# RSC Advances



## PAPER

Cite this: RSC Adv., 2021, 11, 26666

## Transition-metal-free synthesis of aryl 1 thioglycosides with arynes at room temperature†

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A mild, convenient and transition-metal-free protocol for the synthesis of aryl 1-thioglycosides is presented via arynes generated in situ combined with glycosyl thiols in the presence of TBAF(tBuOH)<sub>4</sub>. The methodology provides a general and efficient way to prepare a series of functionalized thioglycosides in good to excellent yields with a perfect control of the anomeric configuration at room temperature. In addition, the reaction conditions tolerate a variety of the pentoses and hexoses, and the reaction also performs smoothly on protected monosaccharides and disaccharides.

rsc.li/rsc-advances

Received 23rd May 2021 Accepted 22nd July 2021 DOI: 10.1039/d1ra04013h

### Introduction Thioglycosides, as the analogue of the O-glycosides, are versatile

intermediates in carbohydrate synthesis and have a wide range of potential applications,<sup>1</sup> and are regularly used as glycosyl donors<sup>2</sup> to construct various oligosaccharides and glycoconjugates. Additionally, they are more stable in both chemical and enzymatic degradations<sup>3</sup> and have also been employed extensively as inhibitors in a great number of biochemical studies.<sup>4</sup> Thioglycoside fragments are also widely used in various drugs, natural products and pharmaceutical active agents.<sup>5</sup> Some examples of the thioglycosides derivatives include the cytotoxic Hsp90 inhibitor,<sup>6</sup> MUS-CB,<sup>7</sup> hSGLT1 inhibitor,<sup>8</sup> lincomycin,<sup>9</sup> clindamycin,<sup>10</sup> as well as irreversible glycoside inhibitors $11$  (Fig. 1). On account of the great significance of S-glycosides, some elegant synthetic protocols have been developed.

Originally, the approaches to prepare the 1-thioglycosides were that glycosyl donors reacted with sulfurs (or thiophenols) in the presence of the stoichiometric Lewis acids $12$  or glycosyl halides reacted with thiolate anions under strong base conditions (Scheme 1a). However, the inferior stereo-selectivity or strong base of those traditional methods limited their application. With the efforts of Sticha, Xue, Messaoudi et al., the synthesis of 1-thioglycosides achieved great advances in recent years. As shown in Scheme 1b, the functionalization of glycosyl thiols had been catalysed by the Ni,<sup>13</sup> Cu<sup>14</sup>, Pd<sup>15</sup> transition metal

via the Buchwald–Hartwlg–Migita coupling reaction. These methods required an expensive catalyst or a high temperature (above 100 $\degree$ C in some protocols of the Cu or Pd catalyst) and long reaction time, which limit the universality of the reaction to some extent. In 2019, Messaoudi and co-workers developed a protocol for the synthesis of aryl 1-thioglycosides via a Ni/ photoredox dual catalyzed cross-coupling reaction<sup>16</sup> (Scheme 1c). Recently, Messaoudi's group described the first electrochemical method for coupling various anomeric glycosyl thiols with aryl bromides<sup>17</sup> (Scheme 1c). The reaction didn't need to perform in a strong base environment, and demonstrated superiority to synthesize highly complex thioglycosides under mild conditions. However, some functional groups are not compatible with this procedure. PAPER<br>
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> Arynes are highly reactive transient intermediates and are also useful synthons in organic synthesis due to the unique triple bonds, and have been extensively used in the synthesis of natural products, drug molecules and functional materials.<sup>18</sup> Arynes have made remarkable achievements in nucleophilic reactions,<sup>19</sup> pericyclic reactions,<sup>20</sup> transition metal catalyzed reactions<sup>21</sup> and multicomponent reactions.<sup>22</sup> In recent years, N-arylation of carbohydrate amines<sup>23</sup> and O-arylation of carbohydrates<sup>24</sup> have been reported via the aryne insertion reactions with glycosyl



Fig. 1 Examples of biologically active thioglycosides.

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<sup>†</sup> Electronic supplementary information (ESI) available: Additional experimental details and complete NMR spectral data for all synthesized compounds. See DOI: 10.1039/d1ra04013h

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thiols. The methodology of using the aryne generated in situ to realize the functionalized thioglycosides was not developed yet. On the basis of our previous research in carbohydrates, $25$  we report a novel protocol for the preparation of thioglycosides via a tandem intermolecular nucleophilic coupling of glycosyl thiols with arynes (Scheme 1c).

### Results and discussion

We started by optimizing the reaction conditions with tetra-Oacetylated 1-thio- $\beta$ -p-glucopyranose 1a (1.0 equiv.),

Table 1 Optimization conditions for the synthesis of  $S$ -glycosides<sup>a</sup>

