




Cite this: *RSC Adv.*, 2021, 11, 21301

Rigid 3D-spiro chromanone as a crux for efficient antimicrobial agents: synthesis, biological and computational evaluation†

F. H. Elghareeb,^a E. M. Kandil,^a M. Abou-Elzahab,^a M. Abdelmoteleb ^b and M. A. Abozeid ^{*a}

The development of new and effective antimicrobial agents with novel chemical skeletons and working mechanisms is highly desirable due to the increased number of resistant microbes. Different new compounds based upon a 3D-spiro chromanone scaffold such as Mannich bases **2** and **3** in addition to azo dye **4** were synthesized. Besides, the condensation reactions of the hydrazide-spiro chromanone **8** with different ketonic reagents led to the synthesis of pyrazoles (**9** & **10**) and anils (**11** & **13**). Moreover, the methoxyl substituted spiro chromanone **14** was condensed with different hydrazines and hydrazides to give the corresponding hydrazones **15–18** in up to 85% yields. The condensation of the hydrazone **18** with salicylaldehyde yielded coumarinyl spiro chromanone **19** in an excellent yield, whereas its reaction with benzaldehyde followed by hydrazine afforded aminopyrazole derivative **21** in 82% yield. The antimicrobial evaluation suggested that hydrazide **8** has a substantial activity against different microbes (*S. aureus*: *D* = 22 mm, MIC = 1.64 μM; *E. coli*: *D* = 19 mm, MIC = 1.64 μM; *C. albicans*: *D* = 20 mm, MIC = 6.57 μM). Moreover, promising antimicrobial activities were observed for azo dye **4** (*D* = 13–19 mm, MIC = 5.95–11.89 μM), hydrazone **17** (*D* = 17–23 mm, MIC = 1.88–3.75 μM), and aminopyrazole **21** (*D* = 14–19 mm, MIC = 2.24–8.98 μM). The molecular docking revealed that compounds **4**, **8**, **17**, and **21** had good to high binding affinities with different microbial targets such as penicillin-binding proteins (−7.4 to −9.9 kcal), DNA gyrase (−7.8 to −9.0 kcal), lanosterol 14- α demethylase (−8.2 to −11.2 kcal), and exo- β -1,3-glucanase (−8.2 to −11.9 kcal). The QSAR analysis ascertained a good correlation between the antimicrobial activity of 3D-spiro chromanone derivatives and their structural and/or physicochemical parameters.

Received 5th May 2021
Accepted 10th June 2021

DOI: 10.1039/d1ra03497a

rsc.li/rsc-advances

Introduction

Chromone derivatives constitute one of the main families of heterocyclic compounds.¹ Particularly, 4-chromanones have attracted significant attention because of their wide natural occurrence such as sterubin,² morellin,³ and blennolide⁴ (Fig. 1A). They are the precursors for the synthesis of many naturally occurring compounds such as ripariochromene, hematoxylin, and brazilin.⁵ In addition, many synthetic chromanone derivatives revealed diversified biological effects such as anti-HIV,⁶ antibiotic,⁷ antidiabetic,⁸ antifungal,⁹ antiarrhythmic,¹⁰ and antitumor¹¹ as well as different agonist and antagonist activities (Fig. 1A). As a result, the chromanone nucleus became a privileged scaffold in drug design and tuning

the substitution patterns appeared to be efficient structural screws to build up new biologically active leads.¹²

On the other hand, spirocyclic compounds represent a privileged class of cyclic organic compounds that comprise rings fused through a tetrahedral central atom.¹³ The restricted rotation around the spiro-linked atom accommodates the ring planes to be perpendicularly oriented. Based upon the inherent rigidity, higher stability, and precise three-dimensionality, spirocyclic compounds appear as an attractive platform for different applications such as synthesis of chiral ligands^{14a} and organocatalysts^{14b,c} as well as the design of organic optoelectronics.¹⁵ Furthermore, spiro compounds were found in a plethora of naturally occurring compounds that manifested diverse biological activities.¹⁶ Currently, spiro compounds became a catchy scaffold in the modern drug design because of their intrinsic rigidity and precise 3D-dimensionality which maximize H-bonding, π – π stacking, and hydrophobic interactions.¹⁷ These skeletal features could improve drug-receptor binding¹⁷ as well as the ADMET score.¹⁸ Within the spirocyclic compounds, the oxygen-bearing counterparts constitute a distinguishable subclass due to their wide natural occurrence

^aDepartment of Chemistry, Faculty of Science, Mansoura University, Mansoura-35516, Egypt. E-mail: dr.abozeid.chem@gmail.com; mabozeid@mans.edu.eg

^bDepartment of Botany, Faculty of Science, Mansoura University, Mansoura-35516, Egypt

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d1ra03497a



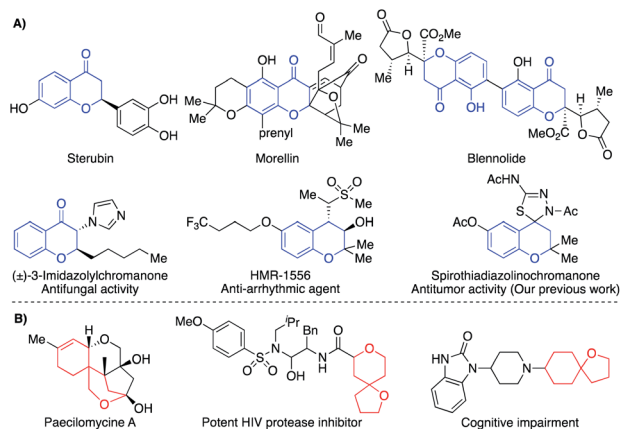


Fig. 1 (A) Representative examples of naturally and biologically active chromanones. (B) Representative examples of natural and bioactive oxaspirocyclic derivatives.

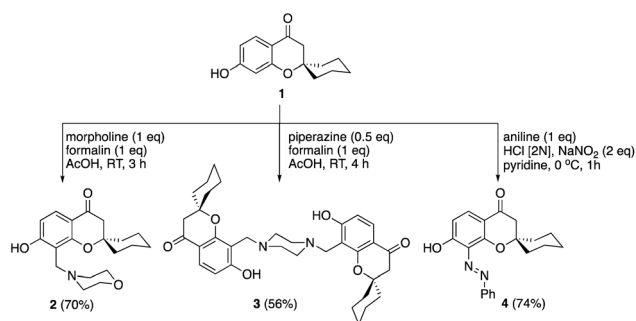
and notable biological activities in addition to the aforementioned skeletal features for spiro skeletons (Fig. 1B).¹⁹

Antimicrobial resistance (AMR) is the ability of microorganisms to grow in the presence of drugs that have been used once against them.^{20a-c} AMR is considered one of the most serious health that leads to persistent infections with high risk of disease transmission. Moreover, increasing the number of multidrug-resistant microbes depicts a cloudy future for the health care systems leading to an urgent need for new antimicrobial agents with novel chemical skeletons as well as working mechanisms.^{20a-c} In our group, we designed different classes of heterocyclic compounds with potent antimicrobial and antiquorum-sensing (QS) activities.^{20d-g} In continuation of our research interest in the synthesis of biologically interesting heterocyclic compounds,^{20d-i} herein we have designed a novel series of spirocyclic chromanone derivatives and evaluated them for their antimicrobial activities with the aid of computational analysis. The construction of a hybrid skeleton combining both chromanone and oxaspiro[5.5]undecane motifs has been the crux of the newly synthesized compounds.

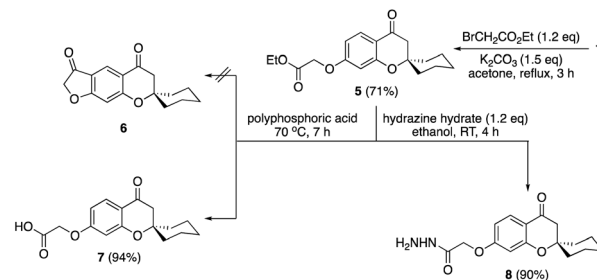
Results and discussion

Chemistry

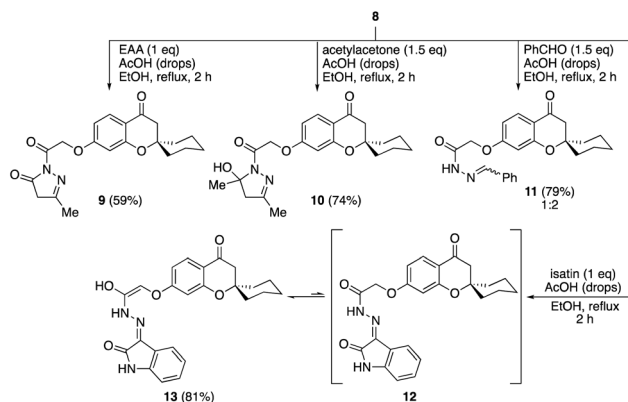
Mannich reaction of the spirocyclic chromanone **1**²¹ with formalin and morpholine afforded the corresponding **8**-



Scheme 1 Synthesis of Mannich bases (**2** & **3**) and azo dye **4**.



Scheme 2 Synthesis of carboxylic acid **7** and hydrazide **8**.

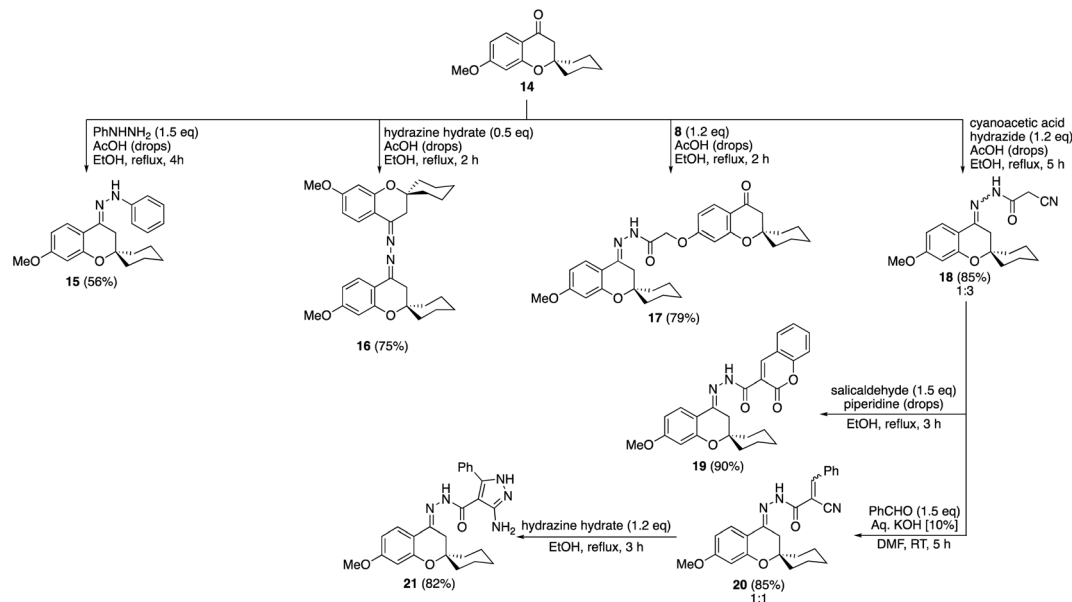


Scheme 3 Reaction of hydrazide **8** with ethyl acetoacetate, acetylacetone, benzaldehyde, and isatin.

morpholinomethyl spirocyclic derivative **2** in a good yield (Scheme 1). Similarly, bis-Mannich base **3** was obtained in 56% yield *via* reaction of spiro chromanone **1** with formalin and piperazine. Moreover, the reaction spiro chromanone **1** with benzene diazonium chloride produced 8-phenyldiazenyl spiro chromanone **4** in 74% yield. The chemical structures of the new compounds **2–4** were secured based on different spectral analyses as shown in the Experimental part.

The chemistry of spiro chromanone **1** was further extended by clipping of ethyl acetate moiety on the pendant phenolic function as in ester **5** (Scheme 2). Treatment of ester **5** with polyphosphoric acid failed to give the desired furanone **6**, and instead, the carboxylic acid **7** was obtained in 94% yield. In addition, the hydrazide **8** was synthesized from the ester **5** in an excellent yield by the reaction with hydrazine hydrate in ethanol at room temperature. The absence of ethoxyl group signals in the ¹H-NMR spectrum of hydrazide **8** in addition to the appearance of NH and NH₂ functionalities at 3303 and 3214 cm⁻¹ in the IR spectrum confirmed its chemical structure.

