


Cite this: *RSC Adv.*, 2021, 11, 20541

Methane production from wheat straw pretreated with CaO₂/cellulase

Zhenmin Li,^a Jishi Zhang,^{ID} *^a Chunduo Kong,^a Wenqing Li,^a Jinwei Wang^b and Lihua Zang^a

There are various lignocellulosic biomass pretreatments that act as attractive strategies to improve anaerobic digestion for methane (CH₄) generation. This study proposes an effective technique to obtain more CH₄ via the hydrothermal coupled calcium peroxide (CaO₂) co-cellulase pretreatment of lignocellulosic biomass. The total organic carbon in the hydrolysate of samples treated with 6% CaO₂ and 15 mg enzyme per g-cellulose was 7330 mg L⁻¹, which represented an increase of 92.39% over the total organic carbon value of samples hydrolyzed with the enzyme alone. The promotion of the anaerobic digestion of wheat straw followed this order: hydrothermal coupled CaO₂ co-cellulase pretreatment > hydrothermal coupled CaO₂ pretreatment > enzymatic pretreatment alone > control group. The sample treated with 6% CaO₂ and 15 mg enzyme per g-cellulose gave the highest CH₄ production with a CH₄ yield of 214 mL g⁻¹ total solids, which represented an increase of 64.81% compared to the control group. The CH₄ yield decreased slightly when the amount of added cellulase exceeded 15 mg enzyme per g-cellulose.

Received 27th March 2021

Accepted 25th May 2021

DOI: 10.1039/d1ra02437j

rsc.li/rsc-advances

Introduction

With the increasing depletion of fossil fuels, there has been a gradual increase in environmental pollution problems caused by modern development. As the world's population grows, the demand for resources is increasing; biomass-based wastes have been studied worldwide in recent decades because they are a potential biomass resource.¹ Wastes usually have the advantages of high abundance, low cost and carbon neutrality.² However, direct incineration is the main technology used to treat biomass, and this treatment method not only generates large amounts of hazardous dust and gases that are harmful to human health and cause air pollution, but also has the potential to cause fires.³ Lignocellulose biomass is an agricultural biomass that provides an important source of energy for the sustainable production of biofuels.⁴ In the area of lignocellulose biomass, wheat straw is an attractive feedstock for large-scale biological production plants due to its low cost, high sugar content and abundance.⁵ Every year, a large amount of wheat straw is produced worldwide, and the annual output of wheat straw in China is about 80 million tons.⁶ Wheat straw is considered to be one of the most promising renewable raw materials for the sustainable production of biomethane (bioCH₄) and biohydrogen (bioH₂) from biomass.^{7,8}

CH₄ gas has a high calorific value, is easy to store, and the only products of its combustion are carbon dioxide and water, which do not cause haze because they do not contain ash. It also has the advantages of high efficiency, cleanliness, economy and safety, and is expected to become one of the most important future resources utilized for clean energy in China. Anaerobic digestion (AD) from waste biomass is an important method to produce CH₄ because the process is cost-effective, efficient and reliable.⁹ Generally speaking, the AD process can be divided into the following four stages: (a) hydrolysis, (b) fermentation (or acidification), (c) acetic acid generation, and (d) CH₄ production.¹⁰ In this process, hydrolysis is considered to be the rate-limiting step.¹¹ Wheat straw is the part of wheat that remains after harvesting the seeds, and its main components include cellulose, hemicellulose, and lignin (about 85% to 95% of the total mass);¹² these three polymers are embedded and cross-linked, and are often called lignocellulose. In addition, wheat straw contains small amounts of crude protein, pectin, fat, ash and moisture, which are linked with each other through various chemical bonds or binding forces.^{13,14} Therefore, wheat straw should be pretreated to destroy or remove the lignin to reduce its wrapping effect on cellulose and hemicellulose, to break the dense structure. This can reduce the degree of polymerization of the cellulose and increase its accessibility of enzymes to achieve effective utilization of the cellulose. This is also a key step to convert lignocellulose biomass into biofuel.^{10,15}

There are a variety of current straw pretreatment technologies, which are categorized as physicochemical, thermal, thermochemical and biological pretreatments, in addition to

^aCollege of Environmental Science and Engineering, Qilu University of Technology (Shandong Academy of Science), Jinan 250353, China. E-mail: lyzhangjishi@163.com; Fax: +86 531 89631680; Tel: +86 531 89631680

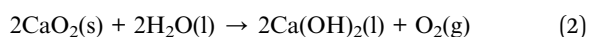
^bWeifang Ensign Industry Co., Ltd, Weifang 250353, China



combinations of these.^{15–18} However, all single pretreatment methods have some disadvantages that result in important additional costs to the process; these costs are often the main obstacle to the commercial pretreatment of lignocellulose waste. Thus, two or more pretreatment methods can be combined based on the characteristics of the different pretreatment techniques to give full play to their respective advantages in order to achieve cost savings, reduce the amount of chemicals used in the pretreatment process, lower the degree of equipment wear, improve pollution problems, and significantly increase the efficiency of straw hydrolysis and develop a low-energy, environmentally friendly and efficient pretreatment technology. Higher temperature can achieve a better pretreatment effect, but overly high temperatures (more than 150 °C) result in the conversion of lignin to phenolic compounds, which are toxic to the microbes in AD systems.¹⁹ Recent reports have revealed that the hydrolysis phase of AD can be accelerated at lower oxygen levels.^{20,21} The reason for this is that micro-aerobic conditions stimulate the secretion of extracellular hydrolytic enzymes such as cellulases and proteases,²² which ultimately improve the enzymatic efficiency of the enzymatic products.

