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Photodegradable antimicrobial agents — synthesis, photodegradation, and biological evaluation†

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Multi-drug resistant (MDR) bacteria are already a significant health-care problem and are making the combat of infections quite challenging. Here we report the synthesis of several new compounds containing an ethanolamine moiety, of which two exhibit promising antimicrobial activity (at the 6 μM level). All the compounds are degraded when exposed to light and form inactive products.

Introduction

Alexander Fleming's discovery of penicillin in 1928 [1] started a revolution in the treatment of infections that over the years has saved millions of lives.[2,3] However, the extensive use of penicillin and other antibiotics is about to make them less effective due to the development of multi-drug resistant (MDR) bacteria.[4–6] Such bacteria are already a widespread problem, and several public-health organisations describe the situation as critical and predict catastrophic consequences with an annual death toll in the millions by 2050.[2,7–10] Thus, there is a need for new antibiotics that both combat existing MDR bacteria and prevent formation of new ones.[11]

There are several causes for the MDR-bacteria problems,[2,7,8] but a most important factor is the excessive use, both in human health care and agriculture,[4] of antibiotics, which enter the environment more or less metabolized through sewage and run-off from farmland.[12–15] A proof of the alarming state of affairs is the increasing levels of antibiotic residues in drinking water, wastewater, ground water, coastal waters, and marine organisms.[12,16–20] In order to stop this development, it is necessary to develop antibiotics based on scaffolds with a structural motif that will enable their degradation under ambient conditions so that accumulation in the biosphere does not take place.
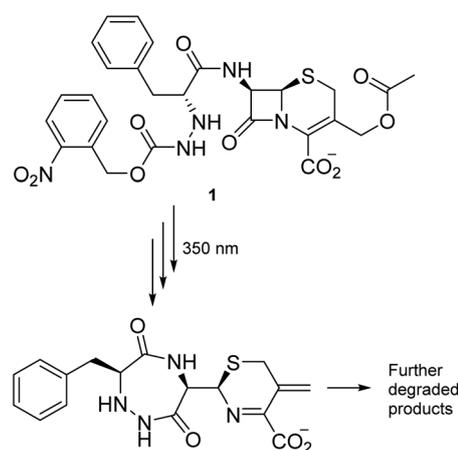
Most antibiotics act inside the body, so the stability of conceivable new agents must necessarily be significant under aqueous and physiological conditions. Our search for degradable moieties was therefore turned to motifs that are known to react with light, another reagent present in abundance in the environment. Light has already been applied repeatedly to turn compounds into derivatives for medical application,[21–23] and irradiation of a carbamate group has successfully been utilized by Lee *et al.* to facilitate the decomposition of the β-lactam moiety embedded in cephalosporanic-acid antibiotic (**1**) (Scheme 1) in a cascade reaction resulting in fragments with no antibiotic activity.[24]

The decomposition illustrated in Scheme 1 appeared to be made possible by the nitrobenzyl group, which has a well-documented reactivity in the excited state.[25] This reactivity has been utilized by Wan and Muralidharan to carry out photo-*retro*-aldol reactions with a variety of 1-(4-nitrobenzyl)-1-alkanols, the result of which was fragmentation and formation of 4-nitrotoluene, 4,4′-dinitrobibenzyl, and the corresponding alkanal in yields that were pH dependent (Scheme 2).[26] We therefore speculated if such alkanols have the potential to function as scaffolds for new photodegradable antibiotics provided substituents that induce biological activity are attached in appropriate positions (Fig. S1†).

Synthesis

The substituents we had in mind were halogenated anilines that were present in some biologically active compounds

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Scheme 1 Light-induced degradation of a cephalosporanic acid analogue.
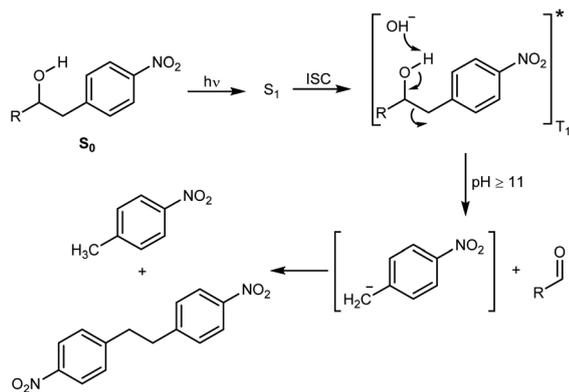
Scheme 2 The photo-*retro*-aldol type reaction of nitrophenethyl alcohols. R = Ph, CH$_3$. Irradiation with a 254, 300, or 350 nm lamp yields the first excited singlet state (S$_1$), which undergoes intersystem crossing (ISC) and formation of triplet-excited (T$_1$) alkanols that suffer fragmentation.

prepared and tested for other purposes (Fig. S2†). In order to make the alkanols as biocompatible as possible, we opted for 2-(arylamino)-1-(nitrobenzyl)ethanol derivatives (**2–5**), which were prepared in quite satisfactory yields by application of standard coupling and oxidation reactions (Scheme 3).

