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Succession of the microbial community during the process of mechanical and biological pretreatment coupled with a bio-filter for removal of VOCs derived from domestic waste: a field study

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Changes in the microbial community can not only reflect the efficiency of waste disposal, but also reveal the effect of odor control during the treatment process. This study aimed to evaluate the removal efficiency of volatile organic compounds (VOCs) by the process of mechanical and biological pretreatment (MBP) coupled with a bio-filter (BF). An interesting phenomenon was found that the VOCs were effectively reduced through the MBP process. To understand the removal mechanism of VOCs, the abundance and diversity of microbial bacteria and fungi in the biological dehydration (BD) process, biological fermentation process, and BF process were explored. The abundance and diversity of microbes in the BF were relatively high, of which the bacteria such as *Lactobacillus*, *Bacillus* and *Candida* were the dominant species for VOCs treatment. The proposed technical process and the positive effects observed in this study indicate that it could be applied to the control of VOCs in the treatment of domestic waste.

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1. Introduction

The rapid growth of domestic waste with high moisture content and large fluctuations in composition has brought tremendous pressure and challenges to its treatment. Disposal methods for domestic waste include stacking, landfilling, composting, incineration and so forth.^{1,2} Since landfill will occupy a large amount of land and incineration will cause pollution such as dioxins, composting has become the main technical method for domestic waste treatment in recent years. Numerous studies have shown that domestic waste is separated before composting to ensure the quality of the composting products.^{3–5} Therefore, mechanical biological treatment (MBT), which is composed of mechanical screening and biological fermentation, has become an emerging technological hotspot. For example, single-stream or separated-stream MBT has been applied in Italy for waste treatment.⁶ Also, in the domestic waste plants in Phitsanulok, Thailand, MBT is used as a process to turn household waste into fertilizers or fuels.⁷ However, the high moisture content of domestic waste in China increases the length of the composting time. Simultaneously, domestic waste has also suffered from dehydration, which ultimately slows down the humification rate. Furthermore, if the front-end mechanical screening effect

is not good, it will seriously restrict the back-end composting effect.

To overcome this difficulty, a quality classification and utilization technology (QCU) that adds the mechanical biological pretreatment (MBP) link on MBT has become an improved new technology adopted by Chinese factories in accordance with the characteristics of Chinese waste. MBP is mainly composed of mechanical sorting and biological dehydration (BD). Mechanical sorting is very necessary due to its advantages of being relatively harmless, showing an obvious reduction effect and having low processing costs.⁸ The water content of domestic waste can be reduced by the BD process, making the final humic acid content derived from humified fermentation higher and fertilizer efficiency better. Then, domestic waste pretreated by MBP is further treated by secondary mechanical refinement and humified fermentation to improve its classification and utilization, and ensure the effective control of secondary pollutants such as volatile organic compounds (VOCs).

Various toxic and harmful gases (such as ammonia, hydrogen sulfide, and toluene) and a large amount of VOCs (e.g., dimethyl disulfide, dimethyl sulfide, benzene, 2-butanone, limonene, and methylene chloride) can be generated during domestic waste treatment,^{9,10} causing environmental pollution in and around the domestic waste plant. Removing inorganic odors and VOCs is important in the quality classification and utilization of domestic waste and secondary pollution control.^{11–13} In the research on the removal of VOCs under the

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bio-filter (BF) treatment, there are numerous reports about the removal rate, as well as the influence of process parameters such as inlet, gas flow, microbes and pressure drop.^{14–16} However, in the process of domestic waste treatment, the types and quantities of VOCs produced at different processing stages have a great influence on the treatment effect of the BF, but there are few relevant studies.

The specific microbes in the BF can mineralize VOCs into carbon dioxide and water,^{17–20} or convert target pollutants into intermediate products and final products that are less harmful.²¹ Some reports have examined the microbial action mechanism in the treatment of VOCs by evaluating microbial population dynamics.^{22,23} Zhang *et al.* used a BF to treat the gases produced by domestic sewage and found that overall or functional microorganisms can explain the difference in fine purification performance.²⁴ Zheng *et al.* used a BF to remove ammonia and sulfurated hydrogen from landfills and found that the microenvironment within the packing materials is an important factor in the removal performance of the BF.²⁵ Therefore, the microbial changes in the processing stage and the BF are critical to the evaluation of the removal potential of VOCs.

This study was performed in a domestic waste treatment plant in Shanghai. The types and quantities of VOCs produced in gases from the MBP process were analysed. The succession of the microbial community was also studied to evaluate the performance of the technology of QCU for the removal of VOCs.

2. Materials and methods

2.1 Sample collection

The QCU used in the domestic waste treatment plant included the following processes: mechanical sorting–biological dehydration–secondary mechanical refinement–humified fermentation (Fig. 1). The cycle of the QCU of domestic waste takes approximately two months. Initial domestic waste underwent BD for 20 days after mechanical sorting, and then the dehydrated waste was further processed by a secondary mechanical refinement to ensure the proportion of organic materials. Finally, the high organic waste was humified fermented for 35 days to produce organic fertilizers. The difference between mechanical sorting and secondary mechanical refinement was the sieve diameter of the trommel screen. The VOCs in the gases from MBP were analysed in this study. Also, the VOCs in the gases produced in the process were collected by the BF. The BF included an upper compartment and a lower compartment, and each compartment had three layers. In each layer, 3 points were randomly selected and mixed into one sample of 300 g. The samples were immediately taken to the laboratory and stored in a refrigerator at $-80\text{ }^{\circ}\text{C}$.²⁶ The collected samples for the process of the QCU and BF are shown in Table 1.

