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Aptamer-functionalized pH-sensitive liposomes for a selective delivery of echinomycin into cancer cells†

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Echinomycin (quinomycin A) is a peptide antibiotic from the quinoxaline family, which has a DNA bifunctional intercalating activity and an inhibitor of hypoxia-inducible factor (HIF1 α). Echinomycin was discovered in 1957 as a potent antitumor agent; however, it was not successful in clinical use due to its low water solubility and short half-life. To revitalize this potent drug, it is important to increase its aqueous solubility and bioavailability. In this study, echinomycin was loaded into PEGylated pH-sensitive liposomes (PEG-Lip_{pH}) and functionalized with anti-nucleolin aptamer (Apt_{NCL}) for selective targeting and pH-responsive release of echinomycin into cancer cells. Echinomycin was complexed with γ -cyclodextrin (EC γ CD) to enhance its water solubility and then encapsulated into pH-sensitive liposomes (PEG-Lip_{pH}-EC γ CD). Then, liposomes were functionalized with Apt_{NCL} (Apt_{NCL}-PEG-Lip_{pH}-EC γ CD) and the successful functionalization was confirmed by dynamic light scattering (DLS) measurements and gel electrophoresis. Cellular uptake for Apt_{NCL}-PEG-Lip_{pH} was evaluated by flow cytometry analysis using MDA-MB-231, MCF7, A549 cancer cell lines with respect to the normal fibroblast cells. The results showed a higher uptake and selectivity for Apt_{NCL}-PEG-Lip_{pH} compared to PEG-Lip_{pH}. The anti-proliferative effects of Apt_{NCL}-PEG-Lip_{pH}-EC γ CD were more potent than PEG-Lip_{pH}-EC γ CD by 3.5, 4, and 5 folds for A549, MDA-MB-231, and MCF7, respectively. Selectivity indices (SI) for Apt_{NCL}-PEG-Lip_{pH}-EC γ CD for the tumor cell lines compared to the normal cell line after 72 h were MDA-MB-231 (43.3), MCF7 (16.9), and A549 (8.5). Furthermore, SI after 3 h for the three cancer cell lines were 4.7, 2.5, 2.8, respectively.

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1. Introduction

Echinomycin (quinomycin A), a hydrophobic DNA bis-intercalator peptide and an inhibitor of Hypoxia-Inducible Factor (HIF1 α) has been proved to have potent anticancer^{1,2} and antibacterial activities.^{3,4} Echinomycin structure is composed of two quinoxaline-2-carboxylic acid moieties, de-sipeptide ring, and a thioacetal bridge.⁵ Echinomycin is an example of promising drug that failed in clinical trials.^{3,6,7} For example, a clinical trial conducted to investigate echinomycin in patients with metastatic soft tissue sarcoma using cremophor as a solubilizing agent and dose escalation on subsequent cycles of therapy produced no clinical responses in the 12 evaluated patients.⁶ Unfortunately, because of severe side effects including severe nausea and vomiting dose escalation occurred in only 5 of 25 treatment cycles.⁶ Most anticancer drugs lack specificity and belong to substances with both low solubility in aqueous fluids and poor cellular uptake, which leads to unwanted side effects.⁸ The development of drug delivery systems implied to reconsider the benefit of such toxic products.^{8,9} In 2015, echinomycin received an orphan drug designation for the treatment of acute myeloid leukemia in the U.S.A.⁹

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Drug delivery systems offer the potential to enhance the therapeutic index of drugs, by increasing the drug concentration, the residence time in target cells, and by decreasing the contact in normal host tissues.^{10,11} Liposomes are widely used nanoparticles for drug delivery mainly due to their biocompatibility, stability, ease in formulation, high drug-loading efficiency,¹² and high bioavailability, and their safe excipients are used in formulations.^{13–15} Moreover, liposome cell-specific targeting can be achieved by decorating the surface of liposomes with ligand moieties such as aptamer, antibody, polymers, small molecules, and peptides.^{16,17} Aptamers are single-stranded oligonucleotides that can be selected for a wide range of targets and can bind its target with high affinity and selectivity. Aptamers were used as a targeting ligand for many drug delivery systems and showed potent and promising targeting efficacy *in vitro* and *in vivo*.^{18,19}

Trigger-release liposomes are smart liposomes that release their payload on demand in response to endogenous (pH, enzymes, redox, or molecules) or external stimuli (heat or light).²⁰ pH-triggered release liposomes undergo bilayer destabilization and leakage of their cargo at low pH inside diseased tissues or cells.^{21,22} These pH-sensitive liposomes are of particular interest for the delivery of anticancer drugs since the extracellular pH of cancer tissues is slightly acidic due to the^{22,23} high metabolic activity of cancer cells^{24,25} in addition to the low pH of some intracellular organelles that enable liposomes to release their loads into the cell cytoplasm.²⁶ These types of liposomes are categorized into different classes, mainly composed of polymorphic lipids, *e.g.*, dioleoyl phosphatidylethanolamine (DOPE), which can be stabilized by PEG and cholesterol.^{27–30}

After internalization inside tumor cells, pH-sensitive liposomes are retained in early endosomes (pH 6.5), which will be developed into late endosomes (pH 5), without being destructed by the lysosomal enzymes.^{31,32} There are three mechanisms for cytoplasmic delivery of the cargo from pH-sensitive liposomes; (i) destabilization of liposomes and drug defuses through the endosomal membrane into the cytoplasm, (ii) liposomes destabilize the endosomal membrane by the formation of pores in the membrane, and (iii) finally, the fusion of liposome with the endosome leading to drug release into the cytoplasm.^{22,33}

In a previous study, echinomycin was complexed with γ -cyclodextrin and inclusion complexes were encapsulated inside PEGylated liposomes and showed promising anti-proliferative and anti-invasive activity against U-87 MG glioblastoma cells.⁹ In the current study, we propose aptamer-guided and stimuli-sensitive liposomes as a smart multifunctional drug delivery system for selective delivery of echinomycin into breast and lung cancer cell lines. First, the echinomycin- γ -cyclodextrin complexes were prepared and loaded into PEGylated and pH-sensitive liposomes. Then, the loaded liposomes were functionalized with anti-nucleolin aptamer using a post-insertion method for targeting nucleolin-expressing cancer cells. Further, the selectivity and anti-proliferative activity of the resulting aptamer-functionalized and echinomycin-loaded pH-sensitive liposomes were tested against three types of nucleolin-expressing cancer cell lines and compared with normal cell line fibroblasts. Finally, the impact of freeze-drying on the stability and functionality of the formulated liposomes was investigated.

2. Materials and methods

2.1. Chemicals

Echinomycin (*N*-(2-quinoxaliny carbonyl)-*O*-[*N*-(2-quinoxaliny carbonyl)-*D*-seryl-*L*-alanyl-3-mercapto-*N,S*-dimethyl-cysteinyl-*N*-methyl-*L*-valyl]-*D*-seryl-*L*-alanyl-*N*-methyl cysteinyl-*N*-methyl *L*-valine (81)-lactone cyclic (37)-thioester) obtained from TOCRIS® bioscience (Bristol, UK). The lipids; DPPC (1,2-dihexadecanoyl-*sn*-glycero-3-phosphocholine), oleic acid, 1,2-dioleoyl-*sn*-glycero-3-phosphoethanolamine (DOPE), DSPE-PEG₂₀₀₀(1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-[methoxy (polyethyl-ene glycol)-₂₀₀₀]), 1,2-Dioleoyl-*sn*-glycero-3-phosphoethanolamine-*N*-(lissamine rhodamine B sulfonyl) (Lissamine) (Rho-PE), and cholesterol were purchased from Avanti Polar Lipids (Alabaster, USA). Calcein dye, γ -cyclodextrin, and trypan blue 0.4% solution were purchased from Sigma-Aldrich (USA). Anti-nucleolin aptamers with the sequence: (5'-GGT GGT GGT GGT GGT TGT GGT GGT GGT GGT TTT TTT TTT TT/3CholTEG/-3') and alexa-labeled anti-nucleolin aptamers (5'-/5Alexa647N-GGT GGT GGT GGT GGT TGT GGT GGT GGT TTT TTT TTT TT/3CholTEG/-3') were purchased from Integrated DNA Technologies Inc. (Coralville, IA, USA). Phosphate buffer saline (PBS) and Tris-base EDTA buffer (TBE) were obtained from LONZA® (USA). 6-Diamidino-2-phenylindole dihydrochloride (DAPI) from Merck Millipore (Germany) and ethidium bromide from Fisher Scientific Ltd (England). All other chemicals and solvents were of analytical grade. All reagents and chemicals were used without further treatment.

2.2. Cells

MCF-7 (ATCC number: HTB-22) and A549 (ATCC Number: CCL-185) cell lines were cultured in RPMI-1640 growth medium (Capricorn Scientific GmbH, Germany), MDA-MB-231(ATCC number: HTB-26) and human skin cells (fibroblasts) (ATCC® PCS-201-012) cell lines were cultured Eagle's Minimum Essential Medium (EMEM) (Euroclone SpA, Italy). Both RPMI-1640 and EMEM medium were supplemented with 10% (v/v) fetal bovine serum (FBS), 1% (v/v) 200 mM *L*-glutamine, and antibiotics, penicillin-streptomycin (100 IU/mL-100 μ g mL⁻¹). Cells were maintained in humidified 5% CO₂ incubator at 37 °C.

2.3. Preparing echinomycin- γ -cyclodextrin (EC γ CD) complex

Based on our previous work,⁹ EC γ CD inclusion complexes were prepared using the co-evaporation dispersion method. Echinomycin solution in chloroform (100 μ L) was added dropwise to 235 μ L of γ CD aqueous solution to get an EC γ CD ratio of 1 : 2 (v/v). The complex solution was shaken for 24 h to achieve equilibrium, then chloroform was evaporated under vacuum for 30 min. The final volume of EC γ CD inclusion was adjusted to 1 mL in PBS (pH 7.4) for loading into liposomes.⁹

2.4. Preparation of liposomes

Liposomes were formulated using molar ratios of 40% of DPPC, 30% of DOPE, 30% of cholesterol, 3% DSPE-PEG₂₀₀₀ (10 mM



total lipids concentration). Liposomes made from 10 mM of DPPC and cholesterol (70 : 30) were used for comparison. PE-rhodamine-labelled liposomes were prepared using PE-rhodamine 0.5% molar ratio subtracted from the DPPC ratio.

