


Cite this: *RSC Adv.*, 2021, 11, 35754

Preparation of a novel bridged bis(β -cyclodextrin) chiral stationary phase by thiol–ene click chemistry for enhanced enantioseparation in HPLC†

Ning Zhang, Siyu Guo and Bolin Gong *

A bridged bis(β -cyclodextrin) ligand was firstly synthesized via a thiol–ene click chemistry reaction between allyl-ureido- β -cyclodextrin and 4-4'-thiobis(phenol), which was then bonded onto a 5 μ m spherical silica gel to obtain a novel bridged bis(β -cyclodextrin) chiral stationary phase (HTCDP). The structures of HTCDP and the bridged bis(β -cyclodextrin) ligand were characterized by the ^1H nuclear magnetic resonance (^1H NMR), solid state ^{13}C nuclear magnetic resonance (^{13}C NMR) spectra spectrum, scanning electron microscope, elemental analysis, mass spectrometry, infrared spectrometry and thermogravimetric analysis. The performance of HTCDP in enantioseparation was systematically examined by separating 21 chiral compounds, including 8 flavanones, 8 triazole pesticides and 5 other common chiral drugs (benzoin, praziquantel, 1-1'-bi-2-naphthol, Tröger's base and bicalutamide) in the reversed-phase chromatographic mode. By optimizing the chromatographic conditions such as formic acid content, mobile phase composition, pH values and column temperature, 19 analytes were completely separated with high resolution (1.50–4.48), in which the enantiomeric resolution of silymarin, 4-hydroxyflavanone, 2-hydroxyflavanone and flavanone were up to 4.34, 4.48, 3.89 and 3.06 within 35 min, respectively. Compared to the native β -CD chiral stationary phase (CDCSP), HTCDP had superior enantiomer separation and chiral recognition abilities. For example, HTCDP completely separated 5 other common chiral drugs, 2 flavanones and 3 triazole pesticides that CDCSP failed to separate. Unlike CDCSP, which has a small cavity (0.65 nm), the two cavities in HTCDP joined by the aryl connector could synergistically accommodate relatively bulky chiral analytes. Thus, HTCDP may have a broader prospect in enantiomeric separation, analysis and detection.

Received 17th June 2021
Accepted 25th October 2021

DOI: 10.1039/d1ra04697g

rsc.li/rsc-advances

1. Introduction

Many studies have shown that the enantiomers of chiral compounds tend to demonstrate obvious differences in biological activity, and pharmacological and toxicological effects,^{1–3} which makes enantioseparation a hot research topic in food chemistry, the pharmaceutical industry and environmental protection.^{4,5} So far, conventional enantiomer resolution methods include chemical resolution, enzyme kinetic resolution, high-performance liquid chromatography (HPLC),⁶ gas chromatography,⁷ and capillary electrochromatography.⁸ Among them, HPLC is widely used because of its high sensitivity, reproducibility, rapid separation speed, large column capacity, and broad applicability. At present, various chiral stationary phases (CSPs) combined with HPLC have been proved to be one of the most effective methods for direct

separation of enantiomers.⁹ Therefore, the development of multifunctional and efficient CSPs is the key to improving the enantiomeric resolution. β -cyclodextrin (β -CD) is a mesa-shaped oligosaccharide formed by connecting 7 D-glucopyranose units via α -1,4 glycosidic bonds. The unique cavity structure of β -CD allows the formation of inclusion complexes with chiral analytes, which has been proved to be one of most important roles in driving enantioseparation.¹⁰ Additionally, the introduction of various substituents through the functionalization of the hydroxyl group on β -CD can also promote the separation of enantiomers. For example, many interactions including hydrogen bonding, π - π conjugation, dipole-dipole, steric repulsion, and static electricity can be introduced through functionalization of the hydroxyl group on β -CD.^{11–14} The introduction of these interactions enables β -CD CSP to enhance the identification and separation of chiral analytes. Therefore, various types of β -CD CSPs are widely prepared and used in chiral separations. Moreover, compared with vancomycin,¹⁵ teicoplanin,¹⁶ cellulose,¹⁷ amylos¹⁸ and polymer CSPs,¹⁹ such multi-functional enantiomeric separation mode is also an important reason why β -CD CSPs have received widespread attention and application.

School of Chemistry and Chemical Engineering, North Minzu University, No. 204 Wenchang North Street, Xixia District, Yinchuan 750021, China. E-mail: gongbolin@163.com

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d1ra04697g



However, the separation ability and stability of β -CD CSPs are significantly affected by the functionalization of its hydroxyl groups and bond chemistry on silica gel.^{20,21} β -CD hydroxyl derivatization includes single,²² partial,¹¹ and complete²³ derivatization. Single-derivatized β -CD CSPs have a simple structure and stable chromatographic performance but limited ability to separate chiral enantiomers.¹⁷ By comparison, partially derivatized β -CD CSPs show improved enantioseparation but the number and position of the derived substituents are difficult to control, resulting in poor chromatographic reproducibility.²⁴ Fully derivatized β -CD CSPs have more functional sites but too many substituents inevitably reduce the inclusion capacity of the β -CD cavity.²⁵ Shuang *et al.*²⁶ prepared a novel of single-derivatized β -CD CSP and separated 9 triazole pesticides. However, the resolution was not high, which may be caused by the low degree of derivatization. Armstrong research group^{27–29} prepared a series of fully derivatized β -CD CSPs. The enantioselectivity of β -CD CSPs was improved through the introduction of carbamate linkages, and successfully resolved chiral alcohols, amino acids and chiral amines. However, fully derivatized β -CD CSPs could not separate crown ether and dansyl amino acid enantiomers, which could be separated on natural β -CD CSPs. According to Stalcup and Chang,³⁰ the fully derivatized β -CD CSPs may be due to the high degree of derivatization blocking the cavity of β -CD. Moreover, the separation of enantiomers is an extremely complex and delicate process that requires multiple recognition sites to cooperate with each other,³¹ with the inclusion effects and hydrogen bonding playing important roles.³² Therefore, it may not be sufficient to rely solely on the functionalization of the hydroxyl group on β -cyclodextrin to improve the enantiomeric selectivity. In recent years, the emergence of bridged β -CD CSPs is a very promising candidate.³³

The Bridged bis(β -CD) is a supramolecular dimer connected by 2 single-derived CD units that has been widely used for molecular recognition and as a targeted drug carrier, photochemical, and asymmetric catalyst^{34–37} because of its greater capacity for synergistic inclusion and molecular recognition compared to single β -CD.³⁸ In recent years, although the bridged β -CD has been widely used in many fields, the CSPs used for the separation of enantiomers are rarely reported. Wang's research group prepared a series of triazolyl-bridged bis(β -CD) CSPs and successfully separated enantiomers such as dansyl amino acids, aryl carboxylic acids and isoxazoline with high resolution.^{39,40} Yuan's research group prepared two bridged β -CD CSPs and separated 4 aromatic compound positional isomers and 5 amino acid enantiomers.⁴¹ Li's research group prepared a series of bridged bis(β -CD) CSPs and separated β -blockers, flavanones, triazoles, dansyl amino acids and other enantiomers with high resolution.^{42–44} Obviously, bridged β -CD CSPs have potential development prospects in improving the resolution of enantiomers. Liu and Chen⁴⁵ also found that the bridged spacer arm of the bridged bis(β -CD) can be used as a pseudo-cavity to provide additional sites of action, thereby improving the ability of molecular recognition. Moreover, thiol-ene click chemistry has been used to bridge spacer arms

between mono- β -CD and silica gel due to its advantages of mild reaction conditions, fast reaction speed, and high yield.⁴⁶

Herewith, we prepared a new bridged bis(β -cyclodextrin) chiral stationary phase (HTCDP) by thiol-ene click chemistry, applying the HTCDP's synergistic inclusion, hydrogen bonds and π - π interaction to the enantiomeric separation of chiral compounds at the same. The chromatographic performance of HTCDP was comprehensively evaluated by using 21 chiral drugs and pesticides as probes, and the chromatographic conditions affecting the enantioseparation were optimized. The results demonstrate that the enantioselectivity of HTCDP was significantly better than that of CDCSP.

