


Cite this: *RSC Adv.*, 2021, **11**, 26493

Modification of a nitrocellulose membrane with nanofibers for sensitivity enhancement in lateral flow test strips†

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Lateral-flow analysis (LFA) is a convenient, low-cost, and rapid detection method, which has been widely used for screening of diseases. However, sensitivity enhancement in LFA is still a focus in this field and remains challenging. Herein, we propose an electrospinning coating method to modify the conventional nitrocellulose (NC) membrane and optimize the liquid flow rate for enhancing the sensitivity of the NC based LFA strips in the detection of human chorionic gonadotropin (HCG) and luteinizing hormone (LH). It can be seen that coating the NC membrane with nitrocellulose fibers could obtain a NC based strip with HCG and LH detection limits of 0.22 and 0.36 mIU mL⁻¹ respectively, and a quantitative linear range of 0.5–500 mIU mL⁻¹. The results show that electrospinning is effective in modifying conventional NC membranes for LFA applications.

Received 6th June 2021

Accepted 23rd July 2021

DOI: 10.1039/d1ra04369b

rsc.li/rsc-advances

1 Introduction

Lateral-flow analysis (LFA) strips, as a typical low-cost, miniaturized, user friendly, robust, and equipment-free method, are an analysis platform that is ideally suited for point of care diagnostics, and has broad applications in many fields, including food safety,^{1–3} water resources,^{4–6} medical safety and body health.^{7–9} At present, several common LFAs, such as ovulation test strips,¹⁰ pregnancy test strips,^{11,12} and urine test strips¹³ are sold in hospitals and pharmacies and are successful cases of LFA commercialization. However, the limited sensitivity of LFA is still a concern and remains challenging in applications.

Research in this field has developed various methods for improving the sensitivity of LFA, including sample pretreatment,^{14–17} modification of labeled antibody nanoparticles,^{18–22} nanozyme signal enhancement,^{23–26} and targeted fixation of antibodies.²⁷ Besides, many researches have been done focusing on modification of the key component of the test strip, *i.e.*, the nitrocellulose (NC) membrane by introducing functional groups onto the surface or increase the specific surface area to improve the sensitivity of the test strip.^{28,29} However, these signal-amplification methods often need additional equipment,

complex operations, or specialized personnel, most of which are time-consuming and unsuitable for onsite detection.

Recently, reducing the flow rate of the sample solution in the detection shows positive to enhance sensitivity of test strips.^{30–35} For example, sponge shunt was integrated into LFA to decrease the fluid flow rate to achieve a 10-fold signal enhancement in nucleic acid testing of HBV.³² Additionally, membrane materials made by electrospinning was applied in LFAs to increase sensitivity.^{36,37} Electrospinning is a simple and cost-effective method of generating fibers and nanomembranes with great porosity and large surface-to-mass ratio. Yew *et al.*³⁸ electrospun polycaprolactone (PCL) fibers on the surface of commercially available nitrocellulose membrane to reduce the liquid flow rate on the membrane, thereby fulfilled the ultimate aims of increasing detection sensitivity.

Herein, a layer of nitrocellulose fibers were electrospun on the surface of commercially available NC membranes to adjust the porosity for improving the protein adsorption performance and reducing the flow rate to increase the opportunity of antigen–antibody reaction. The electrospin-coated nitrocellulose (ENC) membrane was used as a matrix in LFA for detecting antibody/antigen, including human chorionic gonadotropin (HCG) and luteinizing hormone (LH) antigen (Fig. 1). We predicted that this ENC as the support of LFA strips might provide a potential reference for the preparation of membranes with high sensitivity.

2 Experimental

2.1 Materials

Acetone, *N,N*-dimethylformamide (DMF), sucrose, Tween-20, polyethylene glycol 20000 (PEG-20000), chloroauric acid (HAuCl₄)

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† Electronic supplementary information (ESI) available. See DOI: 10.1039/d1ra04369b



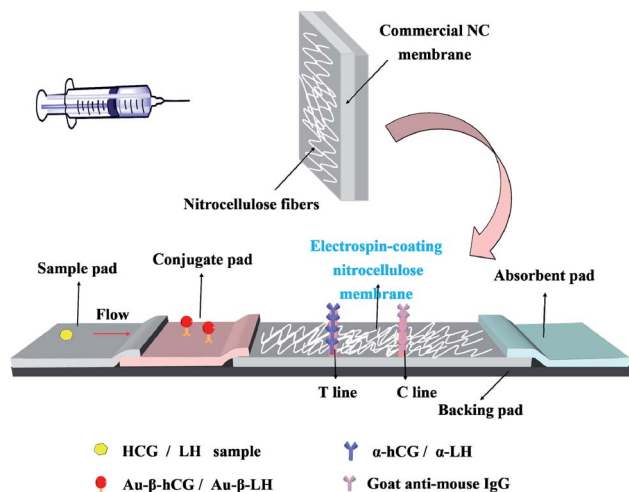


Fig. 1 Schematics of electrospin-coating nitrocellulose membrane on commercial nitrocellulose membrane for LFA strips.

and sodium citrate ($\text{Na}_3\text{C}_6\text{H}_5\text{O}_7$) were purchased from Sinopharm Chemical Reagent Co., Ltd (China). Nitrocellulose (NC) was purchased from Xingping Baotashan Coating Co., Ltd (Shaanxi, China). PVC backing plate, sample pad, conjugate pad, absorbent pad, α -hCG antibody, β -hCG antibody, β -LH antibody, α -LH antibody, goat anti-mouse IgG, commercial nitrocellulose (CNC) membrane and bovine serum albumin (BSA) were purchased from Shanghai JieYi Biotechnology Co., Ltd (Shanghai, China). Human chorionic gonadotropin (HCG), luteinizing hormone (LH) and follicle-stimulating hormone (FSH) were purchased from Shanghai Sangon Biotech Co., Ltd (Shanghai, China).

2.2 Coupling colloidal gold particles and monoclonal antibodies

2.2.1 Preparation of AuNPs. By rapidly adding a certain amount of sodium citrate solution into a sufficiently boiling chloroauric acid solution, narrowly distributed colloidal gold particles (AuNPs) were prepared. Briefly, 0.01% (w/w) chloroauric acid aqueous solution was preheated to 105 °C. Then 3 mL of 1% (w/w) sodium citrate solution was quickly added under stirring. When the color of the solution became red, the heating was stopped for cooling the solution to room temperature under stirring. Finally, the AuNPs were extracted and filtered to obtain a purified AuNPs solution which was stored in a brown bottle at 4 °C for further use.