2.2 Analysis method

2.2.1 Volatile organic compounds. The VOCs of gases generated from the QCU were determined by gas

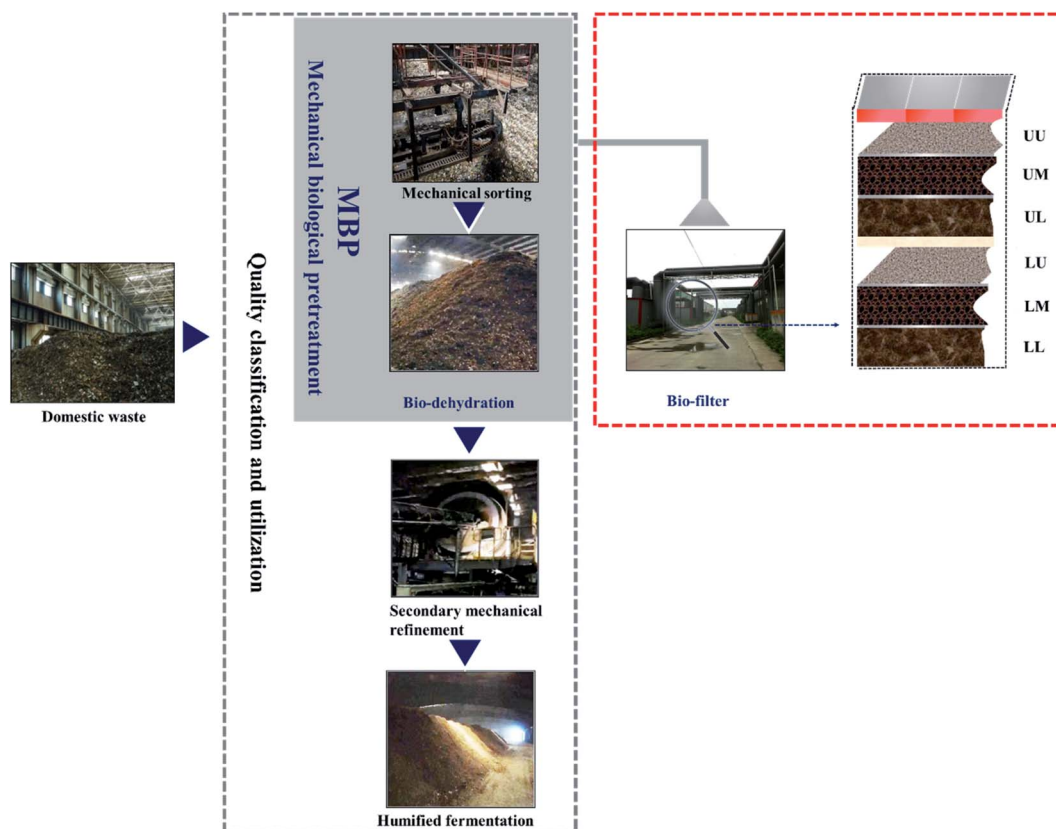


Fig. 1 Quality classification and utilization of domestic waste and BF for odor removal.

Table 2 Classification of the detected VOCs

Category	Volatile organic compounds
Saturated hydrocarbon	Trichloromethane, 1,1-dichloroethane, 1,2-dichloropropane, dichloromethane, cyclohexane, <i>N</i> -heptane, <i>N</i> -hexane
Unsaturated hydrocarbon	<i>cis</i> -1,3-Dichloropropene, hexachloro-1,3-butadiene, styrene, perchloroethylene, <i>trans</i> -1,3-dichloropropene, trichloroethylene
Aromatic hydrocarbon	Benzene, chlorobenzene, <i>M</i> -dichlorobenzene, <i>P</i> -dichlorobenzene, ethylbenzene, toluene, trimethylbenzene, xylene, 1-ethyl-4-methylbenzene
Sulfide	Carbon disulfide
Ketones	Acetone, methyl ethyl ketone, methyl isobutyl ketone, methyl <i>n</i> -butyl ketone
Ester	Ethyl acetate

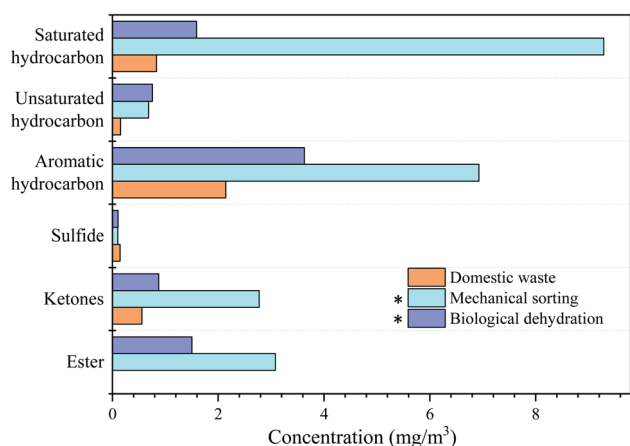


Fig. 2 Changes in the contents of VOCs during QCU. Saturated hydrocarbon (trichloromethane, 1,1-dichloroethane, 1,2-dichloropropane, dichloromethane, cyclohexane, *N*-heptane, and *N*-hexane), unsaturated hydrocarbon (*cis*-1,3-dichloropropene, hexachloro-1,3-butadiene, styrene, perchloroethylene, *trans*-1,3-dichloropropene, and trichloroethylene), aromatic hydrocarbon (benzene, chlorobenzene, *M*-dichlorobenzene, *P*-dichlorobenzene, ethylbenzene, toluene, trimethylbenzene, xylene, and 1-ethyl-4-methylbenzene), sulfide (carbon disulfide), ketones (acetone, methyl ethyl ketone, methyl isobutyl ketone, and methyl *n*-butyl ketone) and ester (ethyl acetate) were detected in VOCs. * Significant difference at $p < 0.05$.

mechanical sorting. For the mechanical motion, the content of saturated hydrocarbons after the mechanical sorting of domestic waste was 5.3 times that of the initial waste.³⁶ The increase in the content of some VOCs in the gases might also be attributable to the photodegradation caused by the operation of the drum screening machine for mechanical sorting.^{37,38}

