


 Cite this: *RSC Adv.*, 2021, **11**, 24669

 Received 25th May 2021
 Accepted 2nd July 2021

 DOI: 10.1039/d1ra04082k
rsc.li/rsc-advances

Development of an ultrafast fluorescent probe for specific recognition of hypochlorous acid and its application in live cells†

 Zhencai Xu,‡^{ab} Xiaofeng Wang,‡^a Tingting Duan,^a Rong He,^a Fangwu Wang^a and Xuejun Zhou  ^{a*}

Hypochlorous acid (HOCl), a highly potent oxidant of reactive oxygen species, plays critical roles in many physiological and pathological processes. In this work, a novel coumarin-based fluorescent probe, Cou-HOCl, was prepared for the detection of HOCl. The probe exhibited good selectivity over other analytes, excellent sensitivity with a detection limit of 16 nM, and fast response within 5 s. And further study demonstrated that the probe could be used not only to image exogenous HOCl in various cells, but also to determine the fluctuating levels of HOCl in macrophage cells during inflammation.

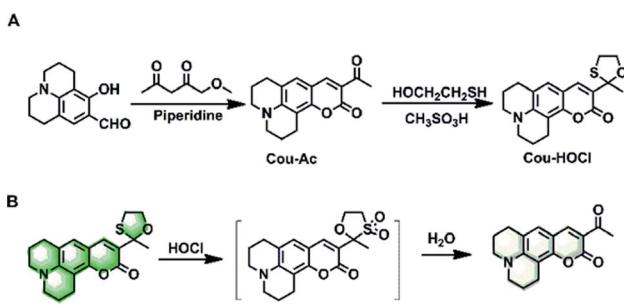
Introduction

Hypochlorous acid (HOCl) is an indispensable reactive oxygen species (ROS) in organisms, which is intimately involved in various pathophysiological processes and has a significant impact on organisms.¹ HOCl is produced in the body by inflammatory cells such as macrophages and neutrophils *via* the reaction of hydrogen peroxide (H₂O₂) and chloride ions (Cl⁻) catalyzed by myeloperoxidase (MPO).^{2,3} HOCl can effectively kill bacteria in cells, which allows HOCl to be used as a powerful tool to resist pathogens that invade the immune system.⁴ However, abnormal production of HOCl may cause oxidative stress, leading to the occurrence of various diseases including inflammation, cardiovascular disease, nervous system disease, and even cancer.⁵⁻⁷ In view of the important role of HOCl in living systems, there is an urgent need to develop tools to study the dynamic changes of HOCl *in vivo*.

Various methods such as electrochemistry, iodometry and colorimetry are available to detect HOCl, but they are very complicated to execute.^{8,9} In recent years, fluorescence imaging technology has developed rapidly and attracted increasing attention because it can be used to study changes of molecules at the cellular level in real time, and do so non-invasively and with high selectivity and sensitivity. Fluorescent probes are the ideal tools for use with fluorescence imaging technology to investigate molecular life activities.¹⁰⁻¹⁴ Many efforts have been

made to develop fluorescent probes of HOCl on the basis of reactions with various sensing moieties including oxime oxidation,^{15,16} selenium oxidation,^{17,18} *p*-aminophenol oxidation,^{19,20} lactam oxidation,²¹⁻²³ double bond oxidation,²⁴⁻²⁶ thioester chlorination,²⁷ and others.²⁸ However, some of these HOCl probes are limited by poor sensitivity, poor photostability, and slow response.^{29,30} Therefore, developing HOCl fluorescent probes featuring advantages such as high sensitivity, high photostability, and rapid response is still very important.

Herein, we describe a simple coumarin-based fluorescent probe, Cou-HOCl, for detecting HOCl based on an HOCl-mediated oxidation deprotection mechanism. Cou-HOCl was obtained by simple condensation of coumarin derivatives and mercaptoethanol (Scheme 1). In the absence of HOCl, Cou-HOCl itself exhibited a strong green fluorescence. After reacting with HOCl, Cou-HOCl showed a dramatic decrease in fluorescence. It was worth noting that Cou-HOCl displayed excellent sensitivity and selectivity. The biological evaluation also demonstrated that Cou-HOCl could be employed to visualize the fluctuating levels of HOCl in cells.