Liposomes were prepared using the conventional thin-film hydration technique.^{34–36} Briefly, lipids were mixed in 3 mL of chloroform in a 50 mL round bottom flask. The thin film was formed by evaporation of chloroform using 20 rpm at 50 °C under a gradual decrease in negative pressure down to 100 psi (IKA RV 05 Basic Rotary Evaporator combined with VacuuBrand CVC2000, Germany). The film was then hydrated with 2 mL of PBS buffer, pH 7.4 at ~50 °C for 30–60 min with vigorous vortexing every 2–3 min.³⁴ The vesicle suspension was successively extruded through a polycarbonate membrane (100 nm, Whatman®) using Mini-Extruder (Avanti Polar Lipids, Inc. USA) at 50 °C for 13 times (same size for more than 13 times) to obtain the final liposomes with low polydispersity and desired size, which then stored at 4 °C for further use.

2.5. Calcein releasing assay

For the calcein-releasing assay, the dry film was dispersed into 2 mL of 50 mM calcein solution of pH 8.8. Then, calcein-loaded vesicles (50 µL) were added to 1 mL PBS solutions with different pHs (4.4, 5.4, 6.4, and 7.4). The mixtures were incubated for 15 min at room temperature under 100 rpm stirring. Then, 10 µL of the samples were diluted to 1 mL and the pH was adjusted to 7.4. Calcein fluorescence was measured using the GloMax-Multi detection system (Promega, USA) before and after the addition of 50% absolute ethanol.

The percentage of calcein release was calculated using the following equation:

$$\% \text{ calcein release} = \frac{(F_t - F_i)}{(F_t - F_i)} \times 100 \quad (1)$$

where F_i is the fluorescence intensity of calcein in the buffer (pH 7.4), F_t is the fluorescence intensity after incubation in the acid buffer, and F_r is the fluorescence intensity after the addition of 50% absolute ethanol. The fluorescence intensity at various pH should be adjusted to the intensity at pH 7.4 since the liposomes were prepared at pH 7.4 and the intensity of calcein fluorescence decreases at acidic pH.

2.6. Simulations of the liposomal lipid bilayer decorated with PEG₂₀₀₀ and Apt_{NCL}

Molecular dynamics (MD) simulations of the liposomal heterogeneous lipid bilayer composed of 37% of DPPC, 30% of DOPE, 30% of cholesterol, 3% DSPE-PEG₂₀₀₀ were performed using the CHARMM-GUI Membrane Builder (<http://www.charmm-gui.org/input/membrane>), a web-based graphical user interface. The initial membrane bilayer was built with 200 Angstroms for the X and Y axes. A replacement method was used for the build system, and 171 positive and 171 negative ions (0.15 M KCL) were added to neutralize the system. The final resulting bilayer contains 225 cholesterol molecules, 285 DPPC, 15 DSPE-PEG₂₀₀₀, and 225 DOPE.

PEG₂₀₀₀ conical smile was imported from PUBCHEM (<https://pubchem.ncbi.nlm.nih.gov>) to Discovery Studio (version 2.5.5, Biovia, San Diego, CA, USA), and the covalent bond forming tool was used to connect it to DSPE. The aptamer was imported from the PDB databank (PDB ID: 2N3M) by removing the first nucleotide at the 5' end and adding an extra 11 thymidine monophosphate (TMP) at the 3' end by Discovery Studio. The resulting aptamer was added by a linker using a “covalent bond forming tool” in Discovery studio to a cholesterol molecule (according to the experimentally calculated ratio). The final structure was minimized several times with the minimization tool in Discovery studio until converging to the most stable state.

2.7. Preparing liposomes loaded with echinomycin-γ-cyclodextrin complexes

pH-Sensitive liposomes composed of DPPC, DOPE, cholesterol, and DSPE-PEG₂₀₀₀ with a molar ratio of 37 : 30 : 30 : 3, respectively, were prepared by a thin-film-hydration extrusion method as described before. Instead of PBS buffer hydration, the thin film was hydrated with 400 µL of ECγCD complexes and continued with the liposomal preparation protocol. The free ECγCD complexes were removed and washed two times with PBS solution using ultrafiltration 100 kDa cutoff amicon filter (Millipore, USA). The ECγCD loaded-liposomes were stored at 4 °C for further use.³⁷

2.8. Differential scanning calorimetry (DSC) for pH-sensitive liposomes

The liposome samples weights in the range of 3–6 mg were analyzed by Netzsch DSC 204 F1 instrument (Germany). The liposome suspension was heated up to 50 °C and was then cooled with liquid nitrogen to –50 °C. The DSC patterns of the samples were obtained by heating liposomes suspension gradually from –50 °C to 200 °C at a heating rate of 5 °C min^{–1} under a constant flow (100 mL min^{–1}) of nitrogen gas.

2.9. Post-insertion of Apt_{NCL} into PEGylated liposomes

Functionalizing liposomes and PEG-liposomes with Apt_{NCL} aptamer was performed using a post-insertion method.^{38,39} Free or loaded different liposome formulations were mixed with cholesterol-linked Apt_{NCL} at (0.3 mg lipid: 10 µg Apt_{NCL}) or Alexa-tagged Apt_{NCL} (0.3 mg lipid: 9 µg Apt_{NCL}: 1 µg Alexa-tagged Apt_{NCL}). The mixture was stirred at room temperature (25 °C) for 1 h. Post-inserted Apt_{NCL} was finally mixed with 2.5 mM MgCl₂ and 140 mM KCl in 1× PBS, pH 7.4 as a folding buffer for an extra 10 min to enable folding.

2.10. Characterization of aptamer-liposomes (Apt_{NCL}-Lip_{pH})

2.10.1. Agarose gel electrophoresis. Agarose gel electrophoresis was performed to confirm the successful conjugation of Apt_{NCL} to the surface of liposomes. Samples of free liposomes, free Apt_{NCL} and Apt_{NCL}-PEG-Lip_{pH} were loaded into 3% agarose gel (Promega, USA) supplemented with 5 µl of 2.5 mg mL^{–1} ethidium bromide followed by running electrophoresis in



1× TBE buffer (pH 8) at 85 V for 15 min. Images for analysis were obtained using the Chemi-Doc™ Bio-Rad gel imaging system (USA).³⁴

2.10.2. Size and zeta potential. The size, PDI and zeta potential of different liposomal formulations, encapsulated liposomes and free liposomes, were measured at 25 °C by dynamic light scattering (DLS) using nano zeta sizer (Malvern Instruments, UK). The different liposome formulations were diluted in PBS 1× to obtain 8 mM sodium chloride final concentration (pH 7.4).³⁴

2.11. Freeze-drying of the liposomes

Freshly prepared liposomal suspensions were freeze-dried in the presence of trehalose and sucrose ratios; 1%, 2.5%, 5%, and 10% (w/v). Liposomes without lyoprotectant were lyophilized as a reference. Liposomes were freeze-dried using an OPERON freeze-dryer (OPERON Co., Ltd). A volume of 1 mL liposomal dispersion was transferred into vials and stored for 24 h at −20 °C. Then, the liposomes were soaked in liquid nitrogen for 10 min. The temperature of the freeze dryer was then lowered to −55 °C. Sublimation of the aqueous solvent was then initiated by decreasing the pressure to 150 μbar for the next 24 h. At the end of the process, the vials were closed with rubber caps and stored at −20 °C until further analysis. The influence of the type and molar ratio of lyoprotectant on the final quality of the product was examined by measuring size and charge before and after lyophilization, with and without lyoprotectants.

2.12. Flow cytometry analysis

Apt_{NCL}-Lip_{pH} binding selectivity and cellular uptake against four cell lines (fibroblast, A459, MCF7, and MDA-23) were evaluated by measuring the mean fluorescence intensity (MFI) using a flow cytometer (BD FACSCanto™ II). Approximately, 2.5 × 10⁵ cells of each cell line were seeded in 12-well plates and incubated for 24 h at 37 °C to reach 80% confluency. After that, cells were washed with PBS and incubated in 500 μL of fresh media and 200 nM tagged Alexa-Apt_{NCL}-Rhod-Lip_{pH} was incubated with cells for 3 h at 37 °C. The unbound Alexa-Apt_{NCL}-Rhod-Lip_{pH} was washed three times with PBS and detached by 100 μL of EDTA-accutase for 5 min. After detachment, cells were transferred into flow cytometry tubes and 10 000 events were counted for each sample using y FACS Canto II and analyzed using BD FACSDiva™ software version 8.0 (BD, USA). Cells treated with lipo-Rhod and Alexa-tagged aptamers were used for comparison. Cells with media have been used as negative controls.

2.13. *In vitro* stability of PEG-Lip_{pH}-ECγCD and Apt_{NCL}-PEG-Lip_{pH}-ECγCD

The stability assay of PEG-Lip_{pH}-ECγCD and Apt_{NCL}-PEG-Lip_{pH}-ECγCD was performed by measuring the mean hydrodynamic diameter, zeta potential, and PDI in PBS at pH 7.4 with a storage period of 10 at 4 °C and 3 days at 37 °C. The releasing assay of echinomycin from PEG-Lip_{pH}-ECγCD and Apt_{NCL}-PEG-Lip_{pH}-ECγCD was performed at 37 °C in PBS at pH 7.4 and 5.4. Different samples of 700 μL were incubated at different time

intervals (0, 10, 20, 30,60, and 720 min). The free ECγCD inclusions at each time point were removed by ultrafiltration using Amicon® filters (cut-off of 100 kDa) (Millipore, Germany). The concentration of echinomycin in the liposomal fraction was quantified by HPLC.