Many investigations of hydrolysis using bases such as sodium hydroxide (NaOH) or calcium hydroxide (Ca(OH)₂) have been carried out by scholars worldwide.^{23,24} The main advantage of alkali pretreatment is the breakdown of lignin, the presence of which greatly limits the role of enzymes in the AD process.^{25,26} Alkali conditions are efficient not only for breaking the ester bonds between polysaccharides and lignin,²⁷ but also for dissolving some of the hemicellulose and cellulose.²⁸ Calcium peroxide (CaO₂) is one of the safest and most versatile solid inorganic peroxide compounds and is considered to be the solid form of H₂O₂.²⁹ When CaO₂ is dissolved in an aqueous solution, it can react with water and decompose at a controlled rate to slowly produce oxygen (O₂), calcium hydroxide (Ca(OH)₂) and hydrogen peroxide (H₂O₂), as shown in eqn (1) and (2).^{30–32} Due to these properties, CaO₂ can be used as an alternative additive to supply oxygen to the AD process and create micro-oxygen demand conditions to facilitate the hydrolysis of the substrate. CaO₂ can be used as a highly effective additive for pollutant removal due to the reactive oxygen species in it.^{30,33} CaO₂ can enhance the hydrolytic acidification of activated sludge by increasing the activity of hydrolases and acid-producing enzymes.³¹ In addition, the reaction of CaO₂ with water produces Ca(OH)₂, which fixes CO₂ (Ca(OH)₂(l) + CO₂(g) → CaCO₃(s) + H₂O(l)), thus increasing the CH₄ content of the biogas. At the same time, the alkalinity of the AD system can be increased by the generation of Ca(OH)₂ or CaCO₃, which facilitates the production of volatile fatty acids (VFAs) and soluble chemical oxygen demand (SCOD).³⁴ However, the OH[−] produced by CaO₂ has a significant negative effect on CH₄-producing bacteria.³⁵ Therefore, when using CaO₂ as an additive, the amount of CaO₂ must be controlled to improve the efficiency of AD. The hydrolysis temperature of wheat straw is related to the biomass pretreatment performance.³⁶ Based on this pretreatment, further enzymatic hydrolysis is considered in this work. Cellulase is a specific and highly efficient biotype catalyst for the hydrolysis of cellulose. It is a collective term for a number of enzymes that degrade the fibrous raw material dextran, and the different component enzymes act

synergistically with each other.³⁷ The purposes of this study of the pretreatment of wheat straw with CaO₂ and cellulase were to (1) investigate the synergistic effects of CaO₂, cellulase, and CaO₂ reacted with cellulose; (2) evaluate the physicochemical properties of the pretreated straw hydrolysate and solid residues; (3) reveal the effects of the pretreatment conditions on the CH₄ production performance of AD from lignocellulose materials; and (4) estimate the biodegradability and CH₄ production potential of each substrate using the modified Gompertz kinetic model.



Materials and methods

Feedstock and inoculum

The AD substrate used in this study was wheat straw from an agricultural processing plant in Shandong, China. Wheat straw must be pretreated prior to AD for CH₄ production. After natural drying at 30–35 °C, the straw was chopped into small pieces of 2.5–3.5 cm, further homogenized using a plant disintegrator, crushed into straw powder of about 40 mesh (0.45 mm), and then placed in sample bags and sealed for later use. Its main elemental components were as follows: total N 0.45 ± 0.05%, total C 46.4 ± 0.4% and H 6.2 ± 0.02%.

Inoculated sludge was obtained from the laboratory of Qilu University of Technology (Shandong Academy of Science). The inoculated sludge was cultured with glucose (0.3 g L^{−1}) for 20 d at 36 ± 1 °C until no gas was produced and then further concentrated by gravity separation, from which the AD inoculum was obtained. In addition, the cellulase used in this work was purchased from Novozymes (Denmark). The cellulase activity was 200 000 U mL^{−1}, and its optimum temperature and pH value were 55–60 °C and 4.5–5.0, respectively. Moreover, the cellulase contained some metal ions such as Na⁺, Cu²⁺, Ca²⁺ and Zn²⁺, further promoting its activity. The main properties of the wheat straw and inoculum are shown in Table 1.

Hydrothermal and enzymatic co-treatment for wheat straw

5 g straw samples were weighed out into seven 500 mL bioreactors numbered 1–7. To bioreactors no. 1–6, 6% CaO₂ by mass

Table 1 The characteristics of the wheat straw and inoculum

Characteristic	Wheat straw	Inoculum
pH	6.69	7.2
Moisture content (%)	7.63	90.06
Total solids (TS, %)	92.37	9.94
Volatile solids (VS, % of TS)	95.15	95.93
C (% of TS)/TOC (mg L ^{−1})	43.53	1950
N (% of TS)/NH ₄ ⁺ -N (mg L ^{−1})	0.231	136.1
H (% of TS)	6.182	—
C/N ratio	188.4	—



was added, and bioreactor no. 7 was the control group without CaO₂. The deionized water used in this study was obtained by two-stage reverse osmosis, and was added to all the bioreactors to give a solid to liquid ratio (m straw/v deionized water) of 1 : 12. After the samples were thoroughly wetted, all the bioreactors with straw samples were placed in a 35 °C incubator at 150 rpm for 2 h. Bioreactors no. 1–6 were then taken out and placed into the reaction kettle, kept at a constant temperature at 121 °C for 60 min, and then removed and subsequently cooled to room temperature (20–25 °C) to obtain a mixture of hydrolytic liquids and solid residue. The pretreated samples of bioreactors no. 1–5 and 7 were adjusted to pH 4.5–5.0 with 1 mol L^{−1} HCl and placed in a constant temperature incubator set at 55 °C with shaking at 120 rpm. When the temperature was reached, the treatment was carried out by adding the appropriate amount of acidic cellulase, which was 0.1 mL, 0.2 mL, 0.3 mL, 0.4 mL, and 0.5 mL for reactors no. 1–5, and 0.3 mL for no. 7. The pretreated samples were further digested/hydrolyzed for 72 hours and then cooled to room temperature.³⁸

AD design for methane production

Eight 500 mL serum bottles were numbered R1–R8 and the pretreated wheat straw mixtures numbered 1–7 were transferred into the serum bottles numbered R1–R7, respectively. The R6 group was pretreated with CaO₂ alone, the R7 group was pretreated with cellulase alone, and in the R8 group, the wheat straw was directly fermented without additives to produce CH₄ as a control experiment. Then, 150 mL of inoculated sludge was added to each bottle and distilled water was added to make up the volume to 400 mL. The top of each bioreactor was flushed with N₂ for 1 min to remove O₂ before the serum bottle was quickly sealed with a rubber stopper. Finally, all the bioreactors were placed in an electric-heated thermostatic water bath and digested at 36 ± 1 °C for 42 days. The above experiments were conducted in three parallel experiments. During the AD processes, each bioreactor was manually mixed by gently shaking it once a day and gas production data were recorded once a day. The extracted fermentation broth from each reactor was centrifuged at 5000 rpm for 5 minutes using a medical centrifuge, and the liquid parameters, such as the pH, volatile fatty acids (VFAs), total organic carbon (TOC), total carbon (TC), inorganic carbon (IC) and ammonium nitrogen (NH₄⁺-N), were measured in the supernatant. The CH₄ volume was converted to standard temperature and pressure (STP) through correction of the water vapor pressure at 25 °C, which was based on eqn (3).³⁹

$$V_{\text{gas}} (\text{mL, STP}) = V_{\text{gas}} (\text{mL, } T) \times \frac{273}{273 + T} \times \frac{101\,325 - W}{101\,325} \quad (3)$$

where $T = 25$ °C (room temperature), $W = 3167$ Pa (water vapour pressure at 25 °C), and V_{gas} = volume of gas at STP (mL).

