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Bimetallic cobalt–iron diselenide nanorod modified glassy carbon electrode: an electrochemical sensing platform for the selective detection of isoniazid†

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The increasing demand of a sensitive and portable electrochemical sensing platform in pharmaceutical analysis has developed widespread interest in preparing electrode materials possessing remarkable properties for the electrochemical determination of target drug analytes. Herein, we report the synthesis, characterization and application of bimetallic cobalt–iron diselenide (FeCoSe₂) nanorods as electrode modifiers for the selective detection of a commonly used anti-tuberculosis drug Isoniazid (INZ). We prepared FeCoSe₂ nanorods by a simple hydrothermal route and characterized these by X-ray diffraction (XRD), scanning electron microscopy (SEM), transmission electron microscopy (TEM), energy dispersive X-ray spectroscopy (EDX) and temperature-programmed reduction (TPR) techniques. The electrochemical characterization of FeCoSe₂ modified GCE was performed by cyclic voltammetry (CV) and square wave anodic stripping voltammetry (SWASV). Under optimized experimental conditions, a linear current-concentration response was obtained for INZ in the range of 0.03–1.0 μM, with very low limit of detection 1.24 × 10^{−10} M. The real applicability of the designed FeCoSe₂/GCE sensing platform was adjudicated by the detection of INZ in biological samples.

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Introduction

Isoniazid (INZ) chemically named as pyridine-4-carboxylic acid hydrazide is the most commonly used anti-tuberculosis drug.¹ Tuberculosis is an infectious bacterial disease caused by *Mycobacterium tuberculosis* (MBT) and INZ has a great potency to suppress the activity of MBT.² In this regard, it is highly important to monitor the INZ level in biological fluid to control and regulate the level of this drug for effective treatment.^{3,4} The chemical structure of INZ is shown in Fig. 1.

A variety of analytical approaches such as high-performance liquid chromatography, Raman spectroscopy, chemiluminescence, colorimetry, capillary electrophoresis, fluorimetry, chemometry, titrimetry, polarography, and spectrophotometry are used for INZ analysis.^{5–9} All these

techniques have their own advantages but every technique has some limitations. Therefore, efforts are underway to search for effective tools on case to case basis. Electrochemical techniques are attracting attention owing to their peculiar features of being cheaper, easy to operate, portable, sensitive and selective. Many research teams have employed modified electrodes for INZ detection.^{10–15} But the modified electrodes suffer from serious fouling effects that can be minimized by the right selection of supporting electrolyte and type of material used for electrode modification.^{16–19}

Fig. 1 Chemical structure of isoniazid (INZ).

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Metal chalcogenides and specifically metal diselenides (MSe_2) have attained great attention for ongoing research.^{20–22} The higher metallic character of Se in chalcogenides might attributed to their extraordinary physical and electrochemical properties. In recent years, various metal selenides MSe_2 ($\text{M} = \text{Co}, \text{Fe}, \text{Mo}, \text{Cu}, \text{and Ni}$) have been extensively applied as effective electrocatalysts in electrochemical applications.^{23–26} Among various metal diselenides, FeSe_2 and CoSe_2 have been mostly investigated due to their excellent electrical conductivity, low diffusion length for ion/electron, high specific capacitance, mechanical stability, low charge transfer resistance, and significant recycling ability.^{27–30} In fact, 3d electrons with a low spin in Co metal facilitate the metallic characteristic of CoSe_2 which leads to increase in the charge transfer during the electrocatalytic process. The electrocatalytic activity of CoFeSe_2 alloy could be enhanced due to the synergistic effect between Fe and CoSe_2 . The hetero-metallic bond formation in bimetallic alloys results in the tuning of the bonding patterns of catalyst surface with the reactant molecules (INZ).³¹ The strong metal–metal interactions in bimetallic systems also modify the electrical and mechanical properties which make them favorable for different electrocatalytic applications.

Cobalt diselenide nanobelts grafted on carbon fiber presented a robust electrochemical performance that can be correlated to the synergistic effect between CoSe_2 and carbon fiber besides 3D cathode architecture providing catalytic stability.³² Liu *et al.* successfully fabricated 3D $\text{Ni}_{(1-x)}\text{Co}_x\text{Se}_2$ mesoporous nanosheet on Ni foam with tunable stoichiometry.³³ The collective effects of special morphological design and electronic structure engineering enable the integrated highest catalytic activity and outstanding stability in a wide pH range. Recently, Zheng *et al.*, studied phase transition of CoSe_2 from cubic to orthorhombic with appropriate doping of phosphorus.³⁴ The striking electrocatalytic performance can be attributed to the favorable electronic structure and local coordination environment created by this doping-induced structural phase transition strategy. In literature, most of the studies are focused on electrochemical

oxidation/reduction reactions using cobalt diselenide, whereas electrochemical studies using composite materials of cobalt diselenide are rather limited.

