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Synthesis and biological evaluation of novel artemisone–piperazine–tetronamide hybrids†

 Meng-Xue Wei,  Jia-Ying Yu, Xin-Xin Liu, Xue-Qiang Li,* Jin-Hui Yang, Meng-Wei Zhang, Pei-Wen Yang, Si-Si Zhang and Yu He

For the first time, six novel artemisone–piperazine–tetronamide hybrids (**12a–f**) were efficiently synthesised from dihydroartemisinin (DHA) and investigated for their *in vitro* cytotoxicity against some human cancer cells and benign cells. All the targets showed good cytotoxic activity *in vitro*. Hybrid **12a** exhibited much better inhibitory activity against human liver cancer cell line SMMC-7721 ($IC_{50} = 0.03 \pm 0.04 \mu\text{M}$ for 24 h) than the parent DHA ($IC_{50} > 0.7 \mu\text{M}$), and two references, vincristine (VCR; $IC_{50} = 0.27 \pm 0.03 \mu\text{M}$) & cytosine arabinoside (ARA; $IC_{50} = 0.63 \pm 0.04 \mu\text{M}$). Furthermore, hybrid **12a** had low toxicity against human benign liver cell line LO2 ($IC_{50} = 0.70 \pm 0.02 \mu\text{M}$ for 24 h) compared with VCR, ARA, and DHA *in vitro*. Moreover, the inhibitory activity of hybrid **12a** was obviously enhanced when human liver cancer cell line MHCC97H absorbed Fe^{2+} *in vitro*.

1. Introduction

Artemisinin (ARS, Fig. 1) obtained from *Artemisia annua* is used to treat chills and fever.¹ In the last century, it was found that ARS has anticancer activity,² and a large volume of studies have highlighted the potential of ARS as a novel therapeutic agent for cancer.^{3–7} Cancer is often life-threatening and hinders health and social progress.⁸ Artemisinin and its derivatives are highly selective and cytotoxic to tumor cells.⁹ ARS combined with cisplatin, can induce apoptosis of ovarian cancer cells.¹⁰ Dihydroartemisinin (DHA, **1**) exhibited high anticancer activity in lewis lung carcinoma (LLC) cell lines *in vitro*.¹¹ In 2006, Haynes *et al.* reported the discovery, synthesis and preclinical evaluation of artemisone (**3**) as an effective oral antimalarial drug.¹² In 2011, the same group reported **3** combined with oxaliplatin can enhance anticancer drug activity in breast cancer cells MCF-7, colon cancer cells HCT116, and SW480.¹³ The mechanism of action of artemisinin-type drugs is still under research. Our group also reported a series of dihydroartemisinin ramifications, some of which showed good cytotoxic activity against cancer cells.¹⁴ Tetronamides are an important class of β -hetero-substituted butenolides and has extensive biological activities.¹⁵ In previous research, our group has synthesised some tetronamides¹⁶ and some of them have good anticancer activity. The combination of artemisinin-type drugs with anticancer

drugs can enhance the activity of the anticancer drugs *in vitro* or *in vivo*.^{9,10,17,18}

Given the therapeutic properties of tetronamides and the lack of artemisone–piperazine–tetronamide hybrids reported in

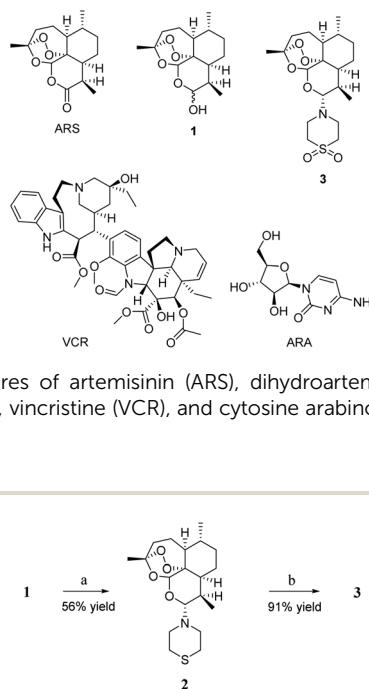


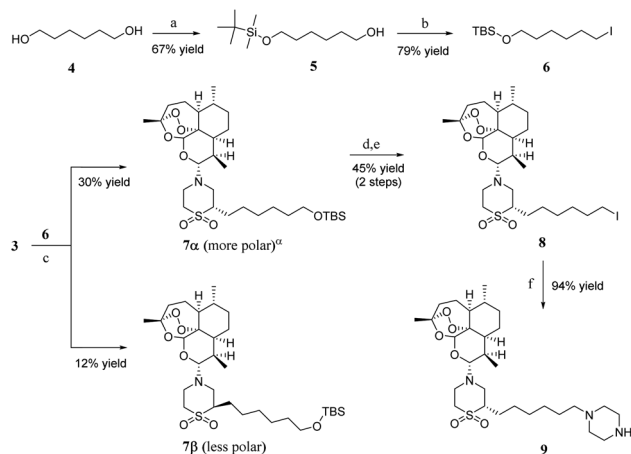
Fig. 1 Structures of artemisinin (ARS), dihydroartemisinin (DHA, **1**), artemisone (**3**), vincristine (VCR), and cytosine arabinoside (ARA).

State Key Laboratory of High-efficiency Utilization of Coal and Green Chemical Engineering, Ningxia Engineering Research Center for Natural Medicine, National Demonstration Center for Experimental Chemistry Education, College of Chemistry and Chemical Engineering, Ningxia University, 489 Helanshan West Road, Yinchuan 750021, China. E-mail: lixq@nxu.edu.cn

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Scheme 1 Synthesis of artemisone **3**. Reagents and conditions: (a) dry DMSO, oxalyl chloride, dry xylene, rt, 7–10 min, then drop in dry DCM solution for thiomorpholine, rt; (b) Na_2WO_4 , H_2O_2 , THF: water = 4 : 1, rt. DMSO = dimethyl sulfoxide; DCM = dichloromethane; THF = tetrahydrofuran.





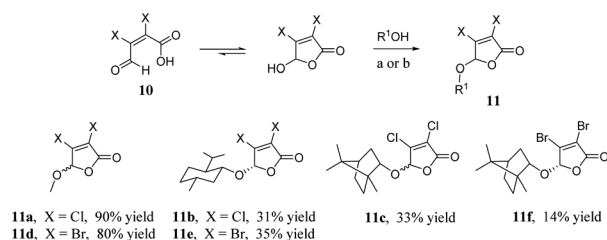
Scheme 2 Synthesis of artemisone–piperazine **9**. Reagents and conditions: (a) *tert*-butyldimethylsilyl chloride, imidazole, dry DMF, rt; (b) triphenylphosphine, imidazole, dry THF, rt, then add iodine, keep in dark place; (c) LDA, dry THF, $-78\text{ }^{\circ}\text{C}$ to rt; (d) TBAF, THF, rt; (e) triphenylphosphine, imidazole, dry THF, rt, then add iodine, keep in dark place; (f) piperazine, DIPEA, dry THF; dry MeOH = 20 : 1, rt to reflux. DMF = *N,N*-dimethylformamide; LDA = lithium diisopropylamide; TBAF = tetrabutylammonium fluoride; DIPEA = diisopropylethylamine. ^a The stereochemistry of **7α** was assigned on the basis of X-ray analysis of a closely related artemisone derivative prepared by the same method. These findings will be reported in a future publication.

the literature, we designed and synthesised a series of hybrid tetronamide derivatives possessing an artemisone–piperazine moiety (**12a–f**, Scheme 4) to evaluate their *in vitro* cytotoxicity and compare with VCR and ARA. Then we also research the effects of iron ions (Fe^{2+}) on cytotoxic activity.^{19–22}