Analysis methods

Total solid (TS) and volatile solid (VS) were determined based on a previous work.⁴⁰ Additionally, the ammonia nitrogen (NH₄⁺-N), TOC and pH in the liquid samples were measured according

to a previous study.⁴¹ The total carbon (C) and nitrogen (N) of wheat straw were also determined according to the method previously described by Zhang *et al.*⁴¹

In addition, the topography of the straw samples was measured using a scanning electronic microscope (SEM) (JEOL JSM-5600 LV, Japan) according to the method previously reported by Zhang *et al.*⁴¹ The changes in the functional groups in the wheat straw samples before and after treatment with CaO₂ were evaluated using a Fourier transform infrared (FTIR) spectrophotometer (IR Prestige-21, Japan). The lignin, cellulose, and hemicellulose contents of the samples before and after treatment were measured based on a previous study.⁴¹ Additionally, an X-ray diffractometer (D8 Advance, Bruker AXS GmbH, Germany) was employed to obtain the X-ray diffractometry (XRD) curves of the cellulase- and CaO₂-treated wheat straw samples.⁴¹ Moreover, the crystallinity index (CrI) of each sample was estimated based on the previous report.⁴¹ The bioCH₄ yield was determined by the saturated NaOH drainage technique, and the CH₄ content was measured using gas chromatography (GC-2014C/TCD, Shimadzu, Japan) with a thermal conductivity detector (TCD). The biogas volumes were further determined under STP conditions (273 K, 101.325 kPa).

Kinetic analysis of methane production

The CH₄ production potential (MPP) was estimated by fitting a cumulative CH₄ production (CMP) curve with the modified Gompertz model, which is described in eqn (4).⁴² The software Origin 9.0 was used to determine the P_m , R_m and λ values and evaluate the correlation coefficient (R^2).

$$P(t) = P_m \times \exp \left\{ -\exp \left[\frac{R_m \times e}{P_m} (\lambda - t) + 1 \right] \right\} \quad (4)$$

where $P(t)$ is the CMP (mL) at time t (d), P_m is the MPP (mL), R_m is the maximum methane production rate (mL d^{−1}), e is 2.72, λ is the lag period (d), and t is the fermentation time (d).

Results and discussion

Effects of CaO₂/cellulase pretreatment on the hydrolysis products

The effect of hydrothermally coupled CaO₂-coordinated cellulase pretreatment on the characteristics of the straw hydrolysis products was mainly evaluated in terms of the TOC of the pretreated hydrolysis products and the changes in the content of the three substances in the hydrolyzed straw residue.⁴¹ As shown in Tables 2 and 3, the addition of cellulase to the hydrothermally coupled CaO₂ pretreatment technique for wheat straw increased the TOC of the hydrolysis products in all cases (Table 2). The TOC content of the hydrolysate from the control group was 1997 mg L^{−1}. As the amount of cellulase used was increased from the 5 mg per g-cellulose to 25 mg per g-cellulose group, the TOC of the hydrolysis products increased, which indicated that the hydrolysis effect was enhanced by increasing the amount of cellulase added. However, the TOC value did not show a linear increase with the amount of cellulase added. As the amount of added cellulase was increased from 5 mg per g-



Table 2 The indicators of straw hydrolysate after enzymatic pretreatment

Sample	Enzyme (mg per g-cellulose)	pH	NH ₄ ⁺ -N (mg L ⁻¹)	TOC (mg L ⁻¹)
1	6% CaO ₂ + 5 enzyme	5.8 ± 0.2	36 ± 2.0	5180 ± 15
2	6% CaO ₂ + 10 enzyme	5.7 ± 0.2	41 ± 2.5	6250 ± 15
3	6% CaO ₂ + 15 enzyme	5.7 ± 0.2	79 ± 2.5	7330 ± 20
4	6% CaO ₂ + 20 enzyme	5.6 ± 0.1	117 ± 3.0	7450 ± 15
5	6% CaO ₂ + 25 enzyme	5.4 ± 0.2	110 ± 2.5	7590 ± 35
6	6% CaO ₂ + 0 enzyme	8.9 ± 0.2	16 ± 2.5	4865 ± 15
7	0% CaO ₂ + 15 enzyme	4.8 ± 0.1	67 ± 2.5	3810 ± 15
8	Control group	6.7 ± 0.1	86.12 ± 2.0	1997 ± 15

Table 3 Lignin, cellulose, and hemicellulose content levels before and after pretreatment

Sample	Cellulose (%)	Hemicellulose (%)	Lignin (%)
Wheat straw	38.8 ± 0.1	27.9 ± 0.1	20.1 ± 0.1
CaO ₂ /cellulase	30.2 ± 0.1	25.6 ± 0.3	18.6 ± 0.2
CaO ₂	45.1 ± 0.3	26.1 ± 0.3	18.8 ± 0.1
Cellulase	36.8 ± 0.2	27.3 ± 0.3	20.0 ± 0.2

