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Appraisal of the greenness profile of a chromatographic method for the simultaneous estimation of carbamazepine and oxcarbazepine, along with two potential impurities and three formulation excipients

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Structurally related carbamazepine (CBZ) and oxcarbazepine (OX) are two of the most commonly used antipsychotic drugs. The main impurities of CBZ, as described in both the USP and the BP, are iminodibenzyl (IMD) and iminostilbene (IST). Meanwhile, for non-pharmacopeial OX, the declared impurities include CBZ and IST. Prescribed oral suspensions of CBZ and OX contain additives including methyl paraben (MP), propyl paraben (PP) and sorbic acid (SA) as preservatives. An HPTLC method was introduced and developed for resolving the interference between CBZ, OX, their impurities, and the suspension additives in a single run, in addition to their quantitation with a high sensitivity that satisfies the USP requirements for the detection and quantitation of drug impurities. In the developed HPTLC method, CBZ and OX were measured in the range of 40–4000 ng per band, while IMD, IST, MP, PP and SA were in the range of 20–2000 ng per band, using a mixture of hexane : ethylacetate : formic acid : acetic acid (8 : 2 : 0.5 : 0.3, by volume) and UV scanning at 254 nm. The greenness profile of the method was evaluated by two different tools, the analytical Eco-Scale and the Green Analytical Procedure Index (GAPI), then a comparison between their results was conducted. This is the first time that the studied drugs, along with their impurities and suspension additives, were analyzed by a HPTLC method in a single run and within the limits required by the USP guidelines.

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1. Introduction

Antipsychotics are a class of drugs known as neuroleptics and are used mainly for the management of psychotic disorders like paranoia, hallucinations or delusions, primarily in epilepsy, schizophrenia, and bipolar disorders (along with mood stabilizers).¹ Some of the most commonly used antipsychotic drugs are carbamazepine (CBZ) and oxcarbazepine (OX), that are structurally close to tricyclic antidepressants.²

Carbamazepine (CBZ) is officially a dibenzazepine derivative with psychotropic properties.² It is listed in both the British Pharmacopeia (BP)³ and the United States Pharmacopeia (USP).⁴ Additionally, CBZ has different impurities and related substances^{3,4} including iminodibenzyl (IMD) and iminostilbene (IST). OX is a CBZ derivative with similar actions.² OX is a non-official drug that has been found to have different impurities such as CBZ and IST.⁵ Moreover, CBZ and OX are individually

formulated in tablet and suspension dosage forms, marketed as Tegretol® and Trileptal®, respectively. The marketed oral suspensions of CBZ and OX, as labeled by the manufacturer, contain methyl paraben (MP), propyl paraben (PP) and sorbic acid (SA) as preservatives. The chemical structures of all of the studied compounds are given in Fig. 1.

Upon reviewing the literature, it was observed that CBZ can be measured in different biological fluids and in tablet dosage form by different methods that were summarized in the review article published by Datar⁶ in 2015. Recently, it was analyzed in its pharmaceutical formulation by HPLC^{7,8} and LC-MS^{9–12} methods. Likewise, CBZ was analyzed by different HPLC methods in the presence of its impurities.^{13,14} OX was estimated in tablets by TLC-densitometric¹⁵ and different HPLC^{16,17} methods. It was also determined in the presence of its degradation products or related substances by TLC-densitometric,¹⁸ HPLC^{5,19–21} and LC-MS/MS²² methods. Additionally, it was analyzed along with its metabolites by HPLC^{23,24} and LC-MS/MS^{25–27} methods. Similarly, different methods including HPLC,²⁸ LC-MS/MS,^{29,30} and supercritical fluid³¹ chromatographic methods were used for the determination of both CBZ and OX simultaneously in the presence of their metabolites.

