


 Cite this: *RSC Adv.*, 2021, **11**, 16246

Light-driven selective aerobic oxidation of (iso)quinoliniums and related heterocycles†

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Selective C1–H/C4–H carbonylation of *N*-methylene iminium salts, catalyzed by visible-light photoredox and oxygen in the air, has been reported. A ruthenium complex acts as a chemical switch to conduct two different reaction pathways and to afford two different kinds of products. In the absence of the ruthenium complex, the Csp²–H bonds adjacent to the nitrogen atoms are oxidized to α -lactams by the *N*-methyleneiminium substrates themselves as photosensitizers. In the presence of the ruthenium complex, the oxidation reaction site of quinoliniums is switched to the C4 region, resulting in the formation of 4-quinolones. The use of two transformations directly introduces oxygen into the nitrogen heterocyclic skeletons under an air atmosphere.

 Received 15th February 2021
 Accepted 23rd March 2021

DOI: 10.1039/d1ra01226f

rsc.li/rsc-advances

Introduction

The biomimetic characteristics of isoquinolone and quinolone rings, reminiscent of the rigidly fixed skeleton of *N*-heterocycles, frequently emerge in many natural products and pharmaceuticals.¹ For example, the alkaloids doryanine and sarcomejine have been isolated from *Cryptocarya chinensis* (Lauraceae) and *Sarcomelicope megistophylla* (Rutaceae), respectively.² The isoquinolone scaffold is widely found as the core structure in topoisomerase I (Top I) inhibitors.³ Elvitegravir (EVG), which contains the elementary unit of the corresponding quinolone, is a potent inhibitor of human immunodeficiency virus type 1 (HIV-1) integrase (Fig. 1).⁴ Due to their diverse biological activities, tremendous effort has been devoted to the effective synthesis of (iso)quinolones. The traditional methods usually require stoichiometric amounts of K₃Fe(CN)₆ as an oxidant, and this inevitably produces a considerable number of toxic by-products such as K₄Fe(CN)₆.⁵ Other methods, such as Conrad–Limpach and Niementowski reactions, have proven to be powerful tactics using the condensation of amines with carboxyl derivatives followed by cyclization to generate quinolone analogs.⁶

However, most of these syntheses suffer from harsh reaction conditions (high temperature, Eaton reagent, *etc.*), poor conversion of the substrates and the use of excessive oxidants or additives. Consequently, considerable efforts have been devoted to establishing mild, environmentally friendly and sustainable procedures for these valuable scaffolds.

Using O₂ (or ambient air) as an oxidant is the most ideal methodology for direct oxygenation.⁷ Moreover, visible-light-promoted aerobic oxidations have been demonstrated in the synthesis of biologically active molecules in green and environmentally friendly conditions.⁸ Recently, Fu *et al.* successfully achieved the visible-light mediated *ortho* C–H oxidation of *N*-alkyliminiums using Eosin Y as the photocatalyst and air as the oxidant, obtaining isoquinoline-1-ones (Scheme 1a).⁹ Deng and co-authors highlighted the photocatalytic oxidation and subsequent cyclization protocol to transform indoles to quinolones.¹⁰ The attractive properties of these photocatalytic reactions were their mild reaction conditions, low catalyst loading, ubiquity and versatility functionality of directing C–H activation with O₂ (or air). In our designed method, the isoquinolinium salts (**1**) formed excited isoquinolinium salts (**1***) under visible-light irradiation without other photosensitizers,¹¹ and then underwent photoinduced electron transfer (PET) to transform the α -carbon radicals (**A**), which were captured by O₂ to obtain isoquinolones (**2**) (Scheme 1b). This process sets the stage for

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† Electronic supplementary information (ESI) available: ¹H, ¹⁹F and ¹³C NMR spectra of compounds **2** and **3**. See DOI: 10.1039/d1ra01226f

