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Vlasoulides A and B, a pair of neuroprotective C₃₂ dimeric sesquiterpenes with a hexacyclic 5/7/5/5/(5)/7 carbon skeleton from the roots of *Vladimiria souliei*†

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Vlasoulides A and B (**1** and **2**), a pair of epimeric C₃₂ sesquiterpene lactone dimers, featuring a 5/7/5/5/(5)/7 ring system were isolated from the roots of *Vladimiria souliei*. Their chemical structures were determined by comprehensive analysis of spectroscopic data, including HRESIMS and 1D and 2D NMR spectroscopic data. Their absolute configurations were established by Mosher's method and ECD experiments. Furthermore, biological studies showed that compound **1** showed prominent neuroprotective effects against glutamate-induced neurotoxicity in PC-12 cells, with EC₅₀ values of 13.54 ± 0.33 μM, while, the EC₅₀ value of compound **2** is greater than 30 μM.

Introduction

The large genus *Vladimiria* belonging to the family Asteraceae comprises about 12 species. Most of them are widely distributed in East Asia, especially in the Sichuan Province, China.^{1,2} The roots are usually applied in traditional Chinese medicine to treat a number of diseases, showing antitumor, analgesic, anti-inflammatory, gastric ulcer resistant and antibacterial activities.^{3–6} Furthermore, according to previous phytochemical studies, they were found to contain many sesquiterpenes, steroids, phenylpropanoids, flavonoids, and triterpenes.^{7–9} Aiming to search for new and rare SLDs with unique skeletons and novel bioactivities from this genus *Vladimiria*, we investigated the chemical constituents of the roots of *Vladimiria souliei*. Over the past few years, some rare sesquiterpene lactone dimers (SLDs), with influence on NO production and the activation of NF-κB pathway,^{10–14} were isolated from the title plant. At the same time, we also have separated some rare dimeric sesquiterpenes exhibiting potential neuroprotection activity from *Vladimiria souliei* in our previous study.^{15,16} Therefore, as a continuing investigation on this plant, two C₃₂ sesquiterpene dimers, vlasoulides A and B (**1** and **2**), were isolated from the same plant. Herein, we describe the isolation, structural elucidation and neuroprotective activities of two new compounds (Fig. 1).

Experimental

General experimental procedure

Column chromatography (CC): silica gel H (10–40 μm; Marine Chemical Factory, Qingdao, P. R. China); MCI gel CHP-20P; Sephadex LH-20 (Pharmacia Fine Chemicals, Piscataway, NJ, USA); RP-C₁₈ gel (40–63 μm; Daiso, Co, Japan) were used for column chromatography. TLC: silica gel plates, visualization by spraying with 10% H₂SO₄ in EtOH and dragendorff's reagent. Semi-preparative HPLC: Agilent 1260 series with a Zorbax SB-C₁₈ (5 μM, 9.4 mm × 25 cm) column. Melting point: X-4B apparatus and was uncorrected. IR spectra: Bruker Vector 22 (KBr pellets). Optical rotation: Autopol VI (serial no. 90079, manufactured by Rudolph Research Analytical, Hackettstown, NJ). UV spectra were obtained by the DAD detector of HPLC.

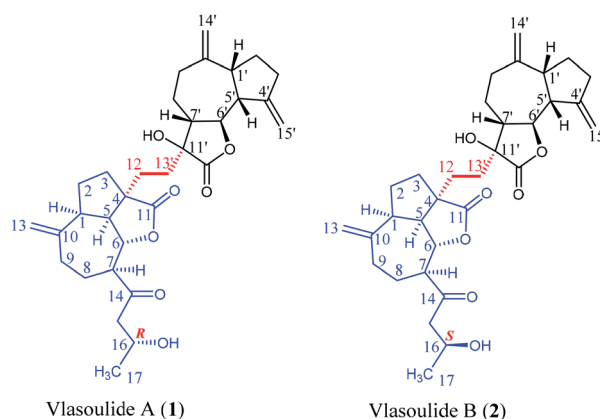


Fig. 1 Chemical structures of **1** and **2**.

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(Agilent 1260). NMR Spectra: Bruker Ascend-500 spectrometer (500 MHz); δ in ppm with SiMe₄ as internal standard. MS: Agilent MSD-Trap-XCT (for ESI) and Q-Tof micro mass spectrometer (for HR-ESI).