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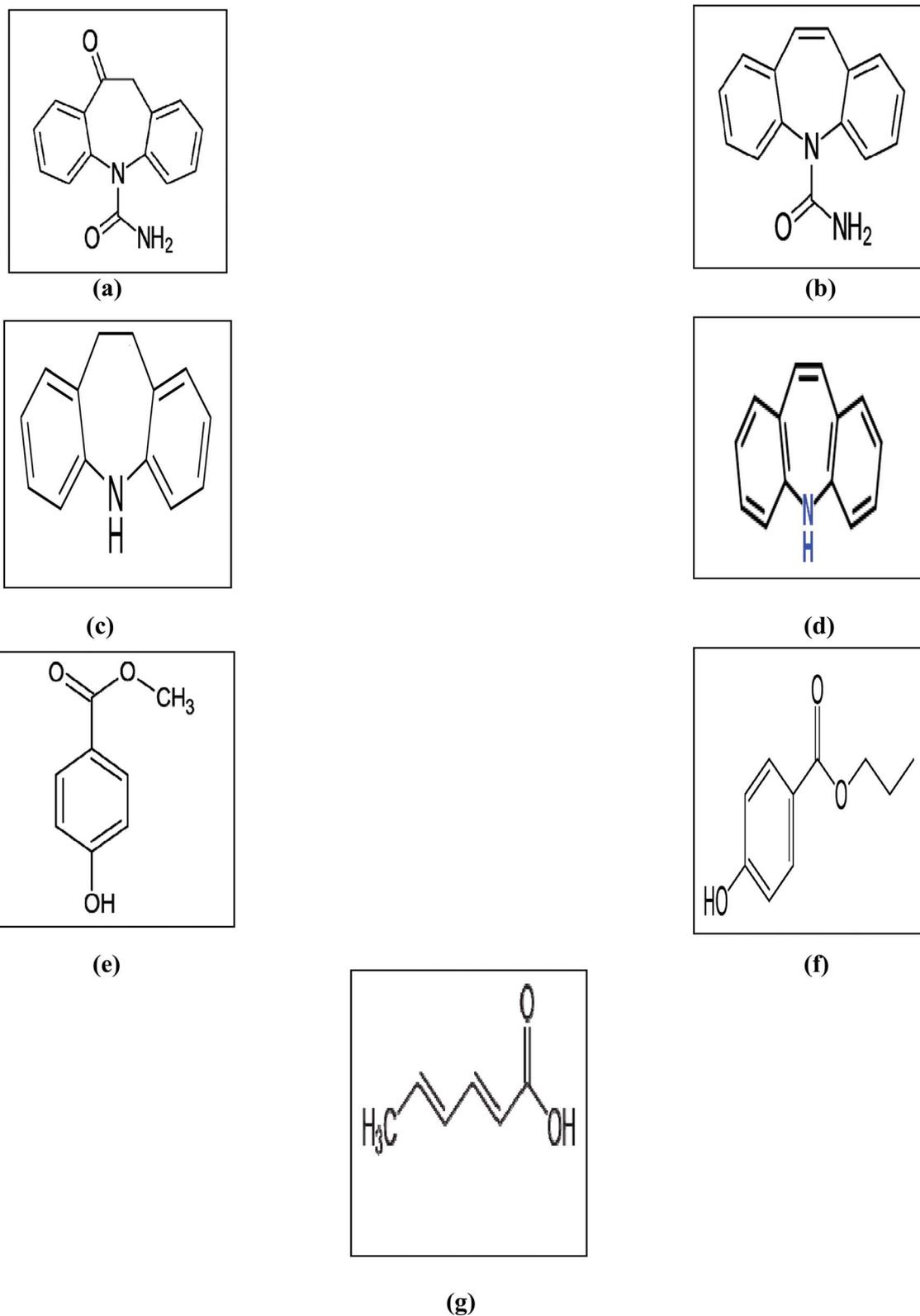


Fig. 1 The chemical structures of (a) OX, (b) CBZ, (c) IMD, (d) IST, (e) MP, (f) PP and (g) SA.

Pharmaceutical safety and efficacy are important matters in drug therapy. It has been reported that the negative effects caused by drug impurities significantly affect drug safety.

Therefore, the issues concerning the analytical estimation of impurities in pharmaceuticals are some of the most important subjects in modern pharmaceutical analysis.³² In the



USP,⁴ it is specified that the total impurities should not exceed a maximum limit of 0.5% relative to the main drug. HPTLC methods are widely used planar chromatographic methods, which are applied to analyze several samples in parallel. In these chromatographic methods, disposable stationary phases are used which do not need a lot of sample clean up.³³ Furthermore, the addition of MP, PP, and SA to oral suspensions was found to be a big problem during the analysis of active ingredients in pharmaceutical formulations.

Only one method has been previously developed in our laboratory for the determination of the studied components using a LC-MS-MS method.³⁴ Although it attains high sensitivity and selectivity, it needs expensive instruments and chemicals. Similarly, all the published methods analyzed either CBZ or OX in tablets only, even though interference due to the suspension excipients is expected. All of this, in addition to the pharmaceutical importance of the studied drugs, persuaded us to develop a selective and sensitive method that is capable of simultaneously resolving and quantifying the two drugs (CBZ and OX), along with IMD and IST (as impurities), and the formulation excipients (MP, PP, and SA).

Nowadays, the analytical community is interested in protecting human health and the environment through reducing the harmful impacts of hazardous chemicals by using more clean methodologies. Chemists have paid attention to the application of the concepts of green analytical chemistry (GAC) in different analytical methods. Furthermore, the compromise between increasing the quality of the results and decreasing the environmental impact of a method is considered to be a great challenge. Recently, different tools have been applied to assess the greenness profile of any developed method, of which the analytical Eco-Scale and Green Analytical Procedure Index (GAPI) tools are widely used.

The proposed HPTLC method is the first established stability indicating chromatographic method for the quantitation of the seven components with minimal analysis costs and time. Besides, the method has the advantage of its high sensitivity, especially for the analyzed impurities, which met the requirements stated in the USP⁴ recommendations.