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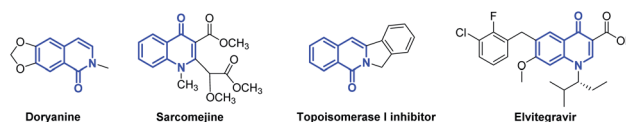
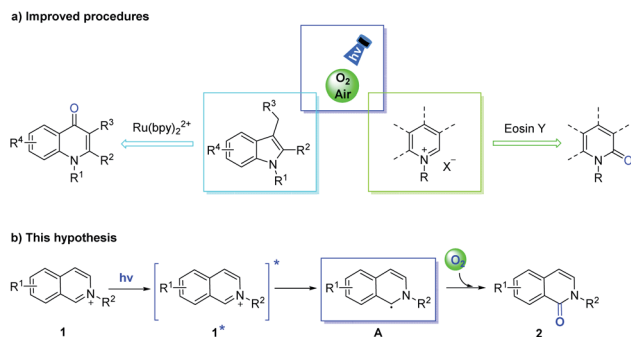


Fig. 1 Representative examples of natural products and pharmaceuticals containing the (iso)quinolone structural motif.





Scheme 1 The representative photocatalysis of oxidative approaches to the construction of *N*-substituted isoquinolin-1(2*H*)-ones and 4-quinolones (a), and our hypothesis (b).

carbonylation to the oxidation onto the C1-position of isoquinoliniums, and another successful strategy that obtains quinoliniums by *para*(C4)-selective oxidation will significantly broaden the potential of remote site-selective functionalization. Herein, we delineate the successful realization of different reactive sites in (iso)quinoliniums and related heterocycles through visible-light sources, orchestrating a Ru-photocatalyst, which controls the *ortho*- or *para*-position of oxidation.

Experimental section

General methods

Unless otherwise specified, commercial reagents and solvents were used without further purification. All manipulations were performed in air at room temperature. For flash column chromatography, 100–200 mesh and 300–400 mesh silica gels were used by eluting with petroleum ether/ethyl acetate. ¹H and ¹³C spectra were recorded on a 400 MHz spectrometer. The chemical shifts were reported in ppm. ¹H NMR spectra were referenced to DMSO (2.50 ppm) or CDCl₃ (7.26 ppm), and ¹³C NMR spectra were referenced to DMSO (39.5 ppm) or CDCl₃ (77.0 ppm). Peak multiplicities were designated by the following abbreviations: s, singlet; d, doublet; t, triplet; m, multiplet, and J is the coupling constant in Hz. HRMS spectra were recorded with a Micromass QTOF2 Quadrupole/time-of-flight tandem mass spectrometer using electron spray ionization.

The synthesis of halogenated *N*-methylene iminiums (1a as the example)

A solution of isoquinoline (5 mmol), benzyl bromide (10 mmol) and CH₃CN (15 mL) was refluxed for 12 hours and cooled to room temperature. After the addition of ethyl acetate, isoquinoline salt was precipitated rapidly into a solid, and this was then filtered and washed with ethyl acetate to obtain a white solid **1a**. The volatile solvents were removed by vacuum evaporation.

The synthesis of α -lactams

A solution of isoquinoline (5 mmol), halogenated *N*-methylene iminiums **1** (0.2 mmol), and DABCO (0.4 mmol), was added to a 10 mL quartz tube in CH₃CN (2 mL), and then the mixture was irradiated with green LEDs (15 W) (approximately 1.5 cm away

from the light source) at room temperature in an air atmosphere. After complete conversion of the substrate (monitored by TLC), the reaction mixture was diluted with 20 mL of EtOAc and the solution was filtered by flash chromatography (petroleum ether/ethyl acetate 10 : 1). The filtrate was evaporated by rotary evaporator and the residue was purified by silica gel column chromatography to give the desired product **2**.

The synthesis of 4-quinolones

Halogenated quinoliniums **1ag–1aq** (0.2 mmol), DABCO (0.4 mmol), and Ru(bpy)₃Cl₂ (0.004 mmol) were added to a 10 mL quartz tube in CH₃CN (2 mL), and then the mixture was irradiated with blue LEDs (15 W) (approximately 1.5 cm away from the light source) at room temperature in an air atmosphere. After complete conversion of the substrate (monitored by TLC), the reaction mixture was diluted with 20 mL of EtOAc, and the solution was filtered by flash chromatography (petroleum ether/ethyl acetate 2 : 1). The filtrate was evaporated by rotary evaporator and the residue was purified by silica gel column chromatography to give the desired product **3**.