To this end, a Suzuki–Miyaura cross-coupling reaction with allylboronic acid pinacol ester, using a method described by Kotha and co-workers,[27] yielded the allylated product (Scheme 3). Subsequent treatment with *m*CPBA gave the corresponding epoxide and a Lewis acid-promoted epoxide ring-opening reaction using 5 M lithium perchlorate-diethyl ether (LPDE) solution gave aminol **2**. The remaining three compounds were prepared by a selective meta nitration followed by a Stille cross-coupling reaction with allyltributylstannane yielding allylbenzene. Subsequent epoxidation with *m*CPBA gave the required

epoxide and a Lewis acid-promoted epoxide ring opening with anilines **A**, **B**, and **C** gave aminols **3–5**.

Photodecomposition

The photodegradation of compounds **2–5** was studied at pH 8 and 13 following the procedure reported by Wan and Muralidharan[26] using a low-pressure mercury-vapor lamp emitting light mainly at 254 nm (Fig. S3†). The reaction was monitored by proton NMR spectroscopy, which proved that almost complete conversion was achieved after 24 hours, for compounds **2–4** at pH 13 and compounds **3** and **5** at pH 8 (Table S1†). Control experiments show that compounds **2–5** are stable for more than 24 hours in the dark at pH 8 and compound **2** is even stable at pH up to 13 for 24 hours. The changes that took place are illustrated by three spectra associated with photolysis of aminol **2**. Sharp, intense peaks in the spectrum of **2** (A in Fig. 1) are essentially absent in the spectrum of the reaction mixture after irradiation for 24 hours (B in Fig. 1), which on the other hand, contains strong peaks due to aniline **A** (Scheme 3; C in Fig. 1) formed during the degradation of **2** (see also Fig. S4†). MS analysis of the decomposition mixture also disclosed a mass equivalent to *para*-nitrobenzoic acid, which must have derived from the corresponding aldehyde formed under the photodecomposition.

Antibacterial activity

With compounds **2–5** and the corresponding crude degradation mixtures (**2d–5d**) at hand, the minimum inhibitory concentration (MIC) was determined by screening the compounds against several laboratory strains of Gram-positive and Gram-negative bacteria, including *E. faecalis*, *E. coli*, *P. aeruginosa*, *S. aureus*, *S. agalactiae*, and *S. epidermidis* (Tables 1 and S2†). As the results in Table 1 show, compounds **2** and **5** inhibited the growth of *S. agalactiae* at 6.3 μM while compounds **3** and **4**
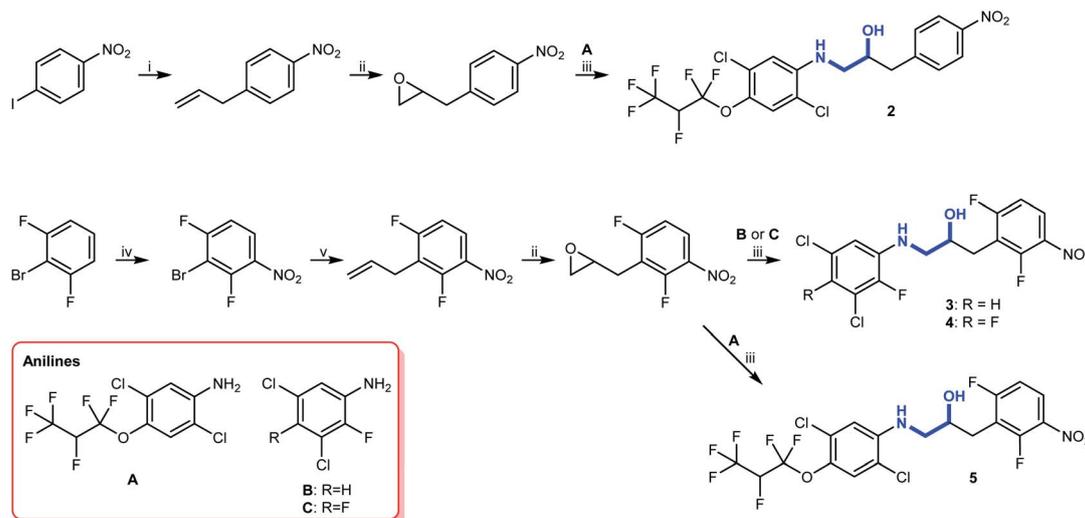
Scheme 3 Synthesis of aminols **2–5**. Reagents and conditions: (i) Pd(PPh$_3$)$_4$, CsF, AllylBpin, THF, reflux;[27] (ii) *m*CPBA, DCM, rt; (iii) 5 M LPDE, 40 °C; (iv) H$_2$SO$_4$, HNO$_3$, 0 °C; (v) Pd(PPh$_3$)$_4$, Bu$_3$SnAllyl, DMF, 110 °C.
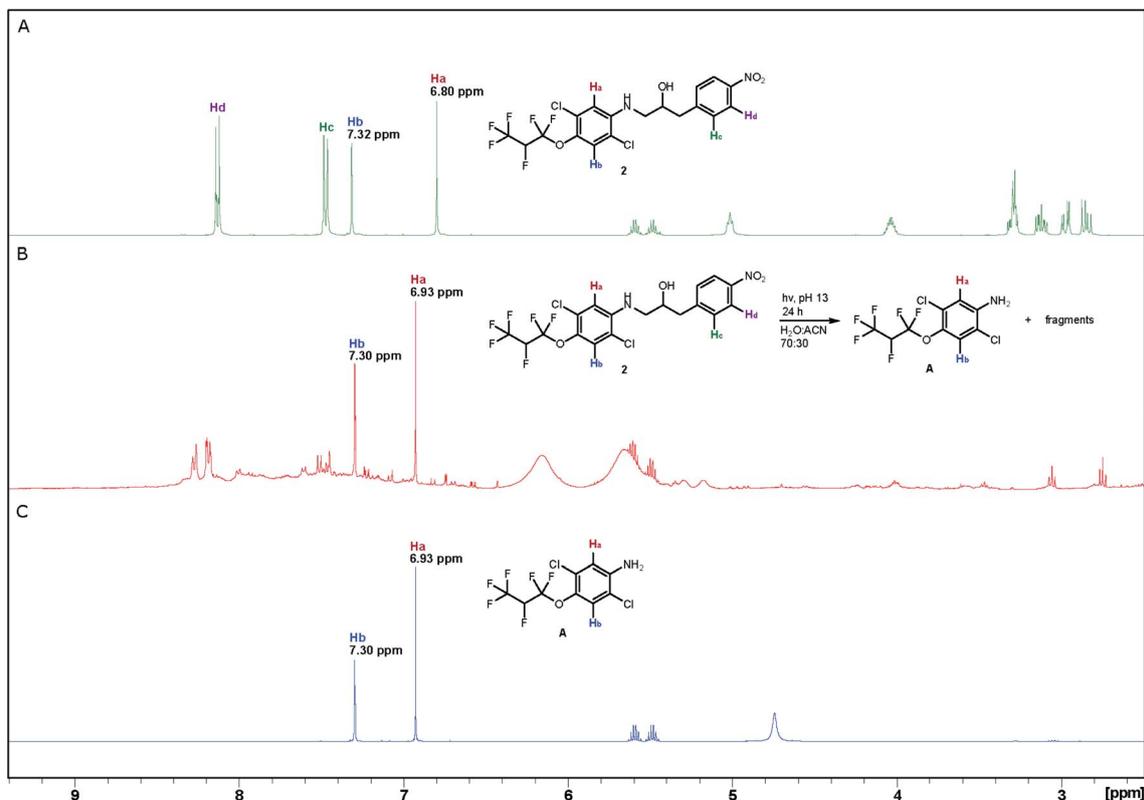
Fig. 1 ^{1}H NMR spectra (A) compound **2** (0.7 mM) before irradiation; (B) reaction mixture after photolysis of compound **2** for 24 hours; (C) decomposition product (A). The NMR analyses were recorded in CD$_3$CN at 400 MHz.