Plant material

The whole plant of *Vladimiria souliei* was collected at Dajin, Sichuan Province of China, in October, 2019, and authenticated by Prof. Bao-Kang Huang of Second Medical Military University. Currently A voucher specimen (No. 20191001) is deposited in School of Pharmacy, Second Military Medical University.

Extraction and isolation

The dried and chipped roots of *V. souliei* (50.0 kg) were extracted by maceration with 95% ethanol overnight at room temperature (3 × 60 L). After remove of solvent, the ethanol extract (5.60 kg) was partitioned between water and petroleum ether (PE)/ethyl acetate (EtOAc), successively, to give PE, EtOAc and water extracts. EtOAc extract (1.15 kg) was segmented by MCI column chromatography (MeOH/H₂O, 30 : 70 to 100 : 0) to give 8 fractions (Fr. 1–8). Fraction 5 (52.5 g) was further isolated by ODS column chromatography (MeOH/H₂O, 30 : 70 to 90 : 10) to obtain 8 subfractions (Fr. 5.1–5.8). Subfraction 5.4 (5.5 g) was further purified by Sephadex LH-20 column chromatography (PE : EtOAc : MeOH, 10 : 10 : 1) to give 6 subfractions (Fr. 5.5.1–5.5.6). Subfraction 5.5.4 (252 mg) was purified by semi-preparative RP-C₁₈ HPLC (CH₃CN/H₂O, 70 : 30) to produce compounds **1** (7.2 mg) and **2** (6.5 mg). Above all, compounds **1** (7.2 mg) and **2** (6.5 mg) were obtained.

Compound characterization of **1** and **2**

Vlasoulide A (1). White powder; $[\alpha]_D^{25} -1.00$ (*c* 0.08, CH₃OH); UV (CH₃CN/H₂O) λ_{\max} 210; IR (KBr) ν_{\max} 3438, 3079, 2933, 2869, 1770, 1710, 1639, 1448, 1382, 1351, 1268, 1222, 1153, 1272, 1020, 995, 896, cm⁻¹; ¹H and ¹³C-NMR data (600 MHz/150 MHz), see Table S1;† ESIMS *m/z* 561.4 ([M + Na]⁺); positive HRESIMS *m/z* 561.2828 ([M + Na]⁺, calcd 561.2823).

Vlasoulide B (2). White powder; $[\alpha]_D^{25}$ 0.00 (*c* 0.05, CH₃OH); UV (CH₃CN/H₂O) λ_{\max} 210; IR (KBr) ν_{\max} 3440, 3079, 2929, 2869, 1772, 1752, 1639, 1448, 1400, 1382, 1353, 1282, 1257, 1224, 1068, 1018, 995, 896, cm⁻¹; ¹H and ¹³C-NMR data (600 MHz/150 MHz), see Table S1;† ESIMS *m/z* 561.5 ([M + Na]⁺); positive HRESIMS *m/z* 561.2829 ([M + Na]⁺, calcd 561.2823).

(R)-and (S)-MTPA esters of compounds **1** and **2**

To each compounds **1** and **2** (each 1.5 mg) in pyridine-*d*₅ (130 μ L) was separately added (R)-(–)-MTPA (5 μ L) and (S)-(+)-MTPA (5 μ L) at room temperature, followed by stirring at 40 °C for 8 h, and each reaction mixture was transferred into a 1.7 mm NMR tube.

(R)-MTPA ester of 1. ¹H NMR (pyridine-*d*₅, 600 MHz): δ_H 1.20 (3H, d, H₃-17), 2.88 (1H, m H-15a), 2.91 (1H, m, H-15b), 4.30 (1H, t, H-6), 2.91 (1H, m, H-7), 2.21 (1H, m, H-8a), 1.58 (1H, m, H-8b).

(S)-MTPA ester of 1. ¹H NMR (pyridine-*d*₅, 600 MHz): δ_H 1.32 (3H, d, H₃-17), 2.83 (1H, m H-15a), 2.85 (1H, m, H-15b), 4.20 (1H, t, H-6), 2.88 (1H, m, H-7), 2.19 (1H, m, H-8a) 1.56 (1H, m, H-8b).

(R)-MTPA ester of 2. ¹H NMR (pyridine-*d*₅, 600 MHz): δ_H 1.32 (3H, d, H₃-17), 2.71 (1H, m H-15a), 2.74 (1H, m, H-15b), 4.21 (1H, t, H-6), 2.93 (1H, m, H-7), 2.14 (H, m, H-8a) 1.54 (1H, m, H-8b).