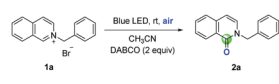
In vitro cardiovascular activities

The MOVAS (mouse aorta vascular smooth muscle cells), HUVEC (human umbilical vein endothelial cells) and AC16 (human cardiomyocytes cells) cell lines were all obtained from the Institute of Biochemistry and Cell Biology, China Academy of Sciences. The cells were cultured in DMEM (Sigma), which was supplemented with 10% fetal bovine serum in a humidified atmosphere of 5% CO₂/95% air at 37 °C. All compounds were dissolved in phosphate buffered saline (PBS) with 1% DMSO to obtain various concentrations in 96-well plates, whilst control wells contained supplemented media with 1% DMSO, and the wells continued to be incubated for 48 h at 37 °C in a 5% CO₂ atmosphere. MTT (5 mg mL⁻¹) was added into the wells and the plates were incubated at 37 °C for 4 h. The MTT assay was stopped by adding dimethyl sulfoxide (100 μ L per well) and mixing for 10 min vigorously before measuring the absorbance at 490 nm in a multi-well plate reader. The cytotoxicity was estimated based on the percentage cell survival in a dose dependent manner relative to the negative control. The final IC₅₀ (drug concentration that kills 50% of the cells) values were calculated by the Bliss method. All target compounds were tested on MOVAS, HUVEC and AC16 cell lines, respectively.

Results and discussion

We started to investigate the reactivity of *N*-benzylisoquinolinium **1a** as a model substrate in an air atmosphere and at room temperature (Table 1; more details in the ESI†). The optimal reaction was achieved with substrate **1a** as the photosensitizer, 1,4-diazobicyclo[2.2.2]octane (DABCO) as a base, and air as an oxidant in CH₃CN under the irradiation of 15 W blue light-emitting diodes (LEDs), thereby affording the carbonylation product **2a** (entry 1). Inhibition of the reactivity was observed in the presence of other solvents (entries 2 and 3). Other bases, such as Et₃N and KO^tBu, were also effective for this C–H



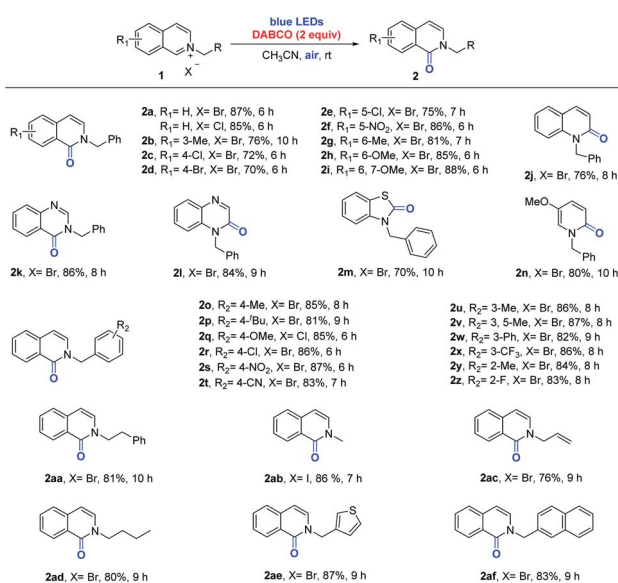
Table 1 Model studies of *ortho* C–H oxidation^a


Entry	Substrate	Variation from standard conditions	2a ^b (%)
1	1a	None	87
2	1a	DMF instead of CH ₃ CN	22
3	1a	THF instead of CH ₃ CN	58
4	1a	Et ₃ N instead of DABCO	54
5	1a	KO ^t Bu instead of DABCO	67
6	1a	Addition 2 mol% Ru(bpy) ₃ Cl ₂	63
7	1a	Without light	21
8	1a	Without base	NR


^a Reaction conditions: *N*-methylene iminiums **1** (0.2 mmol), DABCO (2.0 equiv.), LED lamp as light source, room temperature, under air atmosphere. ^b Isolated yield.

oxidation transformation, but in inferior yields (entries 4 and 5). There was a negative effect on the reaction with the addition of the photocatalyst Ru(bpy)₃Cl₂ or without light (entries 6 and 7). The experiments showed that no desired product was formed in the absence of base, revealing the crucial role of the base in this reaction (entry 8).

