

Cite this: *Chem. Sci.*, 2021, 12, 1392

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 7th October 2020
Accepted 25th November 2020

DOI: 10.1039/d0sc05557c

rsc.li/chemical-science

Discovery of a size-record breaking green-emissive fluorophore: small, smaller, HINA†

Rui Kang,^a Laura Talamini,^b Elisa D'Este,^c Bianca Martins Estevão,^b Luisa De Cola,^{ab} Wim Klopper^{ad} and Frank Biedermann^{ab}

Astonishingly, 3-hydroxyisonicotinaldehyde (HINA) is despite its small size a green-emitting push–pull fluorophore in water (QY of 15%) and shows ratiometric emission response to biological relevant pH differences ($pK_{a2} \sim 7.1$). Moreover, HINA is the first small-molecule fluorophore reported that possesses three distinctly emissive protonation states. This fluorophore can be used in combination with metal complexes for fluorescent-based cysteine detection in aqueous media, and is readily taken up by cells. The theoretical description of HINA's photophysics remains challenging, even when computing Franck–Condon profiles *via* coupled-cluster calculations, making HINA an interesting model for future method development.

Introduction

Fluorescence is both fundamentally fascinating and practically relevant. Countless examples of fluorescent compounds have been described and hundreds of dyes have found commercial use.^{1–4} Decade-long debates were spurred about the origin of the dual fluorescence of DMABN, a small, weakly emissive push–pull system (Scheme 1a).^{5–7} For comparison, the GFP chromophore is, discounting the surrounding protein pocket, one of the smallest green-emitting fluorophores of biological origin.⁸ However, synthetic protein-free GFP analogues suffer from emission quenching by protic solvents.^{8–10} Structurally related polymethine or squarylium push–pull dyes consist of at least two methylene-bridged aromatic moieties.^{11–15} Nitrobenzoxadiazole-derivatives (NBDs) are typical push–pull fluorophores that display good quantum yields (QY) of $\sim 10\%$ in hydrophobic environments but are not emissive in water.^{16–18} Thus, rhodamines, fluoresceins, coumarins, BODIPYs (Scheme 1a) and other polyaromatic dyes are commonly utilized as water-soluble green-emitting dyes.^{11–13,19,20} However, motivated by the need for smaller, non-disturbing fluorescent labels,^{16,21} the search for compact fluorophores that are applicable to

aqueous media continues.^{13,22} Furthermore, luminescent compounds that possess a rich photophysical behaviour also provide valuable test cases for theoretical electron-dynamics method development.^{23–25}

We have stumbled over the unexpected emission of 3-hydroxyisonicotinaldehyde (HINA, see Scheme 1b) in aqueous media. Surprisingly, while the absorbance properties of HINA have been known for decades,^{26–28} its unique fluorescent properties were completely overlooked.²⁹ HINA consists of only 14 atoms (123 g mol^{-1}); for comparison, the similar-sized fluorophores naphthalene and azulene (both 18 atoms, 128 g mol^{-1}) are blue emitting, $\lambda_{\text{em,max}} = 327$ and 373 nm in methanol, respectively.^{30–32} Typical green-emitting dyes are much larger in terms of atom number, molecular weight and size than HINA, see Scheme 1a.

Results

Systematic investigations by absorbance and emission spectroscopy were carried out to elucidate the photophysical behaviour of HINA (Fig. 1). At $3.8 < \text{pH} < 7.0$, HINA occurs in its neutral form and is blue emissive ($\lambda_{\text{em,max}} = 382 \text{ nm}$) with a respectable QY of 7% (Table 1). Upon addition of NaOH, a new absorbance centered at 385 nm arose while the band at 325 nm decreased (Fig. 1a). Simultaneously, the blue emission of neutral HINA vanished and an even stronger green emission ($\lambda_{\text{em,max}} = 525 \text{ nm}$) with a QY of 15% appeared;³³ the corresponding colour change can be seen by the naked eye (Fig. 1b and c). The pK_a value for deprotonation of HINA was obtained by absorbance-based titration, $pK_{a2} = 7.05 \pm 0.01$, and agreed with the value from pH-meter recordings, see Fig. 1d and S10.† Coincidentally, this pK_a value lies perfectly in the biologically relevant range. Interestingly, very similar pK_{a2} values (~ 7) were