2. Experimental

2.1. Instruments

– The TLC Scanner (CAMAG) was controlled by WINCATS software. Scanning was performed in the absorbance mode with a 20 mm s⁻¹ speed. A Linomat IV applicator was used for the band application with a 100 μL syringe (CAMAG, Muttenz, Switzerland). And the source of radiation used was a deuterium lamp. It was operated with a band width of 5 mm, and 3 × 0.45 mm were selected as the slit dimensions. The chromatogram output data were obtained as integrated peak areas. HPTLC plates with a 0.25 mm coat of silica gel 60F₂₅₄ (Merck, Germany) with dimensions of 15 × 20 cm were utilized.

– A Sonix TV ss-series ultra-sonicator (USA) was used.

2.2. Samples

2.2.1. Pure samples. Carbamazepine and oxcarbazepine were obtained from Novartis Co. with purities of 99.51% and 99.76%, respectively, according to their reported methods of analysis.^{4,5} IMD and IST were bought from Sigma-Aldrich (Chemie GmbH., Germany) with purities of 99.26% and 98.26%; respectively; according to their analysis certificates. MP, PP and SA were obtained from CID Co. Chemical Industries Development, Giza, Egypt with reported purities of 99.25%, 99.76%, and 99.25%, respectively, according to their analysis certificates.

2.2.2. Marketed samples. – Tegretol® tablets (Batch No. Y0487) and Tegretol® suspension (Batch No. Y1008) made by Novartis Pharma Co. were used. The labels stated that each tablet contained 200 mg of CBZ and the oral suspension contained 2% CBZ.

– Trileptal® tablets (Batch No. T1887) and Trileptal® suspension (Batch No. (W/V) H6725) were manufactured by Delpharm Huningue S.A.S., Huningue, France and licensed for Novartis Pharma AG, Basle, Switzerland. Their labels stated that they contained 300 mg of OX per tablet and 60 mg of OX per 1 mL suspension.

2.3. Solvents and chemicals

Analytical grade methanol, hexane, glacial acetic acid, formic acid, and ethyl acetate were obtained from El Nasr Pharmaceutical Chemicals Co., Abo-Zabaal, Giza, Egypt.

2.4. Stock standard solutions

– **Stock solutions of CBZ, OX, IMD, IST, MP, PP, and SA (0.5 mg mL⁻¹).** 50 mg of each component was accurately and separately weighed into seven separate 100 mL volumetric flasks, then dissolved in methanol.

2.5. Procedure

2.5.1. Linearity. Concentrations equivalent to 0.04–4 mg each of CBZ and OX, and 0.02–2 mg each of IMD, IST, MP, PP and SA were accurately and separately transferred from their particular stock solution (0.5 mg mL⁻¹) to seven individual 10 mL volumetric flasks. The volume was made up to 10 mL using methanol. Then, triplicates (10 μL) of each prepared solution were applied to HPTLC aluminum plates using an auto-sampler equipped with a syringe (100 μL) with an application rate of 0.1 μL s⁻¹. The used band width was adjusted to 5 mm and the space between the bands was 8.9 mm, 10 mm from the bottom edge of the plate. The chromatographic run was carried out in a glass jar saturated for 20 min with a mixture of hexane : ethyl acetate : formic acid : acetic acid (8 : 2 : 0.5 : 0.3, by volume) to a distance of about 8 cm. Subsequently, the developed plates were dried and scanned at 254 nm. The radiation source applied was a deuterium lamp and the software was WINCATS. The peak areas of the studied components were recorded and the calibration graphs were established using the calculated integrated peak areas and the corresponding concentrations of each component.



2.5.2. Analysis of the marketed samples

– For the tablet dosage forms (Tegretol® and Trileptal® tablets).

Five tablets of each were accurately and separately weighed and ground. 50 mg of each of the powdered tablets was accurately transferred into two separate 100 mL volumetric flasks. After that, 75 mL methanol was added and the solutions were sonicated for about 15 min, then filtered, and the residue was washed with 5 mL methanol twice (2×5 mL) and the washing solution was then added to the filtrate. Finally, the volume of the prepared solutions was made up with methanol to obtain solutions of 0.5 mg mL^{-1} of either CBZ or OX.

– For the suspension dosage forms. 1.25 mL of the Tegretol® suspension and 0.5 mL of the Trileptal® suspension were accurately and separately transferred into two 50 mL volumetric flasks. After that, methanol was used to make up the volume to the mark then these were shaken well to prepare stock solutions of 0.5 and 0.6 mg mL^{-1} of CBZ and OX, respectively.

– Suitable dilutions within the linearity ranges of both drugs were made after proper filtration of the prepared solutions. Then, the procedure was followed to construct the calibration curves. The previously computed regression equations for CBZ and OX were employed to obtain the recovered concentrations in the analyzed dosage forms.

– The technique of standard addition was also used by producing mixtures of standard CBZ or OX solutions with their respective tablet and suspension solutions. Subsequently, the proposed HPTLC method was adopted and the concentrations of CBZ and OX were calculated using their relative regression equations.

3. Results and discussion

Carbamazepine and OX are widely prescribed tricyclic antipsychotics that are available in several dosage forms such as tablets and suspensions.