According to the readily available optimization conditions, we significantly extended the scope of *ortho*-selective oxidation to synthesize α -lactams. As shown in Scheme 2, isoquinolinium chloride (X = Cl) showed good reaction activity and the target product **2a** was obtained in 85% yield. Moderate yields were realized when the pyridine ring contained Me, Cl and Br functional groups (**2b–2d**). Substituting the benzene ring with electron-donating groups, such as methyl and methoxy groups, and with electron-withdrawing groups, such as chloro- and nitro-groups, proceeded smoothly to give isoquinolones **2e–2i**



Scheme 2 The *ortho* C–H selective oxidation reaction using *N*-methylene iminium salts.^a Reaction conditions: *N*-methylene iminiums **1** (0.2 mmol), DABCO (2.0 equiv.), blue LED (15 W), room temperature, under air atmosphere. ^b Isolated yield.

Table 2 Model studies of C4-selective oxidation^a


Entry	Substrate	Variation from standard conditions	3a ^b (%)
1	1ag	None	68
2	1ag	Eosin Y instead of Ru(bpy) ₃ Cl ₂	NR
3	1ag	1,4-Dioxane instead of CH ₃ CN	33
4	1ag	Et ₃ N instead of DABCO	NR
5	1ag	Without light	Trace
6	1ag	Without base	NR

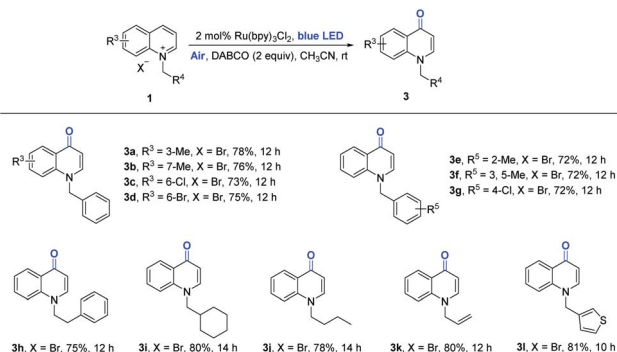
^a Reaction conditions: *N*-methylene iminiums **1** (0.2 mmol), DABCO (2.0 equiv.), Ru(bpy)₃Cl₂ (2.0 mol%), LED lamp as light source, room temperature, under air atmosphere. ^b Isolated yield.

in 75–88% yields. *N*-Benzylquinolinium was transformed into the corresponding quinoline-2-one **2j**. It is worth noting that heteroarene derivatives, such as quinazolinium, quinoxalinium, benzothiazolium and 3-methoxypyridinium, were produced smoothly by our standard protocol as products **2k–2n**. Substituent R represents the phenyl group, such as the various electron-donating groups (Me, ^tBu, OMe, and Ph) and electron-withdrawing groups (Cl, NO₂, CN, CF₃, and F), that can be transformed smoothly to obtain corresponding products (**2o–2z**). Many functional groups, including benzyl and alkyl, were well tolerated (**2aa–2ad**). Among them, isoquinolinium halides (X = Cl and I) also proceeded in good yields (**2q** and **2ab**). In addition, the heterocyclic substrates were well tolerated in the oxidative reactions, and target products **2ae** and **2af** were obtained with yields of 87% and 83%, respectively.

When 3-methylquinolinium (**1ag**) instead of *N*-benzylisoquinolinium bromide (**1a**) was applied to this system, 4-quinolone **3a** was successfully achieved in a 68% yield using the Ru-catalyst (Table 2, entry 1). Further screening of the reactions could not be effectively performed with other photocatalysts, solvents and bases (entries 2–4). No aerobic oxidation occurred in the absence of light or base, demonstrating that these components were essential factors (entries 5 and 6).

Subsequently, we assessed the reaction of quinolinium substrates with a ruthenium catalyst, and this protocol straightforwardly photocatalyzed the carbonylation of the Csp²–H bond to afford the desired products with excellent C4-regiocontrol, as illustrated in Scheme 3. The quinoline ring bearing various groups, such as methyl, chloro- and bromo-groups, were successfully engaged in the current photocatalytic oxidation reaction (**3b–3d**). The benzene ring with different electronic groups, connected to the nitrogen atom, maintained selectivity for the C4-site (**3e–3g**). The substrate with the benzyl group was still suitable for the reaction, and the product **3h** was obtained in a 75% yield. Other alkyl quinoliniums, such as cyclohexane, *n*-propyl, and vinyl, were also well-tolerated and provided the corresponding products (**3i–3k**) in 78%–80% yields. Thiophene quinolinium was tolerated to give the desired product **3l** in a good yield.