Scheme 1 (A) Synthesis of Cou-HOCl. (B) Reaction of Cou-HOCl with HOCl.

^aDepartment of Otolaryngology, Head and Neck Surgery, The First Affiliated Hospital of Hainan Medical University, Haikou 570102, China. E-mail: xuejunzhouent@hainmc.edu.cn

^bGuanyun People's Hospital, Lianyungang 222000, China

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d1ra04082k

‡ These two authors contributed equally to this work.



Results and discussion

Spectral properties of Cou-HOCl

With the probe in hand, UV-Vis and fluorescence spectra were first acquired. As shown in Fig. 1A, upon being excited with light having a wavelength of 410 nm, Cou-HOCl displayed an emission peak at 510 nm. As the HOCl concentration was increased from 0 to 50 μ M, the fluorescence emission showed a prompt decrease in intensity and a slight red shift. The fluorescence intensity of Cou-HOCl at about 510 nm and the concentration of HOCl (0–50 μ M) showed a predominantly linear relationship ($F_{510\text{ nm}} = -17.99 \times [\text{HOCl}]/\mu\text{M} + 979.83, R^2 = 0.9987$). Based on the standard method of $3\sigma/k$, the detection limit of HOCl was calculated to be 16 nM (Fig. S1†). Free Cou-HOCl yielded a maximum absorption peak at 410 nm, as shown in Fig. 1B. Upon increasing the concentration of HOCl from 0 to 50 μ M, a new absorption peak at 470 nm emerged and increased in intensity with a well-defined isobestic point at 430 nm. The fluorescence intensity changes at 510 nm over time are shown in Fig. 1C. After adding 50 μ M HOCl to the solution of Cou-HOCl, the reaction was basically completed within 5 s, and the fluorescence intensity remained stable over the course of 30 s. The time-dependent changes in fluorescence indicated the ability of the probe to quickly respond to HOCl, and in fact do so more quickly than most of the previously reported HOCl probes.³⁰ Next, we investigated the ability of Cou-HOCl to recognize HOCl in conditions with different pH values from 2.5 to 10.0. As shown in Fig. 1D, the fluorescence intensity at 510 nm remained unchanged in the pH range 6.0–10.0, showing the excellent stability of Cou-HOCl. Throughout the pH range 2.5–10.0, the fluorescence intensity of Cou-HOCl was less in the presence of HOCl than in its absence. The results implied that

Cou-HOCl could be deployed for the detection of HOCl in the physiological pH range of living systems. Various competing analytes such as reactive sulfur species, metal ions, reactive nitrogen species and reactive oxygen species were applied together with fluorescence spectroscopy to investigate the selectivity of Cou-HOCl toward HOCl. As shown in Fig. S2,† the presence of HOCl triggered an apparent fluorescence decrease, while other analytes such as GSH, Cys, Hey, Fe^{2+} , Fe^{3+} , Zn^{2+} , Cu^{2+} , NO , KO_2 , H_2O_2 , $^1\text{O}_2$, HO^\cdot , and *t*-BuOOH did not cause such fluorescence changes. All of these results showed the excellent sensing performance of Cou-HOCl toward HOCl over other competing analytes. To verify the reaction mechanism, we acquired ^1H NMR spectra of Cou-HOCl to which HOCl was added in $\text{CD}_3\text{CN}/\text{H}_2\text{O}$ (v/v = 9/1). As depicted in Fig. S3,† Cou-HOCl was observed to convert to Cou-Ac.

Imaging application of Cou-HOCl

The excellent spectral properties of Cou-HOCl motivated us to investigate its potential applications in live cells. Prior to the cell imaging, the cytotoxic activity of Cou-HOCl was first evaluated by performing Cell Counting Kit-8 (CCK-8) assays. As can be seen from Fig. S4,† Cou-HOCl showed inconspicuous cytotoxicity even at a concentration of 100 μ M in various cells, which indicated that the probe could be applied for cell imaging. Cou-HOCl was incubated with different types of cells including A549, HepG 2 and HeLa cells for 30 minutes and in each case a remarkable green fluorescence was seen (Fig. 2). However, following incubation with 50 μ M of HOCl, the intensity of the green fluorescence decreased rapidly. The results confirmed that Cou-HOCl could sensitively detect HOCl in biological systems.