The condensation of the hydrazide **8** with ethyl acetoacetate (EAA) in the presence of a catalytic amount of acetic acid in refluxing ethanol gave the pyrazolone **9** in 59% yield (Scheme 3). Similarly, the reaction of **8** with acetylacetone afforded dihydropyrazole **10** in 74% yield. The condensation of hydrazide **8** with benzaldehyde afforded the corresponding hydrazone **11** as an *E/Z*-mixture in a very good yield. Additionally, the isatin analogue **13** was obtained by refluxing ethanolic solution of the hydrazide **8** and isatin in presence of few drops of acetic acid.



Scheme 4 Synthesis of compounds 15–21 via condensation reactions of methyl protected spiro chromanone 14.

The condensation of the methyl protected spiro chromanone 14²² with phenylhydrazine afforded the corresponding hydrazone 15 in 56% yield (Scheme 4). Moreover, the acid-mediated synthesis of the bis-hydrazone 16 was accomplished by reaction of 14 with hydrazine hydrate in refluxing ethanol. Reacting precursor 14 with the previously synthesized hydrazide 8 gave hydrazone 17 clipped with two spiro chromanone motifs in 79% yield. Similarly, cyanoacetic acid hydrazide successfully was

condensed with the spiro chromanone 14 under similar conditions to produce the cyanoacetyl hydrazone 18 in 85% yield (Scheme 4). The reaction of hydrazone 18 with salicylaldehyde in the presence of a catalytic amount of piperidine gave coumarinyl substituted spiro chromanone 19 in 90% yield. Moreover, the basic condensation of cyanoacetyl hydrazone 18 with benzaldehyde afforded benzylidene 20 which gave aminopyrazole 21 via reaction with hydrazine hydrate in 82% yield.

Table 1 Evaluation of antimicrobial activity

Compound	<i>S. aureus</i> ^a			<i>E. coli</i>			<i>C. albicans</i>			
	<i>D</i> ^b (mm)	AI ^c (%)	MIC ^d (μM)	<i>D</i> ^b (mm)	AI ^c (%)	MIC ^d (μM)	<i>D</i> ^b (mm)	AI ^c (%)	MIC ^d (μM)	IC ₅₀ ^e (mM)
2	10	41.7	96.56	5	19.2	—	9	33.3	—	23.86 ± 1.9
3	NA ^f	—	—	NA ^f	—	—	2	7.4	—	65.63 ± 3.7
4	18	75	11.89	13	50	5.95	19	70.4	11.89	7.31 ± 0.5
5	8	33.3	—	2	7.7	—	12	44.4	100.51	41.22 ± 2.8
7	4	16.7	—	NA ^f	—	—	5	18.5	—	32.74 ± 2.5
8	22	91.7	1.64	19	73.1	1.64	20	74.1	6.57	23.86 ± 1.9
9	16	66.7	10.80	11	42.3	10.80	15	55.5	21.60	8.90 ± 0.7
10	15	62.5	20.70	8	30.8	—	13	48.1	41.40	19.04 ± 1.5
11	11	45.8	40.77	4	15.4	—	7	25.9	—	57.73 ± 3.4
13	NA ^f	—	—	NA ^f	—	—	NA ^f	—	—	81.08 ± 4.2
15	15	62.5	11.89	10	38.5	23.78	14	51.8	23.78	10.53 ± 0.9
16	NA ^f	—	—	NA ^f	—	—	NA ^f	—	—	48.58 ± 3.0
17	20	83.3	1.88	17	65.4	1.88	23	85.2	3.75	54.67 ± 3.3
18	5	20.8	—	2	7.7	—	10	37	97.74	62.49 ± 3.5
19	7	29.2	—	NA ^f	—	—	4	14.8	—	29.15 ± 2.2
20	2	8.3	—	NA ^f	—	—	3	11.1	—	76.57 ± 3.9
21	19	79.2	4.49	14	53.8	2.24	18	66.7	8.98	14.92 ± 1.2
Ciprofloxacin	24	100	1.51	26	100	1.51	NA ^f	—	—	—
Clotrimazole	NA ^f	—	—	NA ^f	—	—	27	100	2.90	—

^a Compounds 2–21 revealed weak or no activity against methicillin-resistant *Staphylococcus aureus* (MRSA) compared to ciprofloxacin. ^b *D*: diameter of inhibition zone. ^c AI: activity index (%) = (diameter of the inhibition zone of the tested compound/diameter of the inhibition zone of the reference drug) × 100. ^d MIC: Minimum Inhibitory Concentration. ^e Cytotoxicity against hepatocellular carcinoma (HePG-2), IC₅₀ is the concentration required to inhibit 50% of cell growth, values were represented as mean ± SD of triplicate readings. ^f NA: no activity.



Table 2 Binding affinities against different microbial targets

Compound	Binding affinity (kcal mol ⁻¹)			
	1VQQ	5MMN	5TZ1	3N9K
1	-7	-5.4	-7.2	-8.4
2	-7.5	-6.7	-8.2	-7.0
3	-9.5	-8.7	-8.8	-7.5
4	-8.4	-8.1	-9.4	-8.5
5	-6.8	-7.6	-7.7	-8.7
7	-6.9	-7.2	-8.0	-9.2
8	-7.4	-8.0	-8.2	-8.2
9	-8.7	-9.0	-9.0	-10.0
10	-8.8	-8.5	-9.0	-9.9
11	-8.8	-7.8	-9.1	-9.1
12	-9.8	-9.0	-10.6	-10.5
13	-9.8	-8.9	-10.7	-10.7
14	-6.6	-6.6	-7.2	-8.5
15	-7.7	-8.0	-9.1	-9.5
16	-8.5	-7.4	-10.1	-10.2
17	-9.9	-9.0	-10.7	-11.9
18	-7.8	-7.5	-8.2	-9.6
19	-9.5	-8.3	-10.7	-11.1
20	-8.4	-8.1	-10.8	-10.1
21	-8.8	-7.8	-11.2	-11.6
Ciprofloxacin	-6.9	-7.0	-8.0	-8.1
Clotrimazole	-7.3	-6.1	-8.0	-5.7

All the compounds were well characterized by spectroscopic methods (ESI material 1†).

Prediction of biological activities and physicochemical parameters

Biological activities of the synthesized derivatives were predicted using PASS online software *via* comparing the descriptors derived from their structures with descriptors of biologically active drugs in the database.^{20d-f,23} Results illustrated that some compounds might act as promising anti-infective drugs including antibacterial, antibiotic, antifungal, and antiviral activities (ESI material 2†). Other activities such as anti-inflammatory and anti-cancer in addition to membrane integrity agonists were also anticipated. Moreover, the pharmacokinetics and drug-likeness prediction were calculated using an *in silico* ADMET (ESI material 3†).²⁴ Compounds **8** and **18** might have better interaction abilities as they have eight rotatable bonds and ten hydrogen bond acceptors. Most of the synthesized derivatives gave log *P* values < 5 suggesting good cell membrane permeabilities. The compounds with good water solubility will have a good delivery in small doses. All compounds had a bioactivity score of 0.55 indicating higher biological activities in the next steps of drug discovery. Toxicity assessment by AdmetSAR-2 online predictor suggested that the derivatives were likely have no toxicity risk.

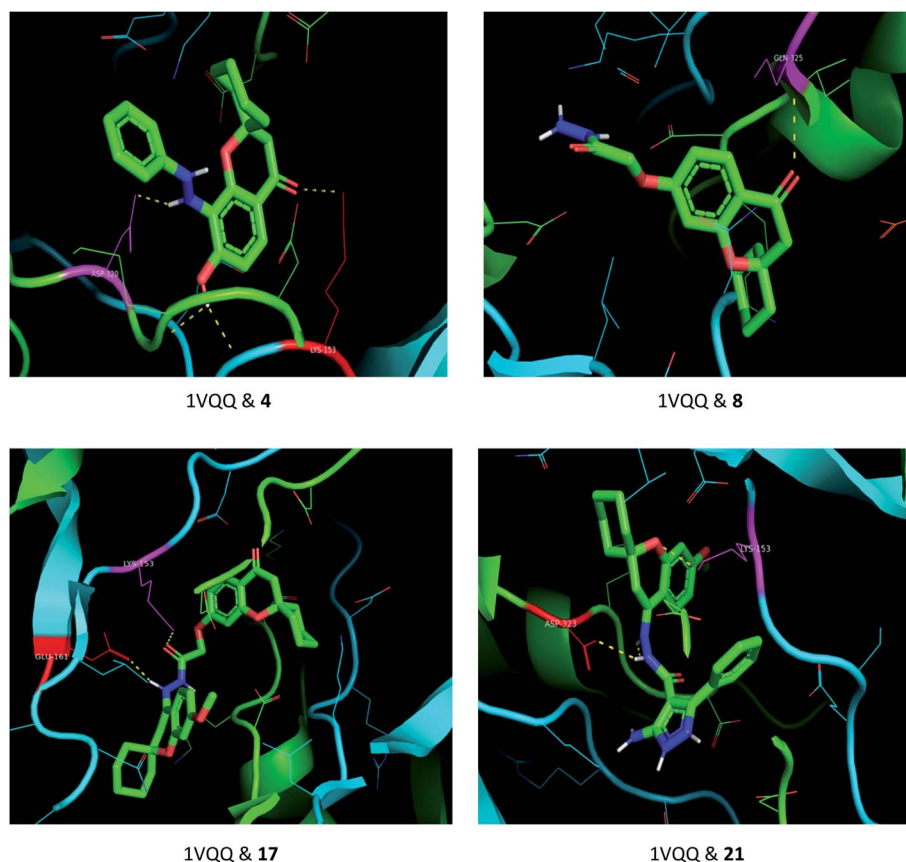


Fig. 2 Binding modes between *S. aureus* penicillin-binding protein (1VQQ) and compounds **4**, **8**, **17** and **21**.



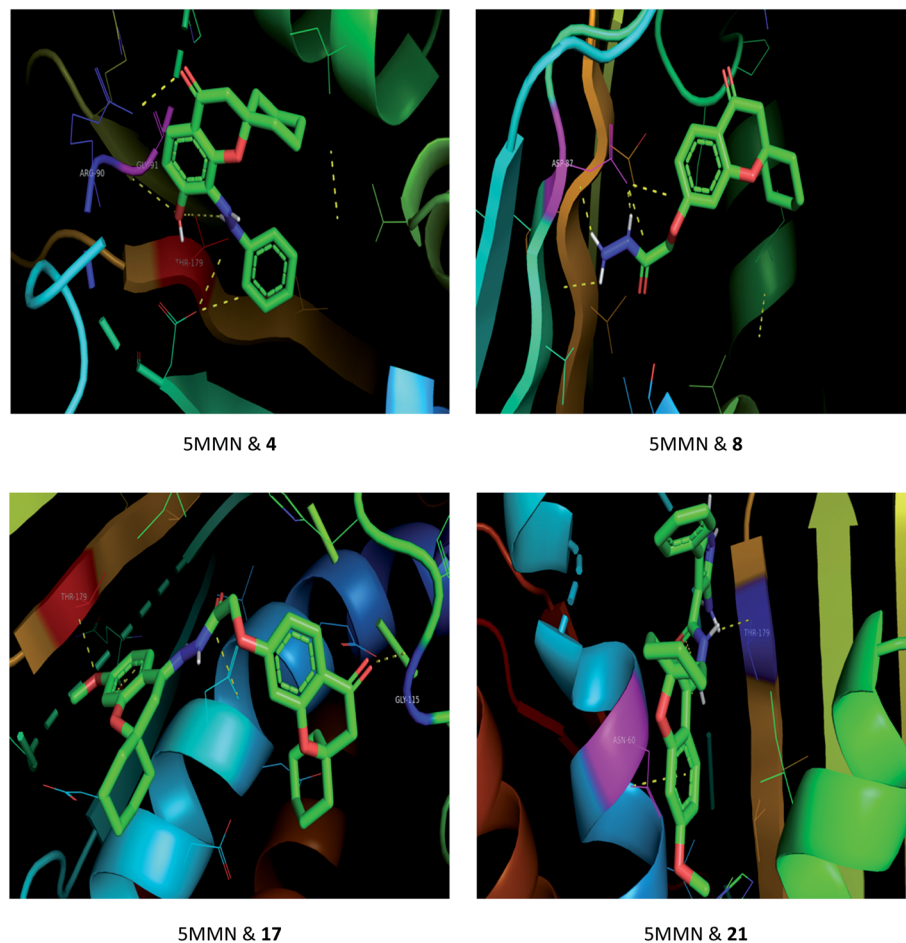


Fig. 3 Binding modes between *E. coli* DNA gyrase (5MMN) and compounds 4, 8, 17 and 21.