2.14. Cytotoxicity (MTT) assay of Lip_{pH}-ECγCD and Apt_{NCL}-Lip_{pH}-ECγCD

The cytotoxicity of free echinomycin and liposomal preparations was measured using the cell viability MTT assay (micro-culture tetrazolium). Cells were seeded at a density of 5 × 10⁴ cells per well in a microtiter 96-well plate (Costar, USA) in 100 μL complete culture medium and incubated for 24 h in a humidified atmosphere of 5% CO₂ at 37 °C. Then, stock solutions of free echinomycin (free EC), PEG-Lip_{pH}-ECγCD, and Apt_{NCL}-PEG-Lip_{pH}-ECγCD were serially diluted in the medium of concentrations range from 0.02 to 50 nM in 100 μL medium and added to seeded cells. After that, treated cells were incubated in a humidified atmosphere of 5% CO₂ at 37 °C for 72 h. For comparison with the 72 h treatment time, cells were treated with similar concentrations of free EC, PEG-Lip_{pH}-ECγCD, and Apt_{NCL}-PEG-Lip_{pH}-ECγCD for 3 h and replaced with 100 μL fresh medium and incubated in a humidified atmosphere of 5% CO₂ at 37 °C for 72 h. The solutions were then removed and replaced with 100 μL fresh media and 15 μL of MTT solution were added to each well, followed by incubation at 37 °C for 3 h. After that, MTT-media solution was removed and 50 μL of DMSO was added to solubilize the dark blue formazan crystals. The absorbance of formazan was measured at 570 nm with a microplate reader Synergy™ HTX by BioTek Instruments Inc. (USA) and then IC₅₀ values were calculated according to the following equation:

$$\text{cell viability \%} = \frac{\text{average of samples' OD}}{\text{average of control}} \quad (2)$$

$$\text{selectivity index (SI)} = \frac{\text{IC}_{50} \text{ of normal cells}}{\text{IC}_{50} \text{ of cancer cells}} \quad (3)$$

3. Results and discussion

3.1. pH-sensitive liposomes preparation and characterization

3.1.1. Calcein dye releasing assay. Dioleoylphosphatidyl ethanolamine (DOPE) can form an inverted hexagonal (H_{II}) phase above 10–15 °C in neutral and acidic media.⁴⁰ Phosphatidyl ethanolamine lipids have a smaller head group to acyl chain ratio, when the smaller heads get close to each other, the broader the acyl chains leading to a positive curvature.⁴¹ On the other hand, the phosphatidylcholine lipids are packed and organized into lamella without any curvature.⁴²

To optimize a stable pH-sensitive liposomal formulation with a good release under acidic pHs, DPPC was used as the bulk constituent of the formulation (≥40%), while the DOPE molar ratio was ≤30%. DPPC can form a stable lamellar phase (L_β) and has a transitional temperature slightly higher than



physiological temperature (41 °C).⁴³ This stable lipid can decrease the intermolecular interaction of the headgroups of DOPE and increase surface hydration.³² Cholesterol, at a molar ratio up to 30%, increased the stability of nanoparticles.⁴⁴ Cholesterol fills up gaps between the packing of other lipid species. It is a membrane constituent widely found in biological systems and serves a unique purpose of modulating membrane fluidity, elasticity, as well as permeability.⁴⁴ Cholesterol serves the same purpose in model membranes.⁴⁵ These lipid combinations are consistent with a previous literature^{10,33} in which the pH-sensitive liposomes should contain the very low transition temperature phospholipid, DOPE, as a basic component in addition to cholesterol as a stability lipid.⁴³

The calcein-releasing experiment revealed that the release at physiological pH (pH \approx 7.4) was (8.82 \pm 2.5%), while the release at pH \approx 5.4 was (46.65 \pm 6.3%) (Fig. 1A). Based on these results, the pH-sensitive liposomal formulation is stable at physiological pH and releases approximately 50% of its content at the endosomal pH. Moreover, the incorporation of DSPE-PEG₂₀₀₀ into DOPE-containing liposomes decreased the release of calcein (24.7 \pm 1.2%) from DOPE-containing liposomes at acidic pHs, which indicates higher stability and lower pH-sensitivity. Besides, liposomes containing DPPC and cholesterol only showed low and stable release at different pHs.

The stability and pH sensitivity results of the developed formula were consistent with previous studies. For example, Ghanbarzadeh *et al.*, (2014) developed a liposomal formula composed of DPPC: DOPE: CHO: with a molar ratio of 50 : 30 : 20, respectively, and had a better antiproliferative effect on MCF-7 cells compared to conventional liposomes.³³

3.1.2. Differential scanning calorimetry (DSC). The DSC method was used to determine the phase changes in the pH-sensitive liposomes' transitional temperature (T_m) due to the presence of DOPE in different formulations. DSC analysis depends on the thermodynamic concept of heat flow differences among samples and the reference. Thus, the calorimeter measures the amount of heat absorbed or released during material transformation.⁴⁶

Fig. 1B shows a clear endothermic peak between 10–14.6 °C for the T_m of DOPE in the intended liposomes. Another broad and highly endothermic peak at 100–130 °C, which indicates the evaporation of water or melting of other constituents. The slight shift in DOPE T_m indicates the effect of different formulations on the DOPE transition temperature. Despite these slight shifts in DOPE T_m , the results of DSC and calcein release at acidic pH indicate that the release is pH-sensitive rather than thermo-sensitive.

3.2. Apt_{NCL} post-insertion into liposome surface and characterization of Apt_{NCL}-PEG-Lip_{pH}

In this study, PEGylated pH-sensitive liposomes (PEG-Lip_{pH}) were functionalized with anti-nucleolin cholesterol-modified AS1411 aptamer (Apt_{NCL}-PEG-Lip_{pH}) to selectively target nucleolin-expressing cancer cells. The liposomes were stealthed with 3% DSPE-PEG₂₀₀₀ and encapsulated with calcein to investigate release and pH-sensitivity or with EC γ CD complexes to investigate the anti-proliferative effect on different cancer cell lines. The pegylated Lip_{pH} was grafted with Apt_{NCL} using a post-insertion method that utilized cholesterol (CHO) as an anchoring unit to incorporate Apt_{NCL} on the surface of preformed and loaded liposomes (Fig. 2A).³⁸

3.2.1. Agarose gel electrophoresis. The post-insertion of Apt_{NCL} on the surface of pH-sensitive liposomes was also confirmed by agarose gel electrophoresis (Fig. 2B). Free aptamers and blank liposomes were used as controls for comparison. Apt_{NCL}-PEG-Lip_{pH} appeared to be entrapped in the well where a very low signal was obtained for blank Lip_{pH}. As a negatively charged single-strand DNA, free Apt_{NCL} can transfer freely through the gel from the negative to the positive electrode, while the large molecular weight of Apt_{NCL}-PEG-Lip_{pH} prevents Apt_{NCL} from free migration and was trapped in the well.

3.2.2. Size and zeta potential. The effect of Apt_{NCL} post-insertion on the size and zeta potential of preformed pH-sensitive liposomes was examined and compared to blank liposomes as illustrated in Fig. 2C. The mean hydrodynamic size

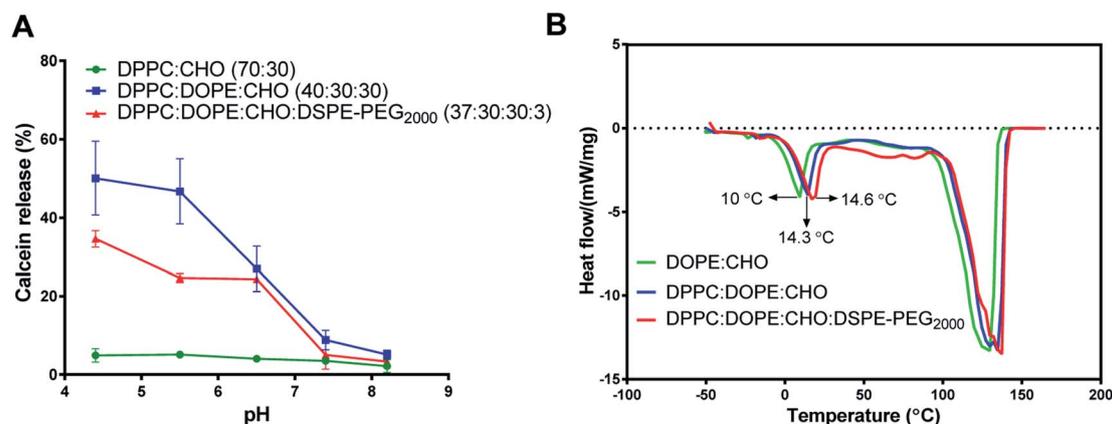


Fig. 1 Effect of lipid constituents on pH sensitivity and transitional temperature (T_m) of different liposomal formulations. (A) Calcein dye leakage at different pHs from (DPPC:CHO), (DPPC:DOPE:CHO), and (DPPC:DOPE:CHO:DSPE-PEG₂₀₀₀) liposomal formulations. (B) Differential scanning calorimetric (DSC) analysis for (DOPE:CHO), (DPPC:DOPE:CHO), and (DPPC:DOPE:CHO:DSPE-PEG₂₀₀₀). (mean \pm SD, $n = 3$, CHO was 30% in all formulations).



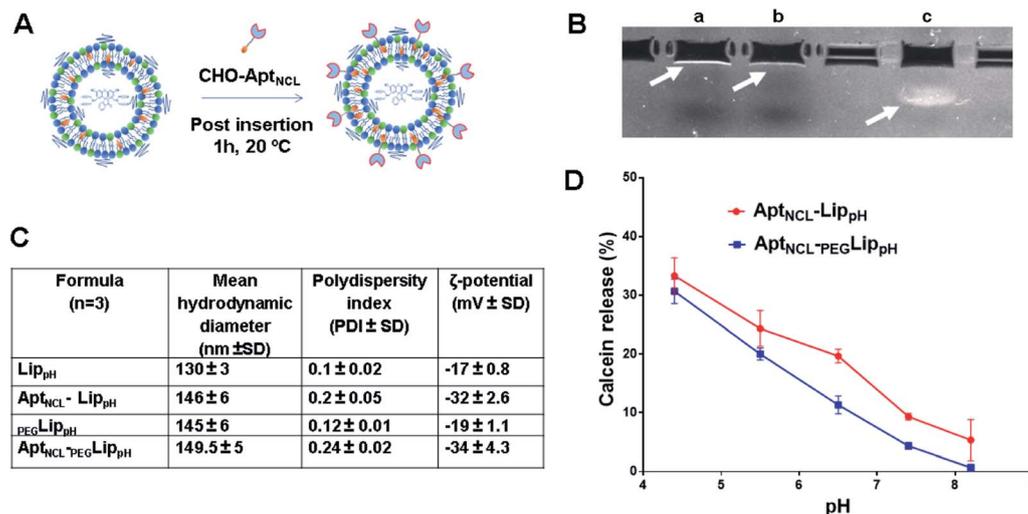


Fig. 2 Apt_{NCL} post-insertion and pH sensitivity of Apt_{NCL}-Lip_{pH} and PEGylated Apt_{NCL}-PEG-Lip_{pH} formulations. (A) Schematic representation of post-insertion of cholesterol-conjugated Apt_{NCL} on the surface of Lip_{pH} and PEG-Lip_{pH}. (B) and (C) Characterization of liposomes before and after Apt_{NCL} post insertion on the surface of Lip_{pH} and PEG-Lip_{pH}. (B) Agarose gel electrophoresis. Lanes: a, Apt_{NCL}-PEG-Lip_{pH}; b, Free PEG-Lip_{pH}; c, free Apt_{NCL}. (C) the change of hydrodynamic diameter and the polydispersity index of Lip_{pH}, Apt_{NCL}-Lip_{pH}, PEG-Lip_{pH}, and Apt_{NCL}-PEG-Lip_{pH}. (D) calcein dye releasing assay from Apt_{NCL}-Lip_{pH} and PEGylated Apt_{NCL}-PEG-Lip_{pH} formulations. (mean ± SD, n = 3).

and PDI of Apt_{NCL}-Lip_{pH} (146 ± 6 nm and 0.2 ± 0.05) and Apt_{NCL}-PEG-Lip_{pH} (149.5 ± 5 nm and 0.24 ± 0.2) were found to be slightly higher than the Lip_{pH} (130.5 ± 3 nm and 0 ± 0.02) and PEG-Lip_{pH} (145 ± 6 nm and 0.12 ± 0.01). Moreover, the zeta potential of Apt_{NCL}-Lip_{pH} (-32 ± 2.6 mV) and Apt_{NCL}-PEG-Lip_{pH} (-34 ± 4.3 mV) were decreased compared to Lip_{pH} (-17 ± 0.8 mV) and PEG-Lip_{pH} (-19 ± 1.1 mV) in diluted PBS buffer (8 mM NaCl, pH 7.4).