Subsequently, we set out to investigate the ability of using Cou-HOCl to detect HOCl in inflammatory cells. Different samples of live RAW264.7 cells were first pretreated with different concentrations of HOCl (0, 5, 10, 15, 20, 30, and 50 μ M, respectively), and then each incubated with Cou-HOCl. As shown in Fig. 3A, in the absence of HOCl, the cells loaded with Cou-HOCl showed a strong green fluorescence signal. As the concentration of the exogenous HOCl was increased, the intracellular green fluorescence gradually weakened in a concentration-dependent manner. Compared with the control group, the addition of 50 μ M HOCl reduced the fluorescence intensity at least five fold (Fig. 3B), which indicated that Cou-HOCl could effectively penetrate the cell membrane and be used to detect changes in the level of exogenous HOCl in RAW264.7 cells. To investigate the feasibility of using Cou-HOCl to image endogenous HOCl in inflammatory cells, we chose LPS (an inflammation activator) to stimulate RAW264.7 cells to produce endogenous HOCl. As displayed in Fig. 3C and D, the green fluorescence of the LPS-treated group was weaker than that of the control group. Myeloperoxidase (MPO) is a well-known peroxidase that could catalyze the conversion of hydrogen peroxide and chloride ions to HOCl. To investigate the mechanism of the catalysis of the production of HOCl by MPO, different samples of the cells were treated with MPO, $\text{H}_2\text{O}_2 + \text{Cl}^\cdot$, and MPO + $\text{H}_2\text{O}_2 + \text{Cl}^\cdot$, respectively. For the MPO and H_2O_2

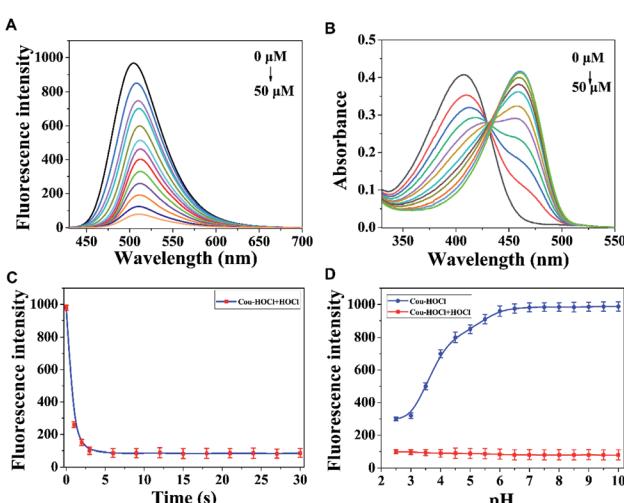


Fig. 1 (A) Fluorescence response of Cou-HOCl (10 μ M) toward HOCl (0–50 μ M). (B) Absorption spectra of Cou-HOCl (10 μ M) with HOCl (0–50 μ M). (C) Time course of the fluorescence intensity of Cou-HOCl before and after adding 50 μ M HOCl. (D) Fluorescence responses of Cou-HOCl toward HOCl at various pH levels. All of the data were collected from the samples in $\text{CH}_3\text{CN}/\text{PBS}$ solutions (10 mM, v/v = 3/7, pH = 7.4). $\lambda_{\text{ex}} = 410$ nm.