inhibited bacteria growth at 50 μM. The corresponding degradation product mixtures exhibited no antimicrobial activity whatsoever and showed no toxicity towards MRC5 and HepG2 cell lines either (Table 1). However, the two most active antimicrobial agents, *viz.* compounds **2** and **5**, were somewhat toxic (Table 1), but at a much higher concentration then that required to reveal antimicrobial activity.

Table 1 Minimum inhibitory concentration (MIC) in μM for compounds **2–5** against Gram-positive bacteria and toxicity tests against MRC5 and HepG2 cell lines. The activity was screened at the following concentrations: 100, 75, 50, 25, 12.5, 6.3, 3.1, and 1.6 μM[a]

	MIC			Tox	
Compound	*S. epidermidis*	*S. aureus*	*S. agalactiae*	MRC5	HepG2
2	I	I	6.3	50	75
3	50	50	50	50	75
4	I	50	I	50	50
5	I	I	6.3	25	50
2d	I	I	I	I	I
3d	I	I	I	I	I
4d	I	I	I	I	I
5d	I	I	I	I	I

[a] I = inactive at the tested concentrations.

Experimental

General information

IR spectra were recorded on an Agilent Cary 630 FT-IR spectrophotometer equipped with an attenuated total reflectance (ATR) attachment. Samples were analysed neat on a ZnSe crystal and the absorption frequencies are given in wave numbers (cm^{-1}).

UV-vis spectra were obtained on an Agilent 8453 single-beam UV-vis spectrophotometer with a deuterium-discharge lamp for the UV range and a tungsten lamp for the visible wavelength range. Samples were analysed in an Agilent open-top UV quartz cell (10 mm, 3.0 mL) with ethanol as solvent. The wavelengths are reported in nm and molar attenuation coefficients in M^{-1} cm^{-1}.

