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Determination of 12 insect growth regulator residues in foods of different matrixes by modified QuEChERS and UPLC-MS/MS†

 Ke Wang,^{*ab} Lingzhi Zhao,^a Can Zhang,^c Hong Zhang^{ab} and Kaoqi Lian^{id}^{*c}

An analytical method was developed and validated for the simultaneous determination of 12 insect growth regulators (IGRs) (buprofezin, cyantranilprole, flubendiamide, flonicamid, tolfenpyrad, chlorantranilprole, RH-5849, methoxyfenozide, chromafenozide, tebufenozide, pyriproxyfen and fenoxycarb) in foods collected from different matrixes by modified QuEChERS and ultraperformance liquid chromatography-tandem mass spectrometry (UPLC-MS/MS). The samples were ultrasonically extracted with acetonitrile containing 0.5% formic acid, and different QuEChERS purification conditions were optimized for different matrixes (vegetable oil, fruit and tea). 12 IGRs were separated on a Plus C₁₈ column, and detected by MS/MS under multiple reaction monitoring (MRM) mode. The developed method was validated in terms of linearity, matrix effect, accuracy and precision. Acceptable recoveries of IGRs in three different substrates (vegetable oil, tea and fruit) at three spiked levels were in the range of 65.47–95.17%, 80.55–110.15%, and 62.02–96.50%, respectively, with RSDs less than 11.58%. The method showed a good linearity ($R^2 \geq 0.9994$) for all analytes in the range of 0.2–200 $\mu\text{g L}^{-1}$. The LODs (S/N = 3) and LOQs (S/N = 10) of the method were 0.04–0.40 $\mu\text{g kg}^{-1}$, and 0.13–1.24 $\mu\text{g kg}^{-1}$, respectively. Owing to the advantages of simple operation, high accuracy and sensitivity, this method is suitable for the rapid and simultaneous detection of 12 IGRs in vegetable oil, tea and fruit.

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1. Introduction

Insect growth regulators (IGRs) are chemicals that disrupt the life cycle of insects and lead to their death. According to the insecticidal mechanism, IGRs are divided in three major classes including juvenile hormone analogs, ecdysteroid agonists, and chitin synthase inhibitors.^{1,2} Because of their mode of action, IGRs can efficiently control many arthropod pests in many crops (fruits and vegetables, tea, cereals, *etc.*), and they are particularly effective against many lepidopteran pests while exhibiting good selectivity with low toxicity to nontarget arthropods. They act by hindering the biosynthesis and deposition of chitin, reducing the hardness of new epidermis, and hindering the growth and development of chitin.^{3,4} IGRs are advantageous over conventional insecticides as they are more potent, less toxic to mammals, and biodegradable in soil and water.⁵ Pyriproxyfen and fenoxycarb are two new insecticides of

the class juvenile hormone analogs; their characteristics include high efficiency, low dosage, and long duration.^{2,6,7} RH-5849, tebufenozide, methoxyfenozide and chromafenozide are new generation of diacylhydrazide IGRs and ecdysone analogs. They mainly induce lepidoptera pests to molt and mature prematurely by reducing or inhibiting the feeding ability of larva and adult, resulting in incomplete development of insects and reduction in oviposition; thus, the pests die in a few days, thus achieving insecticidal effect by hindering insect reproduction.^{8,9} Buprofezin, chloroanilide, flubendiamide, flonicamid, tolfenpyrad, and cyananilide are all chitin synthesis inhibitors that activate ryanodine receptors, stimulating the uncontrolled release of calcium ions from the sarcoplasmic reticulum of muscle cells and resulting in feeding cessation, lethargy, muscle paralysis, and death.^{10,11} With the wide application of IGRs as insecticides, it has been found that RH-5849 is a moderately toxic insecticide, with a certain degree of damage to the genetic material of human peripheral blood lymphocytes;¹² excessive use of fenoxycarb will produce a certain reproductive toxicity to animals and lead to anemia in mammals.^{13,14}

To ensure food safety and human health, many countries and regions have set the maximum residue limit (MRL) standards for IGRs in food. The United States, EU, and Japan have published thousands of maximum residue limits (MRLs) for pesticides.¹⁵ The European Union has set the MRLs for

^aShijiazhuang Center for Disease Control and Prevention, Shijiazhuang 050011, China. E-mail: wkecdc@163.com

^bShijiazhuang Technology Innovation Center for Chemical Poison Detection and Risk Early Warning, Shijiazhuang 050011, China

^cHebei Key Laboratory of Environment and Human Health, School of Public Health, Hebei Medical University, Shijiazhuang, 050017, China. E-mail: liankq@hebmh.edu.cn

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methoxyfenozide, chromafenozide and flonicamid in tea as 50, 50, and 100 $\mu\text{g kg}^{-1}$, respectively.¹⁶ The Japanese positive list provides the MRLs for methoxyfenozide, chromafenozide, tebufenozide in vegetables as 30 000, 5000, and 20 000 $\mu\text{g kg}^{-1}$, respectively.¹⁷ Chinese national food safety standard GB 2763-2019 stipulates the MRLs for buprofezin, tebufenozide, tolfenpyrad, cyantraniliprole, pyriproxyfen, chlorantraniliprole, and flonicamid in foods (fruits, vegetables, tea, cereals, *etc.*) as 10–10 000, 10–20 000, 500–50 000, 30–20 000, 10–3000, 10–40 000 and 200–1000 $\mu\text{g kg}^{-1}$, respectively.¹⁸ Therefore, it is very important to determine the level of IGR residues in agricultural products and monitor the safety of agricultural products.

Several analytical methods have been developed to determine IGRs by in food and environmental samples by high-performance liquid chromatography (HPLC),^{19,20} HPLC coupled with tandem mass spectrometry (HPLC-MS/MS),^{4,21} gas chromatography (GC),¹⁰ gas chromatography coupled with tandem mass spectrometry (GC-MS/MS),²² and enzyme-linked immunosorbent assays (ELISAs).²³ Among them, because of the advantages of HPLC-MS/MS and ultraperformance liquid chromatography with tandem mass spectrometry (UPLC-MS/MS) in multiple reaction monitoring (MRM) mode, such as high sensitivity, selectivity, and specificity, it has been widely applied to detect pesticide residues. For sample preparation, various pretreatment methods were used for pesticide analysis, such as liquid–liquid extraction (LLE),^{22,24} solid-phase extraction (SPE),²⁵ magnetic solid-phase extraction (MSPE),^{26,27} dispersive liquid–liquid microextraction (DLLME)²⁸ and solid-phase microextraction (SPME),²⁹ and so on. Compared with other pretreatment technique, quick, easy, cheap, rugged, effective, and safe (QuEChERS)^{21,30–32} method has the advantages of simple and fast operation, requirement of less solvent in the extraction process, and less environmental pollution. This is followed by a salting-out step, causing the partitioning of two liquid phases. A clean-up step, usually dispersive solid-phase extraction, is then added to remove interferences.

