



Cite this: *RSC Adv.*, 2021, 11, 11583

Received 8th February 2021
Accepted 12th March 2021

DOI: 10.1039/d1ra01070k

rsc.li/rsc-advances

An efficient and scalable synthesis of 2,4-di-*N*-acetyl-L-altrose (L-2,4-Alt-diNAC)[†]

Anna Niedzwiecka, Carita Sequeira, Ping Zhang and Chang-Chun Ling *

Bacterial nonulosonic acids such as pseudaminic acids and others constitute a family of 9-carbon monosaccharides that contain a common 3-deoxy-2-ketoacid fragment but differ in their stereochemistries at 5 stereogenic centers between C-4 to C-8. Their unique structures make them attractive targets for use as antigens in vaccinations to combat drug-resistant bacterial infections and their challenging stereochemistries have attracted considerable attention from chemists. In this work we report the development of an improved synthesis for 2,4-di-*N*-acetyl-L-altrose (L-2,4-Alt-diNAC), which is a key hexose required for the chemical and chemoenzymatic synthesis of pseudaminic acids. Using L-fucose as a starting material, our synthesis overcomes several pitfalls in previously reported syntheses.

Introduction

Bacterial infection is becoming an increasing cause of mortality and morbidity world-wide as traditional antibiotics continuously lose their effectiveness to pathogen resistance mechanisms.¹ There are currently very few antimicrobial drug candidates undergoing clinical trials,² creating an urgent need for alternative antibacterial strategies. The vaccination approach has demonstrated long-lasting beneficial effects, while resistance to it has rarely been observed.³ The success of a vaccine critically depends on the choice of proper antigens. Pseudaminic acid (Pse, **1**, Fig. 1), along with legionaminic acid (**2**) and its 8- or 4-epimers (**3–4**), are members of a class of unique 9-carbon sugars called nonulosonic acids, commonly distinguished by their 3-deoxy-2-ketoacid fragment.⁴

Pse and its derivatives are structural components of flagella and pili,^{5,6} exclusive to several Gram-positive bacteria: among them, *Pseudomonas aeruginosa* is the main cause of mortality for people suffering from cystic fibrosis, and *Acinetobacter baumannii* is a common cause of ventilator associated pneumonia (VAP).⁴ These pathogens thrive in mucous-rich environments, relying on flagella and pili as efficient motility systems for their virulence.^{5,7,8} Studies have shown that mutants with the inability to incorporate Pse into their cell structures lose mobility, thus their ability to colonize and infect hosts.⁹ In addition to their structural significance, these types of carbohydrates could function as interesting antigens for immunogenic vaccine-induced antibodies because of their unique L-altro-

configuration between C5–C9, combined with their acylamido substitution at both 5,7-positions and their 9-deoxygenation^{10,11} (Fig. 1).

In nature, the acyl groups attached to the two amido groups could either be identical (commonly acetyl) or different, depending on the bacterial strain.⁴ For example, *P. aeruginosa* O7a produces a version of Pse that has a formamido group at C-7 and an acetamido group at C-5, while the related O9a strain produces another version with a (*R*)-3-hydroxybutanamido at C-7 and the same acetamido group at C-5; for *Vibrio cholera* O:2, another variant of Pse with the C-5 amido substituted by an acetamidoyl group is produced. In the biosynthesis of Pse, the intermediate 2,4-dideoxy-2,4-diacetamido-L-altrose (L-Alt-diNAC, **5**) is enzymatically extended *via* an aldol condensation with

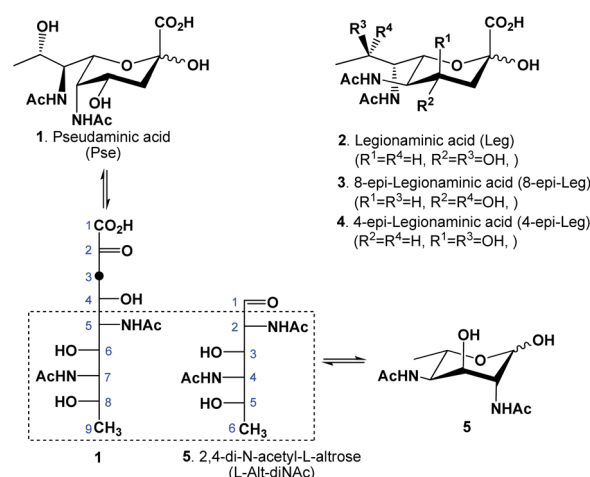


Fig. 1 Structure of pseudaminic acid and its biosynthetic precursor L-Alt-diNAC (**5**).

Department of Chemistry, University of Calgary, 2500 University Dr NW, Calgary, AB, T2N 1N4, Canada. E-mail: ccling@ucalgary.ca

[†] Electronic supplementary information (ESI) available. See DOI: 10.1039/d1ra01070k



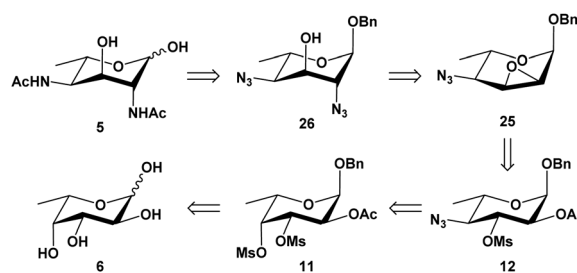
phosphoenolpyruvate (PEP).^{12,13} Analogously, suitably protected L-Alt-diNAc substrates have also been used for the chemical synthesis of Pse, typically by reaction with methyl 2-(bromomethyl)acrylate *via* indium-mediated allylation,¹⁴ followed by an ozonolysis to cleave the alkene, revealing the desired 3-keto functionality. There have to date been several chemical syntheses of Pse reported in literature with varying success; some suffered from long reaction sequences, others from low overall yield, or difficulty in scaling up. For example, in 2001, Zähringer reported¹⁵ a synthesis from L-rhamnose which took advantage of its 6-deoxygenation and L-configuration. Although the use of L-sugar¹⁶ represented an advantage because of a configurational match at the C-5 position, the conversion to a 2-epimer of L-Alt-diNAc was achieved in a long and rather tedious reaction sequence (16 steps), as the introduction of the two acetamido groups required two double inversions, which was inefficient. Before that, in early 1970, Liav and Sharon reported¹⁷ a synthesis of L-Alt-diNAc from β -L-fucopyranoside, by first converting it to a 2,3-anhydro-4-O-mesyl- β -L-talopyranoside intermediate that then underwent a nucleophilic attack by azide; unfortunately, this reaction resulted in a mixture that included the 2,4-diazido-2,4,6-trideoxy- β -L-altropyranoside in very poor yield (16%) and 2,4-diazido-2,4,6-trideoxy- β -L-idopyranoside, also in very poor yield (23%). Ito and coworkers later reported an improved synthesis of Pse¹⁸ *via* the L-Alt-diNAc intermediate from the readily available N-acetyl-D-glucosamine starting material. Their synthesis benefited from the initial configurations of the acetamido group at the C-2 position and the hydroxyl group at C-3 of the hexose, which matched those of the desired compound. The reaction sequence in their case was also lengthy (14 steps to L-Alt-diNAc) due to the required 6-deoxygenation, the inversion of configurations at both C-4 and C-5 centers and the introduction of an acetamido group to C-4. More recently Li and coworkers¹⁹ established an elegant *de novo* synthesis of L-Alt-diNAc and Pse from L-threonine: their synthesis included an initial multistep transformation of L-threonine to a suitably protected 2-amino-2,4-dideoxy-L-erythrose that underwent a key aldol condensation step with glycine thioester isonitrile to generate a diastereomeric mixture with the desired L-*altro*-thioester isolated as a major product. Although Li's synthesis of L-Alt-diNAc was quite efficient, the synthesis included a C-C bond-forming step necessitating subsequent separations of the formed diastereomers. To eliminate C-C bond formation, one other strategy was reported using the readily available 9-carbon N-acetylneuraminic acid (Neu5Ac) as a starting material.^{20–24} Although attractive, there were complications associated with this strategy, as it generally entailed stereo- and regioselective conversions at several key stereocenters, resulting in low overall yields. Here we wish to report an improved synthesis of L-Alt-diNAc (5) from L-fucose (6).

Results and discussion

The hexose L-fucose (6) was chosen as an ideal starting material because it is conveniently deoxygenated at the C-6 position and has the same L-configuration at C-5 as the target compound, while the configurations at the remaining three centers, C-2, C-3

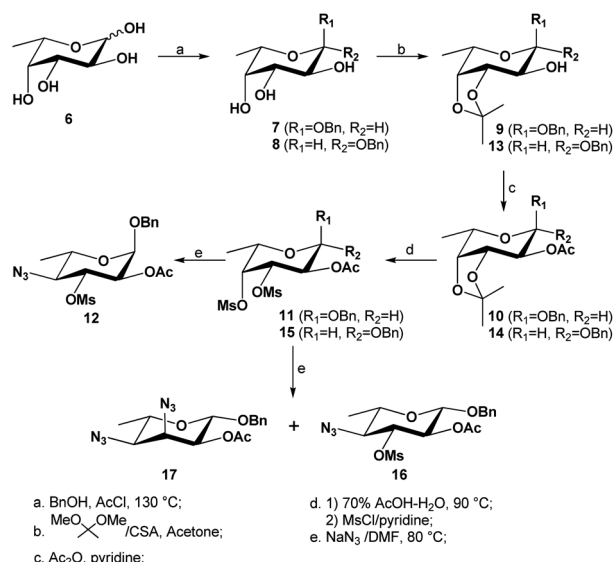
and C-4, could be selectively inverted. This tactic allowed for the introduction an acetamido group for both C-2 and C-4 positions, which could be done *via* nucleophilic substitution using a nitrogen-based nucleophile such as azide. Unlike Liav and Sharon's approach, where a β -L-talopyranoside intermediate that simultaneously contained a 2,3-anhydro and 4-O-mesyl group was utilized, our strategy took advantage of the significant difference in reactivity between secondary sulfonyl esters in a polysulfonylated pyranoside during a nucleophilic substitution. Previously, McGeary *et al.* reported^{25,26} that nucleophiles such as benzoate or azide could be used to regioselectively displace the 4-tosylate from 2,3,4-tritosylated pyranosides of L-arabinose, D-ribose, D-lyxose and D-xylose. As shown in Scheme 1, our strategy involved using a di-O-mesylated substrate such as the benzyl 2-O-acetyl-3,4-di-O-mesyl- α -L-fucopyranoside (11) as the key substrate for the reaction with sodium azide; this was predicted to afford regioselectively the C-4-monosubstituted product (12), which would then undergo further transformations to form a 2,3-anhydro-4-azido- α -L-allopyranoside (25) that would advantageously invert the configuration at C-3 center. Following the Fürst-Plattner rules, another ring opening of the 2,3-anhydro functionality by an azide nucleophile was expected to regioselectively yield the desired 2,4-diazido-2,4,6-trideoxy- α -L-altropyranoside (26). Obtaining the final compound (5) was devised by a reduction of both azides, followed by an *in situ* N-acetylation and lastly a deprotection of the anomeric benzyl group.

Our synthetic route to compound 12 from L-fucose (6) is presented in Scheme 2. The synthesis of the 2-O-acetyl-3,4-O-isopropylidened intermediate 10²⁷ and its β -anomeric glycoside 14¹⁷ had previously been reported, and so we focused on optimizing the experimental procedures to allow the synthesis in a multigram scale. Starting with L-fucose 6 (30 g), a Fischer glycosylation (90 °C, 24 hours) with benzyl alcohol (150 mL, 5 mL g⁻¹ L-fucose) in the presence of HCl (generated *in situ* by adding acetyl chloride to the benzyl alcohol) gave the desired α -isomer 7 as a majority due to anomeric effect. The benzyl α -fucopyranoside (7) was isolated in almost pure form by a simple precipitation in hexanes (65% yield, contaminated with <5% of β -anomer 8). The pure α -isomer 7 was isolated by a recrystallization in ethyl acetate and hexanes, and its 3,4-*cis*-diol was regioselectively protected *via* an isopropylidene to afford compound 9 in excellent yield (93%). After a simple acetylation, compound 10 was isolated in quantitative yield. Alternatively,



Scheme 1 Retrosynthesis of L-Alt-diNAc (5).