The domestic waste was further subjected to BD to achieve the purpose of waste volume reduction. The content of total VOCs in the gases detected after BD was 3.81 mg m^{-3} , which was 2.2 times lower than that of the initial waste. As shown in Fig. 2, aromatic hydrocarbons, saturated hydrocarbons, and ketones were the key pollutants that should be controlled during this process. After BD, the content of saturated hydrocarbons decreased to one sixth of its initial value and the content of aromatic hydrocarbon was reduced to half of that of

mechanical sorting. However, BD did not contribute much to sulfide degradation but caused a slight increase in its content. The volatile sulfur compounds were the most important aromatic components produced by biodegradation.³⁹ Aromatic hydrocarbons are usually observed in transfer stations, landfills, and aerobic fermentation plants. Studies have shown that aromatic hydrocarbons were not produced through biodegradation but result from the chemically synthesized materials and household solvents contained in wastes.⁴⁰ BD had a better effect in controlling the release of VOCs compared with mechanical sorting. The results showed that in the MBP process, the content of target pollutants such as saturated hydrocarbons, aromatic hydrocarbons, ketones and esters had been reduced. However, the removal of VOCs in the gases from the MBP process was a synergistic BD and BF, which is the key aspect that needs to be explored through microbial changes.

3.2 Denatured gradient gel electrophoresis profile analysis

The 16S rDNA and 18S rDNA gene fragments of bacteria and fungi were revealed by denatured gradient gel electrophoresis (DGGE) patterns (Fig. 3). The distribution of microbes corresponding to different samples can be judged according to the position, number and brightness of the bands. As illustrated in Fig. 3, bands r and cc were detected among all padding filters of the BF, while band d was detected throughout the BD and humified fermentation. In addition, bands m and o appeared only in the lower compartment of the BF, while band n was detected only in the upper compartment. The results also revealed that the bacterial diversity decreased with the increase in the height of the padding filter, which was consistent with previous findings.⁴¹ Regarding BD and humified fermentation, band v and bands j, l, and gg appeared separately in humified fermentation and BD. The dehydration and fermentation were important for organic matter degradation, thereby accelerating rotting.^{33,42,43} Significant differences between classification, utilization, and biofiltration were found in the bacterial profiles. It can be seen from the bacterial DGGE map that compared to BD, the BF got a better biodiversity.

As shown in Fig. 3b, bands l–r were located in the lower part of the DGGE profile, signifying that these fungi had a higher G + C content³³ and better deodorization ability. The numbers of bands were in the order of resource utilization > biofiltration,



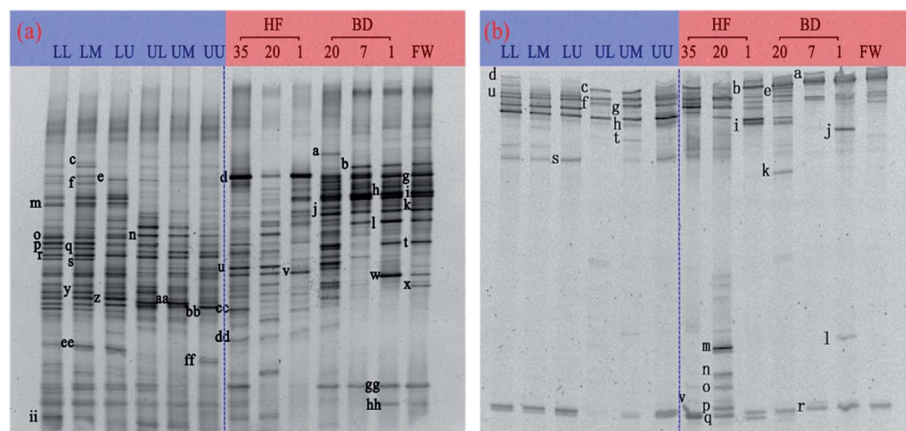


Fig. 3 DGGE maps of bacterial (a) and fungal (b) communities at different locations (purple area) and stages (red area). Samples of fresh waste (FW), biological dehydration for 1 day (BD1), biological dehydration for 7 days (BD7), biological dehydration for 20 days (BD20), humified fermentation for 1 day (HF1), humified fermentation for 20 days (HF20), humified fermentation for 35 days (HF35), up layer from the upper compartment of the BF (UU), middle layer from the upper compartment of the BF (UM), low layer from the upper compartment of the BF (UL), up layer from the lower compartment of the BF (LU), middle layer from the lower compartment of the BF (LM) and low layer from the lower compartment of the BF (LL) were detected.

which was consistent with the bacterial DGGE profile. The bands of the fungal DGGE maps of BD and BF were similar, and there was a small difference in the number of bands. The results showed that resource utilization could increase the amounts and types of the dominant fungi in the humified fermentation and BD. Regarding bands a–k from the top of the DGGE profile, band f appeared in both biofiltration and resource utilization, while band h occurred only in biofiltration. The number of bands in BD20 and HF20 was significantly higher than that in the other lanes. Among these, band k in BD20 and bands m–q in HF20 were the dominant species in their lane. Further analysis of the bands in the BF showed that bands u and s were typical in the lower compartment, and the types and numbers of these bands were in the order of lower compartment > upper compartment. The VOCs discharged by the QCU of the four-stage compartment preferentially passed through the lower compartment of the BF, where their air inflows were higher than those in the upper compartment of the BF. This is because VOCs are introduced into the BF, which has an inner effect on the microbes in the filler.⁴⁴

3.3 Dominant microbial communities of biofiltration

The number of bands and Shannon index of bacteria were more than those of fungi (Table 3). The number of bands in BD20 of bacteria was 26, which was 3.71 times that of fungi. The diversity (H) of bacteria was 3.19, which was 1.72 times that of fungi. The greater the diversity, the more abundant the species.⁴⁵ The diversity and abundance of the species decreased with the period of humified fermentation. Regarding biofiltration, the numbers of bands of lower padding filters were much more than those of upper padding filters, but the Pielou index of each padding layer did not show much difference, suggesting that the more abundant the species diversity, the more stable the biological community, which was consistent with the data in the PCR-DGGE analysis. The Shannon index of bacteria in the

lowest layer was 0.35 times higher than that in the highest layer. The filter material with the strongest bacterial diversity was the LL layer with a value of 3.48; however, for fungi, the filter material with the strongest fungal diversity was the LU layer with a value of 2.27. From the perspective of microbial abundance and diversity, bacteria in the BF and fungi in BD20 were more abundant.