The increase in liposome size (16 nm) and a decrease in zeta potential (-15 mV) confirm the successful post-insertion of the negatively charged aptamer to our liposomes. Moreover, our results indicated that aptamer conjugation has increased liposomal stability by decreasing aggregation through the process of aptamer immobilization onto the surface of Lip_{pH} as reported in previous reports.⁴⁷

3.3. pH-Sensitivity and calcein release from the Apt_{NCL}-PEG-Lip_{pH}

The calcein releasing assay was performed to demonstrate the effect of aptamer and PEG₂₀₀₀ on the pH sensitivity and stability of liposome formulation. Fig. 2D shows higher stability of Apt_{NCL}-PEG-Lip_{pH} liposomes than Apt_{NCL}-Lip_{pH} at different pHs. These results evidenced the role of PEG and aptamer in increasing liposome stability.

3.4. Cellular uptake and selectivity of Apt_{NCL}-Lip_{pH} using flow cytometry

The *in vitro* selectivity and cellular uptake of Apt_{NCL}-PEG-Lip_{pH} against MDA-MB-231, MCF7, A549, and normal skin fibroblast cells were examined by measuring the mean fluorescence intensity (MFI) of the fluorescent signal obtained from Alexa 647-labeled Apt_{NCL} and rhodamine-labeled liposomes in free Apt_{NCL}, PEG-Lip_{pH}, and Apt_{NCL}-PEG-Lip_{pH} (Fig. 3A–D).

All examined cells showed higher MFI of rhodamine signals when treated with Rhod-labelled Apt_{NCL}-PEG-Lip_{pH} compared to

rhod-labeled PEG-Lip_{pH} and untreated cells, whereas all cells showed a slight increase in the MFI of Alexa Fluor 647 signals when treated with Alexa- and Rhod-labelled Apt_{NCL}-PEG-Lip_{pH} compared to Rhod labeled PEG-Lip_{pH} and untreated cells. The cellular uptake results proved the role of the aptamer for liposomal selective targeting and accumulation into the cancer cells by at least by two folds *versus* the plain liposome in cancer cells. The same conclusion can be obtained from the flow cytometry histogram analysis as reported in Fig. 3A–D. Interestingly, fibroblasts showed a higher uptake for the free Alexa-Apt_{NCL} and Alexa- and Rhod-labelled Apt_{NCL}-PEG-Lip_{pH} compared to cancer cell lines. Previous reports have explained that the higher uptake of the anti-nucleolin aptamer by the normal cells compared to cancer cells is due to different uptake mechanism.^{48,49} In cancer cells, the uptake was mediated by macropinocytosis (fluid-phase endocytosis) that stimulates further macropinocytosis by nucleolin-dependent mechanism while in normal cells the uptake is mediated by a different mechanism (nonmacropinocytic pathway).

3.5. Encapsulation of echinomycin-γCD into pH-sensitive liposomes (PEG-Lip_{pH}-ECγCD)

In our previous study,⁹ echinomycin-γCD (ECγCD) was prepared and characterized using ¹H-NMR spectroscopy, the phase solubility diagram, and the Benesi–Hildebrand method. In the current work, ECγCD was prepared and encapsulated into PEGylated pH-sensitive liposome formulation composed of DPPC : DOPE : CHO : DSPE-PEG₂₀₀₀ with 37 : 30 : 30 : 3 molar ratio (Fig. 4A–D). It is worth mentioning that the current formula is different from the previously investigated liposomes, which were composed of DPPC, and cholesterol.

Fig. 5A shows DLS measurements of PEG-Lip_{pH}, PEG-Lip_{pH}-ECγCD, and Apt_{NCL}-PEG-Lip_{pH}-ECγCD in addition to



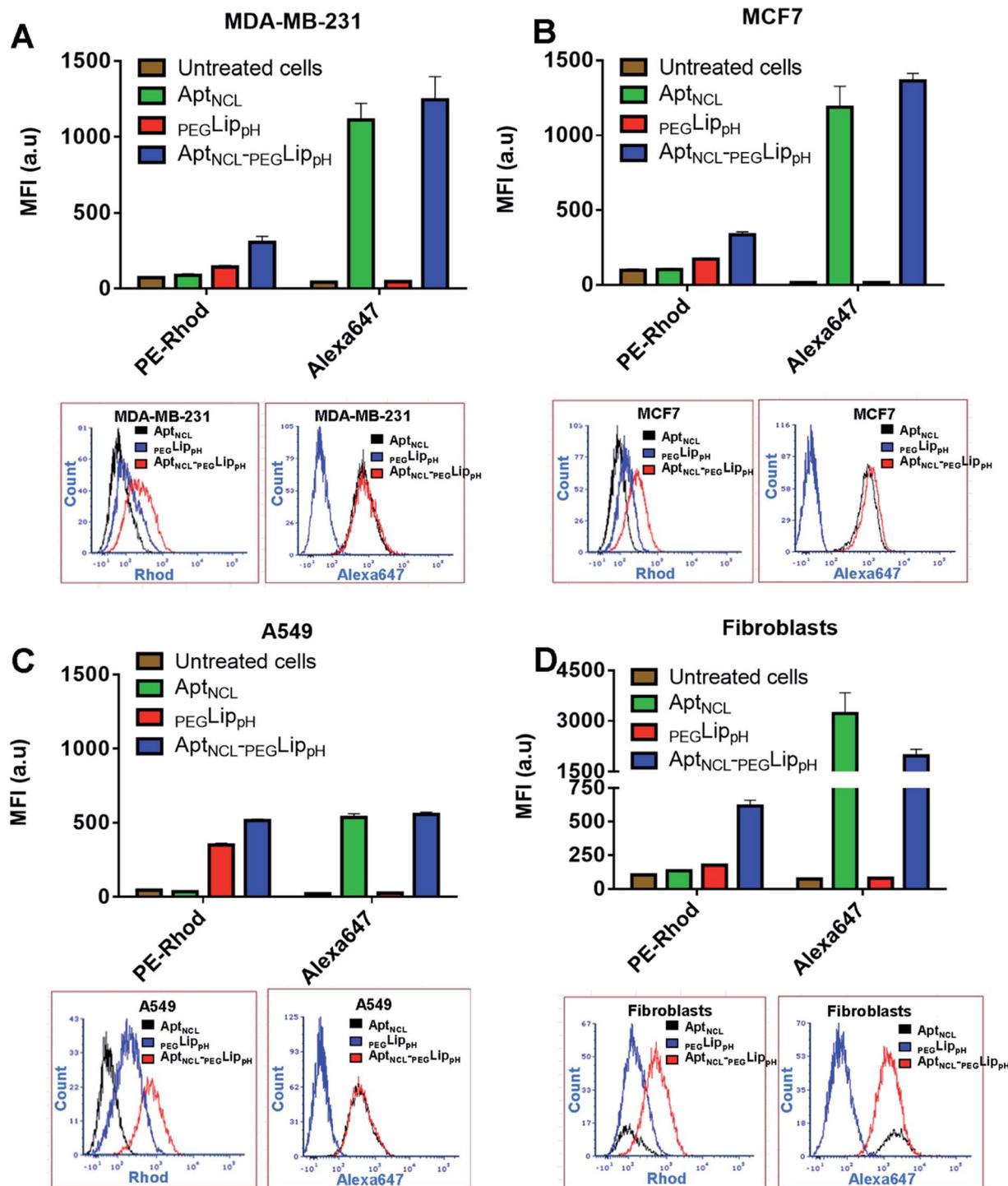


Fig. 3 Flow cytometry analyses for the uptake of free Apt_{NCL}, PEG_{Lip}_{PH}, and Apt_{NCL}-PEG_{Lip}_{PH} in cultured cells; for each cell line (A) MDA-MB-231, (B) MCF7 (C) A549, (D) fibroblasts; (upper part of A, B, C, D) show the mean fluorescence intensity (MFI) measurements after treatment with free Apt_{NCL}, PEG_{Lip}_{PH}, and Apt_{NCL}-PEG_{Lip}_{PH}. (Lower part of A, B, C, D) show flow cytometry histogram analysis after treatment with free Apt_{NCL}, PEG_{Lip}_{PH}, and Apt_{NCL}-PEG_{Lip}_{PH}. Cells were treated with free Apt_{NCL} (200 nM), or PEG_{Lip}_{PH} (120 mM), or Apt_{NCL}-PEG_{Lip}_{PH} (200 nM Apt_{NCL}/120 mM PEG_{Lip}_{PH}) and compared to untreated cells. Apt_{NCL} labeled with Alexa647 and PEG_{Lip}_{PH} labeled with rhodamine. (mean \pm SD, $n = 3$).

encapsulation efficiency. The average size of PEG_{Lip}_{PH} was 134 ± 10 nm and was 120 ± 16 nm for PEG_{Lip}_{PH}-EC γ CD with no significant differences. Both liposome preparations showed a similar polydispersity index (~ 0.1), which demonstrates the presence of a monodisperse distribution. The zeta potential of

PEG_{Lip}_{PH} was -12 ± 2 mV with values similar to the one observed for PEG_{Lip}_{PH}-EC γ CD (-15 ± 2 mV). The encapsulation efficiency of echinomycin was found to be around $6.3 \pm 0.4\%$ and a drug loading of 0.03% (wt/wt), which is similar to our previous work (5.1 ± 0.8).⁹