cellulose to 15 mg per g-cellulose, the TOC value of the pretreated hydrolysis products increased significantly and rapidly. However, with further increasing the amount of cellulase, the increase in the TOC values became less pronounced, which was probably due to the inhibition of cellulase when the concentration of reducing sugars in solution was too high. The results showed that the samples without hydrothermally coupled calcium oxide pretreatment were the least hydrolysed, with an improvement of 90.79% over the control group. Due to the complex physical and chemical structure of wheat straw, it was difficult for cellulase to reach the cellulose surface directly for enzymatic hydrolysis. The TOC of the sample pretreated only with hydrothermally coupled CaO₂ was 4865 mg L⁻¹, which was 27.45% and 143.62% higher than those of the enzymatic-treatment and control groups, respectively. The TOC value of the hydrolysis product of the samples treated with 6% CaO₂ plus 15 mg enzyme per g-cellulose was 7330 mg L⁻¹, which was 92.39% and 267.05% higher than those from the enzymatic hydrolysis and control groups, respectively. The TOC from the sample treated with 6% CaO₂ and 25 mg enzyme per g-cellulose was 7590 mg L⁻¹, showing a 99.21% increase in the TOC value compared to enzymatic hydrolysis alone. In order to determine whether the effects of different pretreatment conditions on the hydrolysis process of straw were statistically significant, a one-way analysis of variance (ANOVA) and a *post hoc* test were carried out on the TOC values of the hydrolysis products of each group of samples using the analysis software SPSS, and a *p*-value of 0.00 (less than 0.05) was obtained, which indicated that different pre-treatment conditions had a significant effect on the TOCs of the hydrolysate samples. The co-pretreatment using 6% CaO₂ and 15 mg enzyme per g-cellulose was optimal based on the combined effect of pre-hydrolysis and cost considerations.

On the other hand, due to the different pretreatment conditions, the contents of cellulose, hemicellulose and lignin varied considerably after pretreatment hydrolysis. The most significant changes were observed in the samples pretreated with CaO₂ co-cellulase; details are described in Table 3. The weight loss of the wheat straw was due to the dissolution of components such as cellulose, hemicellulose and lignin.^{42,43} The cellulose, hemicellulose and lignin contents of the raw wheat straw samples were 38.8%, 27.9% and 20.1%, respectively. The levels of the three substances in the samples subjected to direct enzymatic hydrolysis and in the samples pretreated with CaO₂ did not change significantly, whereas the heat and CaO₂ co-pretreatment was more effective in degrading hemicellulose to obtain soluble oligomers and monomeric sugars, and also in removing lignin, which was consistent with previous work.⁴¹ Consequently, the hemicellulose and lignin contents decreased, and the relative cellulose content increased. The contents of cellulose, hemicellulose and lignin in the sample under the conditions of 6% CaO₂ and 25 mg enzyme per g-cellulose were 30.2%, 25.6% and 18.6%, respectively (Table 3). Additionally, hemicellulose and lignin were removed; the complex structure of the wheat straw was disrupted. Consequently, the straw sample was hydrated and swollen, exhibiting a loose and soft structure, which facilitated the accessibility of the cellulose and increased the adsorption sites for cellulase. The findings were in line with a previous report.⁴¹

Effects of CaO₂-coordinated cellulase treatment on the wheat straw structure

The physical and chemical properties of the straw under different pretreatment conditions were analyzed and compared with those of the untreated wheat straw samples to investigate the differences in the surface structure, functional groups and crystal structure of the wheat straw before and after CaO₂-coordinated cellulase treatment. The morphologies of the wheat straw samples under different pretreatment conditions were observed under a scanning electronic microscope (SEM). As shown in Fig. 1, the surface of unprepared wheat straw is smooth, dense, rigid and structurally intact (Fig. 1a); these properties do not facilitate access to the cellulose interior by cellulase during subsequent AD.³² After the pretreatment, the waxy surface coating of the wheat straw is removed, the surface



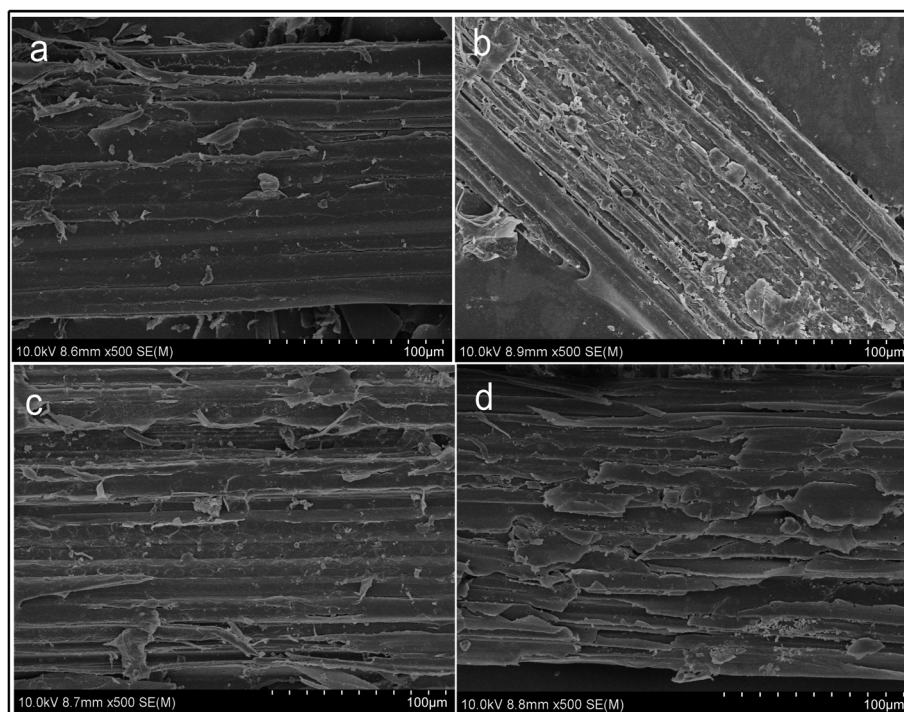


Fig. 1 The morphologies of wheat straw under different pretreatment conditions: control group (a), CaO_2 plus enzyme treatment (b), CaO_2 treatment (c), and enzyme treatment (d).

roughness is significantly increased and the surface structure becomes porous and loose. Due to the alkalinity and strong oxidation of CaO_2 , part of the lignin was destroyed and part of the hemicellulose and cellulose was dissolved, allowing the surface structure of wheat straw to begin to separate and be converted into cellulose filaments (Fig. 1c), which was similar to the findings from Kang *et al.*⁴² and Solé-Bundó *et al.*⁴⁴ The enzymatic hydrolysis of samples pretreated with enzyme alone resulted in a less damaged surface structure than for those pre-treated with hydrothermally coupled CaO_2 . This is mainly due to the complex structure of the straw, which prevents the effective utilization of cellulase due to its low accessibility (Fig. 1d). The SEM images of

samples that were first pretreated with hydrothermally coupled CaO_2 and then further hydrolyzed using cellulase showed that the surface structure of the wheat straw was greatly damaged under these conditions. Their morphologies revealed that the pre-treatment process allowed the wheat straw to hydrate and swell, and that the liquid penetrated into the wheat straw cells, effectively destroying the complex linkages between the various straw components. The combination treatment with CaO_2 and cellulase was the most damaging to the surface structure of the wheat straw, followed by the hydrothermal coupled CaO_2 pretreatment, and finally the cellulase hydrolysis alone.