2. Experimental

2.1. Materials

Mono-(6-amino-6-deoxy)- β -cyclodextrin (NH_2 - β -CD, purity $\geq 98\%$) was purchased from Shandong Binzhou Zhiyuan Biotechnology Co (Binzhou, China). 4,4'-Thiobisbenzenethiol (purity $\geq 98\%$), 2,2'-azobis(2-methylpropionitrile) (AIBN, purity $\geq 98\%$), 3-isocyanatopropyltriethoxysilane (purity $\geq 98\%$), formic acid (FA), acetic acid (HOAc), and triethylamine (TEA) were purchased from Shanghai Aladdin Bio-Chem Technology Co (Shanghai, China). Allyl isocyanate (purity $\geq 98\%$) was obtained from Alfa Aesar (Tewksbury, MA, USA). Pyridine and acetone were provided by Sinopharm Chemical Reagent Co (Shanghai, China). Methanol (MeOH) and acetonitrile (ACN) (HPLC grade) purchased from Tedia (Shanghai, China). Sephadex C-25 was provided by Rui Da Heng Hui Science Technology Development Co (Beijing, China). Triazole racemic standards (purity $\geq 98\%$) were purchased from Shanghai Pesticide Research Institution (Shanghai, China). Flavanones, benzoin, praziquantel, 1,1-bi-2-naphthol, Tröger's base, and bicalutamide racemic standards were obtained from Sigma-Aldrich (St. Louis, MO, USA). Spherical silica gel (5 μm , 25 nm) was provided by Lanzhou Institute of chemical physics (Lanzhou, China). Pyridine was treated with calcium hydride and distilled before use. The water was of ultrapure grade.

2.2. Apparatus

All chiral analytes were separated on an LC-20AT high-performance liquid chromatograph (Shimadzu, Kyoto, Japan) equipped with an SPD-20A UV-VIS detector (Shimadzu). The ^{13}C NMR spectra were collected on Bruker Advance III WB 400M (Bruker Daltonics, Bremen, Germany). The ^1H NMR spectrum was collected on the AvanceII 400 MHz NMR spectrometer (Bruker, Switzerland). Scanning electron micrograph were obtained using Zeiss Sigma 300 field emission scanning electron microscopy (Zeiss, Germany). Thermogravimetric analysis was performed on an STA 449C synchronous thermogravimetric analyzer (Netzsch, Selb, Germany). Mass spectrometry (MS) analysis was performed on a 6540 TOF high-resolution mass spectrometer (Agilent, Santa Clara, CA, USA). C, H, N, and S contents were determined using a VarioEL III element analyzer (Elementar Analysensysteme GmbH, Langenselbold, Germany). IR analysis was performed on an FTIR-8400S Fourier Transform



IR Spectrometer (Shimadzu). The synthesized bridged bis(β -CD) chiral ligand was packed using a CGY-100B chromatographic column packing machine (Beijing Fusiyan Machinery Processing Department, Beijing, China). Ultrapure water was obtained using the VE-AS water purification system (Shenzhen Vamia Environmental Protection Co, Shenzhen, China).

2.3. Preparation of HTCDP and CDCSP

The synthetic route of HTCDP was displayed in Fig. 1. NH_2 - β -CD (2 g, 1.76 mmol) was dissolved in 40 mL of anhydrous pyridine. The anhydrous pyridine solution of allyl isocyanate (10 mL anhydrous pyridine contained 0.234 g allyl isocyanate) was slowly dripped at ice bath temperature (0–5 °C) and stirred for 30 min, then the mixed solution was reacted at 80 °C for 24 h under the protection of nitrogen. At the end of the reaction, the unreacted anhydrous pyridine and allyl isocyanate were evaporated by vacuum distillation, and a white precipitate was obtained by adding acetone to the residual liquid. The precipitate was purified with a Sephadex C-25 column and dried under vacuum at 50 °C for 24 h to obtain 2.09 g of allyl-ureido- β -CD with the yield of 61%. ESI-MS (m/z), $[\text{M} + \text{H}]^+$ was calculated for 1218.48 found at 1218.41 exhibited in Fig. S1a.†

Allyl-ureido- β -CD (2 g, 1.64 mmol), 4,4'-thiobisbenzenethiol (0.103 g), and AIBN (30 mg) were dissolved in 40 mL of anhydrous pyridine with stirring for 30 min, and reacted at 70 °C for 24 h under the protection of nitrogen. After the reaction, part of the anhydrous pyridine was evaporated by distillation under reduced pressure and acetone was added to the remaining liquid to obtain a pale yellow solid. The pale yellow solid was purified with a Sephadex C-25 column and dried under vacuum at 50 °C for 24 h to obtain 1.32 g of 4,4'-thiobis(phenylene) bridged bis(β -CD) with the yield of about 60%. ESI-MS (m/z), $[\text{M}$

$+ \text{H}]^+$ was calculated for 2686.38 found at 2686.35 exhibited in Fig. S1b.† As shown in Fig. S2,† the ^1H NMR results (DMSO- d_6 , 400 MHz, ppm) were found as: δ 6.04 (2H, $-\text{NH}-\text{CO}-\text{NH}$), 5.87–5.50 (28H, OH, 2,3-position of CD), 5.29–4.87 (8H, phenyl), 4.83 (12H, 6-position of CD), 4.69–4.18 (14H, C (1)H of CD), 3.61 (56H, C (3,5,6) H of CD), 3.36–3.14 (28H, C (2,4) H of CD).

4,4'-Thiobis(phenylene) bridged bis(β -CD) (1.2 g, 0.45 mmol) was dissolved in 40 mL of anhydrous pyridine, and 0.23 mL of 3-isocyanatopropyltriethoxysilane was added dropwise with stirring for 30 min. The temperature was increased to 80 °C and the solution was reacted for 12 h under nitrogen protection. After cooling to room temperature, silica gel (3 g) was added to the mixture, which was reacted at 115 °C for 24 h under nitrogen protection. After filtration, the solid product was successively washed several times with pyridine, MeOH, and acetone, and dried under vacuum at 50 °C for 24 h to obtain HTCDP.

CDCSP was prepared in a manner similar to HTCDP. β -CD (1 g, 0.88 mmol) was dissolved in 30 mL of anhydrous pyridine; 0.44 mL of 3-isocyanatopropyltriethoxysilane was added dropwise with stirring for 30 min, and the mixture was reacted at 80 °C and for 12 h under nitrogen protection. The temperature was lowered to room temperature and silica gel (3 g) was added to the mixture, which was reacted at 115 °C for 24 h under nitrogen protection to obtain CDCSP.

2.4. Packing of the chromatographic column

HTCDP and CDCSP were packed into stainless steel columns (150 \times 4.6 mm inner diameter) using a standard slurry method. A 2.5 g of HTCDP or CDCSP was dispersed in 50 mL of MeOH and dispersed by ultrasound for 5 min, then poured into the homogenizing tank; with MeOH used as the displacement liquid, the tank was filled at 40 MPa pressure for 35 min. Finally,