Currently, studies on IGRs have focused on samples of fruits, vegetables, and grains, often testing for one or more pesticides, there are few reports of simultaneous detection of multiple IGRs in different matrices. For QuEChERS pretreatment, although the standard operating procedures were used in most cases, further study should be conducted on the pretreatment modification and quantitative evaluation, especially for complex sample matrix. The aim of this study was to develop simultaneous determination of 12 IGRs involving four ecdysteroid agonists (RH-5849, methoxyfenozide, chromafenozide, and tebufenozide), six chitin synthase inhibitors (buprofezin, cyantraniliprole, flubendiamide, flonicamid, tolfenpyrad, and chlorantraniliprole), and two juvenile hormone analogs (pyriproxyfen and fenoxycarb) in vegetable oil, tea, and fruit by ultra-performance liquid chromatography-tandem mass spectrometry (UPLC-MS/MS). To guarantee the accuracy and sensitivity of detection of different substrates, this study explored the improvement of QuEChERS method, optimization of detection conditions, and improvement of the pretreatment conditions for different matrices. A UPLC-MS/MS method was developed for the simultaneous determination of 12 IGRs in different

substrates by using modified QuEChERS and UPLC-MS/MS techniques. This method might provide technical support for food safety and risk assessment.

2. Material and methods

2.1. Chemicals and reagents

Standards of flonicamid (99% purity; CAS Registry no. 158062-67-0), RH5849 ((1,2-dibenzoyl-1-*tert*-butylhydrazine), 99.0% purity; CAS Registry no. 112225-87-3), chlorantraniliprole (97.8% purity; CAS Registry no. 500008-45-7), methoxyfenozide (99.0% purity; CAS Registry no. 161050-58-4), chromafenozide (99.9% purity; CAS Registry no. 143807-66-3), fenoxycarb (99.5% purity; CAS Registry no. 72490-01-8), tebufenozide (99.6% purity; CAS Registry no. 112451-23-8), flubendiamide (99.2% purity; CAS Registry no. 272451-65-7), buprofezin (99.9% purity; CAS Registry no. 69327-76-0), tolfenpyrad (99.3% purity; CAS Registry no. 129558-76-5), pyriproxyfen (99.9% purity; CAS Registry no. 95737-68-1) were obtained from Dr Ehrenstorfer GmbH (Augsburg, Germany); cyantraniliprole was purchased from ANPEL Laboratory Technologies (Shanghai) Inc. HPLC-grade acetonitrile (ACN) was obtained from Merck (Darmstadt, Germany), methanol, acetone and ethyl acetate were purchased from Thermo Fisher Co., Ltd. (USA). Hexane, formic acid and ammonium acetate were purchased from Dikma (USA). Superior pure petroleum ether was purchased from Tianjin Kemiou Chemical Reagent Co., Ltd (China). Sodium chloride (NaCl) and magnesium sulfate anhydrous were purchased from Tianjin Yongda Chemical Reagent Co., Ltd (Tianjin, China) and were of analytical grade. Primary secondary amine (PSA) and graphitized carbon black (GCB) were supplied by Dikma (USA). C₁₈ was purchased from Welch (40–50 μm , USA). Ultra-pure water was prepared using a Millipore Milli-Q Gradient Water Purification System (USA).

2.2. Standard solution preparation

Individual standard solutions of 12 IGRs were prepared in ACN at a concentration of 100 $\mu\text{g mL}^{-1}$. A working solution (1 $\mu\text{g mL}^{-1}$) was prepared by mixing a certain amount of individual standard solution and methanol. The solvent calibration standard solutions were prepared by mixing the working standard solutions with additional methanol to reach the different concentration. Matrix-matched calibration standard solutions were prepared in blank sample containing different analytes contents. All standards solutions were stored at $-20\text{ }^{\circ}\text{C}$ in a bottle and freshly prepared solvent or matrix standards were used before each analysis.

2.3. UPLC-MS/MS analysis conditions

A UPLC-MS/MS instrument (Exion-TRIPLE QUAD 5500, AB SCIEX, USA) was used to analyze the 12 target compounds. The LC unit was used to separate the target compounds using an Agilent ZORBAX Eclipse Plus-C₁₈ column (2.1 mm \times 100 mm, 1.8 μm) held at 40 $^{\circ}\text{C}$. The injection volume and flow rate were 3 μL and 0.3 mL min^{-1} , respectively. The mobile phase was composed of 0.1% formic acid solution (A) and methanol (B),



and a linear-gradient elution program was set as follows: 45% B (0–0.1 min), 45–71% B (0.1–0.2 min), 71% B (0.2–2.8 min), 71–63% B (2.8–2.9 min), 63% B (2.9–3.8 min), 63–82% B (3.8–3.9 min), 82% (3.9–6.9 min), 82–45% (6.9–7.0 min), and 45% B (7.0–10.0 min). Ionization was carried out using an electrospray ionization (ESI) source in the positive mode, and mass spectrometric analysis was carried out in MRM mode. The parameters for MS/MS were set as follows: spray voltage, 5500 V; vaporizer temperature, 550 °C; nebulizing gas pressure, 55 psi; auxiliary gas pressure, 55 psi; and curtain gas pressure, 40 psi. Table 1 showed the optimized MRM data acquisitions.

2.4. Sample preparation

Vegetable oil, fruit, and tea samples were purchased from local supermarkets of Shijiazhuang. For fruit, the edible part of sample was uniformly crushed using a proofer and placed in a closed container, then it was stored in a refrigerator at 4 °C away from light. The tea sample was ground in a mortar and placed in a clean self-sealing bag until use. Blank samples were used for validation studies and matrix-matched standard calibrations. Samples for recovery studies were spiked with a known amount of fortification standard and left for 10 min before the extraction.

The general procedures for extraction and purification were presented separately as follows.

Extraction: 2.00 g samples were accurately weighed in a 50 mL centrifuge tube, 2 mL water was added, vortexed for 30 s. Then, 10 mL acetonitrile containing 0.5% formic acid was added. After vortex mixing and ultrasonication for 10 min,

800 mg NaCl was added and mixed by vortexing for 30 s. The mixture was centrifuged at 4000 rpm for 9 min and subjected to purification.

Purification: 8 mL of the supernatant was transferred to a 15 mL centrifuge tube containing different sorbents. Then, the mixture was vortexed for 30 s, left standing for 2 min, centrifuged for 5 min at 8000 rpm, and 5 mL of the supernatant was evaporated to dryness under nitrogen at 40 °C. The residue was dissolved in 1 mL methanol, vortexed for 30 s, and passed through a 0.20 μm nylon filter membrane before LC-MS/MS analysis.