Antimicrobial activity and toxicity

The antibacterial activities of the synthesized compounds were evaluated against Gram-positive bacteria such as *Staphylococcus aureus* and methicillin-resistant *Staphylococcus aureus* (MRSA) in addition to Gram-negative bacteria such as *Escherichia coli* using agar-diffusion protocol compared with ciprofloxacin as reference antibiotic.^{25,26} Similarly, the antifungal activity was assessed against *Candida albicans* compared with clotrimazole as a standard antifungal drug. The results were reported as the average diameter of inhibition zones (*D*) in millimeters. Compared to the inhibition zones for both standard drugs, the activity indices were calculated according to the equation: activity index (%) = (diameter of the inhibition zone of the tested compound/diameter of the inhibition zone of the reference drug) × 100. The minimal inhibitory concentrations (MICs) for the synthesized compounds that revealed inhibition zones with diameters of ≥10 mm were determined using the two-fold dilution method.²⁷ The diameter of inhibition zones, activity indices, and MICs were summarized in Table 1.

However different compounds were active against all tested microbes, MRSA revealed high antimicrobial resistance (ESI material 1†). In the case of *S. aureus*, hydrazide 8 showed the best antibacterial activity with 91.7% AI and 0.5 µg mL⁻¹ MIC.

Besides, azo dye 4, hydrazone 17, and aminopyrazole 21 revealed noticeable activities with MICs (1–4 µg mL⁻¹). In the case of *E. coli*, hydrazide 8 was the only strong candidate with 73.1% AI and 0.5 µg mL⁻¹ MIC. In the case of *C. albicans*, azo dye 4, hydrazide 8, and hydrazone 17 were the best compounds with an activity index of more than 70% and minimal inhibitory concentration of 2–4 µg mL⁻¹. Other compounds showed low to moderate activities against *S. aureus*, *E. coli*, and *C. albicans*.

The presence of hydrazide functionality was essential to impart strong antimicrobial activity to compound 8 compared to less active analogues 5, 7, and 9–11 as seen in the previously published literature.²⁸ The combination of spiro chromanone and isatin moieties was very deleterious to give the inactive compound 13. Phenylazo was more fruitful substituent than morpholinomethyl as clearly observed by comparing the antimicrobial activities of compounds 2 and 4. Symmetrical bis-compounds bearing two spiro chromanone moieties such as 3 and 16 were proven to be inactive as antimicrobial. Unsymmetrical bis-compound bearing two spiro chromanone motifs 17 showed very good antibacterial and antifungal activities. Comparing the structures of compounds 18–21 highlighted the role of aminopyrazole moiety in compound 21 to have a selective antibacterial activity for *S. aureus*.²⁹

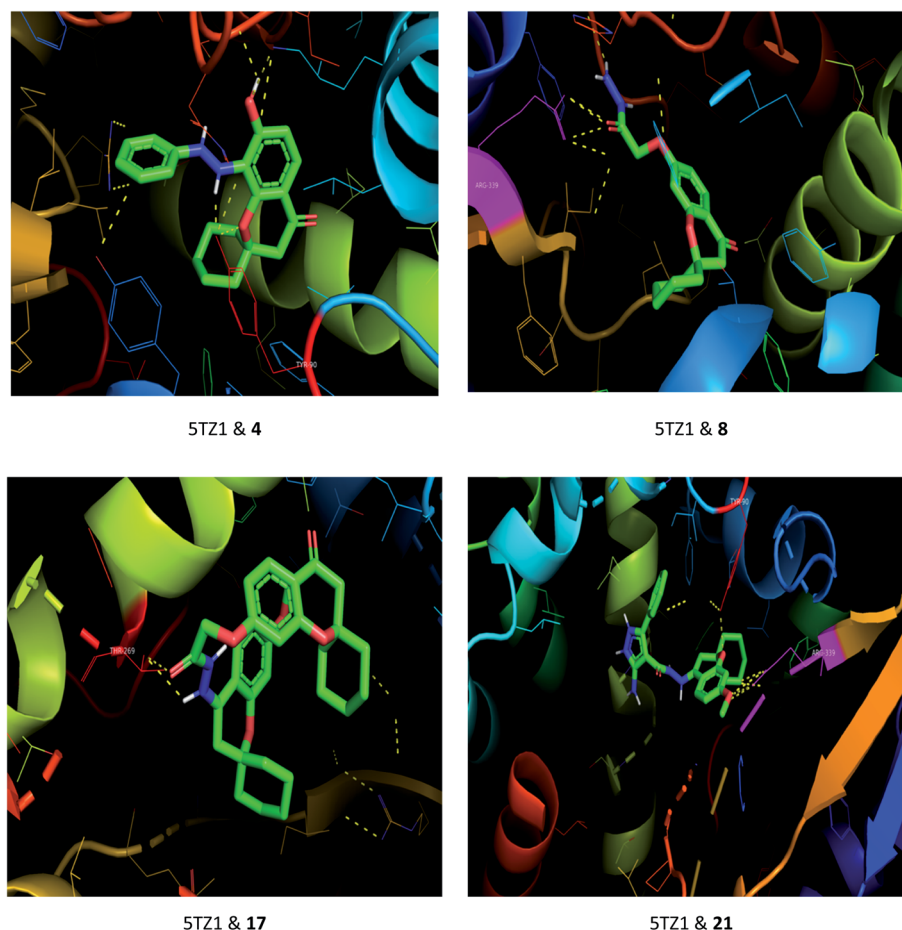


Fig. 4 Binding modes between lanosterol 14- α demethylase (5TZ1) and compounds 4, 8, 17 and 21.

The cytotoxicity of the synthesized compounds was evaluated against hepatocellular carcinoma (HePG-2) using MTT assay. The values of IC_{50} which is the required concentration to suppress 50% of the cell growth were presented in Table 1. Mostly, IC_{50} of the compounds 4, 8, 17, and 21 were higher than their MICs against *Staphylococcus aureus*, *Escherichia coli*, and *Candida albicans*. This observation indicated that the antimicrobial activity is not resulted from the general toxicity, but it could be rationalized to the selective inhibitory action against the tested microbes.³⁰ Thus, compounds 4, 8, 17, and 21 could be promising antimicrobial candidates.

Molecular docking

The drug–target interactions were investigated using molecular docking analysis in order to clarify the exact binding sites and to understand the mode of action for the best biologically active compounds.³¹ Penicillin-binding protein from *S. aureus* (PDB ID: 1VQQ)³² and DNA gyrase from *E. coli* (PDB ID: 5MMN)³³ were used as biological targets to simulate the mechanism of antibacterial activities, while lanosterol 14- α demethylase (PDB ID: CYP51)³⁴ and *exo*-beta-1,3-glucanase (PDB ID: 3N9K)³⁵ from *C. albicans* were the fungal targets. Protein–ligand docking scores were presented in Table 2. Compounds 4, 8, 17, and 21

revealed good to high binding affinities scores against the four targets. In the case of *S. aureus* penicillin-binding protein (1VQQ), compounds 4, 8, 17, and 21 with binding affinities from -7.4 to -9.9 kcal mol⁻¹ revealed polar interactions with ASP-320 and LYS-153; GLN-325; GLU-161; ASP-323 and LYS-153, respectively (Fig. 2). The binding affinity of compounds 4, 8, 17, and 21 (-7.8 to -9.0 kcal mol⁻¹) with *E. coli* DNA gyrase (5MMN) was rationalized to the polar interactions with amino acid residues: ARG-90, GLY-91 and THR-179; ASP-87; THR-179 and GLY-115; THR-179 and ASN-60, respectively (Fig. 3). In addition, compounds 4, 8, 17 and 21 revealed enhanced binding affinities (-8.2 to -11.2 kcal mol⁻¹) with lanosterol 14- α demethylase (5TZ1), and this was explained *via* polar interactions with TYR-90; ARG-339; THR-269; TYR-90 and ARG-339 residues as shown in Fig. 4. In the case of *exo*-beta-1,3-glucanase (3N9K), compounds 4, 8, 17, and 21 also generated polar interactions with HIS-225 and HIS-252; HIS-134, ASN-145, TYR-254 and SER-291; ASN-145 and TRP-276; ASN-145 and GLU-191, respectively, which were in agreement with the binding affinities (-8.2 to -11.6 kcal mol⁻¹) (Fig. 5). The promising antimicrobial activities of compounds 4, 8, 17, and 21 might be rationalized to abilities to inhibit the biosynthesis of fungal sterols and cell wall branched β -1,3-D-glucans.



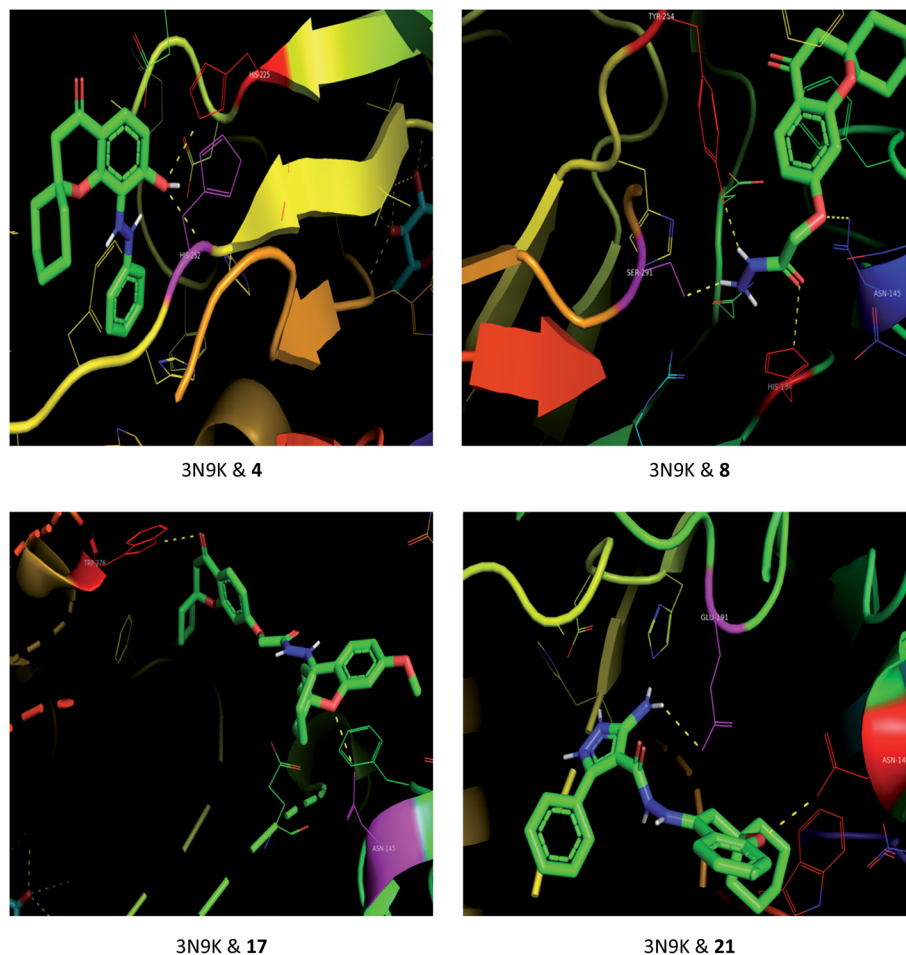


Fig. 5 Binding modes between exo-beta-1,3-glucanase (3N9K) and compounds 4, 8, 17 and 21.

Quantitative structure–activity relationship (QSAR)

The QSAR analysis³⁶ was conducted in order to interpret the observed antimicrobial activity based on the structural characteristics of the synthesized spiro chromanone derivatives. The number of calculated descriptors through the PaDEL-Descriptor was 770.³⁷ Then, the dataset of descriptors was imported into QSARINS.³⁸ To mitigate redundancy in the dataset, descriptors with 95% pairwise correlations and 80% sharing the same values were deleted. The whole dataset consisting of all compounds was divided into training and test sets using a random selection of 25% of the total compounds. The selected descriptors were used as independent variables while antibacterial and antifungal activities (diameter of inhibition zone) were used as response variables.

The activity trend against *S. aureus* was evaluated using the following QSAR equation [R_2 : 0.5279, R_2 adj: 0.4421, $R_2 - R_2$ adj: 0.0858, RMSE tr: 5.4764, MAE tr: 4.8993, CCC tr: 0.691, s : 6.1782, F : 6.1508, Q2LOO: 0.2923 (R_2 = coefficient of determination; R_2 adj = adjusted coefficient of determination; RMSE tr = root mean square error for training set; CCC tr = concordance correlation coefficient for training set; s = standard error of estimate; F = Fischer's value; Q2LOO = cross-validation coefficient/Leave-One-Out)]. The activity against *E. coli* was evaluated

using this QSAR equation [R_2 : 0.5449, R_2 adj: 0.469, $R_2 - R_2$ adj: 0.0759, RMSE tr: 5.3517, MAE tr: 4.2506, CCC tr: 0.7054, s : 5.9834, F : 7.1839, Q2LOO: 0.354]. Both models were internally and externally validated and selected based on the best fitting (highest R_2), and robustness (highest Q2LOO) values. Similarly, the activity against *C. albicans* was described by following equation [R_2 : 0.5815, R_2 adj: 0.5118, $R_2 - R_2$ adj: 0.0697, RMSE tr: 5.3328, MAE tr: 4.3787, CCC tr: 0.7354, s : 5.9622, F : 8.3374, Q2LOO: 0.4029]. High values of R_2 and Q2LOO for the equation illustrated good fitting and stability of the QSAR model. Internal and external predictive ability in addition to randomization of the developed models was ascertained.