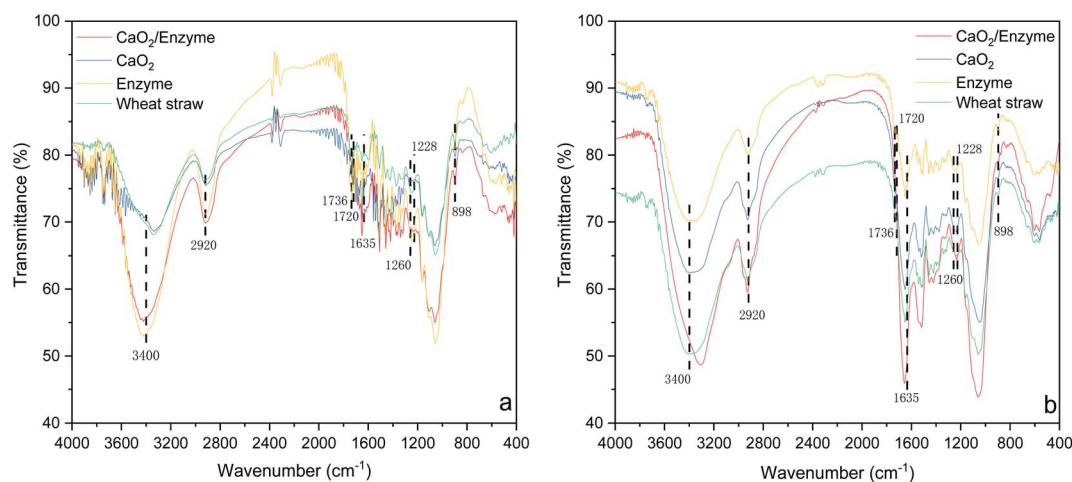


Fig. 2 Changes in the surface functional groups before (a) and after (b) anaerobic digestion.

The FTIR spectra of the raw straw samples, the CaO_2 -coordinated cellulase-pretreated samples, the CaO_2 -treated samples and the cellulase-treated samples are described in Fig. 2. The peaks of methyl ($-\text{CH}_3$) or methylene ($-\text{CH}_2$) groups occurred at 2920 cm^{-1} and the absorption peak at 3400 cm^{-1} was related to alcohol hydroxyl and phenolic hydroxyl ($-\text{OH}$) groups. In the pretreated samples, the absorption of this peak was weaker than in the original straw sample, indicating that the hydroxyl interactions decreased after the pretreatment. The hydrogen bonding was mainly present in the crystalline zone of cellulose, which indirectly indicated that the crystalline zone of cellulose was somewhat damaged and the degree of crystallinity decreased. The spectral bands at 1260 cm^{-1} (C–O bond, related to guaiac-based lignin) and 1635 cm^{-1} (related to the C=O vibration of the conjugate carbonyl group of lignin) were weakened by the removal or degradation of lignin.^{45,46} The degradation of hemicellulose was indicated by a decrease in the peak intensities at 1228 cm^{-1} and 1720 cm^{-1} , which correspond to the C–O and ester/carboxylic acid bonds of the hemicellulose acetyl unit. In addition, the peak intensity at 898 cm^{-1} associated with the stretching vibration of the cellulose cyclic β -1,4-glycosidic bond was reduced and the absorption peak of the cellulose C–H bending vibration signature in the sample after CaO_2 co-cellulase pretreatment was significantly lower than that of the raw material, which demonstrated the removal of cellulose from the samples after this co-pretreatment and further confirmed the degradation of amorphous cellulose.⁴⁷ Similarly, the peak at 1736 cm^{-1} was related to the C=O stretching vibration, which reflected the variation in hemicellulose content. After the pretreatment process, the peaks were weakened, suggesting a decrease in hemicellulose content. In addition, the decrease in peak intensity at 1603 and 1511 cm^{-1} indicated the degradation of lignin.⁴⁸ Co-cellulase pretreatment with CaO_2 was more likely to give the maximum utilization of cellulose than heat and CaO_2 co-treatment, resulting in improved straw biodegradability.

In addition, the XRD characterization of the wheat straw raw material and the samples treated under various treatment conditions is shown in Fig. 3, and their crystallinity is also

Table 4 Crystallinity properties for different CaO_2 and enzyme pretreatment conditions

Sample	Crystallinity (%)	Grain size (nm)	Strain
Wheat straw	33.12	48.2 ± 2.5	0.678 ± 0.020
CaO_2 /enzyme	27.37	7.5 ± 1.5	0.72 ± 0.018
CaO_2	54.55	20.3 ± 1.4	0.196 ± 0.012
Enzyme	31.41	28.5 ± 1.31	0.636 ± 0.101

presented in Table 4. The XRD profiles of the pre-treated samples and wheat straw were similar (Fig. 3). The presence of diffraction peaks indicated a relatively high degree of crystallinity in the structure of the fibers. After the alkali-treatment process, the cellulose chain segments were folded to form a new crystal structure.⁴⁹ Natural cellulose consists of type I cellulose crystals, which undergo a structural change from type I to type II cellulose after alkali treatment.⁵⁰ As revealed in Fig. 3, the XRD spectra of both the raw wheat straw and the pre-treated wheat straw samples had a clear and strong band at $2\theta = 22.6^\circ$. There were two weak diffraction bands at $2\theta = 14.7^\circ$ and 16.4° , which indicated cellulose I.⁵¹ The XRD pattern of wheat straw before pretreatment had two strong diffraction bands at $2\theta = 16^\circ$ and 22° , but the intensities of the diffraction peaks were reduced by the pretreatment (Fig. 3). The crystallinity of the wheat straw was 33.12%. In the wheat straw sample treated with 6% CaO_2 , the band at $2\theta = 29^\circ$ shifted to a slightly higher angle and appeared as an intense and sharp diffraction peak. The half-width of the diffraction peak (002) became smaller and its intensity increased, and the crystallinity was 54.55%. The alkaline oxygen pretreatment under hydrothermal conditions effectively removed lignin and degraded part of the hemicellulose and amorphous cellulose, causing enrichment of the cellulose. Under the optimal conditions (6% CaO_2 + 25 mg cellulase per g-cellulose), the crystallinity of the pretreated sample was 27.37%. SEM and FTIR spectroscopy of the samples demonstrated that the surface morphology and functional groups changed considerably after pretreatment with the different methods, which could be supported and corroborated by changes in the content of the three substances in the wheat straw and the composition of the hydrolysis product.