2.2.2 Preparation of AuNPs labeled β -hCG/ β -LH monoclonal antibody (Au-mAb) conjugates. Electrostatic adsorption method was used to prepare Au-mAb conjugates. The freshly prepared AuNPs solution with β -hCG antibody was shaken for 2 h. Then, 5% BSA and 10% PEG 20000 was added, centrifuged (11 000 rpm, 10 min) and dissolved in Au-mAb storage solution. Finally, the conjugate was stored at 4 °C for further use. Similarly, AuNPs labeled β -LH monoclonal antibody conjugate was obtained.

2.3 Modification of CNC membrane by electrospinning coating with NC

In a typical experiment, a certain amount of fully dried nitrocellulose raw material was added into a mixed solvent of

acetone and DMF, and stirred at room temperature for 24 h to obtain a clear yellowish solution with a concentration of 26%. CNC membranes were carefully placed on the receiver and a syringe containing the spinning solution was mounted on the syringe pump. The electrospinning process was carried out at room temperature for a given time. According to the spinning time, including 1 min, 2 min, 3 min, 6 min, the electrospinning coated CNC (ENC) membranes were named ENC-1, ENC-2, ENC-3, ENC-6, respectively.

2.4 Performance testing of the membranes

2.4.1 Flow rate measurement. AuNP solution was used to test the flow rate of the membranes (4 cm \times 0.5 cm) by recording the time of the solution flowing through the membrane, which was repeated three times. According to the time, the suitable membrane was selected to assemble the strip.

2.4.2 Evaluation of protein capacity on the membranes. The protein adsorption rate of NC membranes were determined by Lowry method. The values of different NC membranes are different, which are determined by a UV-Vis spectrophotometer using the following formula (eqn (1)).

$$\text{Adsorption rate (\%)} = \frac{\bar{A}_0 - \bar{A}}{\bar{A}_0} \times 100\% \quad (1)$$

where \bar{A}_0 and \bar{A} are the absorbance of membranes before and after adsorption, respectively.

2.5 Preparation of the LFA strips

The LFA strip consists of sample pad, conjugate pad, ENC membrane, absorbent pad, and PVC backing plate. With the use of a gold spraying instrument (HM3030, Shanghai Jinbiao Biotechnology Co., Ltd.), α -hCG/ α -LH antibody and goat anti-mouse IgG antibody were immobilized in a defined test line (T-line) and control line (C-line) on the ENC membrane, respectively. Then, Au-mAb conjugate was dispensed on the conjugate pad. The strips were cut into 3.3 mm and stored in dry conditions.

2.6 Assay procedure

PBS buffer solution was used to dilute the standard HCG and LH samples into different concentrations (0.25, 0.5, 1, 5, 25, 50, 100, 500 mIU mL⁻¹). 100 μ L HCG/LH dilution samples were dropped on the sample pad and observed for the detection results after 15 min. The lowest detection limit (LOD) was defined as 3 times the blank/slope standard deviation ($S/N = 3$).

2.6.1 Sensitivity test. In order to get the sensitivity results, different concentrations of HCG sample solutions (0.25, 0.5, 1, 5, 25, 50, 100, 250, 500 mIU mL⁻¹) and LH sample solutions (0.25, 0.5, 1, 5, 25, 50, 100, 250, 500 mIU mL⁻¹) were used for testing. The images were captured by camera of mobile phone.

2.6.2 Specificity test. Three proteins, human chorionic gonadotropin (HCG), luteinizing hormone (LH), follicle-stimulating hormone (FSH) were selected for specific tests, and the concentrations of which were all the same.



2.7 Testing and characterization

The morphology of the membranes was investigated using a S-4800 field emission scanning electron microscope (Hitachi Japan) at an accelerating voltage of 5 kV. To increase the conductivity of the sample, the test sample was sputter-coated with gold before testing. Water contact angles (CA) of the NC membranes were measured with a deionized water droplet of 5 μ L on a video optical contact angle system (OCA 20, Data-physics, Germany), and images were captured with a digital camera. The morphology and particle size of the AuNPs and Au-mAb conjugate were measured using FEI Tecnai G2 F20 S-TWIN Transmission Electron Microscope (US FEI Company). The characteristic absorption peaks of the prepared AuNPs and Au-mAb conjugate were characterized by a UV-Vis-NIR

spectrophotometer (Cary 5000 Agilent USA). The porosity and OD value, which were analyzed by Image J software. The coverage fraction of the electrospinning fibers on the ENC membranes were obtained by using Photoshop software.

3 Results and discussion

3.1 NC membranes morphology

Fig. 2a–d shows that increasing the electrospin-coating time increased the number of fibers on the surface of the membrane. The fiber diameters are uniform, while the CNC membrane shows porous structure (Fig. 2e). The coverage fraction as well as the mass of NC fibers on the ENC membranes also increased with the increase of electrospinning time (Fig. 2f). The covering of NC fibers might change the liquid flow channel of the CNC