3. Results and discussion

3.1. Optimization of MS/MS conditions

A tandem mass detector has high selectivity and sensitivity and provides effective detection. First, a needle pump was used for continuous sampling. A parent ion scan (Q1 MS) was performed to obtain the parent ion of target, and then the fragment ion was obtained through product ion scan. A detection ion pair was formed by the parent ion and two fragment ions with strong response signals. In addition to the $[M + H]^+$ of flubendiamide with weak response signal, other 11 IGRs showed the $[M + H]^+$ ion as the precursor ion. The intensities of addition peaks $[M + Na]^+$ and $[M + NH_4]^+$ of flubendiamide were relatively good. However, because the stability of addition peaks of $[M + Na]^+$ was significantly affected by the matrix, the addition ions $[M + NH_4]^+$ were selected as the parent ions. The parent ions, product

Table 1 Mass spectrometric parameters of 12 kinds of IGRs

Compound	Retention time (min)	Precursor ion (m/z)	Declustering potential (V)	Product ion (m/z)	Collision energy (eV)
Flonicamid	1.23	230.2	130.2	203.1	23.8
			81.9	176.2	32.1
Cyantraniliprole	2.73	475.1	71.0	286.0 ^a	18.9
			71.9	443.9	22.6
RH-5849	2.87	297.2	73.3	105.1	24.8
			62.1	241.2	11.0
Chlorantraniliprole	3.25	484.2	75.1	453.0 ^a	26.1
			69.0	286.1	19.2
Methoxyfenozide	3.96	369.4	54.1	149.2 ^a	23.6
			67.1	313.2	11.7
Chromafenozide	4.41	395.2	87.9	175.2 ^a	20.9
			81.8	339.3	10.7
Fenoxycarb	5.57	302.1	80.7	116.0 ^a	15.3
			95.1	256.1	17.7
Tebufenozide	5.79	353.3	66.0	133.2 ^a	23.0
			63.6	297.4	11.7
Flubendiamide	6.07	700.2	63.0	408.0 ^a	18.9
			65.9	273.7	42.1
Buprofezin	7.38	306.2	60.8	201.0 ^a	16.8
			60.1	116.2	22.4
Tolfenpyrad	7.82	384.3	77.2	197.2 ^a	34.1
			77.2	170.8	28.7
Pyriproxyfen	8.12	322.2	90.4	227.0 ^a	20.4
			89.3	185.1	28.9

^a Quantifying ions.



ions, declustering potential, and collision energy of IGRs using MRM were shown in Table 1.

3.2. Optimization of UPLC conditions

Despite the high specificity of MS/MS detection, chromatographic separation (resolution) is decisive for improving the sensitivity and peak shape of analytes.⁴ The chromatographic conditions of compounds were affected by the packing, particle size, and mobile phase. To obtain the best chromatographic conditions for the target compounds, the separation effects of chromatographic columns with three different specifications were compared. It was found that 12 types of target compounds could not be well separated using Zorbax Eclipse XDB-C₁₈ (2.1 × 50 mm, 1.8 μm) and Zorbax Eclipse XDB-C₁₈ (4.6 × 50 mm, 1.8 μm) column. The separation effect was better on Zorbax Eclipse Plus C₁₈ (4.6 × 100 mm, 1.8 μm) column, and the response of each substance was higher.

Regarding the optimization of chromatographic conditions, mobile phase was another important factor affecting chromatographic peak-to-peak type and their response. In the experiment, two mobile phases A and B were optimized. Methanol and acetonitrile were selected and compared as the organic phase B. Flonicamid showed poor peak shape and split peak in acetonitrile as the mobile phase. While methanol was used as the mobile phase, flonicamid exhibited good peak shape and 4 times higher response than that in acetonitrile; meanwhile, chlorantraniliprole showed about 2 times higher response. In addition, the other nine types of compounds showed better response in methanol except buprofezin. Therefore, methanol was selected as the organic phase of mobile

phase B. Because the mass spectrum peak of flubendiamide showed an additional ammonia peak, two aqueous phases were compared: 2 mmol L⁻¹ ammonium acetate, 0.1% formic acid aqueous solution containing 2 mmol L⁻¹ ammonium acetate. The [M + NH₄]⁺ signal intensity of flubendiamide did not increase significantly after adding ammonium acetate to aqueous phase, and the response signals of other some compounds were suppressed. When the aqueous phase A contained 0.1% formic acid, the separation effect of 12 IGRs was better, the response signal had a higher intensity, and the peak shape was sharp. Therefore, 0.1% formic acid aqueous solution and methanol was finally selected as the mobile phase, and separation was carried out by gradient elution procedure. Within 9 min, 12 compounds were well separated (Fig. 1).

3.3. Optimization of modified QuEChERS for different food matrix samples

3.3.1. Optimization of extraction. The extraction efficiency was expressed by the spiked recovery, which was calculated by matrix-matched standard curves.

In this experiment, soybean oil was used as a vegetable oil substrate. Various extraction conditions such as the extractant, extractant volume, ultrasonic extraction time were examined, 3 replicates were done for every test. Because of the complexity of sample matrix, the extractant should not only solubilize the target compound well, but also avoid the extraction of impurities. Currently, acetonitrile, acetone and ethyl acetate are the most used solvents for extraction in QuEChERS. Moreover, acidified MeCN showed good extraction efficiency for many analytes.³³ Therefore, acetonitrile, acetone, acetonitrile with

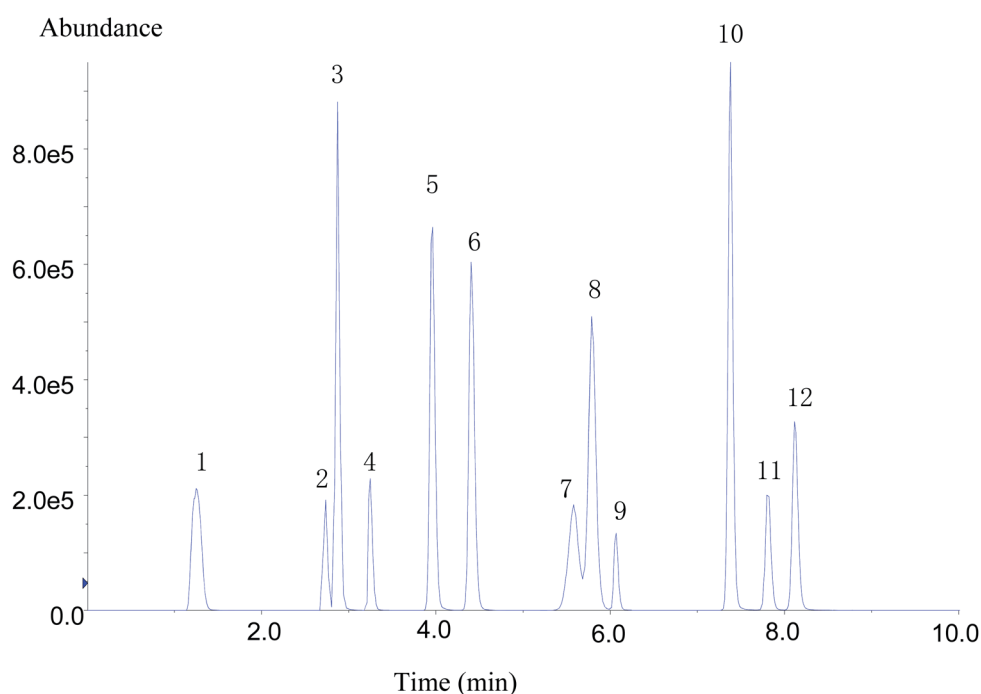


Fig. 1 Chromatograms of 12 kinds of IGRs (50 μg L⁻¹). 1. Flonicamid, 2. cyantraniliprole, 3. RH-5849, 4. chlorantraniliprole, 5. methoxyfenozide, 6. chromafenozide, 7. fenoxycarb, 8. tebufenozide, 9. flubendiamide, 10. buprofezin, 11. tolfenpyrad, 12. pyriproxyfen.