Undoubtedly, pharmaceuticals intended for human consumption should be as fully characterized as possible. Assurance of a drug's quality and safety is attained by monitoring and controlling its impurities. The presence of such impurities prohibits the accurate determination of the drug. Therefore, there is a demand for developing and validating a highly sensitive and selective analytical method for the analysis of these drugs without interference from their impurities. Synthesis-related impurities are expected to be found in the bulk forms and pharmaceutical formulations of CBZ and OX. Pharmacopoeias set strict limits for the purity of most drugs to ensure their efficacy and safety. The USP⁴ declared that the maximum concentration of any individual impurity should not be more than 0.2%, while the concentration of all impurities should not be more than 0.5% of the active pharmaceutical ingredient. Routine analysis of drug purity necessitates a simple and robust analytical method that provides sufficient resolution, accuracy and sensitivity.

In this manuscript, efforts were made to develop a sensitive, specific and accurate HPTLC method for the resolution of the active drugs, their impurities, and the labeled suspension

additives. In addition, the environmental and health hazards of the developed method were reduced. The developed HPTLC method is more specific and economical than the official HPLC^{3,4} methods and any previously published chromatographic methods. In addition, it is the first developed method for resolving and quantifying the seven components using a single developing system and scanning wavelength. Moreover, it is the first developed method that has been applied to the determination of both CBZ and OX in their available suspension dosage forms, along with their suspension additives, MP, PP and SA.

One of the challenges in developing a specific method for resolving the cited mixture is the structural similarity between several pairs of the chosen components, like (CBZ and OX), (IMD and IST) and (MP and PP), leading to similar polarities for the members of each pair, and thus the same affinity with the stationary phase. In order to optimize the developed HPTLC method, different separation conditions were studied, such as the developing system composition, saturation time, different instrumental parameters, and the height of the used HPTLC plates.

The initial optimization steps started with the choice of the most appropriate developing system that affected the partition of the separated components between the stationary and mobile phases. Different systems were tested, like hexane : methanol (8 : 2, v/v), hexane : ethyl acetate (8 : 2, v/v), hexane : ethanol (8 : 2, v/v), and hexane : acetone (8 : 2, v/v). Unresolved spots between (CBZ and OX), (MP and PP) and (SA, IMD and IST) were obtained in all trials except when using hexane : ethyl acetate, which resulted in a slight separation between the studied components. The composition of the chosen developing system was then optimized by trying different ratios of hexane and ethyl acetate (from 7 : 3 to 9 : 1, v/v) where the best ratio was 8.5 : 1.5, v/v. The effect of the developing system pH was then studied using different volumes of acetic acid, formic acid, and ammonium hydroxide (0.2–0.5 mL each). No significant effect on the separation was found when using ammonium hydroxide (33%) solution. On the contrary, it was found that acetic acid was necessary to differentiate between the adjacent bands of OX and CBZ, while formic acid was essential for the good separation of MP, PP, SA, IMD and IST. Different combined ratios of formic acid and acetic acid were tried, and a developing system of hexane : ethyl acetate : formic acid : acetic acid (8.5 : 1.5 : 0.5 : 0.3, by volume) gave the greatest possible separation between the seven components. For the further improvement of the chromatographic resolution, HPTLC plates with different lengths were examined (10, 12, and 15 cm). The plate height was observed to significantly affect the separation efficiency and the chromatographic resolution between the seven components. The HPTLC plate of 15 cm length was the most suitable one. The effect of the saturation time of the stationary phase with the developing system was tested (15 and 30 min) and no significant effect of saturation time on the resolution was seen. Finally, different scanning wavelengths (215, 225, and 254 nm) were used in order to meet the detection and quantitation limits stated by the USP.⁴ Upon