Herein, we report FeCoSe_2 nanorods modified GCE for electrochemical detection of INZ in neutral media. Our objective is to use comparatively cheaper electrode modifier for more sensitive detection of INZ. The morphological features of metal nanorods provide excellent electrochemical sensing platforms due to high surface area and higher exposure of specific crystal facets. To the best of our knowledge, no previous work is reported on the use of $\text{FeCoSe}_2/\text{GCE}$ for the electroanalytical detection of INZ. To get a robust electroanalytical signal for INZ on the $\text{FeCoSe}_2/\text{GCE}$, we have optimized experimental conditions. We have validated the real-world applicability of $\text{FeCoSe}_2/\text{GCE}$ in the real samples.

Experimental section

Chemicals and apparatus

Isoniazid ($\text{C}_6\text{H}_7\text{N}_3\text{O}$, Merck, $\geq 99\%$), iron(II) chloride tetrahydrate ($\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$, Merck, $\geq 99\%$), cobalt(II) chloride hexahydrate ($\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$, Merck, 98%), sodium hydroxide (NaOH , Merck, $\geq 97\%$), EDTA ($\text{C}_{10}\text{H}_{14}\text{N}_2\text{Na}_2\text{O}_8 \cdot 2\text{H}_2\text{O}$, Merck, 99%), boric acid (H_3BO_3 , Merck, $\geq 99.5\%$), and phosphoric acid (H_3PO_4 , Merck, $\geq 99\%$) were purchased and used as received without any purification. The Britton Robinson buffer (BRB) of pH 7 was used as supporting electrolyte which was prepared by mixing 0.04 M H_3BO_3 , 0.04 M H_3PO_4 , and 0.04 M CH_3COOH . The pH of BRB was adjusted to 7 by adding 0.2 M NaOH solution. All the solutions were made in doubly distilled water.

Instrumentation

XRD analysis was performed on Rigaku D/max 2500 with a radiation source Cu $\text{K}\alpha 1$ ($\lambda = 1.54056 \text{ \AA}$). SEM and TEM images of the prepared samples were recorded on electron microscope JEM 2100 (USA) equipped with EDS unit for chemical analysis. The accelerating voltage was kept at 10 kV and 100

Fig. 2 Schematic representation of the synthesis of CoFeSe_2 modification.

kV for SEM and TEM images, respectively. TPR analysis of the synthesized nanocomposite was performed on ChemiSorb 2750 (Micromeritics-USA) to evaluate the different metallic phases and alloying effects of constituent metals. The surface area analysis was performed using Sorptometer Kelvin 1042 (Costech Instruments) at liquid N₂ temperature (−196 °C) after degassing the samples at 200 °C for 2 h.

Catalyst preparation

At first, CoSe₂ was synthesized by following the hydrothermal route as shown in Fig. 2. The solution labeled as A was prepared by mixing 0.8 mmol of CoCl₂·6H₂O with 2 mL of 0.5 M ethylene diamine tetra-acetic acid (EDTA). Another solution labeled as B was prepared by mixing 0.162 mmol of Se powder in 8 mL of 3.3 M NaOH. Both solutions A and B were then mixed *via* ultrasonication. After that, the resultant mixture was shifted to autoclave and kept at 180 °C for 18 h for the hydrothermal reaction. Similarly, FeCoSe₂ bimetallic catalyst was prepared by adding 0.4 mmol of FeCl₂·4H₂O to the above mentioned solution A. In each case, the black product was collected and washed with deionized water followed by air drying. The total quantity of metal sources (Fe + Co) was kept very low and constant (0.8 mmol). Finally, feeding ratios of metals was further confirmed by EDX.

Electrode fabrication

Before each electrochemical analysis, GCE was polished using alumina powder (0.05 μm) and carefully washed in an ultrasonic bath with doubly distilled water, followed by a mixture of acetone and nitric acid (1 : 1) mixture, and finally washed by doubly distilled water and dried in a desiccator. The 1 mg of FeCoSe₂ was dispersed in 1 mL of isopropanol *via* ultrasonication for 30 minutes to achieve uniform dispersion. Then, 5 μL of FeCoSe₂ solution was drop casted over the polished surface of bare GCE (0.07 cm²). The fabricated electrode was first dried at room temperature for 20 minutes and then completely dried in the oven at 50 °C for 30 minutes. The FeCoSe₂/GCE was then employed for electrochemical analysis of INZ under optimized conditions.