OAc <b>OAc</b> TMS F-, Solvent AcC Ac( AcC $T(^{\circ}C)$ , $t(h)$ OAc OTf OAc 2a 3a 1a					
Entry	sources $_{\rm F}^{-}$	Solvent	Time/h	$T$ /°C	Yield $(\% )$
1	CsF	CH <sub>3</sub> CN	2.0	25	$70(64^{b})$
2	AgF	CH <sub>3</sub> CN	2.0	25	51
3	KF	CH <sub>3</sub> CN	2.0	25	42
$\overline{4}$	ZnF <sub>2</sub>	CH <sub>3</sub> CN	2.0	25	35
5	$TBAF \cdot 3H_2O$	CH <sub>3</sub> CN	2.0	25	63
6	TBAF(THF)	CH <sub>3</sub> CN	2.0	25	67
7	$TBAF \cdot (tBuOH)_4$	CH <sub>3</sub> CN	2.0	25	86
8	$TBAF \cdot (tBuOH)_4$	<b>DCM</b>	2.0	25	78
9	$TBAF·($ <i>tBuOH</i> $)_4$	<b>THF</b>	2.0	25	40
10	$TBAF \cdot (tBuOH)_4$	Toluene	2.0	25	33
11	$TBAF·($ <i>tBuOH</i> $)_4$	MeOH	2.0	25	37
12	$TBAF·($ <i>tBuOH</i> $)_4$	<b>DMSO</b>	2.0	25	26
13	$TBAF \cdot (tBuOH)_4$	<b>DMF</b>	2.0	25	30
14	$TBAF·($ <i>tBuOH</i> $)_4$	CH <sub>3</sub> CN	2.0	40	85
15	$TBAF \cdot (tBuOH)_4$	CH <sub>3</sub> CN	2.0	60	60
16	$TBAF \cdot (tBuOH)_4$	CH <sub>3</sub> CN	3.0	25	$83^c$
17	$TBAF \cdot (tBuOH)_4$	CH <sub>3</sub> CN	4.0	25	82

 $a$  Standard conditions: 1a (0.1 mmol, 1.0 equiv.), 2a (0.11 mmol, 1.1 equiv.), TBAF $\cdot$ (tBuOH)<sub>4</sub> (0.2 mmol, 2.0 equiv.), CH<sub>3</sub>CN (1.5 mL) as solvent, r.t., 2 h. <sup>b</sup> 1a (0.1 mmol, 1.0 equiv.), 2a (0.1 mmol, 1.0 equiv.), source (0.2 mmol, 2.0 equiv.), CH<sub>3</sub>CN (1.5 mL).  $c$  1a (0.1 mmol, 1.0 equiv.), 2a (0.11 mmol, 1.1 equiv.), TBAF  $(tBuOH)_4$  (0.3 mmol, 3.0 equiv.),  $CH<sub>3</sub>CN$  (1.5 mL) as solvent, r.t., 2 h.

 $(t$ rimethylsilyl $)$ aryl triflates 2a  $(1.0 \text{ equiv.})$  as model substrate. Phenyl-2,3,4,6-tetra-O-acetyl-1-thio-B-p-glucopyranoside 3a was isolated after 2 hours in 64% yield at room temperature (entry 1, Table 1). When the equivalent of aryne precursor increased to 1.1 equiv., the yield of desired product increased to 70% (entry 1, Table 1). In spired by this result, other fluorides, such as AgF,  $ZnF_2$ , KF, more widely used tetrabutyl ammonium fluoride (TBAF) and its analogs (entries  $2-7$ , Table 1) were tested. To our delight, the yield of 3a increased to 86% when TBAF $\cdot$ (tBuOH)<sub>4</sub> (ref. 26) was used as a fluorine source (entry 7, Table 1). Next, we also investigated the solvent effect by using varies polar solvents and non-polar solvents (entries 8–13, Table 1). Nucleophilic solvents such as tetrahydrofuran, methanol, N,N-dimethylformamide and dimethyl sulfoxide exhibited low yields (26–40%), probably because the competitive reaction between nucleophilic solvents and the substrates. The yield of 3a was decreased to 60% when the temperature increased to 60  $\degree$ C (entry 15). We found that the yield of the target product was not influenced with extended reaction time. Puper<br>
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With the optimized condition in hand, the scopes of the glycosyl thiols were further investigated and the results are presented in Scheme 2. Various acetyl-protected  $\beta$ -glycosyl thiols 1c, 1d reacted with arynes under the standard conditions and the corresponding  $\beta$ -configuration thioglycosides products (3c, 3d) were obtained in good yields (85%, 75%). This conversion was also applicable for the  $\alpha$ -configuration glycosyl thiol substrate 1b and 1e, providing corresponding arylthioglycoside products 3b and 3e in 80% and 77% yield, respectively. Moreover, triacetyl protected-pentose glycosyl thiols 1f, 1g participated in this reaction smoothly and the yields of the corresponding thioglycoside products were 82% and 81%, respectively. The reaction was not limited to monosaccharides but also extended to disaccharides. Acetyl-protected cellobiose 1h and benzoyl-protected cellobiose 1i were reacted with arynes



Scheme 2 Scopes of glycosyl thiols 1 reacted with aryne 2a. Standard conditions: 1 (0.1 mmol, 1.0 equiv.), 2a (0.11 mmol, 1.1 equiv.), TBAF $\cdot$ (tBuOH)<sub>4</sub> (0.2 mmol, 2.0 equiv.), dry CH<sub>3</sub>CN (1.5 mL).

successfully and obtained the  $\beta$ -disaccharide in good yields, indicated the electronic-effect of the protecting groups did not have significant influence for the reaction efficiency. Meanwhile, we also found the  $\alpha$ -glycosyl thiols with OBn protected group can reacted with 2a when the temperature improved to 45 °C and the yield of corresponding product 3j was 78%. The sugar 1k with OPiv group provided 3k in 71% yield under optical condition. However, the reactivity of unprotected glycosyl thiol was decreased and the yield of product was only 30%.