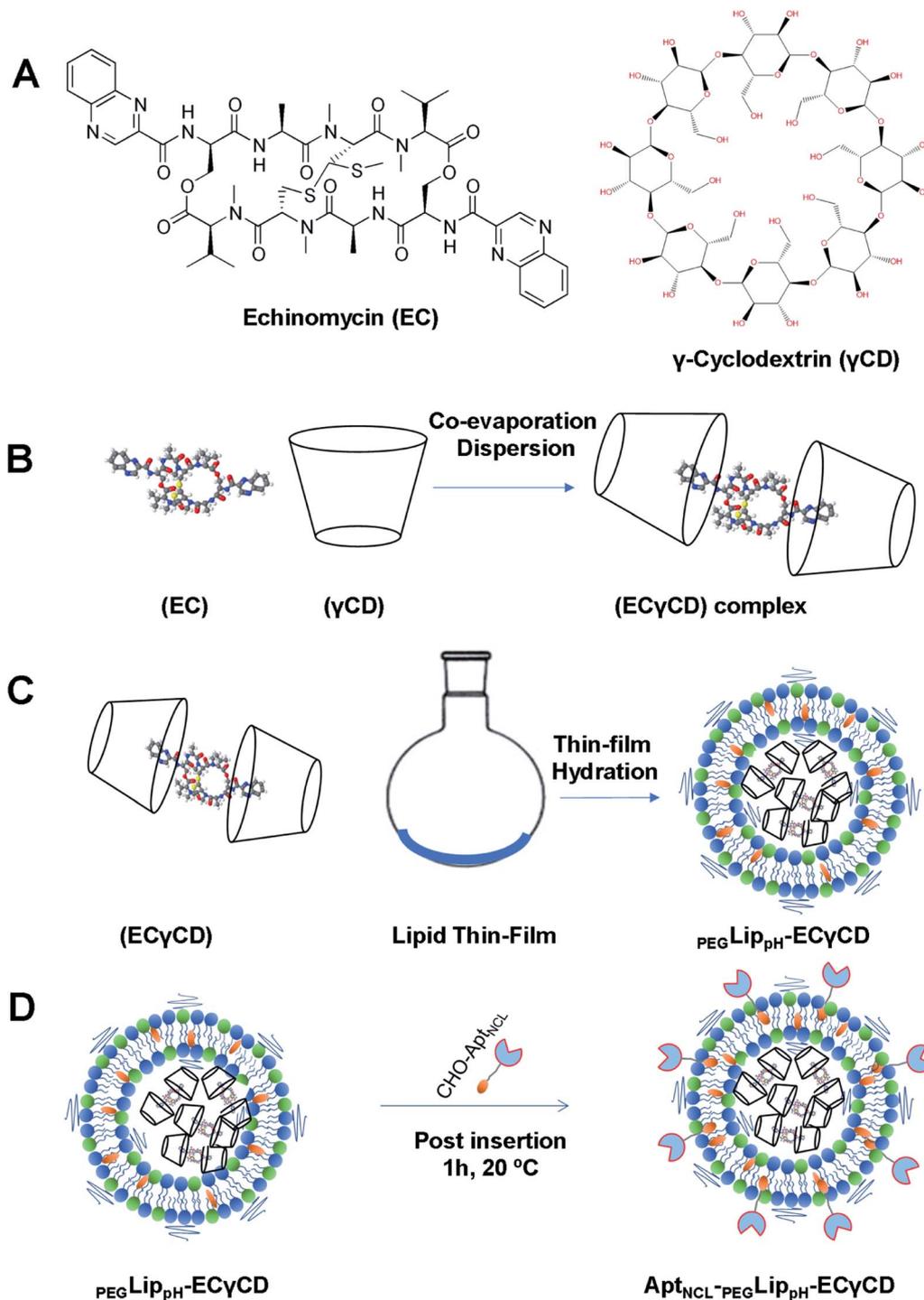


Fig. 4 Schematic representation of (A) the chemical structure of echinomycin and γ -cyclodextrin (EC and γ CD) (B) formation of EC γ CD inclusion complexes (C) encapsulation of EC γ CD complex into PEG Lip_{pH} nanoparticles (D) decoration of PEG Lip_{pH}-EC γ CD with Apt_{NCL} by post-insertion at room temperature for 1 h.

3.6. *In vitro* releasing assay of EC from PEG Lip_{pH}-EC γ CD and Apt_{NCL}-PEG Lip_{pH}-EC γ CD

The *in vitro* release of echinomycin from PEG Lip_{pH}-EC γ CD and Apt_{NCL}-Lip_{pH}-EC γ CD was assessed in physiological PBS buffer (pH 7.4 and pH 5) at 37 °C over a period of 12 h at different intervals.

As illustrated in Fig. 5B, echinomycin release from PEG Lip_{pH}-EC γ CD was higher than Apt_{NCL}-PEG Lip_{pH}-EC γ CD at both pHs. This can be explained by the fact that the Apt_{NCL} was post inserted and immobilized onto the surface of PEGylated Lip_{pH} utilizing CHO as anchoring units. Extra CHO molecules increase the rigidity of the vesicles while the presence of DSPE-



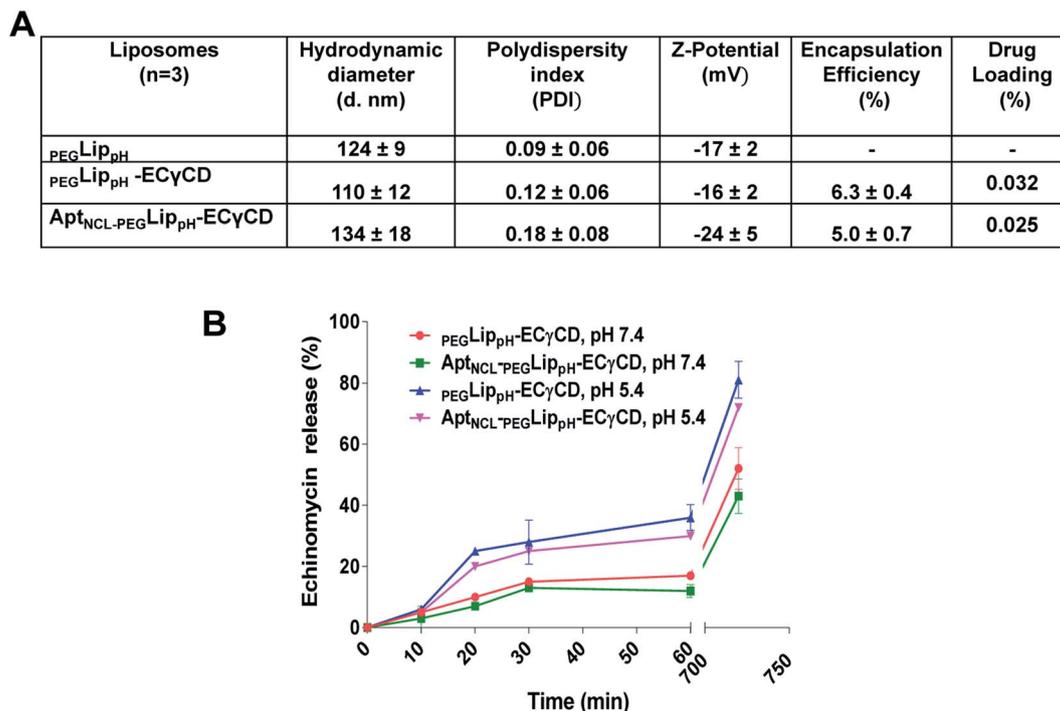


Fig. 5 (A) Characterization using DLS measurements, drug loading, and encapsulation efficiency for the PEG_{pH}Lip_{pH}, PEG_{pH}Lip_{pH}-ECyCD, and Apt_{NCL}-PEG_{pH}Lip_{pH}-ECyCD liposomes before and after Apt_{NCL} post-insertion. (B) Calcein dye releasing assay of Lip_{pH} formulation before and after CHO-Apt_{NCL} post insertion at pH (7.4 and 5.4).

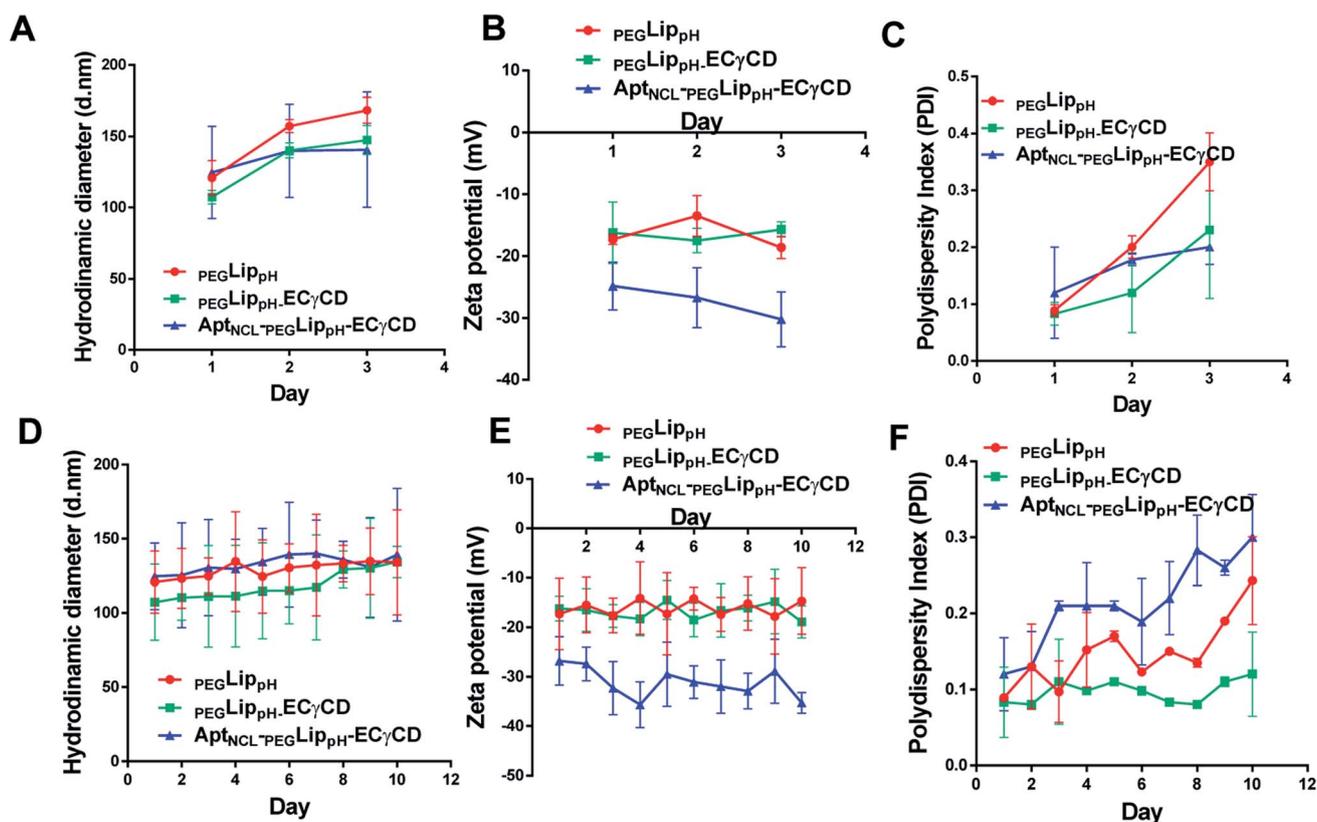


Fig. 6 Colloidal stability of blank Lip_{pH}, Lip_{pH}-ECyCD and Apt_{NCL}-Lip_{pH}-ECyCD in terms of DLS measurements at 37 °C for three days (A) the hydrodynamic diameter, (B) zeta potential and (C) polydispersity index, and at 4 °C for ten days (D) the hydrodynamic diameter, (E) zeta potential and (F) polydispersity index, (mean ± SD, n = 3).