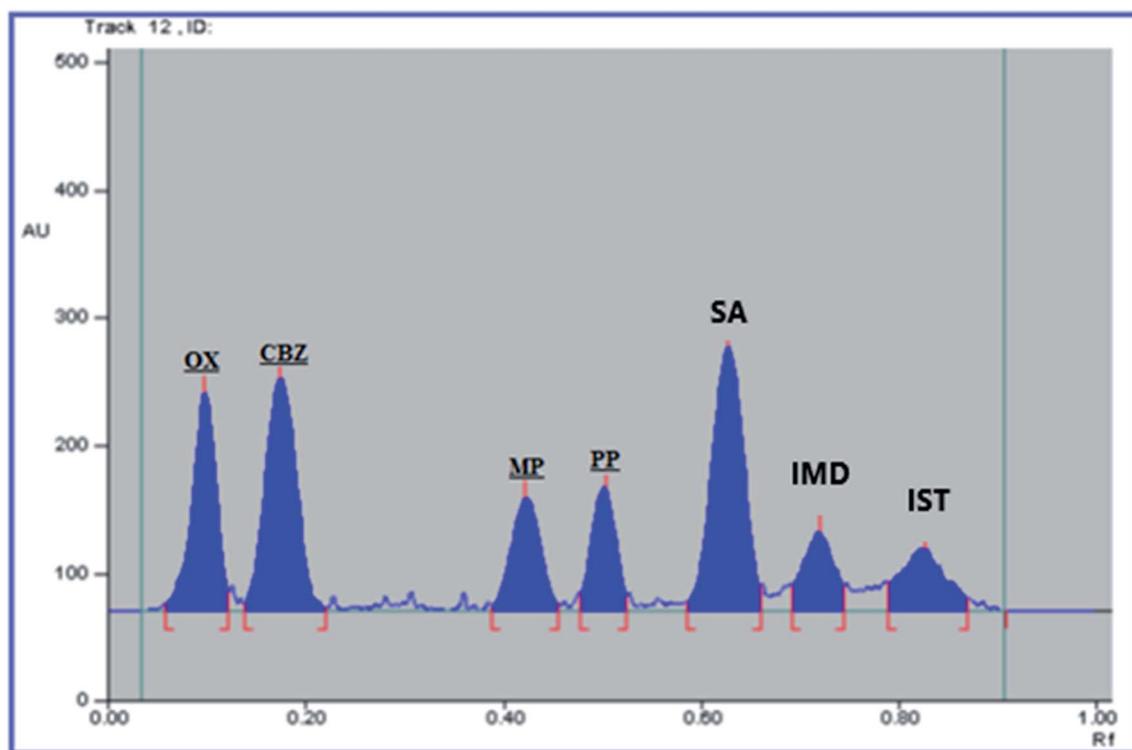


Fig. 2 HPTLC densitogram of a mixture of OX (500 ng per band), CBZ (500 ng per band), IMD (200 ng per band), IST (200 ng per band), MP (200 ng per band), PP (200 ng per band) and SA (200 ng per band).

using 254 nm as a scanning wavelength, the highest sensitivity for all the studied components was attained.

At the end, the optimum chromatographic conditions were: the developing system: hexane : ethyl acetate : formic acid : acetic acid (8.5 : 1.5 : 0.5 : 0.3, by volume), saturation time: 15 min, HPTLC plate height: 15 cm and scanning wavelength: 254 nm. The obtained R_f values were 0.08, 0.16, 0.43, 0.55, 0.62, 0.72, and 0.83 for OX, CBZ, MP, PP, SA, IST, and IMD, respectively. The HPTLC densitogram is shown in Fig. 2.

3.1. Application of the method

After the proposed method was optimized, it was applied to analyze different samples of the pure components at different concentration ranges to check the method's linearity. Linearity was achieved at different ranges: 40–4000 ng per band (for OX and CBZ) and 20–2000 ng per band (for IMD, IST, MP, PP, and SA). The regression equations were constructed using polynomial regression which gave a wider linear range for all the

Table 1 Regression and analytical parameters and results for the developed HPTLC method

Parameters	OX	CBZ	IMD	IST	MP	PP	SA
Linearity range (ng per band)	40–4000		20–2000		20–2000		
Slope	−237.84 ^a 3807.90 ^b	−615.48 ^a 6069.10 ^b	−1769 ^a 6937.4 ^b	−1512.10 ^a 11 537 ^b	−2265.70 ^a 13 314 ^b	−2631.60 ^a 12 313 ^b	−4209.80 ^a 18 288 ^b
Intercept	226.12	205.20	571.47	293.40	118.14	323.50	1050.50
Correlation coefficient (r)	0.9999	0.9996	0.9997	0.9998	0.9998	0.9997	0.9997
Accuracy (mean ± % RSD) ^c	99.84 ± 1.38	100.10 ± 1.82	99.81 ± 1.11	99.94 ± 2.04	99.49 ± 1.47	98.96 ± 1.52	97.66 ± 0.38
Precision (% RSD)							
Repeatability ^{d,e}	0.98	1.08	1.39	0.64	1.46	1.64	2.07
Intermediate precision ^{d,f}	1.81	1.16	2.09	2.04	2.40	2.26	2.27
LOD (ng per band) ^g	12.50	13.00	6.00	6.40	6.60	6.60	6.65
LOQ (ng per band) ^g	38.00	39.00	18.50	19.25	19.75	19.90	20.00

^a Coefficient 1. ^b Coefficient 2. ^c Accuracy: mean of 9 concentrations for each component. ^d Average of 3 experiments. ^e Standard deviation of 3 concentrations of each component: (100, 1000 and 3000 ng per band) for OX and CBZ, and (100, 500 and 1500 ng per band) for IMD, IST, MP, PP, and SA, on the same day. ^f Standard deviation of 3 concentrations of each component: (100, 1000 and 3000 ng per band) for OX and CBZ, and (100, 500 and 1500 ng per band) for IMD, IST, MP, PP, and SA, on three successive days. ^g Where $LOD = 3.3 \times SD/S$ and $LOQ = 10 \times SD/S$, where SD is the standard deviation of the intercept and S is the slope of the calibration curve.