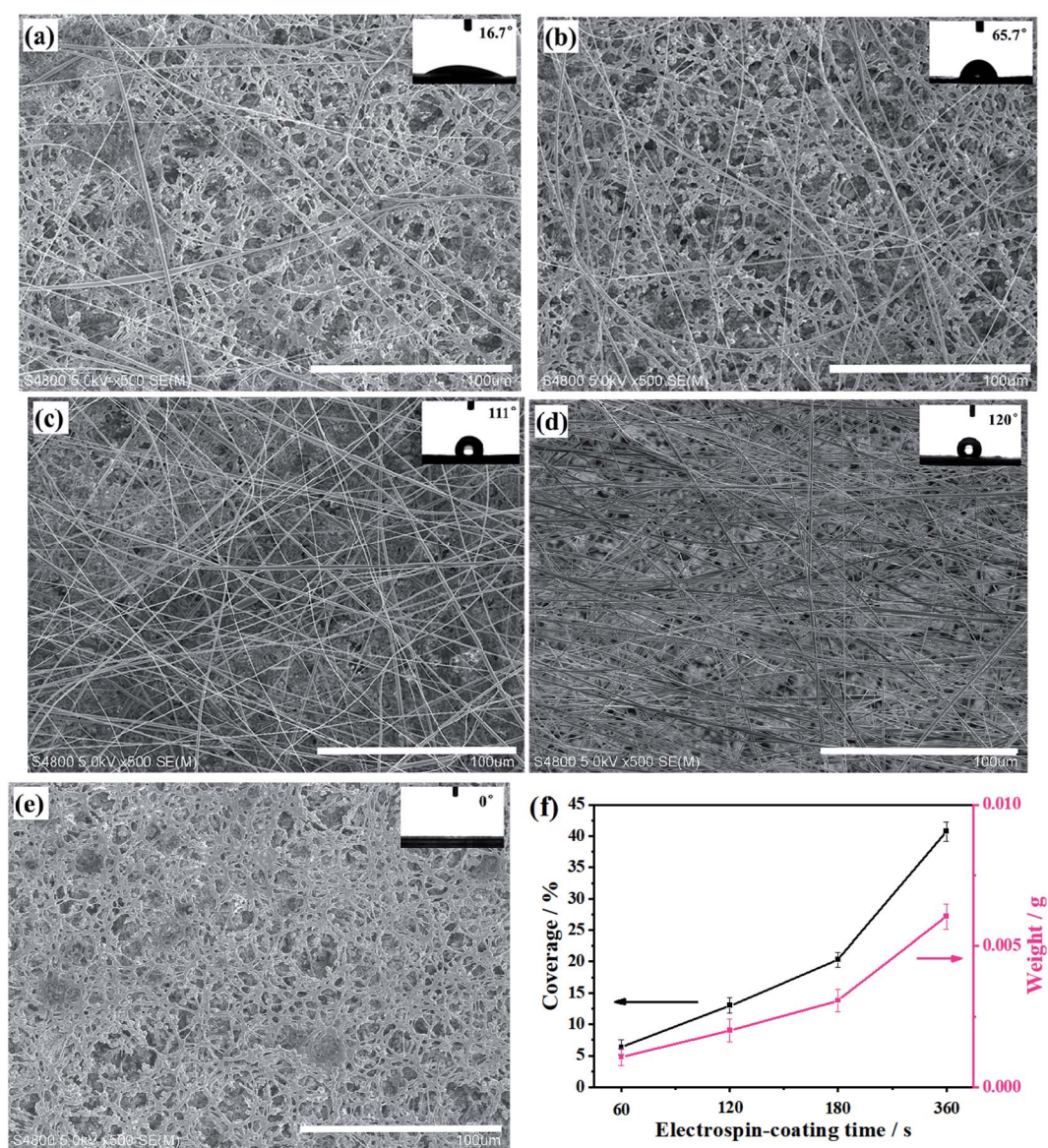


Fig. 2 SEM images of the ENC membranes and the corresponding diameter distributions: (a) ENC-1, (b) ENC-2, (c) ENC-3, (d) ENC-6; (e) SEM image of the CNC membrane; (f) the relationship between the electrospin-coating time of the membrane fibers and the coverage fraction and weight increase of membrane, respectively.

membranes and the surface properties. It is recognized that water droplets spread easily on the CNC membrane due to its hydrophilicity and the capillary effect. However, water CA increased with increasing the electrospinning time (insets in Fig. 2a–e), it was found that the CA duration of 0 s, 60 s, 120 s, 180 s, 360 s were 0°, 16.7°, 65.7°, 111°, 120°, indicating that the membranes were changed from hydrophilic to hydrophobic.

3.2 Flow rate measurement of NC membrane

In traditional immunoassay, due to such limitations of CNC membrane as flow rate, porosity, protein adsorption rate and so on, the amount of antibody adsorbed on surface of the membrane is very less. And when the protein flows through the CNC membrane, it has not enough time for antibody get specific reaction, which will affect the sensitivity of immune reaction. Therefore, we expect that electrospin-coating of fibers on the membrane would increase the porosity and decrease the liquid flow rate, thus increase the protein adsorption, and ends up to enhance the sensitivity of the strips (Fig. 3a).

Fig. 3b shows the relationship between electrospinning time and porosity. When the electrospinning time is 0, *i.e.*, the sample is CNC membrane, the porosity is close to 40%. When the time increases, the fiber accumulation increases, the porosity reached 55% in 2 min. However, as the time goes on,

the porosity increases first and then decreases, this is because overly accumulated fibers covered or blocked the pores of the membrane.

The histogram in Fig. 3c shows the difference of membrane velocity between CNC and ENC membrane. It can be seen that it takes 60 s for the liquid to cross the whole CNC membrane, while it takes more than 60 s for the ENC membrane. This indicates that the time of chromatography of ENC membrane is longer than that of the CNC membrane, which corresponds to the porosity changes in Fig. 3b. Fig. 3d shows that the adsorption rate of CNC membrane is 5.3%, and ENC-2 membrane is close to 12%, which is the highest in protein capacity and protein adsorption performance, it provides great convenience for immunoassay of ICT bands.

3.3 Test performance for HCG and LH detection

3.3.1 Influence of the electrospinning coating on the signal strength of the strips. To research the effect of electrospinning time on LFA signal strength, the LFA strips were assembled with different spinning time membranes. By detecting HCG and LH antigen with the concentration of 25 mIU mL⁻¹, and the different OD values of T-line were obtained by Image J software. Fig. 4 shows the signal strength of the strips after detecting HCG and LH as well as their corresponding OD values. It was