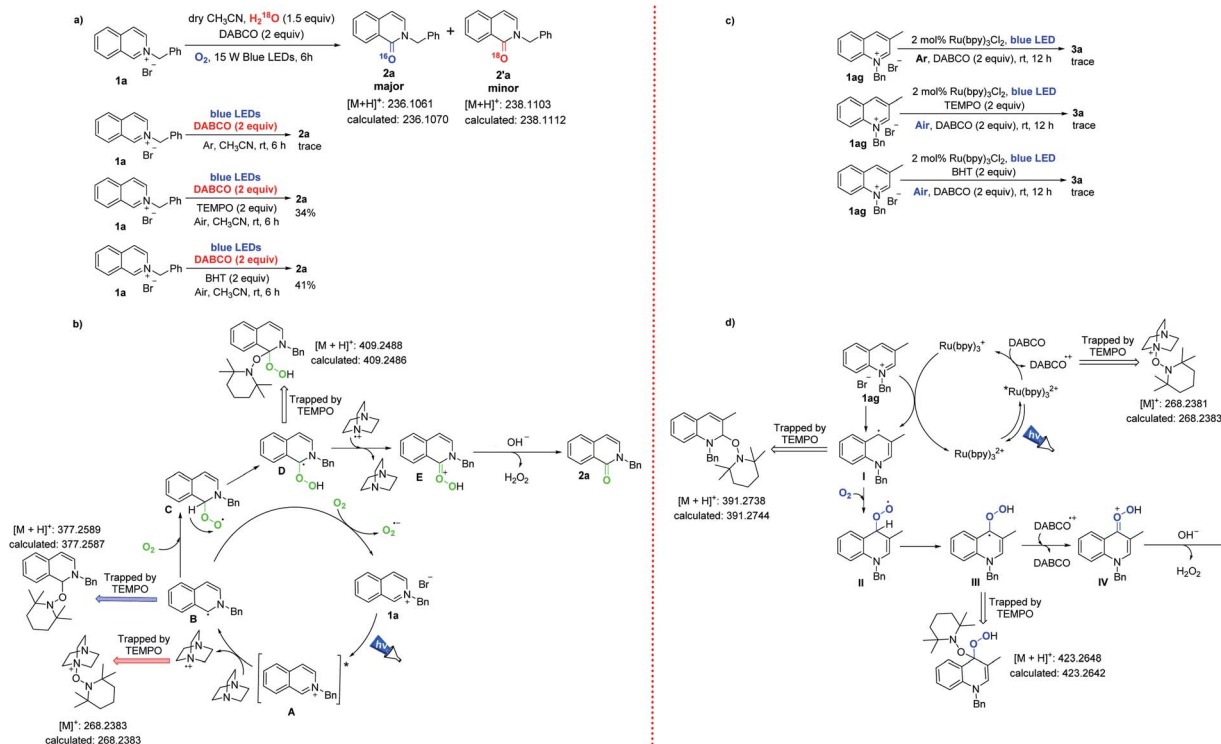


Scheme 3 The C4-selective oxidation reaction using *N*-benzylquinolinium salts. ^a Reaction conditions: quinoliniums **1ag**–**1ar** (0.2 mmol), DABCO (2.0 equiv.), Ru(bpy)₃Cl₂ (2.0 mol%), blue LED (15 W), ambient temperature, under air atmosphere. ^b Isolated yield.

Having established our developed methods and scope for a wide range of applications, we investigated the proposed mechanism. Isotope experiments indicated that the oxygen atom in the target products was mainly derived from oxygen in the atmosphere (Scheme 4a). Only a trace amount of isoquinolinone **2a** was observed in an argon atmosphere. Under the optimal reaction conditions, the reaction was clearly inhibited by the addition of 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO) or 2,6-di-*tert*-butyl-4-methylphenol (BHT), indicating that the conversion could have undergone a free radical process. It is well known that *N*-functionalized onium salts act as photosensitizers.¹¹ Hence, the excited state **A** was formed

under continuous visible light irradiation. Subsequently, the nitrogen-centered cation **A** could undergo photoinduced electron transfer to generate the α -amino radical **B** (trapped by TEMPO),¹² and DABCO was simultaneously oxidized to DABCO⁺ (trapped by TEMPO). The radical **B** was coupled with oxygen in the air to produce the peroxide radical **C**.¹³ The other route for radical **B** involved being oxidized to the ground state isoquinolinium **1a**, while O₂ was reduced to O₂⁻, and so the photocatalytic cycle was completed. Intermediate **C** continued to form the carbon radical **D** (trapped by TEMPO) through hydrogen atom transfer (HAT), and then **D** was oxidized to the oxygen cation **E** by DABCO⁺, and was then simultaneously reduced to DABCO. Finally, treatment of the cation **E** with NaOH and Br⁻ afforded the desired adduct **2a** (Scheme 4b).¹⁴