The scatter plots of the experimental antibacterial and antifungal activities (exp. endpoint) versus calculated activities by QSAR models [pred. by model eq. & pred. by Leave-One-Out (LOO)] were drawn and depicted in Fig. 6. These plots showed that compound 8 had a strong agreement between the experimental antimicrobial activity and those predicted by QSAR models and lied close to ciprofloxacin and clotrimazole along the regression line. In addition, compounds 4, 17, and 21 were the nearest to ciprofloxacin and clotrimazole compared to the other compounds. Therefore, the predictive ability of the QSAR

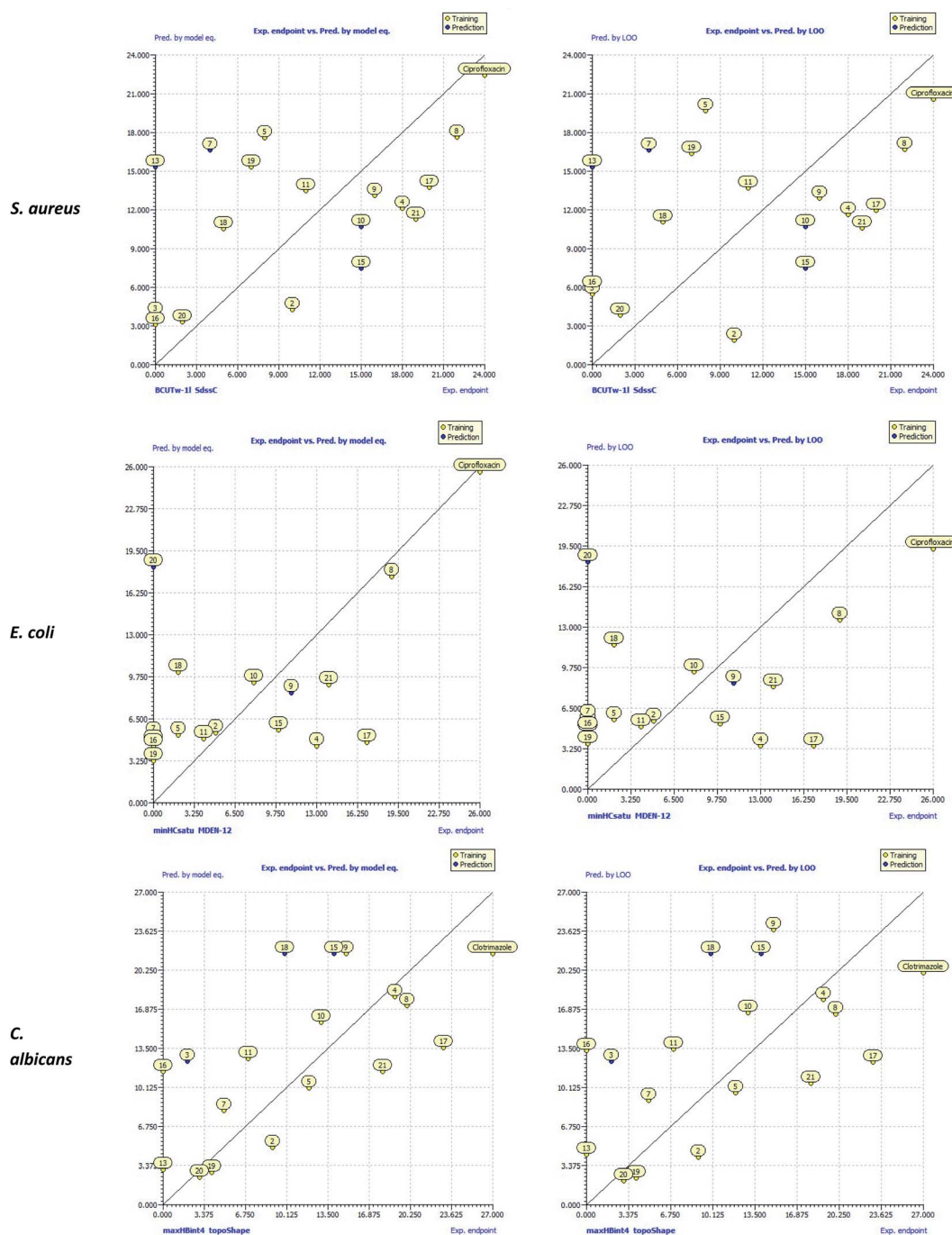


Fig. 6 Scatter plots of the experimental antibacterial and antifungal activities (exp. endpoint) versus the calculated activities by QSAR models [pred. by model eq. & pred. by Leave-One-Out (LOO)].

model supported the experimental antibacterial and antifungal activities for these compounds.

Conclusion

Novel 3D-spiro chromanone derivatives such as Mannich bases (2 & 3), azo dye 4, pyrazolines (9 & 10), and hydrazones (11 & 13) were efficiently synthesized by adapting the versatile 7-hydroxy-spiro[chromane-2,1'-cyclohexan]-4-one (1) as a precursor.

Moreover, the condensation of methoxylspirochromanone 14 with different hydrazines and hydrazides gave the corresponding hydrazones 15–18 in yields up to 85%. The coumarinyl spiro chromanone 19 was furnished by the reaction of the hydrazone 18 with salicylaldehyde, whereas the aminopyrazole 21 was synthesized indirectly from the same precursor *via* the reaction with benzaldehyde followed by hydrazine. The antimicrobial evaluation against different microbes revealed the potent activity of hydrazone 8 (*S. aureus*: $D = 22$ mm, MIC = 1.64 μ M; *E.*



coli: $D = 19$ mm, MIC = 1.64 μM ; *C. albicans*: $D = 20$ mm, MIC = 6.57 μM). Additionally, promising activities of azo dye **4** ($D = 13$ – 19 mm, MIC = 5.95–11.89 μM), hydrazone **17** ($D = 17$ – 23 mm, MIC = 1.88–3.75 μM), and aminopyrazole **21** ($D = 14$ – 19 mm, MIC = 2.24–8.98 μM) were observed. These noticeable antimicrobial activities were compatible with the good to high binding affinities (-7.4 to -11.9 kcal) towards different targets as seen in molecular docking analysis. In addition, QSAR studies highlighted the role of structural and/or physicochemical parameters of 3D-spirochromanones to predict antimicrobial activity. Further studies synthesis of more effective 3D-spirochromanone derivatives and screening against more challenging microbes such as Methicillin-resistant *Staphylococcus aureus* (MRSA) in addition to enzyme inhibitory studies are currently underway and will be reported in the due time.

Experimental section

General

Melting points for all new compounds were measured using a Gallenkamp apparatus. The IR spectra for all newly synthesized compounds were recorded using KBr method on a Mattson 5000 FT-IR spectrophotometer (Faculty of Pharmacy, Mansoura Uni., Egypt) or Thermo Fisher Nicolet iS 10, USA (Faculty of Science, Mansoura Uni., Egypt). The ^1H - and ^{13}C -NMR spectra were measured on Bruker Avance III 400 MHz (Faculty of Pharmacy, Mansoura Uni., Egypt), and/or JEOL ECA II 500 MHz (NMR Unit, Faculty of Science, Mansoura Uni., Egypt). The signal multiplicities were reported as: s = singlet, brs = broad singlet, d = doublet, dd = doublet of doublets, and m = multiplet. ^{13}C NMR spectra were measured on JEOL ECA II 125 MHz (NMR Unit, Faculty of Science, Mansoura Uni., Egypt). CDCl_3 , $\text{DMSO}-d_6$ were used as NMR solvents, and the value of chemical shifts (δ) was in ppm unit. The EI-Mass analyses were performed on Thermo Fisher scientific DSQ II GC/MS with Kratos MS-70 ev spectrometer (Faculty of Pharmacy, Al-Azhar Uni., Egypt). The LC-MS analysis was performed on LCMS-2020-Single Quadrupole mass spectrometer from Shimadzu (Faculty of Pharmacy, Al-Azhar Uni., Egypt). High resolution mass spectrometry (HRMS) measurements were carried out using Thermo Fisher Scientific Q Exactive Hybrid Quadrupole-Orbitrap Mass Spectrometer (National Research Centre, Egypt). Elemental analyses (C, H, and N) were performed at the microanalytical unit, Cairo Uni., Egypt. Reaction progress and product mixtures were routinely monitored by thin layer chromatography (TLC) on silica gel plates. Antibacterial and antifungal screenings were performed at the Faculty of Pharmacy, Mansoura Uni., Egypt.

Synthesis

7-Hydroxy-8-(morpholinomethyl)spiro[chromane-2,1'-cyclohexan]-4-one (2). To a solution of **1** (4 mmol, 0.93 g) in acetic acid (20 mL), formalin (1 eq, 4 mmol, 0.14 mL) and morpholine (1 eq, 4 mmol, 0.34 mL) were added. After stirring for 3 h at room temperature, the reaction mixture was poured into crushed ice and neutralized with liquid ammonia. The

precipitate was filtered off and purified using column chromatography (eluent: petroleum ether : ethyl acetate = 3 : 1) to give **2** as colorless crystals. Yield (0.93 g, 70%); m.p. = 130–131 $^\circ\text{C}$; IR (KBr): ν/cm^{-1} = 3430 (OH), 2860–2939 ($\text{CH}_{\text{Aliphatic}}$), 1670 (CO); ^1H NMR ($\text{DMSO}-d_6$, 400 MHz) δ (ppm): 1.23–1.71 (m, 10H, $(\text{CH}_2)_5$), 2.57 (s, 2H, CH_2), 2.41–2.51 (m, 4H, $\text{N}(\text{CH}_2)_2$), 3.48–3.57 (m, 4H, $\text{O}(\text{CH}_2)_2$), 3.62 (s, 2H, CH_2), 6.42 (d, 1H, Ar-H, $J = 8.80$ Hz), 7.48 (d, 1H, Ar-H, $J = 8.80$ Hz), 9.72 (brs, 1H, OH); MS (EI, 70 eV): m/z (%) = 331 [M^+ , 27.7%], 298 [100.0%], 248 [89.7%], 229 [75.8%], 206 [57.4%], 180 [69.7%], 99 [67.4%]; anal. calcd for $\text{C}_{19}\text{H}_{25}\text{NO}_4$ (331.18): C, 68.86; H, 7.60; N, 4.23%. Found: C, 68.92; H, 7.51; N, 4.29%.

8,8''-(Piperazine-1,4-diylbis(methylene))bis(7-hydroxyspiro[chromane-2,1'-cyclohexan]-4-one) (3). A mixture of **1** (4 mmol, 0.93 g), formalin (1 eq, 4 mmol, 0.14 mL) and piperazine (0.5 eq, 2 mmol, 0.17 g) in acetic acid (20 mL) was stirred for 4 h at room temperature. After that, the reaction mixture was poured into crushed ice and neutralized with liquid ammonia. The formed precipitate was filtered off and recrystallized from ethanol to yield **3** as pale grey crystals. Yield (1.3 g, 56%); m.p. 190–191 $^\circ\text{C}$; IR (KBr): ν/cm^{-1} = 3415 (OH), 2825–2942 ($\text{CH}_{\text{Aliphatic}}$), 1678 (CO); ^1H NMR ($\text{DMSO}-d_6$, 500 MHz) δ (ppm): 1.45–1.85 (m, 20H, $(\text{CH}_2)_5$, $(\text{CH}_2)_5$), 2.07 (s, 8H, 4 CH_2), 2.74 (s, 4H, 2 CH_2), 3.15 (s, 4H, 2 CH_2), 6.52 (d, 2H, Ar-H, $J = 9.00$ Hz), 7.42 (d, 2H, Ar-H, $J = 9.00$ Hz), 10.21 (brs, 1H, 2 OH); MS (EI, 70 eV): m/z (%) = 574 [M^+ , 74%], 556 [36.9%], 511 [51.4%], 471 [72%], 413 [63.3%], 345 [72.3%], 288 [72.1%], 242 [68.6%], 199 [59.9%], 146 [100.0%], 113 [48.2%]; anal. calcd for $\text{C}_{34}\text{H}_{42}\text{N}_2\text{O}_6$ (574.30): C, 71.06; H, 7.37; N, 4.87%. Found: C, 71.26; H, 7.19; N, 4.72%.