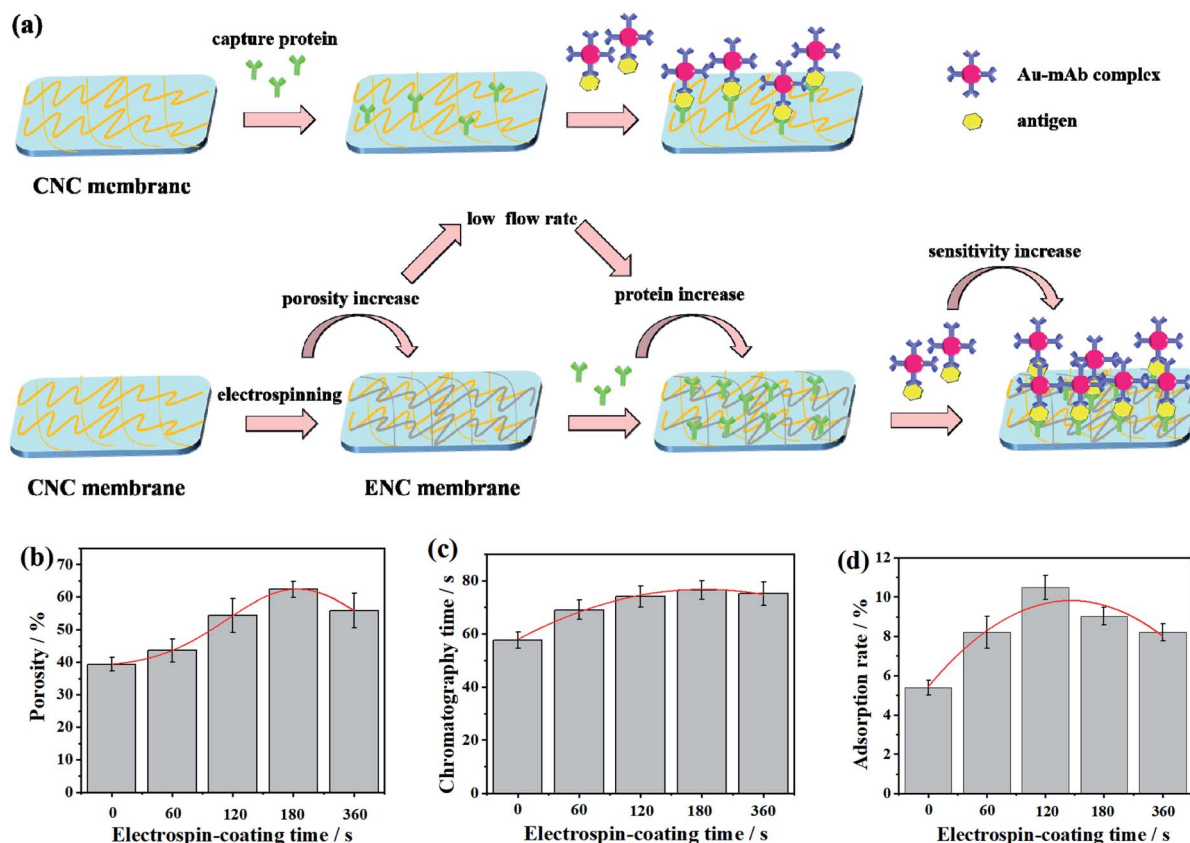


Fig. 3 (a) A sandwich immunoassay with a conventional immunochromatographic strip (top) and an immunochromatographic strip with electrospin-coating NC membrane using antibody-conjugated Au nanoparticles (AuNPs) (bottom). (b) The relationship between the electrospin-coating time and porosity. (c) The relationship between the electrospin-coating time and chromatographic time. (d) The relationship between the electrospin-coating time and protein adsorption rate.



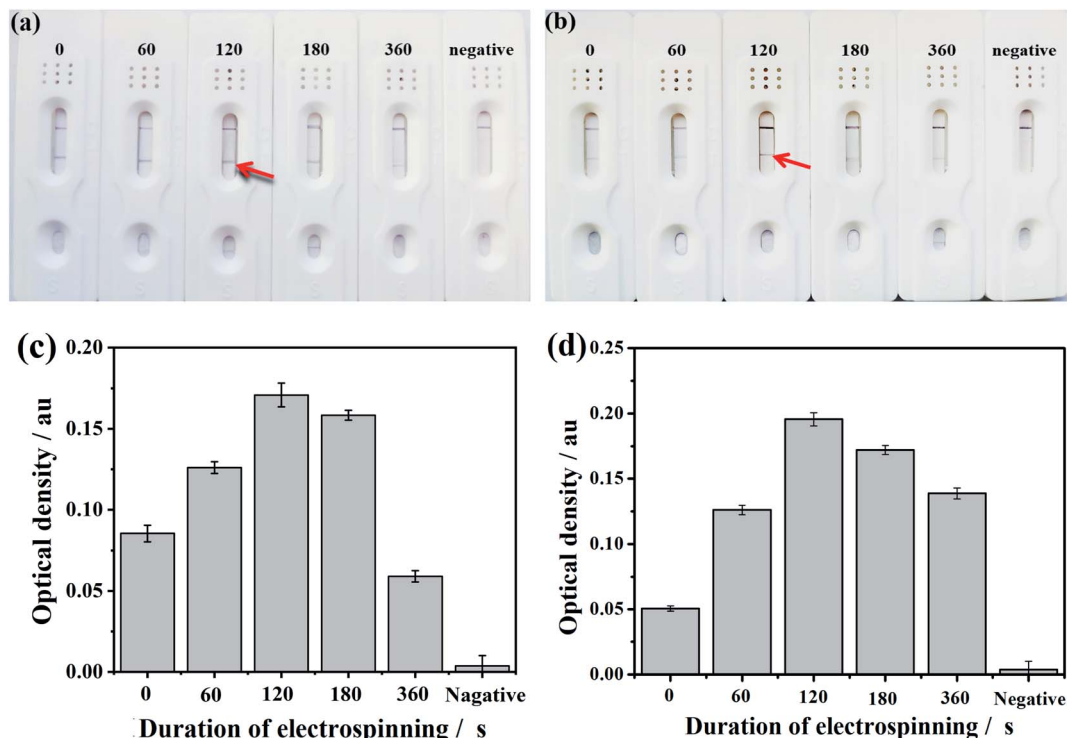


Fig. 4 (a) CNC and ENC based LFA test strips after detecting HCG in which the number means the electrospinning time. (c) OD values corresponding to the T-lines in (a). (b) CNC and ENC based LFA test strips after detecting LH in which the number means the electrospinning time. (d) OD values corresponding to the T-lines in (b). The upper signal in the test strip represents the C-line, and the lower signal represents the T-line.

found that the test strip using the ENC-2 membrane with electrospinning coating time of 120 s shows the strongest signal as well as the highest OD value of 0.17 and 0.20 a.u. separately, which indicates that this kind of membrane is the most suitable

for assembling LFA test strip, because the stronger the OD signal is, the easier the test result can be observed by naked eyes.