PEG₂₀₀₀ decreases the release by decreasing the pH sensitivity of DOPE.⁵⁰ Moreover, the aptamers post-inserted to the surface of liposomes may decrease the pH sensitivity of the PEGLip_{pH}-EC γ CD. In general, these findings confirm the pH sensitivity for Apt_{NCL}-PEGLip_{pH}-EC γ CD, which might be beneficial for the drug delivery in the tumor microenvironment.

3.7. Colloidal stability for PEGLip_{pH}-EC γ CD and Apt_{NCL}-PEGLip_{pH}-EC γ CD

The stability of blank PEGLip_{pH}, PEGLip_{pH}-EC γ CD, and Apt_{NCL}-PEGLip_{pH}-EC γ CD was investigated by measuring the changes in the mean size and polydispersity index (PDI) over a period of three days in physiological buffer at 37 °C (physiological temperature) (Fig. 6A–C) and at 4 °C (storage temperature) (Fig. 6D–F).

The results showed acceptable stability of PEGLip_{pH}, PEGLip_{pH}-EC γ CD, and Apt_{NCL}-PEGLip_{pH}-EC γ CD at tested temperatures as indicated by minimal changes in the mean size, zeta potential, and PDI.

The lipid bilayer of liposomes grafted with PEG₂₀₀₀ and Apt_{NCL} was simulated to investigate the stability, distribution, and packing of lipids. The top view, side view, and tilt of the bilayer for the primary sheet composed of different types of lipids showed stable and uniform distribution of DPPC, DOPE, cholesterol, DSPE-PEG₂₀₀₀, and CHO-Apt_{NCL} (Fig. 7A–C), respectively.

3.8. Cytotoxicity study

MTT assay was performed to investigate the cytotoxic effects of free EC, PEGLip_{pH}-EC γ CD, and Apt_{NCL}-PEGLip_{pH}-EC γ CD in terms

of the absolute IC₅₀ after 72 h treatment incubation for MDA-MB-231, MCF7, A549, and normal fibroblast cells.

Fig. 8A–D represents the viability dose–response curve and the absolute IC₅₀ of free EC, PEGLip_{pH}-EC γ CD, and Apt_{NCL}-PEGLip_{pH}-EC γ CD for 72 h treatment incubation of MDA-MB-231, A549, MCF7 cancer cell lines, and fibroblast cells, respectively. The cytotoxicity results showed a higher anti-proliferative effect for Apt_{NCL}-PEGLip_{pH}-EC γ CD compared to PEGLip_{pH}-EC γ CD and were 1.5, 2, and 5 folds for A549, MCF7, and MDA-MB-231, respectively, after 72 h treatment incubation Fig. 8E.

Moreover, Fig. 8A–D showed the selective localization of Apt_{NCL}-PEGLip_{pH}-EC γ CD *versus* PEGLip_{pH}-EC γ CD after adding the treatment for 3 h to allow binding of EC loaded nanoparticles to the cell surface and to be internalized into the cell cytoplasm. The remaining free treatment was removed, and the cells were incubated for 72 h.

These results were consistent with the previously described cellular uptake study of unloaded Apt_{NCL}-PEGLip_{pH} and PEGLip_{pH} (≥ 2 folds for all cells) using flow cytometry. However, selectivity values in terms of cell viability were higher, which reflects the high potency of echinomycin drug.

The IC₅₀ values of free EC and PEGLip_{pH}-EC γ CD were around 1 nM for all investigated cancer cell lines and were around 6 nM for fibroblast cells. The IC₅₀ for PEGLip_{pH}-EC γ CD and Apt_{NCL}-PEGLip_{pH}-EC γ CD were about 7.5 nM for the fibroblast cells. Furthermore, the IC₅₀ of Apt_{NCL}-PEGLip_{pH}-EC γ CD for fibroblast was higher than PEGLip_{pH}-EC γ CD IC₅₀ by 1.5 folds (Fig. 8E)

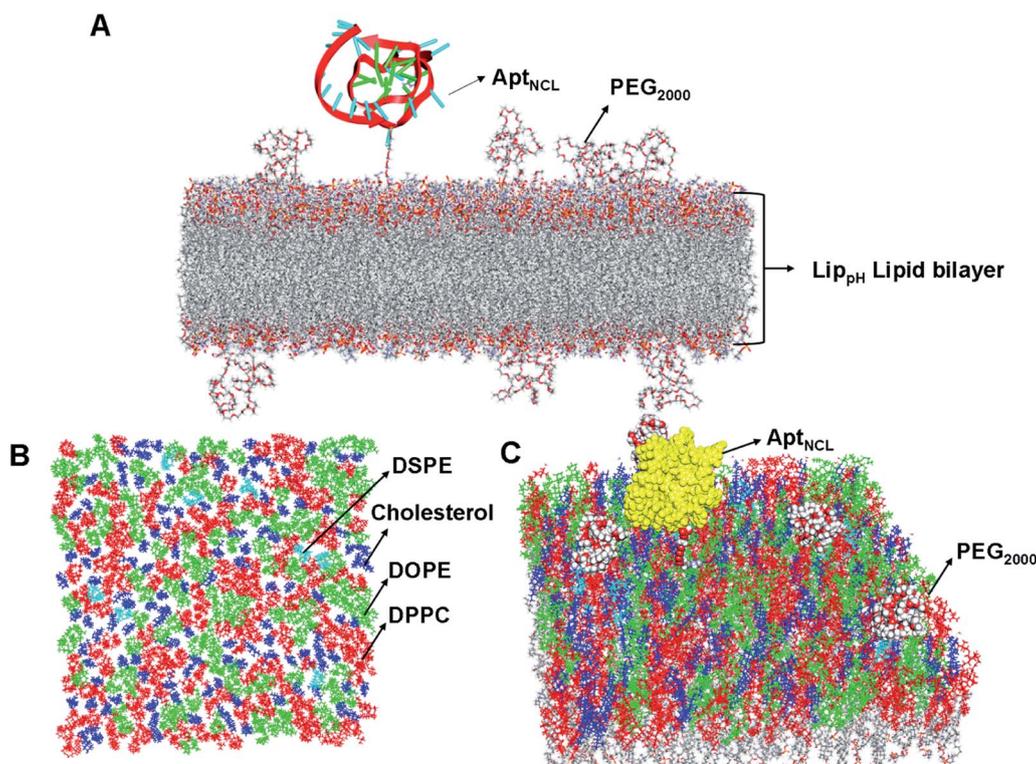


Fig. 7 Distribution and packing of different lipids (DPPC, DOPE, DSPE, and CHO), PEG₂₀₀₀, and Apt_{NCL} in the computationally designed bilayer. (A) The top view of the bilayer for the primary sheet composed of different types of lipids, DPPC, DOPE, cholesterol, and DSPE are shown in red, green, blue, and cyan, respectively. (B) Side view of the bilayer composed of different lipids, PEG₂₀₀₀, and the aptamer. (C) Tilt view of the bilayer after full minimization for the most stable state, the yellow color is used for the aptamer, other color codes as described before.



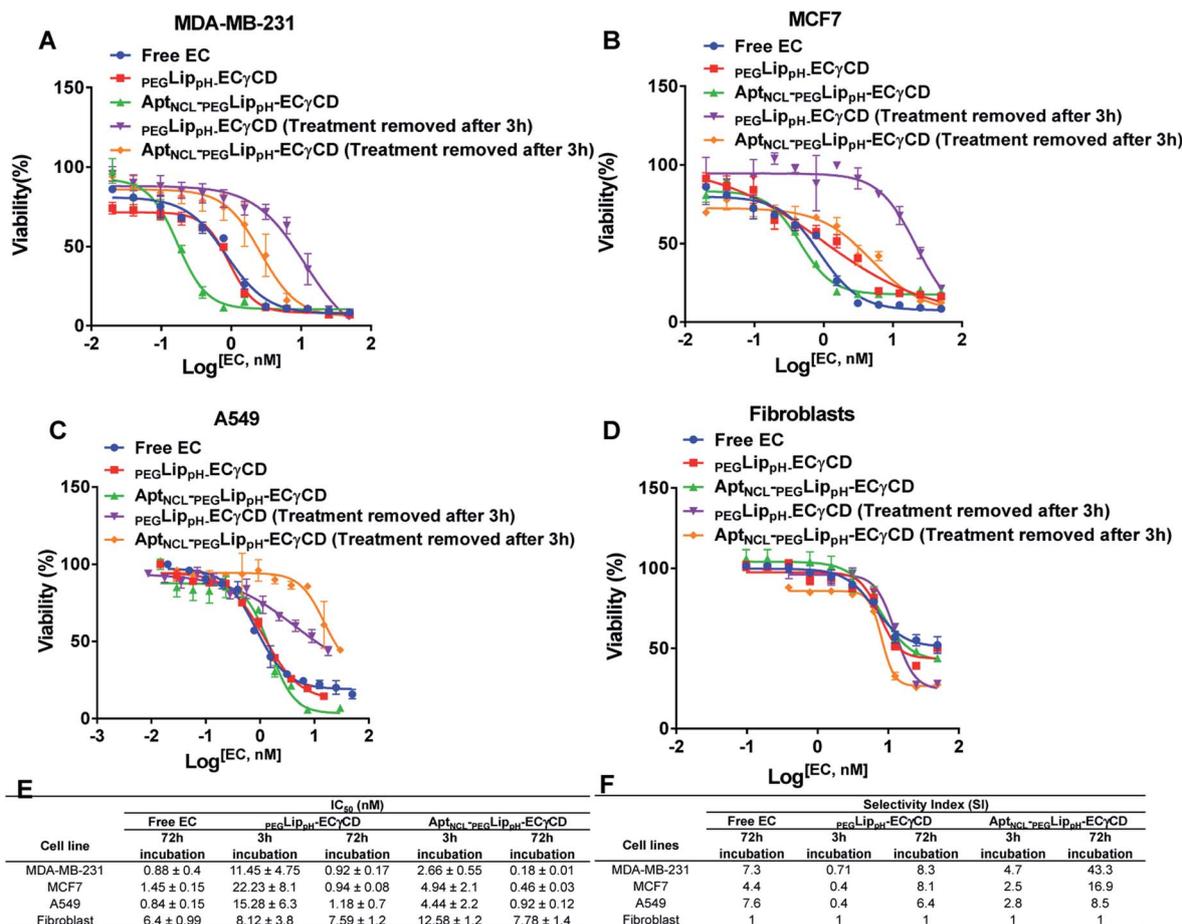


Fig. 8 Viability assay of both breast cancer cells, (A) MDA-MB-231 and (B) MCF-7 (C) small cell lung cancer (A549) and (D) normal cell fibroblast when treated with free EC, Lip_{pH}-EC_γCD and Apt_{NCL}-Lip_{pH}-EC_γCD at different concentrations of EC range from 0 to 50 nM for 72 h and 3 h. (E) and (F) Table of IC₅₀ and selectivity index for the four cell types. (mean ± SD, n = 3).