(S)-MTPA ester of 2. ¹H NMR (pyridine-*d*₅, 600 MHz): δ_H 1.22 (3H, d, H₃-17), 2.77 (1H, m H-15a), 2.78 (1H, m, H-15b), 4.27 (1H, t, H-6), 2.98 (1H, m, H-7), 2.16 (1H, m, H-8a), 1.55 (1H, m, H-8b).

Neuroprotection assay

The PC-12 cells were purchased from Shanghai Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences (Shanghai, China) and cultured in Dulbecco's modified eagle's medium (DMEM) containing 10% FBS supplemented with 100 μ g mL⁻¹ penicillin and 100 μ g mL⁻¹ streptomycin in a humidified atmosphere containing 5% CO₂ at 37 °C. Compounds **1** and **2** and vitamin E were dissolved in dimethyl sulfoxide (DMSO) and freshly prepared each time before use. Then, the PC-12 cells were seeded in 96-well culture plates at 8 × 10³ cells per mL at 37 °C for 12 h. Then the cells were incubated with glutamate for an additional 24 h and the drugs (10, 15, 20 μ M, respectively) were pretreated for 1 h before treated with glutamate. Cell viability was determined by the CCK8 assay, after treatment, 10 μ L of CCK8 were added to each well and incubated at 37 °C for 4 h. The optical density (OD) was spectrophotometrically measured at 450 nm (CCK8) using a microplate reader, respectively (BioTek Instruments, Inc.).

Results and discussion

Vlasoulide A (**1**) was isolated as white power, possessing the molecular formula C₃₂H₄₂O₇ determined by positive HRESIMS *m/z* 561.2828 ([M + Na]⁺, calcd 561.2823), indicating 12 degrees of unsaturation.

Comprehensive analysis of the ¹H and ¹³C NMR spectra of **1**, 32 carbon signals of the ¹³C NMR spectrum were classified by HSQC spectrum as one CH₃, fourteen CH₂ (including three sp² carbons), nine CH (two oxygenated ones as well as one hydroxy methine), and eight quaternary carbons (assigned as two carbonyl groups, two sp³ and six sp² carbons respectively) (ESI, Tables S1†). Therefore, through a detailed analysis of 1D NMR spectra of **1** indicated that the compound **1** should be a dimeric sesquiterpene lactone.

The planar structure of **1** was determined by the analysis of ¹H–¹H COSY and HMBC spectra. Two proton-bearing structural fragments: H-5/H-6/H-7/H₂-8/H₂-9 and H₂-15/H-16/H₃-17 were observed by analyzing the ¹H–¹H COSY spectra, together with the HMBC correlations of H₂-13/C-1, C-9 and C-10; H-5/C-3 and C-12; H-7/C-14 and C-15 as depicted with arrows from H to C. The above conjectured that the unit A should be deduced as a ring-opening guaianolide moiety. Furthermore, a five-membered ring newly generated fused with the seven-



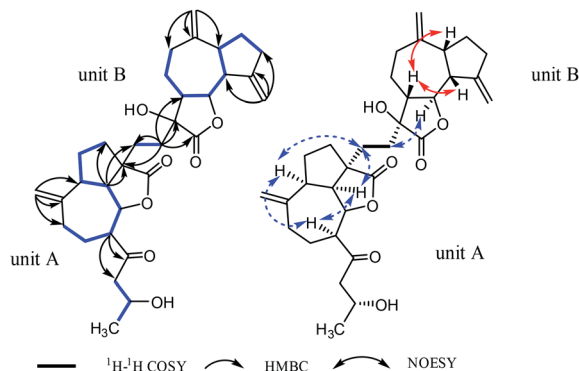


Fig. 2 Selected NMR correlations of **1**.