7-Hydroxy-8-(phenyldiazenyl)spiro[chromane-2,1'-cyclohexan]-4-one (4). A stirred solution of aniline (2 mmol, 0.18 mL) in HCl [2 N] (3 mL) in an ice-salt bath was diazotized with a cold solution of sodium nitrite (2 eq, 4 mmol, 0.28 g) in a least amount of water. This cold diazotized amine was added in a dropwise manner to a cold solution of **1** (1 eq, 2 mmol, 0.46 g) in pyridine (10 mL). The stirring was continued for further 30 minutes in the ice bath and for further 2 h at room temperature. The formed precipitate was filtered off, washed with water and recrystallized from diluted ethanol to yield **4** as pink crystals. Yield (0.5 g, 74%); m.p. 168–169 $^\circ\text{C}$; IR (KBr): ν/cm^{-1} = 3122 (OH), 2854–2936 ($\text{CH}_{\text{Aliphatic}}$), 1653 (CO); ^1H NMR ($\text{DMSO}-d_6$, 400 MHz) δ (ppm): 1.44–1.83 (m, 10H, $(\text{CH}_2)_5$), 2.61 (s, 2H, CH_2), 6.27 (d, 1H, Ar-H, $J = 8.50$ Hz), 7.22–7.26 (m, 2H, Ar-H), 7.43–7.56 (m, 4H, Ar-H), 10.50 (brs, 1H, OH); MS (EI, 70 eV): m/z (%) = 336 [M^+ , 24%], 292 [33.9%], 232 [91.4%], 180 [32%], 115 [73.3%], 99 [100.0%], 54 [52.3%]; anal. calcd for $\text{C}_{20}\text{H}_{20}\text{N}_2\text{O}_3$ (336.15): C, 71.41; H, 5.99; N, 8.33%. Found: C, 71.32; H, 6.12; N, 8.41%.

Ethyl 2-((4-oxospiro[chromane-2,1'-cyclohexan]-7-yl)oxy)acetate (5). A mixture of **1** (2 mmol, 0.46 g) and ethyl bromoacetate (1 eq, 2 mmol, 0.22 mL) in acetone (20 mL) was refluxed for 3 h in the presence of K_2CO_3 (1.5 eq, 2.5 mmol, 0.34 g) the reaction mixture was then cooled to room temperature and poured into crushed ice. The separated product was filtered off and purified using column chromatography (eluent: petroleum ether : ethyl acetate = 3 : 1) to give **5** as a colorless powder. Yield (0.45 g, 71%); m.p. 73–74 $^\circ\text{C}$; IR (KBr): ν/cm^{-1} = 2863–2942



(CH_{Aliphatic}), 1764 (CO), 1669 (CO); ¹H NMR (DMSO-*d*₆, 400 MHz) δ (ppm): 0.98 (t, 3H, CH₃, *J* = 6.80 Hz), 1.03–1.62 (m, 10H, (CH₂)₅), 3.11 (s, 2H, CH₂), 3.94 (q, 2H, CH₂, *J* = 6.80 Hz), 4.63 (s, 2H, CH₂), 6.27–6.37 (m, 2H, Ar-H), 7.41 (d, 1H, Ar-H, *J* = 8.50 Hz); ¹³C NMR (CDCl₃, 125 MHz) δ (ppm): 14.1, 21.4, 25.1, 34.8, 47.8, 61.6, 65.2, 80.5, 102.3, 109.0, 115.4, 128.4, 161.4, 164.1, 168.1, 191.1; MS (EI, 70 eV): *m/z* (%) = 318 [M⁺, 6.1%], 297 [46.1%], 287 [100.0%], 240 [58.1%], 187 [88.1%], 160 [37.3%], 123 [63.1%], 46 [82.1%]; LCMS: *m/z*: 319 [(M + H)⁺]; HRMS (ESI): *m/z* calcd for C₁₈H₂₃O₅ [(M + H)⁺] 319.1540, found 319.1528; anal. calcd for C₁₈H₂₂O₅ (318.15): C, 67.91; H, 6.97%. Found: C, 67.85; H, 6.99%.

2-((4-Oxospiro[chromane-2,1'-cyclohexan]-7-yl)oxy)acetic acid (7). The ester **5** (3 mmol, 0.95 g) was added to a freshly prepared polyphosphoric acid (PPA) [prepared by portion wise addition of P₂O₅ (4.5 g) to phosphoric acid (85%) (10 mL) under heating on a water bath till the formation of viscous solution]. After that, the reaction mixture was stirred at 70 °C for 6 h. After complete consumption of ester **5**, the reaction mixture was poured onto crushed ice. The formed precipitate was filtered off then purified using column chromatography (eluent: petroleum ether : ethyl acetate 3 : 1) to yield **7** as a colorless powder. Yield (0.82 g, 94%), m.p. 140–141 °C; IR (KBr): ν /cm⁻¹ = 3420–3217 (COOH), 2866–2938 (CH_{Aliphatic}), 1767 (CO), 1641 (CO); ¹H NMR (DMSO-*d*₆, 500 MHz) δ (ppm): 1.46–1.82 (m, 10H, (CH₂)₅), 2.67 (s, 2H, CH₂), 4.76 (s, 2H, CH₂), 6.46 (d, 1H, Ar-H, *J* = 3.0 Hz), 6.57 (dd, 1H, Ar-H, *J* = 2.00, 8.00 Hz), 7.62 (d, 1H, Ar-H, *J* = 8.00 Hz), 13.14 (brs, 1H, COOH); ¹³C NMR (DMSO-*d*₆, 125 MHz) δ (ppm): 21.0, 24.6, 33.9, 47.0, 64.7, 80.2, 102.1, 109.3, 114.5, 127.5, 160.8, 164.2, 169.6, 190.3; MS (EI, 70 eV): *m/z* (%) = 290 [M⁺, 10.6%], 143 [42.8%], 115 [87.3%], 91 [28.7%], 76 [53.2%], 43 [100.0%]; LCMS: *m/z*: 291 [(M + H)⁺]; HRMS (ESI): *m/z* calcd for C₁₇H₁₉O₅ [(M + H)⁺] 291.1227, found 291.1215; anal. calcd for C₁₆H₁₈O₅ (290.12): C, 66.20; H, 6.25%. Found: C, 66.02; H, 6.48%.

2-((4-Oxospiro[chromane-2,1'-cyclohexan]-7-yl)oxy)aceto-hydrazide (8). To a stirred solution of **5** (3 mmol, 0.96 g) in ethanol (15 mL) in an ice-salt bath, hydrazine monohydrate (1 eq, 3 mmol, 0.15 mL) was added. The stirring was continued for further 4 h at room temperature. The produced product was filtered off and recrystallized from ethanol giving **8** as yellow needles. Yield (0.82 g, 90%); m.p. 194–195 °C; IR (KBr): ν /cm⁻¹ = 3303 (NH), 3270, 3214 (NH₂), 2857–2926 (CH_{Aliphatic}), 1687 (CO); ¹H NMR (DMSO-*d*₆, 500 MHz) δ (ppm): 1.45–1.85 (m, 10H, (CH₂)₅), 2.67 (s, 2H, CH₂), 4.34 (brs, 2H, NH₂), 4.55 (s, 2H, CH₂), 6.51 (d, 1H, Ar-H, *J* = 2.00 Hz), 6.61 (dd, 1H, Ar-H, *J* = 2.00, 9.00 Hz), 7.63 (d, 1H, Ar-H, *J* = 9.00 Hz), 9.37 (brs, 1H, NH); ¹³C NMR (DMSO-*d*₆, 125 MHz) δ (ppm): 21.0, 24.6, 33.9, 47.0, 66.3, 80.3, 102.2, 109.5, 114.6, 127.5, 160.8, 164.2, 166.0, 190.3; MS (EI, 70 eV): *m/z* (%) = 304 [M⁺, 41.2%], 263 [40.5%], 220 [51.5%], 183 [100%], 147 [61.2%], 114 [66.3%], 58 [70.2%]; LCMS: *m/z*: 305 [(M + H)⁺]; HRMS (ESI): *m/z* calcd for C₁₆H₂₁N₂O₄ [(M + H)⁺] 305.1496, found 305.1481; anal. calcd for C₁₆H₂₀N₂O₄ (304.14): C, 63.14; H, 6.62; N, 9.20%. Found: C, 63.21; H, 6.67; N, 9.18%.

5-Methyl-2-(2-((4-oxospiro[chromane-2,1'-cyclohexan]-7-yl)oxy)acetyl)-2,4-dihydro-3H-pyrazol-3-one (9). To a solution of **8** (2 mmol, 0.61 g) in absolute ethanol (25 mL) containing few

drops of glacial acetic acid, ethyl acetoacetate (1 eq, 2 mmol, 0.25 mL) was added. The reaction mixture was heated under reflux for 2 h, concentrated to 5 mL and purified using column chromatography (eluent: petroleum ether : ethyl acetate 3 : 1) to give **9** as colorless crystals. Yield (0.44 g, 59%); m.p. 288 °C; IR (KBr): ν /cm⁻¹ = 2858–2931 (CH_{Aliphatic}), 1689 (CO), 1665 (CO); ¹H NMR (DMSO-*d*₆, 500 MHz) δ (ppm): 1.41–1.81 (m, 10H, (CH₂)₅), 1.93 (s, 3H, CH₃), 2.64 (s, 2H, CH₂), 3.54 (s, 2H, CH₂), 4.52 (s, 2H, CH₂), 6.60 (d, 1H, Ar-H, *J* = 2.50 Hz), 6.72 (dd, 1H, Ar-H, *J* = 2.50, 8.50 Hz), 7.62 (d, 1H, Ar-H, *J* = 8.50 Hz); MS (EI, 70 eV): *m/z* (%) = 370 [M⁺, 23.4%], 352 [31.7%], 207 [40.8%], 161 [60.4%], 94 [80.7%], 59 [100.0%], 46 [56.6%]; anal. calcd for C₂₀H₂₂N₂O₅ (370.15): C, 64.85; H, 5.99; N, 7.56%. Found: C, 64.74; H, 6.04; N, 7.61%.

7-(2-(5-Hydroxy-3,5-dimethyl-4,5-dihydro-1H-pyrazol-1-yl)-2-oxoethoxy)spiro[chromane-2,1'-cyclohexan]-4-one (10). A mixture of **8** (2 mmol, 0.61 g) and acetylacetone (1 eq, 2 mmol, 0.2 mL) was refluxed in absolute ethanol (25 mL) containing few drops of glacial acetic acid as a catalyst for 2 h. After that, the reaction mixture was concentrated to 5 mL the formed product was collected and purified using TLC (eluent: petroleum ether : ethyl acetate 3 : 1) yielding **10** as colorless crystals. Yield (0.57 g, 74%); m.p. 222–223 °C; IR (KBr): ν /cm⁻¹ = 3446–3351 (OH), 2859–2947 (CH_{Aliphatic}), 1672 (CO), 1655 (CO); ¹H NMR (DMSO-*d*₆, 500 MHz) δ (ppm): 1.45–1.57 and 1.76–1.84 (m, 10H, (CH₂)₅), 1.73 (s, 3H, CH₃), 1.98 (s, 3H, CH₃), 2.65 (s, 2H, CH₂), 2.79 (d, 1H, CH, *J* = 18.50 Hz), 2.91 (d, 1H, CH, *J* = 18.50 Hz), 4.96 (s, 2H, CH₂), 6.38 (d, 1H, Ar-H, *J* = 2.50 Hz), 6.54 (dd, 1H, Ar-H, *J* = 3.00, 9.00 Hz), 7.61 (d, 1H, Ar-H, *J* = 9.00 Hz); MS (EI, 70 eV): *m/z* (%) = 386 [M⁺, 16.1%], 368 [58.6%], 292 [49.2%], 228 [28.5%], 167 [41.2%], 85 [100.0%], 45 [32.6%]; anal. calcd for C₂₁H₂₆N₂O₅ (386.18): C, 65.27; H, 6.78; N, 7.25%. Found: C, 65.37; H, 6.61; N, 7.32%.