when treatment was removed after 3 h incubation. These findings are in favor of echinomycin (either free or formulated) as more cytotoxic for cancer cells than normal cells.

It is revealed from Fig. 8E that the *in vitro* cytotoxicity of Apt_{NCL}-PEG-Lip_{pH}-EC_γCD in terms of IC₅₀ against MDA-MB-231, MCF7, A549 were 0.18 ± 0.01, 0.46 ± 0.03 and 0.92 ± 0.12, respectively, compared to PEG-Lip_{pH}-EC_γCD, which were 0.92 ± 0.17, 0.94 ± 0.08 and 1.18 ± 0.7, respectively. The viability results provided a promising potential for selectivity and higher toxicity of Apt_{NCL}-PEG-Lip_{pH}-EC_γCD over non functionalized formulations into cancer cells.

The cytotoxicity results and conclusions are consistent with previous studies. For example, *in vivo* and an *in vitro* study conducted by Xing *et al.*, (2013) showed aptamer-liposomes bind with high affinity and specificity to nucleolin cell surface receptors, which overexpressed on MCF-7.⁵¹ Moreover, Liao *et al.*, (2015) conjugated liposomes encouraged the specific binding of liposomes to the nucleolin receptors, improving the affinity and cellular uptake by tumor cells more than plain liposomes.⁵²

The selectivity index (SI) of anticancer drugs, measures the window between toxicity against normal cells and anticancer activity. The higher the SI ratio, the more effective and safer

a drug during *in vivo* treatment for anti-cancer drugs.⁵³ An SI value greater than 2 indicates that the compound is more than twice cytotoxic to the tumor cell line compared to the normal cell line, while SI < 2 means that the drug has general toxicity.⁵³

Selectivity indices are shown in Fig. 8F for the tested PEG-Lip_{pH}-EC_γCD, and Apt_{NCL}-PEG-Lip_{pH}-EC_γCD against the cancer cell lines (A549, MDA-MB-231, and MCF7) and normal fibroblast cells. The selectivity index (SI) of Apt_{NCL}-PEG-Lip_{pH}-EC_γCD for tumor cell lines compared to the normal cell line after 72 h was 43.3 (MDA-MB-231), 16.9 (MCF7) and 8.5 (A549). Furthermore, SI after 3 h were found 4.7, 2.5, 2.8, respectively. These results confirmed the selectivity of aptamer-guided drug delivery systems, especially SI after 3 h. In addition, the selectivity index (SI) of PEG-Lip_{pH}-EC_γCD for tumor cell lines compared to the normal cell line after 72 h was 8.3 (MDA-MB-231), 8.1 (MCF7), and 6.4 (A549). Furthermore, SI after 3 h were found to be 0.71, 0.4, 0.4, respectively. Based on the SI of PEG-Lip_{pH}-EC_γCD after 72 h, selectivities were comparable for the three cancer cell lines and it is proposed that PEG-Lip_{pH}-EC_γCD is more toxic on cancer cells than on normal cells. Whereas, after 3 h, the SI indicated that PEG-Lip_{pH}-EC_γCD was retained in the cell and then released in a controlled manner



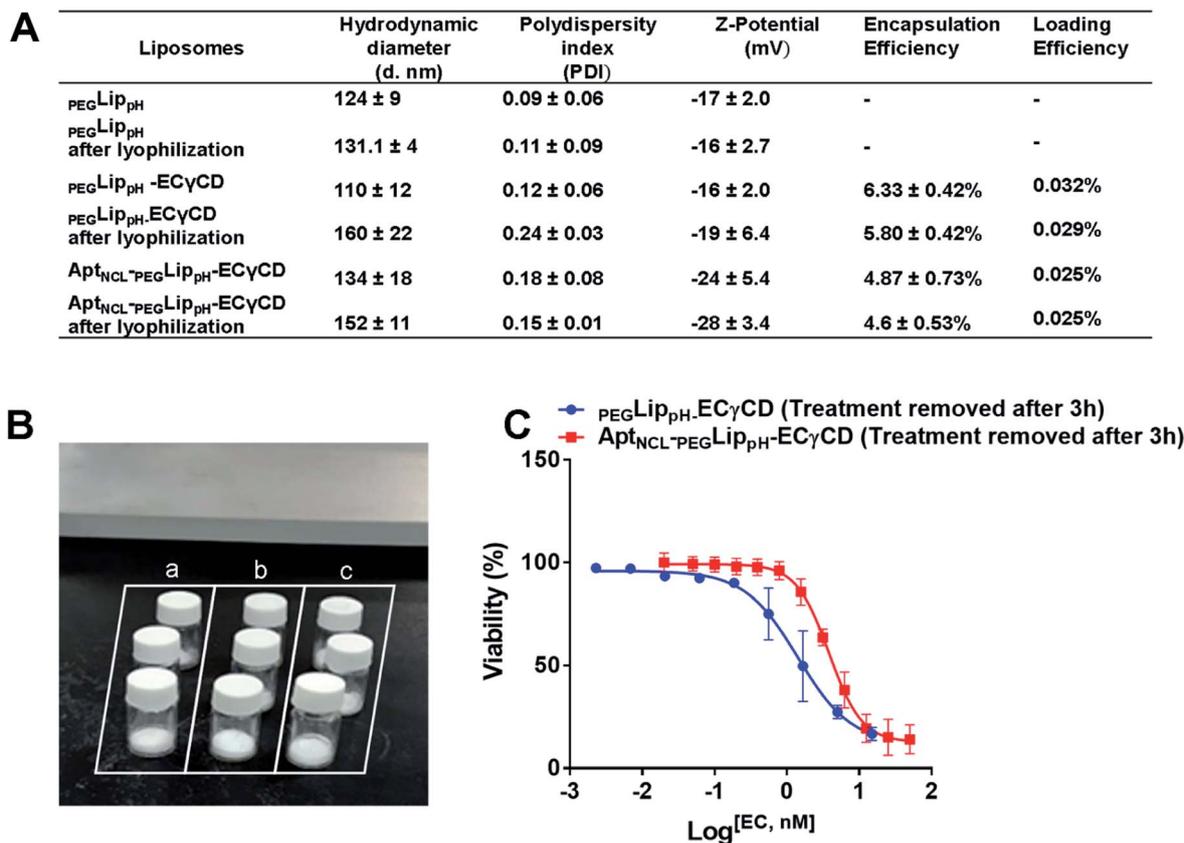


Fig. 9 (A) Table showing the effect of lyophilization on the stability of PEG-Lip_{pH}, PEG-Lip_{pH}-ECγCD, and Apt_{NCL}-PEG-Lip_{pH}-ECγCD in terms of mean hydrodynamic diameter, PDI, and zeta potential, encapsulation efficiency, and drug loading (B) successfully prepared lyophilized liposomes. Labels: a, PEG-Lip_{pH}; b, PEG-Lip_{pH}-ECγCD; and c, Apt_{NCL}-PEG-Lip_{pH}-ECγCD (C) the dose-response curve for MDA-MB-231 breast cancer cells treated with lyophilized Apt_{NCL}-PEG-Lip_{pH}-ECγCD and PEG-Lip_{pH}-ECγCD (treatment was removed after 3 h). (mean ± SD, *n* = 3).

over a longer time. In general, both PEG-Lip_{pH}-ECγCD and Apt_{NCL}-PEG-Lip_{pH}-ECγCD showed higher selectivity toward cancer cells than normal cells.

3.9. Lyophilization of the liposomal formulation

Lyophilization of liposomal formulations is a commonly used method to increase liposomal stability and shelf life. Lyoprotectants such as sucrose, maltose, lactose, and trehalose are needed to prevent liposomal collapse during the lyophilization process and vesicle fusion upon rehydration.⁵⁴ Freshly prepared liposomal suspensions were freeze-dried in the presence of either trehalose or sucrose in ratios; 1%, 2.5%, 5%, and 10%. It is worth mentioning that protected liposomes were significantly lower in size and PDI than unprotected liposomes, which means that both sugars protect liposomes from fusion and become larger in size after rehydration, (Table S1†). Sucrose and trehalose were used as cryoprotectants. Both sugars resulted in stable particle size upon reconstitution. However, sucrose was slightly better than trehalose. This can be explained due to the condition of the freeze dryer, which was used, and the lowest temperature and pressure that can be reached.⁵⁴

Lyophilization of the final formulation was successfully performed using 5% sucrose as a cryoprotectant. This was

illustrated in (Fig. 9A and B). The stability of the final lyophilized formulations was investigated by measuring the changes in the mean size, size distribution (PDI), Z-potential, and EE% blank PEG-Lip_{pH}, PEG-Lip_{pH}-ECγCD, and Apt_{NCL}-PEG-Lip_{pH}-ECγCD after rehydration (Fig. 9A). Moreover, the results showed acceptable stability of both blank and loaded liposomes indicated by minimal changes in the mean size, PDI, and EE%. In addition, PEG-Lip_{pH}-ECγCD and Apt_{NCL}-PEG-Lip_{pH}-ECγCD retained their cytotoxic and targeting functions by measuring their anti-proliferative activity on the MDA-MB-231 breast cancer cell line (Fig. 9C). Indeed, the toxicity of Apt_{NCL}-PEG-Lip_{pH}-ECγCD was 2.7 fold higher than that of PEG-Lip_{pH}-ECγCD.