Similarly, a trace amount of 4-quinolone **3a** was observed in an argon atmosphere (Scheme 4c). The reaction was inhibited by adding the free radical inhibitors TEMPO or BHT under the optimal reaction conditions, indicating that the conversion involves a radical process. Based on these control experiments, we speculated the following mechanism (Scheme 4d). Photo-activated Ru(II)* could be reduced to Ru(I) species by DABCO. The redox potential of Ru(bpy)₃Cl₂ was $E_{1/2}$ Ru(II)* / Ru(I) = 0.77 V vs. SCE in CH₃CN,¹⁵ and this was higher than $E_{1/2}$ DABCO = 0.6 V vs. SCE.¹⁶ Therefore, the photoredox reactions could occur spontaneously between Ru(bpy)₃Cl₂ and DABCO. 3-Methylquinolinium (**1ag**) could accept a single electron from the Ru(I) species to form the C4-site quinoline radical **I**,^{11b,17} which was coupled with oxygen to form the peroxy radical **II**. The C4-position radical **III** was generated from **II**, which was



Scheme 4 Control experiments (a and c) and the proposed mechanism (b and d).



Table 3 IC₅₀ values of the tested compounds^a

Compound	IC ₅₀ ^b (μM)		
	MOVAS	HUVEC	AC16
2g	59.14 ± 2.31	>100	>100
2t	94.93 ± 4.57	>100	>100

^a IC₅₀ represents the concentration that inhibits 50% of cell proliferation. ^b Each value is expressed in μM and represents the mean of three data sets.

promoted by a hydrogen atom transfer (HAT) process. This was similar to the above oxidation of isoquinoliniums, in that intermediate **III** was oxidized to cation **IV** by DABCO⁺ (radical cation) and then reacted with base to remove H₂O₂ to obtain target product **3a**.

The *in vitro* cardiovascular activities of all the synthesized compounds were tested on several cell lines, including MOVAS (mouse aorta vascular smooth muscle cells), HUVEC (human umbilical vein endothelial cells) and AC16 (human cardiomyocytes cells). As reported in Table 3, the biological activities were expressed as IC₅₀ values. The table shows that two products demonstrated moderate activities of MOVAS cells with IC₅₀ values of 59.14 ± 2.31 and 94.93 ± 4.57, respectively. The results showed that these products only had potential effects on mouse aorta vascular smooth muscle cells. Unfortunately, the products did not inhibit the activity of HUVEC and AC16 cells.

Conclusions

In summary, we have developed the selective oxidation of Csp²-H functionalization by employing oxygen under light-driven conditions. Clearly, the ruthenium complex was used as a chemical switch to trigger two different oxidation pathways of the *N*-onium salts. In the absence of the Ru-complex, the *N*-functionalized onium ions were irradiated from the α -amino radicals of tertiary amines to the corresponding amides using air as an oxidant. On the other hand, the quinoliniums underwent single electron transfer to form the C4-site quinoline radical using the Ru(bpy)₃Cl₂ catalyst, and this was shown to be smoothly carried out by the formation of 4-quinolones. The outstanding advantages of these transformations are that carbonylation at different sites of the *N*-methyleneiminiums could be directly oxidized to obtain diverse ketones under air and at room temperature.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This study was supported by the National Natural Science Foundation of China (21977021 and 81760626), the Ministry of Education Innovation Team Fund (IRT_16R15, 2016GXNSFGA380005), and the Natural Science Foundation of

Guangxi Province (AB17292075) and Guangxi Funds for Distinguished Experts.