Electrochemical characterization

Cyclic voltammetry and SWASV measurements were performed at room temperature on potentiostat Gamry interface 1000. The three electrode setup was used for electrochemical studies. Ag/AgCl (3 M KCl), a platinum wire and modified GCE were used as reference, counter and working electrodes respectively. Measurement of pH of solutions was done using INOLAB pH meter. The INZ concentration was quantified by assessing oxidation peak current values using the SWASV technique. SWASV voltammograms were recorded under optimized conditions (*i.e.*, deposition voltage of 0.0 V and deposition time of 90 s by using both pure GCE and modified FeCoSe₂/GCE electrodes. For voltammetric investigations experimental conditions were optimized by varying: scan rate, volume of catalyst suspension for electrode fabrication, pH, and buffer solution. These optimizations were done and the best current response was obtained in a medium of pH 7 using catalyst suspension of 5 μL for electrode modification. Prior to each measurement, pre-

conditioning was done in consecutive 20 runs in the respective potential window to reduce signal to noise ratio.

Results and discussion

XRD analysis

Fig. 3 illustrates the XRD pattern of the synthesized FeCoSe₂ bimetallic nanocatalyst. The sharp and intense peaks of XRD show that the synthesized material has high crystallinity. The diffraction peaks at 2θ ~30.71°, 34.37°, 35.8°, 47.8°, 50.41°, 53.33°, and 55.3° in the XRD spectra of FeCoSe₂ correspond to (101), (110), (120), (211), (002), (031), and (131) *hkl* planes, respectively (Powder Diffraction Standard Database: JCPDS # 21-0432).

Surface area analysis

To assess the surface properties such as the specific surface area and pore particularity, CoFeSe₂ nanocomposite was further explored by N₂ adsorption-desorption isotherms as shown in Fig. 4. A typical type IV isotherm (Fig. 4A) with H3 hysteresis loop between 0.4 and 0.8 *P/P*₀ reveals the mesoporous nature of the material as per IUPAC classification.³⁵ The BET surface area of CoFeSe₂ nanocatalyst was found to be 93.14 m² g^{−1}. The specific surface area is highly influenced by the surface morphologies and internal porous structures. Besides, the pore size distribution curve (Fig. 4B) was derived from the desorption isotherm by applying the Barret–Joyner–Halenda (BJH) method. Narrow scattering of pores was obtained with the maximum distribution of pores with a size of 5.5 nm, reflecting a unimodal behavior. The large value of the BET area and distinct mesoporous nature contributed towards introducing more active sites which were later manifested by optimum electrochemical sensing of isoniazid over the electrode surface modified with this synthesized catalyst.

Morphological characterizations

SEM images of FeCoSe₂ (Fig. 5(A–C)) indicate the presence of FeCoSe₂ nanorods with the diameter ranging from 40 to 90 nm which is suitable for electrochemical applications. It displayed the uniform distribution of FeCoSe₂ nanorods with no agglomeration high metal dispersion of this optimal catalyst

Fig. 3 XRD pattern of the synthesized FeCoSe₂ bimetallic catalyst.

Fig. 4 (A) N₂ adsorption-desorption isotherm and (B) pore size distribution curve of CoFeSe₂ nanorods using BJH method.

resulted to a better catalytic activity for hydrazine electro-oxidation, later on. Furthermore, the TEM image (Fig. 5D) ensures the presence of rod type structures in the synthesized material with an average size of 40 nm.

EDX analysis

EDX analysis was performed at randomly selected points over the surface of the prepared samples (Fig. S1†). Fig. 6A shows the EDX spectra of CoFeSe₂ with resultant sharp peaks of Co, Fe, and Se elements. The considerable quantitative weight percentage was recorded for Co, Fe, and Se elements of about 19.26%, 1.21%, and 59.91%, respectively. The co-existence of Co, Fe, and Se presented the synergistic effect to form metal alloy NPs. Moreover, EDX mapping (Fig. 6B–E) of CoFeSe₂

depicted the distribution of Co, Fe, and Se metals with the uniform dispersion.

Fig. 7 presents TPR profiles of the prepared nanocomposite. In general, two step reduction of cobalt oxide (Co₃O₄) occurs as follows:³⁶

TPR analysis

TPR profile of monometallic Co catalyst showed the 3 reduction peaks in the range of 150–800 °C, respectively. The appearance of these reduction peaks can be attributed to the reduction of

Fig. 5 (A–C) SEM micrographs at various magnifications and (D) TEM image of synthesized FeCoSe₂ nanorods.

Fig. 6 (A) EDX profile for chemical composition and (B–E) EDX mapping for elemental distribution of the prepared CoFeSe₂ nanocomposite.