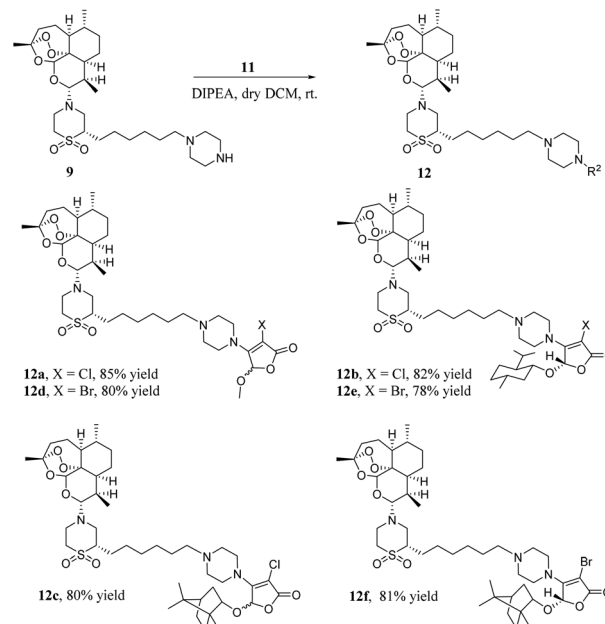
2. Results and discussion

2.1. Chemistry

Artemiside **2** was synthesised through substitution reaction using dry dimethyl sulfoxide (DMSO) as a catalyst (shown in Scheme 1). First, **1** and oxaloyl chloride formed chloroartemisinin. Then the amine of thiomorpholine attacked chloroartemisinin to produce **2**.²³ **2** was converted to **3** by mild oxidation of hydrogen peroxide and Na_2WO_4 , and this reaction rate was affected by temperature; the higher the temperature, the faster the speed, but the more by-products were produced.



Scheme 3 Synthesis of furan ethers **11a–f**. Reagents and conditions: (a) MeOH, 1,2-dichloroethane, H_2SO_4 , rt then reflux; (b) menthol or borneol, cyclohexane, H_2SO_4 , rt then reflux.



Scheme 4 Synthesis of hybrids **12a–f**.

Table 1 Cytotoxic activities against SMMC-7721 and LO2 cell lines for **12a–f**, VCR, ARA and DHA^a

Compounds	IC_{50} (μM)/24 h	
	SMMC-7721	LO2
12a	0.03 ± 0.04	0.70 ± 0.02
12b	0.32 ± 0.03	0.36 ± 0.03
12c	0.07 ± 0.04	0.07 ± 0.03
12d	0.34 ± 0.02	0.37 ± 0.04
12e	0.21 ± 0.02	0.33 ± 0.03
12f	0.22 ± 0.03	0.30 ± 0.03
VCR	0.27 ± 0.03	0.30 ± 0.02
ARA	0.63 ± 0.04	0.43 ± 0.04
DHA	>0.7	0.22 ± 0.04

^a SMMC-7721 = human liver cancer cell lines; LO2 = human benign liver cell lines; VCR = vincristine; ARA = cytosine arabinoside; μM = $\mu\text{mol mL}^{-1}$; IC_{50} values represent the compound concentration (μM) required to inhibit tumor cell proliferation by 50% and were calculated using concentrations from triplicate measurements.

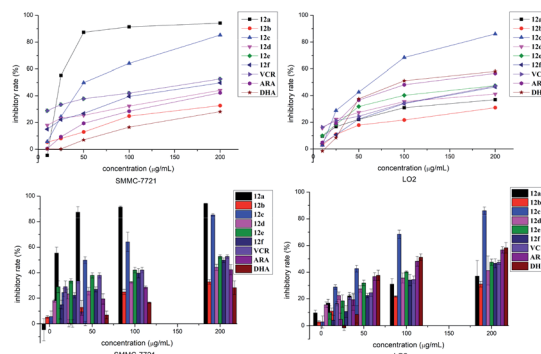


Fig. 2 Inhibitory effect of **12a–f**, VCR, ARA and DHA with various concentrations on SMMC-7721 and LO2 cells at 24 h.



Finally, we performed the reaction at room temperature (rt) with a 91% yield.²⁴

As shown in Scheme 2, one hydroxyl group of 1,6-hexanediol **4** was protected with *tert*-butyldimethylsilyl chloride to produce compound **5**, and then compound **6** was obtained through Appel reaction. The synthesis of **7 α** and **7 β** needed a strong base to promote the reaction at -78 °C, and the yield was low.

Unfortunately, we did not find another way to get a better yield. However, **7 α** and **7 β** could be separated through chromatography on silica gel column; **7 α** made up about 71 percent of the total products **7 α** and **7 β** . We chose **7 α** as the major material of the next reactions. **7 α** was converted to compound **8** through a two-step series reaction. First, compound **7 α** was deprotected with tetrabutylammonium fluoride (TBAF), and then iodine

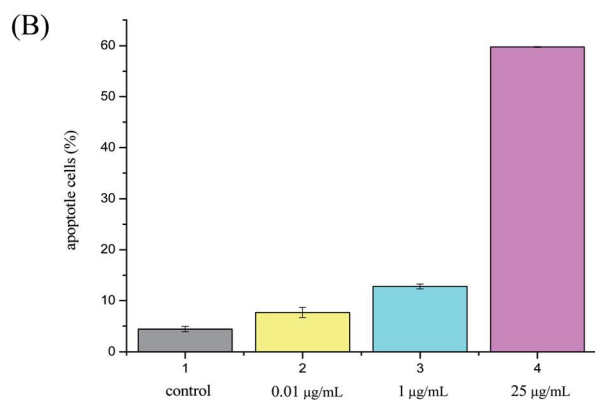
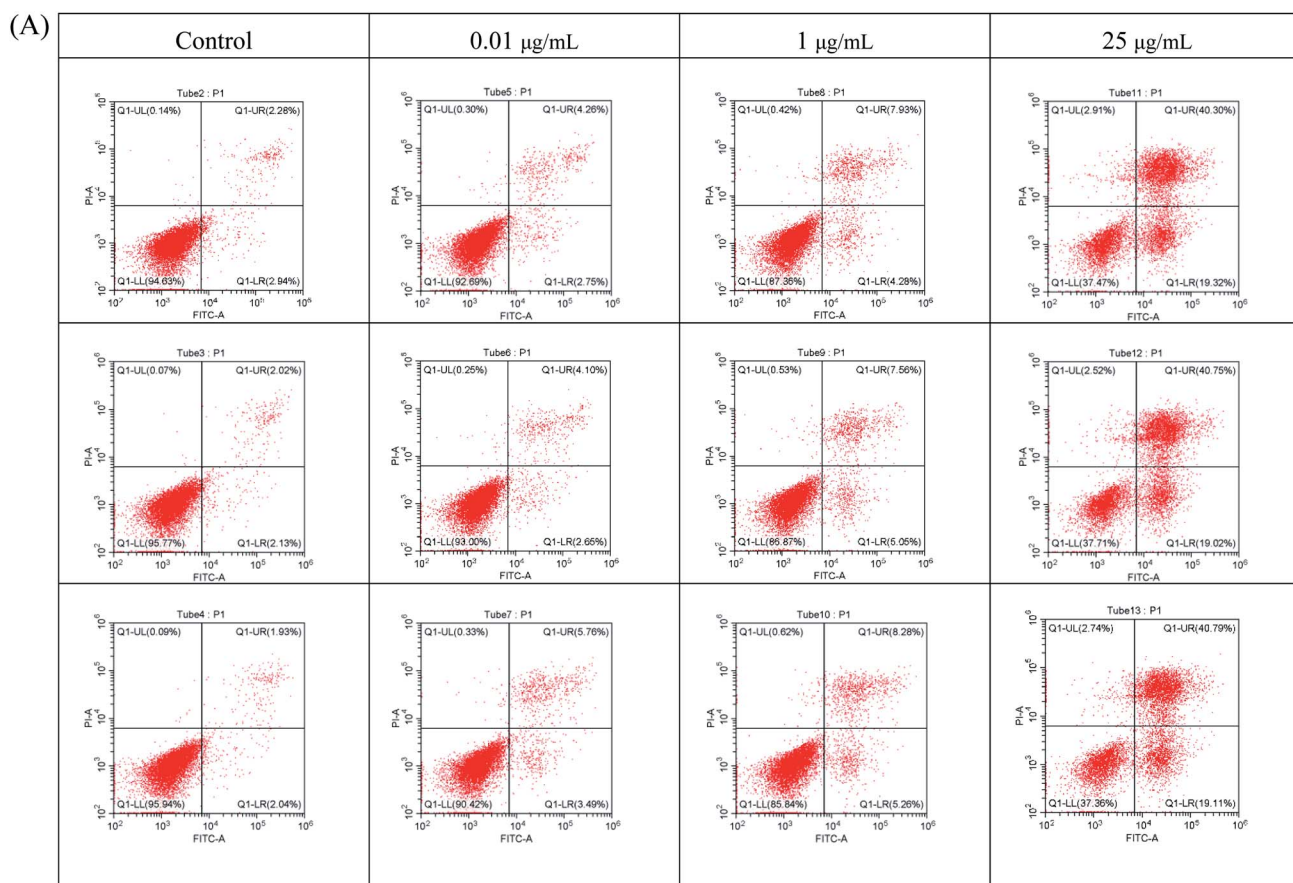
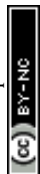