NMR spectra were recorded on a Bruker Ascend™ 400 spectrometer (400.13 MHz for ^{1}H, 100.61 MHz for ^{13}C, 376.46 MHz for ^{19}F) or a Bruker Ascend™ 850 spectrometer (850.13 MHz for ^{1}H and 213.77 MHz for ^{13}C). Coupling constants (*J*) are given in Hz and the multiplicity is reported as singlet (s), doublet (d), triplet (t), sextet (sxt), multiplet (m), and broad singlet (bs). The chemical shift are reported in ppm in the order downfield to upfield and calibration is done using the residual solvent signals for chloroform-*d* (^{1}H 7.26 ppm; ^{13}C 77.16 ppm) or acetonitrile-*d$_3$* (^{1}H 1.94 ppm; ^{13}C 1.32 ppm).[28] Calibration for ^{19}F NMR is done using α,α,α-trifluorotoluene as internal

standard in chloroform-d (−62.61 ppm) and acetonitrile-d_3 (−63.10 ppm).[29]

High-resolution mass spectra were obtained on a JEOL AccuTOF™ T100GC mass spectrometer. The instrument was operated with an orthogonal electrospray ionization source (ESI), an orthogonal accelerated time of flight (TOF) single stage reflectron mass analyzer and a dual micro channel plate (MCP) detector at the following instrumental settings/conditions; ionization mode: positive, desolvating temperature/ion source temperature = 250 °C, needle voltage = 3000 V, desolvation gas flow = 2.0 L min^{-1}, nebulising gas flow = 1.0 L min^{-1}, orifice1 temperature = 120 °C, orifice1 voltage = 24 V, ring lens voltage = 12 V, orifice2 voltage = 6 V, ion guide peak voltage = 800 V, detector voltage = 2300 V, acquisition range = 4–1000 m/z, spectral recording interval = 0.5 s, wait time = 0.03 ns and data sampling interval = 0.5 ns. Mass calibration were performed using the internal standard method and mass drift compensation was performed in each acquisition.

Thin-layer chromatography (TLC) was carried out with silica gel (60 F$_{254}$) on aluminium sheets with solvent systems consisting of various mixtures of petroleum ether, ethyl acetate, and dichloromethane. Visualization was achieved with either UV light (254 and/or 365 nm) or a potassium permanganate stain. Flash chromatography was performed with a hand pump and 230–400 mesh silica gel or an Interchim Puriflash 215 autoflash chromatography system with Biotage Snap Ultra HP-Sphere™ 25 μm silica-gel columns.

General procedure

Lewis-acid promoted epoxide ring opening. Lithium perchlorate was dried under vacuum for 1 h and dissolved in anhydrous diethyl ether to a 5 M solution. The appropriate aniline (1.0–1.3 equiv.), (∼0.2 M) and epoxide (1.0 equiv.), (∼0.2 M) was added and the reaction mixture was stirred at 40 °C under Ar overnight. DCM and water were added, the phases were separated, and the aqueous layer was extracted with DCM (3 × 10 mL). The combined organic phases were evaporated on Celite and subjected to silica-gel flash chromatography (pet. ether : DCM, 3 : 7), and concentration of the relevant fractions yielded the nitrophenethyl alcohols.

Synthesis of 1-allyl-4-nitrobenzene. An oven-dried 25 mL round-bottom flask fitted with a condenser was charged with 1-iodo-4-nitrobenzene (1.00 g, 4.00 mmol), CsF (1.52 g, 10.0 mmol), Pd(PPh$_3$)$_4$ (0.70 g, 15 mol%), THF (20 mL), and water (2 mL). The mixture was stirred at rt under Ar for 30 min, followed by addition of allylboronic acid pinacol ester (1.36 mL, 7.20 mmol) in THF (8 mL). The reaction mixture was refluxed (oil bath, 95 °C) for 22 h. After cooling to rt the product mixture was evaporated onto Celite and purified by silica-gel flash chromatography (pet. ether). Concentration of the relevant fractions (R_f = 0.47 (pet. ether : EtOAc, 80 : 20 v/v)) yielded 1-allyl-4-nitrobenzene (0.35 g, 52%) as a slightly yellow liquid. The spectroscopic data were in accordance with previously reported data.[30]

IR (neat): ν_{max} 3091, 3017, 2918, 1593, 1502; ^{1}H NMR (400.13 MHz, CDCl$_3$): δ 8.15 (d, J = 8.8 Hz, 2H), 7.34 (d, J = 8.8 Hz, 2H),

5.99–5.89 (m, 1H), 5.18–5.09 (m, 2H), 3.49 (d, J = 6.7 Hz, 2H); ^{13}C NMR (100.61 MHz, CDCl$_3$): δ 147.9, 146.7, 135.6, 129.5, 123.8, 117.5, 40.0.