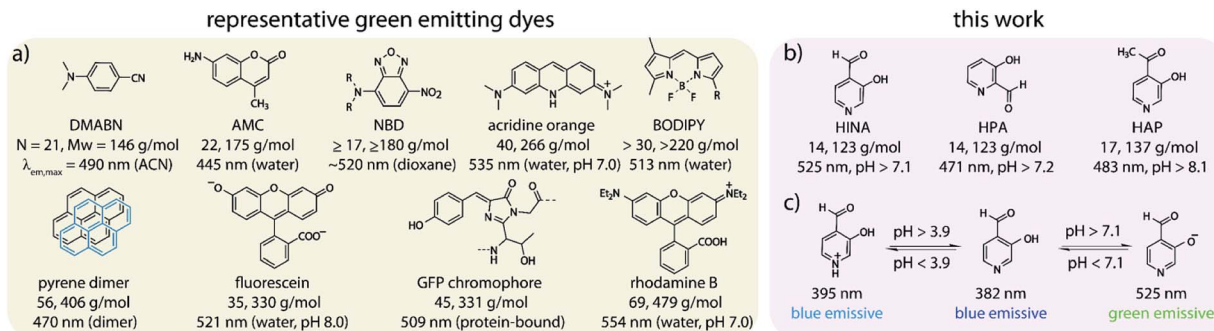
^aInstitute of Nanotechnology (INT), Karlsruhe Institute of Technology (KIT), Hermann-von-Helmholtz Platz 1, 76344 Eggenstein-Leopoldshafen, Germany. E-mail: frank.biedermann@kit.edu; willem.klopper@kit.edu

^bInstitut de Science et d'Ingénierie Supramoléculaires (ISIS), Université de Strasbourg, CNRS, 8Rue Gaspard Monge, 67083 Strasbourg, France. E-mail: decola@unistra.fr

^cOptical Microscopy Facility, Max Planck Institute for Medical Research, Jahnstraße 29, D-69120 Heidelberg, Germany

^dInstitute of Physical Chemistry (IPC), Karlsruhe Institute of Technology (KIT), Fritz-Haber-Weg 6, 76131 Karlsruhe, Germany

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0sc05557c



Scheme 1 Chemical structures, abbreviations, atom numbers (N), molecular weights (M_w) and the peak maximum of emission ($\lambda_{em, max}$) of (a) representative green-emitting dyes, and of (b) herein investigated formyl/acetyl-hydroxy-pyridines. (c) (De)protonation equilibria of HINA in water.

obtained by fluorescence-based titrations regardless of the excitation wavelength, *i.e.* by exciting the neutral form of HINA, the anion or both at an isosbestic point. This indicates that

deprotonation of the neutral form of HINA is not a significant process during the lifetime of its excited state³⁴ despite its largely negative pK_{a2}^* value estimate of -5.5 , which was obtained by a Förster cycle analysis (see the ESI†). Unlike many other hydroxarenes such as phenols and naphthols,^{34,37} HINA does not function as a photoacid. HINA becomes *N*-protonated below pH 3.9, where it remains fluorescent, albeit with a low QY of 0.9%. The emission band of cationic HINA is centred at 395 nm, and thus surprisingly bathochromically shifted compared to neutral HINA (Fig. S1† and Table 1). Conversely, the absorbance spectra showed the expected hypsochromic shift upon addition of acid. The emission lifetimes of the cationic, neutral and anionic forms of HINA identified them as fluorophores (Table 1). Indeed, HINA is a fascinating emissive dye that occurs in three distinct emissive forms. We are unaware of any other small fluorophore with this property.

Strongly solvent-dependent absorbance and emission spectra of HINA were observed. For instance, the characteristic ~ 340 nm absorbance maxima of neutral HINA and a weak blue emission was found in anhydrous acetonitrile (ACN). In contrast, a long-wavelength absorbance band and green emission appeared when water was added to ACN (Fig. S13†). Indeed, the emission of HINA is centred at ~ 400 nm in all anhydrous organic solvents tested (DMSO, MeOH, ethylene glycol, Fig. S14–S18†), but shifts to the 450–600 nm range upon addition of water. Generally, the emission quantum yield of HINA increases upon deprotonation and is higher in polar protic than in polar aprotic solvents, *e.g.* reaching up to 24% in basified methanol (Table S8†).