Table 3 Statistical analysis of the results obtained by the analysis of pure carbamazepine and oxcarbazepine using the developed HPTLC method and the reference ones

Component Method	CBZ		OX	
	HPTLC	Reference ^a	HPTLC	Reference ^b
Mean \pm SD	100.10 \pm 1.83	99.82 \pm 2.03	99.84 \pm 1.38	99.51 \pm 1.75
Variance	3.33	4.13	1.91	3.07
<i>n</i>	7	6	9	6
Student's <i>t</i> -test	0.26 (2.16) ^c		0.40 (2.16) ^c	
<i>F</i> -Test	1.24 (4.39) ^c		1.61 (3.69) ^c	

^a HPLC method using a (4.6 mm \times 250 mm) cyano column with 5–10 μ m silica particles, a mobile phase of water/methanol/tetrahydrofuran (85 : 12 : 3, by volume) containing 0.22 mL formic acid and 0.5 mL triethylamine, flow rate = 1.5 mL min⁻¹, and UV detection at 230 nm.

^b UHPLC method using a C18 column (100 \times 2.1 mm, 1.9 μ m particle size) using a gradient program of water (mobile phase A) and acetonitrile (mobile phase B), flow rate = 0.5 mL min⁻¹, UV detection at 254 nm, and the column temperature was set to 30 $^{\circ}$ C. ^c The values between parentheses correspond to the theoretical values of *t* and *F* (*P* = 0.05).

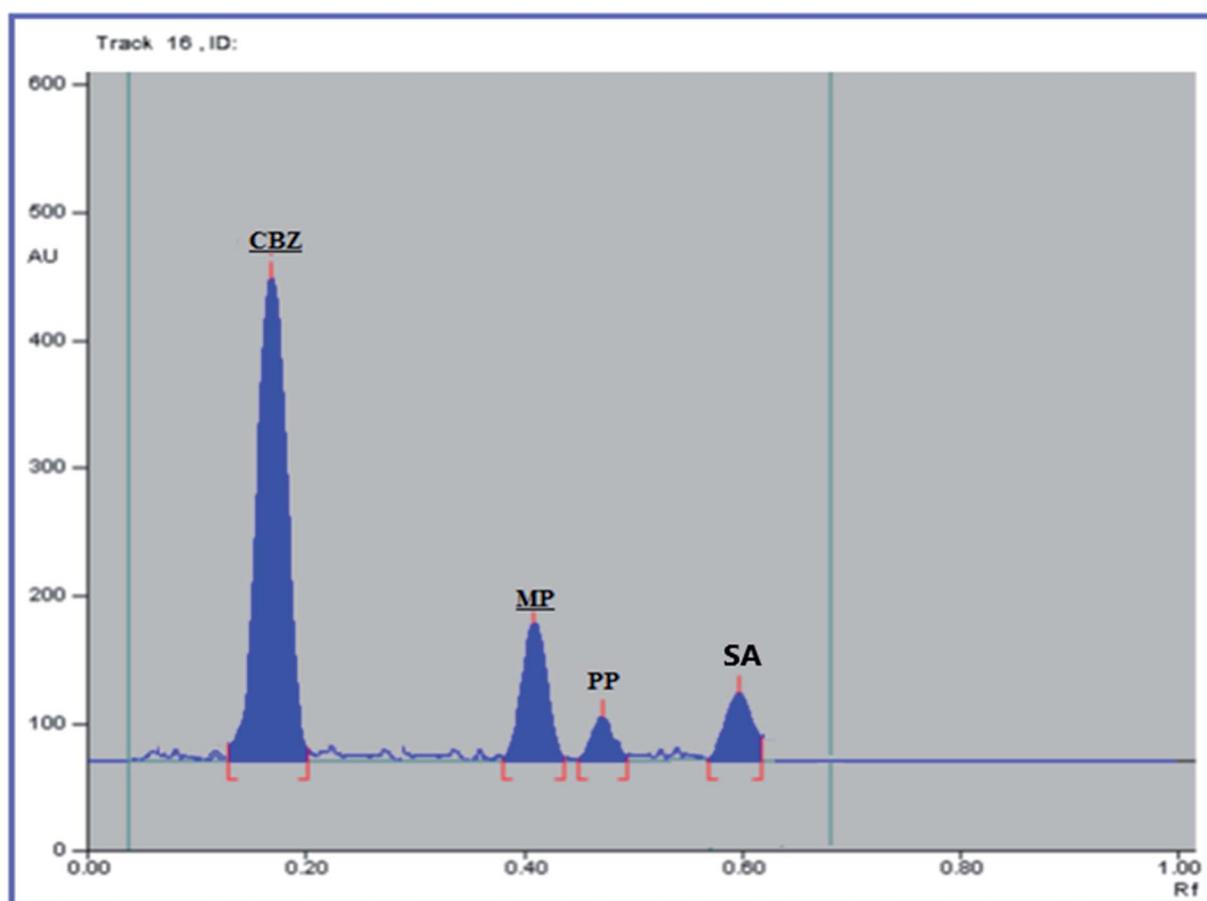


Fig. 3 HPTLC densitogram of the mixture of the Tegretol® suspension (1000 ng per band CBZ).

amount of acetic acid significantly affected the resolution for both CBZ and OX. So, the amount of acetic acid should be exactly adjusted to 0.3 mL (in each 10 mL developing system) in order to obtain reproducible results.