Co₃O₄ to CoO and further reduction of CoO to metallic Co⁰.³⁷ In comparison, the CoSe₂ catalyst presented ease in reduction behavior than that of the monometallic Co sample. In the case of CoFeSe₂, the incorporation of Fe metal even in a small content led to a decline in the reduction temperature of cobalt oxide and reduction peaks were shifted towards lower temperature region. In this aspect, the present study suggested that the alloying effect of the constituent metals (Co, Fe, and Se) may promote the catalytic efficiency of CoFeSe₂/GCE towards electro-sensing of INZ. Overall, the synergistic effect might be accountable for the development in the field of catalysis.

Active surface area measurements

Voltammetric investigations of the FeCoSe₂/GCE were performed using 5 mM potassium ferrocyanide (K₃[Fe(CN)₆]/K₄[Fe(CN)₆]) solution as a redox probe. Fig. S2† shows the cyclic voltammograms recorded by using bare and modified GCE.

There were observed sharp peaks with a narrow peak potential difference and an obvious enhancement in the oxidation-reduction peak currents in the CV of FeCoSe₂/GCE compared to bare GCE, signifying rapid electron transfer process.³⁸

The active surface area of the electrodes was estimated according to the Randles-Sevcik equation: $i_{pa} = 2.69 \times 10^5 n^3/2 AD^{1/2} \nu^{1/2} C$, where A is the active surface area (cm²), i_{pa} is the redox peak current of potassium ferricyanide, n is the number of electrons transferred (2 in this case), D is the diffusion coefficient (0.76×10^{-5} cm² s⁻¹, 25 °C), ν is the scan rate (V s⁻¹) and C is the concentration of analyte (5 mM). The calculated active surface area of bare GCE and modified electrode are found to be 0.071 cm² and 0.117 cm², respectively. The larger active surface area is obtained for the modified electrode shows the greater electrochemical activity of the FeCoSe₂/GCE as compare to bare GCE.

Electrooxidation of INZ at FeCoSe₂/GCE modified electrode

The electrocatalytic oxidation of Isoniazid was evaluated by SWASV technique. Fig. 8A shows the square wave voltammograms for the oxidation of INZ at bare and FeCoSe₂/GCE in the BRB solution of pH 7. At a potential of 0.45 V, an oxidation signal for INZ was obtained by SWASV. The current response for INZ oxidation at FeCoSe₂/GCE is manifestly promoted as compared to bare GCE. This enhancement in the current signal by using FeCoSe₂/GCE is attributed to the larger active surface area, good conductivity, and excellent electro-sensing potency of the FeCoSe₂ nanorods. Fig. 8B shows the schematic presentation for electrochemical oxidation mechanism of INZ over modified FeCoSe₂/GCE.³⁹ These outcomes verified that the projected FeCoSe₂/GCE is efficiently accomplished for the voltammetric sensing of INZ.

Optimization of experimental parameters

The pH of the medium has a pronounced effect on INZ detection and was observed using FeCoSe₂/GCE by changing the pH of the solution from 4 to 8 as shown in Fig. 9A. It was seen that

Fig. 7 TPR profiles of the synthesized nanocatalysts.

Fig. 8 (A) SWASV of (a) bare GCE in 25 μM solution of INZ, (b) FeCoSe₂/GCE in 25 μM solution of INZ solution, and (c) FeCoSe₂/GCE in solvent (BRB of pH 7) containing no INZ at a scan rate of 50 mV s^{-1} , accumulation potential of 0.0 V, and accumulation time of the 90 s, and (B) electrochemical oxidation mechanism of INZ over modified FeCoSe₂/GCE.

the peak current value increases steadily with increasing the pH of the solution up to pH 7 and then a steady fall in current response was recognized by further increasing pH of the solution. Hence, pH 7 was optimized for further experimental work. Furthermore, a shift in peak potential for oxidation of INZ was observed in the negative direction with an increase in pH of the

solution, signifying the direct involvement of the proton in electrode processes. Also, the slope value 52.48 mV per unit pH was obtained from the plot of E_p vs. pH (Fig. 9B) which demonstrates that the oxidation of INZ involves equal no of electrons and protons.

The effect of changing accumulation potential (−0.4 to 0.3) on the oxidation peak current of INZ was studied by SWASV as shown in Fig. S3A.† The oxidation peak current magnitude increases gradually towards positive potential direction until it reached its maximum value at 0.0 V thereafter, it decreased abruptly. So, an accumulation potential of 0.0 V was selected as optimum potential. The effect of accumulation time was also studied as shown in Fig. S3B.† It showed the enhancement in the peak current value up to 90 s, afterward the current value declines with increasing time, and hence accumulation time of 90 s was selected for further experimental work.