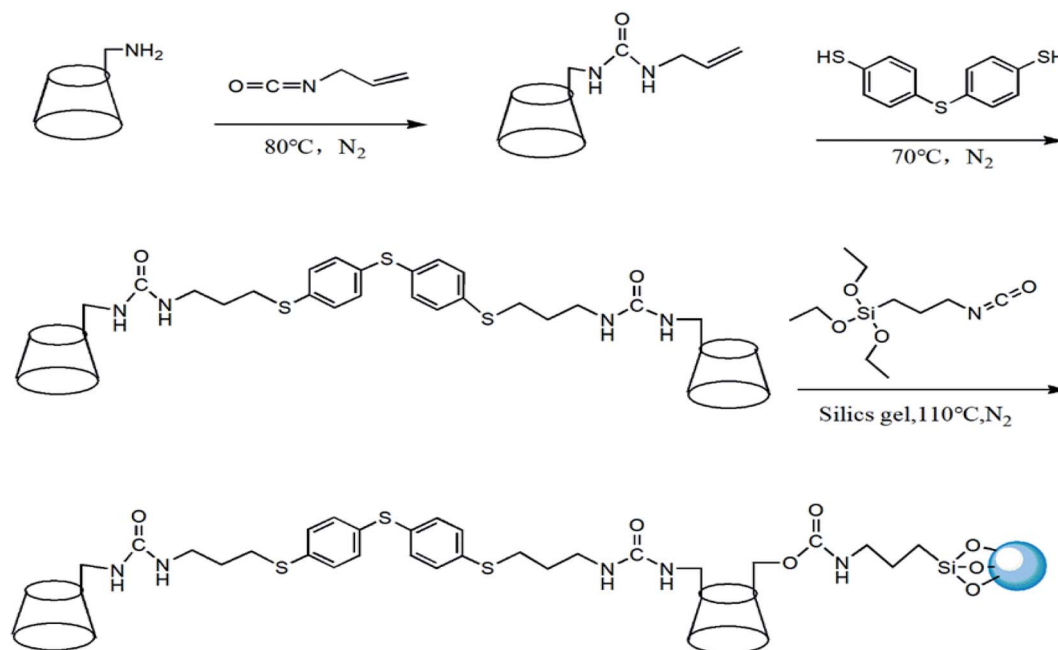


Fig. 1 The synthetic route of HTCDP.



the pressure was gradually released to atmospheric level to complete the packing of the chromatographic column.

2.5. Chromatographic methods

The mobile phase consisted of MeOH or ACN and water. According to the actual separation of analytes, a certain amount of FA or TEA and HOAc was added to the water. The 1% of triethylammonium acetate buffer (1% TEAA) was prepared from TEA and water in a volume ratio of 1 : 100, and the pH was adjusted with HOAc. All chiral drugs and pesticides were dissolved in MeOH to prepare standard solutions of 100–1000 $\mu\text{g mL}^{-1}$. All standard analytes and mobile phases were passed through a 0.22 μm membrane filter and degassed by ultrasound before use. The detection wavelength was set to 214 nm. The flow rate was set to 0.8 mL min^{-1} . The injection volume was 3–10 μL . The column temperature was adjusted according to the separation conditions of analytes. HPLC analysis was performed under isocratic conditions. The storage temperature of the analyzed sample was 4 $^{\circ}\text{C}$.

According to the USP standard, three parameters for evaluating chromatographic performance were calculated. (1) Retention factor (k') = $(t_{\text{R}} - t_0)/t_0$, where t_0 is the retention time of the first baseline interference of the solvent peak (unit: min), and t_{R} is the retention time of each enantiomer (unit: min); (2) separation factor (α) = k_2'/k_1' , where k_1' and k_2' are the retention factors of the two enantiomers eluted before and after; (3) resolution (R_s) = $1.18(t_{\text{R}2} - t_{\text{R}1})/(W_1 + W_2)$, where $t_{\text{R}1}$ and W_1 are the retention time and half-width of the enantiomer that is eluted first, respectively; $t_{\text{R}2}$ and W_2 are the retention time and half-width of the enantiomer that is eluted later, respectively.

3. Results and discussion

3.1. Characterization of HTCDP

The prepared HTCDP was characterized by infrared (IR) spectroscopy, elemental analysis, thermogravimetric analysis, ^{13}C NMR

and scanning electron microscopy. The IR spectrum of the bridged bis(β -CD) ligand (Fig. S3a†) showed a strong peak at 3413.58 cm^{-1} corresponding to the stretching vibration peak of the residual hydroxyl (ν_{OH}) group; the peak at 2928.08 cm^{-1} was related to the stretching vibration of $\nu_{\text{C-H}}$; peaks at 1638.35 and 1710.21 cm^{-1} were attributable to the carbonyl ($\nu_{\text{C=O}}$) stretching vibration in the bridge arms; peak clusters at 1535.51 and 1411.39 cm^{-1} reflected tensile vibration of the benzene ring ($\nu_{\text{C=C}}$); and peaks at 1029.37 and 1238.37 cm^{-1} were related to the $\nu_{\text{C-O}}$ stretching vibration of the bridged bis(β -CD) ligand. These results indicate that the bridged bis(β -CD) ligand was successfully prepared. In the IR spectrum of HTCDP (Fig. S3b†), the strong peak at 3416.38 cm^{-1} corresponded to the stretching vibration of the residual hydroxyl groups (ν_{OH}) on the silica gel and bridged bis(β -CD) ligand; the peak at 1618.39 cm^{-1} was related to the stretching vibration of the carbonyl ($\nu_{\text{C=O}}$) on the bridged bis(β -CD) ligand; and the strong peak at 1114.93 cm^{-1} was attributable to the stretching vibration of Si–O–Si. This demonstrated that the bridging bis(β -CD) ligand was bonded to the silica gel.

The results of the elemental analysis of HTCDP and CDCSP were depicted in Table S1.† HTCDP contained 3.23% C, 0.46% H, 0.24% N, and 0.12% S, whereas CDCSP contained 1.6% C, 0.28% H, and 0.38% N. The loads of HTCDP and CDCSP were calculated based on their carbon contents using the following formula:⁴³ $\text{C\%}/(12 \times N_{\text{c}} \times S) \times 10^6$, where C% is the percentage of carbon in the CSP, 12 is the atomic mass of carbon, N_{c} (about 123) is the number of carbon atoms in each molecule of the CD unit, and S is the specific surface area of silica gel ($60\text{ m}^2\text{ g}^{-1}$). As a result, the loads of HTCDP and CDCSP were comparable at 0.45 and 0.51 $\mu\text{mol m}^{-2}$, respectively.

The thermogravimetric analysis curve of HTCDP was exhibited in Fig. S4.† The weight loss rate of HTCDP was about 9.26% with a heating rate of $10\text{ }^{\circ}\text{C min}^{-1}$ from $30\text{ }^{\circ}\text{C}$ to $800\text{ }^{\circ}\text{C}$. HTCDP began to lose weight significantly at about $300\text{ }^{\circ}\text{C}$, indicating that HTCDP had good chemical and thermal stability.

Table 1 The optimal separation results and chromatographic conditions of 8 flavanone enantiomers^a

No.	Compounds	k_1	k_2	α	R_s	Mobile phases (v/v)	Temp. ($^{\circ}\text{C}$)	CSPs
1	Flavanone	9.17	11.16	1.22	3.06	ACN/0.1% FA (15/85)	20 $^{\circ}\text{C}$	HTCDP
		2.81	—	—	—	ACN/0.1% FA (15/85)	20 $^{\circ}\text{C}$	CDCSP
2	2-Hydroxy flavanone	2.11	2.96	1.40	3.89	ACN/0.1% FA (20/80)	20 $^{\circ}\text{C}$	HTCDP
		2.34	2.96	1.26	1.92	ACN/0.1% FA (15/85)	20 $^{\circ}\text{C}$	CDCSP
3	4-Hydroxy flavanone	2.94	4.36	1.48	4.48	ACN/0.1% FA (20/80)	20 $^{\circ}\text{C}$	HTCDP
		2.19	—	—	—	ACN/0.1% FA (15/85)	20 $^{\circ}\text{C}$	CDCSP
4	6-Hydroxy flavanone	11.76	12.78	1.09	1.26	MeOH/0.3% FA (20/80)	30 $^{\circ}\text{C}$	HTCDP
		1.96	—	—	—	MeOH/0.3% FA (15/85)	30 $^{\circ}\text{C}$	CDCSP
5	6-Methoxy flavanone	10.77	11.79	1.09	1.19	MeOH/0.3% FA (25/75)	30 $^{\circ}\text{C}$	HTCDP
		2.88	—	—	—	MeOH/0.3% FA (15/85)	30 $^{\circ}\text{C}$	CDCSP
6	Naringin	2.73	3.35	1.23	1.82	MeOH/0.1% FA (15/85)	20 $^{\circ}\text{C}$	HTCDP
		2.40	2.76	1.15	1.19	MeOH/0.1% FA (10/90)	20 $^{\circ}\text{C}$	CDCSP
7	Hesperidin	4.06	4.66	1.14	1.50	ACN/0.1% FA (20/80)	20 $^{\circ}\text{C}$	HTCDP
		1.59	—	—	—	ACN/0.1% FA (15/85)	20 $^{\circ}\text{C}$	CDCSP
8	Silymarin	1.50	4.73	3.15	4.34	ACN/0.1% FA (15/85)	20 $^{\circ}\text{C}$	HTCDP
		0.40	0.70	1.75	1.47	ACN/0.1% FA (15/85)	20 $^{\circ}\text{C}$	CDCSP

^a k' , retention factor; α , separation factor; R_s , resolution; temp., temperature ($^{\circ}\text{C}$); CSPs, chiral stationary phase; HTCDP, a novel bridged bis(β -cyclodextrin) chiral stationary phase; CDCSP, native β -cyclodextrin chiral stationary phase; —, no separation.