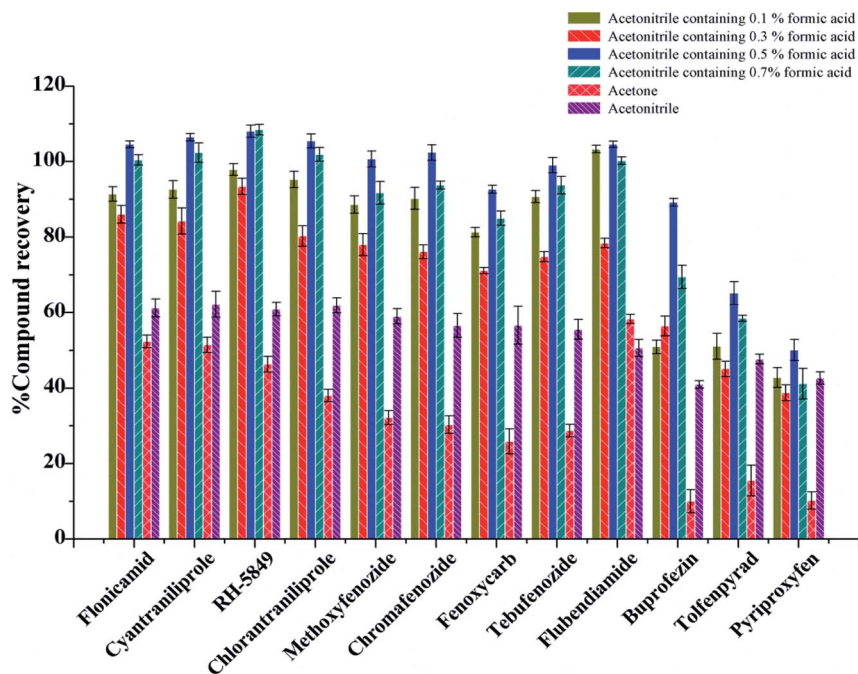


Fig. 2 Recoveries of 12 kinds of IGRs with different extractant.

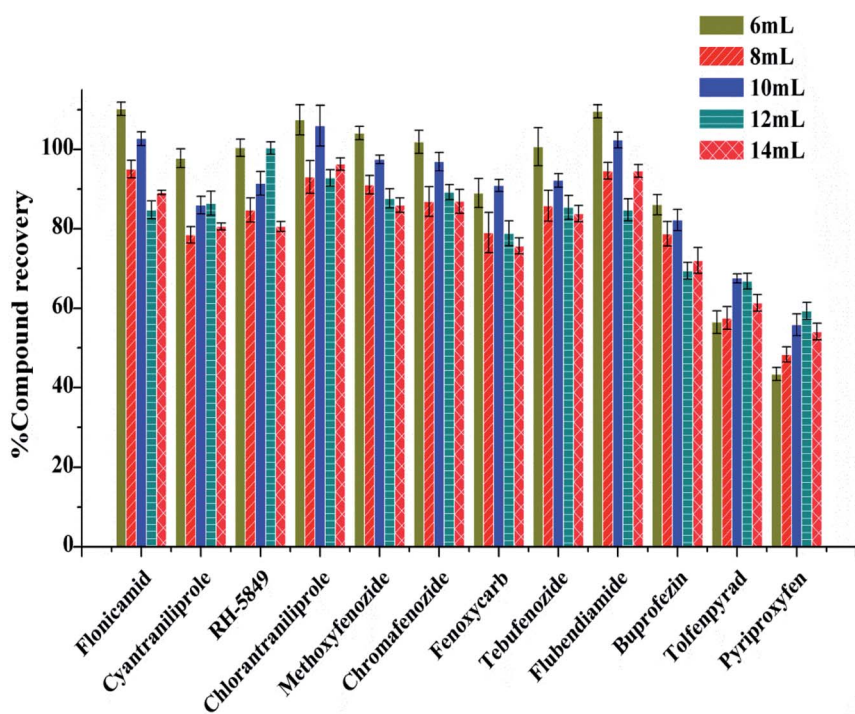


Fig. 3 Recoveries of 12 kinds of IGRs with different extractant volume.

different concentrations of formic acid were investigated in the experiment.

Firstly, 2.00 g blank vegetable oil was spiked at the level of $10 \mu\text{g kg}^{-1}$, the extraction and purification procedure was done as shown in section 2.4 with different extractant, and the 0.1 g PSA + 0.3 g MgSO_4 was used as purifying sorbent. When acetone was

used as an extractant, more impurities were obtained after drying under nitrogen, and the recoveries of most of the substances were below 50% (Fig. 2). The recoveries of 12 IGRs were between 40.99% and 62.21% when pure acetonitrile was used as an extraction reagent. The recoveries of 12 IGRs increased with the amount of formic acid added, and the



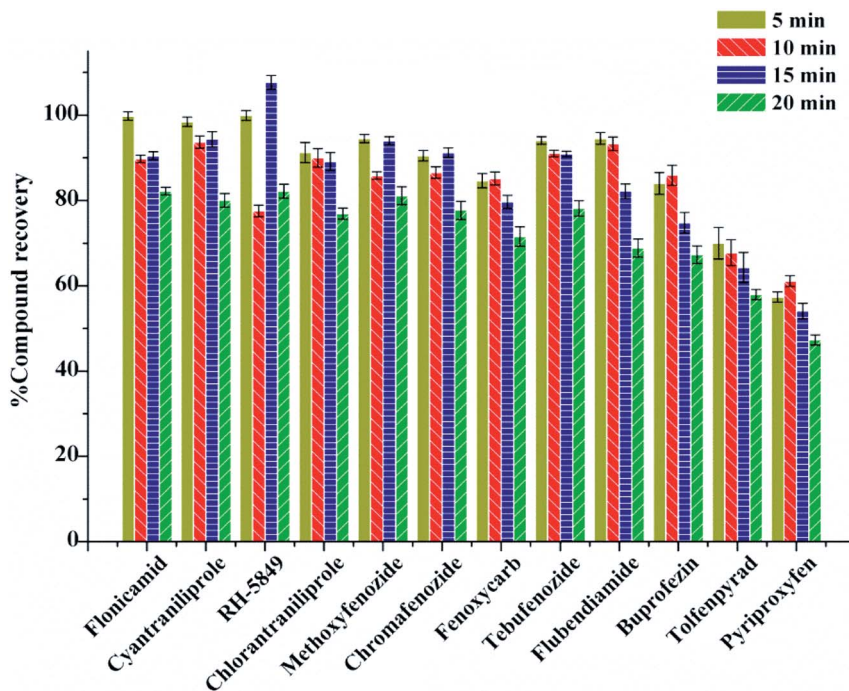


Fig. 4 Recoveries of 12 kinds of IGRs under different ultrasonic extraction time.

extraction effect was the best when the amount of formic acid added was 0.5%. The recoveries of 12 IGRs were 50.11–108.07%; most of them were over 80% (Fig. 2).