Scheme 2 Synthesis of 3,4-di-O-mesylated precursors **11** and **15** and their subsequent substitutions with azide nucleophile.

the crude α/β -fucopyranoside (**7** and **8**) mixture obtained from the initial precipitation could be used directly for the 3,4-O-isopropylideneation, followed by an O-2-acetylation and purification by column chromatography, affording **10** in pure form in 34% yield over 3 steps from L-fucose (**6**). Although the above methods were suitable for preparing the α -anomer **10**, it was found that most of the formed β -fucopyranoside precursor **8** was lost during the post-Fisher glycosylation precipitation. In order to obtain the analogous β -anomer **14**, we developed a one-pot synthesis from **6** by carrying out a Fisher glycosylation using a much reduced amount of benzyl alcohol (2 mL g^{-1} L-fucose) and a trace amount of *p*-toluenesulfonic acid as catalyst. After heating to 130 – 140°C with azeotropic removal of water, the reaction mixture was cooled to room temperature; then, instead of carrying out a precipitation, the reaction mixture was directly subjected to an *in situ* 3,4-O-isopropylideneation with an excess amount of 2,2-dimethoxypropane. The crude reaction mixture was concentrated and acetylated using acetic anhydride and pyridine. After a purification on column chromatography on silica gel, the desired α -glycoside **10** and its β -anomer **14** were isolated in 48% and 20% yield respectively, over three steps from L-fucose. Reducing the amount of benzyl alcohol in our improved protocols thus allowed for a higher-yielding one-pot synthesis of both α/β -fucopyranosides (**10** and **14**).

Deprotection of the isopropylidene from the α -fucopyranoside **10** was achieved in 85% aqueous acetic acid at 80°C , exposing the two hydroxyl groups at C-3 and C-4 centers (Scheme 2). To avoid possible O-deacetylation at C-2, the hydrolysis was carefully monitored and stopped once the desired product was formed (~ 2 hours). The free 3,4-O-diol intermediate was not isolated but subjected to a dimesylation without difficulty, using an excess amount of methanesulfonyl chloride (2.5 equivalents per OH group) in pyridine, giving compound **11** in 90% yield over two steps. The structure of the

3,4-O-dimesylate **11** was confirmed with the appearance of two distinct singlets at 3.12 and 3.19 ppm, integrated to three protons each ($2 \times \text{Ms's}$). The β -glycoside **14** was likewise hydrolyzed, and dimesylated afterwards as above, to afford the corresponding 3,4-di-O-mesylated β -fucopyranoside **15**.

As shown in Scheme 2, when compound **11** was heated in DMF with an excess amount of sodium azide (~ 5.2 equivalents) at 90°C for 40 hours and subsequently at 100°C for 4 hours, the monosubstituted product **12** was obtained in almost quantitative yield; despite the use of a large excess of nucleophile, no disubstituted product was isolated. This azide substitution was carried out in a ~ 20 gram scale without losing the selectivity nor the productivity (98% yield). The 4-O-mesylate of compound **11** was confirmed to have been selectively displaced by ^1H NMR spectrum, because according to ^1H - ^1H COSY spectrum, H-4 was found to shift upfield from 5.04 ppm (compound **11**) to 3.37 ppm (compound **12**); furthermore, the coupling pattern of H-4 of formed product **12** was observed to be a doublet of doublets with two large coupling constants (9.7, 9.9 Hz), confirming that the C-4 center had an axial hydrogen and equatorial azide. Interestingly, when the analogous 3,4-di-O-mesylate of the β -fucopyranoside (**15**) was heated to 90°C for 40 hours, two products were obtained after a purification by column chromatography on silica gel: the major was found to be monosubstituted (60% yield), and the minor was found to be disubstituted (29% yield). The structure of major product was confirmed by ^1H NMR spectrum to be the 4-monosubstituted product **16**, as one mesylate was still present and determined to attach to C-3 center; furthermore, the H-4 resonance was found to have shifted upfield to ~ 3.35 ppm, confirming the regioselective displacement of the O-4-mesylate with an azide. The structure of the minor compound was determined to be the 3,4-diazide **17**, as no mesylate signal was observed in the ^1H NMR spectrum of the compound, and both H-3 (4.34 ppm) and H-4 (3.23 ppm, $J = 3.1, 9.4 \text{ Hz}$) were observed to have shifted upfield.

It was interesting to observe a significant difference in regioselectivity during azide substitutions between the 3,4-di-O-mesylated α/β -fucopyranoside (**11** and **15**). For the α -fucopyranoside **11**, the substitution of the 3-O-mesylate was completely

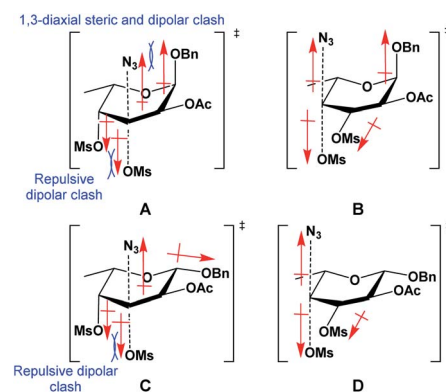


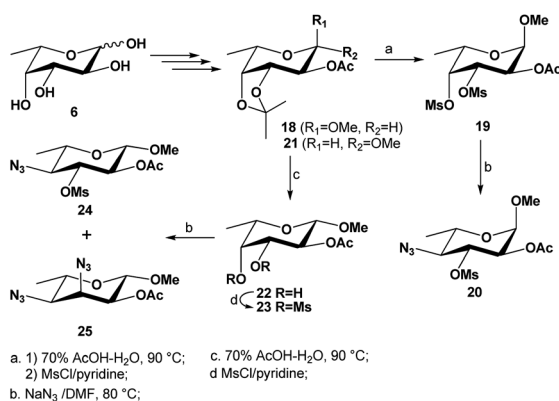
Fig. 2 Transition states formed from the $\text{S}_{\text{N}}2$ displacement of 3- and 4-mesylates of α -fucopyranoside **11** (structures A and B) and β -fucopyranoside **15** (structures C and D), by an azide.



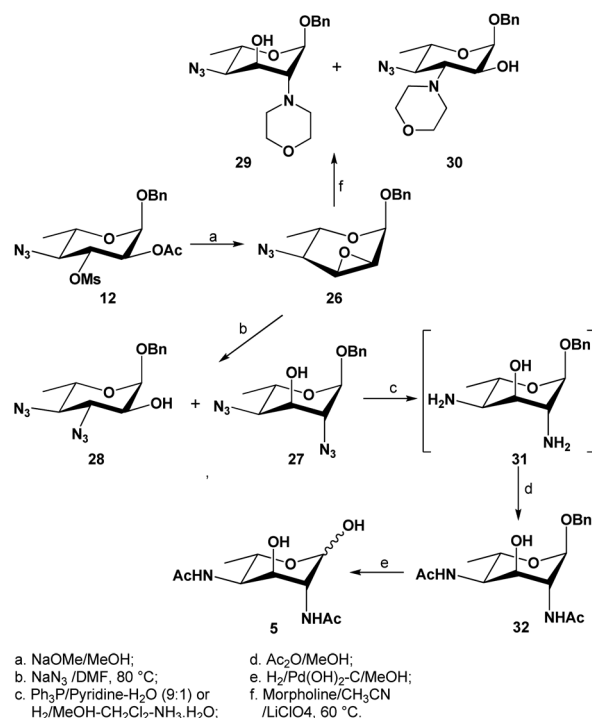
inhibited, while for the β -fucopyranoside **15**, the 3-*O*-mesylate was partially exchanged. This phenomenon was attributed to the dipole-dipole interactions (Fig. 2) explained the Richardson-Hough rules,²⁸ recently updated by Hale *et al.*²⁹

As depicted in Fig. 2, for α -fucopyranoside **11**, the S_N2 displacement of mesylates on pyranosides by azide undergoes two possible transition states (TS); the attack on the 3-*O*-sulfonyl (structure **A**) causes a repulsive alignment between the dipole of the partially bonded departing C-3...OMs and the permanent dipole of axial C-4...OMs, in addition to a 1,3-diaxial dipole clash between the C-1...OBn dipole and the partially bonded incoming C-3...N₃ dipole, making the substitution of C-3-mesylate difficult. On the other hand, the S_N2 substitution of 4-*O*-mesylate can be illustrated by a TS (structure **B**) with misaligned dipoles between the C-3 and C-4 mesylates, where no hindering repulsion effect is present, making the S_N2 displacement of 4-mesylate favorable. In the case of S_N2 attack on the 3-*O*- and 4-*O*-mesylates of β -fucopyranoside **15**, the unfavorable dipole interactions for the substitutions at both positions are markedly reduced in the TS. The attack on 3-*O*-mesylate (structure **C**) is thus made more likely by the elimination of the 1,3-diaxial clash with the incoming C-3...N₃ dipole, while the bottom-face dipole repulsions remain, and in structure **D**, there are no dipole alignments whatsoever in the TS: this explains why both compounds **16** and **17** were formed, but in different yields.

To illustrate the influence of size of aglycone on the nucleophilic substitutions at C-3 and C-4 centers, we also prepared the 3,4-dimesylates of both α/β -anomers of methyl *L*-fucopyranosides **19**³⁰ and **22** from *L*-fucose **6** (Scheme 3). As can be seen, when the same reaction was performed on the methyl 2-*O*-acetyl-3,4-di-*O*-mesyl α -*L*-fucopyranoside **19**, the monosubstituted product **20** was isolated in 68% yield; no 3,4-disubstituted product was observed. However, when the same reaction was carried out with the corresponding 3,4-dimesylate **23** of β -*L*-fucopyranoside, the monosubstituted product **24** was isolated in 24% yield while the 3,4-disubstituted product **25** was isolated in majority (52% yield). These results confirm that both the steric hindrance and strong dipole/dipole repulsions in the TS play a significant role in this type of reaction.

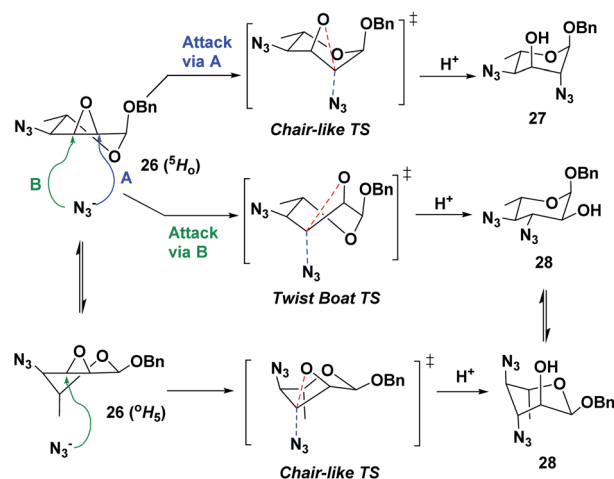


Scheme 3 Synthesis of 3,4-di-*O*-mesylated precursors **19** and **23** and their subsequent substitutions with azide nucleophile.



Scheme 4 Base-mediated epoxidation of compound **12**, subsequent ring opening with nucleophiles and final conversion of 2,4-diazido- α -*L*-altropyranoside **26** to *L*-Alt-diAc **5**.

With large amounts of benzyl 2-*O*-acetyl-4-azido-4,6-dideoxy-3-*O*-mesyl- α -*L*-fucopyranoside **12** at our disposal, a regio- and stereo-selective epoxidation was elegantly performed *via* transesterification in anhydrous methanol in the presence of sodium methoxide to remove the *O*-2-acetyl group, followed by an *in situ* intramolecular ring closure to displace the 3-*O*-mesylate (Scheme 4): the configuration at the C-3 center was thus inverted. The oxirane formation was found to be much slower than expected at room temperature, which required overnight



Scheme 5 Enhanced flexibility between the ⁵H₀ and ⁰H₅ conformations of compound **26** results in poor regio- and stereo-selectivity during azide-mediated opening of the epoxide.