Synthesis of 2-(4-nitrobenzyl)oxirane. An oven-dried 25 mL round-bottom flask charged with 1-allyl-4-nitrobenzene (0.35 g, 2.16 mmol) in anhydrous DCM (12 mL) was cooled (ice/water bath) and stirred for 5 min under Ar followed by addition of mCPBA (0.59 g, 2.63 mmol). The reaction mixture was stirred at 0 °C for 2 h, then at rt for 15 h. More mCPBA (0.12 g, 0.54 mmol) was added and stirring continued for another 31 h before quenching the reaction with 1 M aq. NaOH solution (10 mL). The phases were separated and the aq. phase was extracted with DCM (3 × 15 mL). The combined organic phases were washed with water (20 mL), brine (20 mL), dried (MgSO4), filtered, and concentrated *in vacuo*. The product was isolated by silica-gel autoflash chromatography (pet. ether : EtOAc : DCM, 93 : 2 : 5 → 40 : 55 : 5) and concentration of the relevant fractions (R_f = 0.26 (pet. ether : DCM, 50 : 50 v/v)) yielded 2-(4-nitrobenzyl) oxirane as a yellow oily liquid (0.20 g, 52%). The spectroscopical data were in full accord with the previously reported data.[31]

IR (neat): ν_{max} 2957, 2923, 2853, 1599, 1516, 1345; ^{1}H NMR (400.13 MHz, CDCl$_3$): δ 8.18 (d, J = 8.5 Hz, 2H), 7.43 (d, J = 8.5 Hz, 2H), 3.20–3.16 (m, 1H), 3.06 (dd, J = 14.8 Hz, 4.2 Hz, 1H), 2.90 (dd, J = 14.8 Hz, 6.4 Hz, 1H), 2.84 (dd, J = 4.7 Hz, 4.0 Hz, 1H), 2.55 (dd, J = 4.7 Hz, 2.6 Hz, 1H); ^{13}C NMR (100.61 MHz, CDCl$_3$): δ 147.1, 145.0, 130.0, 123.9, 51.7, 46.8, 38.6.

Synthesis of 1-((2,5-dichloro-4-(1,1,2,3,3,3-hexafluoropropoxy)phenyl)amino)-3-(4-nitrophenyl)propan-2-ol (2). 2,5-Dichloro-4-(1,1,2,3,3,3-hexafluoropropoxy)aniline (111 mg, 0.34 mmol) and 2-(4-nitrobenzyl)oxirane (62 mg, 0.34 mmol) was reacted according to the general procedure for 19 h to yield aminol **2** (R_f = 0.36 (pet. ether : DCM, 3 : 7)) as a white solid (87 mg, 50%, mp. 121–123 °C) along with 40% recovery of epoxide.

IR (neat): ν_{max} 3507, 3420, 3376, 2980, 2918, 2857, 1599; UV/vis (EtOH): λ_{max} 255 nm (ε 18 339 M^{-1} cm^{-1}); ^{1}H NMR (850.13 MHz, CD$_3$CN): δ 8.14 (d, J = 8.7 Hz, 2H), 7.49 (d, J = 8.7 Hz, 2H), 7.33 (s, 1H), 6.81 (s, 1H), 5.55 (dsxt, J = 42.8 Hz, 5.8 Hz, 1H), 5.03 (t, J = 5.6 Hz, NH), 4.06–4.02 (m, 1H), 3.30 (ddd, J = 13.3 Hz, 6.4 Hz, 3.9 Hz, 1H), 3.27 (d, J = 4.4 Hz, OH), 3.15–3.11 (m, 1H), 2.98 (dd, J = 13.8 Hz, 4.5 Hz, 1H), 2.85 (dd, J = 13.8 Hz, 8.4 Hz, 1H); ^{13}C NMR (213.77 MHz, CD$_3$CN): δ 148.2, 147.7, 145.2, 134.5, 131.5, 127.9, 125.6, 124.3, 121.3 (qd, J = 281 Hz, 25 Hz), 119.1 (td, J = 271 Hz, 23 Hz), 117.7, 112.6, 85.4 (dsxt, J = 198 Hz, 35 Hz), 70.8, 49.7, 41.7; ^{19}F NMR (376.46 MHz, CD$_3$CN): δ −75.6–75.7 (m, 3F), −78.4–80.4 (m, 2F), −213.3 (sxt, J = 12 Hz, 1F); HRMS: (ESI/TOF) m/z: [M + Na]$^+$ calcd for C$_{18}$H$_{14}$Cl$_2$F$_6$N$_2$O$_4$Na$^+$ 529.01270; found 529.01286.

Synthesis of 2-bromo-1,3-difluoro-4-nitrobenzene. 2-Bromo-1,3-difluorobenzene (579 mg, 3.00 mmol) was dissolved in 95–97% sulfuric acid (4 mL) and cooled to 0 °C. To this was added an ice-cold mixture of 95–97% sulfuric acid (4 mL) and 65% nitric acid (5.2 mL) dropwise over 15 min. The reaction mixture was stirred at 0 °C for 1 h, poured over ice, and extracted with DCM (3 × 15 mL). The combined organic layers were washed with sat. aq. NaHCO$_3$ solution (20 mL), dried (MgSO$_4$), filtered, and concentrated under reduced pressure to yield 2-bromo-1,3-

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difluoro-4-nitrobenzene as a white crystalline solid (691 mg, 97%, mp 52–53 °C).