Furthermore, different volume of acetonitrile containing 0.5% formic acid were examined. The recoveries of 12 IGRs were 55.83–105.98%, when the extractant volume was 10 mL, which was better than 6 mL, 8 mL, 12 mL and 14 mL (Fig. 3).

The extraction was carried out with the help of ultrasonication in this experiment. The effect of ultrasonication time on the recovery was evaluated at 5 min, 10 min, 15 min, and 20 min. The results show that with the increase of ultrasonication time, the average recoveries increased and reached the best at 10 min with the recoveries of 61.10–93.71% (Fig. 4). Therefore, 10 min was the best ultrasonication time for extraction.

Moreover, for most QuEChERS method, the combination of NaCl and MgSO₄ was used in extraction to make salting-out effect,³³ causing the partitioning of water and acetonitrile. However, no substantial differences were observed for recoveries of 12 IGRs in vegetable oil, when only NaCl or the combination of NaCl and MgSO₄ were used. As a result, only NaCl was used for partitioning of water and acetonitrile.

For vegetable oil, fruit, and tea samples, the sample characteristic showed obvious difference, the optimal extraction procedure for vegetable oil was also investigated and adjusted for fruit and tea. The experiment showed that addition of water in the extraction procedure made more impurities for fruit and tea, which reduced extraction efficiency. Meanwhile, when 10 mL extractant was examined, 2 g tea make it hard to remove 8 mL supernatant, then 1 g tea was chosen for the extraction procedure. For fruit and tea, no addition of water in the

extraction procedure, then less amount of NaCl (500 mg) was used for fruit, and NaCl was not added for tea.

3.3.2. Optimization of purification procedure. Primary secondary amine (PSA), octadecyl silica (C₁₈), graphitized carbon black (GCB) and MgSO₄ were often used as purification sorbents, PSA is used for the removal of fatty acids, sugars, organic acids, lipids and some pigments, and C₁₈ can make an effective removal of lipid. Moreover, MgSO₄ was used to remove water, and GCB was used to remove the co-extracted pigments, which can also remove pesticides with planar structures.³⁴

In the purification experiment, the sorbents and their amount were discussed with vegetable oil, fruit and tea, respectively. The blank sample was spiked at the level of 10 μg kg⁻¹, after extraction with the optimal procedure, the purifying procedure was explored according to Section 2.4 with different sorbents, and the recoveries was calculated by matrix-matched standard curves, the average recoveries based on 5 replicates were used to evaluate the purifying efficiency.

For the fruit sample, the efficiencies of six sets of sorbents (0.1 g PSA + 0.3 g MgSO₄; 0.1 g C₁₈ + 0.3 g MgSO₄; 0.1 g PSA + 0.1 g C₁₈ + 0.3 g MgSO₄; 0.1 g GCB + 0.1 g C₁₈ + 0.1 g PSA + 0.3 g MgSO₄; 0.1 g GCB + 0.1 g C₁₈ + 0.3 g MgSO₄; 0.1 g GCB + 0.1 g PSA + 0.3 g MgSO₄) were compared by purifying the sample extracts spiked with 10 μg kg⁻¹ of the target 12 IGRs. The results are shown in Table S1. † Graphitized carbon black (GCB) showed a strong adsorption capacity to cyanoformamide and chlorantraniliprole. The recoveries of cyanoformamide and chlorantraniliprole were 16.58–27.49% using the purified sorbents containing GCB; at the same time, GCB also adsorbed tolfenpyrad and pyriproxyfen, which can be attributed to the planar structures of analytes.³⁴ The other four sorbents without



Table 2 Matrix effects of vegetable oil, fruit and tea

Compound	Spiked level ($\mu\text{g kg}^{-1}$)			Vegetable oil (%)			Fruit (%)			Tea (%)		
	10	50	100									
Flonicamid	10	50	100	86.45	80.16	83.46	37.52	37.46	43.42	10.19	10.05	9.98
Cyantranilprole	10	50	100	89.96	87.88	96.88	52.81	54.72	63.24	81.17	85.70	87.76
RH-5849	10	50	100	90.34	86.74	89.20	80.58	88.93	90.47	80.89	85.76	87.88
Chlorantranilprole	10	50	100	98.79	94.68	98.18	78.88	79.96	87.36	86.89	91.42	92.82
Methoxyfenozide	10	50	100	91.11	87.09	89.24	102.44	100.01	111.89	91.03	94.45	97.20
Chromafenozide	10	50	100	93.62	89.76	84.95	103.26	100.61	118.31	93.01	94.64	95.79
Fenoxycarb	10	50	100	93.12	91.81	94.57	87.36	90.40	103.32	90.62	93.09	94.32
Tebufenozide	10	50	100	91.67	90.79	89.86	105.17	101.24	112.00	90.95	96.80	96.50
Flubendiamide	10	50	100	112.97	119.19	115.32	121.01	113.26	122.91	87.69	102.93	94.16
Buprofezin	10	50	100	90.16	86.35	83.98	96.37	100.35	116.94	96.43	97.07	98.39
Tolfenpyrad	10	50	100	109.74	96.02	99.10	91.54	96.18	111.34	97.79	95.09	94.99
Pyriproxyfen	10	50	100	90.23	88.07	91.03	91.67	96.88	113.90	90.39	90.28	92.78

Table 3 Linear range, linear equations, LODs and LOQs of 12 IGRs

Compound	Linear range ($\mu\text{g L}^{-1}$)	Calibration curve	R^2	LOD ($\mu\text{g kg}^{-1}$)	LOQ ($\mu\text{g kg}^{-1}$)
Flonicamid	0.2–200	$y = 157\,355.59x + 826.70$	0.9999	0.11	0.36
Cyantranilprole	0.2–200	$y = 7307.53x + 112.64$	0.9998	0.37	1.12
RH-5849	0.2–200	$y = 61\,441.90x + 22\,201.13$	0.9997	0.10	0.30
Chlorantranilprole	0.2–200	$y = 3566.99x + 774.37$	0.9999	0.04	0.13
Methoxyfenozide	0.2–200	$y = 63\,663.93x + 6348.65$	0.9999	0.05	0.17
Chromafenozide	0.2–200	$y = 78\,091.30x + 10\,905.83$	0.9999	0.08	0.26
Fenoxycarb	0.2–200	$y = 51\,842.30x + 2040.27$	0.9994	0.40	1.24
Tebufenozide	0.2–200	$y = 63\,738.42x + 1748.25$	0.9999	0.23	0.74
Flubendiamide	0.2–200	$y = 3869.97x + 108.77$	0.9999	0.21	0.71
Buprofezin	0.2–200	$y = 140\,217.13x + 6772.69$	0.9999	0.04	0.15
Tolfenpyrad	0.2–200	$y = 30\,751.13x + 948.79$	0.9999	0.28	0.94
Pyriproxyfen	0.2–200	$y = 26\,817.48x + 2904.63$	0.9999	0.09	0.31

GCB showed better purification effect on 12 IGRs, and the recovery of each substance was above 79.9%. According to the results shown in Table S1,† the sorbent C_{18} made better recovery for 12 IGRs than PSA or PSA + C_{18} . Then, a mixture of 0.1 g C_{18} and 0.3 g $MgSO_4$ was selected as the sorbent, and the corresponding recoveries of 12 IGRs were 90.12–107.08%.