Table 4 presents the sequencing results of the dominant bands of bacteria, which were obtained after comparing the sequence of the bands with the National Center for Biotechnology Information (NCBI) database. The results showed that *Lactobacillus* and *Bacillus* were the dominant species. The main difference was that some dominant bacteria in upper-compartment samples had a lower abundance compared with that in the lower compartment such as *Lactobacillus* (Fig. 4). For the original domestic waste, *Lactobacillus*, *Bacillus*, and *Lactococcus* were dominant with the respective abundances of 40.71%, 10.65%, and 10.52%. After mechanical sorting, BD, secondary mechanical refinement, and 35 day humified fermentation, the proportion of *Lactobacillus* was 14.85%, which was quite different from that of the original domestic waste. During the deodorization process of the BF, *Bacillus* could effectively degrade sulfurated hydrogen and ammonia in gases.⁴⁶ *Lactobacillus* is anaerobic, facultative anaerobic, or micro aerobic bacteria, which have good removal efficiency for ammonia and other odors.⁴⁶ *Thermobifida*, *Methylobacterium*, *Pseudoxanthomonas*, *Taibaiella*, *Enterococcus*, *Aerococcus*, and *Lactococcus* were also detected besides *Lactobacillus* and *Bacillus*. *Pseudoxanthomonas* and *Thermobifida* secreted xylanase and glucanase, which were involved in the degradation of cellulose and hemicellulose.⁴⁷ *Taibaiella* has an excellent effect on the removal of aniline, and it usually appears in the landfill leachate.⁴⁸ Therefore, during the classification and utilization of FW to HF35, they had always been dominant species. *Enterococcus* and *Lactococcus* were



Table 3 DGGE band diversity profiles of bacteria and fungi

Sample	Bacteria			Fungi		
	Band number	Shannon (<i>H</i>)	Pielou (<i>E</i>)	Band number	Shannon (<i>H</i>)	Pielou (<i>E</i>)
FW	19	2.85	0.97	5	1.57	0.98
BD1	11	2.35	0.98	7	1.87	0.96
BD7	10	2.22	0.97	5	1.47	0.92
BD20	26	3.19	0.98	7	1.85	0.95
HF1	7	1.87	0.96	4	1.35	0.98
HF20	17	2.81	0.99	11	2.37	0.99
HF35	8	2.02	0.97	5	1.52	0.95
UU	24	3.13	0.98	4	1.34	0.96
UM	19	2.90	0.99	7	1.84	0.95
UL	26	3.19	0.98	9	2.14	0.97
LU	30	3.35	0.99	11	2.27	0.95
LM	34	3.47	0.98	7	1.83	0.94
LL	34	3.48	0.99	5	1.56	0.97

pathogenic bacteria, which only existed in the initial stage of composting and disappeared as the temperature rises.⁴⁹ *Enterococcus* and *Lactococcus* were not detected in HF35, which meant that the QCU used in the domestic waste treatment

effectively removed pathogenic bacteria. Therefore, the dominant bacterial genera of BD and BF differ in composition. The dominant bacterial genera of BF are *Lactobacillus*,

Table 4 Sequencing analysis of 16S rDNA from bacteria

Sequences	Accession number	Species with the greatest similarity	Similarity, %
a	NR104932.1	<i>Erwinia billingiae</i> strain billing E63	100
b	NR113334.1	<i>Lactobacillus curvatus</i> strain NBRC 15884	99
c	NR109520.1	<i>Flavobacterium marinum</i> strain SW105	96.48
d	NR104708.1	<i>Aerococcus viridans</i> strain NBRC 12219	100
e	NR165710.1	<i>Mucibacter soli</i> strain R1-15	91.37
f	NR109509.1	<i>Chitinophaga cymbidii</i> strain R156-2	97.44
g	NR104573.1	<i>Lactobacillus plantarum</i> strain CIP	100
h	NR042438.1	<i>Lactobacillus graminis</i> strain G90 (1)	98.49
i	NR044702.2	<i>Lactobacillus amylophilus</i> strain DSM	98.47
j	NR115765.1	<i>Enterococcus faecalis</i> strain ATCC 19433	99.50
k	NR104926.1	<i>Lactobacillus bifementans</i> strain CIP 102811	99.49
l	NR113925.1	<i>Lactococcus lactis</i> subsp. <i>cremoris</i> strain NBRC 100676	100
m	NR109732.1	<i>Fabibacter pacificus</i> strain DY53	91.62
n	NR108906.1	<i>Bacillus eiseniae</i> strain A1-2	96.02
o	NR133876.1	<i>Taibaiella koreensis</i> strain THG-DT86	98.45
p	NR074291.1	<i>Bordetella petrii</i> strain DSM 12804	95.98
q	NR043110.1	<i>Pseudoxanthomonas kalamensis</i> strain JA40	98.49
r	NR114009.1	<i>Galbibacter mesophilus</i> strain NBRC 101624	92.31
s	NR029151.1	<i>Bacillus thermoamylovorans</i> strain DKP	98.99
t	NR029261.2	<i>Lactobacillus sanfranciscensis</i> strain L-12	99.49
u	NR159904.1	<i>Bacillus lacus</i> strain AK74	97.99
v	NR133975.1	<i>Bacillus kokeshiiformis</i> strain MO-04	95.98
w	NR112108.1	<i>Chroococcidiopsis thermalis</i> PCC 7203	89.83
x	NR117506.1	<i>Bifidobacterium longum</i> strain KCTC 3128	99.43
y	NR114899.1	<i>Methylobacterium populi</i> BJ001	98.28
z	NR044309.1	<i>Steroidobacter dentrificans</i> strain FS	96.98
aa	NR148571.2	<i>Chloroflexus islandicus</i> strain isl-2	88.44
bb	NR074876.1	<i>Dictyoglomus thermophilum</i> H-6-12	89.66
cc	NR112015.1	<i>Thermobifida fusca</i> strain IFO 14071	98.95
dd	NR137360.1	<i>Bacillus campisalis</i> strain SA2-6	98.47
ee	NR145530.1	<i>Actinoallomurus bryophytorum</i> strain NEAU-TX1-15 sequence	88.33
ff	NR042035.1	<i>Actinomadura umbrina</i> strain IMSNU 22165	89.27
gg	NR043148.1	<i>Lactobacillus oligofermentans</i> strain AMKR18	99.49
hh	NR113958.1	<i>Lactococcus lactis</i> subsp. <i>hordniae</i> strain NBRC 100931	100
ii	NR156918.1	<i>Luteitalea pratensis</i> strain HEG-639	88.83