3.4. System suitability testing parameters

Different parameters like the symmetry of peaks, the resolution, and selectivity factors were computed in order to test the

system's performance and the separation efficiency. The obtained results were within the acceptable limits, as shown in Table 4, which demonstrated the high efficiency of the chromatographic separation.

3.5. Greenness profile assessment

In this work, the environmental impact of the newly developed chromatographic method was considered for all steps,



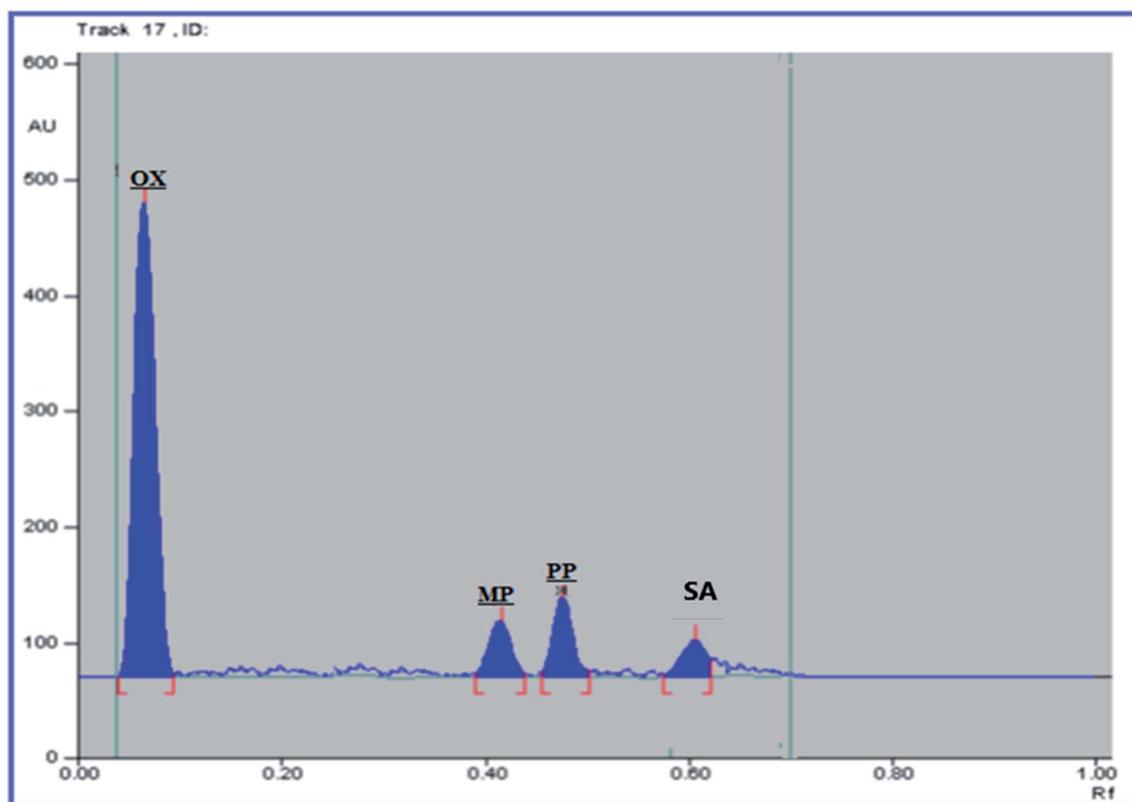


Fig. 4 HPTLC densitogram of the Trileptal® suspension (1000 ng per band OX).

Table 4 Parameters of system suitability for the developed HPTLC method for the determination of the studied components

Parameters	OX	CBZ	MP	PP	SA	IST	IMD	Reference values ³³
R_f	0.08 ± 0.02	0.16 ± 0.02	0.42 ± 0.03	0.55 ± 0.02	0.62 ± 0.03	0.72 ± 0.02	0.83 ± 0.01	—
Capacity factor (K)	11.5	5.25	1.33	0.82	0.61	0.39	0.18	—
Symmetry factor (T)	0.92	1.07	1.04	1.00	0.97	1.00	1.05	~1
Resolution (R_s)	1.13	3.51	1.52	2.15	1.50	1.70	1.70	>1.5
Selectivity (α)	2.19	3.95	1.62	1.34	1.56	2.17	2.17	>1

including sample preparation, method, development, analysis solvents, and the waste produced.^{35–39} In the present study, two approaches were applied to assess the greenness of the method. It is ideal to apply different tools to obtain a deeper view of the greenness of the method.

– **Analytical Eco-Scale.** This is a semi-quantitative tool which is related to a penalty point system.³⁹ It depends on calculating the penalty points of different approaches, including the amount and hazardousness of the used reagents, the generated waste, energy consumption and occupational risks, then subtracting from a base of 100 to acquire the corresponding analytical Eco-Scale score. The higher the score, the greener the developed method. The results given in Table 5 show that the proposed HPTLC method achieved a score of lower than 75 yet higher than 50, thus it can be considered as an acceptable green method.