The synthesized HTCDP was characterized by scanning electron microscope and solid state ^{13}C NMR. The images were shown in Fig. S5 and S6,[†] respectively.

The above characterization results and excellent enantiomeric separation ability indicated that HTCDP and CDCSP were successfully prepared.

3.2. Performance of HTCDP in the chromatographic separation of enantiomers

In the study, 21 common chiral compounds with different chemical structures and properties were selected as analytes to examine the performance of HTCDP in enantioseparation, of which included 8 flavanones, 8 triazole pesticides and 5 other drugs. Moreover, by separating 21 selected analytes under reversed-phase chromatographic mode, some related theories of enantiomeric separation were discussed preliminarily.

3.2.1. Enantioseparation of flavanones. Flavanones are widely distributed in natural plants and their active ingredients are used in medicine for their bactericidal, antioxidant, disinfectant, and anticancer effects. Because flavanone enantiomers often show different biological activities and medicinal value, the separation of their enantiomers is of great significance. In this study, 8 relatively large flavanone molecules (Fig. S7[†]) were selected to evaluate the chiral chromatographic separation performance of HTCDP; the best separation data and chromatographic conditions were listed in Table 1, and representative chromatograms were shown in Fig. 2.

As shown in Table 1, we can find that HTCDP separated all 8 flavanones, with 6 of these completely separated at high resolution (1.5–4.48). In contrast, CDCSP separated only 3 flavanones at a lower resolution and did not separate flavanone, 4-hydroxyflavanone, 6-hydroxyflavanone, 6-methoxyflavanone, and hesperidin. Moreover, most derivatized mono-CD CSPs separate flavanones at a lower resolution than HTCDP,^{17,47}

suggesting that the two bridging cavities of HTCDP promote the recognition of flavanone enantiomers. We also found that the resolution of flavanone, 4-hydroxyflavanone, 6-hydroxyflavanone, 6-methoxyflavanone, and hesperidin was significantly higher on HTCDP than on CDCSP without π -conjugated aryl-bridged bis(β -CD),⁴² indicating that the aryl bond arm of the bridged bis(β -CD) ligand was also important for the recognition of chirality.

On the other hand, hydrogen bonds also play a vital role in the separation of enantiomers. Both HTCDP and CDCSP were able to separate 2-hydroxyflavanone, naringenin, and silymarin. As shown in the Fig. S7,[†] the chiral carbons of the 3 analytes were directly connected to the hydroxyl-containing benzene ring and were potentially responsible for their separation, in line with the previous observation that hydrogen bonding contributes to chiral separation.³² HTCDP separated these 3 analytes at a higher resolution than CDCSP, possibly as a result of the introduction of urea groups into the bridged bonding arms of HTCDP that strengthened the hydrogen bonding. We also found that the resolution of 6-hydroxyflavanone and 6-methoxyflavanone separated by HTCDP was relatively low. This may be because the hydroxyl and methoxy groups at position 6 could not fully enter the hydrophobic cavity of CD, thereby weakening their inclusion.¹⁷ However, 6-hydroxyflavanone had a higher resolution than 6-methoxyflavanone, possibly because the former formed hydrogen bonds with the urea group on the spacer arm of HTCDP and thereby enhanced its chiral recognition.¹⁷ In summary, the two cavities of HTCDP provided synergistic inclusion while the underivatized hydroxyl group engaged in hydrogen bonding and the aromatic ring on the bridge engaged in π - π interactions; the combination of these interactions enhanced the chiral recognition ability of HTCDP, consistent with previous reports.^{43,45}

3.2.1.1. Effect of mobile phase and column temperature conditions on the separation of flavanones. The composition of

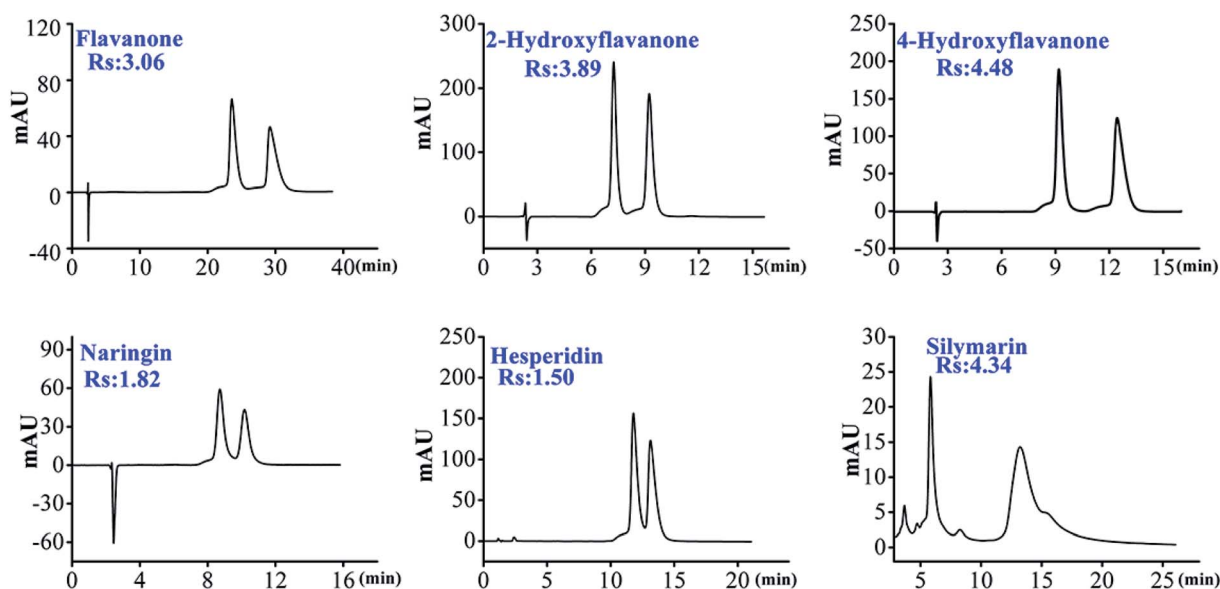


Fig. 2 The representative chromatograms of flavanone enantiomers on HTCDP.