Fig. 3 (A) Flow cytometric analysis of MHCC97H cell lines using different concentrations of hybrid **12a** (0.01 , 1 and 25 $\mu\text{g mL}^{-1}$) for 48 h and each concentration was detected three times in parallel. The percentage of cells positive for AV and/or PI was reported in the quadrants including cells in the lower left quadrant: live cells; lower right quadrant: early apoptotic cells; upper right quadrant: late apoptotic cells; upper left quadrant: necrotic cells. (B) Apoptotic assay by flow cytometry. MHCC97H cell was treated with **12a** at 0.01 , 1 , and 25 $\mu\text{g mL}^{-1}$ for 48 h. Then AV and PI staining were used and performed on flow cytometry.



substituted the hydroxyl through Appel reaction. Compound **8**, piperazine and diisopropylethylamine (DIPEA) were dissolved in methanol and tetrahydrofuran ($v/v = 1 : 20$), then heated to 61 °C, and stirred for about 4 hours to obtain artemisone-piperazine **9** with 94% yield.

As shown in Scheme 3, we synthesised the halogenated furan ethers **11a–f** with mucochloric acid or mucobromic acid **10** and alcohol as raw materials, and concentrated sulphuric acid as a catalyst.²⁵

As shown in Scheme 4, a mixture of artemisone-piperazine **9** and DIPEA in DCM was added to a stirred solution of furan ethers **11** and stirred for 4 hours at room temperature. Then, it was purified by chromatography on a silica gel column to give hybrids **12a–f** in good yields.

2.2. Biological activity

The cytotoxic activities of all the target hybrids **12a–f** were tested. The results are given in Table 1. It is clear that **12a** ($IC_{50} = 0.03 \pm 0.04 \mu\text{M}$, 24 h) shows better cytotoxic activity against human liver cancer cell lines SMMC-7721 than the anticancer drugs VCR ($IC_{50} = 0.27 \pm 0.03 \mu\text{M}$, 24 h), ARA ($IC_{50} = 0.63 \pm 0.04 \mu\text{M}$, 24 h) and DHA ($IC_{50} > 0.7 \mu\text{M}$, 24 h) *in vitro*, and it is gratifying that **12a** ($IC_{50} = 0.70 \pm 0.02 \mu\text{M}$, 24 h) had lower cytotoxicity against human benign liver cells LO2 compared with VCR ($IC_{50} = 0.30 \pm 0.02 \mu\text{M}$, 24 h), ARA ($IC_{50} = 0.43 \pm 0.04 \mu\text{M}$, 24 h) and DHA ($IC_{50} = 0.22 \pm 0.04 \mu\text{M}$, 24 h) *in vitro*. As a whole, hybrid **12a** showed far more cytotoxic against human liver cancer cells SMMC-7721 than benign cells LO2 compared with VCR, ARA and DHA *in vitro*.

As can be seen from Fig. 2, hybrid **12a** showed the best inhibitory activity against SMMC-7721 cell lines, and its inhibitory rate increased with increasing concentration. Moreover, hybrid **12a** was less toxic against LO2 cell lines compared with DHA, ARA, and VCR. As shown in Fig. 3A, the annexin V-FITC (AV) and propidium iodide (PI) staining were used and performed on flow cytometry to explore the apoptotic cells

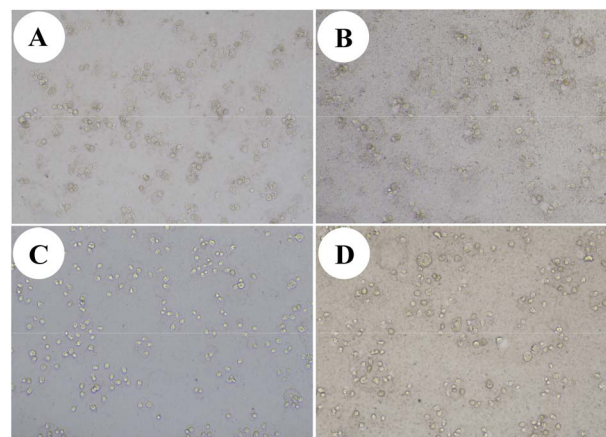


Fig. 4 Morphological changes of human liver cancer cells induced by hybrid **12a** at a concentration of $10 \mu\text{g mL}^{-1}$ after 48 h in (A) MHCC97H cells; (B) MHCC97H cells with Fe^{2+} ; (C) SMMC-7721 cells; (D) SMMC-7721 cells with Fe^{2+} .

mediated by **12a**. MHCC97H cells were treated with **12a** at different concentrations of 0.01, 1, and $25 \mu\text{g mL}^{-1}$, respectively. As can be seen from Fig. 3B, as the concentration of **12a** increased, the percentage of apoptosis increased significantly from 7.67% to 59.76%. Hybrid **12a** could induce the apoptosis of MHCC97H cell lines in an appropriate concentration-dependent manner. The results indicated that the anti-proliferative effect of **12a** might be related to the induction of apoptosis.

In addition, we chose **12a** as a sample to test the cytotoxic activity against other cancer cells and benign cells. The results are shown in Table 2. It's clear that hybrid **12a** had a good cytotoxic activity against human liver cancer cells MHCC97H ($IC_{50} = 0.01 \pm 0.02 \mu\text{M}$), colon cancer cells HCT116 ($IC_{50} = 0.01 \pm 0.04 \mu\text{M}$), breast cancer cells MCF-7 ($IC_{50} = 0.01 \pm 0.04 \mu\text{M}$), and prostate cancer cells DU145 ($IC_{50} = 0.02 \pm 0.04 \mu\text{M}$) for 48 h *in vitro*. The cytotoxicity against liver cancer cells SMMC-7721 of hybrid **12a** had no discernible change as time prolonged to 48 h ($IC_{50} = 0.03 \pm 0.03 \mu\text{M}$; Table 2) compared with that for 24 h ($IC_{50} = 0.03 \pm 0.04 \mu\text{M}$; Table 1). It is reported that some cancer cells can import Fe^{2+} through abnormal iron transporters in cancer cell membranes, while benign cells seldom have abnormal iron transporters.^{19,20} We choose **12a** as a sample to test the cytotoxic activity against human liver cancer cells MHCC97H and SMMC-7721 under an environment with Fe^{2+} for 48 h *in vitro*. Instead of using a medium of the mixture of Fe^{2+} and **12a**, Fe^{2+} was added when culturing cells, which is a good way to simulate the process of cancer cells absorbing Fe^{2+} . Hybrid **12a** showed better cytotoxic activity against human liver cancer cells with Fe^{2+} than without Fe^{2+} . We believe that hybrid **12** may have better anti-tumor activity *in vivo*. The morphological changes of human liver cancer cells induced by hybrid **12a** are shown in Fig. 4. It is clear that hybrid **12a** can effectively inhibit tumor cells' proliferation *in vitro*.