Effects of CaO_2 -coordinated cellulase treatment on methane production

The CH_4 production and yield are two important indexes to evaluate the AD process of wheat straw. Fig. 4 illustrates the relationship between the CMP and fermentation time under different pretreatment conditions, including the relationship between the CMP and AD time under different cellulase conditions (Fig. 4a), and methane production under the optimal conditions with different pretreatments compared to the control group (Fig. 4b). As demonstrated in Fig. 4a, the CMP curves coincided with the lag, log, stability and decay periods of the microbial growth curve. The possible mechanisms of the CaO_2 -coordinated cellulase treatment of wheat straw are shown in Fig. 5.⁵² The addition of cellulase increases the adhesion of

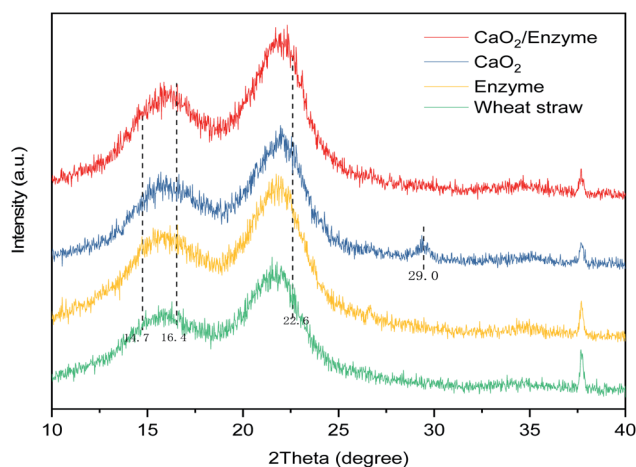


Fig. 3 Changes in the crystallinity of the wheat straw before and after pretreatment.



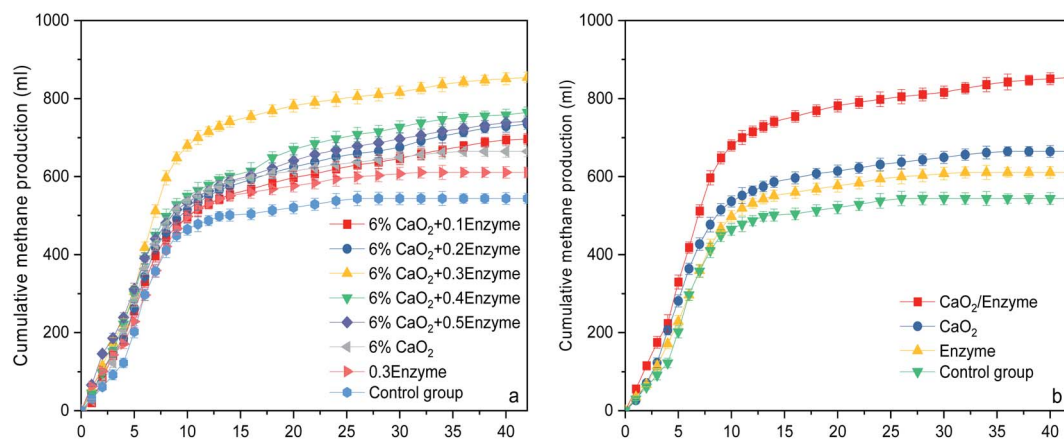


Fig. 4 A comparison of CH_4 production from the CaO_2 /cellulase co-treatment and control groups: (a) CMP under different enzyme conditions; and (b) CMP under the optimal conditions.

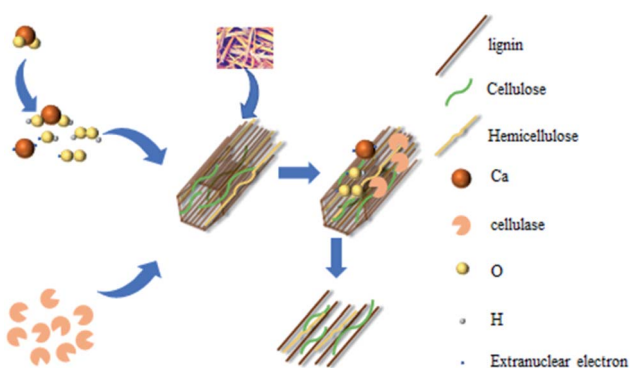


Fig. 5 The possible mechanism involved in the CaO_2 /cellulase treatment of wheat straw (6% CaO_2 coupled with 0.1–0.5 mL of enzyme). Adapted with permission.⁵²

cellulose to the enzyme, so that the TOC and $\text{NH}_4^+\text{-N}$ contents of the hydrolysis product increase and the hydrolysis effect is enhanced. Bolado-Rodríguez *et al.*⁵³ observed that alkali pretreatment promoted the hydrolysis process of wheat straw. Additionally, the addition of CaO_2 released Ca^{2+} , which is a trace element required by methanogens, into the liquid during the dissolution process. Additionally, cellulase addition further promoted the decomposition of hemicellulose, lignin and cellulose into small molecules that were readily attacked and used by different microorganisms. The hydrolytic and fermentative microbes with high anti-oxidative enzyme activity

were capable of scavenging reactive oxygen species and protecting the methanogens. These properties all favored the growth and multiplication of methanogenic bacteria, which in turn increased the CH_4 yield. In the AD system, when the cellulase addition was increased from 0 to 0.3 mL with the addition of 6% CaO_2 pretreatment, the CMP increased progressively compared to that of the control group without pretreatment, from 22.29% to 57.06%. These results of the ANOVA showed a significant difference ($p < 0.0001$) between the total gas productions of straw treated with different amounts of cellulase, indicating that the enzymatic hydrolysis of wheat straw with various amounts of cellulase has a positive effect on the CH_4 production from the wheat straw in the presence of CaO_2 pretreatment. However, when the amount of cellulase added was continually increased, the CH_4 yield revealed a decreasing trend; the addition of 0.4 and 0.5 mL cellulase lowered the CH_4 yield by 10.40% and 13.10% compared to the highest yield, respectively. This was because excess cellulase inhibited the activity of the methanogenic bacteria, thereby lowering the CH_4 production. In the group treated using enzymatic hydrolysis alone, the CH_4 yield was only 12.33% higher than that of the control group (Table 5). After alkali treatment, the lignin was dissolved in the alkali solution, which was a potential toxicant to the microbes in AD system. Thus, a moderate amount of cellulase helped the anaerobes to produce more CH_4 from wheat straw, while excess cellulase inhibited CH_4 production.