N'-Benzylidene-2-((4-oxospiro[chromane-2,1'-cyclohexan]-7-yl)oxy)acetohydrazide (11). A mixture of **8** (2 mmol, 0.61 g) and benzaldehyde (1 eq, 2 mmol, 0.2 mL) in absolute ethanol (25 mL) containing catalytic drops of glacial acetic acid was heated under reflux for 2 h. The reaction mixture was concentrated to 5 mL. The separated product was collected by filtration, washed with petroleum ether and recrystallized from ethanol yielding **11** as colorless needles (mixture of geometrical isomers in a ratio of 1 : 2). Yield (0.62 g, 79%); m.p. 128–129 °C; IR (KBr): ν /cm⁻¹ = 3450 (NH), 2854–2928 (CH_{Aliphatic}), 1678 (CO), 1639 (CO); ¹H NMR (DMSO-*d*₆, 500 MHz) δ (ppm): 1.43–1.85 (m, 10H, (CH₂)₅, major and minor), 2.66 (s, 2H, CH₂, major isomer), 2.68 (s, 2H, CH₂, minor isomer), 4.75 (s, 2H, CH₂, minor isomer), 5.24 (s, 2H, CH₂, major isomer), 6.48 (d, 1H, Ar-H, *J* = 2.50 Hz, major isomer), 6.56 (d, 1H, Ar-H, *J* = 2.00 Hz, minor isomer), 6.60 (dd, 1H, Ar-H, *J* = 2.50, 8.50 Hz, major isomer), 6.66 (dd, 1H, Ar-H, *J* = 2.50, 9.00 Hz, minor isomer), 7.4–7.46 (m, 5H, Ar-H, *E* and *Z*), 7.63 (d, 1H, Ar-H, *J* = 9.00 Hz, major isomer), 7.66 (d, 1H, Ar-H, *J* = 7.00 Hz, minor isomer), 7.68–7.72 (m, 3H, Ar-H, major and minor), 8.00 (s, 1H, CH=N, major isomer), 8.30 (s, 1H, CH=N, minor isomer), 11.59 (brs, 1H, NH, minor isomer), 11.66 (brs, 1H, NH, major isomer); ¹³C NMR (CDCl₃, 125 MHz) δ (ppm): 21.4, 21.5, 25.06, 25.12, 34.7, 34.8, 47.7, 47.8, 65.3, 67.0, 80.5, 80.8, 102.4, 102.5, 109.0, 109.3, 115.3, 115.9, 127.3, 127.9,



128.3, 128.69, 128.72, 128.8, 130.7, 130.9, 133.0, 133.1, 145.8, 149.8, 161.4, 161.6, 163.0, 163.5, 164.7, 169.6, 191.2, 191.3; MS (EI, 70 eV): m/z (%) = 392 [M^+ , 58.2%], 361 [85.3%], 356 [100.0%], 328 [83.6%], 183 [62.8%], 134 [44.8%]; LCMS: m/z : 393 [$(M + H)^+$]; HRMS (ESI): m/z calcd for $C_{23}H_{25}N_2O_4$ [$(M + H)^+$] 393.1809, found 393.1796; anal. calcd for $C_{23}H_{24}N_2O_4$ (392.17): C, 70.39; H, 6.16; N, 7.14%. Found: C, 70.28; H, 6.22; N, 7.24%.

***N'*-(2-Oxoindolin-3-ylidene)-2-((4-oxospiro[chromane-2,1'-cyclohexan]-7-yl)oxy)acetohydrazide (13).** A mixture of **8** (3 mmol, 0.91 g) and isatin (1 eq, 3 mmol, 0.44 g) in absolute ethanol (30 mL) containing a catalytic amount of glacial acetic acid was heated under reflux for 2 h. The product obtained on heating was collected by filtration and washed several times with boiling ethanol to give **13** as a yellow powder. Yield (1.05 g, 81%); m.p. 278–279 °C; IR (KBr): ν/cm^{-1} = 3540–3162 (NH, NH, OH), 2857–2930 ($CH_{Aliphatic}$), 1694 (CO); 1H NMR (DMSO- d_6 , 500 MHz) δ (ppm): 1.26–1.86 (m, 10H, $(CH_2)_5$), 2.69 (s, 2H, CH_2), 4.95 (s, 1H, $OCH=C(OH)NH$), 5.40 (brs, 1H, OH or NH), 6.64–6.69 (m, 2H, Ar-H), 6.94 (d, 1H, Ar-H, J = 7.50 Hz), 7.08–7.11 (m, 1H, Ar-H), 7.37–7.40 (m, 1H, Ar-H), 7.56–7.68 (m, 2H, Ar-H), 11.27 (brs, 1H, OH or NH), 13.66 (brs, 1H, OH or NH); ^{13}C NMR (DMSO- d_6 , 125 MHz) δ (ppm): 21.0, 24.6, 33.9, 47.0, 67.0, 80.3, 102.6, 109.3, 111.2, 115.0, 119.6, 121.0, 122.7, 127.8, 132.0, 138.7, 142.6, 160.8, 162.5, 190.3; MS (EI, 70 eV): m/z (%) = 433 [M^+ , 17.7], 369 [24.7%], 330 [53.7%], 300 [59.3%], 264 [66.5%], 219 [79.3%], 133 [72.3%], 83 [52.6], 42 [100.0%]; LCMS: m/z : 434 [$(M + H)^+$]; HRMS (ESI): m/z calcd for $C_{24}H_{24}N_3O_5$ [$(M + H)^+$] 434.1711, found 434.1691; anal. calcd for $C_{24}H_{23}N_3O_5$ (433.16): C, 66.50; H, 5.35; N, 9.69%. Found: C, 66.61; H, 5.23; N, 9.78%.

1-(7-Methoxyspiro[chromane-2,1'-cyclohexan]-4-ylidene)-2-phenylhydrazine (15). To a solution of **14** (2 mmol, 0.49 g) in ethyl alcohol (20 mL) containing catalytic drops of acetic acid, phenylhydrazine (1 eq, 2 mmol, 0.19 mL) was added, the solution was refluxed for 4 h then left overnight at 0 °C. The formed crystalline product was collected by filtration to give **15** as pale brown crystals. Yield (0.38 g, 56%); m.p. 175 °C; IR (KBr): ν/cm^{-1} = 3347 (NH), 2854–2930 ($CH_{Aliphatic}$), 1600 ($C=N$); 1H NMR (DMSO- d_6 , 400 MHz) δ (ppm): 1.46–1.81 (m, 10H, $(CH_2)_5$), 2.74 (s, 2H, CH_2), 3.75 (s, 3H, OCH_3), 6.42 (d, 1H, Ar-H, J = 2.00 Hz), 6.55 (dd, 1H, Ar-H, J = 2.40, 8.40 Hz), 6.73 (m, 1H, Ar-H), 7.2–7.21 (m, 4H, Ar), 7.86 (d, 1H, Ar-H, J = 8.80 Hz), 9.25 (brs, 1H, NH); MS (EI, 70 eV): m/z (%) = 336 [M^+ , 11.3], 334 [36.8%], 300 [75.1%], 260 [54.5%], 217 [51.6%], 155 [100.0%], 153 [64.1%], 119 [53.2%], 112 [39.7%], 99 [68.2%], 64 [56.6%]; anal. calcd for $C_{21}H_{24}N_2O_2$ (336.18): C, 74.97; H, 7.19; N, 8.33%. Found: C, 74.85; H, 7.29; N, 8.22%.

1,2-Bis(7-methoxyspiro[chromane-2,1'-cyclohexan]-4-ylidene)hydrazine (16). To a solution of **14** (2 mmol, 0.49 g) containing a catalytic amount of acetic acid in ethanol (20 mL), hydrazine monohydrate (1 eq, 2 mmol, 0.1 mL) was added. The reaction mixture was refluxed for 2 h, concentrated to 10 mL and left overnight in an open flask. The formed product was collected and recrystallized from ethanol to give **16** as yellow needles. Yield (0.73 g, 75%); m.p. 175–176 °C; IR (KBr): ν/cm^{-1} = 2856–2935 ($CH_{Aliphatic}$), 1596 ($C=N$); 1H NMR (DMSO- d_6 , 500 MHz) δ (ppm): 1.41–1.78 (m, 20H, two $(CH_2)_5$), 2.95 (s, 2H, 2 CH_2), 3.76 (s, 3H, 2 OCH_3), 6.45 (d, 2H, Ar-H, J = 2.00 Hz), 6.56

(dd, 2H, Ar-H, J = 3.00, 9.00 Hz), 7.93 (d, 2H, Ar-H, J = 8.50 Hz); ^{13}C NMR ($CDCl_3$, 125 MHz) δ (ppm): 21.7, 25.5, 35.0, 35.9, 55.4, 77.4, 101.5, 108.6, 113.8, 126.2, 155.2, 157.1, 162.9; MS (EI, 70 eV): m/z (%) = 488 [M^+ , 31.6%], 478 [66.8%], 430 [71.4%], 396 [48.2%], 332 [100.0%], 250 [62.4%], 202 [59.8%], 158 [62.7%]; LCMS: m/z : 489 [$(M + H)^+$]; HRMS (ESI): m/z calcd for $C_{30}H_{36}N_2O_4$ [$(M + H)^+$] 489.2748, found 489.2759; anal. calcd for $C_{30}H_{36}N_2O_4$ (488.27): C, 73.74; H, 7.43; N, 5.73%. Found: C, 73.59; H, 7.60; N, 5.84%.

***N'*-(7-Methoxyspiro[chromane-2,1'-cyclohexan]-4-ylidene)-2-((4-oxospiro[chromane-2,1'-cyclohexan]-7-yl)oxy)acetohydrazide (17).** To a solution of **14** (1 mmol, 0.24 g) and **8** (1 eq, 1 mmol, 0.3 g) in ethanol (30 mL) a catalytic amount of acetic acid was added, the reaction mixture was refluxed for 2 h and left to cool to room temperature. The formed crystalline product was collected and recrystallized from DMF-ethanol mixture to give **17** as yellow needles. Yield (0.42 g, 79%); m.p. 280–281 °C; IR (KBr): ν/cm^{-1} = 3357 (NH), 2853–2947 ($CH_{Aliphatic}$), 1686 (CO), 1659 (CO), 1600 ($C=N$); (DMSO- d_6 , 400 MHz) δ (ppm): 1.21–1.84 (m, 20H, two $(CH_2)_5$), 2.84 (s, 2H, CH_2), 2.96 (s, 2H, CH_2), 3.81 (s, 3H, OCH_3), 4.71 (s, 2H, CH_2), 6.48 (d, 1H, Ar-H, J = 2.00 Hz), 6.65–7.62 (m, 4H, Ar-H), 7.92 (d, 1H, Ar-H, J = 8.50 Hz), 13.34 (brs, 1H, NH); MS (EI, 70 eV): m/z (%) = 532 [M^+ , 9.5%], 483 [34.7%], 393 [69.3%], 350 [27.6%], 347 [100.0%], 302 [34.5%], 267 [60.1%], 222 [29.3%], 124 [21.2%]; anal. calcd for $C_{31}H_{36}N_2O_6$ (532.26): C, 69.91; H, 6.81; N, 5.26%. Found: C, 70.12; H, 6.89; N, 4.99%.

2-Cyano-*N'*-(7-methoxyspiro[chromane-2,1'-cyclohexan]-4-ylidene)acetohydrazide (18). A solution of **14** (5 mmol, 1.23 g) and cyanoacetic acid hydrazide (1 eq, 5 mmol, 0.49 g) in ethanol (20 mL) containing acetic acid (0.5 mL) was heated under reflux for 5 h and then left to cool. The produced crystals were collected and recrystallized from absolute ethanol to give pure **18** as colorless needles (mixture of geometrical isomers in a ratio of 1 : 3). Yield (1.39 g, 85%); m.p. 256–257 °C; IR (KBr): ν/cm^{-1} = 3192 (NH), 2855–2927 ($CH_{Aliphatic}$), 2260 ($C\equiv N$) 1675 (CO); 1H NMR (DMSO- d_6 , 500 MHz) δ (ppm): 1.35–1.77 (m, 10H, $(CH_2)_5$, major and minor), 2.70 (s, 2H, CH_2 , minor isomer), 2.76 (s, 2H, CH_2 , major isomer), 3.73 (s, 3H, OCH_3 , major isomer), 3.74 (s, 3H, OCH_3 , minor isomer), 3.83 (s, 2H, CH_2 , minor isomer), 4.21 (s, 2H, CH_2 , major isomer), 6.40 (d, 1H, Ar-H, J = 2.50 Hz, major isomer), 6.42 (d, 1H, Ar-H, J = 2.50 Hz, minor isomer), 6.52 (dd, 1H, Ar-H, J = 2.00, 8.50 Hz, major isomer), 6.55 (dd, 1H, Ar-H, J = 2.50, 9.00 Hz, minor isomer), 7.77 (d, 1H, Ar-H, J = 8 Hz, major isomer), 7.81 (d, 1H, Ar-H, J = 9 Hz, minor isomer), 10.65 (brs, 1H, NH, major isomer), 11.01 (brs, 1H, NH, minor isomer); ^{13}C NMR (DMSO- d_6 , 125 MHz): 21.08, 21.15, 24.7, 24.8, 24.9, 33.9, 34.3, 34.4, 35.0, 40.0, 55.3, 76.5, 76.7, 101.4, 101.6, 108.6, 112.4, 116.1, 116.3, 125.5, 125.6, 143.3, 147.7, 155.9, 156.0, 159.0, 162.1, 162.4, 165.5; MS (EI, 70 eV): m/z (%) = 327 [M^+ , 17.1%], 304 [46.4%], 295 [95.8%], 221 [40.9%], 220 [100.0%], 180 [30.7%], 151 [20.7%], 101 [26.1%], 78 [43.6]; LCMS: m/z : 328 [$(M + H)^+$]; HRMS (ESI): m/z calcd for $C_{18}H_{22}N_3O_3$ [$(M + H)^+$] 328.1656, found 328.1644; anal. calcd for $C_{18}H_{21}N_3O_3$ (327.16): C, 66.04; H, 6.47; N, 12.84%. Found: C, 66.21; H, 6.31; N, 13.01%.