IR (neat): ν_{max} 3099, 1916, 1591, 1530; ^{1}H NMR (400.13 MHz, CDCl$_3$): δ 8.12 (ddd, J = 9.4 Hz, 8.0 Hz, 5.5 Hz, 1H), 7.14 (ddd, J = 9.4 Hz, 7.0 Hz, 2.0 Hz, 1H); ^{13}C NMR (100.61 MHz, CDCl$_3$): δ 163.1 (dd, J = 260 Hz, 3 Hz), 154.3 (dd, J = 267 Hz, 5 Hz), 134.8, 126.2 (dd, J = 10 Hz, 2 Hz), 112.1 (dd, J = 24 Hz, 4 Hz), 101.3 (dd, J = 25 Hz, 24 Hz); ^{19}F NMR (376.46 MHz, CDCl$_3$): δ −92.0 (d, J = 9.5 Hz), −104.4 (d, J = 9.5 Hz); HRMS: (EI/TOF) m/z: [M]$^+$ calcd for C$_6$H$_2$BrF$_2$NO$_2$$^+$ 236.92315; found 236.92306.

Synthesis of 2-allyl-1,3-difluoro-4-nitrobenzene. 2-Bromo-1,3-difluoro-4-nitrobenzene (870 mg, 3.66 mmol), Pd(PPh$_3$)$_4$ (634 mg, 15 mol%), and Bu$_3$SnAllyl (1.53 mL, 4.39 mmol) was dissolved in anhydrous DMF (8 mL). 4 Å molecular sieves were added and the reaction mixture was stirred at 100 °C under Ar for 24 h. The solids were filtered off through cotton with the aid of DCM (50 mL), and the volatiles were removed under reduced pressure. The title compound was isolated by silica-gel flash chromatography (pet. ether : EtOAc, 99 : 1) and concentration of the relevant fractions (R_f = 0.32 (pet. ether : EtOAc, 95 : 5, v/v)) yielded 2-allyl-1,3-difluoro-4-nitrobenzene as a colourless liquid (194 mg, 27%).

IR (neat): ν_{max} 3087, 2957, 2925, 2855, 1623, 1596; ^{1}H NMR (400.13 MHz, CDCl$_3$): δ 8.01 (ddd, J = 9.0 Hz, 8.5 Hz, 5.7 Hz, 1H), 7.04–6.99 (m, 1H), 5.95–5.85 (m, 1H), 5.13–5.08 (m, 2H), 3.51–3.48 (m, 2H); ^{13}C NMR (100.61 MHz, CDCl$_3$): δ 164.2 (dd, J = 258 Hz, 8 Hz), 155.2 (dd, J = 266 Hz, 10 Hz), 134.5–134.4 (m), 132.9, 125.4 (dd, J = 11 Hz, 2 Hz), 118.8 (dd, J = 22 Hz, 19 Hz), 117.4, 111.7 (dd, J = 25 Hz, 4 Hz), 26.8 (t, J = 3 Hz); ^{19}F NMR (376.46 MHz, CDCl$_3$): δ −102.5 (d, J = 14.1 Hz), −116.6 (d, J = 14.1 Hz); HRMS: (EI/TOF) m/z: [M]$^+$ calcd for C$_9$H$_7$F$_2$NO$_2$$^+$ 199.04394; found 199.04399.

Synthesis of 2-(2,6-difluoro-3-nitrobenzyl)oxirane. To a stirred solution of 2-allyl-1,3-difluoro-4-nitrobenzene (194 mg, 0.97 mmol) at 0 °C was added mCPBA (425 mg, 1.94 mmol). The reaction mixture was stirred for 2 h at 0 °C and 5 d at rt, followed by addition of a 1 : 1 sat. aq. NaHCO$_3$: 10% Na$_2$S$_2$O$_3$ solution (30 mL). The phases were separated and the aqueous layer was extracted with DCM (3 × 15 mL). The combined organic phases were washed with a 1 : 1 sat. aq. NaHCO$_3$:10% Na$_2$S$_2$O$_3$ solution (30 mL), sat. aq. NaHCO$_3$ (30 mL), water (30 mL), dried (MgSO$_4$), filtered and concentrated under reduced pressure to yield 2-(2,6-difluoro-3-nitrobenzyl)oxirane as a slightly yellow oily liquid (183 mg, 88%). The product was used without further purification.

IR (neat): ν_{max} 3104, 3000, 2926, 1728, 1624; ^{1}H NMR (400.13 MHz, CDCl$_3$): δ 8.05 (ddd, J = 9.2 Hz, 8.5 Hz, 5.7 Hz, 1H), 7.07–7.02 (m, 1H), 3.22–3.17 (m, 1H), 3.16–3.11 (m, 1H), 3.03–2.97 (m, 1H), 2.81–2.79 (m, 1H), 2.56 (dd, J = 4.8 Hz, 2.5 Hz, 1H); ^{13}C NMR (100.61 MHz, CDCl$_3$): δ 164.5 (dd, J = 259 Hz, 8 Hz), 155.5 (dd, J = 266 Hz, 9 Hz), 134.5, 126.1 (dd, J = 11 Hz, 1 Hz), 115.8 (dd, J = 22 Hz, 20 Hz), 111.8 (dd, J = 25 Hz, 4 Hz), 50.1, 46.9, 25.8 (t, J = 2 Hz); ^{19}F NMR (376.46 MHz, CDCl$_3$): δ −101.5 (d, J = 13.6 Hz), −115.7 (d, J = 13.6 Hz); HRMS; (EI/TOF) m/z: [M−C$_2$H$_2$O]$^+$ calcd for C$_7$H$_4$F$_2$NO$_2$$^+$ 172.02046; found 172.02080.