the mobile phase is a key factor affecting the chiral separation of β -CD CSPs. In this study, because most of the selected chiral flavanones contained acidic phenolic hydroxyl groups, FA was added to the mobile phase in order to inhibit the ionization of acidic phenolic hydroxyl groups.²⁹ As shown in Table 1, all 8 flavanones could be separated on HTCDP using MeOH-FA or ACN-FA aqueous solution as the mobile phase. However, we found that MeOH-FA and ACN-FA aqueous solution respectively used as mobile phases caused a huge difference in the chiral resolution of HTCDP. For example, as shown in Table S2,[†] using ACN-formic acid aqueous solution as the mobile phase, the separation factor and resolution of 2-hydroxyflavanone were 1.40 and 3.89, respectively; the separation factor and resolution of 4-hydroxyflavanone were 1.48 and 4.48, respectively. Whereas using MeOH-formic acid aqueous solution as the mobile phase, the separation factor and resolution of 2-hydroxyflavanone were 1.29 and 2.06, respectively; the separation factor and resolution of 4-hydroxyflavanone are 1.31 and 2.68, respectively. This may be because MeOH competes with HTCDP for the hydrogen bonding sites of the analyte, which weakens the retention and separation factors of the analyte on HTCDP, thereby weakening the interaction of the latter.³² Interestingly, the opposite results were obtained in the separation of 6-hydroxyflavanone, naringenin, and 6-methoxyflavanone. Specifically, there was little separation of 6-hydroxyflavanone and 6-methoxyflavanone using ACN-FA aqueous solution as the mobile phase. This may be because MeOH is more polar than ACN and can increase the interaction

time between the them and the β -CD hydrophobic cavity, thereby enhancing the inclusion interaction between the them and the cavity.¹⁷ Moreover, we investigated the influence of MeOH content in the mobile phase on the separation of 6-methoxyflavanone. As shown in Fig. 3a, as the MeOH content increased from 20% to 35%, the resolution of 6-hydroxyflavanone increased before decreasing while the retention time also decreased. The resolution of 6-hydroxyflavanone was maximal (R_s : 1.19) at 25% MeOH. Thus, in order to improve the resolution of enantiomers, the proportioning and type of mobile phase need to be finely optimized.

Column temperature also plays an important role in the chromatographic separation of chiral compounds. Some enantiomers showed decreasing resolution with increasing column temperature while others showed the opposite trend.^{43,48} We examined the effect of column temperature on the resolution of 8 selected flavanones. Using 2-hydroxyflavanone as an example, as shown in Fig. 3b and Table S3,[†] the retention time (k'), separation factor (α), and resolution (R_s) of 2-hydroxyflavanone decreased with increasing column temperature. In order to improve the resolution of flavanones, we selected the lowest possible column temperature. However, the retention times of 6-hydroxyflavanone and 6-methoxyflavanone were longer at 20 °C. Given that raising the temperature can reduce retention time, they were separated at 30 °C.

3.2.1.2. Effect of FA content on the separation of flavanones. As we described earlier, due to most flavanones contain acidic phenolic hydroxyl groups that ionize when resolved by HTCDP.

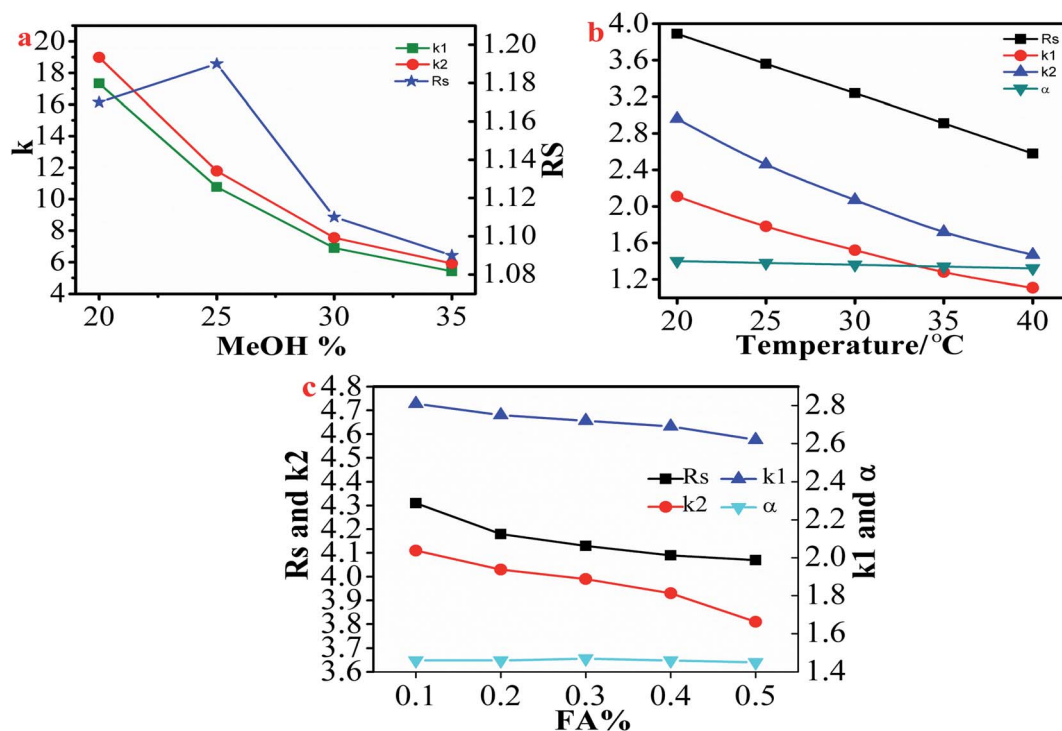


Fig. 3 The effect of methanol content in the mobile phase on the resolution, and retention factor of 6-methoxyflavanone enantiomers (a); the effect of column temperature on the resolution, separation factor and retention factor of 2-hydroxyflavanone enantiomers (b) and the effect of formic acid content in the mobile phase on the resolution, separation factor and retention factor of 4-hydroxyflavanone enantiomers (c).

Table 2 The optimal separation results and chromatographic conditions of 8 triazole pesticide enantiomers^a

No.	Compounds	k_1	k_2	α	R_s	Mobile phases (v/v)	Temp. (°C)	CSPs
9	Hexaconazole	4.48	5.42	1.21	2.19	ACN/H ₂ O (20/80)	20 °C	HTCDP
		3.02	3.50	1.16	1.32	ACN/H ₂ O (15/85)	20 °C	CDCSP
10	Imazalil	5.49	6.76	1.23	2.14	ACN/1% TEAA (10/90)	20 °C	HTCDP
		2.48	—	—	—	ACN/1% TEAA (10/90)	20 °C	CDCSP
11	Tebuconazole	7.46	8.76	1.17	1.91	ACN/H ₂ O (20/80)	20 °C	HTCDP
		9.30	—	—	—	ACN/H ₂ O (10/90)	20 °C	CDCSP
12	Flutriafol	4.80	5.57	1.16	1.84	ACN/H ₂ O (12/88)	20 °C	HTCDP
		3.51	3.85	1.10	0.94	ACN/H ₂ O (12/88)	20 °C	CDCSP
13	Paclobutrazol	7.76	9.64	1.26	1.79	ACN/H ₂ O (28/72)	20 °C	HTCDP
		5.33	5.75	1.08	0.71	ACN/H ₂ O (10/90)	20 °C	CDCSP
14	Diniconazole	6.02	7.04	1.17	1.71	ACN/H ₂ O (20/80)	20 °C	HTCDP
		3.99	4.34	1.09	0.72	ACN/H ₂ O (10/90)	20 °C	CDCSP
15	Uniconazole	15.97	19.40	1.22	1.54	ACN/H ₂ O (25/75)	20 °C	HTCDP
		3.93	—	—	—	ACN/H ₂ O (15/85)	20 °C	CDCSP
16	Triticonazole	8.98	10.60	1.18	1.96	ACN/H ₂ O (20/80)	20 °C	HTCDP
		3.61	3.87	1.07	0.62	ACN/H ₂ O (15/85)	20 °C	CDCSP

^a k' , retention factor; α , separation factor; R_s , resolution; temp., temperature (°C); CSPs, chiral stationary phase; HTCDP, a novel bridged bis(β -cyclodextrin) chiral stationary phase; CDCSP, native β -cyclodextrin chiral stationary phase; —, no separation.