***N'*-(7-Methoxyspiro[chromane-2,1'-cyclohexan]-4-ylidene)-2-oxo-2H-chromene-3-carbohydrazide (19).** A mixture of **18** (2 mmol, 0.65 g) and salicylaldehyde (1 eq, 2 mmol, 0.21 mL) in ethanol (30 mL) containing few drops of piperidine was refluxed for 3 h, the produced product was filtered off while hot, washed with boiling ethanol to remove unreacted starting materials, dried and recrystallized from a mixture of dimethyl formamide/acetone to furnish **19** as yellow crystals. Yield (0.77 g, 90%); m.p. 212–213 °C; IR (KBr): ν/cm^{-1} = 3425 (NH), 2872–2935 ($\text{CH}_{\text{Aliphatic}}$), 1718 (CO), 1670 (CO); ^1H NMR (DMSO- d_6 , 500 MHz) δ (ppm): 1.41–1.60 (m, 10H, $(\text{CH}_2)_5$), 2.72 (s, 2H, CH_2), 3.76 (s, 3H, OCH_3), 6.45 (d, 1H, Ar-H, J = 2.50 Hz), 6.59 (dd, 1H, Ar-H, J = 2.50, 9.00 Hz), 6.84–6.92 (m, 2H, Ar-H), 7.82–7.88 (m, 3H, Ar-H), 8.57 (s, 1H, pyrone), 13.49 (brs, 1H, NH); MS (EI, 70 eV): m/z (%) = 432 [M^+ , 38%], 415 [91.6%], 402 [62.3%], 383 [60.1%], 368 [79.9%], 348 [60.4%], 259 [71.3%], 192 [93.7%], 172 [56.6%], 100 [64.8%], 61 [100.0%]; anal. calcd for $\text{C}_{25}\text{H}_{24}\text{N}_2\text{O}_5$ (432.17): C, 69.43; H, 5.59; N, 6.48%. Found: C, 69.38; H, 5.63; N, 6.53%.

2-Cyano-*N'*-(7-methoxyspiro[chromane-2,1'-cyclohexan]-4-ylidene)-3-phenylacrylohydrazide (20). To a stirred solution of **18** (4 mmol, 1.3 g) in 20 mL dimethyl formamide, 2 mL (10%) KOH was added. The stirring was continued for 30 min at room temperature till a deep yellow solution is formed, benzaldehyde (1 eq, 4 mmol 0.4 mL) was added and the reaction mixture was kept stirred at room temperature for further 5 h, then poured into crushed ice and neutralized using diluted HCl [0.1 N]. The yellow product formed was filtered off and recrystallized from dilute ethanol to give **20** as yellow sheets (mixture of geometrical isomers in a ratio of 1 : 1). Yield (1.42 g, 85%); m.p. 162–163 °C; IR (KBr): ν/cm^{-1} = 3435 (NH), 2855–2928 ($\text{CH}_{\text{Aliphatic}}$), 2213 (CN), 1657 (CO), 1602 ($\text{C}=\text{N}$); ^1H NMR (DMSO- d_6 , 500 MHz) δ (ppm): 1.46–1.80 (m, 10H, $(\text{CH}_2)_5$, *E* and *Z*), 2.78 (s, 2H, CH_2 , *E* or *Z*), 2.85 (s, 2H, CH_2 , *E* or *Z*), 3.73 (s, 3H, OCH_3 , *E* or *Z*), 3.76 (s, 3H, OCH_3 , *E* or *Z*), 6.38–6.58 (m, 2H, Ar-H, *E* and *Z*), 7.59–7.61 (m, 3H, Ar-H, *E* and *Z*), 7.83–8.02 (m, 3H, Ar-H, *E* and *Z*), 8.18 (s, 1H, CH = CCO, *E* and *Z*), 10.88 (brs, 1H, NH, *E* or *Z*), 11.28 (brs, 1H, NH, *E* or *Z*); MS (EI, 70 eV): m/z (%) = 415 [M^+ , 19.5%], 387 [100.0%], 322 [53.2%], 309 [58.4%], 282 [57.7%], 210 [45.5%], 119 [42.6%], 69 [52.8%]; anal. calcd for $\text{C}_{25}\text{H}_{25}\text{N}_3\text{O}_3$ (415.19): C, 72.27; H, 6.07; N, 10.11%. Found: C, 72.18; H, 6.15; N, 10.13%.

3-Amino-*N'*-(7-methoxyspiro[chromane-2,1'-cyclohexan]-4-ylidene)-5-phenyl-1H-pyrazole-4-carbohydrazide (21). To a stirred solution of **20** (2 mmol, 0.83 g) in ethanol (25 mL) at room temperature, hydrazine monohydrate (1 eq, 2 mmol, 0.1 mL) was added. The reaction mixture was stirred for additional 3 h at 80 °C. The formed product on stirring was filtered off and recrystallized from ethanol to give **21** as pale-yellow crystals. Yield (0.73 g, 82%); m.p. 194–195 °C; IR (KBr): ν/cm^{-1} = 3511 (NH or NH_2), 3425 (NH or NH_2), 3337 (NH or NH_2), 3208 (NH or NH_2), 2861–2939 ($\text{CH}_{\text{Aliphatic}}$), 1674 (CO), 1604 ($\text{C}=\text{N}$); ^1H NMR (DMSO- d_6 , 500 MHz) δ (ppm): 1.23–1.78 (m, 10H, $(\text{CH}_2)_5$), 2.69 (s, 2H, CH_2), 3.78 (s, 3H, OCH_3), 6.03 (brs, 2H, NH_2), 6.26–6.32 (m, 2H, Ar-H), 6.54–6.59 (m, 1H, Ar-H), 7.49–7.78 (m, 5H, Ar-H), 11.07 (brs, 1H, NH), 11.72 (brs, 1H, NH); MS (EI, 70 eV): m/z (%) = 445 [M^+ , 26.4%], 330 [78.7%], 235 [26.2%], 133 [39.9%], 90

[62.1%], 77 [100.0%], 63 [86.1%], 50 [38.5%]; anal. calcd for $\text{C}_{25}\text{H}_{27}\text{N}_5\text{O}_3$ (445.21): C, 67.40; H, 6.11; N, 15.72%. Found: C, 67.48; H, 6.18; N, 15.64%.

Antimicrobial evaluation

The antimicrobial activity of the newly synthesized compounds was evaluated against *Staphylococcus aureus* (ATCC® 6535), *Escherichia coli* (ATCC® 8739), and *Candida albicans* (ATCC® 10231). In addition, methicillin-resistant *Staphylococcus aureus* (MRSA) was local isolates from the patients of Mansoura University Hospitals, Egypt. The samples were dissolved in dimethyl sulfoxide (DMSO) and solutions of 1 mg mL^{-1} concentrations were prepared separately. Whatman filter paper discs with a size of 5 mm were cut and then sterilized in the autoclave. After that, the discs were soaked in solutions of the synthesized compounds. To Petri dishes containing agar media [agar (20 g) + beef extract (3 g) + peptone (5 g)] seeded with *Staphylococcus aureus*, *Escherichia coli*, and *Candida albicans*, the previously soaked discs were placed. After incubation of the Petri dishes at 36 °C for 24 h, the inhibition done diameters were measured in millimeter (mm). For each compound, the zone diameter was calculated as the average of three trials. The antimicrobial activities of ciprofloxacin and clotrimazole was assessed as reference drugs under the same conditions. Control experiments using DMSO under the same conditions were run parallel to samples of the synthesized compounds.³⁹ The activity indices were calculated using the following equation:

$$\% \text{ Activity index} = \frac{\text{Diameter of the inhibition zone of the tested compound}}{\text{Diameter of the inhibition zone of the reference drug}} \times 100$$

The minimal inhibitory concentrations (MICs) for compounds with inhibitions zones of ≥ 10 mm diameters were determined at concentration of $64 \mu\text{g mL}^{-1}$. The two-fold dilutions of the solutions were done (64, 32, 16, ..., $0.5 \mu\text{g mL}^{-1}$). The suspensions of different microbes at concentrations of 10^6 (CFU mL^{-1} , colony forming unit per mL) were added to 96-well microliter plate followed by incubation at 36 °C. After incubation for 24 h, MICs were determined as the lowest concentrations that inhibited visible growth of the microorganisms (turbidity).^{40,41}

Evaluation of cytotoxicity⁴²

The cytotoxicity was tested against hepatocellular carcinoma (HePG-2) cell line (ATCC® via VACSERA company for biological products, Cairo, Egypt). The reagents 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT), RPMI-1640 medium, and dimethyl sulfoxide (DMSO) were purchased from Sigma Chemical Company of St. Louis, USA. Fetal bovine serum was purchased from GIBCO, UK. MTT assay depends upon the conversion of a yellow 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) into a purple formazan by the mitochondrial succinate dehydrogenase in viable cells. HePG-2 cell line was cultured in RPMI-1640 medium containing



10% fetal bovine serum and 100 units per mL 1% antibiotic penicillin/streptomycin at 37 °C in a 5% carbon dioxide incubator. After seeding HePG-2 cell line in a 96-well plate (density: 1.0×10^4 cells per well) at 37 °C FOR 48 h under 5% carbon dioxide, the cells were treated with different concentration of the synthesized compounds 2–21 and followed by an incubation period for 24 h. After that, 20 μ L of pre-prepared MTT soln (5 mg mL⁻¹) was added onto each well followed by a second incubation for 4 h. In order to dissolve the generated purple formazan, 100 μ L DMSO was added to each well. The spectrophotometric absorbance for each well was measured at 570 nm using microplate reader (ELx800, USA). All tests were carried out three time. The percentage of cell viability was calculated as $[A_{570\text{nm}} \text{ of treated sample} / A_{570\text{nm}} \text{ of untreated sample} \times 100]$. After plotting the relationship between viable cell percentage and compound concentration, the survival curve for HePG-2 was obtained. The IC₅₀ value which is the required concentration to inhibit a 50% of cell growth was calculated by the nonlinear regression analysis by using Graph pad prism software and shown as mean \pm standard deviation (SD).

Computational evaluation

Prediction of biological activities and physicochemical parameters. The 2-dimensional chemical structures of synthesized compounds were drawn by Marvin Sketch (ChemAxon), and the SMILEYS, (*.sdf), (*.mol), and (*.pdb) mode of structures were saved for further evaluations. PASS online software (<http://www.pharmaexpert.ru/passonline/predict.php>) was used to predict the biological activities for the SMILEYS structures of the synthesized compounds.²³ Moreover, pharmacokinetics and drug-likeness were predicted using SwissADME (<http://www.swissadme.ch/index.php>) and AdmetSAR-2.0 (<http://lmmd.ecust.edu.cn/admetSar2>).²⁴

Molecular docking

To predict the antimicrobial mechanisms and the most suitable targets for the synthesized compounds, two antibacterial and two antifungal targets related to antimicrobial activities were screened. The crystal structures *S. aureus* penicillin-binding protein (PDB ID: 1VQQ), *E. coli* DNA gyrase (PDB ID: 5MMN), *C. albicans* lanosterol 14- α demethylase (PDB ID: 5TZ1), and *exo*-beta-1,3-glucanase (PDB ID: 3N9K) were selected and fetched from Protein Data Bank (www.rcsb.org/pdb). AutoDock 4.2 molecular dockings were performed using PYRX software to predict the binding pose with active sites of the protein targets.⁴³ Water molecules were removed, and hydrogen atoms were added before docking to obtain the correct ionization and tautomeric states for the amino acids. Binding energies of protein–ligand interactions were recorded, and molecular docking was visualized using PYMOL software (version 2.2.3).

Quantitative structure–activity relationship (QSAR)

The dataset containing synthesized compounds was used for making QSAR models. The 2D-structures of compounds were made with MavinSketch and converted into optimized MDL*.mol using same tool. The properties were calculated

PaDEL-Descriptor (29).³⁷ QSARINS v2.0 was used for creating QSAR models with genetic algorithm-variable subset selection (GA-VSS) method (30).³⁸ The dataset was randomly divided into training and test sets (25% of the whole dataset). External validation was carried out in accordance with the reported criteria.

Conflicts of interest

The authors declare no competing financial interests.