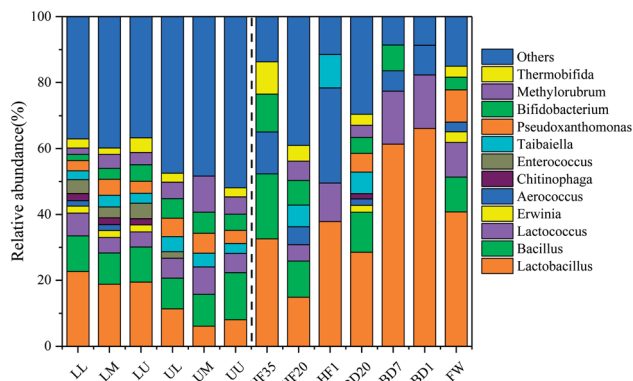


Fig. 4 Relative abundance of bacteria at the genus level. Samples of fresh waste (FW), biological dehydration for 1 day (BD1), biological dehydration for 7 days (BD7), biological dehydration for 20 days (BD20), humified fermentation for 1 day (HF1), humified fermentation for 20 days (HF20), humified fermentation for 35 days (HF35), up layer from the upper compartment of the BF (UU), middle layer from the upper compartment of the BF (UM), low layer from the upper compartment of the BF (UL), up layer from the lower compartment of the BF (LU), middle layer from the lower compartment of the BF (LM) and low layer from the lower compartment of the BF (LL) were detected.

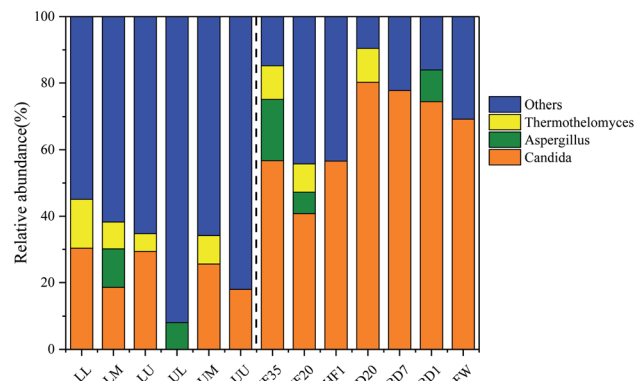


Fig. 5 Relative abundance of fungi at the genus level. Samples of fresh waste (FW), biological dehydration for 1 day (BD1), biological dehydration for 7 days (BD7), biological dehydration for 20 days (BD20), humified fermentation for 1 day (HF1), humified fermentation for 20 days (HF20), humified fermentation for 35 days (HF35), up layer from the upper compartment of the BF (UU), middle layer from the upper compartment of the BF (UM), low layer from the upper compartment of the BF (UL), up layer from the lower compartment of the BF (LU), middle layer from the lower compartment of the BF (LM) and low layer from the lower compartment of the BF (LL) were detected.

Taibaiella, etc. These dominant bacterial genera play a positive role in the removal of target pollutants.

The similar values of sequences a, b, j, s, and u in Table 5 were all more than 99%. The 13 dominant fungal bands were classified as *Candida*, *Aspergillus*, and *Thermothelomyces*. *Candida* has a significant impact on the production and changes of VOCs.⁵⁰ As illustrated in Fig. 5, *Candida* has always been the dominant genus during the QCU process of domestic

waste. It had the highest proportion of 80.27% in BD20, but it was not detected in the UL, indicating that VOCs were not conducive to the survival of *Candida*. *Aspergillus* can promote the removal rate of ammoniacal nitrogen and chemical oxygen demand in the landfill leachate and the activity of cellulolytic enzymes.⁵¹ The concentration of *Thermothelomyces* began to increase on the 20th day of biological dehydration, and it mainly existed in the padding filter of the BF for VOC degradation. *Thermothelomyces* was a common thermophilic genus,