Table 1 Epoxide opening optimizations

Entry	Amount of 26	Solvent	NaN ₃ equivalents	Catalyst	Temp/time	Ratio ^a 27 : 28	Yield ^a
1	2.30 g	DMF	5	—	90 °C, 24 hours	4 : 3	<50% ^b
2	20 mg	EtOH (95%)	4	—	90 °C, 2 days	1 : 1	50%
3	20 mg	DMF	4	NH ₄ Cl (3.5 eq.)	90 °C, 2 days	3 : 2	55%
4	20 mg	EtOH	8	CeCl ₃ (2 eq.)	85 °C, 2 days	4 : 3	25%
5	20 mg	MeCN	2	CeCl ₃ (2 eq.)	85 °C, 2 days	5 : 3	32%
6	20 mg	MeCN	8	Mg(ClO ₄) ₂ (2 eq.)	85 °C, 2 days	—	—
7	20 mg	MeCN–H ₂ O (9 : 1)	8	Fe(ClO ₄) ₃ ·H ₂ O (2 eq.)	85 °C, 2 days	—	—
8	100 mg	MeCN	4	LiCF ₃ SO ₃ (2 eq.)	85 °C, 2 days	5 : 1	70%
9	97 mg	MeCN	5	LiCF ₃ SO ₃ (6 eq.)	80 °C, 2 days	5 : 1	77% ^b
10	20 mg	MeCN	4	LiClO ₄ (2 eq.)	90 °C, 2 days	3 : 1	70%
11	100 mg	MeCN	4	LiClO ₄ (2 eq.)	85 °C, 2 days	5 : 1	80%
12	1.0 g	MeCN	4	LiClO ₄ (3 eq.)	85 °C, 2 days	5 : 1	62% ^b
13	1.0 g	MeCN	4	LiClO ₄ (4 eq.)	85 °C, 2 days	5 : 1	70% ^b
14	20 mg	EtCN	4	LiClO ₄ (4 eq.)	100 °C, 1 day	4 : 1	60%
15	20 mg	MeCN	4	LiClO ₄ (2 eq.)/12-crown-4	85 °C, 2 days	4 : 1	75%
16	21 mg	MeCN	4	LiClO ₄ (2 eq.)/15-crown-5	90 °C, 2 days	3 : 1	60%
17	21 mg	MeCN	4	LiClO ₄ (2 eq.)/TBACl (10 mol%)	90 °C, 2 days	5 : 1	80%
18	5.53 g	MeCN	4	LiClO ₄ (4 eq.)	85 °C, 5 days	5 : 1 ^c	62% ^b

^a Measured by integration in crude ¹H NMR spectra, comparing integration of anomeric peaks, unless specified otherwise. ^b Actual yield % of desired regioisomer (**27**) from products purified by silica gel column chromatography. ^c Measured by TLC.

stirring (~20 hours) to complete, while heating to 55 °C gave the desired compound **26** in large amounts (>10 g) and in good yields (89%) within a few hours. The inversion was confirmed by ¹H NMR spectrum through the change in coupling constants of the H-3 signal (doublet of doublets), from two large *J* coupling constants to two small *J* coupling constants (*J* = 1.6, 4.1 Hz, H-3); furthermore, both H-2 and H-3 signals were observed to exhibit upfield shifts at 3.49 ppm and 3.55 ppm, respectively.

Following the successful synthesis of 4-azido-*l*-allo-epoxide **26**, we proceeded to investigate the oxirane ring opening using sodium azide as the nucleophile (Scheme 4). It was anticipated that the desired 2,4-diazido- α -*l*-altropyranoside (**27**) would be the major isomer from a *trans*-diaxial opening following the well-established Fürst-Plattner rules, where the undesired 3,4-diazido- α -*l*-glucopyranoside (**28**) would be formed in minority due to its unfavorable boat-like TS (Scheme 5).³¹ However, when the reaction was performed under typical conditions, with 5 equivalents of sodium azide in anhydrous DMF at 90 °C, both compounds **27** and **28** were formed with very poor selectivity (**27/28**: 4 : 3, entry 1 in Table 1).³² A change of solvent to ethanol did not increase selectivity, despite an anticipated activation by the media engaging in hydrogen-bonding with the oxirane ring (entry 2). These results are in sharp contrast with those obtained with conformationally restricted pyranosyl substrates that bear a fused 4,6-*O*-benzylidene ring: in those cases, excellent regio and stereoselectivity are normally achieved.^{33–38} In our case, compound **26** is a non-rigid system that undergoes rapid conformational exchanges between the ⁰H₅ and ⁵H₀ conformations, especially at higher temperatures, giving the observed results.

The lack of the C-6 hydroxy handle that allows for the formation of a rigid bicyclic system prompted us to investigate other approaches to address our challenge. It has been reported

that using either a Brønsted^{39,40} or a Lewis acid^{41–47} to coordinate with the ring oxygen of the epoxide could improve reactivity and selectivity in the desired ring opening reaction. We believed that a strong coordination with the oxirane would slow down ring flipping, while the added polarization would likely make the molecule preferentially adopt the ⁵H₀ conformation, thus cancelling its dipole moments (Fig. 3).

It has been reported that ammonium chloride^{39,40} could assist the ring opening of a 2-acetamido-3,4-anhydro- β -*D*-allo-pyranoside, providing the *trans*-diaxially opened product in >70% yield.³⁹ Our attempt to use this catalyst to promote the ring opening of compound **26** in DMF, however resulted in a very slight increase in selectivity (**27/28**: 3 : 2, entry 3 in Table 1). Trimethylsilyl azide has been used in the synthesis of oseltamivir,⁴⁸ and others have successfully used cerium(III) chloride,⁴³ magnesium(II) perchlorate,⁴⁷ zinc(II) perchlorate,^{41,42} ferric perchlorate⁴⁴ and Amberlyst 15 (ref. 49) for the regiochemical control of oxirane openings with amino- or azido-nucleophiles; we have tried some of these metal-catalysed conditions (entries 4–7), with little success. Fortunately, there have also been reports of soluble lithium salts such as lithium perchlorate^{45,46} that improved the regioselective opening of epoxides in carbohydrates; we therefore investigated the oxirane opening of compound **26** using either lithium triflate (2 or 6

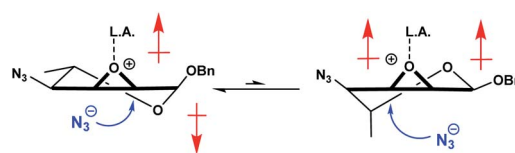


Fig. 3 Coordination of the oxirane could lead to reduced conformational exchange, favoring the ⁵H₀ conformer due to cancellation of dipole moments.



equiv., entries 8–9) or lithium perchlorate (entries 10–14) in either acetonitrile (entries 8–13) or propionitrile (entry 14) at 85–100 °C. To our delight, we observed a significant improvement in selectivity in these trials, with the best ratio of 27/28 to be 5 : 1 (entries 8–9, 11–13), with a much-improved yield as well (60–80% of the desired altropyranoside). While the change of solvent from acetonitrile to propionitrile improved reaction time, because the reaction in the latter was carried out at a higher temperature, it did not help with regioselectivity. We also found that using either lithium triflate or lithium perchlorate gave similar results. The addition of crown ethers such as 12-crown-4 (entry 15) or 15-crown-5 (entry 16) decreased selectivity, presumably due to a competitive binding of crown ethers to lithium cation that led to a decreased coordination of lithium to the oxirane: these results confirmed the importance of lithium coordination.

In an attempt to improve reaction times, we performed a trial by adding tetra-*n*-butylammonium chloride (10 mol%) as a phase catalyst (entry 17): the yield was not significantly improved in the 48 hour timeframe, despite an anticipated increase of solubility of the sodium azide. We concluded that the best condition to obtain compound 27 from compound 26 was to use 4 equivalents of sodium azide and 4 equivalents of lithium perchlorate as an additive in anhydrous acetonitrile at 85 °C. Indeed, under such optimized conditions, we were able to carry out a large-scale conversion of compound 26 (5.53 g) to compound 27 in 62% yield. Another oxirane opening of compound 26 was performed with morpholine as a nitrogen-based nucleophile using similar conditions; the reaction afforded the 2-morpholino-substituted α -L-altropyranoside 29 (63% yield) and the 3-morpholino-substituted α -L-glucopyranoside 30 (15% yield). This result was comparable to the azide-mediated opening of 2,3-epoxide, despite the increased reagent solubility.

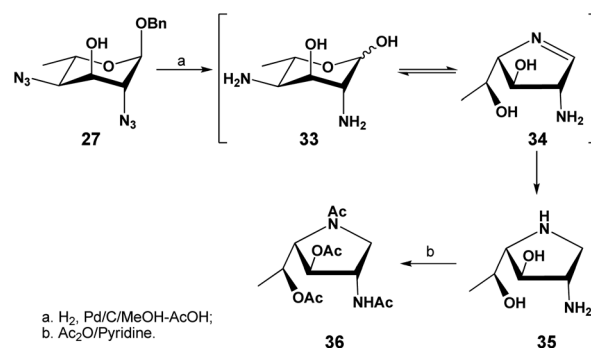
The structures of the two diazido products 27 and 28 were confirmed through ^1H - ^1H gCOSY and ^1H - ^{13}C gHSQC NMR correlation experiments as well as through *O*-acetylation. For compound 27, the ^{13}C signals for the two carbons attached to an azide were observed at 60.8 ppm (C-2) and 61.3 ppm (C-4), while the other carbon with a hydroxyl group attached to was seen at 69.4 ppm (C-3). The shielding effect was also corroborated in the ^1H NMR, where signals for H-2 and H-4 moved significantly upfield (3.85 ppm and 3.24 ppm respectively). Similarly, for compound 28, the ^{13}C signals for the two carbons attached to an azide were observed at 65.8 ppm (C-3) and 66.4 ppm (C-4), while the other carbon with a hydroxyl group attached to was observed at 71.9 ppm (C-2). Again, the shielding effect of the azide groups was seen in the ^1H NMR with H-3 and H-4 shifted upfield (3.58 ppm and 2.91 ppm respectively).

Following the success of synthesizing 2,4-diazido- α -L-altropyranoside 27, the synthesis of 1-2,4-Alt-diNac 5 (Scheme 4) was continued by first reducing the 2,4-diazido functionalities in compound 27. Previously, this transformation had been achieved using lithium aluminium hydride as a reducing reagent, followed by a peracetylation¹⁷ to afford the corresponding fully acetylated product in very poor yields (38%). We thus investigated two alternative methods to achieve the conversion. First, a Staudinger reduction as a mild and chemoselective method to

reduce the two azides was attempted: using triphenylphosphine as a reagent in a mixture of pyridine–water (9 : 1), the two azide functionalities were smoothly reduced at room temperature after 48 hours, and after concentrating the mixture, the crude 2,4-diamine (31) was chemoselectively *N*-acetylated in methanol to give the 2,4-di-*N*-acetylated compound 32 in 59% yield over two steps. The second method involved a controlled hydrogenolysis, where a few drops of ammonia were added as a mild poison against the palladium on charcoal catalyst to reduce its reactivity towards the *O*-benzyl group,⁵⁰ thus selectively reducing the two azide functionalities. Indeed, when compound 27 was subjected to the latter conditions in a mixture of methanol-dichloromethane, the desired selectivity was achieved to afford crude diamine 31 which was further *N*-acetylated as above. This last method afforded the desired compound 32 in much better yield (81%). The formation of 32 was confirmed with the obvious appearance of the two doublets at 6.01 and 6.13 ppm, as well as the presence of two overlapping singlets at 2.02 ppm, attributed to the two NHAc protons. Finally, the anomeric benzyl group was removed using standard catalytic hydrogenolysis conditions to afford the targeted 1-2,4-Alt-diNac 5 in quantitative yield. ^1H NMR spectrum of compound 5 showed the presence of both α - and β -pyranosyl forms. The electrospray high resolution mass spectrometry also confirmed the identity of 1-2,4-Alt-diNac 5 (calc'd m/z for $\text{C}_{12}\text{H}_{21}\text{N}_2\text{O}_6$ [$\text{M} + \text{H}$]⁺: 289.1396; found 289.1394).