Synthesis of 1-((3,5-dichloro-2-fluorophenyl)amino)-3-(2,6-difluoro-3-nitrophenyl)propan-2-ol (3). 3,5-Dichloro-2-fluoroaniline (82 mg, 0.46 mmol) and 2-(2,6-difluoro-3-nitrobenzyl)oxirane (77 mg, 0.36 mmol) was reacted according to the general procedure for 20 h to yield aminol **3** (pet. ether:DCM, 3 : 7 (R_f = 0.36) as a sticky colourless oil (60 mg, 42%) along with 7% recovery of epoxide.

IR (neat): ν_{max} 3565, 3425, 3101, 2926, 2857, 1597; UV/vis (EtOH): λ_{max} 254 nm (ε 16 575 M^{-1} cm^{-1}); ^{1}H NMR (400.13 MHz, CD$_3$CN): δ 8.05 (ddd, J = 9.3 Hz, 8.5 Hz, 5.7 Hz, 1H), 7.14 (ddd, J = 9.3 Hz, 8.5 Hz, 1.9 Hz, 1H), 6.73–6.69 (m, 2H), 4.93 (bs, NH), 4.06–3.98 (m, 1H), 3.34 (d, J = 5.3 Hz, OH), 3.32 (ddd, J = 13.5 Hz, 6.7 Hz, 4.0 Hz, 1H), 3.20–3.13 (m, 1H), 2.99–2.87 (m, 2H); ^{13}C NMR (100.61 MHz, CD$_3$CN): δ 165.5 (dd, J = 256 Hz, 8 Hz), 156.2 (dd, J = 263 Hz, 10 Hz), 146.8 (d, J = 239 Hz), 139.9 (d, J = 12 Hz), 135.5, 130.5 (d, J = 4 Hz), 126.7 (dd, J = 12 Hz, 2 Hz), 121.4 (d, J = 16 Hz), 118.6 (dd, J = 22 Hz, 19 Hz), 116.5 (d, J = 1 Hz), 112.6 (dd, J = 25 Hz, 4 Hz), 111.6 (d, J = 4 Hz), 69.3, 49.5, 29.1; ^{19}F NMR (376.46 MHz, CD$_3$CN): δ −103.0 (d, J = 13.6 Hz, 1F), −117.5 (d, J = 13.6 Hz, 1F), −141.7 (s, 1F); HRMS: (ESI/TOF) m/z: [M + H]$^+$ calcd for C$_{15}$H$_{12}$Cl$_2$F$_3$N$_2$O$_3$$^+$ 395.01716; found 395.01724.

Synthesis of 1-((3,5-dichloro-2,4-difluorophenyl)amino)-3-(2,6-difluoro-3-nitrophenyl)propan-2-ol (4). 3,5-Dichloro-2,4-difluoroaniline (65 mg, 0.33 mmol) and 2-(2,6-difluoro-3-nitrobenzyl)-oxirane (71 mg, 0.33 mmol) was reacted according to the general procedure for 22 h to yield aminol **4** (R_f = 0.30 (pet. ether:DCM, 3 : 7)) as a white solid (45 mg, 33%, mp. 110–112 °C) along with 46% recovery of epoxide.

IR (neat): ν_{max} 3433, 3382, 3103, 2935, 2857, 1625; UV/vis (EtOH): λ_{max} 242 nm (ε 16 289 M^{-1} cm^{-1}); ^{1}H NMR (400.13 MHz, CD$_3$CN): δ 8.04 (ddd, J = 9.2 Hz, 8.6 Hz, 5.7 Hz, 1H), 7.13 (ddd, J = 9.2 Hz, 8.5 Hz, 1.8 Hz, 1H), 6.79 (dd, J = 8.6 Hz, 7.1 Hz, 1H), 4.72 (bs, NH), 4.05–3.97 (m, 1H), 3.34 (d, J = 5.3 Hz, OH), 3.92 (ddd, J = 13.4 Hz, 6.6 Hz, 4.1 Hz, 1H), 3.16–3.10 (m, 1H), 2.98–2.87 (m, 2H); ^{13}C NMR (100.61 MHz, CD$_3$CN): δ 165.5 (dd, J = 256 Hz, 8 Hz), 156.2 (dd, J = 263 Hz, 10 Hz), 146.9 (dd, J = 242 Hz, 1 Hz), 146.3 (dd, J = 237 Hz, 2 Hz), 135.7 (dd, J = 12 Hz, 3 Hz), 135.4 (dd, J = 8 Hz, 3 Hz), 126.7 (d, J = 12 Hz), 118.5 (dd, J = 22 Hz, 20 Hz), 117.3 (dd, J = 18 Hz, 4 Hz), 112.6 (dd, J = 25 Hz, 4 Hz), 111.4–111.0 (m, 1C), 111.2 (d, J = 4 Hz), 69.4, 49.7, 29.1; ^{19}F NMR (376.46 MHz, CD$_3$CN): δ −103.0 (d, J = 14.0 Hz, 1F), −117.5 (d, J = 14.0 Hz, 1F), −133.9 (d, J = 4.0 Hz, 1F), −137.0 (d, J = 4.0 Hz, 1F); HRMS: (ESI/TOF) m/z: [M + H]$^+$ calcd for C$_{15}$H$_{11}$Cl$_2$F$_4$N$_2$O$_3$$^+$ 413.00774; found 413.00790.