In aqueous environments, the aldehyde moiety of HINA equilibrates with its hydrate structure, as confirmed by UV/Vis³⁸ and NMR experiments (ESI, Fig. S42–S46†). At first, we wondered if one of the hydrated forms of HINA is the green-emitting species. However, fluorescence in the 450 to 700 nm wavelength region was also detected when adding anhydrous base or anion-stabilising urea to a solution of HINA in organic solvents (see Fig. S16–S18†), confuting hydrate-contributions. Unlike pyrenes (Scheme 1a) or other aggregation-induced emitting (AIE) systems,^{3,24,39} HINA emission is not caused by aggregate formation (Fig. S27 and S28†).

The photophysical properties of 3-hydroxypicolinaldehyde (HPA, an isomer of HINA), of 3-hydroxy-4-acetylpyridine (HAP,

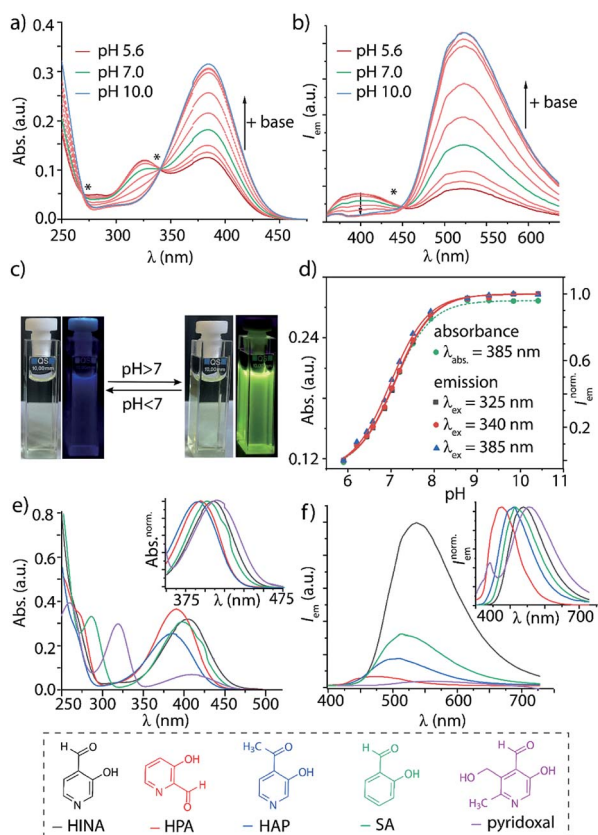


Fig. 1 Titration of an aqueous solution of HINA (50 μ M) with NaOH_{aq} monitored by (a) absorbance and (b) emission spectroscopy ($\lambda_{ex} = 330$ nm). $pK_{a2} = 7.05 \pm 0.01$ and $pK_{a2}^* = 6.98 \pm 0.01$ for absorbance- and emission ($\lambda_{ex} = 385$ nm)-based titration. * Isosbestic or isoemissive point. (c) Photographic images of HINA for its neutral (left) and anionic form (right) under ambient light and UV irradiation. (d) Absorbance ($\lambda_{abs} = 385$ nm) and [0,1]-normalized emission intensity ($\lambda_{ex} = 325, 340$ and 385 nm, $\lambda_{em} = 533$ nm) as a function of pH. Fits shown as dashed green (absorbance) and solid red (emission) lines. Absorbance (e) and emission (f) spectra in water at pH ≥ 9.5 of HINA, HPA, HAP, SA and pyridoxal. Insets show the normalized absorbance and emission spectra at their maximum absorbance and emission wavelength, respectively.

Table 1 Photophysical properties and pK_a values of HINA, HPA and HAP in aerated water at 25 °C

	Cationic form at pH < pK _{a1}			Neutral form at pK _{a1} < pH < pK _{a2}			Anionic form at pH > pK _{a2}			
	λ _{abs} (nm) & [log ε] ^a	λ _{em} ^b (nm) & QY ^c	τ ^d (ns)	λ _{abs} (nm) & [log ε] ^a	λ _{em} ^e (nm) & [QY] ^f	τ ^d (ns)	λ _{abs} (nm) & [log ε] ^a	λ _{em} ^g (nm) & [QY] ^f	τ ^d (ns)	pK _{a1} , pK _{a2} in H ₂ O ^h
HINA	286 [3.71]	395 [0.9%]	0.2	325 [3.32]	382 [7.0%]	0.9	385 [3.74]	525 [15.0%]	1.0	3.9, 7.1
HPA	285 [3.84]	395 [0.4%]	0.2	316 [3.63]	376 [1.2%]	0.4	369 [3.87]	484 [1.4%]	0.2	3.2, 7.2
HAP	318 [3.64]	n.a. ⁱ	—	329 [3.53]	410 [0.1%]	0.2	365 [3.72]	484 [0.8%]	0.9	2.6, 8.1