membered ring and the lactone ring in unit A at C-1, C-4 and C-5 which revealed by the ^1H - ^1H COSY correlations of $\text{H}_2\text{-3}/\text{H}_2\text{-2}/\text{H-1}/\text{H-5}$ and the HMBC cross-peaks from H-5 (δ_{H} 2.78) to C-3 and C-4 (Fig. 2). Meanwhile, the structure of unit B was further determined to be a guaianolide moiety, confirmed by ^1H - ^1H COSY correlations of $\text{H}_2\text{-3'}/\text{H}_2\text{-2'}/\text{H-1'}/\text{H-5'}/\text{H-6'}/\text{H-7'}/\text{H}_2\text{-8'}/\text{H}_2\text{-9'}$ as well as the HMBC correlations of $\text{H}_2\text{-14'}/\text{C-1'}$, C-9' and C-10'; $\text{H}_2\text{-15'}/\text{C-3'}$, C-4' and C-5'; $\text{H}_2\text{-13'}/\text{C-7'}$, C-11' and C-12' (Fig. 2). Finally, the units A and B were linked directly *via* a C-C bond between C-13' and C-12 according to the ^1H - ^1H COSY correlations of $\text{H}_2\text{-13'}/\text{H}_2\text{-12}$, combine with the HMBC correlations of $\text{H}_2\text{-13'}$ (δ_{H} 1.81, 1.69, 1H, respectively)/C-12 (δ_{C} 30.5), C-4 (δ_{C} 54.3), C-7' (δ_{C} 48.7), C-11' (δ_{C} 76.7) and C-12' (δ_{C} 176.8). Thus, the planar structure of **1** was determined as shown in Fig. 1.

The relative configuration of **1** was established by the NOESY spectrum. H-1, H-5, H-7 and H-1', H-5', H-7' in units A and B were on the same face and assigned as α -orientation and β -oriented in **1** based on the similar NOESY correlations of H-1/H-5/H-7 and H-1'/H-5'/H-7'. And, the H-6/H-7 and H-6'/H-7' were in the *trans*-form due to the large coupling constant between H-6/H-7 ($J = 9.8$ Hz) and H-6'/H-7' ($J = 9.5$ Hz) (Fig. 2). In addition, the relative configuration of C-11' and C-4 were resolved by the

NOESY correlations of H-6'/H₂-13' and H₂-12/H-1/H-5 (Fig. 2), exhibiting that the CH₂-13' and CH₂-12 were α -oriented. At the same time, by comparing the calculated and experimental ECD spectra, we found that the experimental ECD spectrum of compound **1** (Fig. 3A) was in good accordance with the calculated curve for the (1*S*,4*R*,5*S*,6*S*,7*S*,16*R*,1'*R*,5'*R*,6'*R*,7'*R*,11'*S*) stereoisomer. Additionally, the observed $\Delta\delta_{\text{H}(S-R)}$ values of the (*S*)- and (*R*)-MTPA esters established the absolute configuration of C-16 in **1** as *R*¹⁷ (Fig. 4), which was consistent with the ECD results. Thus, the structure and absolute configuration of the vlasoulide A (**1**) was fully defined as shown in Fig. 1.

Interestingly, vlasoulide B (**2**), a 16-epimer of **1**, was also an optically active white powder. HRESIMS data at m/z 561.2829 ($[\text{M} + \text{Na}]^+$, calcd 561.2823) of **2** implied that its molecular formula was $\text{C}_{32}\text{H}_{42}\text{O}_7$, same as **1**. The ^1H and ^{13}C NMR data of **2** resembled those of compound **1** (ESI, Tables S1†), suggesting the presence of ring-opening guaianolide and guaianolide units. The analysis of its 2D NMR data (^1H - ^1H COSY and HMBC) revealed that the two compounds had the same 2D structure. Furthermore, inspecting the differences between **1** and **2** on NMR data we found the most obvious change was the C-15 and H₂-15, the chemical shift of C-15 was down field shifted from δ_{C} 50.5 in **1** to δ_{C} 51.6 in **2**, meanwhile, an obvious up field shifted of H₂-15 from δ_{H} 2.67, 2.66 in **1** to δ_{H} 2.63, 2.62 in the ^1H NMR was observe. Thus, these two new compounds represent a pair of stereoisomers possessing opposite configuration at C-16, which was identical to the $\Delta\delta_{\text{H}(S-R)}$ results (Fig. 4). At the same time, the experimental ECD spectrum of **2** also fits well with the calculated spectrum of (1*S*,4*R*,5*S*,6*S*,7*S*,16*S*,1'*R*,5'*R*,6'*R*,7'*R*,11'*S*) **2** (Fig. 3B). Therefore, the structure of vlasoulide B (**2**) was established as shown in Fig. 1.