Synthesis of 1-((2,5-dichloro-4-(1,1,2,3,3,3-hexafluoropropoxy)phenyl)amino)-3-(2,6-difluoro-3-nitrophenyl)propan-2-ol (5). 2,5-Dichloro-4-(1,1,2,3,3,3-hexafluoropropoxy)aniline (151 mg, 0.46 mmol) and 2-(2,6-difluoro-3-nitrobenzyl)-oxirane (100 mg, 0.46 mmol) was reacted according to the general procedure for 18 h to yield aminol **5** (R_f = 0.36 (pet. ether:DCM, 3 : 7)) as a sticky colourless oil (104 mg, 42%) along with 16% recovery of epoxide.

IR (neat): ν_{max} 3550, 3411, 3103, 2937, 1624; UV/vis (EtOH): λ_{max} 254 nm (ε 25 248 M^{-1} cm^{-1}); ^{1}H NMR (850.13 MHz, CD$_3$CN): δ 8.06–8.04 (m, 1H), 7.33 (s, 1H), 7.15–7.13 (m, 1H), 6.86 (s, 1H), 5.55 (dsxt, J = 42.7 Hz, 5.8 Hz, 1H), 5.04 (t, J = 5.7 Hz, NH), 4.07–4.04 (m, 1H), 3.39 (d, J = 5.2 Hz, OH), 3.36 (ddd, J = 13.3 Hz, 6.4 Hz, 4.1 Hz, 1H), 3.22–3.19 (m, 1H), 2.97

(dd, $J = 13.9$ Hz, 4.8 Hz, 1H), 2.93 (dd, $J = 13.9$ Hz, 8.3 Hz, 1H); ^{13}C NMR (213.77 MHz, CD$_3$CN): δ 165.5 (dd, $J = 256$ Hz, 8 Hz), 156.2 (dd, $J = 263$ Hz, 10 Hz), 145.2, 135.5 (d, $J = 5$ Hz), 134.5, 127.9, 126.7 (d, $J = 12$ Hz), 125.6, 121.3 (qd, $J = 281$ Hz, 25 Hz), 119.1 (td, $J = 271$ Hz, 23 Hz), 118.5 (dd, $J = 22$ Hz, 19 Hz), 117.7, 112.73–112.60 (m, 1C), 112.71, 85.4 (dsxt, $J = 198$ Hz, 35 Hz), 69.3, 49.6, 29.2; ^{19}F NMR (376.46 MHz, CD$_3$CN): δ −75.6–75.7 (m, 3F), −78.5–80.4 (m, 2F), −102.9 (d, $J = 14.0$ Hz, 1F), −117.4 (d, $J = 14.0$ Hz, 1F), −213.3 (sxt, $J = 11.2$ Hz, 1F); HRMS: (ESI/TOF) m/z: $[M + H]^+$ calcd for C$_{18}$H$_{13}$Cl$_2$F$_8$N$_2$O$_4^+$ 543.01191; found 543.01196.

Photodegradation reactions at $\sim$0.7 mM for NMR analysis

All reactions were carried out in a 75 mL gas inlet flask ($\sim$0.25–0.35 mM) under a constant stream of nitrogen with a quartz well water-cooled cold finger for 24 hours. The appropriate aminol ($\sim$0.02 mmol) was dissolved in acetonitrile (23 mL), diluted with distilled water adjusted to pH 8 with sodium hydroxide (52 mL), and photolyzed using a 254 nm low-pressure mercury lamp. The aqueous solution was extracted with DCM (3 × 20 mL) and the combined organic layers were dried (MgSO$_4$), filtered, and concentrated to yield a black residue, which was analysed by ^{1}H NMR spectroscopy.

Conclusions

In conclusion, two compounds based on a new structural scaffold, **2** and **5**, possess both antimicrobial activity at the 6 μM level and the ability to decompose upon exposure to light. The decomposition products do not exhibit antimicrobial activity or toxicity. This opens for the development of antibiotics that when released to the environment after treatment will be removed and not contribute to the evolution of MDR bacteria. Both **2** and **5** were not active towards Gram-negative bacteria, but work by Richter *et al.* outlines techniques that can be applied to expand their capability to encompass Gram-negative bacteria as well.[32] Work directed towards this is ongoing in these laboratories.

Author contributions

LKS and MOS had the initial idea; VE, LKS, and MOS developed the idea; VE performed the experimental work; VE drafted the manuscript; VE, LKS, and MOS revised the manuscript.

Conflicts of interest

There are no conflicts to declare.

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