Under optimized conditions calibration plot for INZ was achieved by SWASV in the concentration range of 0.03–1 μM as shown in Fig. 10A. The results obtained showed a direct relation to the oxidation peak current against INZ concentration. The limit of detection (LOD) and limit of quantification (LOQ) for INZ detection were calculated by using eqn (3) and eqn (4), respectively

$$\text{LOD} = 3s/m \quad (3)$$

$$\text{LOQ} = 10s/m \quad (4)$$

Here, “s” is the standard deviation of the peak current of blank solution (three runs) and “m” denotes the slope of current *versus* concentration plot.⁴⁰

Fig. 10B presents the effect of INZ concentration on its peak current response at the modified FeCoSe₂/GCE. From the linear relationship of the peak current and INZ concentration the LOD and LOQ with values of 1.24×10^{-10} M and 4.14×10^{-10} M were evaluated respectively. The regression data and various other parameters obtained from calibration curves are presented in Table S1.†

Stability and reproducibility of fabricated GCE

To check the precision of the proposed method, repeatability and reproducibility of the fabricated GCE was investigated.

Fig. 9 (A) SWASV of 16 μM solution of INZ using FeCoSe₂/GCE in 4–8 pH range and (B) Plots of E_p and I_p vs. pH.

Fig. 10 (A) SWAS voltammograms of INZ in the concentration range of 0.03–1 μM at $\text{FeCoSe}_2/\text{GCE}$ in BRB solution (pH 7) under optimized conditions (scan rate of 50 mV s^{-1} , accumulation potential of 0.0 V, and accumulation time of 90 s and (B) the plot of I_{pa} versus concentration.

The intra-day precision of the $\text{FeCoSe}_2/\text{GCE}$ was checked by performing three repeated measurements within a day and the % RSD was found to be 2.22%. The stability based on inter-day precision was checked by performing three repeated measurements between days by SWASV technique and the % RSD was found to be 0.56%. The relative standard deviation (% RSD) was calculated for isoniazid detection and found to be in acceptable range *i.e.*, less than 5% as shown in Fig. S4.† The results achieved specify that the $\text{FeCoSe}_2/\text{GCE}$ have good precision, thus can be employed for sensitive and selective detection of INZ.

Interference study

In order to study the interference of various interferences, which commonly exist in the biological samples, some

interference experiments were performed as shown in Fig. S5.† It was observed that 500 folds' excess of Ca^{2+} , NH_4^+ , Mg^{2+} , glucose, 200 folds of ascorbic acid and uric acid, showed almost no interference to anodic peak current response of INZ (RSD = 1.59%). The interference experiment showed that the fabricated GCE has the ability of anti-jamming. The comparison of this work with reported modified electrodes for INZ detection is summarized in Table 1.

Practical application of $\text{FeCoSe}_2/\text{GCE}$ for INZ analysis

To check the practical applicability of the $\text{FeCoSe}_2/\text{GCE}$, the analyte INZ was assessed in physiological fluids *i.e.*, in human serum and urine samples by applying SWASV technique. Detection of drugs in physiological fluids can help in following their metabolic fate. The biological fluids were originally taken

Table 1 Comparison of analytical parameters of our modified $\text{FeCoSe}_2/\text{GCE}$ electrode with reported ones for the determination of isoniazid using various techniques

Electrodes	Technique	Linear range (μM)	Medium	LOD	Ref.
GC/Ag-P(MMA-co-AMPS)ESP	Amperometry	0.05–150	0.1 M PBS (pH 7)	10 nM	11
GO/PAG modified GCE	Differential pulse voltammetry (DPV)	20–1400	PBS (pH 7)	2.59 μM	12
Cu/GO/GCE	Amperometry	0.1–500	Phosphate buffer (pH 7)	23.4 nM	13
SPCE/PH electrode	Differential pulse voltammetry (DPV)	0.5–110	Phosphate buffer (pH 7)	17 μM	14
MWCPE	Differential pulse voltammetry (DPV)	1–1000	Acetate buffer (pH 4)	0.5 μM	15
PdNPs/CILE	Cyclic voltammetry (CV)	5–100	Phosphate buffer (pH 7)	0.47 μM	16
ERGO/GCE	LSV	2–70	Phosphate buffer (pH 7)	0.17 μM	17
RGO-Au/GCE	DPV	0.1–1000	Phosphate buffer (pH 7)	0.01 μM	18
CoTRP(dcbpy) $_2$ -Ni/GO	BIA	100–1000	KNO_3 (pH 7)	3.5 μM	19
$\text{FeCoSe}_2/\text{GCE}$	SWASV	0.03–1	BRB (pH 7)	0.12 nM	This work