The successful selective reduction of compound 27 encouraged us to study a complete catalytic hydrogenation of compound 27 in one step, including the *O*-benzyl functionality (Scheme 6). We predicted that the uncontrolled reduction would produce the reactive intermediate 33 that would then self-condense into a 1,4-iminofuranosyl ring (\rightarrow 34), further undergoing a reduction to afford the stable iminofuranoside 35. This was indeed found to be the case when compound 27 was subjected to a palladium-catalyzed hydrogenolysis in methanol in the presence of a few drops of acetic acid. A new polar product, presumably compound 35 was thus formed and after a full acetylation, the iminofuranoside 36 was isolated in 60% yield.

In the ^1H NMR spectrum of the isolated compound, a highly deshielded proton was observed at 6.70 ppm as a doublet, which was assigned to be the single amide proton. 4 singlets



Scheme 6 Conversion of compound 27 to the iminofuranosides 35 and 36 via a complete hydrogenation.



were observed at 2.12, 2.11, 2.08 and 2.03 ppm, corresponding to the 4 acetyl groups present in the molecule. Two highly deshielded protons were observed at 5.45 and 5.35 ppm, attributed to the H-5 and H-3 respectively and confirmed by ^1H - ^1H gCOSY experiment; their large chemical shifts are consistent with an *O*-acetylation on O-5 and O-3, further supporting that O-5 is not involved in cyclization. Two mutually coupled protons (high orders) were observed at 4.24 ppm and 3.19 ppm, attributed to the two diastereotopic protons attached to C-1. These assignments were also confirmed by a ^1H - ^{13}C 2D gHSQC correlation spectrum. The C-1 carbon was observed at a highly shielded region (52.9 ppm), confirming the attachment of C-1 to a less electronegative nitrogen.

Conclusions

In conclusion, we have developed a convenient and readily scalable synthesis toward the challenging 1-2,4-Alt-diNAc (5) from 1-fucose (6) in 10 steps (23% overall yields). Our synthesis has overcome some major bottlenecks in previously reported syntheses.¹⁷ Because of the shorter reaction sequence and much improved transformation in some key steps, our synthesis allows the preparation of key intermediates such as compounds 12, 26, 27 in gram quantities. By further combining with the indium-mediated allylation with ethyl 2-(bromomethyl)acrylate, already attempted by several other groups working in this area, Pse and analogs could be prepared in a straightforward manner.

Experimental section

General methods

All commercial reagents were used as supplied unless otherwise stated. Thin layer chromatography was performed on Silica Gel 60-F254 (E. Merck, Darmstadt) with detection by fluorescence, charring with 5% H_2SO_4 (aq), or a ceric ammonium molybdate solution. Column chromatography was performed on Silica Gel 60 (Silicycle, Ontario) and solvent gradients given refer to stepped gradients and concentrations are reported as % v/v. Organic solutions were concentrated and/or evaporated to dry under vacuum in a water bath (<60 °C). Optical rotations were determined in a 5 cm cell at 20 ± 2 °C; $[\alpha]_D^{20}$ values are given in units of 10^{-1} deg $\text{cm}^2 \text{g}^{-1}$. NMR spectra were recorded on Bruker spectrometers at 400 MHz, and the first-order chemical shifts of ^1H and ^{13}C (DEPT-Q) are reported in δ (ppm) and referenced to residual CHCl_3 (δ_{H} 7.24, δ_{C} 77.23, CDCl_3), or residual HDO (δ_{H} 4.79, external acetone δ_{C} 29.9, D_2O), residual CD_2HOD (δ_{H} 3.31, δ_{C} 49.1, CD_3OD) ^1H and ^{13}C NMR spectra were assigned with the assistance of 2D gCOSY and 2D gHSQC experiments. High-resolution ESI-QTOF mass spectra were recorded on an Agilent 6520 Accurate Mass Quadrupole Time-of-Flight LC/MS spectrometer. All the data were obtained with the assistance of the analytical services of the Department of Chemistry, University of Calgary.

Benzyl α -1-fucopyranoside (7). Acetyl chloride (~5.0 mL) was added to benzyl alcohol (150 mL) at 0 °C and the solution was stirred for 10 minutes. 1-fucose 6 (30.0 g, 183 mmol) was then added into the solution, and the mixture was heated to 90 °C for

24 hours. Cyclohexane (100 mL) was added to the solution, and the solution was distilled off to remove the water azeotrope under reduced pressure at 90 °C. The solution was then poured into hexanes (3.5 L) with vigorous stirring to afford precipitate. Vacuum filtration afforded an off-white powder which consists of a mixture of α/β -anomers (α/β : 95/5; 30.21 g, 65% yield). Some pure α -anomer (7) was isolated by recrystallization of the crude product mixture (~20 g) in ethyl acetate and hexanes (400 mL) to afford white crystals. R_f 0.47 (10% $\text{MeOH}-\text{CH}_2\text{Cl}_2$). ^1H NMR (400 MHz, CD_3OD): δ_{H} 7.44–7.26 (m, 5H, ArH), 4.87 (d, 1H, J = 3.3 Hz, H-1), 4.71 (d, 1H, J = 12.0 Hz, $\text{CH}_a\text{H}_b\text{Ph}$), 4.58 (d, 1H, J = 12.0 Hz, $\text{CH}_a\text{H}_b\text{Ph}$), 3.97 (dq, 1H, J = 6.6, 0.7 Hz, H-5), 3.80 (dd, 1H, J = 10.1, 3.0 Hz, H-3), 3.76 (dd, 1H, J = 10.1, 3.4 Hz, H-2), 3.68 (m, 1H, H-4), 1.20 (d, 3H, J = 6.6 Hz, H-6). ^{13}C NMR data are in agreement with previously reported.⁵¹

Benzyl 3,4-*O*-isopropylidene- α -1-fucopyranoside (9) and benzyl 3,4-*O*-isopropylidene- β -1-fucopyranoside (13)

Method 1. To a mixture of benzyl α -1-fucopyranoside 7 (5.27 g, 20.7 mmol) in acetone (30 mL), was added 2,2-dimethoxypropane (10.0 mL), and 10-camphorsulfonic acid (CSA, 120 mg), and the mixture was heated to reflux for 0.5 h. The mixture was then neutralized with triethylamine (~2 mL) and concentrated under reduced pressure to afford a yellow oil. The mixture was dissolved in a 10 : 1 mixture of methanol and water (220 mL) and heated to reflux for one hour, and concentrated under reduced pressure. The mixture was dissolved in ethyl acetate (50 mL), and the obtained organic solution was washed successively with brine (50 mL), 2 N HCl (50 mL) and saturated aqueous NaHCO_3 (50 mL), dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The crude residue was purified by column chromatography on silica gel using a gradient of 5 \rightarrow 10% ethyl acetate in hexanes as an eluent, to yield the desired compound (9) as a colourless syrup (5.70 g, 93% yield). ^1H NMR (400 MHz, CDCl_3): δ_{H} 7.41–7.30 (m, 5H, ArH), 4.95 (d, 1H, J = 3.9 Hz, H-1), 4.80 (d, 1H, J = 11.8 Hz, $\text{CH}_a\text{H}_b\text{Ph}$), 4.59 (d, 1H, J = 11.8 Hz, $\text{CH}_a\text{H}_b\text{Ph}$), 4.24 (t, 1H, J = 6.2 Hz, H-3), 4.17 (dq, 1H, J = 6.6 Hz, J = 2.3 Hz, H-5), 4.07 (dd, 1H, J = 6.1 Hz, J = 2.3 Hz, H-4), 3.84 (dt, 1H, J = 6.7 Hz, J = 4.0 Hz, H-2), 2.28 (d, 1H, J = 6.9 Hz, OH), 1.53 (s, 3H, Me_{ISP}), 1.37 (s, 3H, Me_{ISP}), 1.32 (d, 3H, J = 6.6 Hz, H-6). ^{13}C NMR data are in agreement with previously reported.²⁷

Method 2. To a mixture of the crude precipitate that contained the benzyl α/β -1-fucopyranosides (7/8, 18.2 g, 62.6 mmol) in acetone (50.0 mL) and 2,2-dimethoxypropane (50 mL), was added CSA (200 mg), and the solution was heated to reflux for 24 hours. The mixture was then neutralized with triethylamine, and solution was concentrated under reduced pressure. The yellow syrup was purified by column chromatography on silica gel using a gradient of 5 \rightarrow 10% ethyl acetate–hexanes as an eluent to yield the pure β -fucopyranoside 13 (3.89 g, 20% yield) and also the α -fucopyranoside 9 (7.17 g, 37% yield), as colourless syrups. Data for compound 13: R_f 0.39 (40% ethyl acetate–hexanes). ^1H NMR (400 MHz, CDCl_3): δ_{H} 7.41–7.23 (m, 5H, ArH), 4.93 (d, J = 11.6 Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 4.58 (d, J = 11.7 Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 4.22 (d, J = 8.3 Hz, 1H, H-1), 4.01 (dd, J = 7.2, 5.5 Hz, 1H, H-3), 3.97 (dd, J = 5.5, 2.2 Hz, 1H, H-4), 3.82 (qd, J = 6.6, 2.2 Hz, 1H, H-5), 3.58 (ddd, J = 8.2, 7.2, 2.5 Hz, 1H, H-2), 2.80 (d,

$J = 2.6$ Hz, 1H, OH-2), 1.53 (s, 3H, CH₃-ISP), 1.44 (d, $J = 6.6$ Hz, 3H, H-6), 1.35 (s, 3H, CH₃-ISP). ¹³C NMR data are in agreement with previously reported.

Benzyl 2-O-acetyl-3,4-O-isopropylidene- α -L-fucopyranoside (10) and benzyl 2-O-acetyl-3,4-O-isopropylidene- β -L-fucopyranoside (14). To a solution of L-fucose **6** (11.0 g, 67.0 mmol) in benzyl alcohol (22 mL), was added *p*-toluenesulfonic acid (0.5 g), and the mixture was stirred and heated to 130 °C for 30 min. Toluene (10 mL) was added and the reaction flask was connected to a Dean-Stark. The temperature was raised to 140 °C to allow azeotropic removal of toluene–water mixture. More toluene (3 × 10 mL) was repeatedly added to reaction mixture, and each time the toluene–water azeotrope was removed. The reaction was cooled to room temperature. To the above reaction mixture, was added 2,2-dimethoxypropane (25 mL, 204 mmol), and the reaction was continued overnight. The solution was concentrated under reduced pressure. Anhydrous pyridine (80 mL) was added to the reaction followed by acetic anhydride (80 mL), and the mixture was heated to 70 °C for 4 hours. The reaction solution was then concentrated under reduced pressure, and co-evaporated with toluene (2 × 100 mL). The mixture was diluted with ethyl acetate (~300 mL), and the organic solution was washed successively with a solution of aqueous 2 N HCl (1 × 100 mL), saturated aqueous NaHCO₃ (1 × 100 mL), 10% brine (1 × 100 mL), dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The mixture was purified by column chromatography on silica gel using a gradient of ethyl acetate–hexanes (0 → 12.5%) to afford the desired α -anomer **10** (10.92 g, 48% over 3 steps), and β -anomer **14** (4.54 g, 20% over 3 steps). Both products can also be recrystallized from hexanes. Data for compound **10**: R_f 0.40 (20% EtOAc in hexanes). [α]_D²⁵ –155.5 (c 1.2, MeOH). ¹H NMR (400 MHz, CDCl₃): δ_H 7.41–7.25 (m, 5H, ArH), 5.00 (d, $J = 3.6$ Hz, 1H, H-1), 4.92 (dd, $J = 8.2$, 3.6 Hz, 1H, H-2), 4.72 (d, $J = 12.3$ Hz, 1H, CH_aH_bPh), 4.53 (d, $J = 12.3$ Hz, 1H, CH_aH_bPh), 4.37 (dd, $J = 8.3$, 5.3 Hz, 1H, H-3), 4.18 (dq, $J = 6.7$, 2.6 Hz, 1H, H-5), 4.10 (dd, $J = 5.4$, 2.6 Hz, 1H, H-4), 2.10 (s, 3H, Ac), 1.54 (s, 3H, CH₃-ISP), 1.37 (d, $J = 6.7$ Hz, 3H, H-6), 1.36 (s, 3H, CH₃-ISP). ¹³C NMR (101 MHz, CDCl₃): δ_C 170.5 (Ac), 137.3 (ArC), 128.4 (ArC), 127.9 (ArC), 127.6 (ArC), 109.3 (C(CH₃)₂), 95.4 (C-1), 76.13 (C-4), 73.38 (C-3), 71.94 (C-2), 69.6 (OCH_aH_bPh), 63.4 (C-5), 26.39 (CH₃-ISP), 20.97 (Ac), 16.25 (C-6). HRMS (ESI-QTOF) calc'd m/z for C₁₈H₂₄O₆ [M + Na]⁺: 359.1465; found: 359.1452. Data for compound **14**: R_f 0.20 (20% EtOAc in hexanes). ¹H NMR (400 MHz, CDCl₃): δ_H 7.42–7.24 (m, 5H, ArH), 5.06 (dd, $J = 8.3$, 7.6 Hz, 1H, H-2), 4.91 (d, $J = 12.5$ Hz, 1H, CH_aH_bPh), 4.62 (d, $J = 12.5$ Hz, 1H, CH_a-H_bPh), 4.37 (d, $J = 8.4$ Hz, 1H, H-1), 4.13 (dd, $J = 7.6$, 5.3 Hz, 1H, H-3), 4.03 (dd, $J = 5.3$, 2.1 Hz, 1H, H-4), 3.86 (dq, $J = 6.6$, 2.2 Hz, 1H, H-5), 2.09 (s, 3H, Ac), 1.60 (s, 3H, CH₃-ISP), 1.47 (d, $J = 6.6$ Hz, 3H, H-6), 1.37 (s, 3H, CH₃-ISP).