Based on the above results, the purification efficiency of PSA, C_{18} , $MgSO_4$, and their different combinations was separately evaluated for vegetable oil and tea. As shown in Table S2,† a mixture of 0.1 g PSA and 0.3 g $MgSO_4$ was the most effective sorbent for the purification of vegetable oil with the recoveries of 60.34–105.85%. For tea, the mixture of 0.1 g PSA, 0.2 g C_{18} and 0.1 g $MgSO_4$ was the optimal purification sorbent, which can obtain the best recoveries of 12 IGRs with 62.97–110.81% (Table S3†).

3.4. Method validation

3.4.1. Matrix effect. Matrix effects (ME) are one of the influencing factors of coextracted pesticides on analytical signals, caused by the type of matrix and efficiency of sample preparation step. ME can be calculated using the following equation: $ME (\%) = A_x/A_s \times 100\%$, where A_s is the peak area of analyte in a pure standard solution, and A_x is the peak area in spiked matrix extract.³⁵ When ME is in the range of 80–120%,

the matrix effect is small and can be ignored. When ME is lower than 80% or higher than 120%, there are different degrees of inhibition or enhancement,³⁶ which should be corrected with matrix-based standard solutions. In the experiment, the samples without target substance were extracted according to “2.4” method to obtain a blank matrix solution. The matrix effect of 12 IGRs in vegetable oil, fruit, and tea was evaluated at concentrations of $10 \mu\text{g L}^{-1}$, $50 \mu\text{g L}^{-1}$, and $100 \mu\text{g L}^{-1}$ (Table 2). The results show that the matrix effect of 12 IGRs in the vegetable oil matrix is 80.16–119.19%, which is small, almost negligible. The ME of flonicamid in fruit and tea matrix were both lower than 50%, which meant that the matrix inhibition effect was severe. Moreover, the fruit matrix also affected cyantranilprole, the ME is 52.81–63.24%. Therefore, for an accurate quantification, matrix-matched standard calibration curves were constructed to quantify the target compounds in fruit and tea samples, which can eliminate the effect of ME on the results.¹⁰

3.4.2. Linearity, limit of detection (LOD), and limit of quantitation (LOQ). The calibration curve was constructed by running solvent standard solutions of different concentrations in methanol (the solvent calibration standards ranged from 0.2 to $200 \mu\text{g L}^{-1}$) (0.2, 0.5, 1, 2, 5, 10, 20, 50, 100, and $200 \mu\text{g L}^{-1}$) shown in Table 3. Correlation coefficient was determined using



Table 4 The recoveries and RSDs of 12 kinds of IGRs in vegetable oil ($n = 7$)

Compound	Spiked level ($\mu\text{g kg}^{-1}$)			Recovery (%)			RSD (%)		
	10	50	100						
Fonicamid	10	50	100	83.20	67.82	65.47	3.16	8.63	3.49
Cyantranilprole	10	50	100	88.02	95.19	85.03	3.27	3.63	3.34
RH-5849	10	50	100	78.45	77.33	80.27	11.09	8.10	6.95
Chlorantranilprole	10	50	100	77.08	78.11	79.87	4.38	3.62	3.45
Methoxyfenozide	10	50	100	80.22	83.25	81.58	5.41	3.28	3.79
Chromafenozide	10	50	100	78.39	80.28	82.53	4.26	5.51	4.06
Fenoxycarb	10	50	100	76.56	78.27	80.51	3.54	5.69	1.93
Tebufenozide	10	50	100	81.26	80.92	83.28	6.24	4.63	4.01
Flubendiamide	10	50	100	78.47	78.01	76.03	11.06	9.40	4.35
Buprofezin	10	50	100	71.94	70.68	74.12	2.60	5.68	1.92
Tolfenpyrad	10	50	100	63.89	66.09	65.15	2.44	6.21	4.29
Pyriproxyfen	10	50	100	67.35	62.75	65.06	3.07	3.21	2.97

Table 5 The recoveries and RSDs of 12 kinds of IGRs in fruit ($n = 7$)

Compound	Spiked level ($\mu\text{g kg}^{-1}$)			Recovery (%)			RSD (%)		
	10	50	100						
Fonicamid	10	50	100	88.70	102.81	96.75	3.65	7.66	3.18
Cyantranilprole	10	50	100	88.02	103.19	85.03	2.95	3.88	3.84
RH-5849	10	50	100	85.82	102.36	88.12	9.68	7.31	8.57
Chlorantranilprole	10	50	100	99.70	105.53	92.02	4.75	3.40	5.07
Methoxyfenozide	10	50	100	93.76	101.49	89.32	6.05	3.41	4.14
Chromafenozide	10	50	100	80.55	99.53	95.78	3.56	5.93	4.16
Fenoxycarb	10	50	100	86.03	94.03	84.45	3.76	5.53	2.79
Tebufenozide	10	50	100	90.88	100.74	102.78	6.54	4.47	5.40
Flubendiamide	10	50	100	89.85	110.15	90.95	11.07	8.91	4.37
Buprofezin	10	50	100	89.65	103.08	95.88	3.22	6.61	3.02
Tolfenpyrad	10	50	100	84.67	96.14	86.57	2.52	5.64	4.40
Pyriproxyfen	10	50	100	84.57	95.66	83.33	3.21	4.30	3.14

a linear regression model. The LOD and LOQ were calculated by analyzing 11 blank sample extracts. LOD was defined as three times the signal-to-noise ratio, and LOQ was defined as ten times the signal-to-noise ratio. The results were shown in Table 3. The LODs and LOQs of 12 IGRs for vegetable oil were 0.04–0.40 $\mu\text{g kg}^{-1}$ and 0.13–1.24 $\mu\text{g kg}^{-1}$, respectively, the satisfactory linearity were obtained with coefficient of determination (R^2) ≥ 0.9994 . For fruit and tea, the matrix-matching standard solutions were used to fit the standard curve, the linearities, LODs and LOQs were shown in Tables S4 and S5.†

3.4.3. Recovery and precision of standard addition. The recovery was used to evaluate accuracy, blank vegetable oil, fruit, and tea samples were spiked with 12 IGRs at three different levels (10.0 $\mu\text{g kg}^{-1}$, 50.0 $\mu\text{g kg}^{-1}$, and 100.0 $\mu\text{g kg}^{-1}$), and seven parallel samples were prepared at each concentration. The solvent calibration curve was used for quantification in vegetable oil, the matrix calibration curve was used for quantification in fruit and tea, based on which the recoveries and precisions were calculated. The recovery results of 12 IGRs in vegetable oil, fruit, and tea were shown in Tables 4, 5 and 6, respectively.