Table 2 Cytotoxic activities of **12a** against some cancer and benign cell lines

Cells	IC_{50} (μM)/48 h
MHCC97H	0.01 ± 0.02
MHCC97H	0.003 ± 0.002^a
SMMC-7721	0.03 ± 0.03
SMMC-7721	0.02 ± 0.03^a
HCT116	0.01 ± 0.04
CCD18CO	0.06 ± 0.04
MCF-7	0.01 ± 0.04
MCF-10A	0.04 ± 0.04
DU145	0.02 ± 0.04
RWPE-1	0.10 ± 0.03

^a Cancer cells were cultured with $100 \mu\text{mol L}^{-1} \text{FeSO}_4$. MHCC97H = human liver cancer cell lines; HCT116 = human colon cancer cell lines; CCD18CO = human benign colon cell lines; MCF-7 = human breast cancer cell lines; MCF-10A = human benign breast cell lines; DU145 = human prostate cancer cell lines; RWPE-1 = human benign prostate cell lines.



3. Conclusions

For the first time, a series of artemisone–piperazine–tetronamide hybrids were synthesised and their cytotoxic activities were represented in this study. Artemisone was synthesised through substitution and oxidation using DHA as the raw material. Halogenated furan ethers were synthesised from mucochloric acid or mucobromic acid, and some chiral furan ethers were obtained through recrystallisation in petroleum ether, water and ethanol. Then artemisone and furan ethers were combined using piperazine as a linker to get the target hybrids through 5 steps reaction. Epimers of **7** can be separated by chromatography on a silica gel column. All target hybrids showed good cytotoxic activity *in vitro*. **12a** was the most active hybrid, with better antiproliferative action against SMMC-7721 cell lines ($IC_{50} = 0.03 \pm 0.04 \mu\text{M}$) and lower toxicity against LO2 cell lines ($IC_{50} = 0.70 \pm 0.02 \mu\text{M}$) compared with VCR, ARA, and DHA for 24 h *in vitro*. The cytotoxicity of hybrid **12a** against human liver cancer cells SMMC-7721 and MHCC97H was enhanced when cancer cells were cultured with Fe^{2+} *in vitro*. The results could provide evidence for the antitumor mechanism of artemisinin-type drugs with Fe^{2+} .

4. Experimental

4.1. General

Thin-layer chromatography (TLC) was carried out on silica GF254 plates (Qingdao Haiyang Chemical Co., Ltd, China). IR spectra were recorded on a FTIR-8400S spectrometer as KBr discs. ^1H NMR and ^{13}C NMR spectra were obtained with a Bruker Avance III 400 MHz spectrometer in chloroform- d (CDCl_3) and tetramethylsilane (TMS) was used as an internal standard. High resolution mass spectrums (HRMS) of targets were recorded on a Thermo Fisher LTQ Orbitrap XL. All chemicals were used as received without further purification unless otherwise stated.

4.2. Synthesis

4.2.1. Synthesis of 4-((3R,5aS,6R,8aS,9R,10R,12aR)-3,6,9-trimethyldecahydro-3H-3,12-epoxy[1,2]dioxepino[4,3-*f*]isochromen-10-yl)thiomorpholine (2). Under the anhydrous condition, anhydrous thiomorpholine (4.2 g, 41 mmol) and dry xylene (35 mL) were added to a flask-3-neck (250 mL), then stirred until completely dissolved. Under the anhydrous condition, dry DCM (25 mL) and DHA (2 g, 7 mmol) were added to another flask-3-neck (200 mL) and stirred at room temperature, within 1 min DMSO (50–70 μL) was added and stirred for another 3 min. Oxalyl chloride (0.6–0.7 mL) was slowly added until no gas was produced and stirred another 8–10 min. The mixture was slowly dropwise in the flask containing thiomorpholine, allowing overnight reaction at room temperature. Following the addition of saturated sodium carbonate (20 mL) and AcOEt (30 mL), the organic phase was separated, and the aqueous layer was extracted with AcOEt (2×30 mL). The combined extracts were washed with brine (20 mL), dried over anhydrous Na_2SO_4 , and concentrated under reduced pressure.

The residue was purified by chromatography on an ammonia-treated silica gel column, eluting with petroleum ether–ethyl acetate (10 : 1). Artemiside **2** was formed in 56% yield as a white solid. ^1H NMR (400 MHz, CDCl_3) δ 5.28 (s, 1H), 4.18 (d, $J = 10.2$ Hz, 1H), 3.45–3.26 (m, 1H), 3.07–2.95 (m, 1H), 2.89 (t, $J = 4.9$ Hz, 3H), 2.55 (ddd, $J = 10.9, 9.4, 5.8$ Hz, 1H), 2.49–2.35 (m, 2H), 2.35–2.26 (m, 2H), 2.06–1.95 (m, 2H), 1.87 (ddt, $J = 10.0, 6.4, 3.4$ Hz, 1H), 1.71 (dq, $J = 10.2, 3.3$ Hz, 1H), 1.56 (dt, $J = 13.7, 4.3$ Hz, 1H), 1.46 (q, $J = 7.4, 4.9$ Hz, 3H), 1.31 (dq, $J = 10.8, 6.4, 5.2$ Hz, 1H), 1.24 (dt, $J = 11.0, 5.5$ Hz, 1H), 1.02 (td, $J = 13.6, 13.2, 3.9$ Hz, 1H), 0.95 (d, $J = 6.2$ Hz, 3H), 0.79 (d, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 105.3, 93.7, 91.3, 80.5, 52.8, 51.8, 47.2, 44.6, 37.2, 35.9, 34.2, 29.6, 25.2, 24.3, 21.8, 20.2, 13.9.

4.2.2. Synthesis of 4-((3R,5aS,6R,8aS,9R,10R,12aR)-3,6,9-trimethyldecahydro-12H-3,12-epoxy[1,2]dioxepino[4,3-*f*]isochromen-10-yl)thiomorpholine 1,1-dioxide (3). Compound **2** (1.87 g, 5.1 mmol), Na_2WO_4 (0.1 g), 60 mL solvent THF–water (4 : 1) were added to a flask (100 mL), stirred at room temperature for 5 min, then H_2O_2 (1.5 mL, 33%) added in 0.5 h and stirred for another 2 h at room temperature. After the compound **2** had disappeared, the mixture was diluted with ethyl acetate (100 mL), the organic phase was separated and washed with brine (3×30 mL), dried over anhydrous Na_2SO_4 and the solvent removed under diminished pressure. The residue was purified by chromatography on a silica gel column, eluting with petroleum ether–ethyl acetate 3 : 1. Compound **3** was formed in 91% yield as a white solid. mp 145.7–146.1 $^\circ\text{C}$; $[\alpha]_D^{20} +34$ (c 0.5, CH_2Cl_2); ^1H NMR (400 MHz, CDCl_3) δ 5.27 (s, 1H), 4.21 (d, $J = 10.3$ Hz, 1H), 3.49 (dt, $J = 14.6, 5.3$ Hz, 2H), 3.37 (dt, $J = 13.8, 5.3$ Hz, 2H), 3.20 (s, 4H), 2.58 (dq, $J = 14.4, 7.1, 4.4$ Hz, 1H), 2.34 (ddd, $J = 14.6, 13.3, 4.0$ Hz, 1H), 2.00 (ddd, $J = 14.6, 5.0, 2.9$ Hz, 1H), 1.87 (ddq, $J = 13.6, 6.8, 3.2$ Hz, 1H), 1.76–1.65 (m, 2H), 1.57 (dt, $J = 13.8, 4.3$ Hz, 1H), 1.47 (dd, $J = 5.1, 2.4$ Hz, 1H), 1.45–1.41 (m, 1H), 1.36 (s, 3H), 1.33 (d, $J = 3.7$ Hz, 1H), 1.24 (dt, $J = 11.1, 5.5$ Hz, 1H), 1.06–0.97 (m, 1H), 0.94 (d, $J = 6.1$ Hz, 3H), 0.79 (d, $J = 7.1$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 103.9, 92.4, 91.6, 80.3, 51.7, 50.5, 45.8, 37.3, 36.3, 34.2, 28.3, 26.0, 24.7, 21.6, 20.3, 13.5.