Subsequently, we also investigated the electronic effects and regioselectivity of this reaction via different aryne precursors and glycosyl thiols 1. Regardless of the electron-donating or -withdrawing properties of the symmetrical aryne precursors 2, the corresponding products were obtained in good yields. The difluoro substituted aryne  $2b$  reacted with 1 gave the thioglycosides derivatives 4a–4h in 71–82% yields. Aryne precursor 2c reacted with a series of glycosyl thiols under standard reaction condition to afford the corresponding products 4i–4l in good yields (72–84%). Also, the dimethyl substituted aryne precursors reacted with 1a under standard condition to give corresponding aryl 1-thioglycoside 4m with 78% yield.

Meanwhile, 3-methoxy non-symmetric aryne precursor 2d exhibited excellent regioselectivity due to the steric effect and electronic effects, and single target thio-glycosides 4n, 4o were obtained in good yields (80%, 78%). However, we found that 4 methylbenzyne precursor 2e reacted with 1a and 1f gave a near equimolar mixture of two inseparable regioisomers, providing



Scheme 3 Scope of aryne precursors 2 reacted with glycosyl thiol. Standard condition: 1 (0.1 mmol, 1.0 equiv.), 2 (0.11 mmol, 1.1 equiv.), TBAF $\cdot$ (tBuOH)<sub>4</sub> (0.2 mmol, 2.0 equiv.), dry CH<sub>3</sub>CN (1.5 mL).



Scheme 4 Gram-scale reaction and plausible mechanism

corresponding products in 82% and 90% yield, respectively. 1,2- Naphthyne precursor 2f under the same conditions also provided a near equimolar mixture of two inseparable regioisomers (Scheme 3).

To demonstrate the synthetic utility of this transformation, we next performed a scale-up reaction of 3a. As shown in Scheme 4a, a gram scale reaction of 1-thio- $\beta$ -D-glucopyranose 1a with aryne precursor 2a proceeded to give 3a in 85% yield. A plausible mechanism was proposed in Scheme 4b, based on the basis of the experimental results and the related report by Jin and co-workers.<sup>27</sup>

### Conclusions

In conclusion, we have developed a convenient method for the preparation of aryl-thioglycosides under mild and metalcatalyst-free conditions via a tandem intermolecular nucleophilic coupling of glycosyl thiols with arynes. Meanwhile, the method is applicable for various monosaccharide and disaccharide substrates, which has wide practical value between biochemistry and medicinal chemistry. In addition, 1,2-transthioglycosides were stereoselectively formed by the reaction of the in situ generated arynes with glycosyl thiols in good to excellent yields. Importantly, we provided a new protocol for the synthesis of functionalized thioglycosides and the possibility for further derivatization of glycosyl donors.

### Experimental

#### General information

All reactions were carried out in dried glassware. The solvent in the reaction were dried use activated  $4 \text{ Å}$  molecular sieve, commercial reagents were used without further purification unless otherwise stated. Purification of reaction products were carried out by flash chromatography on silica gel (200–300 mesh). NMR spectra were measured in CDCl<sub>3</sub> (with TMS as internal standard) on a Bruker AV400 ( $^1\rm H$  at 400 MHz,  $^{13}$ C at 100 MHz,  $^{19}$ F at 376 MHz) magnetic resonance spectrometer.

#### The general procedure for the reaction of glycosyl thiols 1 with aryne precursors 2

The glycosyl thiol 1 (50 mg, 0.14 mmol, 1.0 equiv.), aryne precursor 2 (45 mg, 0.150 mmol, 1.1 equiv.) and TBAF $\cdot$ (tBuOH)<sub>4</sub> (0.274 mmol, 2.0 equiv.) were sequentially added in a clean and dry Schlenk tube, and the tube was then evacuated and back filled with nitrogen (this sequence was repeated three times). Under nitrogen atmosphere, MeCN (1.5 mL) was added to the mixture system, then the mixture was stirred at room temperature for 2.0 hours. Saturated NaCl solution was added to dilute the system and extracted with EtOAc  $(3 \times 2$  mL). The combined organic phase was dried over anhydrous  $Na<sub>2</sub>SO<sub>4</sub>$ , filtered, and evaporated in vacuo. Finally, the crude product was purified via flash column chromatography on silica gel to give the desired product.

#### Phenyl-2,3,5-tri-*O*-acetyl-1-thio-β-D-xylofuranoside (3g)

Purified by flash column chromatography  $R_f = 0.39$  (petroleum ether/AcOEt =  $3:1$ ), white solid (29.8 mg, 81% yield);  $[\alpha]_{\text{D}}^{25} = -55.4$  ( $c = 1.0$ , CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.55–7.46 (m, 2H, ArH), 7.40–7.28 (m, 3H, ArH), 5.18 (t,  $J =$ 8.2 Hz, 1H), 5.06-4.87 (m, 2H), 4.80 (d,  $J = 8.4$  Hz, 1H), 4.28 (dd,  $J = 11.8, 4.9$  Hz, 1H), 3.42 (dd,  $J = 11.8, 8.7$  Hz, 1H), 2.09 (s, 3H), 2.05 (s, 3H), 2.04 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.9, 169.8, 169.3, 132.7, 132.2, 129.1, 128.3, 86.2, 71.9, 69.8, 68.4, 65.2, 20.8, 20.8, 20.7; HRMS (ESI)  $m/z$  calcd for C<sub>17</sub>H<sub>20</sub>O<sub>7</sub>SNa<sup>+</sup>  $(M + Na)^+$  391.0822, found 391.0822.