Table 6 The recoveries and RSDs of 12 kinds of IGRs in tea ($n = 7$)

Compound	Spiked level ($\mu\text{g kg}^{-1}$)			Recovery (%)			RSD (%)		
	10	50	100						
Fonicamid	10	50	100	68.29	62.02	65.13	9.35	4.46	3.18
Cyantranilprole	10	50	100	71.14	69.07	67.86	6.95	2.44	4.19
RH-5849	10	50	100	71.52	65.49	68.45	6.12	3.38	4.31
Chlorantranilprole	10	50	100	78.62	76.45	74.82	11.58	2.05	1.91
Methoxyfenozide	10	50	100	80.54	81.66	91.05	8.93	3.59	4.19
Chromafenozide	10	50	100	64.95	84.86	84.28	7.59	2.48	3.57
Fenoxycarb	10	50	100	95.94	81.24	77.98	10.21	1.30	2.49
Tebufenozide	10	50	100	92.60	85.47	96.50	9.09	2.47	3.23
Flubendiamide	10	50	100	68.69	83.36	91.16	3.49	3.31	6.03
Buprofezin	10	50	100	89.50	81.97	85.64	10.03	3.29	3.06
Tolfenpyrad	10	50	100	97.99	104.86	98.79	8.47	11.29	4.12
Pyriproxyfen	10	50	100	86.75	79.98	81.67	4.50	3.55	1.17



Table 7 Performance comparisons for IGRs determination with other reported analytical methods

Analytical method	Pretreatment method	Analytes	Sample	Recovery (%)	LOQ ($\mu\text{g kg}^{-1}$)	Ref.
LC-MS/MS	DPX (disposable pipette extraction)	8 IGRs	Honey	75–109	1.0–10	4
LC-MS/MS	SPE	Tebufenozide	Vegetable	70–110	4.0	9
LC-MS/MS	SPE	Pyriproxyfen, tolfenpyrad	Citrus	80.6–113	5.0	25
LC-MS/MS	LLE, QuEChERS	6 IGRs	Water, sediment, aquatic products	80–99.7	1.27–3.20	21
LC-MS/MS	QuEChERS	Fonicamid	Paprika	94–101.7	10	37
LC-MS/MS	QuEChERS	Fonicamid	Vegetable	86.8–94.8	4.0	38
LC-MS/MS	QuEChERS	Chlorantraniliprole, buprofezin	Fruit, vegetable	64–115	0.03–0.51	39
LC-MS/MS	QuEChERS	12 IGRs	Vegetable, fruit, tea	62.02–110.15	0.11–3.91	This work

The average recoveries at three spiked levels in three different substrates were in the range of 62.75–95.19% for vegetable oil, 80.55–110.15% for fruit, and 62.02–104.86% for tea, respectively, with relative standard deviations (RSDs, $n = 7$) of 1.17–11.58%. Owing to the advantages of simple operation and high accuracy and sensitivity, this method was suitable for rapid and simultaneous detection of 12 IGRs in vegetable oil, fruit, and tea.

3.5. Application to actual sample analysis

The developed method in this study was applied to the identification and quantification of 12 IGRs in 17 vegetable oils, 16 fruits, and 24 tea samples separately. The samples were obtained from local markets and supermarkets in Shijiazhuang and analyzed following the preparation procedure. Fonicamid ($2.34 \mu\text{g kg}^{-1}$) and RH-5849 ($0.20 \mu\text{g kg}^{-1}$) were observed in one fruit sample, cyantraniliprole ($3.28 \mu\text{g kg}^{-1}$) was observed in one fruit sample, chlorantraniliprole (0.14 – $0.19 \mu\text{g kg}^{-1}$) was observed in four fruit samples, buprofezin (0.32 – $1.26 \mu\text{g kg}^{-1}$) was observed in four fruit samples, tolfenpyrad ($1.63 \mu\text{g kg}^{-1}$) was observed in one fruit sample (Table S6†). The contents of chlorantraniliprole, buprofezin, and tolfenpyrad, in 24 tea samples are 0.74 – $84.72 \mu\text{g kg}^{-1}$, 0.18 – $38.60 \mu\text{g kg}^{-1}$ and 0.34 – $1125.15 \mu\text{g kg}^{-1}$ (Table S7†). The contents of 12 IGRs in edible oil samples were all below the LODs. According to the Chinese GB2763-2019 “National food safety standard—maximum residue limits for pesticides in food”,¹⁸ the above IGRs residue contents in the tested samples did not exceed the standard.

3.6. Method performance comparison

Analytical performances of the developed QuEChERS-UPLC-MS/MS method for the determining IGRs were mainly compared with the reported LC-MS/MS methods. As shown in Table 7, SPE and QuEChERS were the most used pretreatment methods for the detection of IGRs. However, compared with SPE, QuEChERS was more convenient and fast. In these reported methods, most of them were developed to test one or two IGRs,^{9,25,37–39} and/or the testing sample matrix was single. Only a method for 8 IGRs in honey was established,⁴ and another method was used

to detect 6 IGRs in water, sediment and aquatic products.²¹ In comparison, the new QuEChERS-UPLC-MS/MS method was developed for the determination of 12 IGRs in three different sample matrixes with suitable recoveries and the low LOQs, which showed high throughput and high sensitivity.

4. Conclusion

A modified QuEChERS method coupled with UPLC-MS/MS analysis was successfully developed to determine 12 IGRs in foods of different matrixes. The samples were extracted with acetonitrile containing 0.5% formic acid, purified with different sorbents, detected by UPLC-MS/MS with ESI in positive ion mode under MRM mode, and quantified using the external standard method. The extraction and purification methods were optimized for different food matrix samples, and then the method was validated regarding its analytical range, precision, recovery, and ME. Based on the experimental results, the method is simple, fast, sensitive, which can be applied in accurate and high throughout detection of IGRs, providing technical support for relevant food risk assessment. However, fonicamid and cyantraniliprole still show obvious matrix effect in fruit and tea, the further improvement for pretreatment method would be explored to eliminate the matrix effect.

Author contributions

Ke Wang: data curation, formal analysis, methodology, writing – original draft, funding acquisition. Lingzhi Zhao: data curation. Can Zhang: formal analysis, methodology, writing – original draft. Hong Zhang: methodology, validation. Kaoqi Lian: conceptualization, funding acquisition, project administration, resources, writing – review & editing. All authors have read and agreed to the published version of the manuscript.

Conflicts of interest

The authors declare no conflict of interest.