Table 2 INZ determination in real samples (urine and human serum) by proposed SWASV technique

Sample	Analyte	INZ conc. added (μM)	INZ conc. detected (μM)	Recovery (%)	RSD (%) ($n = 3$)
Urine	INZ	5	4.94	98.8	2.5
Human serum	INZ	2	1.96	98.0	3.4

from a healthy volunteer. Each biological sample was diluted up to 15 times by using BRB of pH 7 and then spiked by INZ under commonly used standard addition methods. Each sample was analyzed three times ($n = 3$) to check the reproducibility. The recovery results obtained for spiked INZ samples by using FeCoSe₂/GCE showed a very low RSD value of 2.5% and 3.4% for urine and human serum samples, respectively. This outcome validated that the FeCoSe₂/GCE has its specific extensive reliability towards detection of INZ in biological fluids. The findings of the real sample analyses are listed in Table 2.

Conclusions

In the present work FeCoSe₂ modified GCE was successfully fabricated and used for electrochemical determination of isoniazid. The FeCoSe₂/GCE showed figures of merit in the context of wider linear range, lower limit of detection and strong resilience to interfering agents. Owing to the exceptional properties of FeCoSe₂, it offers an enhanced oxidation response of INZ in neutral medium. The higher sensitivity and lower detection limit are attributed to good conductivity, larger surface area, excellent catalytic properties and amplification effect of FeCoSe₂ nanorods. The FeCoSe₂/GCE also displayed great potential application for INZ detection in human serum and urine samples with acceptable recoveries. The robust performance of FeCoSe₂/GCE bimetallic catalysts would be correlated to the synergistic effect of constituent metals, crystallinity, unique morphology (nanorods), uniform dispersion of alloy particles, narrow pore size distribution, and presence of more active sites as ensured by XRD, SEM, TEM, EDX, and TPR analyses.

Ethical statement

This article does not contain any direct studies on human participants and animals. The experiments were performed in compliance with the relevant laws and institutional guidelines of Quaid-i-Azam University, Islamabad, with prior approval from the ethical committee. Only the assay corresponding to practical testing of the designed sensor includes the use of urine and human serum. The urine sample was obtained with informed consent of the healthy donor and according to institutional guidelines. While the drug free synthetic human serum was purchased from Sigma Aldrich.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 S. Hammi, K. E. Ataouna, K. Bouti, O. Elbouazzi, M. Laine and J. E. Bourkadi, Importance of Therapeutic Drug Monitoring in the Treatment of Active Tuberculosis - A Retrospective Study of 4 Cases, *Int. J. Surg. Med.*, 2016, **3**, 10.
- 2 J. Rosa, T. C. Machado, A. K. D. Silva, G. Kuminek, A. J. Bortolluzzi, T. Caon and S. G. Cardoso, Isoniazid-Resveratrol Cocrystal: A Novel Alternative for Topical Treatment of Cutaneous Tuberculosis, *Cryst. Growth Des.*, 2019, **19**, 5029.
- 3 U. P. Azad and V. Ganesan, Efficient electrocatalytic oxidation and selective determination of isoniazid by Fe(tmphen)³⁺-exchanged Nafion®-modified electrode, *J. Solid State Electrochem.*, 2012, **16**, 2907.
- 4 B. G. Mahmoud, M. Khairy, F. A. Rashwan and C. E. Banks, Simultaneous Voltammetric Determination of Acetaminophen and Isoniazid (Hepatotoxicity-Related Drugs) Utilizing Bismuth Oxide Nanorod Modified Screen-Printed Electrochemical Sensing Platforms, *Anal. Chem.*, 2017, **89**, 2170.
- 5 M. Khuhawar, Liquid chromatographic determination of isoniazid, pyrazinamide and rifampicin from pharmaceutical preparations and blood, *J. Chromatogr. B*, 2002, **766**, 357.
- 6 L. Toit, V. Pillay and Y. Choonara, A Mathematical Approach for the Simultaneous *In Vitro* Spectrophotometric Analysis of Rifampicin and Isoniazid from Modified-Release Anti-TB Drug Delivery Systems, *Curr. Drug Delivery*, 2010, **7**, 5.
- 7 P. R. Chellini, T. O. Mendes, P. H. Franco, B. L. Porto, V. K. Tippavajhala, I. C. César, M. A. Oliveira and G. A. Pianetti, Simultaneous determination of rifampicin, isoniazid, pyrazinamide and ethambutol in 4-FDC tablet by Raman spectroscopy associated to chemometric approach, *Vib. Spectrosc.*, 2017, **90**, 14.
- 8 M. A. Lomillo, R. O. Domínguez and M. Mj. Arcos, Resolution of ternary mixtures of rifampicin, isoniazid and pyrazinamide by differential pulse polarography and partial least squares method, *Anal. Chim. Acta*, 2001, **449**, 167.
- 9 M. Ghoneim, K. El-Baradie and A. Tawfik, Electrochemical behavior of the antituberculosis drug isoniazid and its square-wave adsorptive stripping voltammetric estimation in bulk form, tablets and biological fluids at a mercury electrode, *J. Pharm. Biomed. Anal.*, 2003, **33**, 673.
- 10 X. Yan, X. Bo and L. Guo, Electrochemical behaviors and determination of isoniazid at ordered mesoporous carbon modified electrode, *Sens. Actuators, B*, 2011, **155**, 837.
- 11 P. K. Rastogi, V. Ganesan and U. P. Azad, Electrochemical determination of nanomolar levels of isoniazid in pharmaceutical formulation using silver nanoparticles decorated copolymer, *Electrochim. Acta*, 2016, **188**, 818.
- 12 B. Devadas, S. Cheemalapati, S.-M. Chen, M. A. Ali and F. M. A. Al-Hemaid, Highly sensing graphene oxide/poly-arginine-modified electrode for the simultaneous