Acknowledgements

We would like to deeply thank Prof. Dr Yehia Osman and Dr Sally Elsheikh from the Department of Botany, Faculty of Science, Mansura University, Egypt for their assistance to evaluate the antimicrobial activity of our compounds against Methicillin-resistant *Staphylococcus aureus* (MRSA).

References

- 1 G. P. Ellis, *Chromenes, chromanones and chromones*, John Wiley & Sons, New York, 1977.
- 2 W. Fischer, A. Currais, Z. Liang, A. Pinto and P. Maher, *Redox Biol.*, 2019, **21**, 101089.
- 3 Y. Sukpondma, V. Rukachaisirikul and S. Phongpaichit, *Chem. Pharm. Bull.*, 2005, **53**, 850–852.
- 4 G. Valdomir, S. Senthilkumar, D. Ganapathy, Y. Zhang and L. F. Tietze, *Chem. - Eur. J.*, 2018, **24**, 8760–8763.
- 5 Y. Gao, Q. Ren, H. Wu, M. Li and J. Wang, *Chem. Commun.*, 2010, **46**, 9232–9234.
- 6 Z. Yang, K. Wu, Y. Xu, X. Xia, X. Wang, M. Ge and L. Shao, *J. Antibiot.*, 2020, **73**, 194–199.
- 7 F. Cottiglia, B. Dhanapal, O. Sticher and J. Heilmann, *J. Nat. Prod.*, 2004, **67**, 537–541.
- 8 M. Ceylan-Ünlüsoy, E. J. Verspohl and R. Ertan, *J. Enzyme Inhib. Med. Chem.*, 2010, **25**, 784–789.
- 9 S. Emami, T. Banipoulad, H. Irannejad, A. Foroumadi, M. Falahati, M. Ashrafi-Khozani and S. Sharifynia, *J. Enzyme Inhib. Med. Chem.*, 2014, **29**, 263–271.
- 10 L. Du, M. Li, Q. Yang, Y. Tang, Q. You and L. Xia, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 1477–1480.
- 11 S. I. El-Desoky, F. A. Badria, M. A. Abozeid, E. M. Kandeel and A. H. Abdel-Rahman, *Med. Chem. Res.*, 2013, **22**, 2105–2114.
- 12 (a) H. Chen, J. Xie, D. Xing, J. Wang, J. Tang, Z. Yi, F. Xia, W.-W. Qiu and F. Yang, *Org. Biomol. Chem.*, 2019, **17**, 1062–1066; (b) L. Feng, M. M. Maddox, Md. Z. Alam, L. S. Tsutsumi, G. Narula, D. F. Bruhn, X. Wu, S. Sandhaus, R. B. Lee, C. J. Simmons, Y.-C. Tse-Dinh, J. G. Hurdle, R. E. Lee and D. Sun, *J. Med. Chem.*, 2014, **57**, 8398–8420; (c) N. G. Ghatpande, J. S. Jadhav, R. V. Kaproorath, M. E. Soliman and M. M. Shaikh, *Bioorg. Med. Chem.*, 2020, **28**, 115813.
- 13 R. Rios, *Chem. Soc. Rev.*, 2012, **41**, 1060–1074.
- 14 (a) K. Ding, Z. Han and Z. Wang, *Chem.-Asian J.*, 2009, **4**, 32–41; (b) F. Xu, D. Huang, C. Han, W. Shen, X. Lin and Y. Wang, *J. Org. Chem.*, 2010, **75**, 8677–8680; (c) Y.-H. Yuan, X. Han,

- F.-P. Zhu, J.-M. Tian, F.-M. Zhang, X.-M. Zhang, Y.-Q. Tu, S.-H. Wang and X. Guo, *Nat. Commun.*, 2019, **10**, 3394.
- 15 T. P. I. Saragi, T. Spehr, A. Siebert, T. Fuhrmann-Lieker and J. Salbeck, *Chem. Rev.*, 2007, **107**, 1011–1065.
- 16 E. Chupakhin, O. Babich, A. Prosekov, L. Asyakina and M. Krasavin, *Molecules*, 2019, **24**, 4165.
- 17 (a) Y. Zheng, C. M. Tice and S. B. Singh, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 3673–3682; (b) Y.-J. Zheng and C. M. Tice, *Expet Opin. Drug Discov.*, 2016, **11**, 831–834; (c) P. Saraswat, G. Jeyabalan, M. Z. Hassan, M. U. Rahman and N. K. Nyola, *Synth. Commun.*, 2016, **46**, 1643–1664; (d) A. Ding, M. Meazza, H. Guo, J. W. Yang and R. Rios, *Chem. Soc. Rev.*, 2018, **47**, 5946–5996.
- 18 L. Guan, H. Yang, Y. Cai, L. Sun, P. Di, W. Li, G. Liu and Y. Tang, *RSC Med. Chem.*, 2019, **10**, 148–157.
- 19 (a) G. Mehta, R. Samineni, P. Srihari, R. G. Reddy and S. Chakravarty, *Org. Biomol. Chem.*, 2012, **10**, 6830–6833; (b) N. Abe, R. Onoda, K. Shirahata, T. Kato, M. C. Woods and Y. Kitahara, *Tetrahedron Lett.*, 1968, **9**, 369–373; (c) A. K. Ghosh, K. Krishnan, D. E. Walters, W. Cho, Y. Koo, J. Trevino, L. Holland and J. Buthod, *Bioorg. Med. Chem. Lett.*, 1998, **8**, 979–982.
- 20 (a) F. Micoli, F. Bagnoli, R. Rappuoli and D. Serruto, *Nat. Rev. Microbiol.*, 2021, **19**, 287–302; (b) M. Gajdacs, E. Urbán, A. Stájer and Z. Baráth, *Eur. J. Investig. Health Psychol. Educ.*, 2021, **11**, 71–82; (c) M. Lomazzi, M. Moore, Ap. Johnson, M. Balasegaram and B. Borisch, *BMC Publ. Health*, 2019, **19**, 858; (d) S. I. El-Desoky, E. M. Keshk, A. A. El-Sawi, M. A. Abozeid, L. A. Abouzeid and A. H. Abdel-Rahman, *Saudi Pharmaceut. J.*, 2018, **26**, 852–859; (e) M. A. Abozeid, M. R. El-Kholany, A. H. Abdel-Rahman and S. I. El-Desoky, *Int. J. Mod. Org. Chem.*, 2018, **5**, 1–11; (f) M. A. Abozeid, M. R. El-Kholany, L. A. Abouzeid, A. H. Abdel-Rahman and S. I. El-Desoky, *J. Heterocycl. Chem.*, 2019, **56**, 2922–2933; (g) S. I. El-Desoky, A. A. El-Sawi, M. A. Abozeid, M. Abdelmoteleb, M. Shaaban, E. M. Keshk and A. H. Abdel-Rahman, *Med. Chem. Res.*, 2019, **28**, 1601–1617; (h) M. A. Abozeid, A. A. El-Sawi, M. Abdelmoteleb, H. Awad, M. M. Abdel-Aziz, A. H. Abdel-Rahman and S. I. El-Desoky, *RSC Adv.*, 2020, **10**, 42998–43009; (i) M. A. Abozeid, A. A. El-Sawi, M. R. Elmorsy, M. Abdelmoteleb, A. H. Abdel-Rahman and S. I. El-Desoky, *RSC Adv.*, 2019, **9**, 27996–28005.
- 21 M. Mujahid, R. G. Gonnade, P. Yogeeswari, D. Sriram and M. Muthukrishnan, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 1416–1419.
- 22 S. Y. Dike, J. R. Merchant and N. Y. Spare, *Tetrahedron*, 1991, **47**, 4775–4786.
- 23 S. Parasuraman, *J. Pharmacol. Pharmacother.*, 2011, **2**, 52–53.
- 24 (a) A. Daina, O. Michielin and V. Zoete, *Sci. Rep.*, 2017, **7**, 42717; (b) H. Yang, C. Lou, L. Sun, J. Li, Y. Cai, Z. Wang and Y. Tang, *Bioinformatics*, 2018, **35**, 1067–1069.
- 25 R. D. Pearson, R. T. Steigbigel, H. T. Davis and S. W. Chapman, *Antimicrob. Agents Chemother.*, 1980, **18**, 699–708.
- 26 R. J. Holt, *J. Clin. Pathol.*, 1975, **28**, 767–774.
- 27 J. M. Andrews, *J. Antimicrob. Chemother.*, 2001, **48**, 5–16.
- 28 V. Judge, B. Narasimhan, M. Ahuja, D. Sriram, P. Yogeeswari, E. De Clercq, C. Pannecouque and J. Balzarini, *Med. Chem. Res.*, 2012, **21**, 1451–1470.
- 29 H. F. Rizk, M. A. El-Badawi, S. A. Ibrahim and M. A. El-Borai, *Chin. J. Chem.*, 2011, **29**, 1451–1459.
- 30 H. K. Gençer, U. A. Çevik, S. Levent, B. N. Sağlık, B. Korkut, Y. Özkay, S. İlgin and Y. Öztürk, *Molecules*, 2017, **22**, 507.
- 31 D. B. Kitchen, H. Decornez, J. R. Furr and J. Bajorath, *Nat. Rev. Drug Discovery*, 2004, **3**, 935–949.
- 32 M. Mohamadzadeh, M. Zarei and M. Vessal, *Bioorg. Chem.*, 2020, **95**, 103515.
- 33 B. Fois, Ž. Skok, T. Tomašič, J. Ilaš, N. Zidar, A. Zega, L. P. Mašič, P. Szili, G. Draskovits, Á. Nyerges, C. Pál and D. Kikelj, *ChemMedChem*, 2020, **15**, 265–269.
- 34 E. R. Trösken, M. Adamska, M. Arand, J. A. Zarn, C. Patten, W. Völkel and W. K. Lutz, *Toxicology*, 2006, **228**, 24–32.
- 35 K. Y. Lum, S. T. Tay, C. F. Le, V. S. Lee, N. H. Sabri, R. D. Velayuthan, H. Hassan and S. D. Sekaran, *Sci. Rep.*, 2015, **5**, 9657.
- 36 (a) E. N. Muratov, J. Bajorath, R. P. Sheridan, I. V. Tetko, D. Filimonov, V. Poroikov, T. I. Oprea, I. I. Baskin, A. Varnek, A. Roitberg, O. Isayev, S. Curtalolo, D. Fourches, Y. Cohen, A. Aspuru-Guzik, D. A. Winkler, D. Agrafiotis, A. Cherkasov and A. Tropsha, *Chem. Soc. Rev.*, 2020, **49**, 3525–3564; (b) Danishuddin and A. U. Khan, *Drug Discov. Today*, 2016, **21**, 1291–1302.
- 37 C. W. Yap, *J. Comput. Chem.*, 2011, **32**, 1466–1474.
- 38 P. Gramatica, N. Chirico, E. Papa, S. Cassani and S. Kovarich, *J. Comput. Chem.*, 2013, **34**, 2121–2132.
- 39 J. B. Patel, M. P. Weinstein, G. M. Eliopoulos, S. G. Jenkins, J. S. Lewis II, B. Limbago, A. J. Mathers, T. Mazzulli, R. Patel, S. S. Richter, M. Satlin, J. M. Swenson, M. M. Traczewski, J. D. Turnidge, B. L. Zimmer, *Performance standards for antimicrobial susceptibility testing, CLSI supplement M100*, Clinical and Laboratory Standards Institute, Wayne, PA, 27th edn, 2017.
- 40 J. H. Rex, B. D. Alexander, D. Andes, B. Arthington-Skaggs, S. D. Brown, V. Chaturvedi, M. A. Ghannoum, A. Espinel-Ingroff, C. C. Knapp, L. Ostrosky-Zeichner, M. A. Pfaller, D. J. Sheehan, T. J. Walsh, *Reference method for Broth dilution antifungal susceptibility testing of yeasts; approved standard-third edition. CLSI document M27-A3*, Clinical and Laboratory Standards Institute, 2008.
- 41 J. H. Rex, B. D. Alexander, D. Andes, B. Arthington-Skaggs, S. D. Brown, V. Chaturvedi, A. Espinel-Ingroff, M. A. Ghannoum, C. C. Knapp, M. R. Motyl, L. Ostrosky-Zeichner, M. Pfaller, D. J. Sheehan, T. J. Walsh, *Reference method for Broth dilution antifungal susceptibility testing of filamentous fungi; approved standard-second edition. CLSI document M38-A2*, Clinical and Laboratory Standards Institute, 2008.
- 42 (a) T. Mosmann, *J. Immunol. Methods*, 1983, **65**, 55–63; (b) F. Denizot and R. Lang, *J. Immunol. Methods*, 1986, **22**, 271–277.
- 43 O. Trott and A. J. Olson, *J. Comput. Chem.*, 2010, **31**, 455–461.

