J. Cancer*, 2003, **107**, 721–728; (e) H. D. Jain, C.-C. Zhang, S. Zhou, H. Zhou, J. Ma, X.-X. Liu, X.-B. Liao, A. M. Deveau, C. M. Dieckhaus, M. A. Johnson, K. S. Smith, T. L. Macdonald, H. Kakeya, H. Osada and J. M. Cook, *Bioorg. Med. Chem.*, 2008, **16**, 4626–4651.
- 4 Synthesis of tryprostatin A: (a) T. Gan and J. M. Cook, *Tetrahedron Lett.*, 1997, **38**, 1301–1304; (b) T. Gan, R. Liu, P. Yu, S. Zhao and J. M. Cook, *J. Org. Chem.*, 1997, **62**, 9298–9304; (c) S. Zhao, T. Gan, P. Yu and J. M. Cook, *Tetrahedron Lett.*, 1998, **39**, 7009–7012; (d) C. Ma, X. Liu, X. Li, J. Flippen-Anderson, S. Yu and J. M. Cook, *J. Org. Chem.*, 2001, **66**, 4525–4542; (e) T. Yamakawa, E. Ideue, J. Shimokawa and T. Fukuyama, *Angew. Chem., Int. Ed.*, 2010, **49**, 9262–9265; (f) T. Yamakawa, E. Ideue, Y. Iwaki, A. Sato, H. Tokuyama, J. Shimokawa and T. Fukuyama, *Tetrahedron*, 2011, **67**, 6547–6560.
- 5 For examples of synthesis of tryprostatin B: (a) K. M. Depew, S. J. Danishefsky, N. Rosen and L. Sepp-Lorenzino, *J. Am. Chem. Soc.*, 1996, **118**, 12463–12464; (b) J. M. Schkeryantz, J. C. G. Woo, P. Siliphaivanh, K. M. Depew and S. Danishefsky, *J. Am. Chem. Soc.*, 1999, **121**, 11964–11975; (c) B. Wang, L. Chen and K. Kim, *Tetrahedron Lett.*, 2001, **42**, 1463–1466; (d) S. Zhao, K. S. Smith, A. M. Deveau, C. M. Dieckhaus, M. A. Johnson, T. L. Macdonald and J. M. Cook, *J. Med. Chem.*, 2002, **45**, 1559–1562; (e) E. Caballero, C. Avendaño and J. C. Menéndez, *J. Org. Chem.*, 2003, **68**, 6944–6951; (f) M. Huisman, M. Rahaman, S. Asad, S. Oehm, S. Novin, A. L. Rheingold and M. M. Hossain, *Org. Lett.*, 2019, **21**, 134–137.
- 6 W. Balk-Bindseil, E. Helmke, H. Weyland and H. Laatsch, *Liebigs Ann. Chem.*, 1995, **7**, 1291–1294.
- 7 (a) T. Ueda, M. Inada, I. Okamoto, N. Morita and O. Tamura, *Org. Lett.*, 2008, **10**, 2043–2046; (b) G. Bergonzini and P. Melchiorre, *Angew. Chem., Int. Ed.*, 2012, **51**, 971–974; (c) J. Dugal-Tessier, E. A. O'Bryan, T. B. H. Schroeder, D. T. Cohen and K. A. Scheidt, *Angew. Chem., Int. Ed.*, 2012, **51**, 4963–4967; (d) Y. H. Liu, L. R. Zhang and Y. Jia, *Tetrahedron Lett.*, 2012, **53**, 684–687; (e) T. Ueda, M. Inada, N. Morita and O. Tamura, *Heterocycles*, 2015, **90**, 1179–1195; (f) Y. Lan, Y. Zou, T. Huang, X. Wang, N. L. Brock, Z. Deng and S. Lin, *Sci. China: Chem.*, 2016, **59**, 1224–1228; (g) Y. Zou, Q. Fang, H. Yin, Z. Liang, D. Kong, L. Bai, Z. Deng and S. Lin, *Angew. Chem., Int. Ed.*, 2013, **52**, 12951–12955; (h) M. Herger, P. van Roye, D. K. Romney, S. Brinkmann-Chen, A. R. Buller and F. H. Arnold, *J. Am. Chem. Soc.*, 2016, **138**, 8388–8391.
- 8 (a) T. Huang, X. Wang, W. Guo and S. Lin, Tryptophan-Derived Microbial Alkaloids, in *Comprehensive Natural Products III: Chemistry and Biology*, ed. H.-W. Liu and T. P. Begley, 2020, vol. 2, pp. 393–445; (b) L. M. Alkhalaf and K. S. Ryan, *Chem. Biol.*, 2015, **22**, 317–328; (c) C. M. Santiveri and M. A. Jiménez, *Biopolymers*, 2010, **94**, 779–790.
- 9 (a) W. R. Gutekunst and P. S. Baran, *Chem. Soc. Rev.*, 2011, **40**, 1976–1991; (b) L. McMurray, F. O'Hara and M. J. Gaunt, *Chem. Soc. Rev.*, 2011, **40**, 1885–1898; (c) J. Yamaguchi, A. D. Yamaguchi and K. Itami, *Angew. Chem., Int. Ed.*, 2012, **51**, 8960–9009; (d) D. J. Abrams, P. A. Provencher and E. J. Sorensen, *Chem. Soc. Rev.*, 2018, **47**, 8925–8967; (e) K. Chen and X. Lei, *Current Opinion in Green and Sustainable Chemistry*, 2018, **11**, 9–14; (f) O. Baudoin, *Angew. Chem., Int. Ed.*, 2020, **59**, 17798–17809; (g) N. Y. S. Lam, K. Wu and J.-Q. Yu, *Angew. Chem., Int. Ed.*, 2021, **60**, 15767–15790.