Table 5 Sequencing analysis of 18S rDNA from fungi

No	Accession number	Species with the greatest similarity	Similarity, %
a	NG063407.1	<i>Candida sojae</i> JCM 1644	99.14
b	NG063416.1	<i>Candida quercitrusa</i> JCM 9832	99.43
c	NG064977.1	<i>Tritirachium cinnamomeum</i> IHEM 3714	89.82
d	NG062812.1	<i>Acremonium flavum</i> CBS 596.70	96.78
e	NG062131.1	[<i>Candida</i>] <i>fukazawae</i> JCM 1641	97.93
f	NG063411.1	<i>Candida austromarina</i> JCM 8894	95.42
g	NG064994.1	<i>Thermothelomyces hinnuleus</i> ATCC 52474	93.86
h	NG067668.1	<i>Megasporia hengduanensis</i> BJFC Cui 8076	97.20
i	NG063422.1	<i>Candida edax</i> JCM 9470	93.33
j	NG064802.1	<i>Aspergillus proliferans</i> CBS 121.45	99.14
k	NG065474.1	[<i>Candida</i>] <i>intermedia</i> JCM 1607	97.33
l	NG065472.1	[<i>Candida</i>] <i>pseudointermedia</i> JCM 1592	97.04
m	NG063232.1	<i>Aspergillus cervinus</i> NRRL 5025	80.5
n	NG063232.1	<i>Aspergillus cervinus</i> NRRL 5025	80.28
o	NG064802.1	<i>Aspergillus proliferans</i> CBS 121.45	98.56
p	NG063232.1	<i>Aspergillus cervinus</i> NRRL 5025	80.5
q	NG063419.1	<i>Candida viswanathii</i> JCM 9567	92.84
r	NG063242.1	[<i>Candida</i>] <i>glabrata</i> NRRL Y-65	98.57
s	NG064994.1	<i>Thermothelomyces hinnuleus</i> ATCC 52474	99.12
t	NG064938.1	<i>Aspergillus caninus</i> UAMH 10337	80.5
u	NG063242.1	[<i>Candida</i>] <i>glabrata</i> NRRL Y-65	99.43
v	NG064994.1	<i>Thermothelomyces hinnuleus</i> ATCC 52474	92.75



which has a significant catalytic oxidation effect on polycyclic molecules.⁵² *Thermothelomyces* also appeared in BD20, and the diversity of fungi in BD20 was high, indicating that BD also made a certain contribution to the removal of target pollutants.

4. Conclusions

This study compared the types and contents of VOCs in the gases from the original domestic waste, and illustrated that VOCs can be removed during the MBP process. It was recommended that VOCs produced in 20 days before BD should be monitored. *Lactobacillus* and *Bacillus* among bacteria, and *Candida*, *Aspergillus*, and *Thermothelomyces* among fungi were dominant with a positive role in the disposal of VOCs. In addition, the bacteria in the BF are the key step to remove VOCs in the gases, and the fungi in BD have also made a small contribution in the removal of VOCs. Moreover, inoculating *Lactobacillus*, *Bacillus*, *Candida*, *Aspergillus*, and *Thermothelomyces* into BF is expected to improve the removal efficiency of VOCs.

Author contributions

Jiaqi Hou: Data curation, Writing – original draft preparation, and Funding acquisition; Chengze Yu: Software and Formal analysis; Fanhua Meng: Validation and Software; Xiaosong He: Validation and Software; Yong Wang: Conceptualization and Methodology; Wangmi Chen: Reviewing and Editing; Mingxiao Li: Ideas and Supervision.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 P. Kjeldsen, M. A. Barla, A. P. Rooker, A. Baun, A. Ledin and T. H. Christensen, Present and Long-Term Composition of MSW Landfill Leachate: A Review, *Crit. Rev. Environ. Sci. Technol.*, 2002, **32**(4), 297–336.
- 2 H. Ma, Y. Cao, X. Lu, Z. Ding and W. Zhou, Review of Typical Municipal Solid Waste Disposal Status and Energy Technology, *Energy Procedia*, 2016, **88**, 589–594.
- 3 M. Fuss, M. Vergara-Araya, R. T. V. Barros and W.-R. Pogonietz, Implementing mechanical biological treatment in an emerging waste management system predominated by waste pickers: a Brazilian case study, *Resour. Conserv. Recycl.*, 2020, 162.
- 4 M. X. Paes, G. A. de Medeiros, S. D. Mancini, C. Gasol, J. R. Pons and X. G. Durany, Transition towards eco-efficiency in municipal solid waste management to reduce GHG emissions: the case of Brazil, *J. Clean. Prod.*, 2020, 263.
- 5 V. S. N. S. Goli, D. N. Singh and T. Baser, A critical review on thermal treatment technologies of combustible fractions from mechanical biological treatment plants, *J. Environ. Chem. Eng.*, 2021, **9**(4), 105643.
- 6 L. Rigamonti, G. Borghi, G. Martignon and M. Grosso, Life cycle costing of energy recovery from solid recovered fuel produced in MBT plants in Italy, *Waste Manag.*, 2019, **99**, 154–162.
- 7 W. Punin, S. Maneewan and C. Punlek, The feasibility of converting solid waste into refuse-derived fuel 5 via mechanical biological treatment process, *J. Mater. Cycles Waste Manag.*, 2013, **16**(4), 753–762.
- 8 M. López, M. Soliva, F. X. Martínez-Farré, M. Fernández and O. Huerta-Pujol, Evaluation of MSW organic fraction for composting: separate collection or mechanical sorting, *Resour. Conserv. Recycl.*, 2010, **54**(4), 222–228.
- 9 P. He, J. Tang, D. Zhang, Y. Zeng and L. Shao, Release of volatile organic compounds during bio-drying of municipal solid waste, *J. Environ. Sci.*, 2010, **22**(5), 752–759.
- 10 I. H. Suffet, V. Decottignies, E. Senante and A. Bruchet, Sensory assessment and characterization of odor nuisance emissions during the composting of wastewater biosolids, *Water Environ. Res.*, 2009, **81**(7), 670–679.
- 11 R. Lopez, I. O. Cabeza, I. Giraldez and M. J. Diaz, Biofiltration of composting gases using different municipal solid waste-pruning residue composts: monitoring by using an electronic nose, *Bioresour. Technol.*, 2011, **102**(17), 7984–7993.
- 12 D. P. Komilis, R. K. Ham and J. K. Park, Emission of volatile organic compounds during composting of municipal solid wastes, *Water Res.*, 2004, **38**(7), 1707–1714.
- 13 J. Hu, X. Gao, Q. Fan and X. Gao, Facial controlled synthesis of Pt/MnO₂ catalysts with high efficiency for VOCs combustion, *RSC Adv.*, 2021, **11**(27), 16547–16556.
- 14 G. Á. Mhuireach, L. Dietz, W. Griffiths, P. F. Horve, A. Laguerre, D. Northcutt, R. Vandegrift, E. Gall and K. Van Den Wymelenberg, Differing effects of four building materials on viable bacterial communities and VOCs, *Developments in the Built Environment*, 2021, 7, 100055.
- 15 Y. Zhang, J. Liu, J. Li and T. Yue, Effects of filler voidage on pressure drop and microbial community evolution in fungal bio-trickling filters, *Chemosphere*, 2021, **273**, 129710.
- 16 Z. Yang, J. Li, J. Liu, J. Cao, D. Sheng and T. Cai, Evaluation of a pilot-scale bio-trickling filter as a VOCs control technology for the chemical fibre wastewater treatment plant, *J. Environ. Manag.*, 2019, **246**, 71–76.
- 17 Y. Zhang, J. Liu, H. Xing and J. Li, Performance and fungal diversity of bio-trickling filters packed with composite media of polydimethylsiloxane and foam ceramics for hydrophobic VOC removal, *Chemosphere*, 2020, **256**, 127093.
- 18 S. K. Padhi and S. Gokhale, Treatment of gaseous volatile organic compounds using a rotating biological filter, *Bioresour. Technol.*, 2017, **244**(Pt 1), 270–280.
- 19 H. Wu, H. Yan, Y. Quan, H. Zhao, N. Jiang and C. Yin, Recent progress and perspectives in biotrickling filters for VOCs and