All the isolated compounds were evaluated for their neuroprotective effects against glutamate-induced neurotoxicity in PC-12 cells using CCK8 assay. Compound **1** showed prominent neuroprotective activity against glutamate-induced neurotoxicity in PC-12 cells, with EC_{50} value of 13.54 ± 0.33 μM . While, the EC_{50} values of the compound **2** is greater than 30 μM .

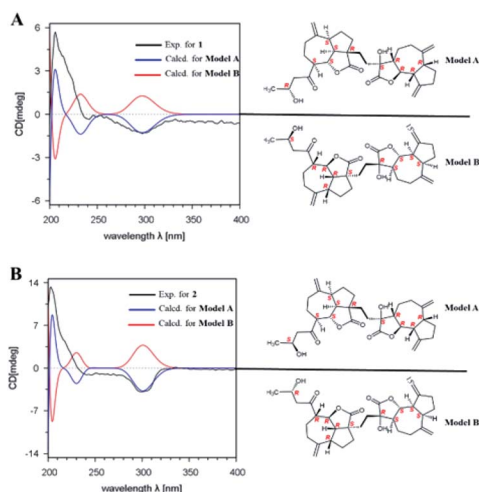


Fig. 3 Experimental and calculated ECD spectra of **1** (A) and **2** (B).

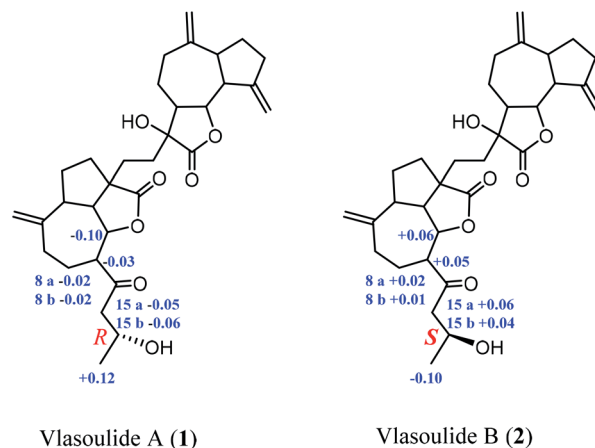


Fig. 4 Chemical shift differences ($\Delta\delta_{\text{H}(S-R)}$) between (*S*)-MTPA and (*R*)-MTPA.



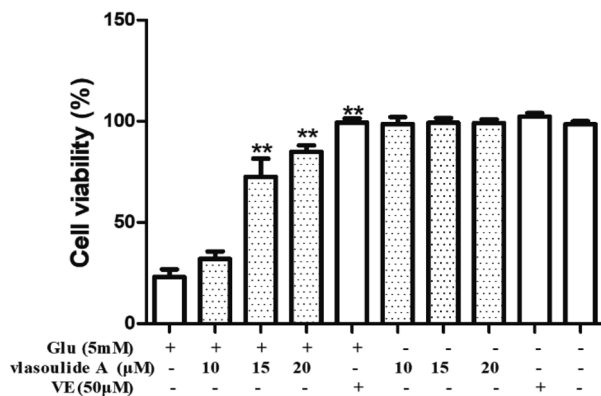


Fig. 5 PC-12 cells were pre-treated with compound 1 and 50 μ M Vitamin E for 1 h, and then co-cultured with or without glutamate (5 mM) for 24 h. Vitamin E as a positive control. The cell viability was detected using CCK8 assay.

Meanwhile, the neuroprotective effects were dose dependent and there were no significant adverse effects on the growth of PC-12 cells with compound 1 solo treatment as shown in Fig. 5. Collectively, these results suggest that compound 1 showed significant neuroprotective activity against glutamate-induced neurotoxicity in PC-12 cells at concentration of 20 μ M.

Conclusions

In conclusion, vlasoulides A and B (1 and 2), two rare and original C₃₂ sesquiterpenoid lactone dimers comprising of two sesquiterpene lactone units, have been isolated and elucidated for the first time from *Vladimiria souliei*. Moreover, compound 1 showed neuroprotective activity against glutamate-induced neurotoxicity in PC-12 cells, with EC₅₀ value of 13.54 ± 0.33 μ M. These results will supply scientific foundation for the scientific research of this plant, as well as might be greatly useful for studying on neuroprotection.

Conflicts of interest

The authors declare that there are no conflicts of interest.

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