^a Absorbance peak maximum and logarithmic extinction coefficient. ^b Emission peak maximum, $\lambda_{ex} = 285$ nm. ^c Emission quantum yield, $\lambda_{ex} = 300$ nm. ^d Emission lifetime, $\lambda_{ex} = 373$ nm. ^e Emission peak maximum, $\lambda_{ex} = 330$ nm. ^f Emission quantum yield, $\lambda_{ex} = 370$ nm. ^g Emission peak maximum, $\lambda_{ex} = 380$ nm. ^h From pH titration experiments. Estimated errors ± 0.1 from different methods, see the ESI. ⁱ Very weak emission.

a ketone analogue of HINA), of salicylaldehyde (SA, a phenyl analogue of HINA) and of pyridoxal (a biological hydroxy-formyl-pyridine) were evaluated to obtain insights into the photophysical mechanism, see Fig. 1e, f and Table 1. HPA showed spectroscopic features that were similar to that of HINA, but its absorbance and emission bands for both the neutral and anionic forms are hypsochromically shifted. Moreover, HPA is a much weaker emitter than HINA. The ketone HAP is also poor emissive in its neutral form and as an anion. HAP requires a higher pH for deprotonation than HINA or HPA. Notably, HAP does not form hydrates in water as was confirmed by 1H NMR experiments (Fig. S55[†]), providing additional evidence that these are not the blue and green-emitting species. Like for HINA, absorbance and emission-based titrations of HPA and HAP yielded essentially overlaying pH-property plots (Fig. S6–S9[†]). Again, effective deprotonation of the neutral form of the hydroxy-pyridines apparently does not occur during the lifetime of the excited state despite the strong acidity of the excited state (pK_{a2}^*) of the fluorophores that was estimated by Förster's method (see ESI[†]).

7-Azaindole (7AI, 15 atoms) and SA (15 atoms) are to the best of our knowledge the smallest green-emitting fluorophores that have been reported so far, but they require the use of an unphysiological basic pH of >13.5 and >8.4 , respectively.^{40–42} SA is structurally closely related to HINA (14 atoms) but shows despite its more electron-rich phenyl ring somewhat counter-intuitively a hypochromic shifted absorbance and emission spectra (Fig. 1e and f). Compared to HINA, SA is an inferior fluorophore because its neutral form is not fluorescent and its anionic form is seven times weaker emissive (QY = 2% at pH 10.0). Pyridoxal is both significantly larger in size and much less emissive than the other formyl-hydroxy-pyridines.

The character of the emitting electronic states has been investigated by computing the Franck–Condon profiles through coupled-cluster (CC) calculations with the def2-TZVPPD basis set, see Table S20.[†] For the anionic form of HINA, the predicted absorption and emissions bands, and the Stokes shift are in good agreement with the experimental findings (Fig. S72 and Tables S20–S22[†]). The $n \rightarrow \pi^*$ transition displayed an almost zero oscillator strength, whereas that of the $\pi \rightarrow \pi^*$ transition is four orders of magnitude larger, resulting in computed RGB values (Table S22[†]) that agreed well with the visually observed green emission. Also, the relative emission wavelength maxima

trends between HINA, HPA and SA were correctly predicted (Table S20[†]). Nevertheless, the theoretical description of HINA and its analogues is not simple, despite the small size of these fluorophores. In fact, electronic states which are very close in energy were present and it was found that vibronic effects^{23,24,43} are crucial for the description of the photophysical properties.⁴⁴

Very large discrepancies with the experiments were encountered when computing the emission transitions for the neutral forms of HINA and its analogues. Indeed, an excited-state intramolecular proton-transfer process (ESIPT)^{23,45–47} likely occurs for these substances (Fig. S73[†]). An intramolecular ESIPT may also explain the large discrepancies seen between the expected pH-dependent deprotonation of the excited state based on calculated pK_{a2}^* values, and the “ground-state like” protonation status observed in fluorescence experiments. Thus, explicit theoretical treatment of the electron dynamics is required for computing the photophysical properties of (neutral) HINA, which is beyond the scope of this work. HINA and its analogues represent challenging test cases for advanced theoretical studies and method developments of the nuclear and electronic dynamics, because they combine fascinating photophysical properties with advantages for carrying out the computations (small number of atoms, electrons, and conformers).