#### 3,4-Difluorphenyl-2,3,4,6-tetra-O-acetyl-1-thio-β-D-gluco-pyranoside (4a)

Purified by flash column chromatography  $R_f = 0.30$  (petroleum ether/AcOEt =  $3:1$ ), white solid (39.1 mg, 82% yield);  $\left[ \alpha \right]_{\text{D}}^{25} = -20.7 \left( c = 1.0, \text{CHCl}_3 \right); \, {}^{1}\text{H NMR} \left(400 \text{ MHz}, \text{CDCl}_3 \right) \delta$  7.42 (ddd,  $J = 10.2, 7.5, 2.2$  Hz, 1H), 7.24-7.19 (m, 1H), 7.10 (dt,  $J = 10.0$ , 8.3 Hz, 1H), 5.21 (t,  $J = 9.4$  Hz, 1H), 5.00  $(t, J = 9.8 \text{ Hz}, 1\text{H})$ , 4.90  $(t, J = 9.6 \text{ Hz}, 1\text{H})$ , 4.63  $(d, J = 10.0 \text{ Hz},$ 1H), 4.19 (d,  $J = 3.8$  Hz, 2H), 3.73 (dt,  $J = 10.1$ , 3.8 Hz, 1H), 2.09  $\left( \text{s, 3H} \right)$ , 2.08  $\left( \text{s, 3H} \right)$ , 2.01  $\left( \text{s, 3H} \right)$ , 1.98  $\left( \text{s, 3H} \right)$ ;  $^{19}$ F  $\{^1\text{H}\}$  NMR  $\left( \text{376} \right)$ MHz, CDCl<sub>3</sub>)  $\delta$  -135.81 (dt, J = 19.7, 9.3 Hz), -136.61 (m); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -135.81 (d, J = 20.9 Hz), -136.61 (d, J  $= 21.0$  Hz). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.6, 170.1, 169.3, 169.2, 150.9 (dd,  $J = 251.0$ , 12.3 Hz), 149.9 (dd,  $J = 251.1$ , 12.5 Hz), 130.3 (dd,  $J = 6.4$ , 3.6 Hz), 127.1 (dd,  $J = 6.2$ , 4.2 Hz), 122.9  $(d, J = 18.2 \text{ Hz})$ , 117.5  $(d, J = 17.6 \text{ Hz})$ , 85.1, 75.9, 73.8, 69.6, 68.0, 62.0, 20.7, 20.6, 20.5; HRMS (ESI):  $m/z$  calcd for C<sub>20</sub>H<sub>22</sub>F<sub>2</sub>O<sub>9</sub>SNa<sup>+</sup>  $(M + Na)^+$  499.0845, found 499.0843.

### 3,4-Difluorphenyl-2,3,4,6-tetra-O-acetyl-1-thio-β-D-galactopyranoside (4b)

Purified by flash column chromatography  $R_f = 0.28$  (petroleum ether/AcOEt =  $3:1$ ), colorless oil (37.1 mg, 78% yield);  $[\alpha]_{\text{D}}^{25} = 6.8 \; (c = 1.0, \text{CHCl}_3); \, {}^{1}\text{H} \; \text{NMR} \; (400 \; \text{MHz, CDCl}_3) \; \delta \; 7.47$ (ddd,  $J = 10.2, 7.5, 2.2$  Hz, 1H), 7.24-7.19 (m, 1H), 7.11  $(dt, J = 10.0, 8.4 Hz, 1H), 5.41 (dd, J = 3.3, 1.1 Hz, 1H), 5.18 (dd, J)$  $= 9.9$  Hz, 1H), 5.03 (dd,  $J = 9.9$ , 3.3 Hz, 1H), 4.64 (d,  $J = 9.9$  Hz,

1H), 4.20-4.09 (m, 2H), 3.95 (td,  $J = 5.8$ , 2.9 Hz, 1H), 2.11 (s, 3H), 2.09 (s, 3H), 2.06 (s, 3H), 1.96 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.4, 169.1, 168.9, 168.3, 149.6 (dd,  $J = 250.8$ , 12.5 Hz), 148.9  $(dd, J = 250.9, 13.0 Hz$ ), 128.5  $(dd, J = 6.2, 3.7 Hz$ ), 127.1  $(dd, J = 1250.9, 13.0 Hz$ 6.1, 4.2 Hz), 121.1  $(d, J = 18.3 \text{ Hz})$ , 116.4  $(d, J = 17.7 \text{ Hz})$ , 84.9, 73.6, 70.8, 66.2, 65.9, 60.8, 19.8, 19.6, 19.5, 19.5; <sup>19</sup>F  ${^1_1}H$  NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  –135.92 (dt, J = 19.8, 9.3 Hz), –137.16 (m); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  –135.92 (d, J = 21.0 Hz), –137.17  $(d, J = 20.9 \text{ Hz})$ . HRMS (ESI): m/z calcd for C<sub>20</sub>H<sub>22</sub>F<sub>2</sub>O<sub>9</sub>SNa<sup>+</sup> (M + Na)<sup>+</sup> 499.0845, found 499.0841.