- odorous gases treatment, *J. Environ. Manag.*, 2018, **222**, 409–419.
- 20 E. R. Rene, B. T. Mohammad, M. C. Veiga and C. Kennes, Biodegradation of BTEX in a fungal biofilter: influence of operational parameters, effect of shock-loads and substrate stratification, *Bioresour. Technol.*, 2012, **116**, 204–213.
 - 21 J. Hu, L. Zhang, J. Chen, Y. Luo, B. Sun and G. Chu, Performance and microbial analysis of a biotrickling filter inoculated by a specific bacteria consortium for removal of a simulated mixture of pharmaceutical volatile organic compounds, *Chem. Eng. J.*, 2016, **304**, 757–765.
 - 22 M. C. Perez, F. J. Alvarez-Hornos, K. H. Engesser, D. Dobslaw and C. Gabaldon, Removal of 2-butoxyethanol gaseous emissions by biotrickling filtration packed with polyurethane foam, *N. Biotechnol.*, 2016, **33**(2), 263–272.
 - 23 F. J. Álvarez-Hornos, C. Lafita, V. Martínez-Soria, J. M. Penya-Roja, M. C. Pérez and C. Gabaldón, Evaluation of a pilot-scale biotrickling filter as a VOC control technology for the plastic coating sector, *Biochem. Eng. J.*, 2011, **58–59**, 154–161.
 - 24 S. Zhang, S. Xia, W. Sang, J. Yuan and W. Guo, Evaluation of functional microbes in explaining biofilter performance during start-up1, *J. Clean. Prod.*, 2020, 277.
 - 25 T. Zheng, L. Li, F. Chai and Y. Wang, Factors impacting the performance and microbial populations of three biofilters for co-treatment of H₂S and NH₃ in a domestic waste landfill site, *Process Saf. Environ. Prot.*, 2021, **149**, 410–421.
 - 26 Y. Yin, J. Gu, X. Wang, Y. Zhang, W. Zheng, R. Chen and X. Wang, Effects of rhamnolipid and Tween-80 on cellulase activities and metabolic functions of the bacterial community during chicken manure composting, *Bioresour. Technol.*, 2019, **288**, 121507.
 - 27 M. Delgado-Rodríguez, M. Ruiz-Montoya, I. Giraldez, R. López, E. Madejón and M. J. Díaz, Use of electronic nose and GC-MS in detection and monitoring some VOC, *Atmos. Environ.*, 2012, **51**, 278–285.
 - 28 Y. Yan, M. Wang, B. Jin, J. Yang and S. Li, Performance evaluation and microbial community analysis of the biofilter for removing grease and volatile organic compounds in the kitchen exhaust fume, *Bioresour. Technol.*, 2021, **319**, 124132.
 - 29 A. Sumi, K. Fukushi, R. Hon Da and K. M. Hassan, *Sustainable Water Resource Management in China: A Great Challenge*, Springer, Netherlands, 2010, ch. 1, 10.1007/978-90-481-9914-3, pp. 3–10.
 - 30 H. Liu, W. Lei, L. Mei, B. Wang, K. Jiang, S. Ma and Q. Li, The intestinal microbial diversity in Chinese shrimp (*Fenneropenaeus chinensis*) as determined by PCR-DGGE and clone library analyses, *Aquaculture*, 2011, **317**(1–4), 32–36.
 - 31 S. Saijai, A. Ando, R. Inukai, M. Shinohara and J. Ogawa, Analysis of microbial community and nitrogen transition with enriched nitrifying soil microbes for organic hydroponics, *Biosci. Biotechnol. Biochem.*, 2016, **80**(11), 2247–2254.
 - 32 M. Das, T. V. Royer and L. G. Leff, Diversity of fungi, bacteria, and actinomycetes on leaves decomposing in a stream, *Appl. Environ. Microbiol.*, 2007, **73**(3), 756–767.
 - 33 B. Xi, X. He, Q. Dang, T. Yang, M. Li, X. Wang, D. Li and J. Tang, Effect of multi-stage inoculation on the bacterial and fungal community structure during organic municipal solid wastes composting, *Bioresour. Technol.*, 2015, **196**, 399–405.
 - 34 C. Wu, J. Liu, S. Liu, W. Li, L. Yan, M. Shu, P. Zhao, P. Zhou and W. Cao, Assessment of the health risks and odor concentration of volatile compounds from a municipal solid waste landfill in China, *Chemosphere*, 2018, **202**, 1–8.
 - 35 L. Zhang, X. Su, Z. Zhang, S. Liu, Y. Xiao, M. Sun and J. Su, Characterization of fly ash from a circulating fluidized bed incinerator of municipal solid waste, *Environ. Sci. Pollut. Res. Int.*, 2014, **21**(22), 12767–12779.
 - 36 J. F. Saldarriaga, R. Aguado and G. E. Morales, Assessment of VOC Emissions from Municipal Solid Waste Composting, *Environ. Eng. Sci.*, 2014, **31**(6), 300–307.
 - 37 M. R. Ras, R. M. Marce and F. Borrull, Volatile organic compounds in air at urban and industrial areas in the Tarragona region by thermal desorption and gas chromatography-mass spectrometry, *Environ. Monit. Assess.*, 2010, **161**(1–4), 389–402.
 - 38 E. Nie, G. Zheng, Z. Shao, J. Yang and T. Chen, Emission characteristics and health risk assessment of volatile organic compounds produced during municipal solid waste composting, *Waste Manag.*, 2018, **79**, 188–195.
 - 39 L. Wenjing, D. Zhenhan, L. Dong, L. M. Jimenez, L. Yanjun, G. Hanwen and W. Hongtao, Characterization of odor emission on the working face of landfill and establishing of odorous compounds index, *Waste Manag.*, 2015, **42**, 74–81.
 - 40 J. Liu, G. Zheng, J. Liu and G. Zheng, Emission of volatile organic compounds from a small-scale municipal solid waste transfer station: ozone-formation potential and health risk assessment, *Waste Manag.*, 2020, **106**, 193–202.
 - 41 H. Benbelkacem, R. Bayard, A. Abdelhay, Y. Zhang and R. Gourdon, Effect of leachate injection modes on municipal solid waste degradation in anaerobic bioreactor, *Bioresour. Technol.*, 2010, **101**(14), 5206–5212.
 - 42 B. Xi, W. Meng, H. Liu, G. Huang, G. Zeng and Q. Wang, The variation of inoculation complex microbial community in three stages MSW composting process controlled by temperature, *Huanjing Kexue*, 2003, **24**(2), 152–155.
 - 43 B. Xi, G. Zhang and H. Liu, Process kinetics of inoculation composting of municipal solid waste, *J. Hazard Mater.*, 2005, **124**(1–3), 165–172.
 - 44 Z. Liang, J. Wang, Y. Zhang, C. Han, S. Ma, J. Chen, G. Li and T. An, Removal of volatile organic compounds (VOCs) emitted from a textile dyeing wastewater treatment plant and the attenuation of respiratory health risks using a pilot-scale biofilter, *J. Clean. Prod.*, 2020, 253.
 - 45 W. Zhang, Y. Mo, J. Yang, J. Zhou, Y. Lin, A. Isabwe, J. Zhang, X. Gao and Z. Yu, Genetic diversity pattern of microeukaryotic communities and its relationship with the environment based on PCR-DGGE and T-RFLP techniques in Dongshan Bay, southeast China, *Contin. Shelf Res.*, 2018, **164**, 1–9.