Benzyl 2-O-acetyl-3,4-di-O-methanesulfonyl- α -L-fucopyranoside (11). A solution of compound **10** (17.2 g, 51.2 mmol) in 85% acetic acid–water (330 mL) was heated to 80 °C. After 2 hours, the solution was concentrated and co-evaporated with toluene (3 × 150 mL) to give an off-white solid (15.1 g, ~100% yield). The hydrolyzed diol was dissolved in anhydrous dichloromethane (100 mL) and pyridine (50 mL) and was cooled to 0 °C;

mesyl chloride (20.0 mL, 0.26 mol) was added, and the reaction was allowed to warm up to room temperature. After stirring for 20 hours, water (20 mL) was added to quench the reaction, and the mixture was diluted with ethyl acetate (200 mL), and washed with 2 N HCl (2 × 100 mL), saturated aqueous NaHCO₃ (100 mL) and brine (100 mL), dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The crude mixture was purified by column chromatography on silica gel using 25% ethyl acetate–hexanes as an eluent to afford compound **11** as a white solid (20.74 g, 90% yield). R_f 0.23 (40% EtOAc–hexanes). [α]_D²⁵ –119.3 (c 0.63, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ_H 7.40–7.28 (m, 5H, ArH), 5.17–5.07 (m, 3H, H-1, H-2, H-3), 5.04 (dd, $J = 1.2$, 3.2 Hz, 1H, H-4), 4.70 (d, 1H, $J = 12.2$ Hz, CH_aH_bPh), 4.57 (d, 1H, $J = 12.2$ Hz, CH_aH_bPh), 4.16 (dq, 1H, $J = 6.5$, 0.8 Hz, H-5), 3.19 (s, 3H, Ms), 3.12 (s, 3H, Ms), 2.09 (s, 3H, OAc), 1.29 (d, 3H, $J = 6.5$ Hz, H-6). ¹³C NMR (101 MHz, CDCl₃): δ_C 169.7 (Ac), 136.6 (ArC), 128.6 (ArC), 128.2 (ArC), 128.0 (ArC), 95.3 (C-1), 80.6 (C-4), 74.4 (C-3), 70.3 (CH₂Ph), 67.7 (C-2), 64.8 (C-5), 39.0 (Ms), 38.6 (Ms), 20.7 (Ac), 16.3 (C-6). HRMS (ESI-QTOF) calc'd m/z for C₁₇H₂₄O₁₀S₂ [M + Na]⁺: 475.0709; found: 475.0703.

Benzyl 2-O-acetyl-4-azido-4,6-dideoxy-3-O-mesyl- α -L-glucopyranoside (12). To a mixture of 3,4-di-O-mesylate **11** (20.74 g, 45.8 mmol) in anhydrous DMF (200 mL), was added NaN₃ (15.53 g, 0.24 mol), and the mixture was heated to 90 °C for 40 hours. The temperature was raised to 100 °C for additional 4 hours. The mixture was concentrated under reduced pressure, and the residue was partitioned between 10% brine (400 mL) and ethyl acetate (300 mL); the aqueous layer was separated and re-extracted with more EtOAc (200 mL). The combined organic solution was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The crude mixture was purified by column chromatography on silica gel using 5% ethyl acetate–hexanes as an eluent to afford **12** as a white solid (17.85 g, 98% yield). R_f 0.53 (40% EtOAc–hexanes). [α]_D²⁵ –172.0 (c 4.6, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ_H 7.61–7.37 (m, 5H, ArH), 5.16 (d, $J = 3.6$ Hz, 1H, H-1), 5.14 (dd, $J = 9.8$, 9.8 Hz, 1H, H-3), 5.03 (dd, $J = 10.0$, 3.7 Hz, 1H, H-2), 4.83 (d, $J = 12.4$ Hz, 1H, CH_aH_bPh), 4.67 (d, $J = 12.4$ Hz, 1H, CH_aH_bPh), 3.90 (dq, $J = 9.9$, 6.2 Hz, 1H, H-5), 3.37 (dd, $J = 9.8$, 9.8 Hz, 1H, H-4), 3.26 (s, 3H, Ms), 2.24 (s, 3H, Ac), 1.48 (d, $J = 6.3$ Hz, 3H, H-6). ¹³C NMR (101 MHz, CDCl₃): δ_C 170.2 (Ac), 136.8 (ArC), 128.6 (ArC), 128.2 (ArC), 128.0 (ArC), 95.3 (C-1), 78.5 (C-3), 70.6 (C-2), 70.1 (CH_aH_bPh), 66.8 (C-4), 66.5 (C-5), 39.1 (Ms), 20.8 (Ac), 18.3 (C-6). HRMS (ESI-QTOF) calc'd m/z for C₁₈H₂₈O₈N [M + NH₄]⁺: 417.1438; found: 417.1455.

Benzyl 2-O-acetyl-3,4-di-O-mesyl- β -L-fucopyranoside (15). Benzyl 3,4-O-isopropylidene- β -L-fucopyranoside **13** (3.39 g, 11.5 mmol) was acetylated in a mixture of acetic anhydride (4.4 mL, 47 mmol) and pyridine (30 mL) in a similar manner as compound **9**. The crude product **14** was then hydrolysed in 80% acetic acid–water (100 mL) to 90 °C for 4 hours. The reaction mixture was concentrated and co-evaporated with toluene (50 mL) to provide crude diol intermediate which was re-dissolved in a mixture of anhydrous dichloromethane (50 mL) and anhydrous pyridine (25 mL). To this solution, mesyl chloride (5.0 mL, 65 mmol) was added, and the mixture was stirred at room temperature for 20 hours. Methanol (~2 mL) was added to quench the reaction, and the reaction mixture was concentrated



and co-evaporated with toluene (100 mL). The residue was partitioned between EtOAc (100 mL) and 1 N HCl (100 mL), and the aqueous phase was extracted with more EtOAc (2 × 100 mL). The combined organic solution was dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The mixture was purified by recrystallization in 30% ethyl acetate in hexanes to afford the desired compound **15**¹⁷ (2.19 g, 42% over 3 steps). ¹H NMR (400 MHz, CDCl₃): δ_H 7.40–7.26 (m, 5H, ArH), 5.27 (dd, 1H, *J* = 10.3, 7.9 Hz, H-2), 4.98 (dd, 1H, *J* = 3.4, 0.7 Hz, H-4), 4.92 (d, 1H, *J* = 12.3 Hz, CH_aH_bPh), 4.78 (dd, 1H, *J* = 10.3, 3.4 Hz, H-3), 4.63 (d, 1H, *J* = 12.4 Hz, CH_aH_bPh), 4.51 (d, 1H, *J* = 7.9 Hz, H-1), 3.78 (dq, 1H, *J* = 6.4, 0.7 Hz, H-5), 3.21 (s, 3H, Ms), 3.10 (s, 3H, Ms), 2.08 (s, 3H, Ac), 1.43 (d, 3H, *J* = 6.4 Hz, H-6).

Benzyl 2-O-acetyl-4-azido-4,6-dideoxy-3-O-mesyl-β-L-glucopyranoside (16) and benzyl 2-O-acetyl-3,4-diazido-3,4,6-trideoxy-β-L-allopyranoside (17). A mixture of 3,4-dimesylate **15** (1.77 g, 3.9 mmol) and NaN₃ (1.27 g, 20 mmol) in anhydrous DMF (15 mL) was heated to 90 °C for 40 h. The reaction mixture was poured into water (50 mL) and the product extracted into EtOAc (3 × 20 mL). The combined organic solutions were dried over anhydrous Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography using a mixture of 10% ethyl acetate–hexanes as the eluent to afford compound **16** (0.8043 g, 60% yield) as a syrup, and compound **17** (0.4467 g, 29% yield) as a white powder. Data for compound **16**: *R*_f 0.58 (40% ethyl acetate–hexanes). [α]_D²⁵ +27.8 (*c* 0.58, MeOH). ¹H NMR (400 MHz, CDCl₃): δ_H 7.38–7.27 (m, 5H, ArH), 5.08 (dd, 1H, *J* = 9.6 Hz, 8.0 Hz, H-2), 4.89 (d, 1H, *J* = 12.4 Hz, CH_aH_bPh), 4.63 (dd, 1H, *J* = 9.5, 9.5 Hz, H-3), 4.61 (d, 1H, *J* = 12.3 Hz, CH_aH_bPh), 4.46 (d, 1H, *J* = 8.0 Hz, H-1), 3.39–3.28 (m, 2H, H-4 and H-5), 3.11 (s, 3H, Ms), 2.09 (s, 3H, Ac), 1.46 (d, 3H, *J* = 5.8 Hz, H-6). ¹³C NMR (101 MHz, CDCl₃): δ_C 169.5 (Ac), 136.7 (ArC), 128.5 (ArC), 128.0 (ArC), 127.7 (ArC), 98.9 (C-1), 80.1 (C-3), 70.8 (C-2), 70.8 (CH₂Ph), 70.6 (C-5), 66.2 (C-4), 38.9 (Ms), 20.8 (Ac), 18.3 (C-6). HRMS (ESI-QTOF) calc'd *m/z* for C₁₆H₂₅N₄O₇S [M + Na]⁺: 422.0998; found: 422.0783. Data for compound **17**: *R*_f 0.20 (40% ethyl acetate–hexanes). [α]_D²⁵ +12.19 (*c* 2.1, CH₂Cl₂). ¹H NMR (400 MHz, CDCl₃): δ_H 7.38–7.27 (m, 5H, ArH), 4.89 (d, 1H, *J* = 12.1 Hz, CH_aH_bPh), 4.84 (m, 2H, H-1 and H-2), 4.62 (d, 1H, *J* = 12.1 Hz, CH_aH_bPh), 4.34 (dd, 1H, H-3), 3.81 (dq, 1H, *J* = 9.7, 6.2 Hz, H-5), 3.23 (dd, *J* = 9.7, 3.1 Hz, 1H, H-4), 2.14 (s, 3H, Ac), 1.37 (d, *J* = 6.2 Hz, 3H, H-6). ¹³C NMR (101 MHz, CDCl₃): δ_C 169.7 (Ac), 137.1 (ArC), 128.4 (ArC), 127.9 (ArC), 127.6 (ArC), 97.1 (C-1), 71.5 (C-2), 70.9 (CH_aH_bPh), 68.8 (C-5), 63.1 (C-4), 62.3 (C-3), 20.6 (Ac), 18.1 (C-6). HRMS (ESI-QTOF) calc'd *m/z* for C₁₅H₂₂N₇O₄ [M + NH₄]⁺: 364.1728; found: 364.1722.