4.2.3. Synthesis of 6-((tert-butyl dimethylsilyl)oxy)hexan-1-ol (5). 1,6-Hexanediol **4** (2.36 g, 20 mmol), *tert*-butyldimethylsilyl chloride (1.8 g, 12 mmol) and imidazole (2.1 g, 30 mmol) were added in 40 mL anhydrous DMF and stirred vigorously at room temperature overnight. Then 40 mL water was added and the mixture was added to ethyl acetate (80 mL). The combined organic layer was washed with brine (3×35 mL), dried by Na_2SO_4 and the solvent removed under diminished pressure. The residue was purified by chromatography on a silica gel column. Get the elution with 16 : 1 petroleum ether–ethyl acetate. Compound **5** was formed in 67% yield as a colourless liquid. ^1H NMR (CDCl_3) δ 3.67–3.57 (m, 4H), 1.63–1.47 (m, 4H), 1.36 (p, $J = 3.4$ Hz, 4H), 0.89 (s, 9H), 0.04 (s, 6H); ^{13}C NMR (CDCl_3) δ 63.6, 62.9, 33.0, 33.1, 26.3, 26.0, 18.7, –4.9.

4.2.4. Synthesis of tert-butyl((6-iodohexyl)oxy) dimethylsilane (6). Compound **5** (1.6 g, 6 mmol), triphenylphosphine (1.83 g, 7 mmol) and imidazole (0.84 g, 12 mmol) were added in dry THF and stirred for 5 min, then add iodine (1.78 g, 7 mmol), kept in a dark place and stirred overnight at



room temperature. Then the mixture was diluted with ethyl acetate 60 mL and washed with saturated $\text{Na}_2\text{S}_2\text{O}_3$ 25 mL and saturated salt solution (2×40 mL), dried over Na_2SO_4 , and the solvent removed under diminished pressure. The residue was purified by chromatography on a silica gel column, eluting with petroleum ether–ethyl acetate 10 : 1. Compound **6** was formed in 79% yield as a colourless liquid. ^1H NMR (400 MHz, CDCl_3): δ 3.593 (t, $J = 6.4$ Hz, 2H), 3.175 (t, $J = 7$ Hz, 2H), 1.773–1.856 (m, 2H), 1.476–1.544 (m, 2H), 1.307–1.436 (m, 4H), 0.883 (s, 9H), 0.038 (s, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 63.0, 33.5, 32.6, 30.3, 26.0, 24.8, 18.4, 7.1, –5.2.

4.2.5. Synthesis of (2S)-2-(6-((tert-butyl)dimethylsilyloxy)hexyl)-4-((3R,5aS,6R,8aS,9R,10R,12aR)-3,6,9-trimethyldecahydro-3H-3,12-epoxy[1,2]dioxepino[4,3-*i*]isochromen-10-yl)thiomorpholine 1,1-dioxide (7 α). Under anhydrous condition and Ar flow, 2,2'-bipyridine (about 1 mg), dry *N,N*-diisopropylamine (0.63 g, 6.3 mmol) and dry THF 10 mL were added to a 100 mL two-neck flask at -40 °C, stirred for 5 min, then add BuLi 4 mL (1.6 M, 6 mmol) stirred for 30 min and then cooled to -78 °C. Compound **3** (1.2 g, 3 mmol) in 10 mL dry THF was added at -78 °C, then stirred for 45 min, and compound **6** (1.19–1.2 g) in 5 mL THF was added slowly. After adding was complete, the reaction mixture was stirred for 3 hours at -78 °C and then the mixture was warmed to room temperature slowly. After the compound **3** disappeared, the mixture was diluted with ethyl acetate (80 mL). The combined organic layer was washed with saturated NH_4Cl (40 mL) and saturated salt solution (2×40 mL), dried over Na_2SO_4 and the solvent removed under diminished pressure. The residue was purified by chromatography on a silica gel column, eluting with petroleum ether–ethyl acetate 6 : 1. Compound **7 α** was formed in 30% yield as a faint yellow liquid. ^1H NMR (400 MHz, CDCl_3) δ 5.28 (s, 1H), 4.18 (d, $J = 10.3$ Hz, 1H), 3.58 (t, $J = 6.6$ Hz, 2H), 3.48–3.26 (m, 5H), 3.02 (t, $J = 12.5$ Hz, 1H), 2.96–2.85 (m, 1H), 2.55 (tt, $J = 11.5, 7.0$ Hz, 1H), 2.39–2.28 (m, 1H), 2.04–1.95 (m, 2H), 1.87 (ddt, $J = 13.6, 6.7, 3.6$ Hz, 1H), 1.71 (dq, $J = 10.2, 3.4$ Hz, 2H), 1.67–1.60 (m, 1H), 1.56 (dt, $J = 13.9, 4.4$ Hz, 1H), 1.51 (s, 1H), 1.36–1.28 (m, 3H), 1.28–1.22 (m, 1H), 1.03 (ddd, $J = 15.5, 10.8, 4.2$ Hz, 1H), 0.95 (t, $J = 5.1$ Hz, 4H), 0.88 (s, 12H), 0.85–0.81 (m, 1H), 0.79 (d, $J = 7.1$ Hz, 3H), 0.04 (s, 7H); ^{13}C NMR (100 MHz, CDCl_3) δ 104.0, 91.7, 90.8, 63.0, 60.4, 51.5, 51.3, 45.4, 37.2, 36.0, 34.0, 32.5, 29.1, 28.8, 26.5, 25.8, 25.7, 25.3, 24.6, 23.1, 21.4, 20.1, 18.2, 13.3, –5.4.

4.2.6. Synthesis of (2S)-2-(6-iodohexyl)-4-((3R,5aS,6R,8aS,9R,10R,12aR)-3,6,9-trimethyldecahydro-3H-3,12-epoxy[1,2]dioxepino[4,3-*i*]isochromen-10-yl)thiomorpholine 1,1-dioxide (8). Compound **7 α** (0.9 g, 1.52 mmol), TBAF (1.42 g, 5.43 mmol), THF (45 mL) were added to a flask and stirred overnight at room temperature. Then 100 mL ethyl acetate was added. The mixture was washed with saturated salt solution (2×40 mL), dried over Na_2SO_4 and the solvent removed under diminished pressure. The residue, triphenylphosphine (0.4 g, 1.52 mmol) and imidazole (0.21 g, 3 mmol) were added in 40 mL dry THF and stirred for 5 min, then iodine (0.39 g, 1.52 mmol) was added in three batches and stirred overnight at room temperature. Then the mixture was diluted with 60 mL ethyl acetate and washed with saturated NaHSO_3 25

mL, saturated salt solution (2×25 mL), dried over Na_2SO_4 and the solvent removed under diminished pressure. The residue was purified by chromatography on a silica gel column, eluting with petroleum ether–ethyl acetate 4 : 1. Compound **8** was formed in 45% yield as a faint yellow liquid for two steps. ^1H NMR (400 MHz, CDCl_3) δ 5.28 (s, 1H), 4.19 (d, $J = 10.3$ Hz, 1H), 3.56–3.26 (m, 6H), 3.17 (t, $J = 7.0$ Hz, 2H), 3.00 (dd, $J = 14.4, 10.9$ Hz, 1H), 2.94–2.84 (m, 1H), 2.54 (ddt, $J = 13.1, 7.1, 3.6$ Hz, 1H), 2.34 (ddd, $J = 14.7, 13.3, 4.0$ Hz, 1H), 2.00 (ddt, $J = 14.3, 5.2, 3.3$ Hz, 2H), 1.92–1.77 (m, 3H), 1.71 (ddq, $J = 13.8, 6.5, 3.5$ Hz, 2H), 1.57 (dt, $J = 13.8, 4.4$ Hz, 1H), 1.48 (dt, $J = 7.5, 5.0$ Hz, 1H), 1.45–1.39 (m, 1H), 1.37 (s, 8H), 1.31 (dt, $J = 15.1, 5.0$ Hz, 1H), 1.23 (dt, $J = 11.2, 5.6$ Hz, 1H), 1.02 (ddd, $J = 15.9, 11.5, 4.2$ Hz, 1H), 0.95 (d, $J = 6.2$ Hz, 3H), 0.91 (t, $J = 7.3$ Hz, 1H), 0.79 (d, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 104.2, 91.9, 91.0, 80.1, 60.4, 51.7, 51.4, 45.6, 37.4, 36.1, 34.1, 33.3, 30.1, 29.0, 28.3, 26.5, 25.9, 24.7, 23.2, 21.6, 20.2, 13.5, 7.1.