- electrochemical determination of buspirone, isoniazid and pyrazinamide drugs, *Ionics*, 2014, **21**, 547.
- 13 T. S. Balamurugan, K. Manibalan, S.-T. Huang, P. Balasubramanian and S. M. Chen, High Sensitive Electrochemical Quantification of Isoniazid in Biofluids Using Copper Particles Decorated Graphene Oxide Nano Composite, *Int. J. Electrochem. Sci.*, 2017, **12**, 9150.
 - 14 M. F. Bergamini, D. P. Santos and M. V. B. Zanoni, Determination of isoniazid in human urine using screen-printed carbon electrode modified with poly-L-histidine, *Bioelectrochemistry*, 2010, **77**, 133.
 - 15 S. Shahrokhian and M. Amiri, Multi-walled carbon nanotube paste electrode for selective voltammetric detection of isoniazid, *Microchim. Acta*, 2006, **157**, 149.
 - 16 G. Absalan, M. Akhond, M. Soleimani and H. Ershadifar, Efficient electrocatalytic oxidation and determination of isoniazid on carbon ionic liquid electrode modified with electrodeposited palladium nanoparticles, *J. Electroanal. Chem.*, 2016, **761**, 1.
 - 17 S. Cheemalapati, S. Palanisamy and S.-M. Chen, Electrochemical determination of Isoniazid at electrochemically reduced graphene oxide modified electrode, *Int. J. Electrochem. Sci.*, 2013, **8**, 3953.
 - 18 Z. Guo, Z.-Y. Wang, H.-H. Wang, G.-Q. Huang and M.-M. Li, Electrochemical sensor for Isoniazid based on the glassy carbon electrode modified with reduced graphene oxide–Au nanomaterials, *Mater. Sci. Eng., C*, 2015, **57**, 197.
 - 19 J. S. Aguirre-Araque, J. M. Goncalves, M. Nakamura, P. O. Rossini, L. Angnes, K. Araki and H. E. Toma, GO composite encompassing a tetra-ruthenated cobalt porphyrin–Ni coordination polymer and its behavior as isoniazid BIA sensor, *Electrochim. Acta*, 2019, **300**, 113.
 - 20 M. F. Bergamini, D. P. Santos and M. V. Zanoni, Determination of isoniazid in human urine using screen-printed carbon electrode modified with poly-L-histidine., *Bioelectrochemistry*, 2010, **77**, 133.
 - 21 S. Mani, S. Ramaraj, S.-M. Chen, B. Dinesh and T.-W. Chen, Two-dimensional metal chalcogenides analogous NiSe₂ nanosheets and its efficient electrocatalytic performance towards glucose sensing., *J. Colloid Interface Sci.*, 2017, **507**, 378.
 - 22 Y.-R. Zheng, M.-R. Gao, Q. Gao, H.-H. Li, J. Xu, Z.-Y. Wu and S.-H. Yu, An Efficient CeO₂/CoSe₂ Nanobelt Composite for Electrochemical Water Oxidation, *Small*, 2015, **11**, 182.
 - 23 L. Mi, H. Sun, Q. Ding, W. Chen, C. Liu, H. Hou, Z. Zheng and C. Shen, 3D hierarchically patterned tubular NiSe with nano-/microstructures for Li ion battery design, *Dalton Trans.*, 2012, **41**, 12595.
 - 24 A. Eftekhari, Molybdenum diselenide (MoSe₂) for energy storage, catalysis, and optoelectronics, *Appl. Mater. Today*, 2017, **8**, 1.
 - 25 S. M. Tan, Z. Sofer, J. Luxa and M. Pumera, Aromatic-Exfoliated Transition Metal Dichalcogenides: Implications for Inherent Electrochemistry and Hydrogen Evolution, *ACS Catal.*, 2016, **6**, 4594.