In order to prevent this process and maintain ionization equilibrium, FA was added to the mobile phase.³² We explored the effect of the ratio of FA to water on the resolution of flavanone compounds using 4-hydroxyflavanone as an example. As can be seen from Fig. 3c, the FA content had no effect on the separation factor of 4-hydroxyflavanone; however, as it increased from 0.1% to 0.5%, the resolution and retention factor of 4-hydroxyflavanone continued to decrease. Therefore, in order to achieve rapid separation and prevent the ionization of phenolic hydroxyl groups, an appropriate amount of FA should be added to the mobile phase. In this study, most selected flavanones were sufficient to achieve rapid and baseline separation with 0.1% FA content. However, given the long retention time of 6-hydroxyflavanone and 6-methoxyflavanone in HTCDP, we increased the FA concentration in the mobile phase from 0.1% to 0.3%, which enhanced the resolution of these 2 analytes.

3.2.2. Enantioseparation of triazole pesticides. Triazole pesticides are widely used as fungicides and growth regulators in agriculture. So far, most of the more than 30 common triazole pesticides are still used in racemic form. Because the enantiomers of triazole pesticides have huge differences in toxicity and biological activity, they enter the human body through the food chain and cause liver and kidney damage in humans. Thus, we selected 8 triazole pesticides as analytes to evaluate the separation performance of HTCDP (Fig. S8†). The best separation data and chromatographic conditions of the pesticides were listed in Table 2, and representative chromatograms were exhibited in Fig. 4.

As shown in Table 2, most of the selected chiral triazole analytes could be separated by HTCDP and CDCSP under simple ACN–H₂O mobile phase conditions, demonstrating that the inclusion effects of HTCDP and CDCSP were the main driving force for their separation.⁴⁹ However, the resolution of the 8 selected chiral triazole analytes on HTCDP and CDCSP differed markedly: HTCDP completely separated all 8 analytes

at high resolution (R_s : 1.54–2.19) whereas CDCSP separated only 5 at a lower resolution (R_s : 0.62–1.32), failing to separate uniconazole, tebuconazole, and imazalil. As HTCDP and CDCSP are both underivatized β -CD CSPs, their different abilities to resolve the 8 analytes can be attributed to their distinct modes of chiral recognition. For example, the resolution of imazalil on HTCDP was as high as 2.14 whereas CDCSP did not separate this compound, possibly because of the lack of hydroxyl groups that could form hydrogen bonds on the chiral carbon of imazalil and the small size of the CDCSP cavity (0.65 nm). However, the two cavities in HTCDP linked *via* an aryl connector synergistically were able to accommodate the bulky imazalil molecule. Moreover, there was a π – π interaction between the aromatic ring of imazalil and the aryl connector of HTCDP. The combination of these two interactions enhanced the separation of imazalil by HTCDP, consistent with the previously reported mechanism.^{43,45}

In addition to the inclusion effect and π – π interaction, hydrogen bonding plays an extremely important role in chiral separation. In addition to the 8 triazole pesticides, we also tried to separate myclobutanil, ornidazole, voriconazole, and lansoprazole (Fig. S9†) on HTCDP. All 4 compounds had a very short retention time and were not separated (Table S4†), unlike the 8 triazole pesticides (Table 1); this is likely because the chiral carbons of the latter compounds have hydroxyl groups that could form hydrogen bonds with HTCDP, thereby promoting their separation. This is consistent with the results previously reported.^{43,44}

3.2.2.1. Effect of column temperature on separation. In order to improve the resolution of the selected triazole analytes, we next studied the effect of column temperature (ranging from 20 °C to 45 °C in increments of 5 °C) on the resolution of all 8 triazole chiral analytes. In the paper, we chose hexaconazole as the example for description. As shown in Fig. 5a and Table S5,† the retention factor, separation factor and resolution of



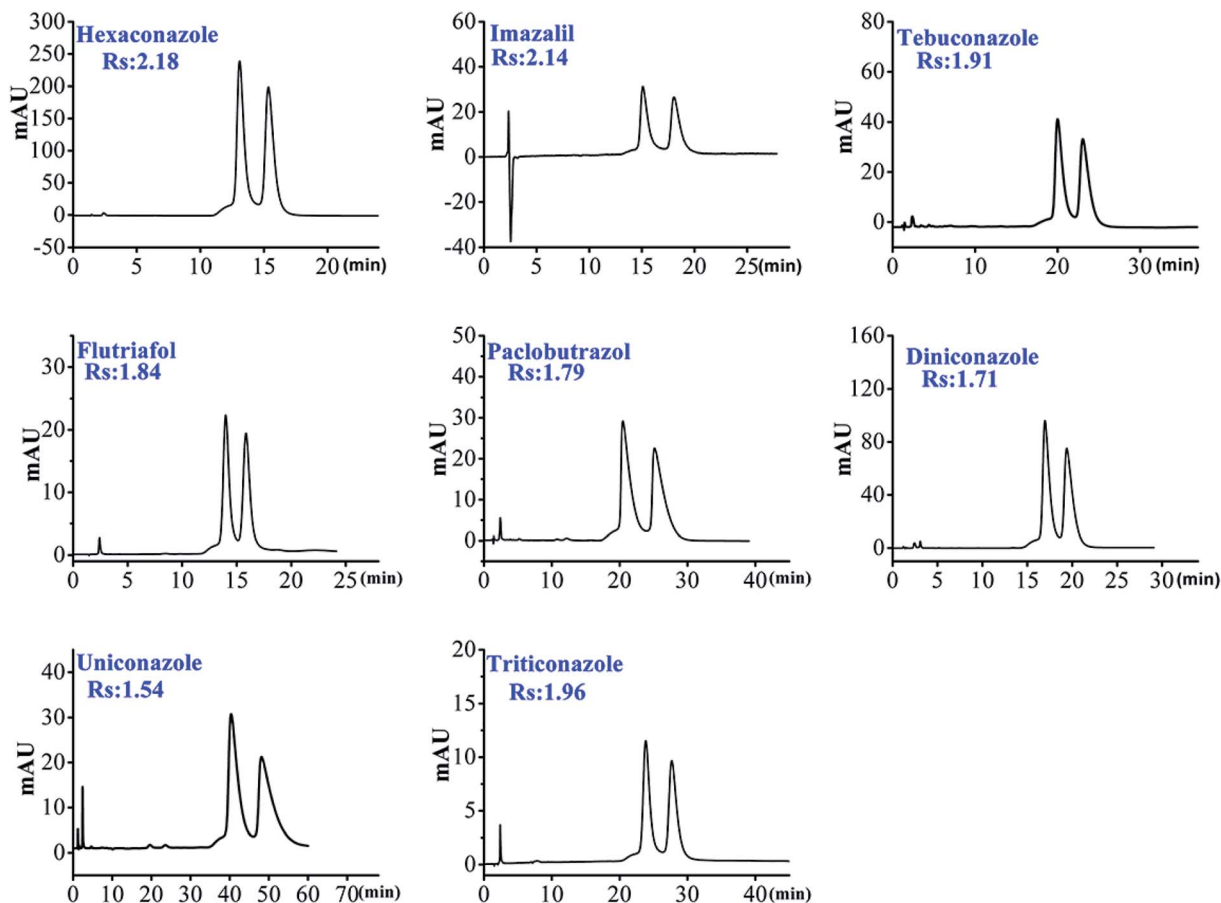


Fig. 4 The chromatogram of 8 triazole pesticide enantiomers on HTCDP.

hexaconazole decreased with the increase of column temperature. However, column temperature from 20 °C to 25 °C, the resolution, separation factor and retention factor of hexaconazole decreased rapidly. Therefore, in order to achieve a higher resolution, the separation of the selected triazole chiral analytes should be carried out at ≤ 20 °C.