The large observed Stokes shifts, *e.g.* 6700, 4600 and 6900 cm^{-1} for protonated, neutral and deprotonated HINA characterize it as a push–pull dye, *e.g.* ~ 5000 cm^{-1} for NBDs and ~ 8400 cm^{-1} for recently reported benzoxazole-thiophenes⁴⁸ and stilbenes.¹⁷ This raises the question if more compact but emissive push–pull fluorophores are even conceivable because single aryl ring-based HINA, HPA and SA already feature some of the smallest but most powerful donor and acceptor moieties, *i.e.* $-O^-$ with a donor resonance const. R^+ of -2.04 , and $-CHO$ with an acceptor resonance const. R^- of 0.70 .⁴⁹ Note that similarly-sized cyanophenols ($R^- = 0.49$ for $-CN$) are not emissive.

Unlike most other push–pull fluorophores, HINA shows a good emission quantum yield in water, which makes it attractive for applications. For instance, it may be used as ratiometric, fluorescent pH indicator dye because its protonated, neutral and deprotonated forms display characteristic emission profiles. Complementary to many other green emitting dyes that interact with hydrophobic species, HINA's high



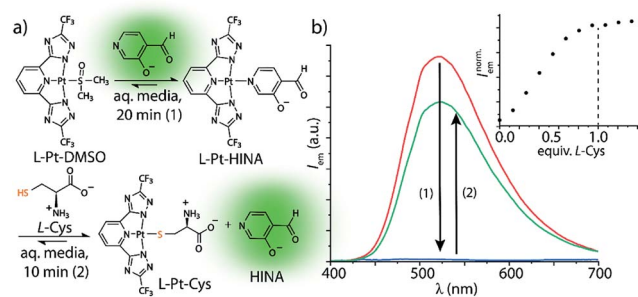


Fig. 2 (a) HINA can be reacted with a Pt(II)-precursor to form a non-emissive L-Pt-HINA complex with a binding constant of $K_a \sim 10^6 \text{ M}^{-1}$, see ESI.† Upon addition of L-Cys, the HINA ligand is displaced and becomes emissive. (b) Emission spectra of HINA (50 μM , red line) upon addition of L-Pt-DMSO (50 μM , blue line), and subsequent addition of L-Cys (50 μM , green line). The inset shows a titration of L-Pt-HINA (50 μM) with L-Cys, 0 to 73 μM ($\lambda_{\text{em}} = 533 \text{ nm}$). All the spectra were obtained in 25 mM NaHCO_3 buffer (pH 9.5) containing 0.9 mM CTAB.

hydrophilicity diminishes its binding to β -cyclodextrin and cucurbit[7]uril as macrocyclic hosts, to serum albumin as a carrier protein, and to polymethacrylate- and polystyrene surfaces.

HINA can also function as a fluorescent indicator in supramolecular sensing assays: we observed that HINA coordinates to Pt- and Pd-complexes, resulting in a full fluorescence switch off (see Fig. 2). From those complexes, the HINA ligand can be readily displaced by a stronger ligand, *e.g.* thiols, yielding in an emission switch-on response in aqueous and organic media. In fact, micromolar concentrations of L-cysteine (L-Cys) can be rapidly ($\sim 10 \text{ min}$) detected through a fluorescence-based indicator-displacement assay in that way (Fig. 2b). Analogously, HINA can be displaced from a Pd-complex by pyridine (Fig. S65†). We believe that HINA-capped metal-organic building blocks could be reacted on demand with stronger ligands to form metal-organic cages or -frameworks,^{50–52} thereby providing an *in situ* fluorescence response for monitoring reactions in real time. Noteworthy, HINA also forms covalent conjugates with L-Cys in aqueous media (Fig. S66–S68†), *via* the formation of a 1,3-thiazolidine ring, which is reminiscent to other aldehyde moiety-containing cysteine-reactive probes.^{53–55} However, like for most reported CHO-based probes, a large excess of L-Cys and long reaction times are needed, making it less attractive for the sensing applications than the HINA-based indicator displacement assay introduced above.