 - 26 M. Gilic, M. Petrovic, J. Cirkovic, N. Paunovic, S. Savic-Sevic, Z. Nikitovic, M. Romcevic, I. Yahia and N. Romcevic, Low-temperature photoluminescence of CuSe₂ nano-objects in selenium thin films, *Process. Appl. Ceram.*, 2017, **11**, 127.
 - 27 C.-H. Ding, J.-J. Tang, S. Chen, Z.-Q. Liu and N. Li, Co_{0.85}Se/ Multi-Walled Carbon Nanotube Composite as Alternative Cathode Catalyst for Microbial Fuel Cells, *J. Nanosci. Nanotechnol.*, 2017, **17**, 1438.
 - 28 A. K. Dutta, S. K. Maji, A. Mondal, B. Karmakar, P. Biswas and B. Adhikary, Iron selenide thin film: peroxidase-like behavior, glucose detection and amperometric sensing of hydrogen peroxide, *Sens. Actuators, B*, 2012, **173**, 724.
 - 29 T. J. Theerthagiri, R. Sudha, K. Premnath, P. Arunachalam, J. Madhavan and A. M. Al-Mayouf, Growth of iron diselenide nanorods on graphene oxide nanosheets as advanced electrocatalyst for hydrogen evolution reaction., *Int. J. Hydrogen Energy*, 2017, **42**, 13020.
 - 30 B. Yu, J. Jin, H. Wu, S. Wang, Q. Xia and H. Liu, Iron and nickel doped CoSe₂ as efficient non precious metal catalysts for oxygen reduction., *Int. J. Hydrogen Energy*, 2017, **42**, 236.
 - 31 A. K. Singh and Q. Xu, Synergistic catalysis over bimetallic alloy nanoparticles, *ChemCatChem*, 2013, **5**, 652.
 - 32 Y.-R. Zheng, M.-R. Gao, Z.-Y. Yu, Q. Gao, H.-L. Gao and S.-H. Yu, Cobalt diselenide nanobelts grafted on carbon fiber felt: an efficient and robust 3D cathode for hydrogen production, *Chem. Sci.*, 2015, **6**, 4594.
 - 33 B. Liu, Y.-F. Zhao, H.-Q. Peng, Z.-Y. Zhang, C.-K. Sit, M.-F. Yuen, T.-R. Zhang, C.-S. Lee and W.-J. Zhang, Electrocatalysts: Nickel–Cobalt Diselenide 3D Mesoporous Nanosheet Networks Supported on Ni Foam: An All-pH Highly Efficient Integrated Electrocatalyst for Hydrogen Evolution, *Adv. Mater.*, 2017, **29**, 19.
 - 34 Y.-R. Zheng, P. Wu, M.-R. Gao, X.-L. Zhang, F.-Y. Gao, H.-X. Ju, R. Wu, Q. Gao, R. You, W.-X. Huang, S.-J. Liu, S.-W. Hu, J. Zhu, Z. Li and S.-H. Yu, Doping-induced structural phase transition in cobalt diselenide enables enhanced hydrogen evolution catalysis., *Nat. Commun.*, 2018, **9**, 1.
 - 35 K. S. W. Sing, Reporting physisorption data for gas/solid systems with special reference to the determination of surface area and porosity (Recommendations 1984), *Pure Appl. Chem.*, 1985, **57**, 603.
 - 36 G. S. Sewell, S. E. Van and C. T. O'Connor, Use of TPR/TPO for characterization of supported cobalt catalysts, *Catal. Lett.*, 1996, **37**, 255.
 - 37 H.-Y. Lin and Y.-W. Chen, The mechanism of reduction of cobalt by hydrogen, *Mater. Chem. Phys.*, 2004, **85**, 171.
 - 38 B. Yu, J. Jin, H. Wu, S. Wang, Q. Xia and H. Liu, Iron and nickel doped CoSe₂ as efficient non precious metal catalysts for oxygen reduction, *Int. J. Hydrogen Energy*, 2017, **42**, 236.
 - 39 R. Chokkareddy, N. K. Bhajanthri and G. G. Redhi, An enzyme-induced novel biosensor for the sensitive electrochemical determination of isoniazid, *Biosensors*, 2017, **7**, 21.
 - 40 M. S. Quintino and L. Angnes, Fast BIA-amperometric determination of isoniazid in tablets, *J. Pharm. Biomed. Anal.*, 2006, **42**, 400.