3.2.2.2. Effect of pH on the imazalil enantiomer resolution. We found that the above-mentioned other chiral triazole analytes could be separated with simple ACN-H₂O as the mobile phase

with the exception of imazalil. In order to achieve the separation of imazalil enantiomers, we used ACN-1% TEAA as the mobile phase to separate imazalil. Interestingly, imazalil was separated to a certain extent by adjusting the pH of the 1% TEAA solution. We evaluated the effect of pH on the resolution of imazalil and found that as the pH value decreased, the resolution first increased before decreasing (Fig. 5 b); the resolution was maximal (1.82) at pH 4.5 and was 1.52 at pH 4. This should be related to the change in charge and polarity of the imazalil at

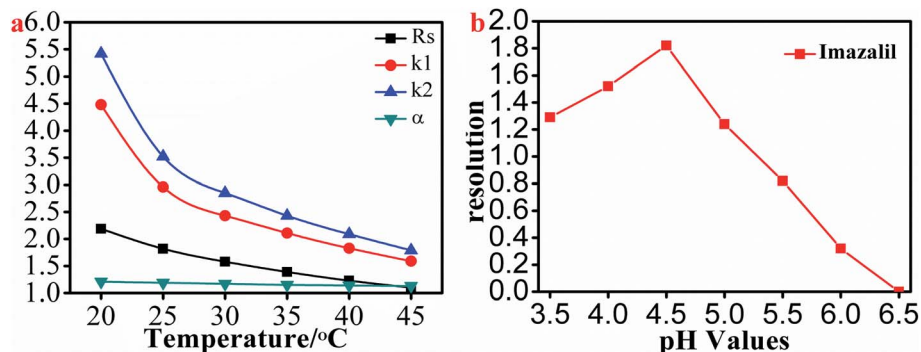
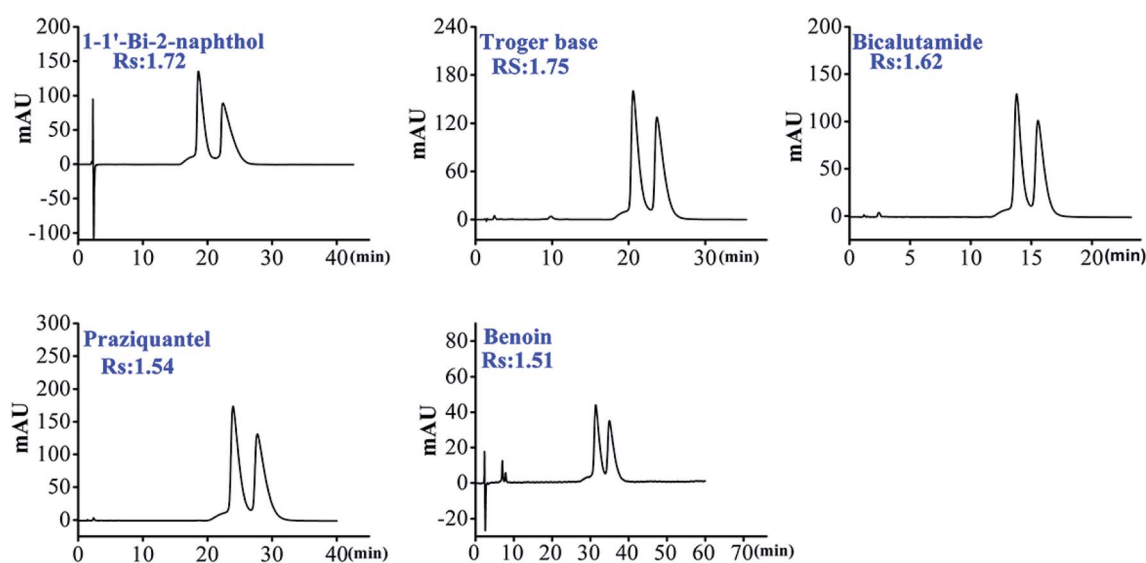


Fig. 5 The effect of column temperature on the resolution, separation factor and retention factor of hexaconazole enantiomers (a) and the effect of pH values in mobile phase on the resolution of imazalil enantiomers (b).

Table 3 The optimal separation results and chromatographic conditions of 5 common enantiomers^a

No.	Compounds	k_1	k_2	α	R_s	Mobile phases (v/v)	Temp. (°C)	CSPs
17	Troger's base	7.42	8.71	1.17	1.75	ACN/H ₂ O (17/83)	20 °C	HTCDP
		3.67	—	—	—	ACN/H ₂ O (15/85)	20 °C	CDCSP
18	1-1'-Bi-2-naphthol	7.24	9.01	1.24	1.72	MeOH/1%TEAA (15/85)	20 °C	HTCDP
		1.85	—	—	—	MeOH/1%TEAA (15/85)	20 °C	CDCSP
19	Bicalutamide	4.71	5.45	1.16	1.62	MeOH/H ₂ O (15/85)	20 °C	HTCDP
		2.07	—	—	—	MeOH/H ₂ O (15/85)	20 °C	CDCSP
20	Praziquantel	9.18	10.80	1.18	1.54	ACN/H ₂ O (15/85)	20 °C	HTCDP
		4.10	—	—	—	ACN/H ₂ O (12/88)	20 °C	CDCSP
21	Benzoin	12.54	14.10	1.12	1.51	ACN/1%TEAA (5/95)	20 °C	HTCDP
		5.53	—	—	—	ACN/1%TEAA (10/90)	20 °C	CDCSP

^a k' , retention factor; α , separation factor; R_s , resolution; temp., temperature (°C); CSPs, chiral stationary phase; HTCDP, a novel bridged bis(β -cyclodextrin) chiral stationary phase; CDCSP, native β -cyclodextrin chiral stationary phase; —, no separation.

**Fig. 6** The chromatograms of 5 common enantiomers on HTCDP.

different pH values, which significantly affected the interaction and separation of the imazalil and HTCDP.³⁸ Thus, the optimal pH to achieve baseline separation of imazalil is between 4 and 4.5. We also determined that imazalil was separated at the highest resolution (2.14) using HTCDP when the proportion of ACN was 10%.

3.2.3. Separation of 5 other common chiral compounds. In order to further investigate the versatility of HTCDP in chromatographic separation, we also tried to separate 5 common chiral compounds, including benzoin (a resin used to treat stroke phlegm convulsion, postpartum blood halo, and infantile convulsion); praziquantel (an anthelmintic used to prevent and treat schistosomiasis, paragonimiasis, echinococcosis, and other diseases); bicalutamide (an anti-androgen drug for the treatment of advanced prostate cancer); Troger's base (a racemate commonly used for molecular recognition, chiral catalysis, and molecular biomimetic receptors); and 1-1'-bi-2-naphthol (an intermediate in asymmetric catalysis and drug

synthesis). Separation data of the 5 compounds were listed in Table 3, and representative chromatograms and chemical structures were shown in Fig. 6 and S10,[†] respectively.

As shown in Table 3, HTCDP completely separated all 5 analytes while none was separated by CDCSP. It is worth noting that the resolution of benzoin on HTCDP was higher than that on all reported bridged β -CD CSPs.^{39,40} The above results indicate that HTCDP exhibited high enantiomeric separation ability to the selected 5 analytes. As can be seen from Fig. S10,[†] the 5 selected analytes were relatively large molecules and most were composed solely of branched chains and benzene rings. As CDCSP is an underivatized β -CD CSP with a relatively small cavity (0.65 nm), there was no inclusion effect on the 5 analytes, which ultimately led to the failure of separation. However, all 5 compounds were separated by HTCDP at higher resolution; this may be because the analytes were included in the two cavities of HTCDP and engaged in π - π interaction with the benzene ring on HTCDP bridge arms.^{43,45}



4. Conclusions

In this paper, a novel HTCDP was prepared through an efficient, simple and rapid thiol–ene click chemistry reaction. The chiral chromatographic performance of HTCDP was systematically studied and evaluated by separating 21 chiral triazole pesticides and drugs in reverse phase chromatographic mode. Moreover, we also studied in detail the effects of formic acid content, column temperature, pH values, and mobile phase composition on enantiomeric resolution. The results showed that in addition to 6-hydroxyflavanone and 6-methoxyflavanone, HTCDP completely separated the other 19 selected analytes with relatively high resolution (1.50–4.48). By constructing a novel bridged bis(β -CD) ligand with a bridging linker (with a urea group and an aromatic ring) and two cavities, many interactions such as π – π , hydrogen bonds, and synergistic inclusion were introduced into the enantiomeric separation, thereby improving the enantiomeric separation and selection capabilities of HTCDP. Thus, HTCDP can potentially be used for quality and safety analyses of chiral drugs in the future.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was financially supported by the Key Research and Development Program of Ningxia (Special Project for Foreign Cooperation) (No. 2019BFH03004), Introduced Team Program of Ningxia (Rouxing) (No. 2019RXTD0004) and Graduate Innovation Project of North Minzu University (No. YCX20141).