Methyl 2-O-acetyl-3,4-di-O-mesyl-α-L-fucopyranoside (19). Crude methyl 2-O-acetyl-3,4-O-isopropylidene-α-L-fucopyranoside **18**³⁰ (0.36 g, 1.4 mmol) was suspended in 80% acetic acid in water (5 mL) and heated to 80 °C for 1 h. The mixture was cooled back to room temperature and the solvent co-evaporated with toluene (30 mL) to give a pale yellow oil (0.35 g). A mixture of crude methyl 2-O-acetyl-α-L-fucopyranoside (0.18 g, 0.81 mmol), dichloromethane (1.0 mL), anhydrous pyridine (0.5 mL), and mesyl chloride (0.31 mL, 4.0 mmol) was cooled to 0 °C and allowed to warm to room temperature over a period 4.5 h. The reaction mixture was quenched with methanol (1 mL) and the

solvent co-evaporated with toluene (30 mL). The residue was then purified by column chromatography (30% ethyl acetate in hexanes) to afford **19** in 0.23 g (74%) yield. ¹H NMR (400 MHz, CDCl₃): δ_H 5.13 (dd, 1H, *J* = 10.6 Hz, *J* = 3.1 Hz), 5.09 (dd, 1H, *J* = 10.6 Hz, *J* = 2.8 Hz), 5.04 (dd, 1H, *J* = 3.0 Hz, *J* = <1 Hz), 4.97 (d, 1H, *J* = 3.1 Hz), 4.12 (dq, 1H, *J* = 6.5 Hz, *J* = <1 Hz), 3.40 (s, 3H), 3.21 (s, 3H), 3.12 (s, 3H), 2.15 (s, 3H), 1.34 (d, 3H, *J* = 6.5 Hz). ¹³C NMR matches literature.³⁰

Methyl 2-O-acetyl-4-azido-4,6-dideoxy-3-O-mesyl-α-L-glucopyranoside (20). A mixture of methyl 2-O-acetyl-3,4-di-O-mesyl-α-L-fucopyranoside **19** (97.8 mg, 0.26 mmol), DMF (2 mL), and sodium azide (94 mg, 1.4 mmol) was heated to 100 °C for 16 h. The reaction mixture was poured into brine (10 mL) and the product extracted into ethyl acetate (3 × 10 mL). The organic fractions were combined, dried over anhydrous sodium sulfate, and concentrated under reduced pressure. The residue was purified by column chromatography (15% ethyl acetate in hexanes) on silica gel to obtain compound **20** as a yellow oil (0.0573 g, 68% yield). *R*_f 0.51 (40% ethyl acetate in hexanes). [α]_D²⁵ −139.4 (*c* 1.8, CHCl₃). ¹H NMR (400 MHz, CDCl₃): δ_H 4.99–4.88 (m, 2H, H-2 and H-3), 4.84 (d, 1H, *J* = 3.5 Hz, H-1), 3.70 (dq, 1H, *J* = 10.0 Hz, *J* = 6.2 Hz), 3.38 (s, 3H, OMe), 3.23 (dd, 1H, *J* = 9.7 Hz, *J* = <1 Hz), 3.12 (s, 3H, OMs), 2.15 (s, 3H, OAc), 1.39 (d, 3H, *J* = 6.2 Hz, H-6). ¹³C NMR (101 MHz, CDCl₃): δ_C 170.2 (OAc), 96.9 (C-1), 78.2 (C-2), 70.5 (C-3), 66.6 (C-4), 66.0 (C-5), 55.4 (OMe), 38.9 (SO₂CH₃), 20.8 (COCH₃), 18.2 (C-6). HRMS (ESI-QTOF) calc'd *m/z* for C₁₀H₁₇N₃O₇S [M + Na]⁺: 346.0679; found: 346.0666.

Methyl 2-O-acetyl-β-L-fucopyranoside (22). A mixture of methyl 3,4-O-isopropylidene-β-L-fucopyranoside **21** (1.51 g, 6.9 mmol), acetic anhydride (2.6 mL, 28 mmol) in anhydrous pyridine (15 mL) was stirred at room temperature for 20 h. The reaction was quenched with water (5 mL), poured into 10% aqueous NaHCO₃ (50 mL) and extracted into CHCl₃ (3 × 30 mL). The combined organic solutions were dried over anhydrous Na₂SO₄, and concentrated under reduced pressure to give a crude solid (1.62 g). The crude mixture was dissolved in 80% acetic acid–water (20 mL), and the solution was heated to 75 °C for 3 hours. After cooled back to room temperature, the reaction solution was concentrated under reduced pressure and co-evaporated with toluene (100 mL). The crude mixture was purified by column chromatography on silica gel using a mixture of 60% ethyl acetate–hexanes to afford the desired compound **22** (1.27 g, 83% yield). *R*_f 0.66 (10% MeOH in CH₂Cl₂). [α]_D²⁵ +3.6 (*c* 1.5, MeOH). ¹H NMR (400 MHz, CD₃-COCD₃): δ_H 4.97 (dd, *J* = 9.6, 8.0 Hz, 1H, H-2), 4.29 (d, *J* = 8.0 Hz, 1H, H-1), 3.75–3.61 (m, 3H, H-3, H-4, H-5), 3.39 (s, 3H, -OMe), 2.01 (s, 3H, Ac), 1.28 (d, *J* = 6.5 Hz, 3H, H-6). ¹³C NMR (101 MHz, CD₃COCD₃): δ_C 101.7 (C-1), 72.2, 72.1, 71.7, 70.4 (C-2,3,4,5), 55.2 (OMe), 20.1 (Ac), 15.8 (C-6). HRMS (ESI-QTOF) calc'd *m/z* for C₉H₁₆O₆ [M + Na]⁺: 243.0839; found 243.0831.

Methyl 2-O-acetyl-3,4-O-mesyl-β-L-fucopyranoside (23). To a solution of compound **22** (0.12 g, 0.54 mmol) in a mixture of anhydrous dichloromethane (1.0 mL) and pyridine (0.5 mL), was added mesyl chloride (0.21 mL, 2.7 mmol) at 0 °C. The temperature was allowed to warm up to room temperature. After stirring for 6 hours, the reaction mixture was quenched with

methanol (~1 mL), and the solvent was removed under reduced pressure, and the residue was co-evaporated with toluene (30 mL). The crude mixture was purified by column chromatography on silica gel using 30% ethyl acetate–hexanes as an eluent to afford compound **23** in pure form (0.20 g, 99% yield). R_f 0.14 (40% EtOAc in hexanes). $[\alpha]_D^{25} -17.93$ (c 0.11, CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ_{H} 5.18 (dd, $J = 10.3$, 7.9 Hz, 1H, H-2), 4.99 (dd, $J = 3.4$, 0.9 Hz, 1H, H-4), 4.82 (dd, $J = 10.3$, 3.4 Hz, 1H, H-3), 4.40 (d, $J = 7.9$ Hz, 1H, H-1), 3.80 (dd, $J = 6.4$, 0.9 Hz, 1H, H-5), 3.52 (s, 3H, OMe), 3.20 (s, 3H, Ms), 3.12 (s, 3H, Ms), 2.13 (s, 3H, Ac), 1.41 (d, $J = 6.4$ Hz, 3H, H-6). ^{13}C NMR (101 MHz, CDCl_3): δ_{C} 101.5 (C-1), 78.19 (C-4), 75.87 (C-3), 68.92 (C-5), 68.14 (C-2), 56.93 (OMe), 38.91 (Ms), 38.71 (Ms), 20.83 (Ac), 16.57 (C-6). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_{11}\text{H}_{20}\text{O}_{10}\text{S}_2$ $[\text{M} + \text{Na}]^+$: 399.0396; found: 399.0411.

Methyl 2-O-acetyl-4-azido-4,6-dideoxy-3-O-mesyl- β -l-glucopyranoside (24) and methyl 2-O-acetyl-3,4-diazido-3,4,6-trideoxy- β -l-glucopyranoside (25). A mixture of 3,4-dimesylate **23** (0.24 g, 0.54 mmol) and sodium azide (0.16 g, 2.5 mmol) in DMF (2.5 mL) was heated to 90 °C for 40 h. The solvent was then removed by evaporation *in vacuo*. The residue was extracted into ethyl acetate (20 mL) and washed with brine (3 \times 10 mL). The organic solution was dried over anhydrous Na_2SO_4 , and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel using 5% ethyl acetate–hexanes as the eluent to afford compound **25** as a white solid (76 mg, 0.28 mmol, 52% yield) and compound **24** (4-azido-3-mesyl) as another white solid (34.0 mg, 20% yield). Data for compound **24**: R_f 0.58 (40% EtOAc–hexanes). $[\alpha]_D^{25} -5.14$ (c 2.5, CH_2Cl_2). ^1H NMR (400 MHz, CDCl_3): δ_{H} 5.01 (dd, $J = 9.7$, 7.9 Hz, 1H, H-2), 4.68 (dd, $J = 9.6$, 9.6 Hz, 1H, H-3), 4.35 (d, $J = 8.0$ Hz, 1H, H-1), 3.51 (s, 3H, OMe), 3.39 (dq, $J = 9.8$, 6.0 Hz, 1H, H-5), 3.30 (d, $J = 9.6$, 9.6 Hz, 1H, H-4), 3.14 (s, 3H, Ms), 2.15 (s, 3H, Ac), 1.47 (d, $J = 6.0$ Hz, 3H, H-6). ^{13}C NMR (101 MHz, CDCl_3): δ_{C} 101.2 (C-1), 80.2 (C-3), 70.8 (C-2), 70.6 (C-5), 66.3 (C-4), 57.0 (OMe), 38.94 (Ms), 20.89 (Ac), 18.29 (C-6). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_9\text{H}_{17}\text{O}_7\text{N}_3\text{S}$ $[\text{M} + \text{Na}]^+$: 346.0679; found: 346.0683. Data for compound **25**: R_f 0.15 (10% ethyl acetate–hexanes). $[\alpha]_D^{25} +25.5$ (c 0.80, MeOH). ^1H NMR (400 MHz, CDCl_3): δ_{H} 4.76 (dd, $J = 8.1$, 3.4 Hz, 1H, H-2), 4.67 (d, $J = 8.1$, 1H, H-1), 4.34 (dd, $J = 3.4$, 3.2 Hz, 1H, H-3), 3.82 (dq, $J = 9.7$, 6.2 Hz, 1H, H-5), 3.52 (s, 3H, OMe), 3.22 (dd, $J = 9.7$, 3.2 Hz, 1H, H-4), 2.18 (s, 3H, Ac), 1.37 (d, $J = 6.3$, 3H, H-6). ^{13}C NMR (101 MHz, CDCl_3): δ_{C} 169.77 (Ac), 98.78 (C-1), 71.49 (C-2), 68.72 (C-5), 63.16 (C-4), 62.27 (C-3), 56.93 (OMe), 20.64 (Ac), 18.08 (C-6). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_9\text{H}_{24}\text{O}_4\text{N}_6$ $[\text{M} + \text{Na}]^+$: 293.0969; found: 293.0971.

Benzyl 2,3-anhydro-4-azido-4,6-dideoxy- α -l-allopyranoside (26). To a solution of benzyl 2-O-acetyl-4-azido-4,6-dideoxy-3-O-mesyl- α -l-glucopyranoside **12** (17.85 g, 44.69 mmol) in anhydrous methanol (200 mL) was added sodium methoxide (2.68 g, 47 mmol), and the mixture was stirred at room temperature for 20 hours under an atmosphere of argon. The reaction solution was concentrated under reduced pressure and the residue was resuspended in ethyl acetate (100 mL). The organic solution was washed with distilled water (3 \times 50 mL), dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The syrupy mixture was purified by recrystallization in a mixture of 5%

ethyl acetate–hexanes (~20 mL) to afford the desired compound **26** as an off-white solid (10.36 g, 89% yield). R_f 0.65 (40% ethyl acetate–hexanes). $[\alpha]_D^{25} -197.5$ (c 0.76, CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ_{H} 7.42–7.28 (m, 5H, ArH), 5.01 (dd, $J = 3.2$ Hz, <1 Hz, 1H, H-1), 4.78 (d, $J = 12.0$ Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 4.65 (d, $J = 12.3$ Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 3.96 (dq, $J = 9.6$, 6.3 Hz, 1H, H-5), 3.55 (dd, $J = 4.1$ Hz, 1.6 Hz, 1H, H-3), 3.49 (dd, $J = 4.1$, 3.2 Hz, 1H, H-2), 3.16 (dd, $J = 9.6$ Hz, 1.5 Hz, 1H, H-4), 1.23, (d, $J = 6.3$ Hz, 3H, H-6). ^{13}C NMR (101 MHz, CDCl_3): δ_{C} 137.4 (ArC), 128.5 (ArC), 128.1 (ArC), 127.9 (ArC), 92.0 (C-1), 69.5 (CH_2Ph), 63.2 (C-5), 61.5 (C-4), 53.7 (C-2), 52.3 (C-3), 17.8 (C-6). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_{13}\text{H}_{22}\text{N}_4\text{O}_3$ $[\text{M} + \text{NH}_4]^+$: 279.1452; found: 279.1460.