Table 5 The kinetic parameters of synergistic CaO_2 /cellulase treated wheat straw in methane production^a

Reagent dosage	P_m (mL)	R_m (mL d ⁻¹)	λ (d)	R^2 (%)	CMP (mL)	CH_4 yield (mL g ⁻¹)	Increase in CMP (%)
6% CaO_2 /0.3 enzyme	817.92	86.32	0.29	99.23	853.75	170.75	57.06
6% CaO_2	641.54	72.62	1.15	99.22	664.72	132.94	22.29
0.3 enzyme	600.74	64.92	1.32	99.69	610.58	122.12	12.33
Control group	535.92	70.04	1.85	99.37	543.58	108.72	—

^a “—” represents the difference between the cumulative methane production of the control group and itself (the difference is 0).



Kinetic analysis of methane production

The length of the delay in the AD process depends on the concentration of substances that are easily digestible by microorganisms in the fermentation broth at the beginning of the reaction.⁵⁴ The lag times of the pretreated samples were short due to the pretreatment, whereas the lag time of the control group without any pretreatment was long. It can be seen from Table 5 that the effect of cellulase addition on CH₄ generation can be well fitted with the modified Gompertz model (eqn (4)), and that the correlation coefficient (R^2) was greater than 98%, indicating that the fitting curve was significantly related to the experimental data. CaO₂-coordinated cellulase treatment had significant effects on the P_m and R_m values. With the 6% CaO₂ pretreatment and the addition of 0.3 mL of cellulase, the P_m value was 817.92 mL, which was 57.06% higher than that of the control group, and the maximum R_m value (86.59 mL d⁻¹) was obtained under these conditions. The CH₄ yield was proportional to the amount of cellulase when the amount of cellulase was below 0.3 mL. Further increasing the amount of cellulase caused a decrease in the R_m values. Therefore, the actual CH₄ production is the result of a combination of factors such as solution pH, lignin, cellulose, AD time and temperature, as well as the inoculum. This further demonstrated that there was a difference between the theoretical and actual CH₄ production potential.

Conclusions

Calcium peroxide (CaO₂) and its combination with cellulase in the pretreatment of wheat straw for bioCH₄ production were investigated in terms of the main characteristics of the wheat straw before and after the pretreatments and the subsequent bioCH₄ generation. The results showed that the promotion of the anaerobic digestion of wheat straw followed the order hydrothermal and CaO₂ co-cellulase pretreatment > hydrothermal and CaO₂ pretreatment > cellulase hydrolysis alone > control group. The samples treated with 6% CaO₂ and 15 mg cellulase per g-cellulose produced the highest CH₄ yields, with a 64.81% increase compared to the control group. Pretreatment with a moderate amount of cellulase also promoted the hydrolysis of wheat straw and the subsequent anaerobic digestion process, while excess cellulase could limit CH₄ production.

Author contributions

Zhenmin Li: data curation, writing – original draft. Jishi Zhang: supervision, conceptualization, writing – review and editing, funding acquisition. Chunduo Kong: methodology, data curation. Wenqing Li: writing – review and editing. Jinwei Wang: methodology. Lihua Zang: funding acquisition, supervision.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This study was financially supported by the Natural Science Foundation of Shandong Province (ZR2016EEM33).

References

- 1 C. Akobi, H. Yeo, H. Hafez and G. Nakhla, *Appl. Energy*, 2016, **184**, 548–559.
- 2 Y. Wang, L. Dai, R. Wang, L. Fan, Y. Liu, Q. Xie and R. Ruan, *J. Anal. Appl. Pyrolysis*, 2016, **119**, 251–258.
- 3 A. Kumar Sakhiya, A. Anand, I. Aier, P. Baghel, V. K. Vijay and P. Kaushal, *Mater. Today: Proc.*, 2020, DOI: 10.1016/j.matpr.2020.08.795.
- 4 P. Ganguly, S. Sengupta, P. Das and A. Bhowal, *Fuel*, 2020, **280**, 118581.
- 5 P. M. Bondesson and M. Galbe, *Biotechnol. Biofuels*, 2016, **222**, 9159.
- 6 C. Huang, X. Wu, Y. Huang, C. Lai, X. Li and Q. Yong, *Bioresour. Technol.*, 2016, **219**, 583–588.
- 7 G. Kumar, P. Sivagurunathan, B. Sen, S. H. Kim and C. Y. Lin, *Bioresour. Technol.*, 2017, **240**, 137–143.
- 8 F. Talebnia, D. Karakashev and I. Angelidaki, *Bioresour. Technol.*, 2010, **101**, 4744–4753.
- 9 C. Huang, X. Wu, Y. Huang, C. Lai, X. Li and Q. Yong, *Bioresour. Technol.*, 2016, **219**, 583–588.
- 10 E. Rouches, I. Herpoël-Gimbert, J. P. Steyer and H. Carrere, *Renewable Sustainable Energy Rev.*, 2016, **59**, 179–198.
- 11 M. Hassan, W. Ding, J. Bi, E. Mehryar, Z. A. Talha and H. Huang, *Bioresour. Technol.*, 2016, **200**, 405–412.
- 12 S. Nadav, T. H. Yeats, S. Heidi, Y. Heather and S. Chris, *BioScience*, 2014, **64**(3), 192–201.
- 13 N. Sorek, T. H. Yeats, H. Szemenyei, H. Youngs and C. R. Somerville, *BioScience*, 2014, **64**, 192–201.
- 14 K. A. Underkofler, R. E. Teixeira, S. A. Pietsch, K. G. Knapp and R. T. Raines, *ACS Sustainable Chem. Eng.*, 2015, **3**, 606–613.
- 15 L. Janke, S. Weinrich, A. F. Leite, F. K. Terzariol, M. Nikolausz, M. Nelles and W. Stinner, *Energy Convers. Manage.*, 2017, **141**, 378–389.
- 16 E. Abdelsalam, M. Samer, Y. A. Attia, M. A. Abdel-Hadi, H. E. Hassan and Y. Badr, *Energy Convers. Manage.*, 2017, **141**, 108–119.
- 17 A. Cesaro and V. Belgiorno, *Chem. Eng. J.*, 2014, **240**, 24–37.
- 18 Y. Zheng, J. Zhao, F. Xu and Y. Li, *Prog. Energy Combust. Sci.*, 2014, **42**, 35–53.
- 19 Z. Sapci, *Bioresour. Technol.*, 2013, **128**, 487–494.
- 20 W. Charles, L. Walker and R. Cord-Ruwisch, *Bioresour. Technol.*, 2009, **100**, 2329–2335.
- 21 J. E. Johansen and R. Bakke, *Water Sci. Technol.*, 2006, **53**, 43–50.
- 22 P. Tsapekos, P. G. Kougias, S. A. Vasileiou, G. Lyberatos and I. Angelidaki, *Bioresour. Technol.*, 2017, **225**, 246–253.
- 23 S. Kim and M. T. Holtzapple, *Bioresour. Technol.*, 2005, **96**, 1994–2006.
- 24 K. E. Kang, G. T. Jeong, C. Sunwoo and D. H. Park, *Bioprocess Biosyst. Eng.*, 2012, **35**, 77–84.