References

- 1 Y. P. Xue, C. H. Cao and Y. G. Zheng, *Chem. Soc. Rev.*, 2018, **47**, 1142.
- 2 S. R. Crowell, W. M. Henderson, J. F. Kenneke and J. W. Fisher, *Toxicol. Lett.*, 2011, **205**, 154.
- 3 L. Z. Chen, D. Q. Zhu and P. Xiang, *Bioanalysis*, 2021, **13**, 493.
- 4 B. Zhang, S. H. Yang, J. P. Deng and K. Pan, *Adv. Sci.*, 2021, **8**, 2003681.
- 5 A. Cavazzini, L. Pasti, A. Massi, N. Marchetti and F. Dondi, *Anal. Chim. Acta*, 2011, **706**, 205.
- 6 O. Horacek, M. Papajova-Janetkova, B. Gruner, L. Lochman, P. SterbovaKovarikova, R. Vespalec and R. Kucera, *Talanta*, 2021, **222**, 121652.
- 7 B. Tang, X. Zhang, L. N. Geng, L. Q. Sun and A. Q. Luo, *J. Chromatogr. A*, 2021, **1636**, 461792.
- 8 Z. T. Li, C. J. Hu, Y. K. Liu, Q. Y. Li, Y. Y. Fu and Z. L. Chen, *J. Chromatogr. A*, 2021, **1643**, 462082.
- 9 M. Lämmerhofer, *J. Chromatogr. A*, 2010, **1217**, 814.
- 10 J. Voskuhl, K. Schaepe and B. J. Ravoo, *Int. J. Mol. Sci.*, 2021, **12**, 4637.
- 11 Y. F. Poon, I. W. Muderawan and I. W. Ng, *J. Chromatogr. A*, 2006, **1101**, 185.
- 12 X. B. Yao, T. T. Y. Tan and Y. Wang, *J. Chromatogr. A*, 2014, **1316**, 80.
- 13 J. Zhou, B. Yang, J. Tang and W. H. Tang, *J. Chromatogr. A*, 2016, **1467**, 169.
- 14 R. Q. Wang, T. T. Ong and S. C. Ng, *Tetrahedron Lett.*, 2012, **53**, 2312.
- 15 Z. A. AlOthman, A. G. Alanazi, M. Suhail and I. Ali, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2020, **1157**, 122335.
- 16 S. Ravichandran, J. R. Collins, N. Singh and I. W. Wainer, *J. Chromatogr. A*, 2012, **1269**, 218.
- 17 L. Li, H. Wang, Y. Z. Shuang and L. S. Li, *Talanta*, 2019, **202**, 494.
- 18 A. Rosetti, C. Villani, M. Pierini and R. Cirilli, *Molecules*, 2021, **26**, 1747.
- 19 S. H. Huang, Z. W. Bai, Q. Chuan, S. R. Li and Z. Q. Pan, *Chirality*, 2007, **19**, 129.
- 20 C. J. Easton and S. F. Lincoln, *Chem. Soc. Rev.*, 1996, **25**, 163.
- 21 X. X. Li, X. Jin, X. B. Yao, X. F. Ma and Y. Wang, *J. Chromatogr. A*, 2016, **1467**, 279.
- 22 L. Li, B. P. Chen, R. D. Zhou, Z. G. Cao, C. Zeng and L. S. Li, *Talanta*, 2017, **174**, 179.
- 23 J. Y. Sun, S. M. Ma, B. B. Liu, J. Yu and X. J. Guo, *Talanta*, 2019, **204**, 817.
- 24 A. M. Stalcup, S. C. Chang and D. W. Armstrong, *J. Chromatogr. A*, 1991, **540**, 113.
- 25 G. K. E. Scriba, *J. Chromatogr. A*, 2016, **1467**, 56.
- 26 Y. Z. Shuang, Z. G. Cao, T. C. Zhang and L. S. Li, *Anal. Lett.*, 2020, **53**(15), 2481.
- 27 S. C. Chang, G. L. R. Iii, S. Chen, C. D. Chang and D. W. Armstrong, *Anal. Chem.*, 1993, **12**, 144.
- 28 P. Sun, C. L. Wang, Z. S. Breitbach, Y. Zhang and D. W. Armstrong, *Anal. Chem.*, 2009, **81**, 10215.
- 29 Q. Q. Zhong, L. F. He, T. E. Beesley, W. S. Trahanovsky, P. Sun, C. L. Wang and D. W. Armstrong, *J. Chromatogr. A*, 2006, **1115**, 19.
- 30 A. M. Stalcup, S. C. Chang and D. W. Armstrong, *J. Chromatogr. A*, 1991, **540**, 113.
- 31 X. Y. Yu and Z. P. Yao, *Anal. Chim. Acta*, 2017, **968**, 1.
- 32 K. E. G. Scriba, *J. Chromatogr. A*, 2016, **1467**, 56.
- 33 L. F. Zhang, J. Zhou and Y. Wang, *Acta Chim. Sin.*, 2015, **73**, 1182.
- 34 M. Nakamura, T. Ikeda, A. Nakamura, H. Ikeda, A. Ueno and F. Toda, *Chem. Lett.*, 1995, **5**, 343.
- 35 A. Ruebner, J. G. Moser, D. Kirsch, B. Spengler, S. Andress and S. Roehrs, *J. Inclusion Phenom. Macrocyclic Chem.*, 1996, **25**, 35.
- 36 D. Q. Yuan, J. Lu, M. Atsumi, J. M. Yan, M. Kai and K. Fujita, *Org. Biomol. Chem.*, 2007, **5**, 2932.
- 37 B. Zhang and R. Breslow, *J. Am. Chem. Soc.*, 1997, **119**, 1676.
- 38 R. Breslow, N. Greenspoon, T. Guo and R. Zarzycki, *J. Am. Chem. Soc.*, 1989, **111**, 8296.
- 39 J. Zhao, X. H. Lu, Y. Wang and J. Lv, *J. Chromatogr. A*, 2015, **1381**, 253.
- 40 J. Zhao, X. H. Lu, Y. Wang and T. T. Y. Tan, *J. Chromatogr. A*, 2014, **1343**, 101.

- 41 P. Ai, L. N. Han, M. Zi, L. Meng, F. T. Zi and L. M. Yuan, *Chin. J. Anal. Chem.*, 2006, **34**, 1459.
- 42 Y. Z. Shuang, Y. Q. Liao, T. C. Zhang and L. S. Li, *J. Chromatogr. A*, 2020, **1619**, 460937.
- 43 Y. Z. Shuang, T. C. Zhang and L. S. Li, *J. Chromatogr. A*, 2020, **1614**, 460702.
- 44 Y. Z. Shuang, Y. Q. Liao, H. Wang, Y. X. Wang and L. S. Li, *Chirality*, 2020, **32**, 168.
- 45 Y. Liu and Y. Chen, *Acc. Chem. Res.*, 2006, **39**, 681.
- 46 M. Chen, X. L. Lu, X. F. Ma, Y. Xiao and Y. Wang, *Analyst*, 2021, **146**, 3025.
- 47 T. C. Zhang, Y. Z. Shuang, H. Zhong, L. Li and L. S. Li, *Anal. Sci.*, 2021, **37**, 1095.
- 48 E. Küsters, V. Loux, E. Schmid and P. Floersheim, *J. Chromatogr. A*, 1994, **666**, 421.
- 49 P. Jandera, *Anal. Chim. Acta*, 2011, **692**, 1.