HINA, HPA and HAP readily permeate through biological membranes^{19,56} as was confirmed for different cancerous and non-cancerous cell lines, see the ESI.† Preliminary results indicate that they localize in the perinuclear region of the cells (Fig. 3a) and the cell toxicity level of the dyes is low (Fig. S69†). Interestingly, a rapid fluorescence photoactivation of HINA-treated cells was observed within seconds during the fluorescence imaging experiments (Fig. 3b and Videos in the ESI†). A similar behaviour was seen in control experiments upon irradiation of HINA solutions with a strong light source, and in the presence of hydrogen peroxide (Fig. S71†).

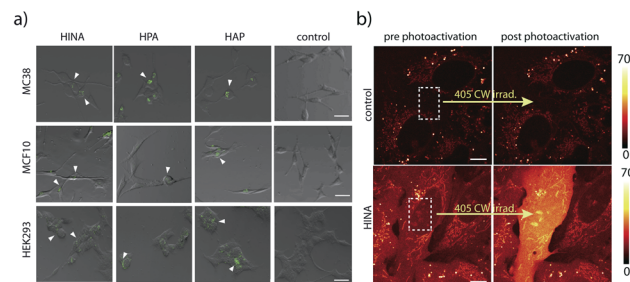


Fig. 3 (a) Microscopy images of the internalization of HINA, HPA, and HAP in MC38, (top row) MCF10 (middle row) and HEK293 (bottom row) cell line after 1 hour of incubation at the concentration of 100 μM . The green signal (white arrowhead) is related to dye molecules and it is mainly localized in the perinuclear region of cells. The control experiment (untreated with dyes) did not show any specific signal (left panel). Scale bar: 20 μm . (b) Confocal images before and after photoactivation of U2OS cells incubated with 2 mM HINA followed by washing (bottom row), and for the control without dye. Dashed boxes indicate the area that was irradiated with a CW 405 light source for three seconds. Colour bars and corresponding minimum and maximum number of counts in displayed in the image are on the right. Scale bars: 10 μm .

Conclusions

In conclusion, 3-hydroxyisonicotinaldehyde (HINA) has been identified as the smallest known green fluorescent dye, and may be reaching the fundamental size-limit. Uniquely, HINA occurs in three different protonation states that are each distinctly fluorescent. The QY of HINA in water is surprisingly high, despite being a push-pull chromophore. HINA's commercial availability, and its favourable photophysical properties (large Stokes shift, pH-dependent ratiometric emission properties that are switching in biorelevant pH 7 regime) will enable future applications, ranging from its use as a fluorescent dye to its function as an indicator in supramolecular assays. The synthesis and investigation of additional hydroxyl-functional pyridine-aldehydes and -ketones will lead to the discovery of novel green-emissive labels with improved photophysical properties. These dyes will like HINA provide excellent test cases to evaluate theoretical predictions of emission spectra using highly advanced computational methods capable of considering both vibronic effects and excited-state intramolecular proton-transfer process (ESIPT).

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

R. K. acknowledges the China Scholarship Council (No. 201806870028). B. M. E. thanks the funding from São Paulo Research Foundation (FAPESP). F. B. thanks the Emmy Noether program of the Deutsche Forschungsgemeinschaft (DFG) for funding. F. B. and W. K. thank the SPP 1807 of the DFG for funding.