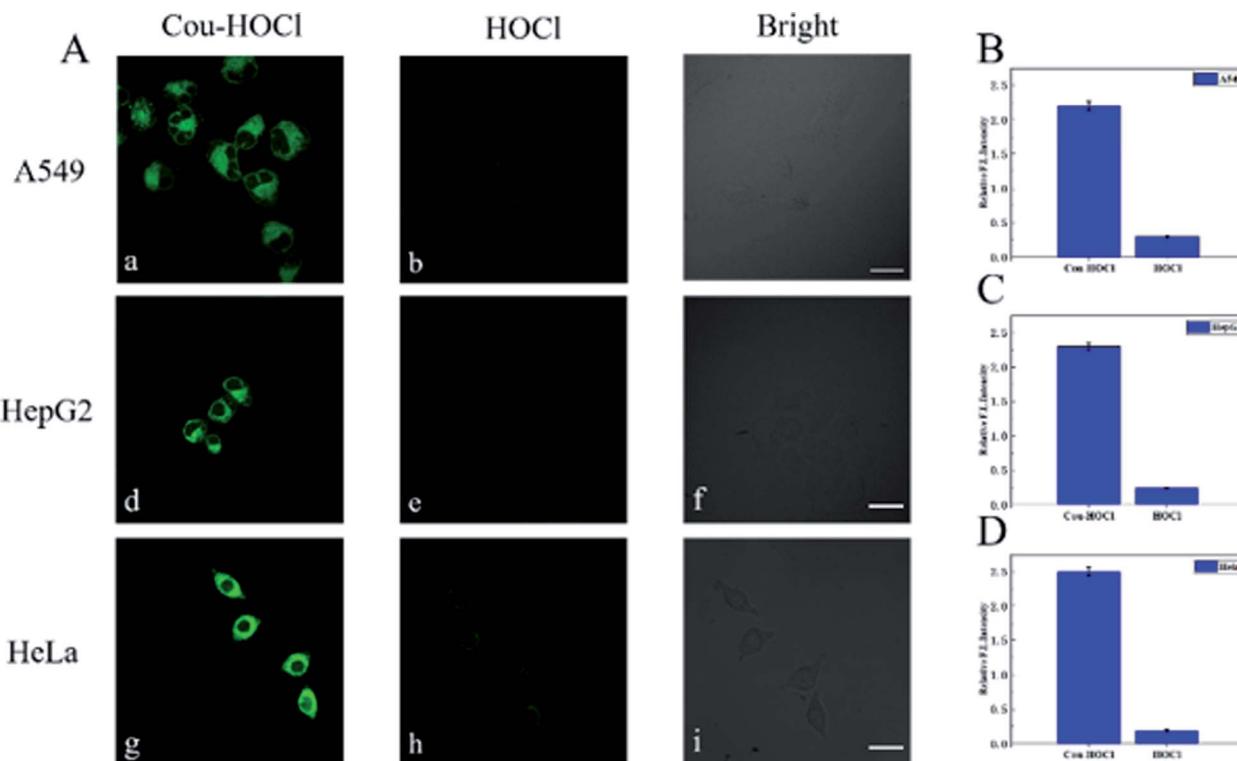


Fig. 2 Fluorescence images of exogenous HOCl in A549, HeLa and HepG2 cells. (A) A549, HeLa and HepG2 cells were pretreated with or without HOCl for 30 minutes, and then treated with Cou-HOCl (10 μM) for 5 minutes. After being washed with PBS, the cells were imaged. (B-D) Quantification of the relative fluorescence intensities of the A549, HepG2 and HeLa cells. The data are shown as mean (\pm SD). Scale bar = 30 μm.

+ Cl⁻ treatment groups, the green fluorescence intensities were slightly reduced compared to the control, while the intensity for the MPO + H₂O₂ + Cl⁻ treatment group was significantly reduced. To further verify the intracellular fluorescence changes caused by endogenous HOCl, different samples of the cells were treated with LPS, 4-aminobenzoic acid hydrazide (ABH) or

salicyl hydroxamic acid (SHA) (two well-known MPO inhibitors), and Cou-HOCl, respectively. The green fluorescence signals of the cells were well maintained in the presence of ABH or SHA. Overall, these results demonstrated that Cou-HOCl was a simple tool capable to detect and image exogenous and endogenous HOCl levels in inflammatory cells.