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Notes and references

- 1 E. A. Gamal, E. E. Anwar, M. E. Ferial, M. E. Hamdi and M. E. Hesham, *Pest Manage. Sci.*, 2003, **69**, 95–102.
- 2 G. C. Zhang, H. Zou, N. N. Geng, N. Ding, Y. J. Wang, J. Zhang and C. Zou, *Pestic. Biochem. Physiol.*, 2020, **163**, 64–75.
- 3 J. D. Fine, C. A. Mullin, M. T. Frazier and R. D. Reynolds, *J. Econ. Entomol.*, 2017, **110**, 1993–2001.
- 4 S. Song, C. Zhang, Z. Chen, F. He, J. Wei, H. Tan and X. Li, *J. Chromatogr. A*, 2018, **1557**, 51–61.
- 5 M. R. Qian, L. Q. Wu, H. Zhang, M. F. Xu, R. Li, X. Y. Wang and C. X. Sun, *Anal. Bioanal. Chem.*, 2012, **402**, 2451–2462.
- 6 D. Šatínský, L. Naibrťová, C. Fernández-Ramos and P. Solich, *Talanta*, 2015, **142**, 124–130.
- 7 X. L. Hu, J. J. Niu, Q. Meng, Y. H. Chai, K. H. Chu and K. M. Chan, *Environ. Pollut.*, 2019, **253**, 89–99.
- 8 J. L. Jiang, Z. J. Shan, X. R. Wang, Y. X. Zhu and J. Y. Zhou, *Environ. Sci. Pollut. Res.*, 2018, **25**, 10730–12739.
- 9 L. Dai, J. Z. Xu, T. Ding, J. J. Zhu, B. Wu, C. Y. Shen and Y. Jiang, *Chin. J. Anal. Chem.*, 2008, **36**, 87–90.
- 10 T. Liu, M. Dong, F. J. Zhou, D. Yang and X. M. Zhang, *Food Sci. Hum. Well.*, 2019, **8**, 362–367.
- 11 A. Kadala, M. Charretona and C. Colleta, *J. Insect Physiol.*, 2020, **125**, 104086.
- 12 S. L. Feng, Z. M. Kong, W. X. Wang, X. M. Wang and P. A. Peng, *Teratog., Carcinog., Mutagen.*, 2002, **14**, 210–213.
- 13 M. Liu, H. Yang, H. Liu, P. Han, X. Wang, S. Zhang and Y. Wu, *J. Sci. Food Agric.*, 2008, **88**, 62–67.
- 14 S. Navis, A. Waterkeyn, L. De Meester and L. Brendonck, *Ecotoxicology*, 2018, **27**, 627–634.
- 15 G.-F. Pang, Chapter 1: Introduction, *Analytical Methods for Food Safety by Mass Spectrometry*, Academic Press, 2018, pp. 1–9.
- 16 Regulation (EC) No. 396/2005 of the European Parliament and of the Council of 23 February 2005 on maximum residue levels of pesticides in or on food and feed of plant and animal origin and amending Council Directive 91/414/EEC (text with EEA relevance).
- 17 Ministry of Health, Labour and Welfare, *The Japanese positive list system for agricultural chemical residues in foods*, Ministry of Health, Labour and Welfare, Tokyo, 2006.
- 18 *National Food Safety Standard, Maximum Residue Limits for Pesticides in Food: GB 2763-2019*, Standards Press of China, Beijing, 2019.
- 19 F. M. Malhat, *Food Anal. Methods*, 2012, **5**, 1492–1496.
- 20 S. Mohapatra, A. K. Ahuja, M. Deepa, G. K. Jagadish, N. Rashmi and D. Sharma, *J. Environ. Sci. Health, Part B*, 2011, **46**, 264–271.
- 21 J. Gan, H. Liu, Y. Chen, J. Peng, T. Liu, J. Chen and L. He, *Ecotoxicol. Environ. Saf.*, 2021, **210**, 111853–111861.
- 22 M. Saitta, G. Di Bella, M. R. Fede, V. Lo Turco, A. G. Potorti, R. Rando, M. T. Russo and G. Dugo, *Food Addit. Contam., Part A*, 2017, **34**, 800–808.
- 23 Y. L. Cui, K. C. Liu, C. Xu, F. Liu, Q. X. Li, S. Z. Liu and B. M. Wang, *Food Chem.*, 2014, **143**, 293–299.
- 24 G. P. de Pinho, A. A. Neves, M. E. L. R. de Queiroz and F. O. Silvério, *Food Control*, 2010, **21**, 1307–1311.
- 25 M. Dong, G. Wen, H. Tang, T. Wang, Z. Zhao, W. Song, W. Wang and L. Zhao, *Food Chem.*, 2018, **269**, 136–141.
- 26 J. Ma, Z. Yao, L. Hou, W. Lu, Q. Yang, J. Li and L. Chen, *Talanta*, 2016, **161**, 686–692.
- 27 J. Ma, L. Hou, G. Wu, L. Wang, X. Wang and L. Chen, *Materials*, 2020, **13**, 5729–5742.
- 28 M. A. Farajzadeh, M. R. A. Mogaddam and H. Ghorbanpour, *J. Chromatogr. A*, 2014, **1347**, 8–16.
- 29 J. Beltran, F. J. Lopez and F. Hernandez, *J. Chromatogr. A*, 2000, **885**, 389–404.
- 30 Y. Wen, Z. Niu, Y. Ma, J. Ma and L. Chen, *J. Chromatogr. A*, 2014, **1368**, 18–25.
- 31 S. Kammoun, B. Mulhauser, A. Aebi, E. A. D. Mitchell and G. Glauser, *Environ. Pollut.*, 2019, **247**, 964–972.
- 32 T. Xu, X. Feng, L. Pan, J. Jing and H. Zhang, *RSC Adv.*, 2018, **8**, 35485–35495.
- 33 R. Perestrelo, P. Silva, P. Porto-Figueira, J. A. M. Pereira, C. Silva, S. Medina and J. S. Camara, *Anal. Chim. Acta*, 2019, **1070**, 1–28.
- 34 L. C. da Cabrera, S. S. Caldas, O. D. Prestes, E. G. Primel and R. Zanella, *J. Sep. Sci.*, 2016, **39**, 1945–1954.
- 35 X. Li, J. Chen, X. He, Z. Wang, D. Wu, X. Zheng, L. Zheng and B. Wang, *Chemosphere*, 2019, **234**, 224–231.
- 36 P. P. Vazquez, A. Lozano, S. Ucles, M. M. Ramos and A. R. Fernandez-Alba, *J. Chromatogr. A*, 2015, **1426**, 161–173.
- 37 A. Y. Ko, A. M. Abd El-Aty, M. M. Rahman, J. Jang, S. W. Kim, J. H. Choi and J. H. Shim, *Food Chem.*, 2014, **157**, 413–420.
- 38 Y. Xu, L. F. Shou and Y. L. Wu, *J. Chromatogr. A*, 2011, **1218**, 6663–6666.
- 39 H. Li, C. Wang, Q. Zhu, H. Du, S. Guan, F. Wang, W. Zhang, W. Fan, Z. Chen, G. Yang and H. Y. Aboul-Enein, *Anal. Methods*, 2016, **8**, 5061–5069.