Notes and references

- 1 L. D. Lavis, *Annu. Rev. Biochem.*, 2017, **86**, 825–843.
- 2 J. Liu, Y. Q. Sun, H. Zhang, H. Shi, Y. Shi and W. Guo, *ACS Appl. Mater. Interfaces*, 2016, **8**, 22953–22962.
- 3 Y. Hong, J. W. Lam and B. Z. Tang, *Chem. Soc. Rev.*, 2011, **40**, 5361–5388.
- 4 F. Pina, M. J. Melo, C. A. Laia, A. J. Parola and J. C. Lima, *Chem. Soc. Rev.*, 2012, **41**, 869–908.
- 5 A. Demeter, S. Druzhinin, M. George, E. Haselbach, J. L. Roulin and K. A. Zachariasse, *Chem. Phys. Lett.*, 2000, **323**, 351–360.
- 6 M. Sayed, F. Biedermann, V. D. Uzunova, K. I. Assaf, A. C. Bhasikuttan, H. Pal, W. M. Nau and J. Mohanty, *Chem.–Eur. J.*, 2015, **21**, 691–696.
- 7 J. Catalán, *Phys. Chem. Chem. Phys.*, 2013, **15**, 8811–8820.
- 8 J. Dong, K. M. Solntsev and L. M. Tolbert, *J. Am. Chem. Soc.*, 2006, **128**, 12038–12039.
- 9 C. Y. Lin, M. G. Romei, L. M. Oltrogge, I. I. Mathews and S. G. Boxer, *J. Am. Chem. Soc.*, 2019, **141**, 15250–15265.
- 10 H. Deng, C. Yu, D. Yan and X. Zhu, *J. Phys. Chem. B*, 2020, **124**, 871–880.
- 11 A. Loudet and K. Burgess, *Chem. Rev.*, 2007, **107**, 4891–4932.
- 12 X. Li, X. Gao, W. Shi and H. Ma, *Chem. Rev.*, 2014, **114**, 590–659.
- 13 R. N. Dsouza, U. Pischel and W. M. Nau, *Chem. Rev.*, 2011, **111**, 7941–7980.
- 14 A. S. Tatikolov, *J. Photochem. Photobiol., C*, 2012, **13**, 55–90.
- 15 B. Tang, Y. Xing, P. Li, N. Zhang, F. Yu and G. Yang, *J. Am. Chem. Soc.*, 2007, **129**, 11666–11667.
- 16 S. Benson, A. Fernandez, N. D. Barth, F. de Moliner, M. H. Horrocks, C. S. Herrington, J. L. Abad, A. Delgado, L. Kelly, Z. Chang, Y. Feng, M. Nishiura, Y. Hori, K. Kikuchi and M. Vendrell, *Angew. Chem., Int. Ed.*, 2019, **58**, 6911–6915.
- 17 X. Liu, Q. Qiao, W. Tian, W. Liu, J. Chen, M. J. Lang and Z. Xu, *J. Am. Chem. Soc.*, 2016, **138**, 6960–6963.
- 18 S. Saha and A. Samanta, *J. Phys. Chem. A*, 1998, **102**, 7903–7912.
- 19 L. Wang, M. Tran, E. D'Este, J. Roberti, B. Koch, L. Xue and K. Johnsson, *Nat. Chem.*, 2020, **12**, 165–172.
- 20 K. Zamojc, W. Wicz, B. Zaborowski, D. Jacewicz and L. Chmurzynski, *Spectrochim. Acta, Part A*, 2015, **136**, 1875–1880.
- 21 C. P. Toseland, *J. Chem. Biol.*, 2013, **6**, 85–95.
- 22 J. R. Lakowicz, *Principles of fluorescence spectroscopy*, Springer Science & Business Media, 2013.
- 23 S. Mai and L. Gonzalez, *Angew. Chem., Int. Ed.*, 2020, **59**, 16832–16846.
- 24 Y. Tu, J. Liu, H. Zhang, Q. Peng, J. W. Y. Lam and B. Z. Tang, *Angew. Chem., Int. Ed.*, 2019, **58**, 14911–14914.
- 25 J. Chmela, J. F. Greisch, M. E. Harding, W. Kloppe, M. M. Kappes and D. Schooss, *J. Phys. Chem. A*, 2018, **122**, 2461–2467.
- 26 K. Nakamoto and A. Martell, *J. Am. Chem. Soc.*, 1959, **81**, 5863–5869.
- 27 C. M. Harris, R. J. Johnson and D. E. Metzler, *Biochim. Biophys. Acta*, 1976, **421**, 181–194.
- 28 D. Sanz, A. Perona, R. M. Claramunt and J. Elguero, *Tetrahedron*, 2005, **61**, 145–154.