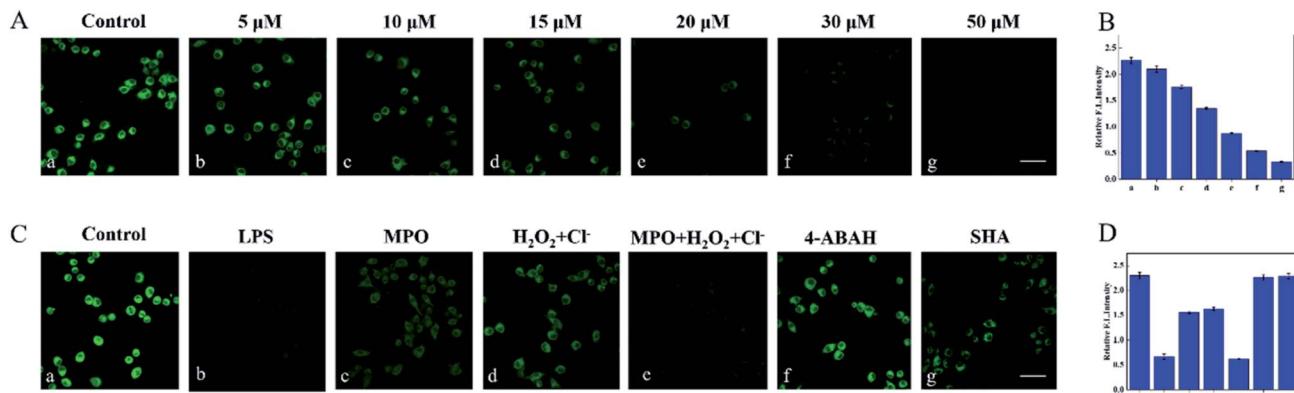


Fig. 3 Fluorescence images of HOCl flux in RAW264.7 cells. (A) Different samples of RAW264.7 cells were pretreated with different concentrations of HOCl (0, 5, 10, 15, 20, 30 and 50 μM, respectively) for 30 minutes, and then each incubated with Cou-HOCl (10 μM) for 30 minutes before being subjected to fluorescence imaging. (B) Quantitative plot of the relative proportion of the fluorescence intensity of the cells in A. (C) Different samples of cells were preincubated with or without LPS (1 μg mL⁻¹, 12 h), MPO (100 ng mL⁻¹, 1 h), H₂O₂ (100 μM, 1 h), Cl⁻ (1 mM, 1 h), 4-ABA (500 μM, 1 h), and SHA (500 μM, 1 h), respectively, and then each treated with Cou-HOCl (10 μM) for another 30 minutes. Then, after being washed with PBS, the cells were imaged. (D) Quantitative plot of the relative proportion of the fluorescence intensity of the cells in C. The data are shown as mean (\pm SD). Scale bar = 30 μm.



Conclusions

In conclusion, we have rationally designed and synthesized a fluorescent probe with high selectivity and sensitivity for HOCl detection. The fluorescence imaging of HOCl in various cell lines demonstrated the potential biological application of Cou-HOCl. Furthermore, Cou-HOCl could be applied for directly visualizing endogenous HOCl in inflammatory cells. Due to the advantages of Cou-HOCl, we believe that it can serve as an important tool for cell biology research and clinical medical diagnosis.

Author contributions

Zhencai Xu: methodology, data curation, writing – original draft. Xiaofeng Wang: validation, data curation. Tingting Duan: data curation. Rong He: validation. Fangwu Wang: writing – original draft. Xuejun Zhou: supervision, project administration, funding acquisition.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the Natural Science Foundation of Hainan Province (819MS121).