4. Conclusions

Smart and multifunctional aptamer-guided and PEGylated pH-sensitive liposomes were designed, formulated, and fully characterized. These liposomes were stable at physiological pH and released their payload at low pH. Particle size, size distribution and charge stability were evaluated at storage and physiological temperatures. These smart-multifunctional liposomes were tested for pH sensitivity and release using calcein dye and echinomycin. Both the dye and the potent anticancer antibiotic drug, echinomycin, were successfully encapsulated inside



liposomes. Apt_{NCL-PEG}Lip_{pH-EC-γCD} exhibited excellent selectivity and cytotoxic activity on three cancer cell lines (MCF7, MDA-MB-231, and A549) compared to normal cells. The current aptamer-guided pH-sensitive liposomes can be used as a delivery system for other therapeutic molecules. Moreover, *in vivo* and mechanistic studies can provide valuable information about the antitumor efficacy and more understanding of the mechanism of selective uptake by cancer cells.

Conflicts of interest

The authors declare no conflict of interest.

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References

- 1 K. R. Fox and E. Kentebe, *Nucleic Acids Res.*, 1990, **18**, 1957–1963.
- 2 Y. Wang, Y. Liu, S. N. Malek, P. Zheng and Y. Liu, *Cell Stem Cell*, 2011, **8**, 399–411.
- 3 Y.-S. Park, W.-S. Shin and S.-K. Kim, *J. Antimicrob. Chemother.*, 2008, **61**, 163–168.
- 4 M. Sato, T. Nakazawa, Y. Tsunematsu, K. Hotta and K. Watanabe, *Curr. Opin. Chem. Biol.*, 2013, **17**, 537–545.
- 5 M. M. Van Dyke and P. B. Dervan, *Science*, 1984, **225**, 1122–1127.
- 6 W. J. Gradishar, N. J. Vogelzang, L. J. Kilton, S. J. Leibach, A. W. Rademaker, S. French and A. B. Benson, *Invest. New Drugs*, 1995, **13**, 171–174.
- 7 H. Shelley and R. J. Babu, *J. Pharm. Sci.*, 2018, **107**, 1741–1753.
- 8 T. Loftsson and D. Duchêne, *Int. J. Pharm.*, 2007, **329**, 1–11.
- 9 W. Alshaer, M. Zraikat, A. Amer, H. Nsairat, Z. Lafi, D. A. Alqudah, E. Al Qadi, T. Alsheleh, F. Odeh and A. Alkaraki, *RSC Adv.*, 2019, **9**, 30976–30988.
- 10 I.-Y. Kim, Y.-S. Kang, D. S. Lee, H.-J. Park, E.-K. Choi, Y.-K. Oh, H.-J. Son and J.-S. Kim, *J. Controlled Release*, 2009, **140**, 55–60.
- 11 Y. Kumar, K. Kuche, R. Swami, S. S. Katiyar, D. Chaudhari, P. B. Katare, S. K. Banerjee and S. Jain, *Int. J. Pharm.*, 2020, **573**, 118889.
- 12 K. Jabin, Z. Husian, M. A. Zahid and P. Kushwaha, *World J. Pharm. Pharm. Sci.*, 2018, **7**, 1307–1319.
- 13 U. Bulbake, S. Doppalapudi, N. Kommineni and W. Khan, *Pharmaceutics*, 2017, **9**, 12.
- 14 C. Chen, D. Han, C. Cai and X. Tang, *J. Controlled Release*, 2010, **142**, 299–311.
- 15 M. R. Rao and L. S. Babrekar, *Indian J. Pharm. Sci.*, 2018, **80**, 1115–1124.
- 16 Y. Ito, *Photochemistry for Biomedical Applications: From Device Fabrication to Diagnosis and Therapy*, Springer, 2018.
- 17 M. Waring and L. Wakelin, *Nature*, 1974, **252**, 653–657.
- 18 H. Sun, X. Zhu, P. Y. Lu, R. R. Rosato, W. Tan and Y. Zu, *Mol. Ther. – Nucleic Acids*, 2014, **3**, e182.
- 19 W. Alshaer, H. Hillaireau and E. Fattal, *Adv. Drug Delivery Rev.*, 2018, **134**, 122–137.
- 20 Y. Lee and D. H. Thompson, *Wiley Interdiscip. Rev.: Nanomed. Nanobiotechnol.*, 2017, **9**, e1450.
- 21 C.-J. Chu and F. C. Szoka, *J. Liposome Res.*, 1994, **4**, 361–395.
- 22 D. Collins, *Liposomes as tools in basic research and industry*, 1995, vol. 13, pp. 201–214.
- 23 Y. Chen, Q. Du, Y. Zou, Q. Guo, J. Huang, L. Tao, X. Shen and J. Peng, *Int. J. Pharm.*, 2020, **584**, 119446.
- 24 R. M. Sawant, J. Hurley, S. Salmaso, A. Kale, E. Tolcheva, T. Levchenko and V. Torchilin, *Bioconjugate Chem.*, 2006, **17**, 943–949.
- 25 R. R. Sawant and V. P. Torchilin, *Soft Matter*, 2010, **6**, 4026–4044.
- 26 A. U. Rehman, Z. Omran, H. Anton, Y. Mély, S. Akram, T. F. Vandamme and N. Anton, *Eur. J. Pharm. Biopharm.*, 2018, **133**, 331–338.
- 27 A. Akbarzadeh, R. Rezaei-Sadabady, S. Davaran, S. W. Joo, N. Zarghami, Y. Hanifehpour, M. Samiei, M. Kouhi and K. Nejati-Koshki, *Nanoscale Res. Lett.*, 2013, **8**, 1–9.
- 28 M. S. Franco, C. A. Silva, E. A. Leite, J. N. Silveira, C. S. Teixeira, V. N. Cardoso, E. Ferreira, G. D. Cassali, A. L. B. de Barros and M. C. Oliveira, *J. Drug Delivery Sci. Technol.*, 2021, 102400.
- 29 S. J. Routledge, J. A. Linney and A. D. Goddard, *Biochem. Soc. Trans.*, 2019, **47**, 919–932.
- 30 A. Sharma and U. S. Sharma, *Int. J. Pharm.*, 1997, **154**, 123–140.
- 31 T. Ishida, Y. Okada, T. Kobayashi and H. Kiwada, *Int. J. Pharm.*, 2006, **309**, 94–100.
- 32 M. Z. Lai, W. J. Vail and F. C. Szoka, *Biochemistry*, 1985, **24**, 1654–1661.
- 33 S. Ghanbarzadeh, A. Khorrami, L. Mohamed Khosroshahi and S. Arami, *Pharm. Biol.*, 2014, **52**, 848–854.
- 34 W. Alshaer, H. Hillaireau, J. Vergnaud, S. Ismail and E. Fattal, *Bioconjugate Chem.*, 2015, **26**, 1307–1313.
- 35 A. D. Bangham and R. Horne, *J. Mol. Biol.*, 1964, **8**, 660.
- 36 F. Odeh, S. I. Ismail, R. Abu-Dahab, I. S. Mahmoud and A. Al Bawab, *Drug Delivery*, 2012, **19**, 371–377.
- 37 A. Laouini, C. Jaafar-Maalej, I. Limayem-Blouza, S. Sfar, C. Charcosset and H. Fessi, *J. Colloid Sci. Biotechnol.*, 2012, **1**, 147–168.
- 38 H. Nsairat, I. S. Mahmoud, F. Odeh, D. Abuarqoub, H. Al-Azzawi, R. Zaza, M. I. Qadri, S. Ismail, A. Al Bawab, A. Awidi and W. Alshaer, *RSC Adv.*, 2020, **10**, 36219–36229.
- 39 W. Alshaer, H. Hillaireau, J. Vergnaud, S. Mura, C. Deloménie, F. Sauvage, S. Ismail and E. Fattal, *J. Controlled Release*, 2018, **271**, 98–106.
- 40 H. Karanth and R. S. Murthy, *J. Pharm. Pharmacol.*, 2007, **59**, 469–483.
- 41 C. Hamai, T. Yang, S. Kataoka, P. S. Cremer and S. M. Musser, *Biophys. J.*, 2006, **90**, 1241–1248.
- 42 I. R. Cooke and M. Deserno, *Biophys. J.*, 2006, **91**, 487–495.
- 43 P. R. Cullis and B. De Kruijff, *Biochim. Biophys. Acta*, 1978, **507**, 207–218.



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- 44 M.-L. Briuglia, C. Rotella, A. McFarlane and D. A. Lamprou, *Drug Delivery Transl. Res.*, 2015, **5**, 231–242.
- 45 S. Kaddah, N. Khreich, F. Kaddah, C. Charcosset and H. Greige-Gerges, *Food Chem. Toxicol.*, 2018, **113**, 40–48.
- 46 C. Demetzos, *J. Liposome Res.*, 2008, **18**, 159–173.
- 47 M. L. Immordino, F. Dosio and L. Cattell, *Int. J. Nanomed.*, 2006, **1**, 297.
- 48 E. M. Reyes-Reyes, Y. Teng and P. J. Bates, *Cancer Res.*, 2010, **70**, 8617–8629.
- 49 E. M. Reyes-Reyes, F. R. Salipur, M. Shams, M. K. Forsthoefel and P. J. Bates, *Mol. Oncol.*, 2015, **9**, 1392–1405.
- 50 S. Simoes, J. N. Moreira, C. Fonseca, N. Duzgunes and M. C. de Lima, *Adv. Drug Delivery Rev.*, 2004, **56**, 947–965.
- 51 H. Xing, L. Tang, X. Yang, K. Hwang, W. Wang, Q. Yin, N. Y. Wong, L. W. Dobrucki, N. Yasui, J. A. Katzenellenbogen, W. G. Helderich, J. Cheng and Y. Lu, *J. Mater. Chem. B*, 2013, **1**, 5288–5297.
- 52 Z. X. Liao, E. Y. Chuang, C. C. Lin, Y. C. Ho, K. J. Lin, P. Y. Cheng, K. J. Chen, H. J. Wei and H. W. Sung, *J. Controlled Release*, 2015, **208**, 42–51.
- 53 C. Bézivin, S. Tomasi, F. Lohézic-Le Dévéhat and J. Boustie, *Phytomedicine*, 2003, **10**, 499–503.
- 54 S. Franze, F. Selmin, E. Samaritani, P. Minghetti and F. Cilurzo, *Pharmaceutics*, 2018, **10**, 139.