Benzyl 2,4-diazido-2,4,6-trideoxy- α -l-altropyranoside (27) and benzyl 3,4-diazido-3,4,6-trideoxy- α -l-glucopyranoside (28)

General method. A solution of benzyl 2,3-anhydro-4-azido-4,6-dideoxy- α -l-allopyranoside (**26**), sodium azide and catalyst in the specified solvent (Table 1) was heated to the specified temperature for the specified amount for time. The solvent was then evaporated under reduced pressure. The residue was extracted into ethyl acetate, and the organic solution was washed with brine, dried over anhydrous Na_2SO_4 , and concentrated under reduced pressure. ^1H NMR experiment was used to directly determine the conversion rates and product ratios (**27/28**). For some experiments, the obtained residue was purified by column chromatography on silica gel using 2.5% ethyl acetate–hexanes as the eluent to yield the desired compounds **27** and **28** respectively. Data for compound **27**: R_f 0.43 (20% ethyl acetate–hexanes, twice). $[\alpha]_D^{25} -116.1$ (c 0.43, CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ_{H} 7.45–7.31 (m, 5H, ArH), 4.89 (s, 1H, H-1), 4.78 (d, 1H, $J = 11.7$ Hz, $\text{CH}_a\text{H}_b\text{Ph}$), 4.59 (d, 1H, $J = 11.8$ Hz, $\text{CH}_a\text{H}_b\text{Ph}$), 4.13–4.02 (m, 2H, H-3 and H-5), 3.85 (dd, 1H, $J = 4.0$ Hz, 1.9 Hz, H-2), 3.45 (d, 1H, $J = 9.8$ Hz, OH), 3.23 (dd, 1H, $J = 9.8$, 3.1 Hz, H-4), 1.38 (d, 3H, $J = 6.3$ Hz, H-6). ^{13}C NMR (101 MHz, CDCl_3): δ_{C} 135.8 (ArC), 128.7 (ArC), 128.5 (ArC), 128.3 (ArC), 96.9 (C-1), 70.3 ($\text{CH}_a\text{H}_b\text{Ph}$), 69.4 (C-3), 63.1 (C-5), 61.3 (C-4), 60.8 (C-2), 18.1 (C-6). HRMS (ESI-TOF) calc'd m/z for $\text{C}_{13}\text{H}_{16}\text{N}_6\text{O}_3$ $[\text{M} + \text{NH}_4]^+$: 322.1626; found 322.1622. Data for compound **28**: R_f 0.37 (20% ethyl acetate–hexanes, twice). $[\alpha]_D^{25} -330.0$ (c 1.7, CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ_{H} 7.43–7.32 (m, 5H, ArH), 4.93 (d, $J = 3.1$ Hz, 1H, H-1), 4.75 (d, $J = 11.7$ Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 4.55 (d, $J = 11.7$ Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 3.69–3.57 (m, 3H, H-2, H-4, and H-5), 2.92 (*high order t*, $J = 9.9$, 9.9 Hz, 1H, H-3) 2.17 (*high order d*, $J = 10.4$ Hz, 1H, OH-2), 1.30 (d, $J = 6.2$ Hz, 3H, H-6). ^{13}C NMR (101 MHz, CDCl_3): δ_{C} 128.7 (ArC), 128.4 (ArC), 128.2 (ArC), 96.8 (C-1), 71.9 (C-2), 70.1 ($\text{CH}_a\text{H}_b\text{Ph}$), 66.6 (C-5), 66.4 (C-3), 65.8 (C-4), 18.2 (C-6). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_{13}\text{H}_{16}\text{N}_6\text{O}_3$ $[\text{M} + \text{NH}_4]^+$: 322.1626; found 322.1617.

Benzyl 3-azido-2,4,6-trideoxy-2-morpholino- α -l-altropyranoside (29) and benzyl 4-azido-3,4,6-trideoxy-3-morpholino- α -l-glucopyranoside (30). A mixture of compound **26** (100 mg, 0.383 mmol), morpholine (0.07 mL, 0.765 mmol) and lithium perchlorate (82 mg, 0.765 mmol) in anhydrous acetonitrile (2.0 mL) was heated to 60 °C for 48 hours. The solution was concentrated under reduced pressure and the residue extracted with ethyl acetate (20 mL) and washed with brine (3 \times 20 mL). The organic solution was dried over anhydrous Na_2SO_4 , and concentrated under reduced pressure. The residue was purified



by column chromatography on silica gel using 10% ethyl acetate–hexanes as the eluent to yield the 2-morpholino product **29** (84 mg, 63% yield), and the 3-morpholino analog **30** (20 mg, 15% yield). Data for compound **29**: R_f 0.41 (40% ethyl acetate–hexanes). $[\alpha]_D^{25}$ –117.2 (c 1.7, CH_2Cl_2). ^1H NMR (400 MHz, CDCl_3) δ_{H} 7.42–7.29 (m, 5H, ArH), 4.98 (d, J = 4.4 Hz, 1H, H-1), 4.76 (d, J = 11.8 Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 4.56 (d, J = 11.8 Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 4.03 (dd, J = 7.6, 4.3 Hz, 1H, H-3), 3.92 (dq, J = 6.5, 6.5 Hz 1H, H-5), 3.65–3.73 (m, 4H, CH_2OCH_2), 3.61 (dd, J = 7.6, 4.2 Hz, 1H, H-4), 2.78 (dd, J = 7.6, 4.4 Hz, 1H, H-2), 2.73 (m, 2H, $\text{CH}_a\text{H}_b\text{NCH}_a\text{H}_b$), 2.55–2.61 (m, 2H, $\text{CH}_a\text{H}_b\text{NCH}_a\text{H}_b$), 1.34 (d, J = 6.5 Hz, 1H, H-6). ^{13}C NMR (101 MHz, CDCl_3) δ_{C} 136.8 (ArC), 128.6 (ArC), 128.2 (ArC), 128.1 (ArC), 95.7 (C-1), 69.7 ($\text{CH}_a\text{H}_b\text{Ph}$), 67.4 (CH_2OCH_2), 66.6 (C-3), 66.0 (C-5), 65.26 (C-2), 63.61 (C-4), 50.3 ($\text{CH}_a\text{H}_b\text{NCH}_a\text{H}_b$), 19.02 (C-6). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_{17}\text{H}_{24}\text{N}_4\text{O}_4$ $[\text{M} + \text{H}]^+$: 349.1880; found: 349.1870. Data for compound **30**: R_f 0.29 (40% ethyl acetate–hexanes). $[\alpha]_D^{25}$ –92.0 (c 1.9, CH_2Cl_2). ^1H NMR (400 MHz, CDCl_3) δ_{H} 7.44–7.31 (m, 5H, ArH), 4.97 (d, J = 3.8 Hz, 1H, H-1), 4.75 (d, J = 11.8 Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 4.59 (d, J = 11.8 Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 3.78–3.68 (m, 5H, H-2 + CH_2OCH_2), 3.65 (dq, J = 9.7, 6.2 Hz, 1H, H-5), 3.02 (dd, J = 10.1, 9.7 Hz, 1H, H-4), 2.94–2.84 (m, 5H, H-3, $\text{CH}_a\text{H}_b\text{NCH}_a\text{H}_b$), 1.28 (d, J = 6.2 Hz, 3H, H-6). δ_{C} ^{13}C NMR (101 MHz, CDCl_3): δ_{C} 137.0 (Ar-C), 128.6 (Ar-C), 128.1 (Ar-C), 128.1 (Ar-C), 97.4 (C-1), 69.9 ($\text{CH}_a\text{H}_b\text{Ph}$), 68.1 (C-2), 67.5 (CH_2OCH_2), 67.3 (C-5), 66.7 (C-3), 63.3 (C-4), 50.1 ($\text{CH}_a\text{H}_b\text{NCH}_a\text{H}_b$), 18.6 (C-6). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_{17}\text{H}_{24}\text{N}_4\text{O}_4$ $[\text{M} + \text{H}]^+$: 349.1870; found 349.1863.

Benzyl 2,4-diacetamido-2,4,6-trideoxy- α -L-altropyranoside (**32**)

Method 1. To a solution of 2,4-diazido compound **27** (80 mg, 0.26 mmol) in a 9 : 1 mixture of pyridine– H_2O (2.0 mL) was added triphenylphosphine (275 mg, 1.05 mmol, 4 equiv.), and the mixture was stirred at ambient temperature for 48 hours. The solution was concentrated under reduced pressure. The residue (**31**) was redissolved in methanol (2.0 mL), and acetic anhydride (250 μL) was added. After stirring for 4 hours, the solution was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel using a gradient of 30 \rightarrow 40% acetone–toluene as the eluent to afford the desired Alt-2,4-DiNHAc compound **32** (51.7 mg, 59% yield).

Method 2. To a solution of 2,4-diazido compound **27** (206 mg, 0.68 mmol) in dichloromethane (5.0 mL) and methanol (5.0 mL), was added concentrated ammonia (30%, 4 drops), and palladium on charcoal (5%, 28 mg) was added; the mixture was purged with hydrogen and stirred continuously under an atmosphere of hydrogen for 5 hours. The reaction solution was filtered over a 0.22 μm membrane disk, and the solution was concentrated under reduced pressure. The residue was redissolved in methanol (5.0 mL), and acetic anhydride (0.5 mL) was added. After stirring for 2 hours, the reaction solution was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel as above to afford compound **32** in pure form (184 mg, 81% yield). R_f 0.55 (10% $\text{MeOH}-\text{CH}_2\text{Cl}_2$). $[\alpha]_D^{25}$ –74.8 (c 0.7, CHCl_3). ^1H NMR (400 MHz, CDCl_3): δ_{H} 7.41–7.26 (m, 5H, Ph), 6.45 (d, J = 8.8 Hz, 1H, NHAc-C₂), 6.25 (d, J = 9.2 Hz, 1H, NHAc-C₄), 4.79 (br s, 1H, H-1), 4.69 (d, J = 11.8 Hz, 1H, $\text{CH}_a\text{H}_b\text{Ph}$), 4.52 (d, J = 11.8 Hz, 1H,

$\text{CH}_a\text{H}_b\text{Ph}$), 4.42 (dd, J = 8.8, 2.8 Hz, 1H, H-2), 4.03 (ddd, J = 10.1, 10.1, 2.9 Hz, 1H, H-4), 3.89–3.78 (m, 2H, H-5 + OH-3), 3.73 (m, 1H, H-3), 1.98 (s, 6H, 2 \times Ac), 1.24 (d, J = 6.3 Hz, 3H, H-6). ^{13}C NMR (101 MHz, CDCl_3): δ_{C} 170.0 (C=O), 169.7 (C=O), 136.1 (Ph), 128.7 (Ph), 128.3 (Ph), 128.1 (Ph), 98.3 (C-1), 69.9 ($\text{CH}_a\text{H}_b\text{Ph}$), 69.1 (C-3), 63.9 (C-5), 50.1 (C-2), 49.7 (C-4), 23.4 (Ac), 23.2 (Ac), 17.8 (C-6). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_{17}\text{H}_{25}\text{N}_2\text{O}_5$ $[\text{M} + \text{H}]^+$: 337.1751; found 337.1758.