- 25 Z. Wang, D. R. Keshwani, A. P. Redding and J. J. Cheng, *Bioresour. Technol.*, 2010, **101**, 3583–3585.
- 26 L. Wu, M. Arakane, M. Ike, M. Wada, T. Takai, M. Gau and K. Tokuyasu, *Bioresour. Technol.*, 2011, **102**, 4793–4799.
- 27 J. Kainthola, A. Podder, M. Fechner and R. Goel, *Bioresour. Technol.*, 2021, **321**, 124397.
- 28 F. Monlau, A. Barakat, J. P. Steyer and H. Carrere, *Bioresour. Technol.*, 2012, **120**, 241–247.
- 29 Y. Qian, X. Zhou, Y. Zhang, W. Zhang and J. Chen, *Chemosphere*, 2013, **91**, 717–723.
- 30 A. Zhang, J. Wang and Y. Li, *Water Res.*, 2015, **71**, 125–139.
- 31 Y. Li, J. Wang, A. Zhang and L. Wang, *Water Res.*, 2015, **83**, 84–93.
- 32 X. Huang, W. Dong, H. Wang, F. Sun and Y. Feng, *J. Cleaner Prod.*, 2019, **228**, 1526–1536.
- 33 A. Goi, M. Viisimaa, M. Trapido and R. Munter, *Chemosphere*, 2011, **82**, 1196–1201.
- 34 H. Wu, J. Gao, D. Yang, Q. Zhou and W. Liu, *Chem. Eng. J.*, 2010, **160**, 1–7.
- 35 J. Wang and Y. Li, *Appl. Energy*, 2016, **183**, 1123–1132.
- 36 W. Tian, Y. Chen, Y. Shen, C. Zhong, M. Gao, D. Shi, Q. He and L. Gu, *Sci. Total Environ.*, 2020, **732**, 139312.
- 37 T. Weide, C. D. Baquero, M. Schomaker, E. Brüggling and C. Wetter, *Biomass Bioenergy*, 2020, **132**, 105442.
- 38 D. Xu, Z. Ding, J. Bai, W. Ke, Y. Zhang, F. Li and X. Guo, *Bioresour. Technol.*, 2020, **310**, 123476.
- 39 Q. Feng and Y. C. Song, *Energy Fuels*, 2016, **30**, 6467–6474.
- 40 G. Kumar, B. Sen, P. Sivagurunathan and C.-Y. Lin, *Fuel*, 2016, **182**, 131–140.
- 41 J. Zhang, C. Kong, M. Yang and L. Zang, *ACS Omega*, 2020, **5**, 9151–9161.
- 42 X. Kang, Y. Sun, L. Li, X. Kong and Z. Yuan, *Bioresour. Technol.*, 2018, **255**, 205–212.
- 43 R. Narinthorn, W. Choorit and Y. Chisti, *Biomass Bioenergy*, 2019, **127**, 105262.
- 44 M. Solé-Bundó, H. Carrère, M. Garfí and I. Ferrer, *Algal Res.*, 2017, **24**, 199–206.
- 45 A. Hiden, *Bioresour. Technol.*, 2017, **230**, 140–142.
- 46 Y. Sheng, X. Tan, Y. Gu, X. Zhou, M. Tu and Y. Xu, *Renewable Energy*, 2021, **163**, 732–739.
- 47 S. Gandolfi, G. Ottolina, R. Consonni, S. Riva and I. Patel, *ChemSusChem*, 2014, **7**, 1991–1999.
- 48 A. M. Mustafa, H. Li, A. A. Radwan, K. Sheng and X. Chen, *Bioresour. Technol.*, 2018, **259**, 54–60.
- 49 C. C. Hernandez, F. F. Ferreira and D. S. Rosa, *Carbohydr. Polym.*, 2018, **193**, 39–44.
- 50 E. Jin, J. Guo, F. Yang, Y. Zhu, J. Song, Y. Jin and O. J. Rojas, *Carbohydr. Polym.*, 2016, **143**, 327–335.
- 51 A. D. French and M. Santiago Cintrón, *Cellulose*, 2013, **20**, 583–588.
- 52 D. Kumari and R. Singh, *Renewable Sustainable Energy Rev.*, 2018, **90**, 877–891.
- 53 S. Bolado-Rodríguez, C. Toquero, J. Martín-Juárez, R. Travaini and P. A. García-Encina, *Bioresour. Technol.*, 2016, **201**, 182–190.
- 54 D. Pečar, F. Pohleven and A. Goršek, *Waste Manage.*, 2020, **102**, 170–178.