#### 3,4-Difluorphenyl-2-acetamido-3,4,6-tri-O-acetyl-2-deoxy-1-thio-ß-D-glucopyranoside (4d)

Purified by flash column chromatography  $R_f = 0.28$  (petroleum ether/AcOEt =  $1:2$ ), white solid (33.6 mg, 71% yield);  $\left[ \alpha \right]_{\text{D}}^{25} = -24.5 \left( c = 1.0, \text{CHCl}_3 \right); \, {}^{1}\text{H} \text{ NMR} \left( 400 \text{ MHz}, \text{CDCl}_3 \right) \delta$  7.43  $(\text{ddd}, J = 10.0, 7.4, 2.1 \text{ Hz}, 1\text{H}), 7.25-7.19 \text{ (m, 1H)}, 7.09 \text{ (dt)}, J =$ 10.1, 8.4 Hz, 1H), 5.76 (d,  $J = 9.1$  Hz, 1H), 5.19 (t,  $J = 9.8$  Hz, 1H), 5.02 (t,  $J = 9.7$  Hz, 1H), 4.77 (d,  $J = 10.4$  Hz, 1H), 4.21-4.13 (m,  $2H$ , 4.00 (q,  $J = 9.8$  Hz, 1H), 3.77-3.67 (m, 1H), 2.08 (s, 3H), 2.02  $(s, 6H)$ , 1.98  $(s, 3H)$ ; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.0, 170.6, 170.1, 169.3, 150.6 (dd,  $J = 251.0$ , 12.6 Hz), 149.9 (dd,  $J = 250.9$ , 12.8 Hz), 129.6 (dd,  $J = 6.3$ , 3.6 Hz), 128.2 (dd,  $J = 6.3$ , 4.2 Hz), 122.1 (d,  $I = 18.2$  Hz), 117.5 (d,  $I = 17.5$  Hz), 86.4, 75.9, 73.5, 68.2, 62.3, 53.2, 23.3, 20.6, 20.6, 20.5; <sup>19</sup>F {<sup>1</sup>H} NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -135.93 (dt, J = 19.9, 9.3 Hz), -137.28 (m); <sup>19</sup>F NMR  $(376 \text{ MHz}, \text{CDCl}_3) \delta -135.93 \text{ (d, } J = 21.4 \text{ Hz}), -137.29$  $(d, J = 21.0 \text{ Hz})$ . HRMS (ESI):  $m/z$  calcd for C<sub>20</sub>H<sub>23</sub>F<sub>2</sub>NO<sub>8</sub>SNa<sup>+</sup> (M  $+$  Na)<sup>+</sup> 498.1005, found 498.1006. Paper<br> **The general procedure for the reaction of gytensyl think 1 with**  $\frac{2\pi}{3}$ **.** This article. This article. This article. This article. This article. This article. This article is licensed to the common a state mult

### 3,4-Difluorphenyl-2,3,4-tri-O-acetyl-1-thio- $\alpha$ -L-rhamno-pyranoside (4e)

Purified by flash column chromatography  $R_f = 0.47$  (petroleum ether/AcOEt =  $3:1$ ), white solid (31.8 mg, 76% yield);  $\left[ \alpha \right]_{\text{D}}^{25}$  =  $-116.0$   $\left( \text{c} = 1.0, \text{CHCl}_3 \right);$  <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.31  $(\text{ddd}, J = 10.0, 7.3, 2.2 \text{ Hz}, 1\text{H}), 7.21-7.17 \text{ (m, 1H)}, 7.11 \text{ (dt)}, J =$ 10.0, 8.3 Hz, 1H), 5.45 (dd,  $J = 3.3$ , 1.7 Hz, 1H), 5.35 (d,  $J = 1.6$  Hz, 1H), 5.21 (dd,  $J = 10.1$ , 3.2 Hz, 1H), 5.14 (t,  $J = 9.8$  Hz, 1H), 4.34– 4.26 (m, 1H), 2.14 (s, 3H), 2.07 (s, 3H), 2.01 (s, 3H), 1.25 (d,  $J =$ 6.2 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 170.1, 170.0, 170.4, 150.5 (dd,  $J = 250.5$ , 12.6 Hz), 150.3 (dd,  $J = 251.8$ , 13.1 Hz), 129.4  $(dd, J = 6.1, 4.1 Hz$ , 128.6  $(dd, J = 6.3, 3.7 Hz$ , 121.2  $(d, J = 18.2$ Hz), 118.0 (d,  $J = 17.7$  Hz), 86.0, 71.1, 71.0, 69.4, 68.1, 21.0, 20.9, 20.8, 17.4; <sup>19</sup>F {<sup>1</sup>H} NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -135.57 (dt, J = 20.9, 9.2 Hz),  $-137.48$  (m); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -135.57  $(d, J = 20.9 \text{ Hz})$ ,  $-137.50$   $(d, J = 21.0 \text{ Hz})$ . HRMS (ESI):  $m/z$  calcd for  $C_{18}H_{20}F_2O_7SNa^+ (M + Na)^+$  441.0790, found 441.0795.