- 46 C. Franke, M. Hilgarth, R. F. Vogel, H. Petermeier and H.-C. Langowski, Characterization of the dynamics of volatile organic compounds released by lactic acid bacteria on modified atmosphere packed beef by PTR-MS, *Food Packag. Shelf Life*, 2019, **22**, 100400.
- 47 X. He, L. Han, B. Fu, S. Du, Y. Liu and G. Huang, Effect and microbial reaction mechanism of rice straw biochar on pore methane production during mainstream large-scale aerobic composting in China, *J. Clean. Prod.*, 2019, **215**, 1223–1232.
- 48 M. El-Fadel, F. Sleem, J. Hashisho, P. E. Saikaly, I. Alameddine and S. Ghanimeh, Impact of SRT on the performance of MBRs for the treatment of high strength landfill leachate, *Waste Manag.*, 2018, **73**, 165–180.
- 49 K. Tsigkou, D. Zagklis, P. Tsafrakidou, C. Zafiri and M. Kornaros, Composting of anaerobic sludge from the co-digestion of used disposable nappies and expired food products, *Waste Manag.*, 2020, **118**, 655–666.
- 50 P. Buzzini, A. Martini, U. M. Pagnoni and P. Davoli, Production of flavoured volatile organic compounds (VOCs) by *Candida oleophila* GK10, *Enzyme Microb. Technol.*, 2003, **33**(5), 668–675.
- 51 M. Gopinath, C. Mohanapriya, K. Sivakumar, G. Baskar, C. Muthukumaran and R. Dhanasekar, Microbial abatement of toluene using *Aspergillus niger* in upflow bioreactor, *Ecotoxicol. Environ. Saf.*, 2016, **134**(Pt 2), 370–376.
- 52 M. J. Furst, S. Savino, H. M. Dudek, J. R. Gomez Castellanos, C. Gutierrez de Souza, S. Rovida, M. W. Fraaije and A. Mattevi, Polycyclic Ketone Monooxygenase from the Thermophilic Fungus *Thermothelomyces thermophila*: A Structurally Distinct Biocatalyst for Bulky Substrates, *J. Am. Chem. Soc.*, 2017, **139**(2), 627–630.