Notes and references

- 1 E. A. Souza, D. Maitra, G. M. Saed, M. P. Diamond, A. A. Moura, S. Pennathur and H. M. Abu-Soud, *PLoS One*, 2011, **6**, e27641.
- 2 D. I. Pattison and M. J. Davies, *Chem. Res. Toxicol.*, 2001, **14**, 1453–1464.
- 3 C. M. Spickett, A. Jerlich, O. M. Panasenko, J. Arnhold, A. R. Pitt, T. Stelmaszyńska and R. J. Schaur, *Acta Biochim. Pol.*, 2000, **47**, 889–899.
- 4 J. Marcinkiewicz, B. Chain, B. Nowak, A. Grabowska, K. Bryniarski and J. Baran, *Inflamm. Res.*, 2000, **49**, 280–289.
- 5 M. S. Petrônio and V. F. Ximenes, *Biochim. Biophys. Acta*, 2012, **1824**, 1090–1096.
- 6 M. L. Aiken, R. G. Painter, Y. Zhou and G. Wang, *Free Radic. Biol. Med.*, 2012, **53**, 2308–2317.
- 7 Y. W. Yap, M. Whiteman and N. S. Cheung, *Cell. Signal.*, 2007, **19**, 219–228.
- 8 N. O. Soto, B. Horstkotte, J. G. March, P. L. López de Alba, L. López Martínez and V. Cerdá Martín, *Anal. Chim. Acta*, 2008, **611**, 182–186.
- 9 O. Ordeig, R. Mas, J. Gonzalo, F. J. Del Campo, F. J. Muñoz and C. de Haro, *Electroanalysis*, 2005, **17**, 1641–1648.
- 10 Q. Yang, T. Lan and W. He, *Dyes. Pigments*, 2021, **186**, 108997.
- 11 H. Liu, M. N. Radford, C. T. Yang, W. Chen and M. Xian, *Br. J. Pharmacol.*, 2019, **176**, 616–627.
- 12 J. L. Kolanowski, F. Liu and E. J. New, *Chem. Soc. Rev.*, 2018, **47**, 195–208.
- 13 R. Lü, *J. Mol. Cell. Cardiol.*, 2017, **110**, 96–108.
- 14 J. Chan, S. C. Dodani and C. J. Chang, *Nat. Chem.*, 2012, **4**, 973–984.
- 15 L. Wang, M. Ren, Z. Li, L. Dai and W. Lin, *Anal. Methods*, 2019, **11**, 1580–1584.
- 16 G. Cheng, J. Fan, W. Sun, K. Sui, X. Jin, J. Wang and X. Peng, *Analyst*, 2013, **138**, 6091–6096.
- 17 G. Cheng, J. Fan, W. Sun, J. Cao, C. Hu and X. Peng, *Chem. Commun.*, 2014, **50**, 1018–1020.
- 18 B. Wang, P. Li, F. Yu, P. Song, X. Sun, S. Yang, Z. Lou and K. Han, *Chem. Commun.*, 2013, **49**, 1014–1016.
- 19 K. Wang, P. Jia, X. Li, X. Zhang, C. Liu, Y. Yu, H. Zhu, Z. Li, W. Sheng and B. Zhu, *Dyes. Pigments*, 2020, **177**, 108310.
- 20 P. Jia, Z. Zhuang, C. Liu, Z. Wang, Q. Duan, Z. Li, H. Zhu, B. Du, B. Zhu, W. Sheng and B. Kang, *Anal. Chim. Acta*, 2019, **1052**, 131–136.
- 21 S. L. Shen, X. Q. Huang, H. L. Jiang, X. H. Lin and X. Q. Cao, *Anal. Chim. Acta*, 2019, **1046**, 185–191.
- 22 T. R. Wang, X. F. Zhang, X. Q. Huang, X. Q. Cao and S. L. Shen, *Spectrochim Acta A*, 2021, **247**, 119115.
- 23 J. T. Hou, M. Y. Wu, K. Li, J. Yang, K. K. Yu, Y. M. Xie and X. Q. Yu, *Chem. Commun.*, 2014, **50**, 8640–8643.
- 24 Y. Gan, G. Yin, X. Zhang, L. Zhou, Y. Zhang, H. Li and P. Yin, *Talanta*, 2021, **225**, 122030.
- 25 H. Ren, F. Huo and C. Yin, *New J. Chem.*, 2021, **45**, 4724–4728.
- 26 B. Wang, F. Zhang, S. Wang, R. Yang, C. Chen and W. Zhao, *Chem. Commun.*, 2020, **56**, 2598–2601.
- 27 H. Zhang, X. Yin, J. Hong, Y. Deng and G. Feng, *Talanta*, 2021, **223**, 121768.
- 28 D. Zhang, S. Guo, L. Li and K. Shang, *Analyst*, 2020, **145**, 7477–7487.
- 29 S. Dong, L. Zhang, Y. Lin, C. Ding and C. Lu, *Analyst*, 2020, **145**, 5068–5089.
- 30 T. Yudhistira, S. V. Mulay, Y. Kim, M. B. Halle and D. G. Churchill, *Chem. Asian. J.*, 2019, **14**, 3048–3084.