Notes and references

- (a) M. J. Fisher, B. P. Gunn, S. Um and J. A. Jakubowski, *Tetrahedron Lett.*, 1997, **38**, 5747–5750; (b) H. Huse and M. Whiteley, *Chem. Rev.*, 2011, **111**, 152–159; (c) D. B. Khadka and W. J. Cho, *Bioorg. Med. Chem.*, 2011, **19**, 724–734; (d) Z. Jin, *Nat. Prod. Rep.*, 2013, **30**, 849–868; (e) B.-R. Kang, S. Li, S. Mao, Y.-X. Cao and S.-Q. Zhang, *Bioorg. Med. Chem.*, 2015, **25**, 5808–5812; (f) Y.-M. Ma, K. Qiao, Y. Kong, M.-Y. Li, L.-X. Guo, Z. Miao and C. Fan, *Nat. Prod. Res.*, 2017, **31**, 951–958; (g) J. Drogosz-Stachowicz, A. Długosz-Pokorska, K. Gach-Janczak, A. Jaskulska, T. Janecki and A. Janecka, *Chem. Biol. Interact.*, 2020, **320**, 109005.
- (a) N. Fokialakis, P. Magiatis, A. L. Skaltsounis, F. Tillequin and T. Sévenet, *J. Nat. Prod.*, 2000, **63**, 1004–1005; (b) T. S. Wu and F. W. Lin, *J. Nat. Prod.*, 2001, **64**, 1404–1407; (c) S. Mitaku, N. Fokialakis, P. Magiatis and F. Tillequin, *Fitoterapia*, 2007, **78**, 169–170; (d) P. H. Coombes, E. M. Mwangi, B. K. Peters, N. R. Crouch and D. A. Mulholland, *Biochem. Syst. Ecol.*, 2009, **37**, 494–496; (e) J.-Y. Cho, S.-H. Bae, H.-K. Kim, M.-L. Lee, Y.-S. Choi, J.-Y. Cho, S.-H. Bae, H.-K. Kim, M.-L. Lee, Y.-S. Choi, B.-R. Jin, H. J. Lee, H. Y. Jeong, Y. G. Lee and J.-H. Moon, *J. Agric. Food Chem.*, 2015, **63**, 3587–3592.
- (a) A. Morrell, S. Antony, G. Kohlhagen, Y. Pommier and M. Cushman, *J. Med. Chem.*, 2006, **49**, 7740–7753; (b) H. T. My Van and W. J. Cho, *Bioorg. Med. Chem.*, 2009, **19**, 2551–2554.
- (a) M. Sato, H. Kawakami, T. Motomura, H. Aramaki, T. Matsuda, M. Yamashita, Y. Ito, Y. Matsuzaki, K. Yamataka, S. Ikeda and H. Shinkai, *J. Med. Chem.*, 2009, **52**, 4869–4882; (b) S. Ramanathan, A. A. Mathias, P. German and B. P. Kearney, *Clin. Pharmacokinet.*, 2011, **50**, 229–244.
- (a) I. M. Khalil, D. Barker and B. R. Copp, *J. Org. Chem.*, 2016, **81**, 282–289; (b) T. T. Fan-Chiang, H. K. Wang and J.-C. Hsieh, *Tetrahedron*, 2016, **72**, 5640–5645.
- (a) J. K. Son, S. I. Kim and Y. Jahng, *Heterocycles*, 2001, **55**, 1981–1986; (b) D. Zewge, C. Y. Chen, C. Deer, P. G. Dormer and D. L. Hughes, *J. Org. Chem.*, 2007, **72**, 4276–4279; (c) J. C. Brouet, S. Gu, N. P. Peet and J. D. Williams, *Synth. Commun.*, 2009, **39**, 1563–1569.
- (a) A. A. Ghogare and A. Greer, *Chem. Rev.*, 2016, **116**, 9994–10034; (b) Y.-F. Liang and N. Jiao, *Acc. Chem. Res.*, 2017, **50**, 1640–1653; (c) Y. Zhang, W. Schilling and S. Das, *ChemSusChem*, 2019, **12**, 2898–2910; (d) W. Schilling, Y. Zhang, P. K. Sahoo, S. K. Sarkar, S. Gandhi, H. W. Roesky and S. Das, *Green Chem.*, 2021, **23**, 379–387.
- (a) Q. Liu and L. Z. Wu, *Natl. Sci. Rev.*, 2017, **4**, 359–380; (b) A. Motaleb, A. Bera and P. Maity, *Org. Biomol. Chem.*, 2018, **16**, 5081–5085; (c) X. Zhang, K. P. Rakesh, L. Ravindar and H. L. Qin, *Green Chem.*, 2018, **20**, 4790–4833; (d) Y. Zhou, W. Liu, Z. Xing, J. Guan, Z. Song and Y. Peng, *Org. Chem. Front.*, 2020, **7**, 2405–2413; (e) W. Schilling, Y. Zhang, D. Riemer and S. Das, *Chem.-Eur. J.*, 2020, **26**, 390–395; (f)