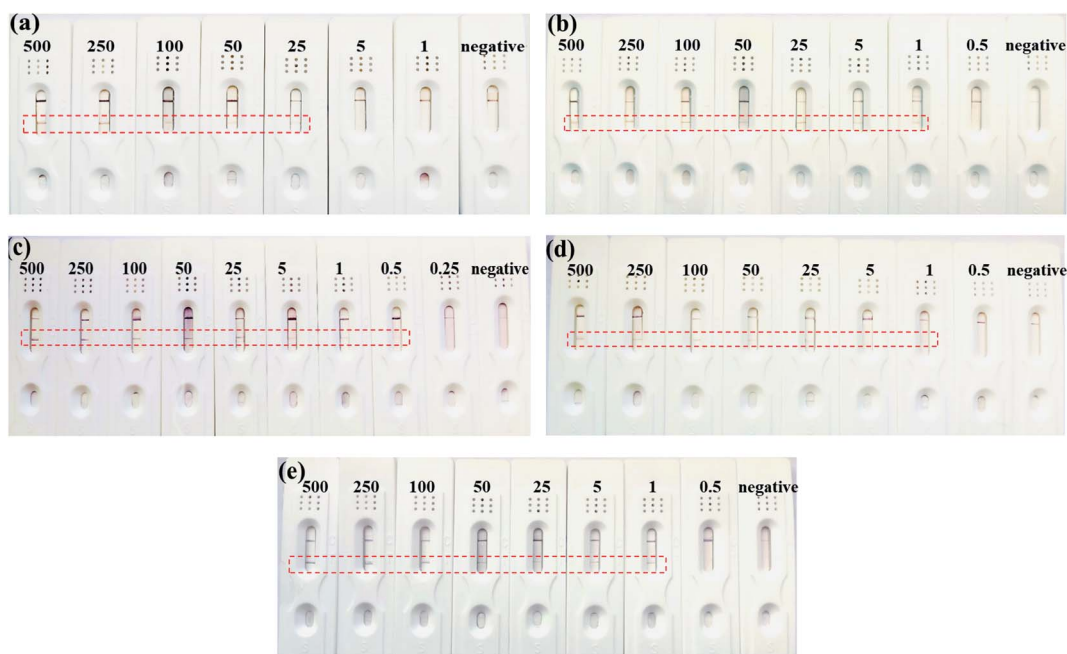


Fig. 5 The sensitivity test of LFA strips based on the (a) CNC, (b) ENC-1, (c) ENC-2, (d) ENC-3, and (e) ENC-6 by detecting HCG antigen, and the number means the concentration of the HCG antigen, with the unit of mIU mL⁻¹. The upper signal in the test strip represents the C-line, and the lower signal represents the T-line.

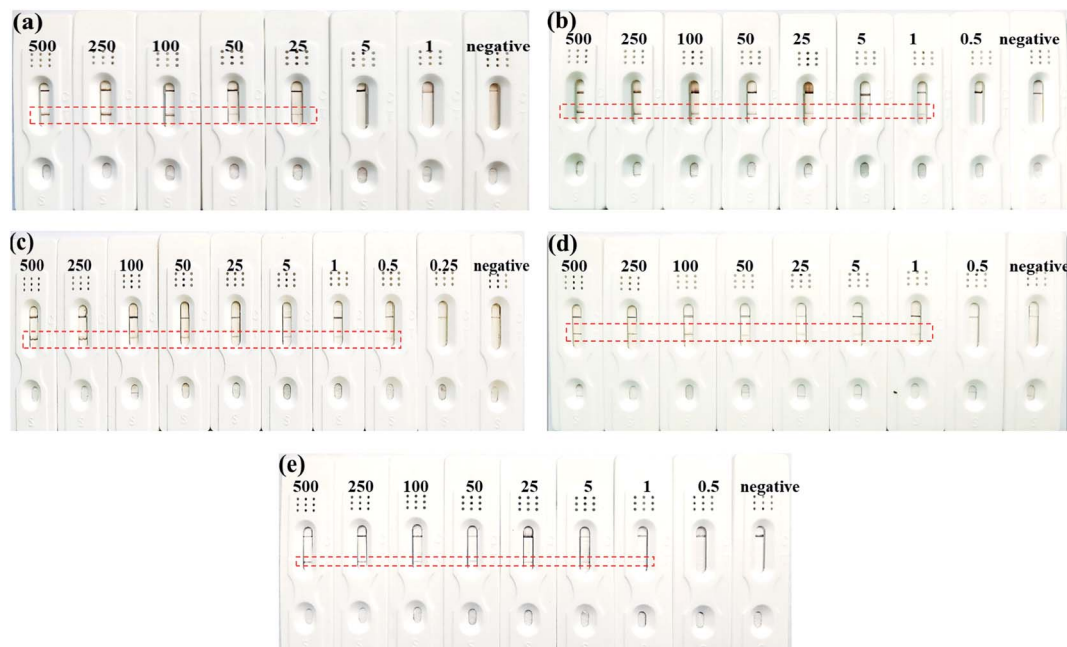


Fig. 6 The sensitivity test of LFA strips based on the (a) CNC, (b) ENC-1, (c) ENC-2, (d) ENC-3, and (e) ENC-6 by detecting LH antigen, and the number means the concentration of the HCG antigen, with the unit of mIU mL^{-1} . The upper signal in the test strip represents the C-line, and the lower signal represents the T-line.

3.3.2 Determination of the concentration of HCG and LH antigen. Fig. 5 show the sensitivity results of LFA strips by detecting HCG antigen. It was found that decreasing the concentration of HCG gradually decreased the color depth of the T-line of all the test strips. The CNC based strips display T-lines when it was decreased to 25 mIU mL^{-1} , while all the ENC based strips display T-lines at 1 mIU mL^{-1} . When it was reduced to 0.5 mIU mL^{-1} , only the T and C lines can be successfully displayed for the ENC-2 test strip. When it was decreased to 0.25 mIU mL^{-1} , the T-line disappeared. Therefore, the minimum detection limit for ENC-2 is 0.5 mIU mL^{-1} . Inspiringly, the sensitivity test of LFA strips based on CNC and ENC membranes by detecting LH antigen shows similar results (Fig. 6) to the detection of HCG. Compared to the minimum detection limit of CNC test strips, the sensitivity is increased by 50 times.

The enhancement of the sensitivity was obviously resulted from the electrospinning coating of NC fibers on the CNC

membrane. This might be because that in one hand, the coating of electrospun NC fibers blocked some of the surface pores and then reduced the porosity of the membrane. In the other hand, the NC fibers coating improved the hydrophobicity of the membrane. Therefore, this coating extends the flow time of the solution on the membrane and increases the specific reaction time of the antigen and antibody in the ENC membrane, which can increase the sensitivity. However, overly loaded NC fibers on the membrane by extending the electrospinning time showed adverse effect. Therefore, the minimum detection limit of ENC-3 and ENC-6 only reached 1 mIU mL^{-1} . This might be because overly loaded NC fibers decreased the protein absorption due to overly increased hydrophobicity.