- 29 The purity of the HINA specimen was confirmed by TLC, HPLC and NMR (ESI[†]).
- 30 R. Weinberger and L. J. C. Love, *Spectrochim. Acta, Part A*, 1984, **40**, 49–55.
- 31 T. Yamaguchi, Y. Kimura and N. Hirota, *J. Chem. Phys.*, 2000, **113**, 2772–2783.
- 32 M. Lamberto, C. Pagba, P. Piotrowiak and E. Galoppini, *Tetrahedron Lett.*, 2005, **46**, 4895–4899.
- 33 Isosbestic points at 270 nm and 341 nm and an isosemmissive point at 450 nm ruled out chemical decomposition of HINA.
- 34 L. M. Tolbert and K. M. Solntsev, *Acc. Chem. Res.*, 2002, **35**, 19–27.
- 35 B. Szczepanik, *J. Mol. Struct.*, 2015, **1099**, 209–214.
- 36 J. Zhou, H. Liu, B. Jin, X. Liu, H. Fu and D. Shangguan, *J. Mater. Chem. C*, 2013, **1**, 4427–4436.
- 37 E. L. Wehry and L. B. Rogers, *J. Am. Chem. Soc.*, 1965, **87**, 4234–4238.
- 38 E. G. Sander and W. P. Jencks, *J. Am. Chem. Soc.*, 1968, **90**, 6154–6162.
- 39 N. Meher, S. Panda, S. Kumar and P. K. Iyer, *Chem. Sci.*, 2018, **9**, 3978–3985.
- 40 F. M. F. Santos, Z. Dominguez, J. P. L. Fernandes, C. Parente Carvalho, D. Collado, E. Perez-Inestrosa, M. V. Pinto, A. Fernandes, J. F. Arteaga, U. Pischel and P. M. P. Gois, *Chem.–Eur. J.*, 2020, **26**, 14064–14069.
- 41 J. E. Wampler and J. E. Churchich, *J. Biol. Chem.*, 1969, **244**, 1477–1480.
- 42 Y. S. Wu, H. C. Huang, J. Y. Shen, H. W. Tseng, J. W. Ho, Y. H. Chen and P. T. Chou, *J. Phys. Chem. B*, 2015, **119**, 2302–2309.
- 43 A. Baiardi, J. Bloino and V. Barone, *J. Chem. Theory Comput.*, 2013, **9**, 4097–4115.
- 44 Computed emission wavelength maxima from non-vibronic DFT and CC-based methods largely deviated from experimental values, even after consideration of solvation effects through COSMO.
- 45 J. Zhao, S. Ji, Y. Chen, H. Guo and P. Yang, *Phys. Chem. Chem. Phys.*, 2012, **14**, 8803–8817.
- 46 C. C. Hsieh, C. M. Jiang and P. T. Chou, *Acc. Chem. Res.*, 2010, **43**, 1364–1374.
- 47 N. Basilio, C. A. Laia and F. Pina, *J. Phys. Chem. B*, 2015, **119**, 2749–2757.
- 48 Z. Gao, Y. C. Hao, M. L. Zheng and Y. Chen, *RSC Adv.*, 2017, **7**, 7604–7609.
- 49 C. Hansch, A. Leo and R. W. Taft, *Chem. Rev.*, 1991, **91**, 165–195.
- 50 D. Samanta, J. Gemen, Z. Chu, Y. Diskin-Posner, L. J. W. Shimon and R. Klajn, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, 9379–9384.
- 51 S. Pullen and G. H. Clever, *Acc. Chem. Res.*, 2018, **51**, 3052–3064.
- 52 A. J. McConnell, C. S. Wood, P. P. Neelakandan and J. R. Nitschke, *Chem. Rev.*, 2015, **115**, 7729–7793.



- 53 H. Peng, W. Chen, Y. Cheng, L. Hakuna, R. Strongin and B. Wang, *Sensors*, 2012, **12**, 15907–15946.
- 54 Y. Zhou and J. Yoon, *Chem. Soc. Rev.*, 2012, **41**, 52–67.
- 55 W. Wang, O. Rusin, X. Xu, K. K. Kim, J. O. Escobedo, S. O. Fakayode, K. A. Fletcher, M. Lowry, C. M. Schowalter, C. M. Lawrence, F. R. Fronczek, I. M. Warner and R. M. Strongin, *J. Am. Chem. Soc.*, 2005, **127**, 15949–15958.
- 56 F. Biedermann, G. Ghale, A. Hennig and W. M. Nau, *Commun. Biol.*, 2020, **3**, 383.