#### 3,4-Difluorophenyl-2,3,5-tri-*O*-acetyl-1-thio-β-D-xylo-furanoside (4g)

Purified by flash column chromatography  $R_f = 0.38$  (petroleum ether/AcOEt =  $3:1$ ), white solid (29.9 mg, 74% yield);  $\lbrack \alpha \rbrack_{\rm D}^{25} = - 56.9 \; (c = 0.5, \, \text{CHCl}_3); \, {}^1\text{H}$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.35  $(ddd, J = 10.0, 7.4, 2.2$  Hz, 1H), 7.24–7.19 (m, 1H), 7.11  $(dt, J = 10.1, 10.1)$ 8.3 Hz, 1H), 5.17 (t,  $J = 8.1$  Hz, 1H), 4.94-4.85 (m, 2H), 4.73 (d,  $J =$ 8.2 Hz, 1H), 4.27 (dd,  $J = 11.8$ , 4.9 Hz, 1H), 3.43 (dd,  $J = 11.8$ , 8.6 Hz, 1H), 2.10 (s, 3H), 2.04 (s, 6H); 13C NMR (100 MHz, CDCl3)  $\delta$  170.0, 169.9, 169.5, 150.9 (dd,  $J = 250.9$ , 12.5 Hz), 150.1 (dd,  $J =$ 251.7, 12.9 Hz), 130.1 (dd,  $J = 5.9$ , 3.8 Hz), 128.0-127.7 (m), 122.6  $(d, J = 18.0 \text{ Hz})$ , 117.8  $(d, J = 17.6 \text{ Hz})$ , 85.9, 71.8, 69.7, 68.3, 65.3, 20.9, 20.9, 20.8; <sup>19</sup>F  ${^4H}$ } NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  –135.81 (dt, *J* = 21.3, 9.2 Hz),  $-136.89$  (m); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -135.81  $(d, J = 21.5 \text{ Hz})$ ,  $-136.90 \, (d, J = 21.4 \text{ Hz})$ . HRMS (ESI):  $m/z$  calcd for  $C_{17}H_{18}F_2O_7SNa^+(M+Na)^+$  427.0634, found 427.0633.

#### 6-Benzo[*d*][1,3]dioxolyl-2,3,6,2′,3′,4′,6′-hepta-*O*-acetyl-1-thiob-cellobioside (4k)

Purified by flash column chromatography  $R_f = 0.41$  (petroleum ether/AcOEt =  $1:1$ , white solid (58.6 mg, 76% yield);  $\alpha_{\rm 1D}^{\rm 225} = -24.5$  ( $c = 0.5$ , CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.01  $(d, J = 1.8$  Hz, 1H), 6.95  $(dd, J = 8.0, 1.8$  Hz, 1H), 6.73  $(d, J = 8.0 \text{ Hz}, 1\text{H}), 5.97 \text{ (s, 2H)}, 5.14 \text{ (dt, } J = 14.9, 9.3 \text{ Hz}, 2\text{H}), 5.05$  $(t, J = 9.6 \text{ Hz}, 1\text{H}), 4.90 \text{ (dd, } J = 9.2, 7.9 \text{ Hz}, 1\text{H}), 4.82$  $(t, J = 9.7 \text{ Hz}, 1\text{H})$ , 4.58 (dd,  $J = 11.9, 2.0 \text{ Hz}, 1\text{H}$ ), 4.50 (dd,  $J =$ 13.5, 9.0 Hz, 2H), 4.37 (dd,  $J = 12.5$ , 4.2 Hz, 1H), 4.07 (dd,  $J = 11.9$ , 5.3 Hz, 1H), 4.01 (dd,  $J = 12.5$ , 2.3 Hz, 1H), 3.69 (t,  $J = 9.5$  Hz, 1H), 3.65–3.56 (m, 2H), 2.12 (s, 3H), 2.08 (s, 3H), 2.06 (s, 3H), 2.01 (s, 3H), 1.99 (s, 3H), 1.99 (s, 3H), 1.97 (s, 3H); 13C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.6, 170.4, 170.4, 169.9, 169.6, 169.4, 169.1, 148.6, 147.9, 128.8, 122.8, 114.8, 108.6, 101.6, 100.9, 85.8, 76.4, 73.8, 73.0, 72.1, 71.7, 70.0, 67.8, 61.9, 61.6, 20.9, 20.8, 20.7, 20.6. RSC Advances  $\frac{1}{2}$  August 2021. Downloaded on 2021. This article is licensed under a creative is licensed under a creative is licensed under a creative Commons Attribution-Noncommercial 3.1.2.27 August 2021. A second

### Conflicts of interest

There are no conflicts to declare.

### Acknowledgements

We thank the Natural Science Foundation of China (21967011) and the Natural Science Foundation of Jiangxi Province (20181BBE50004), and the State Key Laboratory of Catalytic Materials and Reaction Engineering (ZC06070008).