Fresh urine samples from a pregnant woman and a non-pregnant woman were taken for detection respectively. The results of ENC-2 based test strip showed that the urine samples of pregnant women were positive, while the urine samples from

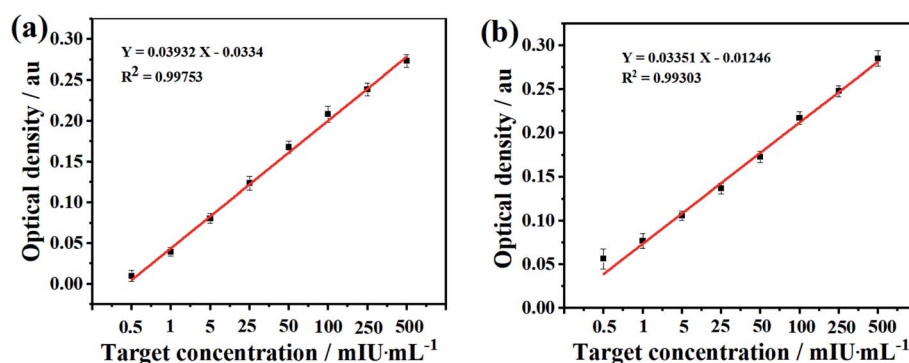


Fig. 7 Correlation between concentration of (a) HCG and (b) LH and its optical density value.



Table 1 An overview on recently reported nanomaterial-based optical methods for HCG

Method applied	Linear range	LODs ^a	References
Electrochemical	10–1000 pg mL ⁻¹	5 ^b	Viet <i>et al.</i> 2019 (ref. 39)
Electrochemical immunoassay	0.0212–530 mIU mL ⁻¹	0.0066 ± 0.02	Roushani <i>et al.</i> 2016 (ref. 40)
Novel immunoassay	0.03–1000 mIU mL ⁻¹	0.19	Chen <i>et al.</i> 2019 (ref. 41)
Enzyme-catalyzed	0.5–400 mIU mL ⁻¹	0.36	Zhang <i>et al.</i> 2019 (ref. 42)
ICAs	9–2304 mIU mL ⁻¹	9	Zhang <i>et al.</i> 2019 (ref. 43)
Transistor immunosensors	1–1 ng mL ⁻¹	1 ^b	Haslam <i>et al.</i> 2018 (ref. 44)
Photothermal immunosensing	0.01–100 ng mL ⁻¹	5.8 ^b	Hong <i>et al.</i> 2019 (ref. 45)
LFA	0.5–500 mIU mL ⁻¹	0.22	This work

^a mIU mL⁻¹. ^b pg mL⁻¹.

non-pregnant women were negative. At the same time, when the urine sample from pregnant women was diluted from 100% to 50%, 25%, 10% and 5%, there were still T-line appeared, which indicated that the test strip is feasible to detect real urine samples.

3.3.3 Linear range and detection limit. In order to further quantify the LOD values of HCG and LH, the detection standard curves of the two antigens were made (Fig. 7), both of which show linearity with the concentration from 0.5 to 500 mIU mL⁻¹. The linear regression equations for HCG is $Y = 0.03932X - 0.0334$ ($R^2 = 0.99753$) and that of LH is $Y = 0.03351X - 0.01246$ ($R^2 = 0.99303$), and the corresponding LOD was 0.22 and 0.36 mIU mL⁻¹, respectively. Obviously, the ENC based test strips shows linear range and LOD on par with or even showing advantage over previous works (Table 1). Importantly, our

method needs no auxiliary instruments to obtain test results. Therefore, it is easy to achieve the purpose of real-time detection.

3.3.4 Specificity analysis. In addition to the sensitivity test, the specific evaluation of the strip is also one of the most important parameters. As shown in Fig. 8a, except for HCG antigen, there is no T-line in other similar antigens, indicating that there is no non-specific binding, so the test strip is indeed specific for HCG measurement. Similarly, as shown in Fig. 8b, except for the LH antigen, the other antigens did not show T-line. Therefore, The ENC membrane can be used to assemble strips to detect HCG and LH separately, which can achieve high specificity.

In order to further show the specificity ability, we detected the OD values of different T-line. As can be seen in Fig. 8c, the

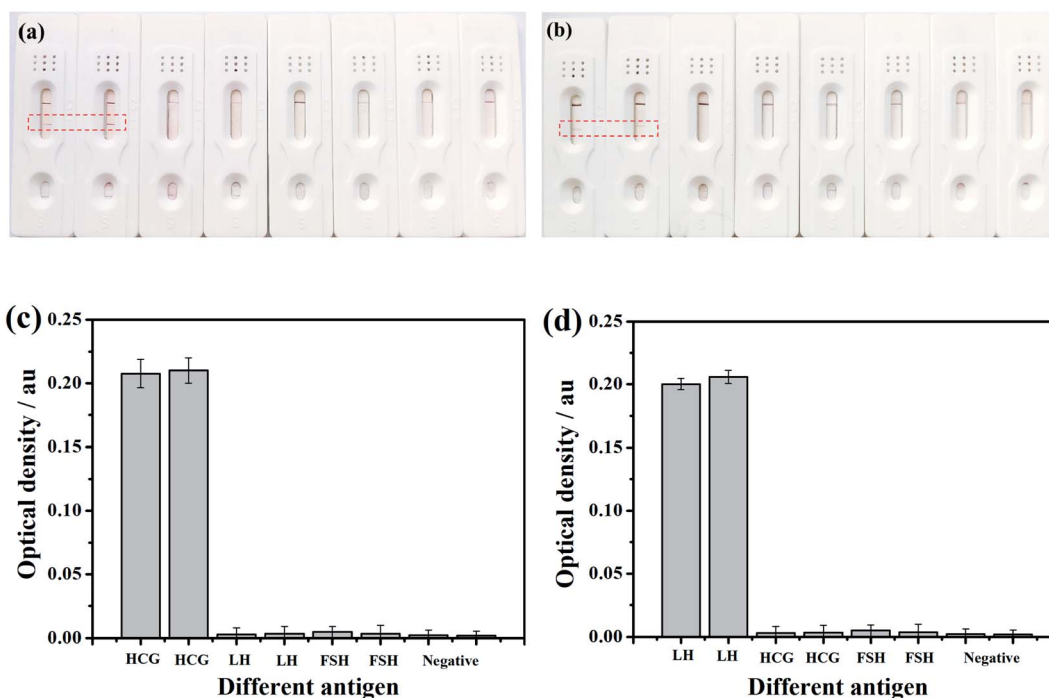


Fig. 8 (a) ENC-2 based LFA test strips was specific for HCG antigen after detecting different antigens. (c) OD values corresponding to the T-lines in (a). (b) ENC-2 based LFA test strips was specific for LH antigen after detecting different antigens. (d) OD values corresponding to the T-lines in (b). The upper signal in the test strip represents the C-line, and the lower signal represents the T-line.