- 10 (a) G. Liao, Q.-J. Yao, Z.-Z. Zhang, Y.-J. Wu, D.-Y. Huang and B.-F. Shi, *Angew. Chem., Int. Ed.*, 2018, **57**, 3661–3665; (b) J. Fan, Q.-J. Yao, Y.-H. Liu, G. Liao, S. Zhang and B.-F. Shi, *Org. Lett.*, 2019, **21**, 3352–3356; (c) S.-L. Fang, M.-X. Jiang, S. Zhang, Y.-J. Wu and B.-F. Shi, *Org. Lett.*, 2019, **21**, 4609–4613; (d) W.-Y. Qi, S.-L. Fang, X. Xu, K. Zhang and B.-F. Shi, *Org. Chem. Front.*, 2021, **8**, 1802–1807.
- 11 K. Chen, S.-Q. Zhang, J.-W. Xu and B.-F. Shi, *Chem. Commun.*, 2014, **50**, 13924–13927.
- 12 For selected reports on Pd(II)-catalysed AQ-directed C-H arylation of  $\alpha$ -amino acids, see: (a) B. V. S. Reddy, L. R. Reddy and E. J. Corey, *Org. Lett.*, 2006, **8**, 3391–3394; (b) L. D. Tran and O. Daugulis, *Angew. Chem., Int. Ed.*, 2012, **51**, 5188; (c) B. Wang, W. A. Nack, G. He, S.-Y. Zhang and G. Chen, *Chem. Sci.*, 2014, **5**, 3952. For reviews on modification of  $\alpha$ -amino acids and peptides via C-H activation, see: (d) A. F. M. Noisier and M. A. Brimble, *Chem. Rev.*, 2014, **114**, 8775–8806; (e) X. Lu, B. Xiao, R. Shang and L. Liu, *Chin. Chem. Lett.*, 2016, **27**, 305–311; (f) W. Wang, M. Lorion, J. Shah, A. R. Kapdi and L. Ackermann, *Angew. Chem., Int. Ed.*, 2018, **57**, 14700–14717; (g) B.-B. Zhan, M.-X. Jiang and B.-F. Shi, *Chem. Commun.*, 2020, **56**, 13950–13958; (h) Z.-B. Bai and H. Wang, *Synlett*, 2020, **31**, 199–204; (i) H. Tong, B. Li, G. Li, G. He and G. Chen, *CCS Chem.*, 2021, **3**, 1797–1820.
- 13 (a) K. Chen and B.-F. Shi, *Angew. Chem., Int. Ed.*, 2014, **53**, 11950–11954; (b) K. Chen, F. Hu, S.-Q. Zhang and B.-F. Shi, *Chem. Sci.*, 2013, **4**, 3906–3911.
- 14 For the pioneering development of 8-aminoquinoline, see: V. G. Zaitsev, D. Shabashov and O. Daugulis, *J. Am. Chem. Soc.*, 2005, **127**, 13154–13155.
- 15 (a) L. Jiao and T. Bach, *J. Am. Chem. Soc.*, 2011, **133**, 12990–12993; (b) L. Jiao, E. Herdtweck and T. Bach, *J. Am. Chem. Soc.*, 2012, **134**, 14563–14572; (c) H. K. Potukuchi and T. Bach, *J. Org. Chem.*, 2013, **78**, 12263–12267.
- 16 (a) L. Cao, Y. Hua, H.-G. Cheng and Q. Zhou, *Org. Chem. Front.*, 2021, **8**, 3883–3914; (b) J. Wang and G. Dong, *Chem. Rev.*, 2019, **119**, 7478–7528; (c) N. Della Ca, M. Fontana, E. Motti and M. Catellani, *Acc. Chem. Res.*, 2016, **49**, 1389–1400; (d) J.-T. Ye and M. Lautens, *Nat. Chem.*, 2015, **7**, 863–870.
- 17 S. Miltsov, L. Rivera, C. Encinas and J. Alonso, *Tetrahedron Lett.*, 2003, **44**, 2301–2303.
- 18 E. Fillion and A. M. Dumas, *J. Org. Chem.*, 2008, **73**, 2920–2923.
- 19 K. Depew, M. Samuel, J. Danishefsky, N. Rosen and L. S. Lorenzino, *J. Am. Chem. Soc.*, 1996, **118**, 12463–12464.
- 20 (a) K. M. Engle, D.-H. Wang and J.-Q. Yu, *J. Am. Chem. Soc.*, 2010, **132**, 14137–14151; (b) K. M. Engle, *Pure Appl. Chem.*, 2016, **88**, 119–138.



- 21 CCDC 2056344 (**20**) contains the ESI† crystallographic data for this paper.
- 22 J. S. Yadav, B. V. Subba Reddy, C. Suresh Reddy and A. D. Krishna, *Tetrahedron Lett.*, 2007, **48**, 2029–2032.
- 23 Compounds **24–27** were all obtained in a 1 : 1 mixture of diastereomers and could be used directly for the total

synthesis. The configuration at C3-position of indolones **24** and **25** was not determined since it would not be retained during the oxidation (**25** → **27**). The configuration of **27** was identified by finishing the total synthesis. See ESI† for details.