4.2.7. Synthesis of (2S)-2-(6-(piperazin-1-yl)hexyl)-4-((3R,5aS,6R,8aS,9R,10R,12aR)-3,6,9-trimethyldecahydro-3H-3,12-epoxy[1,2]dioxepino[4,3-*i*]isochromen-10-yl)thiomorpholine 1,1-dioxide (9). Compound **8** (0.46 g, 0.75 mmol), piperazine (0.43 g, 5 mmol), DIPEA (diisopropylethylamine, 0.2 mL), 20 mL THF and 1 mL methanol were added to a flask and stirred within 5 min, then heated to 61 °C, and stirred for another 4 h. Following the addition of saturated sodium carbonate (15 mL) and AcOEt (25 mL), the organic phase was separated, and the aqueous layer was extracted with AcOEt (2×25 mL). The combined extracts were washed with brine (20 mL), dried over anhydrous Na_2SO_4 , and concentrated under reduced pressure. The residue was purified by chromatography on a silica gel column, eluting with DCM–methanol 15 : 1. Compound **9** was formed in 94% yield as a faint yellow gum. ^1H NMR (400 MHz, CDCl_3) δ 5.29 (d, $J = 7.4$ Hz, 1H), 4.18 (d, $J = 10.2$ Hz, 1H), 3.46–3.25 (m, 5H), 3.08–2.95 (m, 1H), 2.89 (t, $J = 4.9$ Hz, 4H), 2.55 (ddd, $J = 10.9, 9.4, 5.8$ Hz, 2H), 2.37 (d, $J = 4.0$ Hz, 4H), 2.35–2.25 (m, 3H), 2.06–1.94 (m, 3H), 1.92–1.81 (m, 2H), 1.71 (dq, $J = 10.2, 3.3$ Hz, 3H), 1.56 (dt, $J = 13.7, 4.3$ Hz, 4H), 1.48 (d, $J = 9.5$ Hz, 6H), 1.30 (dd, $J = 15.6, 10.8$ Hz, 6H), 1.26–1.18 (m, 1H), 1.12 (d, $J = 6.5$ Hz, 1H), 1.06–0.98 (m, 1H), 0.95 (d, $J = 6.2$ Hz, 3H), 0.79 (d, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 104.2, 91.9, 91.0, 80.1, 59.3, 54.5, 53.2, 51.7, 51.4, 50.8, 46.0, 45.6, 37.4, 36.1, 34.2, 29.4, 29.0, 27.2, 26.6, 26.4, 25.9, 24.8, 23.2, 21.6, 20.2, 13.5.

4.2.8. General procedure for synthesis of furan ethers 11a–f. Furan ethers **11a–f** were synthesized according to ref. 25.

4.2.9. General procedure for synthesis of target hybrids 12a–f. Compound **9** (50 mg, 0.088 mmol), furan ethers **11** (0.09 mmol), DIPEA (0.2 mL), and DCM (10 mL) were added to a flask, and stirred for 4 h at room temperature. The reaction mixture was evaporated under reduced pressure and the residue was purified by chromatography on a silica gel column, eluting with petroleum ether–ethyl acetate 25 : 1 to 4 : 1 to give hybrids **12a–f**.

4.2.9.1. 3-Chloro-4-(4-(6-((2S)-1,1-dioxido-4-((3R,5aS,6R,8aS,9R,10R,12aR)-3,6,9-trimethyldecahydro-3H-3,12-epoxy[1,2]dioxepino[4,3-*i*]isochromen-10-yl)thiomorpholin-2-yl)hexyl)piperazin-1-yl)-5-methoxyfuran-2(5H)-one (12a). Hybrid **12a** was formed in 85% yield (99.4% HPLC purity) as a yellow gum.



$[\alpha]_D^{20} +19$ (c 1.0, EtOH); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 5.69 (s, 1H), 5.29 (s, 1H), 4.20 (d, $J = 10.2$ Hz, 1H), 3.73 (s, 4H), 3.56–3.24 (m, 7H), 3.01 (t, $J = 12.6$ Hz, 1H), 2.96–2.80 (m, 1H), 2.65–2.46 (m, 5H), 2.34 (ddd, $J = 18.2, 11.5, 4.5$ Hz, 3H), 2.01 (dt, $J = 14.3, 4.1$ Hz, 2H), 1.88 (dq, $J = 10.2, 3.3$ Hz, 1H), 1.73 (ddt, $J = 13.5, 6.3, 3.3$ Hz, 2H), 1.62–1.19 (m, 18H), 1.10–0.99 (m, 1H), 0.96 (d, $J = 6.1$ Hz, 3H), 0.80 (d, $J = 7.1$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 171.3, 167.9, 153.6, 104.3, 97.5, 92.1, 91.2, 80.3, 60.6, 58.3, 58.3, 54.9, 54.8, 52.8, 51.9, 51.6, 45.8, 37.6, 36.3, 34.3, 29.8, 29.5, 29.2, 26.9, 26.7, 26.1, 24.9, 23.5, 21.7, 20.4, 14.3, 13.6; IR (KBr) ν/cm^{-1} : 2934, 2869, 1744, 1638, 1438, 1301, 1259, 1041, 848, 746; HRMS m/z : calcd for $\text{C}_{34}\text{H}_{55}\text{ClN}_3\text{O}_9\text{S}$ 716.3342, found 716.3337 $[\text{M} + \text{H}]^+$.

4.2.9.2. (5*S*)-3-Chloro-4-(4-(6-((2*S*)-1,1-dioxido-4-((3*R*,5*aS*,6*R*,8*aS*,9*R*,10*R*,12*aR*)-3,6,9-trimethyldecahydro-3*H*-3,12-epoxy[1,2]dioxepino[4,3-*i*isochromen-10-yl]thiomorpholin-2-yl)hexyl)piperazin-1-yl)-5-(((1*S*,2*S*,5*R*)-2-isopropyl-5-methyl-cyclohexyl)oxy)furan-2(5*H*)-one (12*b*). Hybrid 12*b* was formed in 82% yield (98.6% HPLC purity) as a yellow solid. mp 75.6–76.1 °C; $[\alpha]_D^{20} -2$ (c 1.0, EtOH); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.73 (s, 0.2H), 5.78 (s, 0.8H), 5.29 (s, 1H), 4.20 (d, $J = 10.2$ Hz, 1H), 3.82–3.59 (m, 4H), 3.59–3.27 (m, 5H), 3.01 (t, $J = 12.5$ Hz, 1H), 2.95–2.87 (m, 1H), 2.61–2.29 (m, 8H), 2.28–2.12 (m, 1H), 2.02 (q, $J = 8.3, 6.5$ Hz, 3H), 1.89 (ddd, $J = 14.3, 7.0, 3.6$ Hz, 2H), 1.71 (tdd, $J = 17.9, 8.0, 3.7$ Hz, 5H), 1.51 (tdd, $J = 16.6, 11.5, 6.8$ Hz, 5H), 1.43–1.22 (m, 13H), 1.13 (q, $J = 12.2$ Hz, 1H), 0.93 (dt, $J = 15.2, 7.2$ Hz, 11H), 0.78 (dq, $J = 12.1, 6.8, 5.5$ Hz, 7H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 168.4, 155.0, 104.3, 97.3, 92.1, 91.2, 87.6, 81.0, 80.3, 60.7, 58.5, 52.9, 51.9, 51.6, 48.2, 47.7, 45.8, 42.6, 40.5, 37.6, 36.3, 34.3, 34.0, 31.8, 29.8, 29.5, 29.2, 27.1, 26.8, 26.1, 25.2, 24.9, 23.5, 22.8, 22.4, 21.7, 21.3, 20.4, 15.9, 13.6; IR (KBr) ν/cm^{-1} : 2933, 2888, 2870, 1760, 1634, 1455, 1302, 1126, 974, 879, 746; HRMS m/z : calcd for $\text{C}_{43}\text{H}_{71}\text{ClN}_3\text{O}_9\text{S}$ 840.4594, found 840.4596 $[\text{M} + \text{H}]^+$.