### Notes and references

- 1 For selected reviews for thioglycosides: (a) R. Caraballo, M. Sakulsombat and O. Ramstrom, ChemBioChem, 2010, 11, 1600–1606; (b) N. Ibrahim, M. Alami and S. Messaoudi, Asian J. Org. Chem., 2018, 7, 2026–2038; (c) G. Lian, X. Zhang and B. Yu, Carbohydr. Res., 2015, 403, 13–22; (d) K. Pachamuthu and R. R. Schmidt, Chem. Rev., 2006, 106, 160–187; (e) A. Borbás, Chem.–Eur. J., 2020, 26, 6090–6101; (f) B. Aguilera, J. Jiménez-Barbero and A. Fernández-Mayoralas, Carbohydr. Res., 1998, 308, 19–27.
- 2 (a) Y. Geng, Q. Qin and X.-S. Ye, J. Org. Chem., 2012, 77, 5255– 5270; (b) C.-C. Liu, C. Zhai, X.-J. Zheng and X.-S. Ye, ACS Chem. Biol., 2016, 11, 1702–1709; (c) X. Xiao, Y. Zhao, P. Shu, X. Zhao, Y. Liu, J. Sun, Q. Zhang, J. Zeng and Q. Wan, J. Am. Chem. Soc., 2016, 138, 13402–13407; (d) L. Yang, Q. Qin and X.-S. Ye, Asian J. Org. Chem., 2013, 2, 30–49; (e) L. Yang and X.-S. Ye, Carbohydr. Res., 2010, 345, 1713–1721; (f) N. Basu, S. Kumar Maity and R. Ghosh, RSC Adv., 2012, 2, 12661–12664; (g) Y. Zeng, Z. Wang,

D. Whitfield and X. Huang, *J. Org. Chem.*, 2008, 73, 7952-7962; (h) D. Budhadev and B. Mukhopadhyay, Carbohydr. Res., 2014, 384, 51–55.