OD value of HCG sample reached 0.20 a.u., which means high specificity property. Similarly, the LH test strip also has a good specific test result as shown in Fig. 8d.

3.3.5 Reproducibility test. In addition to sensitivity and specificity tests, the repeatability of the strip was also evaluated. HCG samples with concentrations of 25, 5, 1, 0.5 mIU mL⁻¹ and blank samples were tested for three times. It was found that when the HCG concentration is 25, 5, 1 and 0.5 mIU mL⁻¹ respectively, T-line and C-line can appear, and with the same concentration, the brightness of T-line and C-line of the three strips were similar (Fig. S3†). When the blank sample is taken for testing, only C-line appears on the three test strips, and the brightness is close, which indicates that the test strips are repeatable.

Besides, this method has advantages compared with the other conventional methods for LFA, which utilizes the chemical properties of nanofibers to adsorb protein and adopts electrospinning technology to change the porosity of nanofibers for adjusting the protein load and flow rate, so as to improve the sensitivity. In addition, the electrospinning technology is a mature technology for preparing nano and micro fiber membranes. The membranes have high porosity, large specific surface area and uniform diameter distribution, showing high application value in biomedical,^{46,47} environmental engineering and textile fields. However, the traditional LFA method is different from this method in preparation process, so it has some shortcomings in regulating porosity, flow rate and protein loading.

4 Conclusion

A new AuNPs test strip was developed by electrospinning NC fibers on the surface of the CNC membrane to control flow rate with high sensitivity and selectivity. It was shown that the LODs for HCG and LH were 0.22 and 0.36 mIU mL⁻¹, respectively, and the linear detection range was from 0.5 to 500 mIU mL⁻¹. The sensitivities were significantly improved compared to the strip based on CNC membrane with an increased by 50 times. Therefore, this method has the possibility of greatly reducing the minimum detection limit and effectively increasing the sensitivity of instant detection. The AuNPs test strips are hopeful for satisfying the purpose for point of care testing, especially in resource-limited regions, which has broad application prospects.

Ethical statement

The authors state that all experiments were performed in compliance with the relevant laws and institutional guidelines. The human urine samples were obtained from volunteers at the University of Science and Technology, and informed consent was obtained from all the volunteers.

Author contributions

Xue Wang: responsible for the design, experiment, draft of the manuscript. Chao-Hua Xue: organized, supervised and guided

the whole work. Dong Yang: provided experimental materials and guided the experiment. Shun-Tian Jia: provided instrument guidance and characterization assistance. Ya-Ru Ding: assisted testing and characterization. Lei Lei: assisted testing. Ke-Yi Gao: assisted characterization. Tong-Tong Jia: assisted experiments.

Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This work was supported by Natural Science Foundation of Shaanxi Province (2020JM-506), National Natural Science Foundation of China (51572161), major project of Ministry of Science and Technology of China (2017YFB0307700), the National Natural Science Foundation of China (21505089), and Postdoctoral Research Foundation of China (2017M13038).

References

- 1 H. Duan, X. L. Huang, Y. N. Shao, L. Y. Zheng, L. Guo and Y. H. Xiong, *Anal. Chem.*, 2017, **89**, 7062–7068.
- 2 Y. Chen, L. Guo, L. Liu, S. Son, H. Kuang and C. Xu, *J. Agric. Food Chem.*, 2017, **65**, 8248–8255.
- 3 O. Y. Hui, X. M. Tu, Z. F. Fu, W. Wang, S. Fu, C. Zhu, D. Du and Y. Lin, *Biosens. Bioelectron.*, 2018, **106**, 43–49.
- 4 X. Liu, J. J. Xiang, Y. Tang, X. L. Zhang, Q. Q. Fu, J. H. Zou and Y. Lin, *Anal. Chim. Acta*, 2012, **745**, 99–105.
- 5 K. Kim, H. A. Joung, G. R. Han and M. G. Kim, *Biosens. Bioelectron.*, 2016, **85**, 422–428.
- 6 S. Cinti, L. Fiore, R. Massoud, C. Cortese, D. Moscone, G. Palleschi and F. Arduini, *Talanta*, 2018, **179**, 186–192.
- 7 L. Li, D. Yin, K. Xu, Y. Liu, D. Song, J. Wang, C. Zhao, X. Song and J. Li, *J. Pharm. Biomed. Anal.*, 2017, **141**, 79–86.
- 8 L. Zhang, Y. Chen, N. Cheng, Y. Xu, K. Huang, Y. Luo, P. Wang, D. Duan and W. Xu, *Anal. Chem.*, 2017, **89**, 10194–10200.
- 9 W. B. Wang, L. Q. Liu, S. S. Song, L. G. Xu, H. Kuang, J. P. Zhu and C. L. Xu, *Sci. China Mater.*, 2016, **59**, 665–674.
- 10 A. B. Ayoola, D. Slager, C. Feenstra and G. L. Zandee, *J. Midwifery Womens Health*, 2015, **60**, 604–609.
- 11 D. G. Tincello and D. H. Richmond, *BMJ*, 1998, **316**, 435–437.
- 12 E. Marino, W. R. Threlfall and R. A. Schwarze, *Theriogenology*, 2009, **71**, 877–883.
- 13 J. Hu, L. Wang, F. Li, Y. L. Han, M. Lin, T. J. Lu and F. Xu, *Lab Chip*, 2013, **13**, 4352–4357.
- 14 H. Zhen, S. Hu, Y. H. Xiong, H. Wei, H. Y. Xu, H. W. Duan and W. H. Lai, *TrAC, Trends Anal. Chem.*, 2019, **114**, 151–170.
- 15 S. W. Zhang, T. Q. Yao, S. F. Wang, R. H. Feng, L. Q. Chen, V. V. Zhu, G. P. Hu, H. Zhang and G. W. Yang, *Spectrochim. Acta, Part A*, 2019, **214**, 302–308.
- 16 Q. Chen, M. F. Qie, X. S. Peng, Y. Chen and Y. L. Wang, *RSC Adv.*, 2020, **10**, 3307–3313.