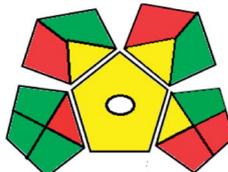
– **Green Analytical Procedure Index (GAPI).** The Green Analytical Procedure Index (GAPI) is a new approach for the

evaluation of a method's greenness profile which is a hybrid of the National Environmental Method Index (NEMI) and the analytical Eco-Scale approaches, and is based on the evaluation of the green characteristics of an entire methodology though all steps, starting from sample collection to the final determination step. When applying this assessment tool, a specific symbol with five pentagrams is used to check the greenness of each step, with color coding from green to yellow to red representing low, medium and high environmental impact, respectively.³⁹ Looking at the GAPI pentagrams in Table 5, they reveal that the proposed method could be considered as an acceptable green method with little environmental impact, since it had six green shaded, four yellow shaded and only five red shaded fields.

The results of the two approaches used agreed with each other, proving the small environmental impact of the proposed HPTLC method.



Table 5 Greenness assessment of the proposed chromatographic method, according to the analytical Eco-Scale and GAPI

Parameters	Developed method	Penalty points		Penalty points			
		Reported HPLC ⁴	Reported HPLC ⁵	Reported HPLC ⁴	Reported HPLC ⁵		
Reagents	Consumed volume = volume of the developing system per run/number of samples on the HPTLC plate	Hexane Consumed volume = 4 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 4	8	Water (Green nonhazardous solvent)	0	Water (Green nonhazardous solvent)	0
	PP of solvent = subtotal PP × number of pictograms × signal word	Ethylacetate Consumed volume = 1 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 2	4	Methanol Consumed volume = 1.8 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 3	6	Methanol Consumed volume = 2.4 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 3	6
		Formic acid Consumed volume = 0.25 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 3	6	Tetrahydrofuran Consumed volume = 0.45 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 3	6	Tetrahydrofuran Consumed volume = 0.6 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 3	6
		Acetic acid Consumed volume = 0.15 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 2	4	Formic acid Consumed volume = 0.033 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 3	6	Formic acid Consumed volume = 0.004 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 3	6
		—	—	Triethylamine Consumed volume = 0.075 mL <i>Subtotal PP</i> = 1 [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 4	8	Triethylamine Consumed volume = 0.01 mL [solvent < 10 mL] <i>Signal word</i> = 2 danger [more severe hazard = 2] <i>No. of pictograms</i> = 4	8
Instruments Energy	≤1.5 kWh per sample	1	≤1.5 kWh per sample	1	≤1.5 kWh per sample	1	
Occupational hazard	Not a closed system	3	Analytical process hermetization	0	Analytical process hermetization	0	
Centrifuge		1	—	—	—	—	
Wastes	<10 mL	3	>10 mL	5	>10 mL	5	
Total penalty points		30		32		32	
Analytical Eco-Scale total score		70		68		68	
Comment	Acceptable green analytical method		Acceptable green analytical method		Acceptable green analytical method		
GAPI assessment							



3.6. Comparison with previously published chromatographic methods

The developed HPTLC method has a higher selectivity than the formerly published TLC-densitometric methods^{15,18} since it is applied for the simultaneous separation and quantitation of the studied drugs, their impurities and the suspension additives. The compared official HPLC methods^{3,4} for the analysis of CBZ and its related substances have similar greenness profiles, as proved by the results in Table 5. Conversely, the developed HPTLC method is greener than the published HPLC methods^{7,19–21} since acetonitrile is the organic modifier used in these methods and it has a high environmental impact. Compared with the other published HPLC methods^{7,8,13,14,16,17,19,20,21} and LC-MS-MS methods^{9–12,22} for the analysis of either CBZ or OX in their tablet formulations, the suggested HPTLC method has the advantage of saving time and money. Additionally, simple sample preparation steps are needed as well as low cost solvents, chemicals and instruments. However, the published LC-MS-MS methods have higher sensitivities for CBZ and OX but were not applied for the quantitation of either their impurities or excipients.

4. Conclusion

For the first time, a selective, specific and accurate HPTLC method with little environmental impact was introduced for the simultaneous determination of the commonly used antipsychotic drugs CBZ and OX in the presence of their reported impurities and formulation excipients, in a single run. The suggested method is superior to any other formerly established chromatographic methods and is able to measure CBZ, OX, IMD, IST, MP, PP and SA with high sensitivity, which follows the USP regulations for drug impurity profiling. The HPTLC method has the advantages of lower solvent consumption, a short analysis time, and several samples can be determined simultaneously in the same run, which are important factors for quality control laboratories. The developed method can be used as an alternative to LC methods in laboratories lacking the facilities for LC instrumentation.

Conflicts of interest

The authors certify that they have no affiliations with or involvement in any organization or entity with any financial interest or non-financial interest in the subject matter or materials discussed in this manuscript.

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