4.2.9.3. 3-Chloro-4-(4-(6-((*S*)-1,1-dioxido-4-((3*R*,5*aS*,6*R*,8*aS*,9*R*,10*R*,12*R*,12*aR*)-3,6,9-trimethyldecahydro-3*H*-3,12-epoxy[1,2]dioxepino[4,3-*i*isochromen-10-yl]thiomorpholin-2-yl)hexyl)piperazin-1-yl)-5-(((2*S*)-1,7,7-trimethylbicyclo[2.2.1]heptan-2-yl)oxy)furan-2(5*H*)-one (12*c*). Hybrid 12*c* was formed in 80% yield (96.9% HPLC purity) as a yellow gum. $[\alpha]_D^{20} +10$ (c 1.0, EtOH); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.73 (s, 0.7H), 5.72 (s, 0.2H), 5.29 (s, 1H), 4.19 (d, $J = 10.3$ Hz, 1H), 3.74 (s, 1H), 3.56–3.27 (m, 8H), 3.02 (t, $J = 12.5$ Hz, 1H), 2.90 (d, $J = 11.1$ Hz, 1H), 2.68–2.17 (m, 8H), 2.05–1.98 (m, 2H), 1.98–1.83 (m, 1H), 1.80–1.67 (m, 4H), 1.62–1.42 (m, 4H), 1.43–1.19 (m, 16H), 1.11–1.00 (m, 1H), 0.96 (d, $J = 6.3$ Hz, 5H), 0.92–0.84 (m, 9H), 0.80 (d, $J = 7.1$ Hz, 3H), 0.61 (t, $J = 7.1$ Hz, 1H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 162.9, 161.8, 132.3, 129.1, 104.3, 92.1, 91.2, 83.5, 80.3, 60.7, 58.5, 53.0, 52.0, 51.6, 49.2, 48.1, 45.8, 44.9, 37.6, 36.4, 34.3, 32.1, 29.8, 29.4, 29.2, 28.5, 28.1, 27.2, 27.1, 26.8, 26.5, 26.1, 24.9, 24.4, 23.5, 22.8, 21.8, 20.4, 19.8, 19.0, 13.6; IR (KBr) ν/cm^{-1} : 2932, 2887, 2867, 1758, 1628, 1449, 1377, 1302, 1127, 1040, 960, 847, 744; HRMS m/z : calcd for $\text{C}_{43}\text{H}_{69}\text{ClN}_3\text{O}_9\text{S}$ 838.4438, found 838.4432 $[\text{M} + \text{H}]^+$.

4.2.9.4. 3-Bromo-4-(4-(6-((2*S*)-1,1-dioxido-4-((3*R*,5*aS*,6*R*,8*aS*,9*R*,10*R*,12*aR*)-3,6,9-trimethyldecahydro-3*H*-3,12-epoxy[1,2]dioxepino[4,3-*i*isochromen-10-yl]thiomorpholin-2-yl)

hexyl)piperazin-1-yl)-5-methoxyfuran-2(5*H*)-one (12*d*). Hybrid 12*d* was formed in 80% yield (98.8% HPLC purity) as a yellow solid. mp 78.2–79.2 °C; $[\alpha]_D^{20} +6$ (c 1.0, EtOH); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 5.74 (s, 1H), 5.29 (s, 1H), 4.16 (dd, $J = 29.7, 8.7$ Hz, 1H), 3.90–3.79 (m, 1H), 3.77 (s, 4H), 3.75–3.63 (m, 1H), 3.48 (s, 1H), 3.47–3.36 (m, 3H), 3.33 (d, $J = 14.4$ Hz, 1H), 3.02 (t, $J = 12.8$ Hz, 1H), 2.95–2.87 (m, 1H), 2.64–2.54 (m, 1H), 2.58–2.51 (m, 4H), 2.44–2.35 (m, 2H), 2.32 (dd, $J = 14.2, 4.3$ Hz, 1H), 2.08–1.95 (m, 2H), 1.89 (ddd, $J = 14.0, 6.8, 3.4$ Hz, 1H), 1.73 (ddt, $J = 13.5, 6.2, 3.4$ Hz, 2H), 1.61–1.44 (m, 6H), 1.42–1.17 (m, 12H), 1.09–0.98 (m, 1H), 0.96 (d, $J = 6.2$ Hz, 3H), 0.80 (d, $J = 7.1$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 171.3, 168.7, 156.6, 104.3, 97.8, 92.1, 91.2, 80.3, 64.0, 60.7, 60.5, 58.4, 53.0, 51.9, 51.6, 47.6, 45.8, 37.6, 36.3, 34.3, 29.4, 29.2, 27.0, 26.8, 26.5, 26.1, 24.9, 23.5, 21.7, 20.4, 15.1, 13.6; IR (KBr) ν/cm^{-1} : 2934, 2892, 2872, 1762, 1636, 1454, 1303, 1127, 974, 646; HRMS m/z : calcd for $\text{C}_{34}\text{H}_{55}\text{BrN}_3\text{O}_9\text{S}$ 760.2837, found 760.2833 $[\text{M} + \text{H}]^+$.

4.2.9.5. (5*S*)-3-Bromo-4-(4-(6-((2*S*)-1,1-dioxido-4-((3*R*,5*aS*,6*R*,8*aS*,9*R*,10*R*,12*aR*)-3,6,9-trimethyldecahydro-3*H*-3,12-epoxy[1,2]dioxepino[4,3-*i*isochromen-10-yl]thiomorpholin-2-yl)hexyl)piperazin-1-yl)-5-(((1*S*,2*S*,5*R*)-2-isopropyl-5-methyl-cyclohexyl)oxy)furan-2(5*H*)-one (12*e*). Hybrid 12*e* was formed in 78% yield as a yellow gum. $[\alpha]_D^{20} -3$ (c 1.0, EtOH); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 5.79 (s, 1H), 5.28 (s, 1H), 4.19 (d, $J = 10.2$ Hz, 1H), 3.86–3.50 (m, 4H), 3.47–3.28 (m, 4H), 3.01 (t, $J = 12.6$ Hz, 1H), 2.90 (d, $J = 11.5$ Hz, 1H), 2.53 (ddt, $J = 21.4, 6.8, 3.9$ Hz, 4H), 2.43–2.28 (m, 4H), 2.25–2.13 (m, 1H), 2.04–1.84 (m, 3H), 1.75–1.63 (m, 4H), 1.59–1.43 (m, 6H), 1.35 (d, $J = 12.6$ Hz, 10H), 1.28–1.05 (m, 6H), 1.03–0.85 (m, 12H), 0.80–0.73 (m, 7H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 168.7, 157.6, 104.2, 98.2, 92.0, 91.1, 80.7, 80.2, 73.8, 58.4, 52.8, 51.8, 51.5, 48.2, 47.8, 45.7, 42.5, 37.5, 36.2, 34.2, 34.0, 31.7, 31.5, 29.4, 29.1, 27.0, 26.7, 26.0, 25.1, 24.8, 23.4, 22.8, 22.3, 21.6, 21.2, 20.3, 19.4, 15.8, 13.5; IR (KBr) ν/cm^{-1} : 2932, 1761, 1635, 1454, 1302, 1126, 934, 879; HRMS m/z : calcd for $\text{C}_{43}\text{H}_{71}\text{BrN}_3\text{O}_9\text{S}$ 884.4089, found 884.4088 $[\text{M} + \text{H}]^+$.