- Y. Zhou, W. Liu, Z. Xing, J. Guan, Z. Song and Y. Peng, *Org. Chem. Front.*, 2020, **7**, 2405–2413.
- 9 Y. Jin, L. Ou, H. Yang and H. Fu, *J. Am. Chem. Soc.*, 2017, **139**, 14237–14243.
- 10 X. Ji, D. Li, Z. Wang, M. Tan, H. Huang and G. J. Deng, *Asian J. Org. Chem.*, 2018, **7**, 711–714.
- 11 (a) P. Hewavitharanage, E. O. Danilov and D. C. Neckers, *J. Org. Chem.*, 2005, **70**, 10653–10659; (b) K. Ohkubo, T. Kobayashi and S. Fukuzumi, *Angew. Chem., Int. Ed.*, 2011, **50**, 8652–8655; (c) J. P. Dinnocenzo, P. B. Merkel and S. Farid, *J. Phys. Chem. A*, 2017, **121**, 7903–7909.
- 12 (a) Y. L. Chow, W. C. Danen, S. F. Nelsen and D. H. Rosenblatt, *Chem. Rev.*, 1978, **78**, 243–274; (b) U. C. Yoon and P. S. Mariano, *Acc. Chem. Res.*, 1992, **25**, 233–240; (c) X. Zhang, S.-R. Yeh, S. Hong, M. Freccero, A. Albin, D. Falvey and P. S. Mariano, *J. Am. Chem. Soc.*, 1994, **116**, 4211–4220; (d) P. Renaud and L. Giraud, *Synthesis*, 1996, **1996**, 913–926.
- 13 (a) Z. Zhang, J. Su, Z. Zha and Z. Wang, *Chem. Commun.*, 2013, **49**, 8982–8984; (b) Q. B. Zhang, Y. L. Ban, D. G. Zhou, P. P. Zhou, L. Z. Wu and Q. Liu, *Org. Lett.*, 2016, **18**, 5256–5259; (c) Y. Zhang, D. Riemer, W. Schilling, J. Kollmann and S. Das, *ACS Catal.*, 2018, **8**, 6659–6664.
- 14 (a) N. Suzuki, Y. Kazui, T. Tsukamoto, M. Kato and Y. Izawa, *Bull. Chem. Soc. Jpn.*, 1983, **56**, 1519–1521; (b) J. Motoyoshiya, H. Inoue, Y. Takaguchi and H. Aoyama, *Heteroat. Chem.*, 2002, **13**, 252–257.
- 15 (a) C. Creutz and N. Sutin, *Inorg. Chem.*, 1976, **15**, 496–499; (b) Y. Matsubara, K. Koga, A. Kobayashi, H. Konno, K. Sakamoto, T. Morimoto and O. Ishitani, *J. Am. Chem. Soc.*, 2010, **132**, 10547–10552; (c) J. M. R. Narayanam and C. R. J. Stephenson, *Chem. Soc. Rev.*, 2011, **40**, 102–113.
- 16 S. F. Nelsen and P. J. Hintz, *J. Am. Chem. Soc.*, 1972, **94**, 7114–7117.
- 17 (a) Y. Su, L. Zhang and N. Jiao, *Org. Lett.*, 2011, **13**, 2168–2171; (b) R. R. Kotha, J. J. Nash and H. I. Kenttämaa, *Eur. J. Org. Chem.*, 2017, **2017**, 1407–1412; (c) D. Ding, H. Jiang, X. Ma, J. J. Nash and H. I. Kenttämaa, *J. Org. Chem.*, 2020, **85**, 8415–8428; (d) J. M. Ganley, P. R. D. Murray and R. R. Knowles, *ACS Catal.*, 2020, **10**, 11712–11738.