2,4-Di-*N*-acetamido-2,4,6-trideoxy-L-altropyranose (5**).** A mixture of benzyl 2,4-di-*N*-acetyl-2,4,6-trideoxy- α -L-altropyranoside **32** (106.1 mg, 0.28 mmol), palladium hydroxide (68 mg) in methanol (10.0 mL), dichloromethane (2.0 mL) and acetic acid (100 μL) was stirred under a hydrogen atmosphere (balloon). After stirring for 20 h, the mixture was filtered through a 0.2 μm PTFE syringe filter and the clear solution was evaporated under reduced pressure to give compound **5** as a white solid (77 mg, ~quantitative yield). Selected ^1H NMR (400 MHz, D_2O): δ_{H} : 5.16 (d, J = 1.93 Hz, H-1 (α -pyranose), 4.93 (d, J = 2.6 Hz, H-1 (β -pyranose), 3.99–3.97 (m, major isomer), 3.94–3.92 (m, minor isomer), 3.87–3.79 (m), 3.70 (dd, J = 10.4 Hz, J = 3.0 Hz), 1.97 (s), 1.91 (s), 1.12 (d, J = 6.5 Hz, H-6 (major isomer)), 1.09 (d, J = 6.2 Hz, (minor isomer)). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_{12}\text{H}_{21}\text{N}_2\text{O}_6$ $[\text{M} + \text{H}]^+$: 289.1396; found 289.1394. ^1H NMR matches literature.¹⁸

2-Acetamido-5-*O*-acetyl-1,4-(*N*-acetylimino)-1,2,4,6-tetradecoxy-L-altritol (36**).** To a solution of 2,4-diazido compound **27** (31 mg, 0.10 mmol) in a 1 : 1 mixture of CH_2Cl_2 – MeOH (5.0 mL) was added ten drops of water and acetic acid (12 μL , 0.20 mmol) and Pd/C on charcoal (42 mg), and the flask was purged with an atmosphere of hydrogen, and kept under a positive hydrogen pressure with stirring for 48 hours. The solids were filtered off using a 0.22 μm membrane filter, and the solution was concentrated under reduced pressure. The crude syrup was acetylated using a mixture of acetic anhydride (1.0 mL) and pyridine (1.0 mL). After stirring overnight, the reaction solution was concentrated and co-evaporated with toluene (3 \times 5 mL). The crude residue was purified by column chromatography on silica gel using 2% methanol–dichloromethane to afford compound **27** as a syrupy product (19 mg, 60% yield). R_f 0.55 (10% $\text{MeOH}-\text{CH}_2\text{Cl}_2$). $[\alpha]_D^{25}$ –9.85 (c 1.3, MeOH). ^1H NMR (400 MHz, CDCl_3): δ_{H} 6.70 (d, J = 4.7 Hz, 1H, NHAc), 5.45 (qd, J = 6.6, 4.1 Hz, 1H, H-5), 5.35 (dd, J = 4.7, 3.9 Hz, 1H, H-3), 4.30–4.23 (m, 2H, H-1a + H-2), 4.22 (dd, J = 4.1, 4.1 Hz, H-4), 3.19 (high order dd, J = 10.3, 14.2 Hz, 1H, H-1b), 2.12 (s, 3H, Ac), 2.11 (s, 3H, Ac), 2.08 (s, 3H, Ac), 2.03 (s, 3H, Ac), 1.28 (d, J = 6.6 Hz, 3H, H-6). ^{13}C NMR (101 MHz, CDCl_3): δ_{C} 171.0 (Ac), 170.5 (Ac), 170.0 (Ac), 169.7 (Ac), 75.9 (C-3), 69.4 (C-5), 63.7 (C-4), 55.0 (C-2), 52.9 (C-1), 23.1 (Ac), 22.3 (Ac), 21.4 (Ac), 20.9 (Ac), 17.0 (C-6). HRMS (ESI-QTOF) calc'd m/z for $\text{C}_{14}\text{H}_{22}\text{N}_2\text{O}_6$ $[\text{M} + \text{Na}]^+$: 337.1376; found 337.1365.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We thank the Natural Sciences and Engineering Research Council of Canada as well as the Canadian Foundation for



Innovation and the Government of Alberta for their support of this project. C. S. thanks the University of Calgary for an Eyes High graduate scholarship.

Notes and references

- 1 K. U. Jansen, C. Knirsch and A. S. Anderson, *Nat. Med.*, 2018, **24**, 10–19.
- 2 U. Theuretzbacher, K. Outtersson, A. Engel and A. Karlén, *Nat. Rev. Microbiol.*, 2020, **18**, 275–285.
- 3 D. A. Kennedy and A. F. Read, *Proc. R. Soc. B*, 2017, **284**, 20162562.
- 4 M. Zunk and M. J. Kiefel, *RSC Adv.*, 2014, **4**, 3413–3421.
- 5 S. M. Logan, *Microbiology*, 2006, **152**, 1249–1262.
- 6 S. L. Howard, A. Jagannathan, E. C. Soo, J. P. M. Hui, A. J. Aubry, I. Ahmed, A. Karlyshev, J. F. Kelly, M. A. Jones, M. P. Stevens, S. M. Logan and B. W. Wren, *Infect. Immun.*, 2009, **77**, 2544–2556.
- 7 J. Horzempa, C. R. Dean, J. B. Goldberg and P. Castirc, *J. Bacteriol.*, 2006, **188**, 4244–4252.
- 8 S. R. Haseley and S. G. Wilkinson, *Eur. J. Biochem.*, 1997, **250**, 617–623.
- 9 P. Guerry, C. P. Ewing, M. Schirm, M. Lorenzo, J. Kelly, D. Pattarini, G. Majam, P. Thibault and S. Logan, *Mol. Microbiol.*, 2006, **60**, 299–311.
- 10 Y. A. Knirel, A. S. Shashkov, Y. E. Tsvetkov, P. E. Jansson and U. Zähringer, *Adv. Carbohydr. Chem. Biochem.*, 2003, **58**, 371–417.
- 11 S. Goon, J. F. Kelly, S. M. Logan, C. P. Ewing and P. Guerry, *Mol. Microbiol.*, 2003, **50**, 659–671.
- 12 W. K. Chou, S. Dick, W. W. Wakarchuk and M. E. Tanner, *J. Biol. Chem.*, 2005, **280**, 35922–35928.
- 13 R. Ménard, I. C. Schoenhofen, L. Tao, A. Aubry, P. Bouchard, C. W. Reid, P. Lachance, S. M. Twine, K. M. Fulton, Q. Cui, H. Hogues, E. O. Purisima, T. Sulea and S. M. Logan, *Antimicrob. Agents Chemother.*, 2014, **58**, 7430–7440.
- 14 K. Pradhan and S. S. Kulkarni, *Eur. J. Org. Chem.*, 2020, **2020**, 6819–6830.
- 15 Y. E. Tsvetkov, A. S. Shashkov, Y. A. Knirel and U. Zähringer, *Carbohydr. Res.*, 2001, **335**, 221–243.
- 16 N. W. See, N. Wimmer, E. H. Krenske and V. Ferro, *Eur. J. Org. Chem.*, 2021, **2021**, 1575–1584.
- 17 A. Liav and N. Sharon, *Carbohydr. Res.*, 1973, **30**, 109–126.
- 18 Y. J. Lee, A. Kubota, A. Ishiwata and Y. Ito, *Tetrahedron Lett.*, 2011, **52**, 418–421.
- 19 H. Liu, Y. Zhang, R. Wei, G. Andolina and X. Li, *J. Am. Chem. Soc.*, 2017, **139**, 13420–13428.
- 20 M. Zunk, J. Williams, J. Carter and M. J. Kiefel, *Org. Biomol. Chem.*, 2014, **12**, 2918–2925.
- 21 J. T. Williams, L. Corcilus, M. J. Kiefel and R. J. Payne, *J. Org. Chem.*, 2016, **81**, 2607–2611.
- 22 B. Dhakal, S. Buda and D. Crich, *J. Org. Chem.*, 2016, **81**, 10617–10630.
- 23 O. Popik, B. Dhakal and D. Crich, *J. Org. Chem.*, 2017, **82**, 6142–6152.
- 24 B. Dhakal and D. Crich, *J. Am. Chem. Soc.*, 2018, **140**, 15008–15015.
- 25 R. P. McGeary, S. Rasoul Amini, V. W. S. Tang and I. Toth, *J. Org. Chem.*, 2004, **69**, 2727–2730.
- 26 R. P. McGeary, K. Wright and I. Toth, *J. Org. Chem.*, 2001, **66**, 5102–5105.
- 27 H. M. Flowers, *Carbohydr. Res.*, 1979, **74**, 177–185.
- 28 L. Hough and A. C. Richardson, in *Rodd's Chemistry of Carbon Compounds*, ed. S. Coffey, Elsevier, Amsterdam, 2nd edn, 1964, pp. 67–595.
- 29 K. J. Hale, L. Hough, S. Manaviazar and A. Calabrese, *Org. Lett.*, 2014, **16**, 4838–4841.
- 30 U. Zehavi and N. Sharon, *J. Org. Chem.*, 1972, **37**, 2141–2145.
- 31 M. Miljković, in *Carbohydrates: Synthesis, Mechanisms, and Stereoelectronic Effects*, ed. M. Miljkovic, Springer, New York, NY, 2009, pp. 221–244.
- 32 D.-Q. Yuan, T. Tahara, W.-H. Chen, Y. Okabe, C. Yang, Y. Yagi, Y. Nogami, M. Fukudome and K. Fujita, *J. Org. Chem.*, 2003, **68**, 9456–9466.
- 33 M. T. C. Walvoort, G.-J. Moggré, G. Lodder, H. S. Overkleeft, J. D. C. Codée and G. A. van der Marel, *J. Org. Chem.*, 2011, **76**, 7301–7315.
- 34 A. Medgyes, E. Farkas, A. Lipták and V. Pozsgay, *Tetrahedron*, 1997, **53**, 4159–4178.
- 35 R. Hevey, A. Morland and C.-C. Ling, *J. Org. Chem.*, 2012, **77**, 6760–6772.
- 36 R. Hevey and C.-C. Ling, *Org. Biomol. Chem.*, 2013, **11**, 1887–1895.
- 37 P. Zhang, R. Hevey and C.-C. Ling, *J. Org. Chem.*, 2017, **82**, 9662–9674.
- 38 R. Hevey and C.-C. Ling, *Carbohydr. Res.*, 2017, **445**, 65–74.
- 39 Y. Cai, C.-C. Ling and D. R. Bundle, *J. Org. Chem.*, 2009, **74**, 580–589.
- 40 H. Okazaki, K. Hanaya, M. Shoji, N. Hada and T. Sugai, *Tetrahedron*, 2013, **69**, 7931–7935.
- 41 B. P. Shivani and A. K. Chakraborti, *J. Org. Chem.*, 2007, **72**, 3713–3722.
- 42 B. P. Shivani and A. K. Chakraborti, *J. Mol. Catal. A: Chem.*, 2007, **263**, 137–142.
- 43 G. Sabitha, R. S. Babu, M. Rajkumar and J. S. Yadav, *Org. Lett.*, 2002, **4**, 343–345.
- 44 P. Salehi, B. Seddighi, M. Irandoost and F. K. Behbahani, *Synth. Commun.*, 2000, **30**, 2967–2973.
- 45 T. Nemcsok, Z. Rapi, P. Bagi and P. Bakó, *Synth. Commun.*, 2019, **49**, 2388–2400.
- 46 J. M. Vega-Pérez, J. I. Candela, M. Vega and F. Iglesias-Guerra, *Carbohydr. Res.*, 1995, **279**, C5–C8.
- 47 M. Chini, P. Crotti and F. Macchia, *Tetrahedron Lett.*, 1990, **31**, 5641–5644.
- 48 Y. Fukuta, T. Mita, N. Fukuda, M. Kanai and M. Shibasaki, *J. Am. Chem. Soc.*, 2006, **128**, 6312–6313.
- 49 V. P. K. Kondapi, O.-M. Soueidan, S. N. Hosseini, N. Jabari and F. G. West, *Eur. J. Org. Chem.*, 2016, **2016**, 1367–1379.
- 50 H. Sajiki, *Tetrahedron Lett.*, 1995, **36**, 3465–3468.
- 51 H.-Y. L. Wang and G. A. O'Doherty, *Chem. Commun.*, 2011, **47**, 10251.