4.2.9.6. (5*S*)-3-Bromo-4-(4-(6-((*S*)-1,1-dioxido-4-((3*R*,5*aS*,6*R*,8*aS*,9*R*,10*R*,12*R*,12*aR*)-3,6,9-trimethyldecahydro-3*H*-3,12-epoxy[1,2]dioxepino[4,3-*i*isochromen-10-yl]thiomorpholin-2-yl)hexyl)piperazin-1-yl)-5-(((2*S*)-1,7,7-trimethylbicyclo[2.2.1]heptan-2-yl)oxy)furan-2(5*H*)-one (12*f*). Hybrid 12*f* was formed in 81% yield (96.8% HPLC purity) as a yellow solid. mp 77.9–78.9 °C; $[\alpha]_D^{20} +11$ (c 1.0, EtOH); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.95 (s, 0.3H), 5.73 (d, $J = 10.1$ Hz, 0.4H), 5.29 (s, 1H), 4.20 (d, $J = 10.3$ Hz, 1H), 3.79–3.73 (m, 2H), 3.67 (s, 1H), 3.54–3.24 (m, 6H), 3.02 (t, $J = 12.7$ Hz, 1H), 2.91 (d, $J = 11.3$ Hz, 1H), 2.63–2.25 (m, 8H), 2.03 (d, $J = 6.9$ Hz, 3H), 1.94–1.83 (m, 2H), 1.79–1.65 (m, 4H), 1.58 (dd, $J = 13.1, 4.5$ Hz, 1H), 1.53–1.46 (m, 7H), 1.36 (d, $J = 11.3$ Hz, 8H), 1.33–1.21 (m, 5H), 1.05 (ddt, $J = 18.1, 13.4, 5.9$ Hz, 2H), 0.96 (d, $J = 6.3$ Hz, 3H), 0.87 (dd, $J = 11.0, 5.2$ Hz, 8H), 0.81 (t, $J = 10.0$ Hz, 3H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 163.3, 162.1, 136.2, 118.9, 104.3, 92.0, 91.1, 83.5, 80.2, 60.7, 58.5, 58.4, 53.0, 52.4, 51.9, 51.6, 49.5, 49.2, 48.0, 47.6, 45.7, 45.0, 44.9, 37.6, 36.3, 34.3, 29.4, 29.1, 28.1, 28.0, 27.1, 26.8, 26.0, 24.9, 23.4, 21.7, 20.4, 19.8, 18.9, 14.2, 13.6; IR (KBr) ν/cm^{-1} : 2932, 2890, 2871, 1760, 1628, 1453, 1304, 1126, 879, 744; HRMS m/z : calcd for $\text{C}_{43}\text{H}_{69}\text{BrN}_3\text{O}_9\text{S}$ 882.3932, found 882.3921 $[\text{M} + \text{H}]^+$.



4.3. Pharmacology

4.3.1. Human liver cancer cell lines SMMC-7721. SMMC-7721 cells (1×10^4 in 100 μL) were seeded on 96 plates in triplicate. Following a 24 h culture at 37 $^\circ\text{C}$, the medium was replaced with fresh medium at various concentrations (10, 25, 50, 100 and 200 $\mu\text{g mL}^{-1}$) of hybrids **12a–f**, DHA, vincristine (VCR), and cytosine arabinoside (ARA) in a final volume of 100 μL . At the same time, the drug-free medium negative control well and the solvent control well were set with the same volume of dimethyl sulfoxide (DMSO). Cells were respectively incubated at 37 $^\circ\text{C}$ for 24 h. Then, 10 μL of 3-(4,5-dime-thylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) (2 mg mL^{-1} in a phosphate buffer solution; PBS) was added to each well, incubated for an additional 4 h, centrifuged at 1000 rpm for 10 min, and then the medium was removed. MTT formazan precipitates were dissolved in 150 μL of DMSO, shaken mechanically for 10 min and then read immediately at 568 nm using a plate reader (Multiskan MK3, Thermo Fisher Scientific, USA).

$$\text{Cell inhibition rate} = [A_{568} (\text{negative control well}) - A_{568} (\text{dosing well})] / A_{568} (\text{negative control well}) \times 100\%$$

4.3.2. Human benign liver cell lines LO2. The method was similar to that of SMMC-7721 cells described above.

4.3.3. Flow cytometry analysis. MHCC97H cells were seeded in on 96 plates in triplicate. Following a 24 h culture at 37 $^\circ\text{C}$, the medium was replaced with fresh medium at various concentrations (0.01, 1 and 25 $\mu\text{g mL}^{-1}$) of hybrid **12a**. At the same time, drug-free medium negative control well and solvent control well were set with the same volume of dimethyl sulfoxide (DMSO). Cells were respectively cultivated at 37 $^\circ\text{C}$ for 48 h, then 0.25% trypsin without ethylene diamine tetra-acetic acid (EDTA) was added to each well. After trypsinisation, the treated cells were stained using Annexin V-FITC/PI apoptosis detection kit (NanJing KeyGen Biotech Co., Ltd, CHN) according to the manufacturer's instructions. After incubation at room temperature for 5–15 min in the dark, the apoptotic cells were immediately analysed by flow cytometry (cyto Flex, BECKMAN COULTER, USA).

4.3.4. The general method for testing the activity of hybrid 12a. Cells (SMMC-7721, MHCC97H, HCT116, CCD18CO, MCF-7, MCF-10A, DU145 and RWPE-1) (1×10^4 in 100 μL) were respective seeded on 96 plates in triplicate. Following a 24 h culture at 37 $^\circ\text{C}$, the medium was replaced with fresh medium at various concentrations (0.01, 0.1, 1, 10 and 25 $\mu\text{g mL}^{-1}$) of hybrid **12a** in a final volume of 100 μL . At the same time, the drug-free medium negative control well and the solvent control well were set with the same volume of dimethyl sulfoxide (DMSO). Cells were respectively incubated at 37 $^\circ\text{C}$ for 48 h. Then, 10 μL of MTT (2 mg mL^{-1} in a phosphate buffer solution; PBS) was added to each well, incubated for an additional 4 h, centrifuged at 1000 rpm for 10 min, and then the medium was removed. MTT formazan precipitates were dissolved in 150 μL of DMSO, shaken mechanically for 10 min and then read

immediately at 568 nm using a plate reader (Multiskan MK3, Thermo Fisher Scientific, USA).

4.3.5. The general method for testing the activity of hybrid 12a in Fe^{2+} environment. Cells (SMMC-7721 and MHCC97H) (1×10^4 in 100 μL) were respectively seeded on 96 plates in triplicate. Following a 12 h culture at 37 $^\circ\text{C}$, add 100 $\mu\text{mol L}^{-1}$ FeSO_4 culture solution, and culture other 12 h at 37 $^\circ\text{C}$. The medium was replaced with fresh medium at various concentrations (0.01, 0.1, 1, 10 and 25 $\mu\text{g mL}^{-1}$) of hybrid **12a** in a final volume of 100 μL . At the same time, the drug-free medium negative control well and the solvent control well were set with the same volume of dimethyl sulfoxide (DMSO). Cells were respectively incubated at 37 $^\circ\text{C}$ for 48 h. Then, 10 μL of MTT (2 mg mL^{-1} in a phosphate buffer solution (PBS)) was added to each well, incubated for an additional 4 h, centrifuged at 1000 rpm for 10 min, and then the medium was removed. MTT formazan precipitates were dissolved in 150 μL of DMSO, shaken mechanically for 10 min and then read immediately at 568 nm using a plate reader (Multiskan MK3, Thermo Fisher Scientific, USA).

Conflicts of interest

There are no conflicts to declare.

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