- 17 M. F. Pan, J. Y. Yang, S. J. Li, G. Z. Wang, J. P. Wang and S. Wang, *Anal. Methods*, 2019, **11**, 2027–2032.
- 18 M. Zhang, T. Bu, Y. Tian, X. Sun, Q. Wang, Y. Liu, F. Bai, S. Zhao and L. Wang, *Food Chem.*, 2020, **332**, 127398.
- 19 C. Wang, C. Wang, X. Wang, K. Wang, Y. Zhu, Z. Rong, W. Wang, R. Xiao and S. Wang, *ACS Appl. Mater. Interfaces*, 2019, **11**, 19495–19505.
- 20 X. F. Ruan, Y. J. Wang, E. Y. Kwon, L. M. Wang, N. Cheng, X. H. Niu, S. C. Ding, B. J. Van Wie, Y. Lin and D. Du, *Biosens. Bioelectron.*, 2021, **184**, 113238.
- 21 X. L. Fu, J. H. Wen, J. W. Li, H. Lin, Y. M. Liu, X. M. Zhuang, C. Y. Tian and L. X. Chen, *Nanoscale*, 2019, **33**, 1–8.
- 22 V. Tran, B. Walkenfort, M. Kçnig, O. Salehi and S. Schlcker, *Angew. Chem., Int. Ed.*, 2019, **58**, 442–446.
- 23 D. W. Bradbury, M. Azimi, A. J. Diaz, A. A. Pan, C. H. Falktoft, B. M. Wu and D. T. Kamei, *Anal. Chem.*, 2019, **91**, 12046–12054.
- 24 Z. Y. Lyu, S. C. Ding, N. Zhang, Y. Zhou, N. Cheng, M. Y. Wang, M. Xu, Z. X. Feng, X. H. Niu, Y. Cheng, C. Zhang, D. Du and Y. H. Lin, *Research*, 2020, **11**, 1–11.
- 25 X. H. Niu, X. Li, Z. Y. Lyu, J. M. Pan, S. C. Ding, X. F. Ruan, W. L. Zhu, D. Du and Y. H. Lin, *Chem. Commun.*, 2012, **77**, 1–17.
- 26 S. C. Ding, Z. Y. Lyu, L. Z. Fang, T. Li, W. L. Zhu, S. Q. Li, X. Li, J. C. Li, D. Du and Y. H. Lin, *Small*, 2021, **17**, 25.
- 27 Y. Ryu, Z. W. Jin, M. S. Kang and H. S. Kim, *BioChip J.*, 2011, **5**, 193–198.
- 28 R. H. Tang, L. N. Liu, S. F. Zhang, A. Li and Z. D. Li, *Microchim. Acta*, 2019, **186**, 831–838.
- 29 B. Zhang, W. Gao, J. Piao, Y. Xiao, B. Wang, W. Peng, X. Gong, Z. Wang, H. Yang and J. Chang, *ACS Appl. Mater. Interfaces*, 2018, **10**, 14549–14558.
- 30 J. R. Choi, K. W. Yong and R. Tang, *Adv. Healthcare Mater.*, 2016, **6**, 1600920–1600928.
- 31 J. R. Choi, Z. Liu, J. Hu, R. Tang, Y. Gong, S. Feng, H. Ren, T. Wen, H. Yang, Z. Qu and F. Xu, *Anal. Chem.*, 2016, **88**, 6254–6264.
- 32 R. Tang, H. Yang, Y. Gong, Z. Liu, X. Li, T. Wen, Z. Qu, S. Zhang, Q. Mei and F. Xu, *Sci. Rep.*, 2017, **7**, 1360–1369.
- 33 C. Parolo, M. S. Mariana and M. Arben, *Lab Chip*, 2013, **13**, 386–390.
- 34 B. J. Toley, B. McKenzie, T. Liang, J. R. Buser, P. Yager and E. Fu, *Anal. Chem.*, 2013, **85**, 11545–11552.
- 35 M. Shen, Y. Y. Chen, M. Zhu and Y. Xu, *Anal. Chem.*, 2019, **91**, 4814–4820.
- 36 S. J. Reinholt, A. Sonnenfeldt, A. Naik, M. W. Frey and A. J. Baeumner, *Anal. Bioanal. Chem.*, 2014, **406**, 3297–3304.
- 37 Y. H. He, S. H. Hong, M. Wang, J. Wang, A. M. Abd El-Aty, J. Wang, H. Ahmet, M. Khan and Y. She, *New J. Chem.*, 2020, **44**, 6026–6036.
- 38 C. T. Yew, P. Azari, J. R. Choi and F. Li, *Anal. Chim. Acta*, 2018, **1009**, 81–88.
- 39 N. X. Viet, N. X. Hoan and Y. Z. Takamura, *Mater. Chem. Phys.*, 2019, **227**, 123–129.
- 40 M. Roushani and A. Valipour, *Microchim. Acta*, 2016, **183**, 845–853.
- 41 J. Chen, L. W. Hao, Y. Q. Wu, T. Lin, X. Li, Y. K. Leng, X. L. Huang and Y. H. Xiong, *Chem. Commun.*, 2019, **55**, 10312–10315.
- 42 Z. Zhang, G. L. Xu, L. Xie and Y. P. Guan, *Microchim. Acta*, 2019, **186**, 581–589.
- 43 W. Zhang, H. Duan, R. Chen, T. Ma, L. Zeng, Y. Leng and Y. Xiong, *Talanta*, 2019, **194**, 604–610.
- 44 C. Haslam, S. Damiani, T. Whitley, P. Davey, E. Ifeachor and S. A. Awan, *Diagnostics*, 2018, **8**, 5–17.
- 45 G. Hong, D. Zhang, Y. He, Y. Yang, P. Chen, H. Yang, Z. Zhou, Y. Liu and Y. Wang, *Anal. Bioanal. Chem.*, 2019, **411**, 6837–6845.
- 46 I. Cruz-Maya, A. Varesano, C. Vineis and V. Guarino, *Polymers*, 2020, **12**, 1671–1679.
- 47 Y. Guo, J. Gilbert-Honick, S. M. Somers, H. Q. Mao and W. L. Grayson, *Biochem. Biophys. Res. Commun.*, 2019, **516**, 1–7.