- 3 (a) C. S. Rye and S. G. Withers, Carbohydr. Res., 2004, 339, 699-703; (b) C. Aydillo, I. Compañón, A. Avenoza, J. H. Busto, F. Corzana, J. M. Peregrina and M. M. Zurbano, J. Am. Chem. Soc., 2014, 136, 789–800; (c) B. B. Metaferia, B. J. Fetterolf, S. Shazad-ul-Hussan, M. Moravec, J. A. Smith, S. Ray, M.-T. Gutierrez-Lugo and C. A. Bewley, J. Med. Chem., 2007, 50, 6326–6336; (d) H. Driguez and J. Thiem, Top. Curr. Chem., 1997, 187, 85– 116.
- 4 (a) K. Pachamuthu and R. R. Schmidt, Chem. Rev., 2006, 106, 160–187; (b) S. Sattin and A. Bernardi, Carbohydr. Chem., 2016, 41, 1–25; (c) K. Qin, H. Zhang, Z. Zhao and X. Chen, J. Am. Chem. Soc., 2020, 142, 9382–9388.
- 5 (a) S. V. Pestova, E. S. Izmest'ev, S. A. Rubtsova, A. V. Polukeev and A. V. Kutchin, Russ. J. Org. Chem., 2018, 54, 1041–1044; (b) S. Biswas, C. V. Garcia De Gonzalo, L. M. Repka and W. A. van der Donk, ACS Chem. Biol., 2017, 12, 2965–2969; (c) T. J. Oman, J. M. Boettcher, H. Wang, X. N. Okalibe and W. A. van der Donk, Nat. Chem. Biol., 2011, 7, 78–80.
- 6 (a) A. Mielczarek-Lewandowska, M. L. Hartman and M. Czyz, Apoptosis, 2020, 25, 12–28; (b) L. Li, N.-N. Chen, Q.-D. You and X.-L. Xu, Expert Opin. Ther. Pat., 2021, 31, 67–80.
- 7 B. K. Barr and R. J. Holewinski, Biochemistry, 2002, 41, 4447– 4452.
- 8 A. Burse, F. Castaneda, W. Boland and R. K.-H. Kinne, Int. J. Med. Sci., 2007, 4, 131–139.
- 9 Z. Kamenik, S. Kadlcik, B. Radojevic, P. Jiraskova, M. Kuzma, R. Gazak, L. Najmanova, J. Kopecky and J. Janata, Chem. Sci., 2016, 7, 430–435.
- 10 B. Keith and S. P. Graham, J. Serb. Chem. Soc., 2000, 65, 691– 694.
- 11 M. Schnabelrauch, A. Vasella and S. G. Withers, Helv. Chim. Acta, 1994, 77, 778–799.
- 12 (a) S. K. Das, J. Roy, K. A. Reddy and C. Abbineni, Carbohydr. Res., 2003, 338, 2237–2240; (b) S. Escopy, Y. Singh and A. V. Demchenko, Org. Biomol. Chem., 2019, 17, 8379–8383.
- 13 E. Brachet, J. D. Brion, M. Alami and S. Messaoudi, Chem.– Eur. J., 2013, 19, 15276–15280.
- 14 (a) A. Chabrier, A. Bruneau, S. Benmahdjoub, B. Benmerad, S. Belaid, J. D. Brion, M. Alami and S. Messaoudi, Chem.–Eur. J., 2016, 22, 15006-15010; (b) P. Nauš, L. Lešetický, S. Smrček, I. Tišlerová and M. Štícha, Synlett, 2003, 2117-2122; (c) X. Yuan, Y. Kou, L. Yu, Z.-X. Zhang and W. Xue, Org. Chem. Front., 2015, 2, 1604–1607; (d) F. Zhu, E. Miller, S. Q. Zhang, D. Yi, S. O'Neill, X. Hong and M. A. Walczak, J. Am. Chem. Soc., 2018, 140, 18140–18150.
- 15 (a) R. A. A. Al-Shuaeeb, D. Montoir, M. Alami and S. Messaoudi, J. Org. Chem., 2017, 82, 6720–6728; (b) S. Benmahdjoub, N. Ibrahim, B. Benmerad, M. Alami and S. Messaoudi, Org. Lett., 2018, 20, 4067–4071; (c) E. Brachet, J.-D. Brion, S. Messaoudi and M. Alami, Adv. Synth. Catal., 2013, 355, 477–490; (d) A. L.-S. RA, G. Galvani, G. Bernadat, J. D. Brion, M. Alami and S. Messaoudi, Org. Biomol. Chem., 2015, 13, 10904–10916.
- 16 M. Zhu, G. Dagousset, M. Alami, E. Magnier and S. Messaoudi, Org. Lett., 2019, 21, 5132–5137.
- 17 M. Zhu, M. Alami and S. Messaoudi, Chem. Commun., 2020, 56, 4464–4467.
- 18 For selected reviews of arynes: (a) V. P. Boyarskiy, N. A. Bokach, K. V. Luzyanin and V. Y. Kukushkin, Chem. Rev., 2015, 115, 2698–2779; (b) A. V. M. Dubrovskiy, N. A. Markina and R. C. Larock, Org. Biomol. Chem., 2013, 11, 191–218; (c) C. M. Gampe and E. M. Carreira, Angew. Chem., Int. Ed., 2012, 51, 3766–3778; (d) M. Giustiniano, A. Basso, V. Mercalli, A. Massarotti, E. Novellino, G. C. Tron and J. Zhu, Chem. Soc. Rev., 2017, 46, 1295– 1357; (e) J. He, D. Qiu and Y. Li, Acc. Chem. Res., 2020, 53, 508–519; (f) P. M. Tadross and B. M. Stoltz, Chem. Rev., 2012, 112, 3550–3577; (g) H. H. Wenk, M. Winkler and W. Sander, Angew. Chem., Int. Ed., 2003, 42, 502–528; (h) E. Yoshioka and H. Miyabe, Tetrahedron, 2012, 68, 179– 189; (i) J. Shi, Y. Li and Y. Li, Chem. Soc. Rev., 2017, 46, 1707-1719; (j) I. Pozo, E. Guitián, D. Pérez and D. Peña, Acc. Chem. Res., 2019, 52, 2472–2481; (k) J. Shi, L. Li and Y. Li, Chem. Rev., 2021, 121, 3892-4044; (l) D. B. Werz and A. T. Biju, Angew. Chem., Int. Ed., 2020, 59, 3385–3398. Paper<br>
So Advances Articles. Published on 2021. Downloaded on 2021. This article. Published on 2021. This article is licensed under a creative Commons Articles are a strength that the common Commons Attribution-Noncommerci
	- 19 (a) H. Yoshida, Comprehensive Organic Synthesis II, ed. P. Knochel, Elsevier, Amsterdam, 2nd edn, 2014, pp. 517–579; (b) S. Ghorai and D. Lee, Org. Lett., 2019, 21, 7390–7393; (c) Y. Sumida, T. Sumida, D. Hashizume and T. Hosoya, Org. Lett., 2016, 18, 5600–5603; (d) K. N. Singh, P. Singh, M. Kaur and E. Sharma, ChemistrySelect, 2017, 2, 2213–2218.
- 20 (a) A. Criado, D. Peña, A. Cobas and E. Guitián, Chem.-Eur. J., 2010, 16, 9736–9740; (b) T. Hosoya, T. Hamura, Y. Kuriyama, M. Miyamoto, T. Matsumoto and K. Suzuki, Synlett, 2000, 520–522; (c) J. Zhang, A. C. S. Page, V. Palani, J. Chen and T. R. Hoye, Org. Lett., 2018, 20, 5550–5553; (d) C.-D. Wang and R.-S. Liu, Org. Biomol. Chem., 2012, 10, 8948–8952; (e) M. Sarmah, A. Sharma and P. Gogoi, Org. Biomol. Chem., 2021, 19, 722–737.
- 21 (a) S. Cacchi, G. Fabrizi and A. Goggiamani, Org. Biomol. Chem., 2011, 9, 641-652; (b) R. Karmakar and D. Lee, Chem. Soc. Rev., 2016, 45, 4459–4470; (c) R. A. Dhokale and S. B. Mhaske, Synthesis, 2018, 50, 1–16; (d) H. Tanaka, H. Kuriki, T. Kubo, I. Osaka and H. Yoshida, Chem. Commun., 2019, 55, 6503–6506; (e) M. Feng and X. Jiang, Synthesis, 2017, 49, 4414–4433.
- 22 (a) H. Yoshida, Y. Ito and J. Ohshita, Chem. Commun., 2011, 47, 8512–8514; (b) S. Ghorai and D. Lee, Synlett, 2020, 31, 750–771.
- 23 K. B. Pal, M. Mahanti and U. J. Nilsson, Org. Lett., 2018, 20, 616–619.
- 24 M. Bhardwaj, N. Hussain, I. A. Zargar, A. K. Dash and D. Mukherjee, Org. Biomol. Chem., 2020, 18, 4174–4177.
- 25 N. Yan, Z.-W. Lei, J.-K. Su, W.-L. Liao and X.-G. Hu, Chin. Chem. Lett., 2017, 28, 467–470.
- 26 D. W. Kim, H.-J. Jeong, S. T. Lim and M.-H. Sohn, Angew. Chem., Int. Ed., 2008, 47, 8404–8406.
- 27 L. Zhang, Y. Geng and Z. Jin, J. Org. Chem., 2016, 81, 3542– 3552.